



Review

The Function of Fish Cytokines

Jun Zou and Christopher J. Secombes *

Scottish Fish Immunology Research Centre, University of Aberdeen, Zoology Building, Tillydrone Avenue, Aberdeen AB24 2TZ, UK; j.j.zou@abdn.ac.uk

* Correspondence: c.secombes@abdn.ac.uk; Tel.: +44-1224-272872

Academic Editor: Brian Dixon

Received: 29 February 2016; Accepted: 17 May 2016; Published: 24 May 2016

Abstract: What is known about the biological activity of fish cytokines is reviewed. Most of the functional studies performed to date have been in teleost fish, and have focused on the induced effects of cytokine recombinant proteins, or have used loss- and gain-of-function experiments in zebrafish. Such studies begin to tell us about the role of these molecules in the regulation of fish immune responses and whether they are similar or divergent to the well-characterised functions of mammalian cytokines. This knowledge will aid our ability to determine and modulate the pathways leading to protective immunity, to improve fish health in aquaculture.

Keywords: cytokines; bioactivity; recombinant proteins; loss- and gain-of-function; interleukins; interferons; colony stimulating factors; tumour necrosis factors; transforming growth factor- β

1. Introduction

It is clear that all of the major cytokine families exist in fish, as outlined in many previous reviews that have described the genes present and when/where they are expressed at the transcript level [1–4]. In many cases, multiple copies (paralogues) of particular genes occur (particularly common in the teleost fish) and independent expansion of gene families in different lineages has taken place from putative common ancestral genes. For example, the classical pro-inflammatory cytokines IL-1 β , TNF- α and IL-6 are present, with multiple paralogues in most species. Similarly, cytokines associated with adaptive immunity are present, including IL-2, IFN- γ , IL-4/13 (with homology to IL-4 and IL-13), IL-10, IL-17A/F (with homology to IL-17A and IL-17F), IL-21, IL-22 and TGF- β 1 (amongst others), also with multiple paralogues. Hence it seems likely that many aspects of the regulation of immune responses in vertebrates are ancient and relatively well conserved, assuming conservation of function based on our understanding of the biological effects of primarily mammalian molecules. Since few antibodies exist to fish cytokines, there is virtually no data on protein expression, with a couple of exceptions (e.g., IFN and TNF- α). Thus most of the functional studies on fish cytokines have either investigated the bioactivities of cytokine recombinant proteins, produced using primarily prokaryotic (e.g., *Escherichia coli*) expression systems and on a few occasions in eukaryotic cell lines (fish and mammalian), or they have used loss- and gain-of-function experiments in model species, such as zebrafish. Such studies, primarily in teleost fish, begin to tell us about the role of these molecules in early vertebrates and whether they are similar or divergent between paralogues and to the well-characterised functions of mammalian cytokines. What is known about cytokine bioactivity in fish will be the focus of this review. The cytokines have been grouped by their molecular structure since this dictates the receptors used and the down-stream signalling pathways that are activated. However, the “open face β sandwich” cytokines (chemokines) are excluded since these are described elsewhere in this special issue [5]. It will become apparent that we now know a lot about the function of many fish cytokines, albeit in relatively few fish species at this point in time.

2. β -Trefoil Cytokines

The beta trefoil cytokine family, also termed the IL-1 family, consists of 11 members (IL-1F1-11) in mammals and are central players in regulating inflammation [6,7]. IL-1F1-4 are also referred to as IL-1 α , IL-1 β , IL-1 receptor antagonist (IL-1Ra) and IL-18, with the other members (IL-1F5-11) assigned as IL-33 (IL-1F11), IL-36 α (IL-1F6), IL-36 β (IL-1F8), IL-36 γ (IL1F9), IL-36 receptor antagonist (IL-36Ra, IL-1F5), IL-37 (IL-1F7) and IL-38 (IL-1F10). Functionally, the IL-1 family cytokines can be classified into two groups, either promoting (IL-1 α , IL-1 β , IL-18, IL-33 and IL-36 α , β , and γ) or suppressing (IL-1Ra, IL-36Ra, IL-37 and IL-38) inflammation.

