

RESEARCH ARTICLE

Characterization of Glucose-Related Metabolic Pathways in Differentiated Rat Oligodendrocyte Lineage Cells

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Although oligodendrocytes constitute a significant proportion of cells in the central nervous system (CNS), little is known about their intermediary metabolism. We have, therefore, characterized metabolic functions of primary oligodendrocyte precursor cell cultures at late stages of differentiation using isotope-labelled metabolites. We report that differentiated oligodendrocyte lineage cells avidly metabolize glucose in the cytosol and pyruvate derived from glucose in the mitochondria. The labelling patterns of metabolites obtained after incubation with [1,2-¹³C]glucose demonstrated that the pentose phosphate pathway (PPP) is highly active in oligodendrocytes (approximately 10% of glucose is metabolized via the PPP as indicated by labelling patterns in phosphoenolpyruvate). Mass spectrometry and magnetic resonance spectroscopy analyses of metabolites after incubation of cells with [1-¹³C]lactate or [1,2-¹³C]glucose, respectively, demonstrated that anaplerotic pyruvate carboxylation, which was thought to be exclusive to astrocytes, is also active in oligodendrocytes. Using [1,2-¹³C]acetate, we show that oligodendrocytes convert acetate into acetyl CoA which is metabolized in the tricarboxylic acid cycle. Analysis of labelling patterns of alanine after incubation of cells with [1,2-¹³C]acetate and [1,2-¹³C]glucose showed catabolic oxidation of malate or oxaloacetate. In conclusion, we report that oligodendrocyte lineage cells at late differentiation stages are metabolically highly active cells that are likely to contribute considerably to the metabolic activity of the CNS.

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Key words: oligodendroglia, energy metabolism, glucose, ¹³C, mitochondria, glycolysis, acetate, pyruvate carboxylation

Introduction

Oligodendrocytes make up a large proportion of the cells in the central nervous system (CNS). Although oligodendrocytes are vulnerable to low energy conditions (Lyons and Kettenmann, 1998; Yan and Rivkees, 2006), their metabolic properties, including their glucose metabolism, have not been investigated in depth (Amaral et al., 2013). In contrast, the metabolic interactions between neurons and astrocytes, have received considerable attention since their discovery in the 1970s (van den Berg and Garfinkel, 1971). Specifically, the shuttling of glutamine–glutamate– γ -Aminobutyric acid (GABA) between astrocytes and neurons is thought to be fun-

damentally important for neuronal function. Because neurons themselves are unable to generate essential precursors of glutamate, GABA, and aspartate, they depend on the supply of glutamine as a precursor from astrocytes for the production of neurotransmitters (glutamate in 90% of the synapses, and GABA in 5%) (Attwell and Laughlin, 2001). In this context, glucose plays a central role as the key molecular building block that is used to synthesize glutamate, GABA, and aspartate.

Glucose is primarily metabolized to pyruvate via glycolysis in the cytosol. Stepwise conversion of a single glucose molecule into two pyruvate molecules generates two molecules of

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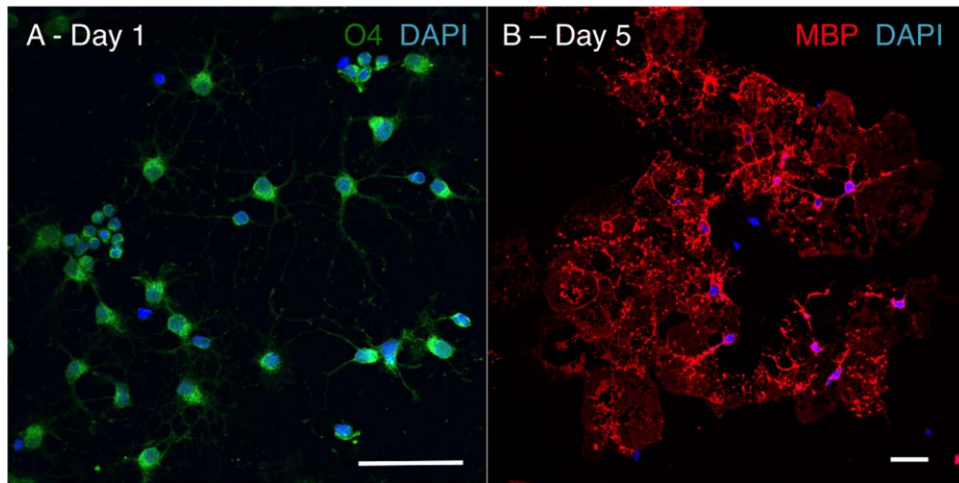


FIGURE 1: Purity of the primary cultures of rat oligodendrocytes. Oligodendrocyte precursor cells were isolated from mixed glia cultures and cultured in Sato's medium + 0.05% FCS to induce differentiation. At day 1 of differentiation, more than 93% of the cells expressed the oligodendroglial lineage marker O4 (A) and at 5 days of differentiation, approximately 65% of the cells expressed myelin basic protein (MBP), a marker of mature oligodendrocytes (B). Scale bars, 50 μm . [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

Adenosine triphosphate (ATP). These reactions are not oxygen dependent. Glucose metabolism can also take an alternative route via a biosynthetic pathway termed *pentose phosphate pathway* (PPP). This complex detour bypasses several steps of glycolysis. In the first, *oxidative phase* of the PPP, NADP^+ is converted into Nicotinamide adenine dinucleotide phosphate (NADPH). NADPH acts as a reducing agent that may participate in lipid and steroid synthesis or in the regeneration of glutathione and thioredoxin, which are involved in the cell's defense mechanism against oxidative stress. In the second phase of the PPP, 5-carbon sugars are nonoxidatively synthesized. The PPP joins the glycolytic pathway at the level of glyceraldehyde-3-phosphate (GA3P) and fructose-6-phosphate (fructose-6P). Fructose-6P is subsequently converted into pyruvate, which constitutes the endpoint of both glycolysis and the PPP.

In the presence of oxygen, the pyruvate produced by glycolysis or by the PPP can be converted to acetyl CoA by the pyruvate dehydrogenase (PDH) complex, and subsequently metabolized in the mitochondrial tricarboxylic acid (TCA) cycle, to further produce ATP via coupling to the mitochondrial electron transport chain. Alternatively, pyruvate can be (reversibly) converted into lactate in the cytosol, which results in the production of NAD^+ from NADH. Net synthesis of TCA cycle intermediates and related compounds, including glutamate and glutamine, depend on *anaplerotic* replenishment of intermediates in the TCA cycle. In the brain, this is mediated by pyruvate carboxylase (PC; Patel, 1974). Pyruvate carboxylation was shown to be absent in neurons, but present in astrocytes (Cesar and Hamprecht, 1995; Hertz et al., 1980; Shank et al., 1985; for review, see Sonnewald and Rae, 2010). Consequently, neurons are thought to depend on astrocytes as an external source of glu-

tamine for the production of neurotransmitters. Conversion of pyruvate by PC generates a "new" molecule of oxaloacetate. Oxaloacetate may subsequently condense with acetyl CoA to synthesize the TCA cycle intermediate citrate, which, after several steps, is converted to α -ketoglutarate, from which glutamate can be formed by transamination or deamination. In a subsequent step, glutamine synthetase, which is known to be expressed in astrocytes (Martinez-Hernandez et al., 1977; Norberg and Martinez-Hernandez, 1979), is able to convert glutamate into glutamine (see Fig. 1 in Amaral et al., 2013).

In the gray matter, glutamate, released from neuronal synapses during glutamatergic neurotransmission, is mainly taken up by astrocytes (Gegelashvili and Schousboe, 1997, 1998). The drain of glutamate from signalling neurons is subsequently compensated by a reverse flow of glutamine from astrocytes back to the neurons. This cross flow of glutamate and glutamine is often referred to as the glutamate–glutamine cycle (McKenna et al., 2012; see Fig. 1 in Amaral et al., 2013). Because glutamine released by astrocytes also functions as a precursor for the production of the inhibitory neurotransmitter GABA via conversion to glutamate (Reubi et al., 1978; Sonnewald et al., 1993b), metabolic interactions between astrocytes and neurons are thought to consist of a glutamate–glutamine and a glutamine–glutamate–GABA cycle.

