

Genome-wide screen identifies host colonization determinants in a bacterial gut symbiont

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Animal guts are often colonized by host-specialized bacterial species to the exclusion of other transient microorganisms, but the genetic basis of colonization ability is largely unknown. The bacterium *Snodgrassella alvi* is a dominant gut symbiont in honey bees, specialized in colonizing the hindgut epithelium. We developed methods for transposon-based mutagenesis in *S. alvi* and, using high-throughput DNA sequencing, screened genome-wide transposon insertion (Tn-seq) and transcriptome (RNA-seq) libraries to characterize both the essential genome and the genes facilitating host colonization. Comparison of Tn-seq results from laboratory cultures and from mono-inoculated worker bees reveal that 519 of 2,226 protein-coding genes in *S. alvi* are essential in culture, whereas 399 are not essential but are beneficial for gut colonization. Genes facilitating colonization fall into three broad functional categories: extracellular interactions, metabolism, and stress responses. Extracellular components with strong fitness benefits in vivo include trimeric autotransporter adhesins, O antigens, and type IV pili (T4P). Experiments with T4P mutants establish that T4P in *S. alvi* likely function in attachment and biofilm formation, with knockouts experiencing a competitive disadvantage in vivo. Metabolic processes promoting colonization include essential amino acid biosynthesis and iron acquisition pathways, implying nutrient scarcity within the hindgut environment. Mechanisms to deal with various stressors, such as for the repair of double-stranded DNA breaks and protein quality control, are also critical in vivo. This genome-wide study identifies numerous genetic networks underlying colonization by a gut commensal in its native host environment, including some known from more targeted studies in other host-microbe symbioses.

type IV pilus | transposon mutagenesis | microbiota | symbiosis | Neisseriaceae

Many animal species, including humans, have highly specific associations with a core gut microbiota consisting of host-adapted bacteria (1). Although many other microbial species are routinely ingested with food, few are able to stably colonize the gut. Considerable progress has been made in identifying the genes and mechanisms enabling bacterial colonization and persistence in hosts (e.g., refs. 2–7), but the extent to which common processes play a role for gut symbionts in different host taxa is unclear.

Honey bees (*Apis mellifera*) resemble mammals in possessing characteristic gut bacteria acquired through social contact; however, the bee gut is dominated by only approximately eight species, in contrast to the hundreds of species typically found in mammalian guts (8). The distribution of these bacteria across related bee species (8) indicates a long evolutionary association, another feature shared with some mammalian gut associates (9). An abundant constituent of the honey bee and bumble bee gut microbiota is *Snodgrassella alvi* (10), a member of Betaproteobacteria that is related to human commensals in the genus *Neisseria* (11). *S. alvi* colonizes the hindgut ileum, a region analogous to the proximal large intestine in humans and is normally found in contact with the intima lining, of the ileum (12, 13).

S. alvi is cultivable in the laboratory, and cultured strains can be used to experimentally colonize germ-free adult worker bees with high efficiency (14) (Fig. 1). Experimental transfers of *S. alvi* strains between honey bee and bumble bee hosts indicate that

strains are specialized to colonize their native host species (14). Although diverse bacteria are initially present in guts of young bees, only core species such as *S. alvi* can stably colonize (8). Neither the basis for host specificity nor the basis for *S. alvi*'s ability to persist in bee guts is known. To determine the genes that enable *S. alvi* to colonize the bee gut, we used high-throughput sequencing of a saturated transposon mutant library (Tn-seq) to screen tens of thousands of random insertion mutants across the *S. alvi* genome during colonization in vivo and in laboratory culture. This technique allowed us to assay the contribution of each gene toward the establishment of this symbiosis. We find that extensive genetic networks involved in extracellular interaction, metabolism, and stress response have strong fitness benefits in vivo and likely play key roles in facilitating gut colonization.

Results and Discussion

To identify genes needed for growth and genes contributing to gut colonization, we generated a transposon insertion library in *S. alvi* strain wkB2 (*SI Appendix*). The library comprised ~74,000 individual mutants, representing on average one insertion per 34 bp across the *S. alvi* genome and thus ensuring that every gene was inactivated by multiple mutants (Fig. 2A and *SI Appendix*, Table S1).