IL-1 β was the first interleukin characterised in bony and cartilaginous fish [8–15]. Later, two other members of the IL-1 family were discovered in fish, namely IL-18 and a teleost specific group termed novel IL-1 family member (nIL-1Fm) [16–18]. Extensive analysis of fish genomes has confirmed that fish lack orthologues to other mammalian IL-1 family members [1]. Teleost fish IL-1 β genes exist as multiple copies that have recently been classified into two groups (type I and II) based on the genomic organisation and gene synteny [11,15,19–25]. Type I IL-1 β genes have a seven exon/six intron organisation, which is conserved among all the jawed vertebrates. In contrast, teleost type II IL-1 β genes appear to be a lineage specific group containing six or five exons and are believed to have arisen from a common ancestral gene that gave rise to the type I IL-1 β genes. However, the phylogenetical tree analysis revealed that the two teleost IL-1 β groups are closely related and form a separate clade relative to IL-1 β genes from other vertebrates. Not all fish species possess both groups, for example, type I but not type II IL-1 β s are found in cypriniformes [20]. A single copy gene of the IL-18 orthologue has been reported in a limited number of fish species but its immune functions have yet to be determined [16,18]. The nIL-1Fms are a teleost specific group of the IL-1 family and have a similar function to mammalian IL-1Ra in antagonising IL-1 β activities [17,22,24]. However, they are not 1:1 orthologues of mammalian IL-1R antagonists.

2.1. IL-1 β

IL-1 β is produced by a wide range of cell types after activation of host pattern recognition receptors (PRRs) by pathogen associated molecular patterns (PAMPs) or danger associated molecular patterns (DAMPs) [26–29]. The IL-1 β is synthesised as a precursor lacking a signal peptide and is required to be processed to release active protein [30]. The IL-1 β precursor can be cleaved either intracellularly or extracellularly [31]. In humans and mice, several proteases such as caspase 1, elastase and cathepsin G are known to recognise specific sequence motifs in the central region of the proIL-1 β protein, separating the mature peptide from the precursor. Caspase 1, also termed IL-1 β converting enzyme (ICE), is the primary protease for processing IL-1 β . However, fish IL-1 β orthologues lack an identifiable ICE cut site [13,14,20,22,25,32–35]. Nevertheless, emerging evidence indicates that teleost IL-1 β can be cleaved by caspases and processing of proIL-1 β into the active form could involve canonical and non-canonical activation of effector caspases, notably caspase 1 and 8 within the inflammasome [22,29,36,37]. Various cut sites within a short region in the middle of the proIL-1 β have been reported in different species [29,37,38]. Using a monoclonal antibody against IL-1 β , a protein of 15 kDa, smaller than the theoretical size of the predicted mature protein, was detected in carp leucocyte cultures activated by PHA [39]. In the rainbow trout monocyte/macrophage cell line RTS-11 transfected with an IL-1 β expression plasmid, an IL-1 β protein of approx. 24 kDa is secreted into the culture medium [40]. Interestingly, zebrafish IL-1 β can be cleaved into two peptides of 22- and 18-kDa by two distinct caspase 1 homologues after infection with a fish-specific bacterial pathogen *Francisella noatunensis*, indicating cleavage at multiple sites is possible and may depend on the immunological environment [29]. Direct evidence to support the involvement of caspase 1 came from a recent study showing that the seabass IL-1 β can indeed be cut by recombinant caspase 1, resulting in a mature peptide of 18.5 kDa. Curiously, the identified cut site for the caspase 1 is located within the predicted first beta sheet of the mature peptide [37], and so it is not certain whether the processed mature IL-1 β is biologically active. In contrast to these reports, cleavage of IL-1 β in seabream macrophages is not

affected by caspase-1 or pan-caspase inhibitors and is independent of caspase-1 [26]. In mammals, caspase 8 has been shown to directly cleave proIL-1 β whilst caspase 11 is required for IL-1 β proteolysis via activation of caspase 1 [41,42]. Some of these caspases have been characterised in fish and are shown to be important for regulation of the inflammatory response [38,43–45]. In addition, neutrophil elastase and some cathepsins may play a role in regulating maturation and release of active IL-1 β , as their human counterparts are shown to be capable of cleaving proIL-1 β [46,47]. Taken together, these observations suggest that the mechanisms for processing of fish IL-1 β are complex and are tightly regulated during inflammation.