How can oligodendrocytes contribute to the metabolic interactions in the CNS? We have argued that, instead of being restricted to closed-loop interactions between astrocytes and neurons, intercellular shuttling of metabolites may occur between all three major cell groups of the CNS: neurons, astrocytes, and oligodendrocytes (Amaral et al., 2013). The limited understanding of the metabolic role of oligodendrocytes in the brain was further highlighted in two recent studies,

which, for the first time, proposed a link between glycolytic metabolism in oligodendrocytes and axonal integrity and function (Funfschilling et al., 2012; Lee et al., 2012).

The aim of this study was to elucidate basic metabolic pathways for glucose catabolism and the anaplerotic replenishment of TCA cycle intermediates in oligodendrocytes. For this purpose, we incubated primary cultures enriched for mature oligodendrocytes in medium containing [1,2-¹³C]glucose, [1,6-¹³C]glucose, [1-¹³C]lactate, or [1,2-¹³C]acetate and analyzed cell extracts and medium using mass spectrometry or nuclear magnetic resonance (NMR) spectroscopy. We found that oligodendrocytes have extensive PPP activity. Furthermore, oligodendrocytes were able to anaplerotically replenish the TCA cycle, via pyruvate carboxylation, and cataplerotically recycle pyruvate. Our data also show that oligodendrocytes are able to convert [1,2-¹³C]acetate into acetyl CoA. Our results establish hallmarks of the cellular metabolism of oligodendrocytes under physiological conditions. They may also be useful for future studies investigating altered oligodendrocyte function and injury in diseases that involve glutamate toxicity and impaired mitochondria function, including, for example, hypoxic-ischemic damage, and multiple sclerosis (MS; Kostic et al., 2013; Mifsud et al., 2014; Pitt et al., 2000; Simonishvili et al., 2013).

Materials and Methods

Materials

Cell culture reagents were purchased from Sigma (Dorset, UK)—Dulbecco's modified eagle's medium (DMEM), minimum essential medium eagle (MEM), L-glutamine, poly-L-lysine (PLL), papain—or Life Technologies (Paisley, UK)—fetal bovine serum (FBS), penicillin-streptomycin (pen-strep), trypsin-Ethylenediaminetetraacetic acid (EDTA), phosphate buffered saline (PBS). ¹³C-labelled compounds were obtained from Cambridge Isotope Laboratories, MA. The mass spectrometry derivatization reagents MTBSTFA (*N*-methyl-*N*-(*tert*-butyldimethylsilyl) trifluoroacetamide), MSTFA (*N*-Methyl-*N*-(trimethylsilyl) trifluoroacetamide) and the *t*-BDMS-Cl (*tert*-butyldimethylchlorosilane) were purchased from Regis Technologies (Morton Grove, IL). All other chemicals were of the purest grade available from Sigma (Dorset, UK).

Preparation of Primary Oligodendrocyte Precursor Cell Cultures

Primary mixed glia cultures were isolated from neonatal Sprague Dawley rat (postnatal day 0–2) forebrains following a standard protocol (Baer et al., 2009). Pups were euthanized according to “Schedule 1” regulations from the Home Office Animal Procedures Committee UK. Cells were cultured for 10–15 days in DMEM supplemented with 10% FBS, 1% pen-strep, and 4 mM glutamine, and kept under a humidified atmosphere at 37°C and 7% CO₂. Oligodendrocyte precursor cells (OPCs) were subsequently isolated using a step-based shake-off protocol and cultured in Sato's medium on PLL-coated plates (Baer et al., 2009). To induce differentiation,

OPCs were cultured in Sato's medium supplemented with 0.5% fetal calf serum (FCS). The cell culture medium was replaced by fresh medium at day two of differentiation. For all *in vitro* experiments, only cultures with >93% purity (determined based on O4 immunostaining) were used (Fig. 1). Quantification of the number of glial fibrillary acidic protein (GFAP)-positive cells after 5 days of differentiation, indicated that astrocytes comprised 7–10% of total cells (data not shown).

Preparation of Rat Cortical Astrocyte Cultures

Astrocytes were prepared from the same mixed glia cultures used for OPC isolation, following an adapted protocol described in Amaral et al. (2014). After the shake-off (which eliminates microglia and most oligodendrocytes from the cultures), the remaining cells, highly rich in proliferative astrocytes, were seeded into new T75 flasks (1:3 dilution) using DMEM supplemented with 10% (v/v) FBS, 4 mM glutamine, and 1% (v/v) pen-strep and allowed to reach confluence (approx. 5–7 days). Then, cells were collected with trypsin-EDTA and seeded in 6 well plates at a density of 5×10^4 cells/well for the [1-¹³C]lactate experiments. Experiments were performed when cells reached confluence.

Incubations with ¹³C-Labelled Compounds

Mature OPC cultures (after 5 days in differentiation medium) were cultured in 6 well plates at a cell density of 4×10^5 cells/well. Prior to incubation, cells were washed once with PBS and incubated with 2 mL Sato's medium prepared from a glucose, glutamine, and pyruvate-free DMEM (Sigma D5030) supplemented with 0.5% FCS and 2 mM [1,6-¹³C]glucose, 2 mM [1,2-¹³C] glucose + 1 mM glutamine, 2 mM [1,2-¹³C]acetate + 1 mM glutamine, or 5 mM [1-¹³C]lactate + 2 mM glucose + 1 mM glutamine for 24 h. Astrocytes were washed once with PBS and incubated with 2 mL DMEM (Sigma D5030) supplemented with 4 mM [1-¹³C]lactate + 2 mM glucose, 1 mM glutamine, 1% pen-strep, and 1% FBS for 24 h. Samples of medium were collected before and after the incubation period and subsequently analyzed by mass spectrometry. After the 24 h incubation period, cells were washed twice with cold PBS and the intracellular metabolites extracted with 70% ethanol (Amaral et al., 2014). Astrocyte cultures were also incubated for 24 h in 2 mL DMEM containing 2 mM D-glucose, 1 mM glutamine, 1% pen-strep, and 1% FBS in order to determine glucose consumption and lactate production rates after 24 h. Experiments were performed on 9–12 samples, which derived from a minimum of three independently generated cultures.

Immunocytochemistry

OPCs were seeded on PLL-coated glass coverslips on 24 well plates (7×10^4 cells/well) and fixed after 5 days of differentiation with 4% paraformaldehyde. Cells were stained with an anti-O4 mouse monoclonal antibody (1:200; Sigma, Dorset, UK), anti-myelin basic protein (MBP) rat polyclonal antibody (1:300; Merck Millipore, Hertfordshire, UK), and anti-GFAP rabbit polyclonal antibody (1/500; Dako, Glostrup, Denmark). Secondary antibodies conjugated with Alexa Fluor 488 and Alexa Fluor 555 were used to visualize positive cells (1:500; Life Technologies, Paisley, UK). Following

immunocytochemistry, cells were mounted with Prolong gold anti-fade mounting medium (Life Technologies, Paisley, UK). To assess purity and differentiation of oligodendroglia lineage cells in the cultures, the percentage of O4-, MBP-, and GFAP-positive cells relative to >100 4',6-diamidino-2-phenylindole (DAPI)-stained nuclei per experiment in randomly selected eye fields was determined. Cells were visualized and digitized at ambient temperature on an LSM 700 confocal microscope (Zeiss, Thornwood, NJ) at 20 \times magnification using Zen Application software (Zeiss).

Assessment of Cell Viability

To assess cell viability under the different experimental conditions, the Dead EndTM Fluorometric Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) System Kit (Promega, Madison, WI) was used according to the manufacturer's instructions. Images of the stained cells were obtained using an INCell Analyzer 2200 Imaging System (GE Healthcare Life Sciences) and processed using the ImageJ software. Ten random fields were acquired per condition using a magnification of 20 \times . Image analysis and quantification was performed on *CellProfiler*, a cell image analysis software developed by the BROAD Institute. The number of DAPI-positive nuclei (total number of cells) and the number of TUNEL-positive nuclei were automatically counted, and the results presented as percentage of apoptotic nuclei of all DAPI-positive nuclei.