Genes Contributing to Growth in Laboratory Culture. We identified genes tolerating far fewer transposon insertions than expected at random during growth in laboratory culture (*SI Appendix*). Of 2,226 protein-coding genes, 519 are essential or highly beneficial under our culture conditions ($P_{\text{adj}} \leq 0.05$) (Fig. 2B and *Dataset*

Significance

Many bacteria are adapted to live in specific locations in animal hosts, even while these same locations exclude colonization by most microorganisms. However, the genetic underpinnings enabling successful colonization are largely unclear. We developed a system to genetically manipulate *Snodgrassella alvi*, a bacterium restricted to bees, and explored the factors permitting *S. alvi* to establish within its natural habitat in the bee digestive tract. Using high-throughput methods that screen the entire genome, we find that host colonization is dependent on genes mediating cell surface interactions (e.g., adhesion), metabolism under nutrient limitation, and responses to various stresses. This study demonstrates the genetic tractability of the bee gut microbiota, an emerging system with parallels to the human microbiome.

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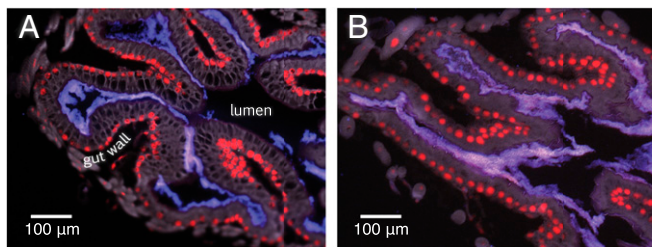


Fig. 1. Cross-sections of the honey bee ileum, showing colonization by *S. alvi* in 4-d-old workers inoculated (A) experimentally with laboratory-cultured *S. alvi* wkB2 and (B) naturally within the hive. *S. alvi* localization (purple) is derived from an overlay of a DNA stain (red) and an rRNA probe specific to *S. alvi* (blue). Methods and *B* are adapted from ref. 14.

SIC). Most of these genes participate in highly conserved cellular functions such as central informational processes (replication, transcription, and translation), respiration (electron transport chain), and biosynthesis of cell constituents (including peptidoglycan and lipopolysaccharide) (*SI Appendix, Fig. S1*). Genes of the gluconeogenesis pathway are essential, reflecting the fact that *S. alvi* lacks the ability to take up exogenous sugars (14). Genes required or highly beneficial for growth in culture included 37 encoding proteins of unknown function.

Genes Promoting Gut Colonization. We compared genome-wide transposon insertion frequencies in our mutant library fed to germ-free adult worker bees to the frequencies obtained from laboratory cultures (*SI Appendix*). This approach allowed us to query the 1,707 protein-coding genes considered “nonessential” in culture, for roles in gut colonization. Of these, 399 genes were underrepresented in the library recovered from guts (cutoff criteria of \log_2 -fold decrease ≥ 1 and $P_{\text{adj}} \leq 0.05$) (Fig. 2C and *Dataset S1B*). Of 399 genes conferring a benefit in vivo, 109 could not be confidently ascribed a function. The remaining 290 genes were inspected and placed into pathways based on similarity to characterized homologs in other bacteria. This analysis reveals the presence of extensive, interconnected gene networks that underlie host colonization (Fig. 3) and gives insight into the demands of the gut environment. Most genes conferring fitness benefits during colonization fall into three broad functional categories: extracellular interaction, metabolism, and stress response.

Extracellular interaction. Extracellular components mediate direct, physical contact between a bacterium and its environment, which potentially includes both host cells and other bacteria (15). In *S. alvi*, as for most Gram-negative bacteria, lipopolysaccharide (LPS) is the major constituent of the outer leaf of the outer membrane, and its synthesis from the carbohydrate products of gluconeogenesis, as well as its export (via the *lpt* gene products) were essential for cell viability under all conditions (Fig. 3). However, in vivo, the LPS core alone was insufficient for colonization. O antigens (polysaccharide additions to LPS) appear crucial; mutants in any of the pathways leading to O-antigen precursor synthesis (dTDP- β -L-rhamnose, UDP- α -D-glucose, and UDP- α -D-galactose) and assembly (*wzx* and *wzy*) were non-competitive in the bee gut. O antigens can confer benefits by enabling evasion of host immunity, binding to host receptors, and promoting cell envelope resilience against external insults (16).

