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Genome Sequence of *Serratia plymuthica* A153, a Model Rhizobacterium for the Investigation of the Synthesis and Regulation of Haterumalides, Zeamine, and Andrimid

Announcements

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The rhizobacterium *Serratia plymuthica* A153 is a Gram-negative bacterium belonging to the family *Enterobacteriaceae*. Here, we present the genome sequence of this strain, which produces multiple bioactive secondary metabolites, including the halogenated macrolide oocydin A, the polyamino antibiotic zeamine, and the bacterial acetyl-CoA carboxylase inhibitor andrimid.

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S*erratia plymuthica* strains are widely distributed, and commonly found associated with plant roots (1). They are effective biocontrol agents and plant growth-promoting bacteria, mainly due to their capacity to produce exoenzymes, phytohormones, and various secondary metabolites, coupled with their ability to induce systemic resistance (1, 2).

Serratia plymuthica A153 was isolated from the rhizosphere of wheat (3) and synthesizes multiple bioactive secondary metabolites, including several antifungal, antioomycete, and anticancer haterumalides/oocydins (4), the bacterial acetyl-CoA carboxylase inhibitor andrimid (5), the polyamino antibiotic zeamine (6), and the broad spectrum antifungal compound pyrrolnitrin (4). Serratia plymuthica A153 was used to identify the biosynthetic cluster encoding synthesis of the haterumalide oocydin A (7). Furthermore, it was used as a model bacterium for the study of the regulation of multiple bioactive non-ribosomal peptides and polyketides (5, 6, 8)—research which was greatly facilitated by the facile genetic tractability of A153 and the isolation of a highly efficient generalized transducing phage, ϕ MAM1 (9).

The sequencing of the genomic DNA of S. plymuthica A153 was performed at the Department of Biochemistry (University of Cambridge) using 454 DNA pyrosequencing technology on a picotiter plate for a Roche Applied Science Genome Sequencer FLX system. The 454 data were de novo assembled using Newbler v2.6. The assembly used 308,585 reads (129 MB of raw data) to give an approximately 22× coverage of the estimated genome size and resulted in a total of 24 contigs larger than 500 bp. The average contig size was 230,980 bp and the largest contig was 1,516,666 bp. The contigs were ordered and oriented based on the wholegenome sequences of the Serratia plymuthica strains AS9 (10), AS12 (11), and 4Rx13 (GenBank accession no. CP006250). Traditional Sanger sequencing was used to close the gaps between contigs. The genome was automatically annotated using NCBI Prokaryotic Genomes Annotation Pipeline (PGAP) version 3.0 (http://www.ncbi.nlm.nih.gov/genome/annotation_prok).

The assembled genome of Serratia plymuthica A153 consists of

2 large contigs and includes 5,475,375 bp, with an overall G+C content of 55.94%. Automated genome annotation predicted 4,809 protein-coding sequences (CDSs), 30 pseudogenes, 21 rRNA operons, 81 tRNA genes, and 11 noncoding RNAs. In addition to the gene clusters responsible for the biosynthesis of oocydin A, zeamine, and andrimid, antiSMASH (12) predicted 6 additional clusters putatively involved in the synthesis of nonribosomal peptides and polyketides. Genome comparison analyses revealed that the genome of A153 shows high sequence homology with the genomes of the Serratia plymuthica strains AS9 (10), AS12 (11), AS13 (13), S13 (14), RVH1 (15), 4Rx13 (GenBank accession no. CP006250), and V4 (16). However, these strains lack several clusters for polyketide and non-ribosomal peptide biosynthesis that are present in A153. The sequencing of the genome of S. plymuthica A153 will enable further research on the biosynthesis and regulation of both the known and putatively novel secondary metabolites produced by this strain.

Nucleotide sequence accession number. The sequences obtained by this whole-genome shotgun project have been deposited in DDBJ/EMBL/GenBank under the accession number LRQU00000000.

