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Review

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Ellen M. Moran, Frank L. Mastaglia

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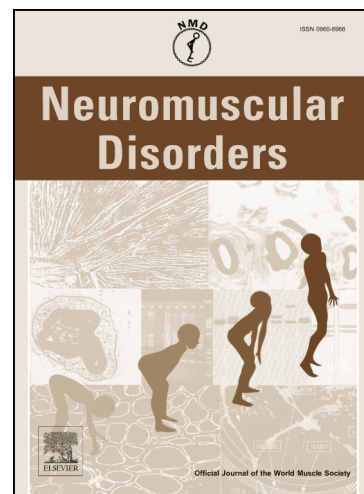
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**The role of Interleukin-17 in immune-mediated inflammatory myopathies and possible
therapeutic implications**

Ellen M. Moran^{1*} and Frank L. Mastaglia^{1,2}

1. Institute for Immunology & Infectious Diseases (IIID), Murdoch University, Murdoch, WA, Australia
2. Western Australian Neuroscience Research Institute, Centre for Neuromuscular & Neurological Disorders, University of Western Australia, Australia.

*Corresponding author. Address: Institute for Immunology & Infectious Diseases (IIID), Building 390, Discovery Way, Murdoch University, Murdoch, Western Australia 6150, Australia. Tel.: +61 8 9360 1368; Fax: +61 8 9360 1366. *Email address: e.moran@iid.com.au*

Abstract

The idiopathic inflammatory myopathies are a heterogeneous group of autoimmune muscle disorders with distinct clinical and pathological features and underlying immunopathogenic mechanisms. Traditionally, CD4⁺Th1 cells or CD8⁺ cytotoxic effector T cells and type I/II interferons have been primarily implicated in the pathogenesis of the inflammatory myopathies. The presence of IL-17A producing cells in the inflamed muscle tissue of myositis patients and the results of *in vitro* studies suggest that IL-17A and the Th17 pathway may also have a key role in these diseases. The contribution of IL-17A to other chronic inflammatory and autoimmune diseases has been well established and clinical trials of IL-17A inhibitors are now at an advanced stage. However the precise role of IL-17A in the various forms of myositis and the potential for therapeutic targeting is currently unknown and warrants further investigation.

Keywords: inflammatory myopathies, dermatomyositis, polymyositis, inclusion body myositis, IL-17A, Th17

1. Introduction

The idiopathic inflammatory myopathies (IIMs) are a heterogeneous group of autoimmune muscle disorders that include dermatomyositis (DM), polymyositis (PM), inclusion body myositis (IBM), and the immune-mediated necrotising myopathies (IMNM)[1]. The association with myositis-specific autoantibodies and certain HLA alleles, and the overexpression of Class I and II major histocompatibility complex molecules by muscle cells and predominance of T cells and B cells in affected muscles has implicated an autoimmune origin despite the fact that the target autoantigen in DM, PM and IBM have yet to be identified. Whilst these conditions share a number of common properties, namely muscle weakness and inflammation, they have distinct clinical and pathological features and are thought to have different underlying immunopathogenic mechanisms. These have been reviewed previously [2-4], as have the underlying cellular and molecular mechanisms that may contribute to the muscle weakness in the IIMs[5]. Dermatomyositis is usually regarded as a CD4⁺ T-cell-driven disease in which a complement-dependent humorally mediated attack on the vascular endothelium results in skin and muscle injury[6, 7]. In contrast, both PM and IBM are thought to share a CD8⁺ T-cell-mediated autoimmune process[8, 9] where muscle fibres expressing MHC-I antigens are invaded by CD8⁺ T lymphocytes which are clonally expanded *in situ*, and drive the induction of cytotoxic necrosis through the liberation of perforins and granzymes[10]. In spite of this, it is recognised that the involvement of these T cell subsets is not necessarily exclusive and that there is a certain amount of overlap in the immunopathological phenotype in the different forms of IIM. Moreover, the demonstration

of a subset of CD28^{null} CD4⁺ and CD8⁺ cells with a natural killer cell phenotype in DM and PM tissues[11] suggests that there may be multiple mechanisms of T cell mediated cytotoxicity. In contrast to the other IIMs, IBM is distinguished by the fact that the inflammatory process is accompanied by myodegenerative changes and abnormal protein aggregation and inclusion body formation in muscle fibres [12].

The majority of research to date has implicated CD4⁺ T-helper (Th1) cells or CD8⁺ cytotoxic effector T cells in the pathogenesis of the inflammatory myopathies, as evidenced by the detection of a type 1 interferon profile in both DM and PM and a Th1 immune profile in IMNM[13, 14]. Interleukin-17 (IL-17) and the Th17 subset of CD4⁺ cells have been strongly implicated in the pathogenesis of a number of other autoimmune diseases including psoriasis, rheumatoid arthritis and multiple sclerosis leading to the development of IL-17 targeted monoclonal antibody therapies. In this review we summarise recent observations on IL-17 and the Th17 pathway in the inflammatory myopathies and their potential as novel therapeutic targets for the treatment of these diseases.

2. Immunopathogenic mechanisms

2.1 Polymyositis and inclusion-body myositis

In PM and IBM CD8⁺ T cells are considered to be the primary effector cells mediating muscle-fibre injury[8, 9, 15] These cytotoxic T cells target muscle fibres expressing MHC Class I molecules and co-stimulatory molecules[16-19] The muscle fibres may therefore act as antigen presenting cells [20, 21] and have been postulated to form immunological synapses with the T - cell receptors (TCRs) of the auto-invasive, clonally expanded CD8⁺ T cells[2]. These T cells have been shown to be activated, expressing ligands specific for the co-stimulatory molecules expressed by the muscle fibres[2]. Matrix-metalloproteinases

(MMPs) are thought to facilitate the transendothelial migration of T cells and their attachment to the surface of the muscle fibres[2]. The T cells also display cytotoxic properties by releasing perforin granules which are directed to the muscle fibres resulting in necrosis[10]. Cytokines such as IFN- γ , IL-1 and TNF secreted by activated CD8⁺ T cells may also contribute to muscle damage through a direct myocytotoxic effect[1].

2.2 Dermatomyositis

The disease process in DM is thought to be initiated by antibodies directed against as yet unknown antigens expressed by the vascular endothelium activating the complement pathway[1]. However, endothelial cell specific autoantibodies have as yet not been identified. Membrane attack complexes (MAC) form on the endomysial capillaries[6, 7] and are thought to cause endothelial cell necrosis leading to capillary depletion and muscle ischaemia[22-24] although the mechanisms leading to capillary loss have yet to be fully elucidated. It is postulated that deposition of immunoglobulins on intramuscular capillaries activates the complement cascade, triggering the production of pro-inflammatory cytokines and chemokines, which in turn up-regulate the expression of adhesion molecules on endothelial cells leading to further recruitment of B cells, T cells and macrophages and interferon- α producing plasmacytoid dendritic cells [1, 25]. The release of other cytokines and soluble mediators such as TNF and NO by the activated T and B cells may further enhance the inflammatory processes taking place in the muscle[4]. Multiple findings indicate that upregulation of the Type 1 interferon pathway plays a prominent role in the disease pathogenesis in DM[26, 27] although it is not specific to DM[29].