Adhesion and biofilm formation are often important for host colonization (15). *S. alvi* grows in a distinct layer adjacent to the gut epithelium, suggesting adherence (12, 13) (Fig. 1). Two of the three trimeric autotransporter adhesin (TAA) genes in *S. alvi*, *staA* and *staB*, appear crucial for gut colonization. TAAs are a family of diverse, but structurally similar, adhesion factors that are widespread in bacteria, including human-associated *Yersinia*, *Neisseria*, and *Bartonella* (17). Consisting of a sticky head for adhering to host cells or extracellular matrices, and connected to the outer membrane by a lengthy stalk, these proteins are encoded by some of the largest

S. alvi genes (~10 kb). Both *staA* and *staB* are also present in *S. alvi* strains from bumble bees (14), suggesting a conserved role in colonization.

S. alvi encodes a complete type IV pilus (T4P) (18), plus a number of accessory or duplicated components. The T4P is a versatile bacterial cellular machine that can facilitate adhesion, biofilm formation, motility, secretion, and DNA uptake (19). In *S. alvi*, only *pilD* (prepilin peptidase) was essential in vitro. However, in vivo, mutations to all core T4P structural components (*pilFGMNOQW* and *tsaP*) were highly detrimental. Among T4P genes with multiple variant copies, such as the pilus retraction/twitch motility motor *pilT*, and the pilus tip adhesin *pilC*, our screen discerned the variant most important for colonization (*Dataset S2*).

The pilus itself is chiefly composed of major pilin subunits, encoded by *pilE*. Pilin expression is triggered by environmental cues; in *S. alvi*, this is likely controlled by the two-component *pilR/S* system, as documented in the related species *Kingella kingae* (20). Transposon insertions in both *pilR* and *pilS* were underrepresented in vivo; although, as in *Kingella*, non-functionalization of the sensor *pilS* was more tolerated than that of *pilR* (20). The *pilE* gene in many bacteria, including commensal *Neisseria* species (21), is directly regulated by the products of *pilR* and *rpoN* (encoding the master transcriptional regulator σ -54), another gene required for gut colonization in our screen. A role for *rpoN* in pilus formation is likely in *S. alvi*, which has the canonical σ -54 promoter upstream of the *pilE* start codon.

Metabolism. *S. alvi* is an obligate aerobe, but potentially experiences periods of suboptimal O₂ concentrations in the gut, as some insect guts have anaerobic regions (22). Correspondingly, the *ntrX/ntrY* two-component oxygen sensor (23), was crucial for gut colonization. Regulatory responses to *ntrX/ntrY* are mediated by σ -54 and potentially lead to the expression of microaerobic/anaerobic respiratory components such as cytochrome *bd*, the nitrite/nitrate antiporter *narK*, and nitrate reductase. These products may accord *S. alvi* flexibility in meeting energy requirements in light of fluctuating gut O₂ levels.

In terms of carbon utilization, genes responsible for acetate and lactate catabolism had strong fitness benefits in the gut (Fig. 3). These compounds likely represent major carbon/energy sources in vivo, being converted to acetyl-CoA and fed into the tricarboxylic acid (TCA) cycle for energy production and generation of biosynthetic precursors (e.g., via gluconeogenesis). All enzymes of the TCA cycle were needed for gut colonization, but not for growth in vitro, possibly due to compensation by TCA intermediates (e.g., succinate, citrate, malate) present in our culture media. The essentiality of TCA-mediated acetate and lactate utilization in vivo was also supported by the fact that mutants in the uptake and synthesis of cofactors for key enzymes in this pathway (e.g., lipolate and thiamine) were similarly strongly selected against in the gut (Fig. 3).

De novo biosynthetic pathways for amino acids and nucleic acids were required for host colonization, further implying low nutrient conditions within the bee hindgut. Most genes for purine and pyrimidine synthesis were nonessential in culture but needed in vivo. Similarly, most amino acid synthesis pathways were not required for growth in vitro, suggesting that *S. alvi* preferentially uptakes exogenous nutrients when available. In the gut, however, insertions in all genes involved in synthesizing 12 of the 20 amino acids were detrimental (Fig. 3). Intriguingly, this set of 12 contains the “essential amino acids” that cannot be synthesized by animals (including bees) (24); their scarcity may be due to efficient host absorption in the midgut. The distal hindgut is nutritionally impoverished in many animals due to upstream absorption. Nonetheless, the largest gut bacterial communities are typically found in hindguts, as exemplified by the human colon (25), the termite paunch (22), and the cecum of certain herbivores (26). The honey bee gut microbiota is also mainly in the hindgut (12), and our results show that a variety