IL-1 β has diverse physiological functions and its roles in regulating the inflammatory process are conserved in fish [1]. Although the native fish IL-1 β proteins have not been purified, recombinant proteins have been produced in bacteria and in general are biologically active [13,32,34,48–52]. To date, most *in vivo* studies have been focused on the transient and local effects of IL-1 β on the immune system. Intraperitoneal administration of IL-1 β into trout increases the number of phagocytes that migrate into the peritoneal cavity and the phagocytic and lysozyme activity of macrophages [48]. The local effect of IL-1 β is also apparent in the elevation of TNF- α and IL-1 β expression in muscle tissue injected with an IL-1 β encoding expression plasmid [52]. When administered directly into the intestine of grass carp, IL-1 β induces severe gut inflammation and expression of TNF- α [32]. Fish IL-1 β s also modulate expression of IL-17 family members, important for antibacterial defence [1,53,54]. In trout head kidney (HK) leucocytes, IL-1 β up-regulates IL-17C2 but not IL-17A/F [55,56]. IL-1 β has also been shown to enhance antibody production when administered with bacterial vaccines, suggesting it may be exploited as an immune-adjuvant for improving vaccine efficacy [13,52].

In a microarray analysis, Martin *et al.* [57] identified a panel of genes that respond to IL-1 β in RTS-11 cells. In other studies, IL-1 β action has been extensively analysed in primary leucocytes and macrophages where induced expression of pro-inflammatory genes is evident. Such genes include TNF- α [58], IL-1 β [20,34], IL-6 [59], IL-8 [59], IL-34 [60], and cyclooxygenase -2 (COX-2) [34,51]. The induced expression of IL-6 and COX-2 by IL-1 β can be inhibited by the stress hormone cortisol [59]. IL-1 β has also been shown to activate expression of genes which are suppressors of the immune response. For example, trout IL-1 β differentially modulates cytokine inducible Src homology 2 (SH2)-containing proteins (CISHs) and strongly enhances CISHa2 expression but interestingly not CISHa1 [61]. In addition, immune suppressive cytokines such as IL-10 and TGF- β 1b can be induced by IL-1 β in primary HK derived macrophages and RTS-11 cells [59,62]. The inducible effect of IL-1 β on TGF- β 1 appears to be mediated via the NF- κ B and MAPK signalling pathways [63].

IL-1 β is a chemoattractant for leucocytes in fish. Chemotaxis of leucocytes is coordinated by a sequential gradient of chemokines and the activation of G protein-coupled receptors. *In vitro* stimulation of freshly isolated trout leucocytes with IL-1 β protein or peptides leads to enhanced cell migration [64]. This may be associated with the rapid release of intracellular Ca⁺ ions [65], as well as the up-regulation of the transcript levels of chemokine receptors on target cells [66]. In addition to its direct impact on target cells, IL-1 β stimulation augments chemokine production in cells at infection sites. Several CXC chemokines, including CXCL8_L1 (IL-8), CXCL11_L1 (gamma IFN inducible protein, γ IP), and CXCL_F4 and _F5 are increased in RTG-2 and RTS-11 cells after IL-1 β stimulation [67].

In addition to its roles in immune regulation, IL-1 β is involved in regulating other physiological processes. Recent studies demonstrate that fish muscle metabolism is affected by IL-1 β . Trout primary muscle cells incubated with IL-1 β exhibit higher expression of both inflammatory genes and genes related to muscle growth and metabolism [68]. In addition, IL-1 β increases expression of atrogenin-1, a key ubiquitin E3 ligase controlling muscle mass, and insulin growth factor binding protein (IGFBP) -6 in salmon muscle cells [69], and induces dilation of isolated steelhead trout coronary microvessels [70].

The IL-1 β functions are controlled at different levels. Interaction of IL-1 β with its heterodimeric receptor is pivotal to downstream signalling and dictates the outcome of the cellular responses. The IL-1 β receptor complex comprises a ligand specific chain (IL-1R1 or IL-1R2) and a common chain (IL-1R accessory protein, IL-1RAcP) shared by different members of the IL-1 family [71].

All the receptors have a similar secondary structure containing 1–3 immunoglobulin (Ig) like domains in the extracellular region, which engage with the ligand (Figure 1). The IL-1R1 serves as an agonistic receptor upon activation by the ligand whilst the IL-1R2 acts as a decoy receptor to block ligand actions. The IL-1R2 has a short intracellular cytoplasmic region lacking the Toll/IL-1 receptor (TIR) homology domain and hence is unable to interact with adaptor proteins for signal transduction. Soluble receptors exist as negative regulators for blocking IL-1 β signalling.