2.3 Myositis Specific Autoantibodies

Myositis-specific autoantibodies (MSA) that target various ubiquitous autoantigens are well documented in both DM and PM, and more recently also in IMNM and IBM, and their potential value as diagnostic biomarkers is being increasingly recognised[30-34]. The best characterised are the antisynthetase antibodies which target a group of cytoplasmic aminoacyl tRNA synthetases[35]the most prevalent of which is anti-Jo-1 (anti-histidyl tRNA synthetase) which is present in about 20% of cases of PM and DM and is associated with the antisynthetase syndrome[36, 37]Other less common antisynthetases include anti-PL-12 (alanyl-tRNA synthetase), anti-PL-7 (threonyl-tRNA synthetase), anti-EJ (glycyl-tRNA synthetase), anti-OJ (isoleucyl-tRNA synthetase), anti-KS (asparaginyl-tRNA synthetase), anti-Zo (phenylalanyl-tRNA synthetase) and anti-Ha (tyrosyl-tRNA synthetase) [38].In DM antibodies to Mi-2, which is a component of the nucleosome remodelling deacetylase complex, has a high specificity, particularly for the adult form of the disease[39, 40].More recently, a number of newer autoantibodies associated with particular subgroups of DM cases have been characterised including anti-TIF-1 α/γ , anti-MDA5 and anti-NXP-2 and these have been reviewed elsewhere[31].

There is increasing evidence that in addition to their role as biomarkers MSA may also play a part in inducing and sustaining the autoimmune process in the IIM. For example, a number of studies have shown that there is a correlation between serum levels of anti-Jo-1 antibodies and both disease activity [42] and muscle damage [41], suggesting that the antibody has a causative role. In addition, histidyl-tRNA synthetase and Mi-2 are over-expressed in muscle tissue in PM and DM[43] and expression of Mi-2 has been shown to

be increased in regenerating fibres in DM muscles and in human myoblasts in culture [31, 44]. These observations have led to the suggestion that enhanced tissue expression of these autoantigens as part of the repair process in muscle may play a critical role in maintaining an autoantibody response and the ongoing autoimmune process in PM and DM [31, 45]. MSA such as histidyl-tRNA synthetase may also play a part in the recruitment of T cells and dendritic cells by activating chemokine receptors on these cells [46].

2.4 Cytokine and Chemokine Involvement

A common pathogenic mechanism in the inflammatory myopathies is the specific migration of T cell subsets to the muscle tissue which involves activation of leukocytes, adhesion to the vasculature and transendothelial migration. Molecules associated with T cell transmigration and cytokine/chemokine signalling have been found to be differentially overexpressed in PM, DM and IBM [1, 47-50]. Furthermore, cultured muscle cells can secrete proinflammatory cytokines upon stimulation in a positive feedback loop mechanism that attracts activated T cells, and Toll-like receptors (TLRs) expressed by muscle tissue from patients with inflammatory myopathies enhance cytokine secretion. Therefore, muscle fibres in addition to being the target of inflammation contribute to creating a proinflammatory environment that drives disease chronicity [48, 51, 52]. Microarray gene expression profiling has detected upregulation of genes for adhesion molecules, cytokines, MHC class I molecules, chemokines and immunoglobulins in the muscles of patients with inflammatory myopathies [4, 53-56]. In particular, the interferons (IFNs) and IFN-inducible genes have been strongly implicated in the pathogenesis of myositis. The enhanced type I IFN gene signature has been associated with disease activity in both PM and

DM[57]. Additionally, immunostaining for type I IFN has been detected in DM in muscle fibres in areas showing perifascicular atrophy, as well as in plasmacytoid dendritic cells and in the capillary endothelial cells [27, 58]. In contrast, overexpression of the type III IFN (IFN- γ) induced genes has been associated with inclusion body myositis, being localised to muscle fibres invaded by CD8⁺ T cells [49, 59].

3. IL-17 and Cellular Sources of IL-17

3.1 Discovery of IL-17

IL-17 is a pro-inflammatory cytokine demonstrated to contribute to several inflammatory and autoimmune diseases [60]. Key sources of IL-17 are the T helper 17 cells (Th17) as well as cells from the innate immune system such as mast cells and neutrophils [61, 62]. IL-17 is the founding member of the IL-17 cytokine family comprising six members: IL-17 (IL-17A), IL-17B, IL-17C, IL-17D, IL-17E (also known as IL-25) and IL-17F. IL-17 was first discovered in 1993 when a molecule termed 'cytotoxic T-lymphocyte-associated antigen 8' (CTLA-8) was isolated from rodent T cell hybridoma clones. Further work subsequently demonstrated its ability to stimulate the production of cytokines from synoviocytes and mesenchymal cells [63, 64]. The results of this work and the identification of a receptor led to the naming of a novel cytokine IL-17 and its receptor IL-17R [65, 66]. IL-17 and IL-17F share a high degree of homology and both bind to the same receptor implying common biological activities. Additionally, IL-17 can exist as a homodimer or a heterodimer with IL-17F [67].

3.2 Th17 cells and other cellular sources of IL-17

Upon its discovery, IL-17 was initially described as a T cell product. Subsequent studies indicated IL-17 secreting cells were Th1/Th0, not TH2, and hinted that IL-17 may define a new subset of Th1 cells[68-72]. Previous to this the pathogenesis of autoimmune diseases had been attributed to TH1 cells in line with the Th1/Th2 model devised by Mossman and Coffman[73]. This hypothesis was challenged when it was demonstrated that mice deficient in IFN- γ expression and signalling were still susceptible to mouse models of multiple sclerosis and inflammatory arthritis[74]. Eventually, a decade after its discovery, IL-17 producing Th17 cells were demonstrated to be a distinct lineage from Th1 or Th2 cells and to be pathogenic in autoimmunity[75-77]. Th17 cells differentiate from naïve precursor T cells (Th0) cells through antigen presentation and co-stimulation and the effects of Th17 cell-polarizing cytokines (TGF- β , IL-21, IL-6 and IL-23)[67](Figure 1). Subsequent research has identified additional non-CD4⁺ cell types that are also key sources of IL-17 in autoimmune and inflammatory diseases, which include CD8⁺ Tc17 cells, $\gamma\delta$ T cells, natural killer cells, neutrophils and mast cells[61, 62, 78]. Studies suggest that these additional IL-17 secreting cell types, in particular CD8⁺ Tc17 cells, may develop under similar inflammatory conditions as Th17 cells and may have common inhibitory control mechanisms[62, 79, 80]. Th17 cells have also been shown to have a similar development pathway but opposing effects in the inflammatory response to CD4⁺CD25⁺Foxp3⁺regulatory T cells (Tregs). It is recognised that there is a fine balance between Th17 and Tregs in maintaining tissue homeostasis and a disruption of this balance can lead to chronic inflammation and autoimmunity. Defects in Treg numbers and function have been well characterised in a number of inflammatory and autoimmune conditions[81, 82].

3.3 Biological effects of IL-17

The main IL-17-responsive cell types are epithelial cells, endothelial cells, fibroblasts, macrophages, and dendritic cells. Furthermore, T cells as well as being producers of IL-17 are also responsive to the cytokine. Upon binding to its receptor, IL-17 stimulates downstream signalling pathways leading to the activation of several common transcription factors (AP-1, NF- κ B, PI3K/JAK, C/EBP), inducing the gene transcription of cytokines, chemokines, growth factors, MMPs and antimicrobial peptides in target cells[83-85]. Additionally, IL-17 can stabilize the mRNA of certain proinflammatory cytokines and chemokines induced by TNF- α [85]. Furthermore, the IL-17 induction of cytokine and chemokine expression can be enhanced by co-stimulation with TNF- α , OncostatinM (OSM), IL-1 β or TLR ligands[72, 84]. IL-17 has been shown to play a key role in driving inflammatory cell migration and invasion by mediating cytoskeletal rearrangement through the activation of integrin-linked/RhoGTPase pathways[83]. Recent studies have also demonstrated that IL-17 is able to regulate lymphocyte function and was shown to modulate the survival and differentiation of antibody producing B cells in mouse models of autoimmune diabetes[86-89]. IL-17 has also been shown to promote the polarization and function of TH1 cells *in vitro* and *in vivo*[90, 91].