Glucose and Lactate Analyses

Glucose and lactate levels in the cell culture medium were analyzed at the Core Biochemical Assay Laboratory, Clinical Biochemistry, Addenbrooke's Hospital using automated assays on a Siemens Dimension RxL analyser. The rate of glucose and lactate net change relative to cells over time ($\mu\text{mol}/10^6$ cells/24 h) was calculated by subtracting the value measured at the end of the experiment ($T = 24$ h) from the one measured in a sample of medium collected at the onset of the incubation, and dividing the resulting value by the amount of cells in each experiment, multiplied by the experimental volume (2 mL). For oligodendrocyte cultures, the cell number considered was the cell number at plating since these cells do not proliferate. For astrocyte cultures (which proliferate to some extent), the cell number considered was determined at the end of the experiment, after collection of cells from two sample wells using trypsin-EDTA.

High Performance Liquid Chromatography

High performance liquid chromatography (HPLC) was used to quantify the total amounts of amino acids in samples of cell extracts and medium. Samples were lyophilized and resuspended in 0.01M HCl and subsequently derivatized with *o*-phthalaldehyde (Geddes and Wood, 1984) using an automated method prior to injection into the HPLC column. The amino acid components were separated with a ZORBAX SB-C18 (4.6 \times 150 mm, 3.55 μm) column from Agilent Technologies (Palo Alto, CA). As eluents, a mixture of 50 mM sodium phosphate buffer (pH 5.9) with 2.5% tetrahydrofuran and a mixture of methanol (98.75%) with tetrahydrofuran (1.25%) were used. The samples were analyzed using a Hewlett Packard 1100 System (Agilent Technologies, Palo Alto, CA) with

fluorescence detection. Amino acid concentrations were determined by comparison to a calibration curve of standard solutions of amino acids run after every 12 samples (Amaral et al., 2014).

Gas Chromatography–Mass Spectrometry

For analysis of percent enrichment of ¹³C in lactate, amino acids (alanine, aspartate, glutamate, and glutamine) and TCA cycle intermediates (citrate and malate) after incubation with different ¹³C-labelled substrates, cell extracts, and samples of medium were lyophilized and resuspended in 0.01M HCl. To move the metabolites of interest into the organic phase in their acid form, the pH was adjusted to pH < 2 with 6M HCl. Samples were dried under atmospheric air (50°C), and metabolites were derivatized with MTBSTFA in the presence of 1% *t*-BDMS-Cl (Mawhinney et al., 1986). The protocol used for analysis of the glycolytic intermediates phosphoenolpyruvate (PEP) and 3-phosphoglycerate (3PG) was based on the protocol reported by Hofmann et al. (2008). Derivatization was performed using a mixture of MSTFA + 1% trimethylchlorosilane and acetonitrile. The samples were analyzed on an Agilent 6890 gas chromatograph connected to an Agilent 5975B mass spectrometer (Agilent Technologies, Palo Alto, CA). The parent ion (*M*) and atom percent excess for one ¹³C atom (*M*+1) values for 3PG, PEP, alanine, aspartate, lactate, citrate, and glutamate were calculated from the gas chromatography-mass spectrometry (GC–MS) data using the MassHunter software supplied by Agilent (Agilent Technologies, Palo Alto, CA) and correcting for the naturally abundant ¹³C using nonenriched standards (Biemann, 1962).

¹³C And ¹H NMR Spectroscopy

¹³C NMR spectroscopy was used to identify the synthesis of particular labelled isotopologues from [1,2-¹³C]glucose metabolism due to the ability of this technique to distinguish between the different carbon positions that are labelled in one molecule. For example, ¹³C NMR spectroscopy enabled to distinguish between the presence of [2,3-¹³C]glutamate (synthesized via pyruvate carboxylation) and [1,2-¹³C]glutamate (synthesized via PDH; see below for further details). In contrast, GC–MS is a more sensitive method that provides information about ¹³C enrichment above natural abundance but lacks the specificity of NMR spectroscopy, as it does not provide information about the location of the ¹³C label. GC–MS, as applied in this study, only enables to distinguish between species that have a different number of ¹³C-labelled carbons. All NMR samples were analyzed using a QCI CryoProbeTM 600 MHz (for proton) ultra-shielded Plus magnet (Bruker BioSpin GmbH, Reinstetten, Germany). ¹H NMR spectra were acquired using a pulse angle of 90°, 12 kHz spectral width with 66 data points, acquisition time of 2.66 s, relaxation delay of 10 s and 128 scans. These spectra were used to quantify the amount of glutamate for correction of natural abundance of [4-¹³C]glutamate (to be used in the calculation of contribution of the PPP to [4-¹³C]glutamate synthesis). Proton decoupled ¹³C NMR spectra were obtained on the same instrument using a 30° pulse angle and 30 kHz spectral width with 98,000 data points using an acquisition time of 1.65 s and a relaxation delay of 0.5 s. The number of scans needed to obtain an appropriate signal to noise ratio was 210,000. TopSpinTM 3.0 software (Bruker BioSpin

GmbH, Reinstetten, Germany) was used for acquisition, integration, and quantification. Relevant peaks in the spectra were assigned and quantified from the integrals of the peaks using ethylene glycol as an internal standard with known amount of ^{13}C . Corrections for natural abundance as well as nuclear Overhauser enhancement and relaxation effects, relative to the internal standard, were applied to all relevant integrals from ^{13}C spectra.

Statistical Analysis

Statistical analysis was conducted using unpaired two-tailed student's *t*-tests (confidence interval = 95%).

Results

Characterization of Late Differentiation-Stage Oligodendrocyte Lineage Cells In Vitro

To study metabolic reactions in mature oligodendroglia cultures, highly enriched primary rat OPCs (Fig. 1A) were differentiated in Sato's differentiation medium. At 5 days of differentiation, the cells displayed the characteristically branched morphology of late oligodendrocyte lineage cells with approximately 65% of the cells expressing MBP (Fig. 1B). To investigate the relative activity of different metabolic pathways in mature oligodendrocytes, cells were incubated with one of the following ^{13}C -labelled substrates: $[1,6-^{13}\text{C}]$ glucose, $[1,2-^{13}\text{C}]$ glucose, $[1-^{13}\text{C}]$ lactate, or $[1,2-^{13}\text{C}]$ acetate.

Differentiated Oligodendrocyte Lineage Cells Display a Significant Activity of the PPP

To investigate the relative activity of the glycolytic pathway versus the PPP, $[1,2-^{13}\text{C}]$ glucose was added to the medium (Brekke et al., 2014; Dusick et al., 2007). Following 24 h of incubation, cell extracts were collected and analyzed using GC-MS and ^{13}C and ^1H NMR spectroscopy. If $[1,2-^{13}\text{C}]$ glucose is metabolized via the glycolytic pathway, $[2,3-^{13}\text{C}]$ 3PG, $[2,3-^{13}\text{C}]$ PEP, and $[2,3-^{13}\text{C}]$ pyruvate are formed (Fig. 2A). $[2,3-^{13}\text{C}]$ pyruvate can enter the mitochondria to be converted into $[1,2-^{13}\text{C}]$ acetyl CoA. Condensation of $[1,2-^{13}\text{C}]$ acetyl CoA with unlabelled oxaloacetate leads to the formation of the TCA cycle intermediate $[1,2-^{13}\text{C}]$ citrate and then, following several steps, α - $[4,5-^{13}\text{C}]$ ketoglutarate, which is subsequently converted into $[4,5-^{13}\text{C}]$ glutamate. $[1,2-^{13}\text{C}]$ glucose metabolism via the PPP gives rise to $[3-^{13}\text{C}]$ 3PG, $[3-^{13}\text{C}]$ PEP, $[3-^{13}\text{C}]$ pyruvate, and $[2-^{13}\text{C}]$ acetyl CoA (Fig. 2B). $[2-^{13}\text{C}]$ acetyl CoA can then be converted into $[4-^{13}\text{C}]$ glutamate via the TCA cycle (Fig. 2B).