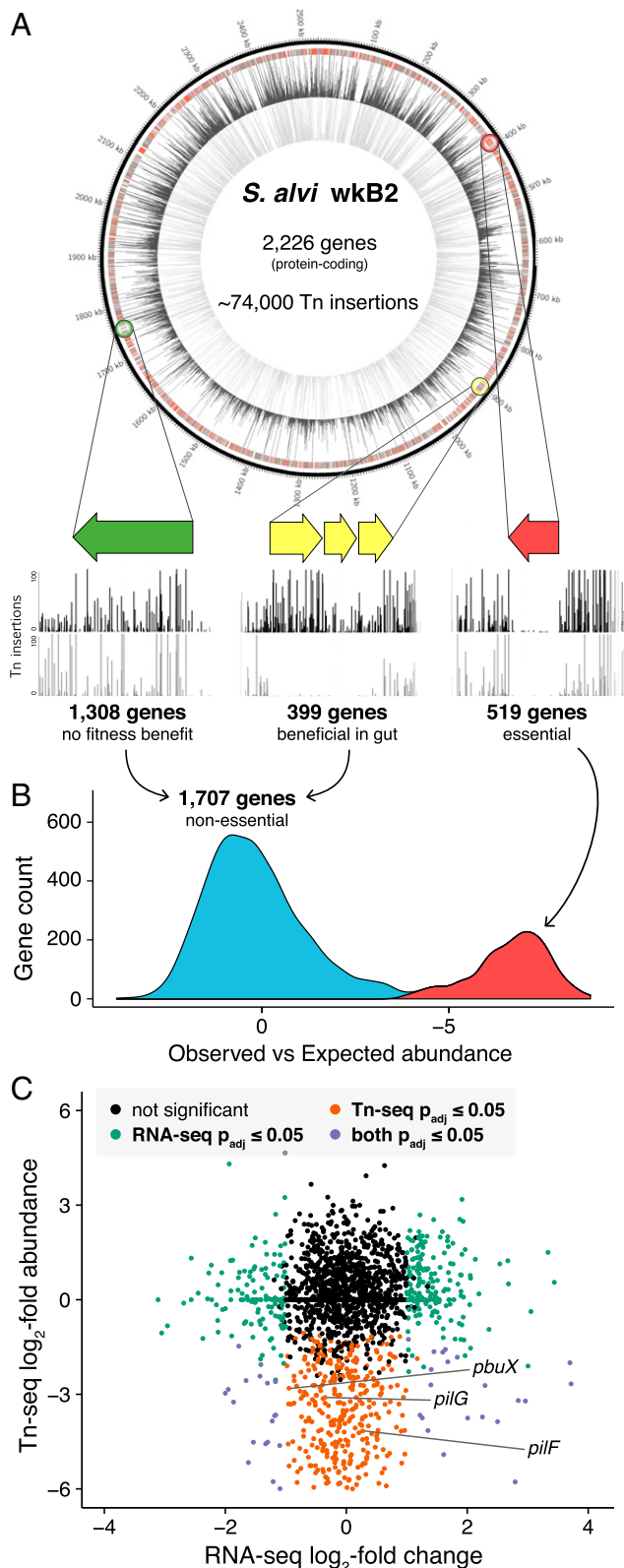


Fig. 2. Determining the *S. alvi* gene set essential for growth and beneficial for gut colonization. (A) Genome of *S. alvi* strain wkB2 with locations of transposon insertions in bee gut samples (inside track) and on culture media (outside track). Callouts illustrate three categories of genes identified from this approach. (B) The essential genome of *S. alvi* wkB2 on laboratory culture plates. (C) Tn-seq and RNA-seq results for *S. alvi* genes in the bee gut compared with culture plate growth. For Tn-seq, only genes with significant fitness benefit are highlighted.

of pathways are needed for tolerating the limited nutrient availability there.