4. Potential role for IL-17 in immunopathogenesis of myositis

4.1 Expression of IL-17 in inflammatory myopathies

Several studies have investigated the presence of IL-17 in muscle tissue or blood in patients with inflammatory myopathies, however these have focused mainly on DM and PM and few

on IBM. Immunohistochemistry has detected IL-17 producing cells in Tcell rich areas of DM and PM tissue, although the number of IL-17 positive cells was low compared to IFN- γ producing cells[92-94]. In addition, increased expression of IL-17 mRNA has been demonstrated in DM, PM and IBM muscle. The detection of IL-17 was also shown to be strongly associated with increased levels of expression of TLR, IFN- γ and MyD88 in these studies [95, 96]. Increased protein expression of another Th17 cytokine IL-22 has been observed in the tissues of DM and PM patients and correlated with markers of myositis activity. Furthermore a number of these IL-22+ T cells co-expressed IL-17 [97]. Increased expression of IL-17 has also been demonstrated in whole blood samples from PM and DM patients, with a subset of DM patients showing an enhanced IL-17 gene signature rather than type I IFN[98].

A number of studies have reported increased serum levels of IL-17 in patients with DM or PM[99-101] and levels significantly correlated with disease activity[100-102]. One study in DM and PM patients found that serum levels of IL-17 were not significantly higher than in healthy controls, although this may have been due to the ELISA sensitivity[103]. Additionally, in an analysis of peripheral blood mononuclear cells (PBMC) from IBM patients the percentage of CD3⁺CD4⁺ IL-17⁺ cells was not higher than in healthy controls[104]. However, activated PBMC from patients with early PM and DM were shown to secrete significantly higher amounts of IL-17 than those with established disease[103]. The frequency of Th17 cells[94, 105] in PM and DM PBMC and Th17 cytokines (IL-6, IL-23, IL-1 β and TGF- β) in DM have also been shown to be increased in comparison to healthy controls[105]. IMNM has been characterized by a Th1 immune response however this study did not assess IL-17/Th17 expression [14].

4.2 Actions of IL-17 on muscle

A limited number of studies have explored the direct effects of IL-17 on muscle cells (Figure 2). The first in 2003 examined the effect of IL-17 alone and in combination with IL-1 β on the expression of cell-adhesion molecules and cytokine/chemokine production by normal human myoblasts and muscle tissue samples *in vitro*[92]. IL-17 alone and in combination with IL-1 β stimulated IL-6 production by myoblasts and cultured muscle samples. While IL-17 alone had no effect on CCL20 production, IL-17 amplified the effects of IL-1 β on IL-6 and CCL20 secretion by the muscle explants. In another study the expression of HLA class I, C-Fos, NF- κ B and C-Jun was also found to be increased by IL-17 stimulation[106]. A recent study demonstrated the ability of IL-17 to inhibit myoblast migration and myogenic differentiation in an ERK1,2 dependent manner[107]. In a further study IL-17 was shown to mediate this effect through the inhibition of urokinase type plasminogen inhibitor (uPA), a key molecule in skeletal myogenesis[108].

5. Strategies for targeting IL-17 therapeutically and relevance to IIM

5.1 Indirect targeting

IL-17 has been targeted indirectly in myositis patients using the IL-1 receptor antagonist anakinra[109]. The study cohort was heterogeneous and included DM, PM and IBM patients. IL-17 expressing CD4⁺ T cells were reduced in three out of six patients after 6 months treatment and a shift in T cell differentiation from Th17 to Th1 was observed in 5 out of 6 patients. This was confirmed by *in vitro* studies that showed a reduction in PBMC IL-17A secretion by PBMCs 92-98%. The TNF inhibitor adalimumab was also tested but showed

no inhibitory effect[109]. Treatment with rituximab did not result in a decrease of Th17 cells in two studies of refractory PM and DM patients[102, 110]. Interestingly, in one study a significant increase in Th17 cytokine levels was observed[102].

Administration of intravenous immunoglobulin (IVIG) has been shown to be effective in patients with DM and PM who are unresponsive to glucocorticoids or immunosuppressive drugs[111-114]. Expression of IL-17 and IFN- γ was quantified in muscle biopsies from a cohort of 13 myositis patients (11PM and 2DM) pre- and post-IVIG treatment, seven of whom were defined as being responders. IL-17 and IFN- γ positive cells were detected in 11/13 patients. The numbers of IFN- γ positive cells and the ratio of IFN- γ /IL-17-producing cells were significantly higher in non-responders than responders, but there was no differential expression of IL-17 producing cells in the two groups. These observations suggest that the Th1/Th17 cell balance is important in determining the response to IVIG therapy in myositis[94].

5.2 Induction of regulatory T cells

A number of autoimmune or inflammatory diseases are characterized by a decrease in Treg cell numbers and/or function. Foxp3+ T regulatory cells have been detected in patients with DM, PM and IBM and their numbers have been found to inversely correlate with the severity of disease activity[115]. The number of circulating Tregs in patients with IBM and DM has been documented to be lower than in healthy controls[116, 117]. Furthermore, the serum levels of the Treg associated cytokines TGF- β and IL-10 are significantly reduced in DM patients[116]. The induction/and or expansion of Tregs is currently being examined as a therapeutic strategy in the treatment of inflammatory diseases, in particular as an inhibitor of Th17-mediated autoimmunity[118, 119]. The balance between Th17 and Treg cells in

myositis patients has not been well characterised. One study in IBM patients described an enhanced Th1 profile with decreased Treg numbers compared to controls, however no increase in Th17 cells was found[117].

5.3 Targeting of Th17 related molecules

IL-6 is a key factor in the differentiation of CD8⁺ Tc17 and Th17 cells[67] and studies suggest the cytokine plays an important role in myositis, in particular in PM[120]. Overexpression of IL-6 has been detected in the serum and muscle tissue of patients with IBM, PM and DM[121-123]. Furthermore, in mouse models of myositis inhibiting IL-6 by gene knockout or with an anti-IL-6 receptor antibody ameliorated myositis severity. Tocilizumab, a humanized anti-IL-6 receptor antibody, has been shown to be successful in the treatment of two patients with resistant PM[124].

The differentiation of IL-17 secreting T cells such as Th17 and CD8 Tc17 cells requires the expression of the 'retinoic acid receptor-related orphan nuclear receptor' ROR γ t[124, 125]. A number of ROR γ t small-molecule inhibitors have been identified and have been shown to inhibit the differentiation and function of Th17 and CD8Tc17 cells[80, 126]. This approach is thought to be particularly advantageous as targeting ROR γ t inhibits not only Th17 differentiation and IL-17 secretion but also the production of other proinflammatory cytokines secreted by Th17 cells. Furthermore, ROR γ t agonists could potentially be delivered orally in comparison to the other injectable biologic therapies. A number of these molecules are currently in clinical development [127, 128].