Because PEP and 3PG are localized downstream of the intersection between glycolysis and the PPP, their ^{13}C isotopologues can be used to calculate the ratio between glycolysis and the PPP. Analysis of PEP and 3PG enables detection of isotopologues that derive from the re-entry of metabolic intermediates (fructose-6P and GA3P) into glycolysis from the PPP. The intermediates of the PPP lose the ^{13}C in the C1

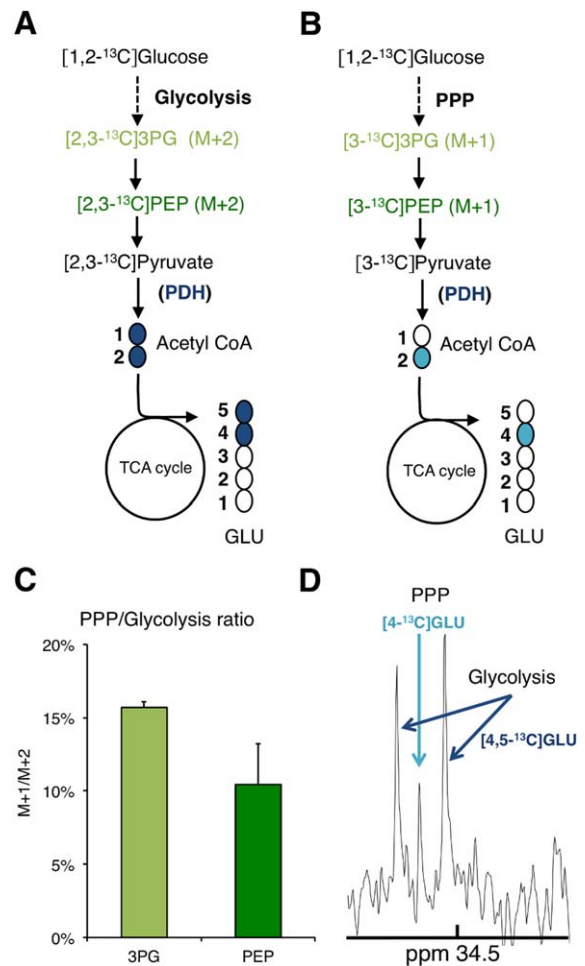


FIGURE 2: Evidence of pentose phosphate pathway (PPP) activity in mature oligodendrocytes in culture after incubation with $[1,2-^{13}\text{C}]$ glucose. Labelling patterns derived from $[1,2-^{13}\text{C}]$ glucose metabolism via glycolysis (A) and the PPP (B). The % enrichment of ^{13}C in M+1 and M+2 isotopologues for 3PG and PEP was determined using GC-MS analysis of cell extracts after 24-h incubation with $[1,2-^{13}\text{C}]$ glucose. The ratio shown in (C) between M+1 (labelling from the PPP) and M+2 (labelling from glycolysis) enables to estimate the contribution of the PPP versus glycolysis to the formation of the glycolytic intermediates 3PG and PEP in mature oligodendrocyte (mean \pm s.e.m.; $n = 3$). (D) ^{13}C nuclear magnetic resonance spectrum of a cell extract from cultures incubated in medium containing $[1,2-^{13}\text{C}]$ glucose for 24 h. The C-4 region of glutamate at 34.5 ppm (GLU) is shown. The doublet peak represents $[4,5-^{13}\text{C}]$ glutamate ($[4,5-^{13}\text{C}]$ GLU) which derives from glucose metabolism via glycolysis only. The singlet peak corresponds to $[4-^{13}\text{C}]$ glutamate ($[4-^{13}\text{C}]$ GLU) which is produced after glucose metabolism via the PPP. Abbreviations: 3PG, 3-phosphoglycerate; GLU, glutamate; PEP, phosphoenolpyruvate; PDH, pyruvate dehydrogenase; M+1, parent ion with one ^{13}C atom; M+2, parent ion with two ^{13}C atoms. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

position via decarboxylation of $[1,2-^{13}\text{C}]$ gluconate-6-phosphate by 6-phosphogluconate dehydrogenase and 1/3 of the molecules will re-enter glycolysis with only one labelled (M+1) carbon instead of two (M+2)—see Fig. 2 and Brekke et al. (2012) for further details. Based on GC-MS results, the

M+2/M+1 ratios of PEP and 3PG were calculated, indicating that 10–15% of glucose is metabolized via the PPP in mature oligodendrocytes (Fig. 2C). Of note, this ratio is likely to underestimate the contribution of the PPP (for details, see Brekke et al., 2012).

An alternative way to investigate the relative contribution of glycolysis and PPP to glucose metabolism is to apply ^{13}C NMR spectroscopy to differentiate between [4,5- ^{13}C] and [4- ^{13}C]glutamate (Fig. 2D). In the ^{13}C NMR spectrum, glycolysis-derived [4,5- ^{13}C]glutamate is represented as a doublet in the C-4 region of glutamate (34.5 ppm), whereas PPP-derived [4- ^{13}C]glutamate is represented as a singlet. The contribution of glucose metabolized via the PPP relative to glucose metabolized via glycolysis to the total glutamate synthesis was calculated by dividing the area of the [4- ^{13}C]glutamate peak (after correction for natural abundance using ^1H NMR spectroscopy) by the area of the doublet peak [4,5- ^{13}C]glutamate. We found that $10 \pm 0.2\%$ (mean \pm s.e.m.; $n = 3$) of the total glutamate comes from glucose metabolized in the PPP. Our results confirm that the PPP is active in oligodendrocytes and that it contributes to glutamate synthesis.

Evidence for PDH and Mitochondrial Activity in Oligodendrocytes

To investigate the extent of oxidative metabolism in oligodendrocytes, cells were incubated with ^{13}C -labelled glucose or [1,2- ^{13}C]acetate for 24 h (for labelling patterns, see Fig. 3A, B). We confirmed that after 24 h, all [1,6- ^{13}C]glucose-derived metabolites, except for glutamine, reached a steady-state of labelling (Supp. Info. Fig. 1). Hence, we decided to investigate labelling patterns in cell lysates after 24 h in all subsequent experiments. Assessing cell viability using TUNEL staining showed no differences between the various experimental conditions (Fig. 3C). Metabolism of [1,6- ^{13}C]glucose yields two molecules of [3- ^{13}C]alanine and [3- ^{13}C]pyruvate. The latter is then converted into [2- ^{13}C]acetyl CoA, which can condense with nonlabelled oxaloacetate to form monolabelled (M+1) compounds in the first turn of the TCA cycle (described in Fig. 3A). [2- ^{13}C]acetyl CoA can also condense with labelled oxaloacetate and give rise to the formation of double-labelled (M+2) compounds in a combination of the first and second turn of the cycle (Fig. 3A). GC–MS analysis of cell extracts incubated with [1,6- ^{13}C]glucose (Fig. 3D), showed that mitochondrial metabolism was prominent in mature oligodendrocytes. TCA cycle intermediates and amino acids were highly enriched ranging from 15 to 25% in citrate, malate, glutamate, and glutamine and 10% in aspartate (Fig. 3D). Moreover, a substantial enrichment was also observed with M+2 isotopologues and even M+3 (data not shown), typical of the second and third turns of the TCA cycle (see

Fig. 3D). However, the second turn of the TCA cycle is underestimated since labelled oxaloacetate can condense with unlabelled acetyl CoA and give rise to single labelled (M+1) compounds in the second turn. Second turn isotopologues were also observed when [1,2- ^{13}C]glucose was used as substrate (Fig. 3D). It is important to note that only half of the pyruvate molecules are labelled from [1,2- ^{13}C]glucose and, therefore, the maximum enrichment levels will be half of those obtained from [1,6- ^{13}C]glucose. Alanine is obtained from pyruvate transamination and is generally considered to be a metabolite related to glycolysis. The expected isotopologue of [1,2- ^{13}C]glucose is [1,2- ^{13}C]alanine. However, we observed M+1 alanine in addition to M+2-labelled alanine (Fig. 3D). This is evidence for the presence of partial pyruvate recycling, which can be performed via decarboxylation of malate or oxaloacetate into pyruvate (Sonnewald, 2014).