Another vital micronutrient is iron, which is scarce in host-associated environments (27). That *S. alvi* possesses multiple systems for iron uptake (Fig. 3 and *SI Appendix, Fig. S2*) is indicative of its importance, particularly for iron bound in soluble complexes, the dominant form in aerobic environments. Three iron importers, for heme-bound Fe^{2+} and siderophore-bound Fe^{3+} , and associated genes (*tonB*, *exeBD*, and *fepA*), were necessary in vivo, as were the exporters *barAB* and *tolC* to avoid toxic buildup of siderophores (28). Siderophore synthesis, however, was not essential, presumably because mutants can “cheat” and take up siderophores secreted by nonmutants in the community. Indeed, mutants for siderophore synthesis were overrepresented in guts (*Dataset S1H*), suggesting that such cheaters prosper when they are at low frequencies by avoiding the costs of production.

Stress response. Many genes involved in stress responses confer strong fitness benefits in vivo, suggesting that the bee gut is a challenging environment in which *S. alvi* is exposed to toxins, low nutrient availability, and fluctuating O_2 , pH, and temperature. Among genes required in vivo were those underlying the ppGpp stringent response and Lrp, a master regulator that is induced by ppGpp upon amino acid starvation in other bacteria (29). In turn, Lrp can modulate expression of various amino acid synthesis enzymes and transporters to ensure cell survival (30).

Twenty-three *S. alvi* genes beneficial in the gut function to modify rRNA (16S and 23S rRNA subunits) and tRNA (e.g., via queuosine incorporation), with the likely effect of improving translational efficiency and fidelity. In other bacteria, these genes have been shown to enhance fitness under nutrient and temperature stress (31) (*Dataset S2*). Protein quality control, involving genes for protein recycling (*clpS*, *htpX*, *lon*, and *tldD*) and stabilization (*dsbC* and *pcm*), also appeared important in vivo, as were genes for the synthesis (*gshAB*) and activity (*gstA*) of glutathione, an antioxidant that contributes to general stress tolerance and detoxification (32).

A gene set with strong fitness benefits in the gut was that for the repair of double-stranded DNA breaks. This pathway includes genes for SOS response (*recA* and *recX*), recombinational repair (*recBCD*), D-loop extension (*priA*, DNA pol IV), resolution of Holliday junctions (*ruvABC* and *recG*), and postrepair chromosomal separation (*xerCD*) (Fig. 3 and *SI Appendix, Fig. S3*). DNA breaks may be induced by reactive oxygen species released by the dual oxidase component of the host immune system, which regulates gut microbiota in *Drosophila* (33) and necessitates DNA repair in the human gut bacterium *Helicobacter pylori* (34).

Response to membrane stress (e.g., via temperature, osmotic, ionic, pH shock, or LPS perturbation) was another critical feature for *S. alvi* gut colonization. A central component of this response appears to be the σ -24 factor (*rpoE*), which regulates, among other targets, protein quality control and LPS synthesis and export (35). The *mli* system, for maintaining outer membrane LPS integrity, and the poorly characterized membrane organizational protein, AsmA, were also important in vivo. Finally, a number of genes involved in cell division were intolerant of mutation in vivo; these may play a role in determining appropriate conditions for cellular replication, the regulation of which is likely crucial under stressful conditions (e.g., for *zapE*) (36).

We note that some mutants, such as those affecting genes underlying siderophore production, were overrepresented in the gut, suggesting that these genes hinder gut colonization under our experimental conditions (upper points of Fig. 2C and *Dataset S1H*).

Changes in Gene Expression upon Gut Colonization. In addition to the Tn-seq analysis, we used transcriptome sequencing to identify genes differentially expressed in the bee gut relative to our culture conditions. A total of 369 protein-coding genes were significantly up- or down-regulated twofold or higher (*Dataset S1D*).

Only 22 of these were among the 399 genes found to promote gut colonization by Tn-seq (Fig. 2C). These include genes underlying branched-chain amino acid synthesis, iron acquisition, and short-chain fatty acid utilization (*SI Appendix*, Fig. S1). Other up-regulated genes include genes coding for transporters involved in nutrient acquisition, and the type VI secretion system, implicated in interbacterial competition.