5.4 Direct Targeting of IL-17

A number of monoclonal antibodies directly targeting IL-17A and its receptor are currently undergoing clinical trials for the treatment of a variety of inflammatory and autoimmune diseases including psoriasis, rheumatoid and psoriatic arthritis, ankylosing spondylitis, multiple sclerosis and asthma [67, 128] but as yet have not been trialled in inflammatory myopathies. Clinical trials assessing IL-17 inhibitors for the treatment of psoriasis are the most advanced, with the IL-17A inhibitor secukinumab having completed phase III trials late last year and are expected to receive approval. Two other inhibitors targeting IL-17A (Ixekizumab) and its receptor IL-17R (Brodalumab) are currently in phase III trials in psoriasis patients [128]. Results from trials to date suggest that the success of IL-17 inhibition is dependent on the specific tissue, cellular and immunological context. For example anti-IL-17A antibodies are showing success in psoriasis and multiple sclerosis but not in Crohn's disease [60].

6. Conclusions

IL-17A has been firmly established as a key mediator in the pathogenesis of a number of chronic inflammatory and autoimmune diseases such as psoriasis and rheumatoid arthritis. The investigations into the efficacy of pharmacological inhibitors targeting the IL-17 pathway are at an advanced stage and the approval of at least one anti-IL-17A drug for the treatment of these conditions is expected imminently. The detection of IL-17A in muscle tissue of myositis patients and *in vitro* studies suggest that IL-17A also plays an important role in the pathogenesis of the inflammatory myopathies and may be a potential therapeutic target in cases resistant to treatment. To date, these studies have been limited and have

mainly focused on PM and DM rather than IBM. Further studies examining the expression profile of IL-17A in the different inflammatory myopathies, as well as the cellular sources of IL-17A are necessary to fully elucidate the contribution of this proinflammatory cytokine to disease pathogenesis.

Search Strategy and selection criteria

Pubmed was searched from 1990 to 2014 using the terms inflammatory myopathies, dermatomyositis, polymyositis, inclusion body myositis, muscle, IL-17, Th17, cytotoxic T cells and cellular sources of IL-17. We also identified further articles through searches of reference lists from these articles. We reviewed only articles published in the English language.

Figure Captions

Figure 1. Differentiation pathways, cytokine profiles and functions of Th1, Th2, Th17 and Treg cell subsets

Figure 2. Putative mechanisms of action of IL-17 on myoblasts and downstream effects. IL-17RA and IL-17RC represent the A and C chains of the IL-17 receptor; TLR: Toll-like receptors; GPCR: G-protein coupled receptor; HLA-I: Human Leukocyte Antigen Class 1; NF- κ B; nuclear factor kappa-light-chain-enhancer of activated B cells.

References

- [1] Dalakas MC. Pathogenesis and therapies of immune-mediated myopathies. *Autoimmun Rev* 2012;11:203-6.
- [2] Dalakas MC. Mechanisms of disease: signaling pathways and immunobiology of inflammatory myopathies. *Nat Clin Pract Rheumatol* 2006;2:219-27.
- [3] Greenberg SA. A gene expression approach to study perturbed pathways in myositis. *Curr Opin Rheumatol* 2007;19:536-41.
- [4] Zong M, Lundberg IE. Pathogenesis, classification and treatment of inflammatory myopathies. *Nat Rev Rheumatol* 2011;7:297-306.
- [5] Loell I, Lundberg IE. Can muscle regeneration fail in chronic inflammation: a weakness in inflammatory myopathies? *J Intern Med* 2011;269:243-57.
- [6] Kissel JT, Halterman RK, Rammohan KW, Mendell JR. The relationship of complement-mediated microvasculopathy to the histologic features and clinical duration of disease in dermatomyositis. *Arch Neurol* 1991;48:26-30.
- [7] Kissel JT, Mendell JR, Rammohan KW. Microvascular deposition of complement membrane attack complex in dermatomyositis. *N Engl J Med* 1986;314:329-34.
- [8] Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopathies. III: Immunoelectron microscopy aspects of cell-mediated muscle fiber injury. *Ann Neurol* 1986;19:112-25.
- [9] Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopathies. V: Identification and quantitation of T8+ cytotoxic and T8+ suppressor cells. *Ann Neurol* 1988;23:493-9.
- [10] Goebels N, Michaelis D, Engelhardt *Met al.* Differential expression of perforin in muscle-infiltrating T cells in polymyositis and dermatomyositis. *J Clin Invest* 1996;97:2905-10.
- [11] Fasth AE, Dastmalchi M, Rahbar *Aet al.* T cell infiltrates in the muscles of patients with dermatomyositis and polymyositis are dominated by CD28null T cells. *J Immunol* 2009;183:4792-9.
- [12] Askanas V, Engel WK, Nogalska A. Pathogenic considerations in sporadic inclusion-body myositis, a degenerative muscle disease associated with aging and abnormalities of myoproteostasis. *J Neuropathol Exp Neurol* 2012;71:680-93.
- [13] Cappelletti C, Baggi F, Zolezzi *Fet al.* Type I interferon and Toll-like receptor expression characterizes inflammatory myopathies. *Neurology* 2011;76:2079-88.

- [14] Preusse C, Goebel HH, Held *et al.* Immune-mediated necrotizing myopathy is characterized by a specific Th1-M1 polarized immune profile. *Am J Pathol* 2012;181:2161-71.
- [15] Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopathies. IV: Cell-mediated cytotoxicity and muscle fiber necrosis. *Ann Neurol* 1988;23:168-73.
- [16] Behrens L, Kerschensteiner M, Misgeld T, Goebels N, Wekerle H, Hohlfeld R. Human muscle cells express a functional costimulatory molecule distinct from B7.1 (CD80) and B7.2 (CD86) in vitro and in inflammatory lesions. *J Immunol* 1998;161:5943-51.
- [17] Murata K, Dalakas MC. Expression of the costimulatory molecule BB-1, the ligands CTLA-4 and CD28, and their mRNA in inflammatory myopathies. *Am J Pathol* 1999;155:453-60.
- [18] Wiendl H, Mitsdoerffer M, Schneider *et al.* Muscle fibres and cultured muscle cells express the B7.1/2-related inducible co-stimulatory molecule, ICOSL: implications for the pathogenesis of inflammatory myopathies. *Brain* 2003;126:1026-35.
- [19] Schmidt J, Rakocevic G, Raju R, Dalakas MC. Upregulated inducible co-stimulator (ICOS) and ICOS-ligand in inclusion body myositis muscle: significance for CD8+ T cell cytotoxicity. *Brain* 2004;127:1182-90.
- [20] Goebels N, Michaelis D, Wekerle H, Hohlfeld R. Human myoblasts as antigen-presenting cells. *J Immunol* 1992;149:661-7.
- [21] Wiendl H, Lautwein A, Mitsdorffer *et al.* Antigen processing and presentation in human muscle: cathepsin S is critical for MHC class II expression and upregulated in inflammatory myopathies. *J Neuroimmunol* 2003;138:132-43.
- [22] De Visser M, Emslie-Smith AM, Engel AG. Early ultrastructural alterations in adult dermatomyositis. Capillary abnormalities precede other structural changes in muscle. *J Neurol Sci* 1989;94:181-92.
- [23] Emslie-Smith AM, Engel AG. Microvascular changes in early and advanced dermatomyositis: a quantitative study. *Ann Neurol* 1990;27:343-56.
- [24] Gitiaux C, Kostallari E, Lafuste P, Authier FJ, Christov C, Gherardi RK. Whole microvascular unit deletions in dermatomyositis. *Ann Rheum Dis* 2013;72:445-52.
- [25] Dalakas M, Hohlfeld R. Polymyositis and dermatomyositis. *Lancet* 2003;362:971-82.
- [26] Greenberg SA, Pinkus JL, Pinkus *et al.* Interferon-alpha/beta-mediated innate immune mechanisms in dermatomyositis. *Ann Neurol* 2005;57:664-78.
- [27] Salajegheh M, Kong SW, Pinkus *et al.* Interferon-stimulated gene 15 (ISG15) conjugates proteins in dermatomyositis muscle with perifascicular atrophy. *Ann Neurol* 2010;67:53-63.
- [28] Englund P, Nennesmo I, Klareskog L, Lundberg IE. Interleukin-1alpha expression in capillaries and major histocompatibility complex class I expression in type II muscle fibers from polymyositis and dermatomyositis patients: important pathogenic features independent of inflammatory cell clusters in muscle tissue. *Arthritis Rheum* 2002;46:1044-55.
- [29] Eloranta ML, Barbasso Helmers S, Ulfgren AK, Ronnblom L, Alm GV, Lundberg IE. A possible mechanism for endogenous activation of the type I interferon system in myositis patients with anti-Jo-1 or anti-Ro 52/anti-Ro 60 autoantibodies. *Arthritis Rheum* 2007;56:3112-24.
- [30] Love LA, Leff RL, Fraser *et al.* A new approach to the classification of idiopathic inflammatory myopathy: myositis-specific autoantibodies define useful homogeneous patient groups. *Medicine (Baltimore)* 1991;70:360-74.
- [31] Casciola-Rosen L, Mammen AL. Myositis autoantibodies. *Curr Opin Rheumatol* 2012;24:602-8.
- [32] Mohassel P, Mammen AL. Statin-associated autoimmune myopathy and anti-HMGCR autoantibodies. *Muscle Nerve* 2013;48:477-83.
- [33] Larman HB, Salajegheh M, Nazareno *et al.* Cytosolic 5'-nucleotidase 1A autoimmunity in sporadic inclusion body myositis. *Ann Neurol* 2013;73:408-18.
- [34] Pluk H, van Hoeve BJ, van Dooren *et al.* Autoantibodies to cytosolic 5'-nucleotidase 1A in inclusion body myositis. *Ann Neurol* 2013;73:397-407.