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REFERENCES

- De Vleesschauwer D, Hofte M. 2007. Using Serratia plymuthica to control fungal pathogens of plants. CAB Rev 2:1–12. http://dx.doi.org/ 10.1079/PAVSNNR20072046.
- 2. Saha D, Purkayastha GD, Saha A. 2012. Biological control of plant diseases by *Serratia* species: a review or a case of study, p. 99–115. In Goyal A, Maheshwari P (ed.), Frontiers on recent developments in Plant Science, vol 1, chapter 7. Bentham Science Publishers, United Arab Emirates.
- Åström B, Gerhardson B. 1988. Differential reactions of wheat and pea genotypes to root inoculation with growth-affecting rhizosphere bacteria. Plant Soil 109:263–269. http://dx.doi.org/10.1007/BF02202093.
- 4. Levenfors JJ, Hedman R, Thaning C, Gerhardson B, Welch CJ. 2004. Broad-spectrum antifungal metabolites produced by the soil bacterium Serratia plymuthica A153. Soil Biol Biochem 36:677-685. http:// dx.doi.org/10.1016/j.soilbio.2003.12.008.
- Matilla MA, Nogellova V, Morel B, Krell T, Salmond GPC. 23 February 2016. Biosynthesis of the acetyl-CoA carboxylase-inhibiting antibiotic, andrimid, in *Serratia* is regulated by Hfq and the LysR-type transcriptional regulator, AdmX. Environ Microbiol. http://dx.doi.org/10.1111/1462 -2920.13241.
- 6. Hellberg JEEU, Matilla MA, Salmond GPC. 2015. The broad-spectrum antibiotic, zeamine, kills the nematode worm *Caenorhabditis elegans*. Front Microbiol 6:137. http://dx.doi.org/10.3389/fmicb.2015.00137.
- Matilla MA, Stöckmann H, Leeper FJ, Salmond GPC. 2012. Bacterial biosynthetic gene clusters encoding the anti-cancer haterumalide class of molecules: biogenesis of the broad spectrum antifungal and antioomycete compound, oocydin A. J Biol Chem 287:39125–39138. http:// dx.doi.org/10.1074/jbc.M112.401026.
- 8. Matilla MA, Leeper FJ, Salmond GPC. 2015. Biosynthesis of the antifungal haterumalide, oocydin A, in *Serratia*, and its regulation by quorum sensing, RpoS and Hfq. Environ Microbiol 17:2993–3008. http:// dx.doi.org/10.1111/1462-2920.12839.
- 9. Matilla MA, Salmond GPC. 2014. The Viunalikevirus, bacteriophage φ MAM1, is a broad host range, high efficiency generalised transducing phage that infects environmental and clinical isolates of the enterobacteria, *Serratia* and *Kluyvera*. Appl Environ Microbiol 80:6446–6457. http://dx.doi.org/10.1128/AEM.01546-14.

- Neupane S, Högberg N, Alström S, Lucas S, Han J, Lapidus A, Cheng JF, Bruce D, Goodwin L, Pitluck S, Peters L, Ovchinnikova G, Lu M, Han C, Detter JC, Tapia R, Fiebig A, Land M, Hauser L, Kyrpides NC, Ivanova N, Pagani I, Klenk HP, Woyke T, Finlay RD. 2012. Complete genome sequence of the rapeseed plant-growth promoting *Serratia plymuthica* strain AS9. Stand Genomic Sci 6:54–62. http://dx.doi.org/10.4056/ sigs.2595762.
- Neupane S, Finlay RD, Alström S, Goodwin L, Kyrpides NC, Lucas S, Lapidus A, Bruce D, Pitluck S, Peters L, Ovchinnikova G, Chertkov O, Han J, Han C, Tapia R, Detter JC, Land M, Hauser L, Cheng JF, Ivanova N, Pagani I, Klenk HP, Woyke T, Högberg N. 2012. Complete genome sequence of *Serratia plymuthica* strain AS12. Stand Genomic Sci 6:165–173. http://dx.doi.org/10.4056/sigs.2705996.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0-a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43: W237–W243. http://dx.doi.org/10.1093/nar/gkv437.
- 13. Neupane S, Finlay RD, Kyrpides NC, Goodwin L, Alström S, Lucas S, Land M, Han J, Lapidus A, Cheng JF, Bruce D, Pitluck S, Peters L, Ovchinnikova G, Held B, Han C, Detter JC, Tapia R, Hauser L, Ivanova N, Pagani I, Woyke T, Klenk HP, Högberg N. 2012. Complete genome sequence of the plant-associated *Serratia plymuthica* strain AS13. Stand Genomic Sci 7:22–30. http://dx.doi.org/10.4056/sigs.2966299.
- 14. Müller H, Fürnkranz M, Grube M, Berg G. 2013. Genome sequence *of Serratia plymuthica* strain S13, an endophyte with germination- and plant-growth-promoting activity from the flower of styrian oil pump-kin. Genome Announc 1(4):e00594-13. http://dx.doi.org/10.1128/genomeA.00594-13.
- Van Houdt R, Van der Lelie D, Izquierdo JA, Aertsen A, Masschelein J, Lavigne R, Michiels CW, Taghavi S. 2014. Genome sequence of Serratia plymuthica RVH1, isolated from a raw vegetable-processing line. Genome Announc 2(1):e00021-14. http://dx.doi.org/10.1128/genomeA.00021-14.
- 16. Cleto S, Van der Auwera G, Almeida C, Vieira MJ, Vlamakis H, Kolter R. 2014. Genome sequence of *Serratia plymuthica* V4. Genome Announc 2(3):e00340-14. http://dx.doi.org/10.1128/genomeA.00340-14.