Testing Candidates from Tn-Seq for Roles in Gut Colonization. We tested colonization abilities of three insertion mutants with strong fitness detriment in the gut, by comparing their colonization success to that of a neutral transposon mutant, c10, in one-on-one competitions with wild-type strain wkB2. Two T4P structural mutants (*pilF*⁻ and *pilG*⁻) were examined in detail. In vitro evaluations showed that both *pilF*⁻ and *pilG*⁻ had reduced abilities to produce surface biofilms compared with wild-type *S. alvi* or to c10 (Fig. 4A and B and *SI Appendix* for explanation of screen). Scanning electron microscopy showed biofilms of multilayered composition along with prolific extracellular matrix in wild-type and c10, whereas the T4P mutants had small and isolated aggregations of cells (Fig. 4C). Lack of T4P-mediated adhesion was detrimental in vivo, as *pilF*⁻ and *pilG*⁻ mutants introduced to the bee gut had lower proportional recovery compared with the c10 mutant (Fig. 4D).

A mutant of a third gene, *pbuX*, was tested. This poorly characterized gene encodes a permease that is likely involved in xanthine/purine metabolism. Results verified that disruption of *pbuX* reduced colonization ability (Fig. 4E).

Comparison of *S. alvi* wkB2 Gene Sets to Those of Related Bacteria with Different Hosts. To better understand the basis for host specificity in *S. alvi* strains, we compared wkB2 to *S. alvi* wkB12, a strain restricted to *Bombus* hosts that is unable to colonize *A. mellifera* (14) (*Dataset S1G*). Of 2,226 wkB2 genes, 80% (1,762) are found in wkB12; of 519 essential wkB2 genes, 95% (497) are in wkB12; and of the 399 wkB2 genes beneficial for host colonization, all but 40 are present in wkB12. These 40 genes are potential determinants of host specificity. Most (26 of 40) are uncharacterized, but one of these is the major pilin subunit, *pilE* (SALWKB2_RS07815), which is highly expressed by wkB2 and significantly up-regulated in the gut.

We compared the genes of *S. alvi* wkB2 to those of *Neisseria gonorrhoeae* MS11, a human pathogen that is a close relative, for which a similar transposon mutant dataset is available (37) (*Dataset S1F*). Only 49% (1,085) of *S. alvi*'s 2,226 genes have orthologs in *N. gonorrhoeae*, but 90% (465) of the 519 *S. alvi* wkB2 essential genes are present, with 337 scored as essential. Among *S. alvi* genes important in gut colonization, 126 genes lack orthologs in *N. gonorrhoeae*, and these are candidates for mechanisms enabling colonization of insect guts.

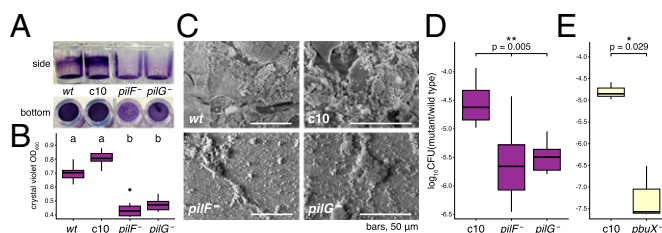


Fig. 4. In vivo competitive ability of mutants implicated in Tn-seq screen as having roles in gut colonization. (A–D) Roles of *pilF* and *pilG* in biofilm formation and host colonization. (A) Crystal violet-stained surface biofilms in polypropylene wells. (B) Mean solubilized crystal violet-stained biofilm, as absorbance at OD₆₀₀ ($n = 16$ wells per strain, $P = 1.91 \times 10^{-13}$, Kruskal–Wallis multiple comparisons). (C) Scanning electron micrographs of surface biofilms at the air/liquid interface of strains grown in vitro. (D and E) Ratio of viable transposon mutants to wild-type following coinoculation of bee guts. * $P < 0.029$, ** $P < 0.005$. (D) $n = 7$ bees per condition (Kruskal–Wallis test). (E) $n = 4$ bees per condition (Mann–Whitney test).

Conclusions

Our results provide a genome-scale view of pathways that enable host colonization by a specialized gut symbiont. We find that colonization depends on distinct processes, including cell surface modifications and biofilm formation, metabolic capabilities enabling growth in low nutrient conditions, and responses to diverse stressors. Some of these capabilities reflect the particular conditions of the bee gut ileum; for example, many amino acids appear to be scarce, possibly due to upstream host absorption, requiring de novo biosynthesis on the part of *S. alvi*. The retention of genes for operating under low O₂ conditions (e.g., the cytochrome *bd* complex, nitrate reductase, and the anaerobic *hemN* heme synthesis pathway) may likewise be explained by fluctuating O₂ availability at the gut wall, where *S. alvi* lives.