- [35] Mathews MB, Bernstein RM. Myositis autoantibody inhibits histidyl-tRNA synthetase: a model for autoimmunity. *Nature* 1983;304:177-9.
- [36] Hengstman GJ, Brouwer R, Egberts WT *et al.* Clinical and serological characteristics of 125 Dutch myositis patients. Myositis specific autoantibodies aid in the differential diagnosis of the idiopathic inflammatory myopathies. *J Neurol* 2002;249:69-75.
- [37] Hengstman GJ, van Brenk L, Vree Egberts WT *et al.* High specificity of myositis specific autoantibodies for myositis compared with other neuromuscular disorders. *J Neurol* 2005;252:534-7.
- [38] Gunawardena H, Betteridge ZE, McHugh NJ. Myositis-specific autoantibodies: their clinical and pathogenic significance in disease expression. *Rheumatology* 2009;48:607-12.
- [39] Targoff IN, Reichlin M. The association between Mi-2 antibodies and dermatomyositis. *Arthritis Rheum* 1985;28:796-803.
- [40] Robinson AB, Reed AM. Clinical features, pathogenesis and treatment of juvenile and adult dermatomyositis. *Nat Rev Rheumatol* 2011;7:664-75.
- [41] Rider LG, Lachenbruch PA, Monroe JB *et al.* Damage extent and predictors in adult and juvenile dermatomyositis and polymyositis as determined with the myositis damage index. *Arthritis Rheum* 2009;60:3425-35.
- [42] Stone KB, Oddis CV, Fertig N *et al.* Anti-Jo-1 antibody levels correlate with disease activity in idiopathic inflammatory myopathy. *Arthritis Rheum* 2007;56:3125-31.
- [43] Casciola-Rosen L, Nagaraju K, Plotz P *et al.* Enhanced autoantigen expression in regenerating muscle cells in idiopathic inflammatory myopathy. *J Exp Med* 2005;201:591-601.
- [44] Mammen AL, Casciola-Rosen LA, Hall JC, Christopher-Stine L, Corse AM, Rosen A. Expression of the dermatomyositis autoantigen Mi-2 in regenerating muscle. *Arthritis Rheum* 2009;60:3784-93.
- [45] Tournadre A, Miossec P. A critical role for immature muscle precursors in myositis. *Nat Rev Rheumatol* 2013;9:438-42.
- [46] Howard OM, Dong HF, Yang D *et al.* Histidyl-tRNA synthetase and asparaginyl-tRNA synthetase, autoantigens in myositis, activate chemokine receptors on T lymphocytes and immature dendritic cells. *J Exp Med* 2002;196:781-91.
- [47] Figarella-Branger D, Civatte M, Bartoli C, Pellissier JF. Cytokines, chemokines, and cell adhesion molecules in inflammatory myopathies. *Muscle Nerve* 2003;28:659-82.
- [48] De Bleeker JL, De Paepe B, Vanwalleghem IE, Schroder JM. Differential expression of chemokines in inflammatory myopathies. *Neurology* 2002;58:1779-85.
- [49] Raju R, Vasconcelos O, Granger R, Dalakas MC. Expression of IFN-gamma-inducible chemokines in inclusion body myositis. *J Neuroimmunol* 2003;141:125-31.
- [50] Carstens PO, Schmidt J. Diagnosis, pathogenesis and treatment of myositis: recent advances. *Clin Exp Immunol* 2014;175:349-58.
- [51] Wiendl H, Hohlfeld R, Kieseier BC. Immunobiology of muscle: advances in understanding an immunological microenvironment. *Trends Immunol* 2005;26:373-80.
- [52] Rayavarapu S, Coley W, Nagaraju K. An update on pathogenic mechanisms of inflammatory myopathies. *Curr Opin Rheumatol* 2011;23:579-84.
- [53] Greenberg SA. DNA microarray gene expression analysis technology and its application to neurological disorders. *Neurology* 2001;57:755-61.
- [54] Raju R, Dalakas MC. Gene expression profile in the muscles of patients with inflammatory myopathies: effect of therapy with IVIg and biological validation of clinically relevant genes. *Brain* 2005;128:1887-96.
- [55] Tezak Z, Hoffman EP, Lutz J *et al.* Gene expression profiling in DQA1*0501+ children with untreated dermatomyositis: a novel model of pathogenesis. *J Immunol* 2002;168:4154-63.
- [56] Choi YC, Dalakas MC. Expression of matrix metalloproteinases in the muscle of patients with inflammatory myopathies. *Neurology* 2000;54:65-71.