Acetate Metabolism in Oligodendrocytes

Conversion of acetate into acetyl CoA is known to take place in astrocytes, but not in neurons (Sonnewald and Rae, 2010). To investigate whether oligodendrocytes are able to convert acetate into acetyl CoA and oxidize it in the TCA cycle, cells were incubated with [1,2- ^{13}C]acetate for 24 h and the extra- and intracellular metabolites were analyzed by GC–MS (Fig. 3B, D). Both [1,2- ^{13}C]acetate and [1,2- ^{13}C]glucose identically yield [1,2- ^{13}C]acetyl CoA (Fig. 3B). Overall, the extent of labelling from [1,2- ^{13}C]acetate was much lower than that from [1,2- ^{13}C]glucose. Enrichment of the most abundant metabolites citrate and glutamate from [1,2- ^{13}C]acetate was approximately half of enrichment obtained from [1,2- ^{13}C]glucose; the less-abundant metabolites, aspartate and glutamine, were poorly enriched. The abundance of M+1 isotopologues (typical of the second turn of the TCA cycle) derived from [1,2- ^{13}C]acetate was very low (Fig. 3D), suggesting that acetate-derived acetyl CoA is metabolized in a different compartment than pyruvate-derived acetyl CoA. Alanine enrichment from [1,2- ^{13}C]acetate (albeit at low levels), confirms that oligodendrocytes have active partial pyruvate recycling, as observed in cells incubated with [1,2- ^{13}C]glucose. In the medium, labelling from [1,2- ^{13}C]acetate was only detectable in citrate, in which M+2 amounted to $10 \pm 2\%$ (mean \pm s.e.m., $n = 12$).

Glucose Consumption and Release of Metabolites to the Culture Medium

Analysis of glucose consumption and lactate release rates (Fig. 3E–I, II) indicates that oligodendrocytes metabolize glucose to an extent comparable to astrocytes ($3.2 \pm 0.06 \mu\text{mol}/10^6$ cells/24 h; mean \pm s.e.m.; $n = 6$). However, oligodendrocytes release less lactate than astrocytes ($5.3 \pm 0.16 \mu\text{mol}/10^6$ cells/24 h; mean \pm s.e.m.; $n = 6$). Although glucose consumption

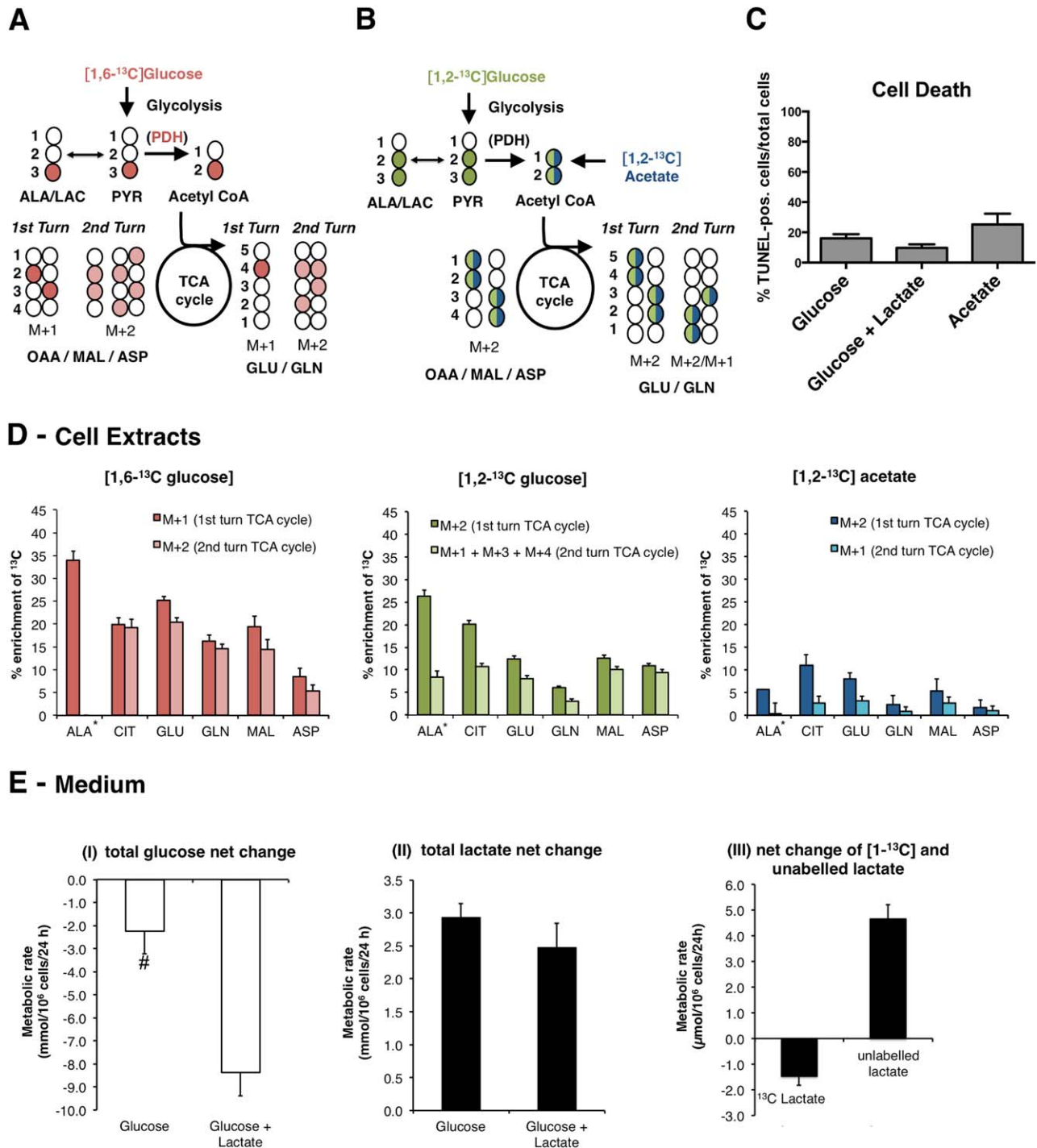


FIGURE 3: Evidence of high TCA cycle activity and acetate metabolism in mature oligodendrocytes in culture. Oligodendrocytes were differentiated for 5 days and incubated for 24 h in medium containing [1,6-¹³C]glucose (A), [1,2-¹³C]glucose (B), or [1,2-¹³C]acetate (B), followed by GC-MS analysis of samples of cell culture medium and cell extracts. A and B describe the labelling patterns deriving from the metabolism of these ¹³C-labelled substrates. The isotopologues formed in the second turn of the TCA cycle result from condensation of labelled oxaloacetate (OAA) with labelled or unlabelled acetyl CoA. (C) Quantification of TUNEL positive/total cell number (given by DAPI staining) in cells incubated with glucose alone (mean ± s.e.m., *n* = 12), glucose and lactate (mean ± s.e.m., *n* = 10), or acetate (mean ± s.e.m., *n* = 8); (D) % enrichment of ¹³C in intracellular alanine and TCA cycle-related metabolites derived from each of the substrates (mean ± s.e.m., *n* = 8 for [1,6-¹³C]glucose, mean ± s.e.m., *n* = 6 for [1,2-¹³C]glucose and mean ± s.e.m., *n* = 12 for [1,2-¹³C]acetate). (E) Glucose (I) and lactate (II) net change in the medium in experiments performed in the presence of glucose alone or glucose + [1-¹³C]lactate. For the experiment where [1-¹³C]lactate was used, the net change of ¹³C-labelled and unlabelled lactate is shown (III) (mean ± s.e.m., *n* = 9). #—significantly different from the glucose + [1-¹³C]lactate group (*P* < 0.05, Student's *t*-test). Abbreviations: ALA, alanine; ASP, aspartate; CIT, citrate; GLN, glutamine; GLU, glutamate; MAL, malate; PYR, pyruvate; M+1, parent ion with one ¹³C atom; M+2, parent ion with two ¹³C atoms; M+3, parent ion with three ¹³C atoms; M+4, parent ion with four ¹³C atoms. *The enrichment detected in alanine derives directly from ¹³C-labelled pyruvate and not from the TCA cycle when [1,6-¹³C]glucose is the precursor. When [1,2-¹³C]glucose is used, M+2 is not derived from the TCA cycle but M+1 alanine is. When [1,2-¹³C]acetate is in the medium, both alanine isotopologues are derived from the TCA cycle. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

in the presence of 4 mM [1-¹³C]lactate (Fig. 3E-I) was increased, the net release of total lactate remained unchanged (Fig. 3E-II). Due to the presence of [1-¹³C]lactate in the culture medium, it was possible to distinguish between the uptake of [1-¹³C]lactate and the release of endogenous (unlabelled) lactate (Fig. 3E-III). This analysis showed that the presence of lactate in the medium increased the release of endogenous lactate (Fig. 3E-III). The release of unlabelled lactate was at least 3-fold larger than the amount of [1-¹³C]lactate consumed. By comparing lactate release rates and glucose consumption rates, it was possible to estimate the fraction of glucose metabolized to lactate, versus the oxidation of pyruvate in the mitochondria. In oligodendrocytes cultured with glucose alone, the ratio of lactate production to pyruvate oxidation was 60:40 (and 80:20 in astrocytes). In the presence of lactate, the ratio changed to 30:70, indicating

increased mitochondrial activity. Figure 3C shows that the viability of cells was not affected by the presence of lactate.