The ability of *S. alvi* to form resident populations in a persistent physical layer along the ileum is likely facilitated by T4P. T4Ps are known to play central roles in host colonization for other proteobacterial taxa, including close relatives of *Snodgrassella* such as *Neisseria meningitidis* and *N. gonorrhoeae*, via surface adhesion and motility, and are thus fundamental to biofilm organization and attachment to the host epithelium (19). Two other major cell surface components, trimeric auto-transporters and LPS O antigens, are also likely candidates for participation in adhesion, as well as in specific interactions with host receptors or evasion of host immune mechanisms (16, 17). These latter functions may give rise to the observed host specificity of particular *S. alvi* strains to particular host species (14).

The incongruence between the RNA-seq and Tn-seq results was not surprising, as transcriptional responses correlate poorly with gene essentiality (38), potentially reflecting either constitutive expression or very transitory expression of many genes. For instance, genes for adhesion might only be highly expressed during an initial critical stage of laying down biofilm. Nonetheless, our transcriptomic results reinforce the importance of nutrient uptake and amino acid synthesis by *S. alvi* in the gut. Some genes, such as RTX toxins and type VI secretion systems, are not essential for gut colonization, but were up-regulated in vivo. These features may affect colonization in the presence of other members of the bee gut microbiome by mediating interbacterial interactions. Studying the responses of *S. alvi* as part of the wider gut community is a logical next step, as other bacterial species may considerably influence the gut environment by liberating nutrients from pollen (39), eliciting host responses (40), altering pH and O₂ concentration, or engaging in direct competition.

To date, there have been few genome-wide studies of the factors underlying colonization by gut commensals in their native host environment. Direct experimental approaches are not feasible for human gut symbionts, but rodent models have been used with considerable success (2–7). Several pathogens have been examined using Tn-seq in vivo, within different tissue types (38, 41–43). Some critical mechanisms for *S. alvi* colonization of the bee gut, including LPS modifications, membrane integrity components, the stringent response, proteases, and DNA break repair, are also important for infection by human pathogens (41–43). Similarly, colonization of light-producing organs by the squid symbiont *Vibrio fischeri* depends on genes dictating biofilm formation and cellular stress responses (44). In aquatic *Vibrio* colonizing zebra fish intestines, genes underlying cell envelope, chemosensory and motility functions seem crucial (45). Hence, many animal-associated bacteria, from pathogens to mutualists, likely face common challenges and rely on a similar set of genes to establish in the host. Whereas many of these genes are part of well-characterized pathways, others remain poorly described despite their importance across a range of host–microbe systems. Examples of enigmatic genes that strongly contribute to *S. alvi* fitness in the bee gut, and also in other animal symbionts, include the *ntrXY* two-component system, the T4P-associated *fimV*, the RNA helicase *hmpA*, the DNA repair regulator *yebC*, the protein stabilizer *dsbC*, and *asmA* (*Dataset S2*).

High-throughput screening of mutant libraries derived from bacterial colonizers in vivo opens a window into the gut milieu as

well as the adaptations required to live under such conditions. We uncovered gene networks involved in extracellular interaction, metabolism, and stress responses that appear critical for gut colonization; these genes may be of general importance to host-associated microbiota, and are ideal candidates for future analyses. Finally, our study demonstrates the genetic tractability of *S. alvi*, a species only recently discovered (10), and paves the way for new applications and engineering of the bee microbiome.

Materials and Methods

Detailed protocols are available in *SI Appendix*. Strains are listed in *SI Appendix*, Table S2. *S. alvi* wkb2 was mutagenized with a Mariner-derived transposon bearing a gentamycin resistance marker, delivered by a counterselectable *Escherichia coli* strain via conjugation with plasmid pBT20 (*SI Appendix*, Figs. S4 and S5). Experiments verified that this transposon made single random insertions in the wkb2 genome and that strains carrying this marker can colonize the bee gut to the same abundance as the wild type (*SI Appendix*, Fig. S6).

A saturated transconjugant library was constructed and harvested from replicates from rich agar-based media (control group) or from the ileums of pooled cohorts of worker bees 5 d after inoculation (experimental group).

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