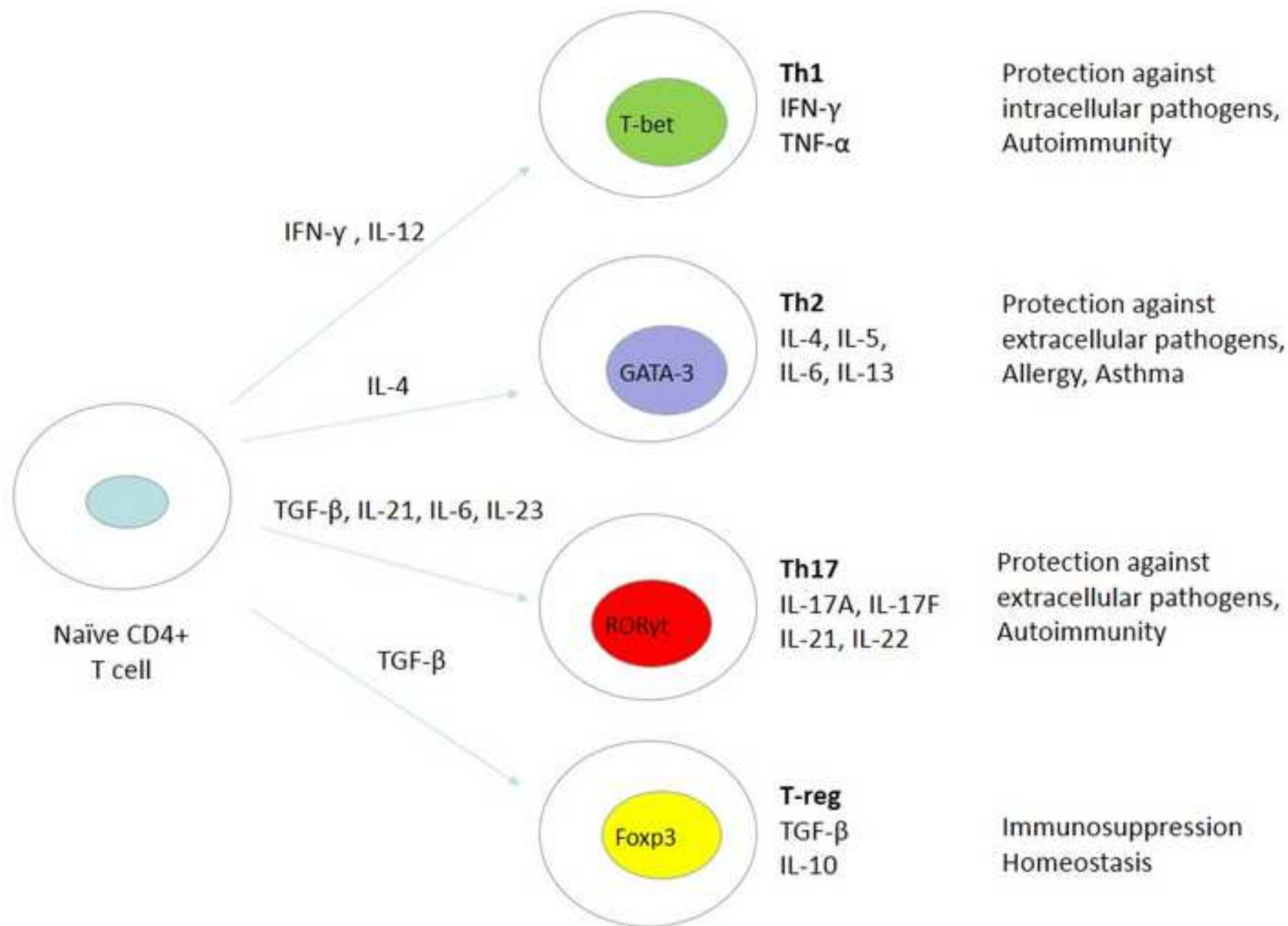
- [57] Walsh RJ, Kong SW, Yao *et al.* Type I interferon-inducible gene expression in blood is present and reflects disease activity in dermatomyositis and polymyositis. *Arthritis Rheum* 2007;56:3784-92.
- [58] Wenzel J, Scheler M, Bieber T, Tuting T. Evidence for a role of type I interferons in the pathogenesis of dermatomyositis. *Br J Dermatol* 2005;153:462-3; author reply 3-4.
- [59] Ivanidze J, Hoffmann R, Lochmuller H, Engel AG, Hohlfeld R, Dornmair K. Inclusion body myositis: laser microdissection reveals differential up-regulation of IFN-gamma signaling cascade in attacked versus nonattacked myofibers. *Am J Pathol* 2011;179:1347-59.
- [60] Baeten DL, Kuchroo VK. How Cytokine networks fuel inflammation: Interleukin-17 and a tale of two autoimmune diseases. *Nat Med* 2013;19:824-5.
- [61] Moran EM, Heydrich R, Ng *et al.* IL-17A expression is localised to both mononuclear and polymorphonuclear synovial cell infiltrates. *PLoS One* 2011;6:e24048.
- [62] Kenna TJ, Brown MA. The role of IL-17-secreting mast cells in inflammatory joint disease. *Nat Rev Rheumatol* 2013;9:375-9.
- [63] Miossec P. Interleukin-17 in fashion, at last: ten years after its description, its cellular source has been identified. *Arthritis Rheum* 2007;56:2111-5.
- [64] Miossec P. Interleukin-17 in rheumatoid arthritis: if T cells were to contribute to inflammation and destruction through synergy. *Arthritis Rheum* 2003;48:594-601.
- [65] Yao Z, Painter SL, Fanslow *et al.* Human IL-17: a novel cytokine derived from T cells. *J Immunol* 1995;155:5483-6.
- [66] Fossiez F, Djossou O, Chomarat *et al.* T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines. *J Exp Med* 1996;183:2593-603.
- [67] Miossec P, Kolls JK. Targeting IL-17 and TH17 cells in chronic inflammation. *Nat Rev Drug Discov* 2012;11:763-76.
- [68] Aarvak T, Chabaud M, Miossec P, Natvig JB. IL-17 is produced by some proinflammatory Th1/Th0 cells but not by Th2 cells. *J Immunol* 1999;162:1246-51.
- [69] Chabaud M, Aarvak T, Garnero P, Natvig JB, Miossec P. Potential contribution of IL-17-producing Th(1) cells to defective repair activity in joint inflammation: partial correction with Th(2)-promoting conditions. *Cytokine* 2001;13:113-8.
- [70] Infante-Duarte C, Horton HF, Byrne MC, Kamradt T. Microbial lipopeptides induce the production of IL-17 in Th cells. *J Immunol* 2000;165:6107-15.
- [71] Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. *J Biol Chem* 2003;278:1910-4.
- [72] Happel KI, Zheng M, Young *et al.* Cutting edge: roles of Toll-like receptor 4 and IL-23 in IL-17 expression in response to *Klebsiella pneumoniae* infection. *J Immunol* 2003;170:4432-6.
- [73] Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J Immunol* 1986;136:2348-57.
- [74] Weaver CT, Hatton RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. *Annu Rev Immunol* 2007;25:821-52.
- [75] Park H, Li Z, Yang *et al.* A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol* 2005;6:1133-41.
- [76] Harrington LE, Hatton RD, Mangan *et al.* Interleukin 17-producing CD4⁺ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 2005;6:1123-32.
- [77] Langrish CL, Chen Y, Blumenschein *et al.* IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med* 2005;201:233-40.
- [78] Menon B, Gullick NJ, Walter *et al.* IL-17+CD8⁺ T-cells are enriched in the joints of patients with psoriatic arthritis and correlate with disease activity and joint damage progression. *Arthritis Rheumatol* 2014;66:1272-81.