Despite the substantial % enrichment of ¹³C in the aforementioned amino acids observed in the cell extracts, only negligible release of alanine and glutamine was detected (data not shown).

Pyruvate Carboxylation in Oligodendrocytes

Pyruvate carboxylation is an important anaplerotic pathway known to operate in astrocytes but not in neurons (McKenna et al., 2012). The presence of this pathway was probed in mature oligodendrocytes using either [1,2-¹³C]glucose or [1-¹³C]lactate (Fig. 4). [1-¹³C]lactate is a valuable substrate for this purpose since only via pyruvate carboxylation the ¹³C label from [1-¹³C]lactate can be found in the TCA cycle intermediate citrate (PDH removes carbon number 1 from pyruvate generating unlabelled acetyl CoA, and therefore, the ¹³C label is lost via PDH; Fig. 4A). In order to evaluate the significance of pyruvate carboxylation in oligodendrocytes, also astrocytes (the cells known to carboxylate pyruvate in the brain) were incubated with [1-¹³C]lactate. Both of these cell types were shown to oxidize lactate to a large extent in culture (Sanchez-Abarca et al., 2001). [1-¹³C]Pyruvate carboxylation was apparent in the labelling of citrate in the medium in both cell types (Fig. 4B). However, in the cell extracts, this was only evident in astrocytes. A possible explanation for the

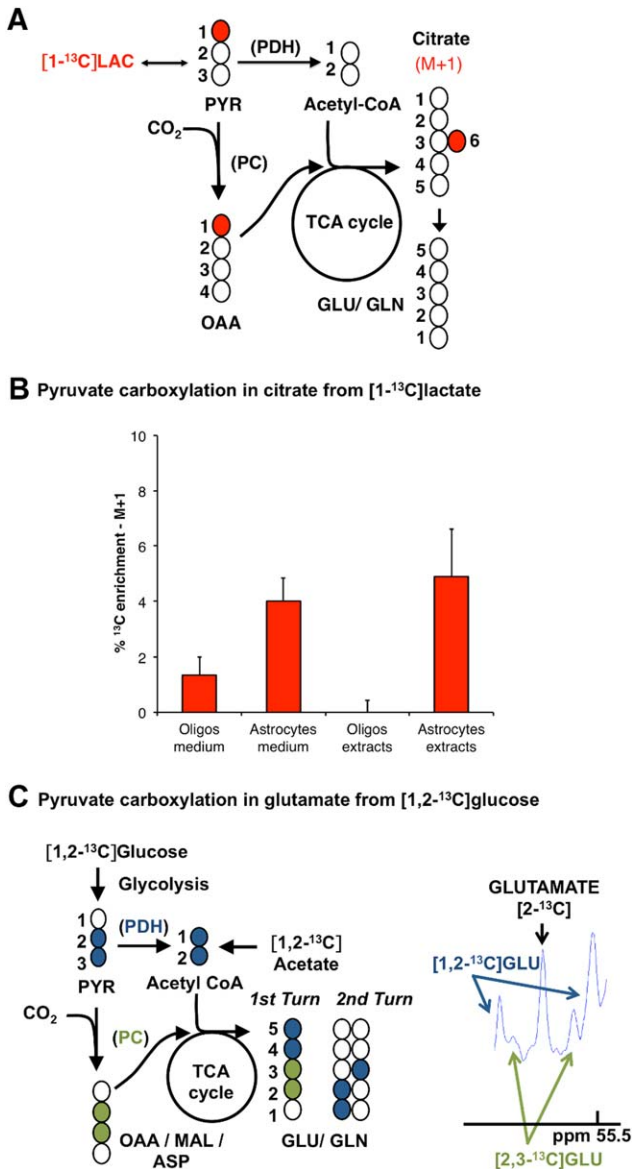


FIGURE 4: [1-¹³C]lactate metabolism and evidence for pyruvate carboxylation in oligodendrocytes. **(A)** Labelling patterns resulting from the metabolism of [1-¹³C]lactate (¹³C atoms are identified as filled circles): [1-¹³C]lactate (LAC) is converted into [1-¹³C]pyruvate, which can be converted into acetyl CoA via PDH or into [1-¹³C]oxaloacetate (OAA) via pyruvate carboxylase (PC). The first carbon of pyruvate is lost in its conversion to acetyl CoA via PDH. Therefore, the ¹³C label can only be observed downstream of pyruvate if PC is active. [1-¹³C]oxaloacetate can condense with acetyl CoA to form [6-¹³C]citrate, which also leads subsequently to the formation of unlabelled α -ketoglutarate by loss of carbon 6. **(B)** The % enrichment of ¹³C in citrate indicates the contribution of pyruvate carboxylation to citrate formation from [1-¹³C]lactate. The % of M+1 citrate was assessed using GC-MS in samples of cell extracts and culture medium from differentiated oligodendrocytes and also in cultures of cortical astrocytes after 24 h of incubation with [1-¹³C]lactate (oligodendrocytes—mean \pm s.e.m., $n = 9$; and astrocytes—mean \pm s.e.m., $n = 6$). **(C)** Contribution of pyruvate carboxylation to glutamate synthesis from [1,2-¹³C]glucose. Oligodendrocytes were incubated in medium containing [1,2-¹³C]glucose for 24 h, extracted, and analyzed using ¹³C-magnetic resonance spectroscopy for the presence of isotopologues of glutamate formed via pyruvate carboxylation. The C-2 region of glutamate (GLU) at 55.5 ppm is shown. The doublet representing [2,3-¹³C]glutamate is formed via pyruvate carboxylation and the doublet representing [1,2-¹³C]glutamate is formed via pyruvate dehydrogenation. Abbreviations: ASP, aspartate; GLN, glutamine; GLU, glutamate; MAL, malate; OAA, oxaloacetate; PC, pyruvate carboxylase; PDH, pyruvate dehydrogenase; PYR, pyruvate. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

absence of intracellular citrate enrichment in oligodendrocytes is a potential compartmentation of pyruvate metabolism, which has also been shown for astrocytes and neurons (Bak et al., 2007; Bakken et al., 1997).

To further investigate the presence of pyruvate carboxylation, oligodendrocytes were also incubated with medium containing [1,2-¹³C]glucose. The C-2 region of glutamate (around 55.5 ppm) in the ¹³C NMR spectrum of cell extracts was analyzed to confirm the presence of pyruvate carboxylation (Fig. 4C). From the scheme depicted in Fig. 4C, it emerges that only pyruvate carboxylation will lead to the formation of [2,3-¹³C]glutamate, whereas [1,2-¹³C]glutamate (and [3-¹³C]glutamate, which is not shown) is formed from pyruvate dehydrogenation (and pyruvate carboxylation, if the label stays in the cycle for an additional turn). The spectrum shown in Fig. 4C, which reflects the presence of [2,3-¹³C]glutamate, indicates that oligodendrocytes carboxylate pyruvate.