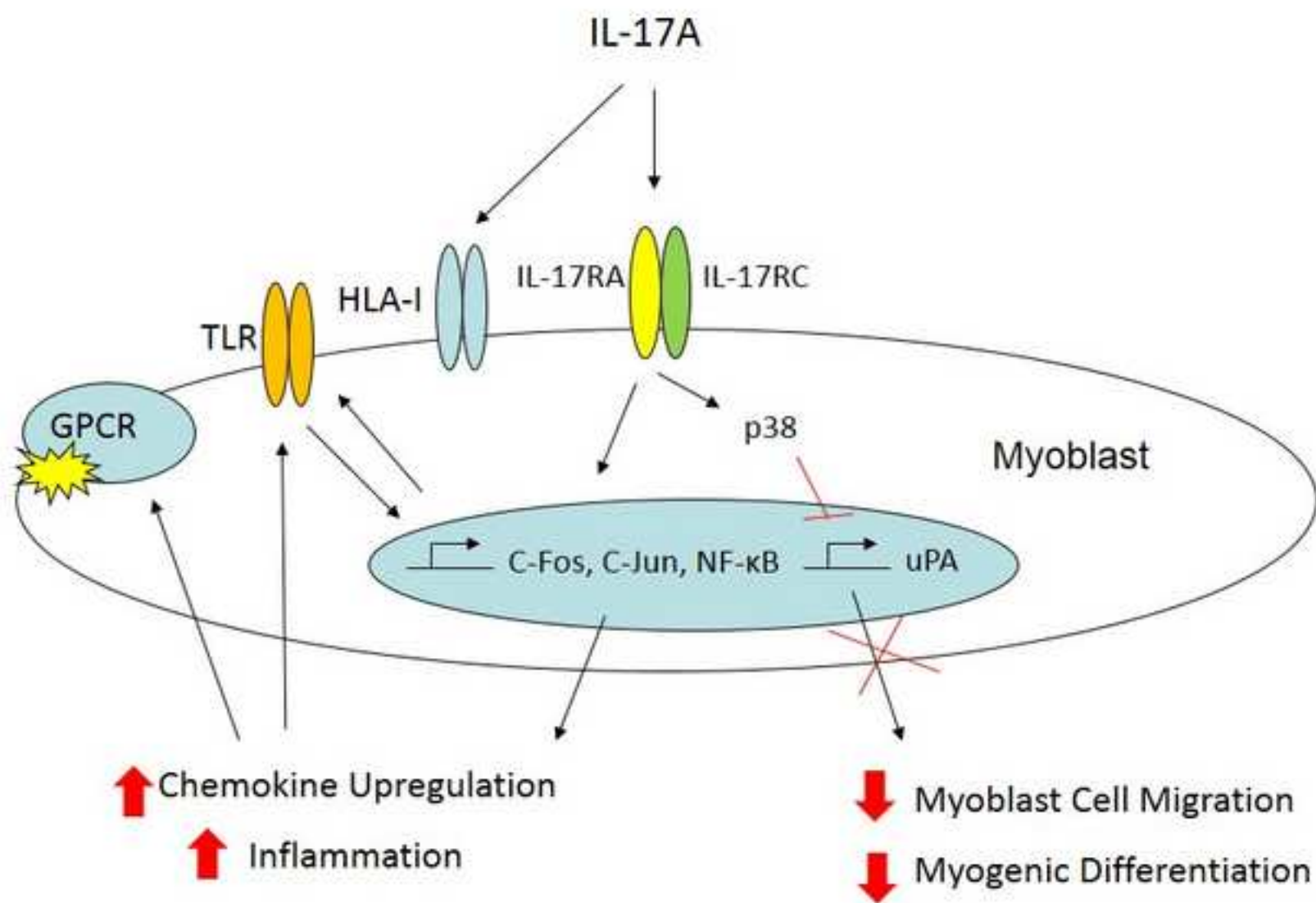
- [79] Huber M, Heink S, Grothe *et al.* A Th17-like developmental process leads to CD8(+) Tc17 cells with reduced cytotoxic activity. *Eur J Immunol* 2009;39:1716-25.
- [80] Skepner J, Ramesh R, Trocha *et al.* Pharmacologic Inhibition of ROR γ Regulates Th17 Signature Gene Expression and Suppresses Cutaneous Inflammation In Vivo. *J Immunol* 2014;192.
- [81] Abe M, Hiasa Y, Onji M. T helper 17 cells in autoimmune liver diseases. *Clin Dev Immunol* 2013;2013:607073.
- [82] Chen X, Oppenheim JJ. Th17 cells and Tregs: unlikely allies. *J Leukoc Biol* 2014;95:723-31.
- [83] Moran EM, Connolly M, Gao W, McCormick J, Fearon U, Veale DJ. Interleukin-17A induction of angiogenesis, cell migration, and cytoskeletal rearrangement. *Arthritis Rheum* 2011;63:3263-73.
- [84] Moran EM, Mullan R, McCormick *et al.* Human rheumatoid arthritis tissue production of IL-17A drives matrix and cartilage degradation: synergy with tumour necrosis factor-alpha, Oncostatin M and response to biologic therapies. *Arthritis Res Ther* 2009;11:R113.
- [85] Xu S, Cao X. Interleukin-17 and its expanding biological functions. *Cell Mol Immunol* 2010;7:164-74.
- [86] Mountz JD, Wang JH, Xie S, Hsu HC. Cytokine regulation of B-cell migratory behavior favors formation of germinal centers in autoimmune disease. *Discov Med* 2011;11:76-85.
- [87] Wang JH, New JS, Xie *et al.* Extension of the germinal center stage of B cell development promotes autoantibodies in BXD2 mice. *Arthritis Rheum* 2013;65:2703-12.
- [88] Xie S, Li J, Wang *et al.* IL-17 activates the canonical NF-kappaB signaling pathway in autoimmune B cells of BXD2 mice to upregulate the expression of regulators of G-protein signaling 16. *J Immunol* 2010;184:2289-96.
- [89] Doreau A, Belot A, Bastid *et al.* Interleukin 17 acts in synergy with B cell-activating factor to influence B cell biology and the pathophysiology of systemic lupus erythematosus. *Nat Immunol* 2009;10:778-85.
- [90] Lin Y, Ritchea S, Logar *et al.* Interleukin-17 is required for T helper 1 cell immunity and host resistance to the intracellular pathogen *Francisella tularensis*. *Immunity* 2009;31:799-810.
- [91] O'Connor W, Jr., Kamanaka M, Booth *et al.* A protective function for interleukin 17A in T cell-mediated intestinal inflammation. *Nat Immunol* 2009;10:603-9.
- [92] Chevrel G, Page G, Granet C, Streichenberger N, Varennes A, Miossec P. Interleukin-17 increases the effects of IL-1 beta on muscle cells: arguments for the role of T cells in the pathogenesis of myositis. *J Neuroimmunol* 2003;137:125-33.
- [93] Page G, Chevrel G, Miossec P. Anatomic localization of immature and mature dendritic cell subsets in dermatomyositis and polymyositis: Interaction with chemokines and Th1 cytokine-producing cells. *Arthritis Rheum* 2004;50:199-208.
- [94] Tournadre A, Porcherot M, Cherin P, Marie I, Hachulla E, Miossec P. Th1 and Th17 balance in inflammatory myopathies: interaction with dendritic cells and possible link with response to high-dose immunoglobulins. *Cytokine* 2009;46:297-301.
- [95] Kim GT, Cho ML, Park *et al.* Expression of TLR2, TLR4, and TLR9 in dermatomyositis and polymyositis. *Clin Rheumatol* 2010;29:273-9.
- [96] Brunn A, Zornbach K, Hans VH, Haupt WF, Deckert M. Toll-like receptors promote inflammation in idiopathic inflammatory myopathies. *J Neuropathol Exp Neurol* 2012;71:855-67.
- [97] Ciccia F, Rizzo A, Alessandro *et al.* Activated IL-22 pathway occurs in the muscle tissues of patients with polymyositis or dermatomyositis and is correlated with disease activity. *Rheumatology (Oxford)* 2014.
- [98] Higgs BW, Zhu W, Richman *et al.* Identification of activated cytokine pathways in the blood of systemic lupus erythematosus, myositis, rheumatoid arthritis, and scleroderma patients. *Int J Rheum Dis* 2012;15:25-35.