Discussion

Although oligodendrocytes make up a significant proportion of brain cells, their metabolic properties remain largely unknown. In a recent review, we have called for a systematic study of metabolic pathways in oligodendrocytes (Amaral et al., 2013). In this work, we study aspects of glucose, lactate, and acetate metabolism in oligodendrocytes, and specifically address the activity of the PPP and whether oligodendrocytes are able to conduct anaplerotic and cataplerotic reactions based on carboxylation of pyruvate and decarboxylation of malate or oxaloacetate.

Primary rat OPCs were purified from perinatal rat-mixed glial cultures (McCarthy and de Vellis, 1980). This approach results in high yields of relatively pure (>93%) cultures. Culture of the O4+, A2B5+, O1-, CNP-, and MBP- OPCs in Sato's medium triggers a highly predictable series of morphological and transcriptional events and results in the formation of mature oligodendrocytes with complex-branched processes and membrane sheets, which express late stage markers, including MBP. Although a significant proportion of cells (approximately 60%) reach a mature oligodendrocyte stage, the cultures also include late stage progenitors. Genomic studies comparing acutely isolated cells at distinct developmental stages with the culture system used in this study confirmed that primary OPC cultures faithfully represent their *in vivo* counterparts (Dugas et al., 2006).

Our results demonstrate that oligodendrocytes have extensive glucose-derived metabolism. In fact, we found that the rate of glucose consumption in oligodendrocytes is comparable to the one in astrocytes. Although Sanchez-Abarca et al. (2001) previously concluded that oligodendrocytes use more glucose than astrocytes, they also showed that oligodendrocytes metabolize a larger proportion of glucose via PDH

than astrocytes. We obtained a similar result by comparing the ratios of lactate release to glucose consumption in both cell types. Furthermore, we observed that both glucose consumption and the proportion of glucose-derived pyruvate metabolized in the mitochondria were increased in the presence of exogenous lactate. This suggests that lactate may act as metabolic activator in oligodendrocytes, fitting with the concept that lactate can act as signalling molecule (Rinholm and Bergersen, 2014). Furthermore, our results confirm that oligodendrocytes are able to release substantial amounts of lactate *in vitro* as has been reported *in vivo* by Funfschilling et al. (2012). Also, it is possible that, at least, a proportion of the [1-¹³C]lactate taken up was oxidized for lipogenesis, as proposed by (Sanchez-Abarca et al., 2001). In spite of the presence of 4-mM exogenous lactate, a net production of lactate was observed indicating that glycolysis in oligodendrocytes is not inhibited by extracellular lactate.

The PPP is a glucose shunt, which is thought to be active in neurons and astrocytes (Almeida et al., 2002; Amaral et al., 2010; Bolanos and Almeida, 2010; Brekke et al., 2012; Garcia-Nogales et al., 2003). Using ¹⁴C tracing techniques, Edmond et al. (1987) showed active PPP in oligodendrocyte lineage cells. Sanchez-Abarca et al. (2001) reported PPP activity in immature OPC cultures as being 2-fold higher than in astrocytes and 4-fold higher than in neurons. Our approach measured the contribution of the PPP to the synthesis of glycolytic intermediates and glutamate synthesis based on ¹³C-tracing techniques. In our study, the labelling of the glycolytic intermediates PEP and 3PG indicate that mature oligodendrocytes use approximately 10–15% of glucose in the PPP compared with glycolysis. These values are in the range of previously published data for cortical astrocyte cultures in a metabolic modelling study also using ¹³C-labelled substrates (Amaral et al., 2011a), which contrasts with the report by Sanchez-Abarca et al. (2001). It is possible that our cultures contain more mature oligodendrocytes than those used by Sanchez-Abarca et al., and therefore, have a lower PPP activity, more closely resembling the rate in astrocytes. On the basis of ¹⁴C tracing experiments, Sykes et al. (1986) reported that, although the PPP oxidizes only <3% of the glucose consumed, it produces more CO₂ than the TCA cycle in primary oligodendrocyte lineage cells, due to its close link to de novo synthesis of fatty acids and cholesterol. Since we did not measure the contribution of glucose to lipid synthesis, it is possible that our approach underestimated the total use of glucose via the PPP.

For the first time, our experiments demonstrate that pyruvate generated from glucose via the PPP contributes to the synthesis of acetyl CoA for oxidation and generation of metabolites in the TCA cycle in oligodendrocytes. We found that approximately 10% of the glutamate produced derives

from glucose metabolized in the PPP. Estimation of the PPP activity on the basis of glutamate isotopomers indicated that the PPP accounted for approximately 6% of glucose metabolism in cortical neurons and approximately 4% in cerebellar neurons (Brekke et al., 2012). Similar to reports in neurons (Brekke et al., 2012), we found that oligodendrocytes incorporate ^{13}C label in glutamate produced by $[1,2-^{13}\text{C}]$ glucose metabolism via the PPP.

The PPP may also play a role in diseases that affect oligodendrocytes. For example, increased activity of the transaldolase, an enzyme which forms part of the nonoxidative branch of the PPP and is involved in lipid and nucleotide synthesis, has been reported in oligodendrocytes in brains of MS patients, compared with healthy controls (Banki et al., 1994). NADPH produced in the PPP is fundamentally important for the synthesis of glutathione, which is thought to protect myelin sheaths from oxidative stress. Banki and colleagues proposed that autoantibodies against transaldolase found in MS patients, may cause destruction of oligodendrocytes via depletion of transaldolase. Altered PPP activity was also reported in a study of patients that suffered a traumatic brain injury (TBI; Dusick et al., 2007). It is possible that, at least, a proportion of the PPP alterations observed in the study by Dusick et al. (2007) could be linked to the extensive demyelination that is thought to occur in TBI patients (Armstrong et al., in press).

Using different forms of ^{13}C -labelled glucose, we demonstrate that our cultures also exhibit a high rate of mitochondrial metabolism, as previously suggested by Sanchez-Abarca et al. (2001). Whereas functional mitochondria seem to be important for OPC differentiation (Schoenfeld et al., 2010; Ziabreva et al., 2010), it was recently proposed that myelinating oligodendrocytes are not dependent on mitochondrial activity *in vivo* (Funfschilling et al., 2012). The most likely explanation for the discrepancy between the high mitochondrial demands of our cultures and the findings *in vivo* is the significant presence of premyelinating (MBP-negative) OPCs that have not yet reached fully mature stages *in vitro*. Our data thus may reflect the prominent role of oxidative metabolism at the late stages of OPC differentiation, including the premyelinating and early myelinating stages.

Another important question with respect to mitochondrial metabolism in oligodendrocytes is whether they can replenish TCA cycle intermediates via anaplerosis. It is well established that neurons depend on astrocytes for replenishing their TCA cycle intermediates. As neurons cannot carboxylate pyruvate, external TCA cycle intermediates are required for the synthesis of amino acid neurotransmitters (McKenna et al., 2012). Whether and to which extent oligodendrocytes are self-sufficient with respect to the production of anaplerotic substrates remained unknown. By incubating cells with

$[1,2-^{13}\text{C}]$ glucose, we found that pyruvate carboxylation indeed takes place in oligodendrocytes as shown by the production of $[2,3-^{13}\text{C}]$ glutamate. This was further confirmed by label incorporation in citrate in the medium of cells incubated with $[1-^{13}\text{C}]$ lactate, which is only possible via pyruvate carboxylation. Comparable label incorporation from $[1-^{13}\text{C}]$ lactate was found in citrate in the medium of astrocyte cultures. This suggests that lactate is metabolized in a similar way in the TCA cycle of both cell types. However, it is possible that a fraction of the lactate taken up by our cultures was used for lipogenesis (Sanchez-Abarca et al., 2001). It must also be noted that the overall contribution of pyruvate carboxylation to oligodendrocyte and astrocyte metabolism is underestimated in the experiments based on $[1-^{13}\text{C}]$ lactate metabolism because $[1-^{13}\text{C}]$ pyruvate derived from $[1-^{13}\text{C}]$ lactate competes with unlabelled pyruvate derived from glucose, which is also present in the incubation medium.

Pyruvate can be carboxylated to oxaloacetate by PC or to malate and NADP⁺ by malic enzyme. Whether PC or malic enzyme is responsible for pyruvate carboxylation in oligodendrocytes remains to be established. Murin et al. (2009) reported PC expression in cultured oligodendroglia. Whether oligodendrocytes express malic enzyme remains unknown. In neurons and astrocytes, malic enzyme only contributes to pyruvate production (McKenna et al., 1995, 2000). However, PC is thought to be the most important anaplerotic enzyme in the brain (Patel, 1974). Irrespective of which enzyme is responsible for pyruvate carboxylation, the detection of carboxylation (and thus anaplerosis) has consequences for oligodendrocyte metabolism. If oligodendrocytes (similarly to neurons) were not capable of anaplerosis, they would depend on the provision of glutamine by astrocytes, which have a net production of glutamine via pyruvate carboxylation (Gamberino et al., 1997; Waagepetersen et al., 2001).

An alternative route to replenish the TCA cycle in oligodendrocytes is to use aspartate, liberated from *N*-acetyl aspartate (NAA), which in turn is supplied by neurons (Moffett et al., 2007). However, aspartate released by NAA hydrolysis in oligodendrocytes could potentially be sent back to neurons, thus avoiding the depletion of anaplerotic substrates in neurons (NAA synthesis in neurons is dependent on glutamine entry from astrocytes). Our results indicate that oligodendrocytes are capable of anaplerosis, which suggests that they are potentially independent of astrocytic pyruvate carboxylation. Whether the level of anaplerosis in oligodendrocytes is sufficient to meet their entire requirements is not known at present. It is also unclear whether aspartate is shuttled back to neurons or whether it is metabolized in oligodendrocytes. Both possibilities have previously been suggested but evidence is lacking (Baslow and Guilfoyle, 2006; Moffett et al., 2007).

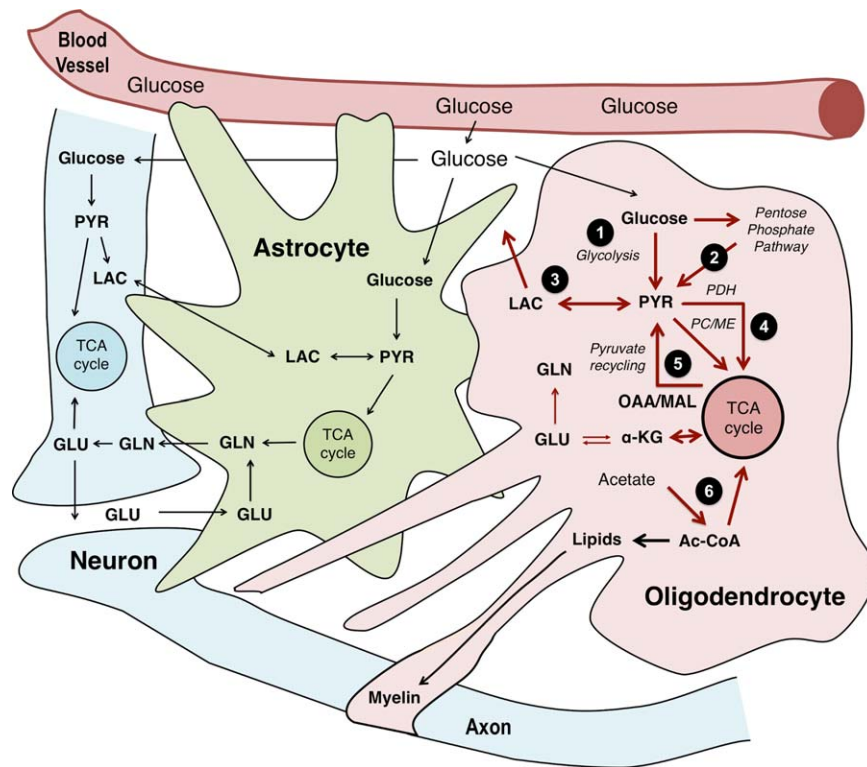


FIGURE 5: Integration of metabolic pathways operating in oligodendrocytes in the context of metabolic interactions with astrocytes and neurons/axons. The pathways investigated in this study are highlighted in red in the oligodendrocyte: glucose is taken up and subsequently metabolized either via glycolysis only (1) or also via the pentose phosphate pathway (2); the resulting pyruvate (PYR) produced can be reduced to lactate (LAC) (3) which can be released and taken up by cells with lower lactate concentration. Moreover, pyruvate can be carboxylated via PC or malic enzyme (ME) into oxaloacetate (OAA) or malate (MAL) or enter the TCA cycle after being converted to acetyl CoA (Ac-CoA) via PDH (4). The TCA cycle intermediate α -ketoglutarate (α -KG) gives rise to glutamate (GLU) and, subsequently, glutamine (GLN), but none of these aminoacids appear to be significantly released. Pyruvate can be further completely oxidized if it is decarboxylated via ME, a pathway called pyruvate recycling (5), which also seems to be present in this cell type. Oligodendrocytes can also metabolize acetate into acetyl CoA (6) that can be then incorporated into lipids or oxidized in the TCA cycle. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

Acetyl CoA is an essential molecule in the TCA cycle. Most acetyl CoA derives from pyruvate via PDH. However, oligodendrocytes are known to express the enzyme aspartoacylase, which catalyzes the hydrolysis of NAA into aspartate and acetate (Moffett et al., 2011). NAA-derived acetate significantly contributes to myelin lipid synthesis in the CNS (Chakraborty et al., 2001) and is also thought to support oxidative metabolism during myelination (Francis et al., 2012). Furthermore, oligodendrocytes express acetyl CoA synthetase-1, which catalyzes the synthesis of acetyl coenzyme A from acetate and coenzyme A, indicating that acetate may contribute to lipid synthesis, especially during postnatal brain development (Ariyannur et al., 2010). So far only astrocytes were shown to convert acetate into acetyl CoA whereas neurons do not seem able to do so (Muir et al., 1986; Sonnewald et al., 1993a). Consequently, acetate has been used extensively to assess astrocyte metabolism in the context of astrocytic-neuronal interactions *in vivo* (e.g., Melo et al., 2005; Morken et al., 2014; Nilsen et al., 2014), in neurospheres (Sa Santos

et al., 2011), and in culture (Sonnewald et al., 1993a). However, potential metabolic contributions from oligodendrocytes were not considered in these studies. This work demonstrates that mature oligodendrocyte cultures are able to convert acetate into acetyl CoA and oxidize it in the mitochondria as shown by the incorporation of ^{13}C label from $[1,2-^{13}\text{C}]$ acetate into the TCA cycle intermediates malate and citrate and the amino acids glutamate and glutamine. Labelling from $[1,2-^{13}\text{C}]$ acetate was not as pronounced as labelling from $[1,2-^{13}\text{C}]$ glucose but, nevertheless, significant and comparable to that observed in astrocytes (data not shown). However, astrocytes label glutamine extensively from ^{13}C -labelled acetate (Hassel et al., 1995), whereas oligodendrocytes do not. Surprisingly, we found that alanine was also labelled from $[1,2-^{13}\text{C}]$ acetate, and that alanine M+1 enrichment was detected in oligodendrocytes incubated with $[1,2-^{13}\text{C}]$ glucose. Both isotopologues of alanine could not have been produced without the participation of the TCA cycle and malic enzyme or pyruvate kinase and phosphoenolpyruvate carboxykinase

(Cruz et al., 1998). This indicates that pyruvate recycling, a catabolic pathway (Amaral et al., 2011b; Cerdan et al., 1990; Haberg et al., 1998; Kunnecke et al., 1993; Olstad et al., 2007), is not only active in astrocytes and neurons but also in oligodendrocytes.

Conclusion

Our results show that oligodendrocyte lineage cells at late stages of differentiation are metabolically active cells and have distinct metabolic properties. We found that the cells were able to conduct all the metabolic functions that were investigated and, therefore, demonstrate a high degree of cellular independence. Figure 5 summarizes the findings of the present series of experiments and integrates our data with the known pathways linking astroglial and neuronal metabolism. Our results indicate that metabolic functions of oligodendroglia need to be considered in studies investigating glucose metabolism in CNS tissue or whole brain studies. This work reinforces the emerging role of oligodendrocyte metabolism with respect to neuronal–glial interactions.

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