- [99] Szodoray P, Alex P, Knowlton Net *al*. Idiopathic inflammatory myopathies, signified by distinctive peripheral cytokines, chemokines and the TNF family members B-cell activating factor and a proliferation inducing ligand. *Rheumatology (Oxford)* 2010;49:1867-77.
- [100] Hatice Bilgic SRY, Kelly T. McNallan, Joseph C. Wilson, Thearith Koeuth¹, Jason W. Bauer, Kevin G. Moder, Shreyasee Amin, Clement J. Michel, Erik J. Peterson,, Emily C. Baechler, Ann M. Reed. Abstract # 2053 IL-17 and IFN-Regulated Genes and Chemokines as Biomarkers of Disease Activity in Inflammatory Myopathies. In *American College of Rheumatology (ACR)*. 2008. San Francisco.
- [101] Reed AM, CSC, MSHet *al*. Cell Depletion In Refractory Adult and Juvenile Dermatomyositis (DM) and Adult Polymyositis (PM) - The RIM (Rituximab in Myositis) Trial. *Arthritis & Rheumatism* 2013;65:S877-S8.
- [102] Lopez de Padilla C, Crowson CS, Hein MSet *al* (2013) *Interferon-Regulated Chemokine and Innate Cytokine Scores Identify Refractory Myositis Patients That Respond Better To Rtuximab Therapy*. *Arthritis & Rheumatism* 65, S887-S8 10.1002/art.38216: 10.1002/art.38216
- [103] Shen H, Xia L, Lu J, Xiao W. Interleukin-17 and interleukin-23 in patients with polymyositis and dermatomyositis. *Scand J Rheumatol* 2011;40:217-20.
- [104] Allenbach Y, Chaara W, Rosenzwaig Met *al*. Th1 response and systemic treg deficiency in inclusion body myositis. *PLoS One* 2014;9:e88788.
- [105] Tang X, Tian X, Zhang Yet *al*. Correlation between the frequency of Th17 cell and the expression of microRNA-206 in patients with dermatomyositis. *Clin Dev Immunol* 2013;2013:345347-54.
- [106] Chevrel G, Granet C, Miossec P. Contribution of tumour necrosis factor alpha and interleukin (IL) 1beta to IL6 production, NF-kappaB nuclear translocation, and class I MHC expression in muscle cells: in vitro regulation with specific cytokine inhibitors. *Ann Rheum Dis* 2005;64:1257-62.
- [107] Kocic J, Santibanez JF, Krstic Aet *al*. Interleukin 17 inhibits myogenic and promotes osteogenic differentiation of C2C12 myoblasts by activating ERK1,2. *Biochim Biophys Acta* 2012;1823:838-49.
- [108] Kocic J, Santibanez JF, Krstic A, Mojsilovic S, Ilic V, Bugarski D. Interleukin-17 modulates myoblast cell migration by inhibiting urokinase type plasminogen activator expression through p38 mitogen-activated protein kinase. *Int J Biochem Cell Biol* 2013;45:464-75.
- [109] Zong M, Dorph C, Dastmalchi Met *al*. Anakinra treatment in patients with refractory inflammatory myopathies and possible predictive response biomarkers: a mechanistic study with 12 months follow-up. *Ann Rheum Dis* 2013;73:913-20.
- [110] Reed AM, Crowson CS, Hein MSet *al* (2013) *Biomarker and Serologic Predictors Of Clinical Improvement After B Cell Depletion In Refractory Adult and Juvenile Dermatomyositis (DM) and Adult Polymyositis (PM) - The RIM (Rituximab in Myositis) Trial*. *Arthritis & Rheumatism* 65, S877-S8 10.1002/art.38216: 10.1002/art.38216
- [111] Amemiya K, Semino-Mora C, Granger RP, Dalakas MC. Downregulation of TGF-beta1 mRNA and protein in the muscles of patients with inflammatory myopathies after treatment with high-dose intravenous immunoglobulin. *Clin Immunol* 2000;94:99-104.
- [112] Basta M, Dalakas MC. High-dose intravenous immunoglobulin exerts its beneficial effect in patients with dermatomyositis by blocking endomysial deposition of activated complement fragments. *J Clin Invest* 1994;94:1729-35.
- [113] Dalakas MC. Controlled studies with high-dose intravenous immunoglobulin in the treatment of dermatomyositis, inclusion body myositis, and polymyositis. *Neurology* 1998;51:S37-45.
- [114] Dalakas MC, Illa I, Dambrosia JMet *al*. A controlled trial of high-dose intravenous immune globulin infusions as treatment for dermatomyositis. *N Engl J Med* 1993;329:1993-2000.

- [115] Waschbisch A, Schwab N, Ruck T, Stenner MP, Wiendl H. FOXP3+ T regulatory cells in idiopathic inflammatory myopathies. *J Neuroimmunol* 2010;225:137-42.
- [116] Antiga E, Kretz CC, Klembt *Ret al.* Characterization of regulatory T cells in patients with dermatomyositis. *Journal of Autoimmunity* 2010;35:342-50.
- [117] Allenbach Y, Rosenzweig M, Prevel *Net al.* P5.27 Evidence for the Implication of Th-1 and Treg cells but not Th-17 in sporadic Inclusion Body Myositis. *NMD* 2011;21:732.
- [118] Keijzer C, van der Zee R, van Eden W, Broere F. Treg inducing adjuvants for therapeutic vaccination against chronic inflammatory diseases. *Front Immunol* 2013;4:245.
- [119] Noack M, Miossec P. Th17 and regulatory T cell balance in autoimmune and inflammatory diseases. *Autoimmunity Reviews* 2014;13:668-77.
- [120] Venalis P, Lundberg IE. Immune mechanisms in polymyositis and dermatomyositis and potential targets for therapy. *Rheumatology (Oxford)* 2014;53:397-405.
- [121] Lundberg I, Ulfgren AK, Nyberg P, Andersson U, Klareskog L. Cytokine production in muscle tissue of patients with idiopathic inflammatory myopathies. *Arthritis Rheum* 1997;40:865-74.
- [122] Lepidi H, Frances V, Figarella-Branger D, Bartoli C, Machado-Baeta A, Pellissier JF. Local expression of cytokines in idiopathic inflammatory myopathies. *Neuropathol Appl Neurobiol* 1998;24:73-9.
- [123] Gabay C, Gay-Croisier F, Roux-Lombard *Pet al.* Elevated serum levels of interleukin-1 receptor antagonist in polymyositis/dermatomyositis. A biologic marker of disease activity with a possible role in the lack of acute-phase protein response. *Arthritis Rheum* 1994;37:1744-51.
- [124] Narazaki M, Hagihara K, Shima Y, Ogata A, Kishimoto T, Tanaka T. Therapeutic effect of tocilizumab on two patients with polymyositis. *Rheumatology* 2011;50:1344-6.
- [125] Ivanov, II, McKenzie BS, Zhou *Let al.* The orphan nuclear receptor ROR γ directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell* 2006;126:1121-33.
- [126] Huh JR, Littman DR. Small molecule inhibitors of ROR γ : targeting Th17 cells and other applications. *Eur J Immunol* 2012;42:2232-7.
- [127] Williams SC. Flurry of deal-making surrounds new autoimmunity target. *Nat Med* 2013;19:1078.
- [128] Tse MT. IL-17 antibodies gain momentum. *Nat Rev Drug Discov* 2013;12:815-6.

Figure 1





Highlights

- CD4⁺ Th1 cells or CD8⁺ cytotoxic T cells have been associated with myositis
- IL-17 and Th17 cells have been implicated in autoimmunity resulting in IL-17 targeted therapies
- We review the IL-17/Th17 axis in myositis and the therapeutic potential
- Detection of IL-17 in myositis and *in vitro* studies suggest a key role in myositis

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