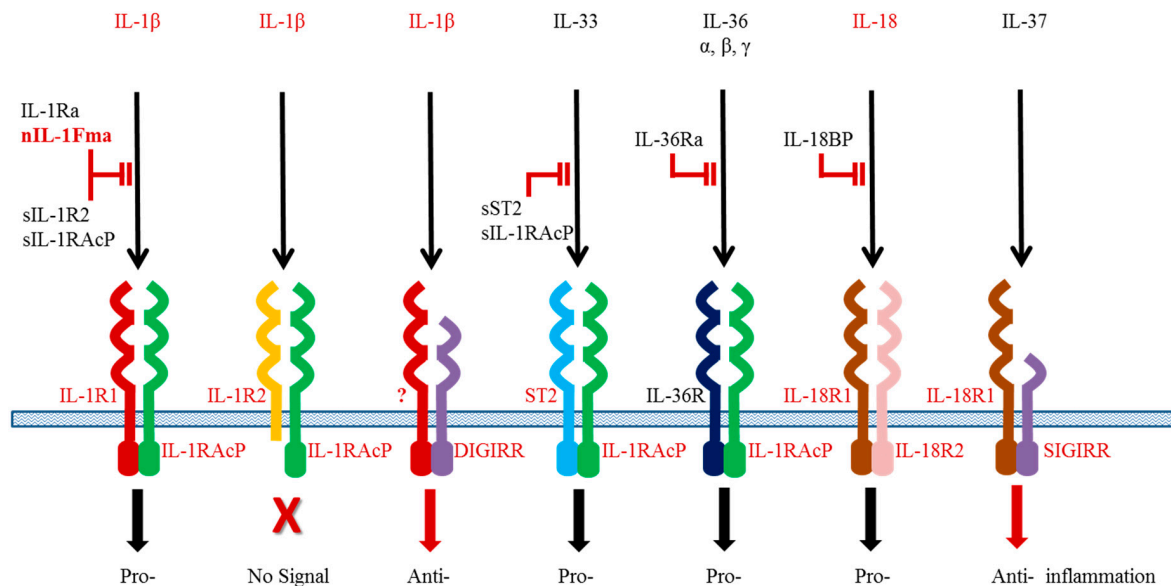


Figure 1. Interaction of the IL-1 family cytokines and receptors. The homologues identified in fish are indicated in red.

The first fish IL-1R homologue was identified as an IL-1R related protein in Atlantic salmon [72], and later confirmed as the orthologue of IL-1R1 [73]. It has a typical arrangement of three conserved Ig domains in the extracellular region. Like its mammalian orthologues, salmon IL-1R1 forms a heterodimer with its co-receptor IL-1RAcP. A soluble form of IL-1R1 has recently been identified in orange-spotted grouper [74]. It can be induced by high concentrations of IL-1 β and is suggested to be a potential negative regulator to dampen excessive IL-1 β effects.

Teleost IL-1R2 has a well-conserved protein structure as a decoy receptor for antagonising IL-1 β functions, and contains three Ig domains in the extracellular region and a short cytoplasmic region lacking a TIR domain [75–77]. One exception is the report of a two Ig domain-containing IL-1R2 in seabream, which is probably due to a prediction error of the software used [75]. The seabream IL-1R2 is expressed at the cell surface of transfected human HEK293 cells, and has been shown to bind to IL-1 β [75]. Direct interaction between IL-1 β and soluble IL-1R2 has also been confirmed using recombinant proteins produced in bacteria [78]. As expected, the soluble IL-1R2 protein inhibits the effect of IL-1 β to induce expression of inflammatory genes in grass carp primary HK cells [78]. At the transcript level, the IL-1R2 is expressed in all tissues of healthy fish and LPS stimulation or bacterial infection (*Vibrio anguillarum* and *Aeromonas hydrophila*) results in a significant increase of gene expression [75,78,79]. Interestingly, IL-1R2 expression is not altered in amoebic gill disease affected fish or down-regulated in fish with parasitic infections (*Ichthyophthirius multifiliis*) [75,76,80].

A novel double Ig IL-1R related molecule (DIGIRR) (Figure 1) has been added to the teleost IL-1R family [81]. It has a close phylogenetic relationship with the mammalian Single Domain IL-1R related (SIGIRR) molecule and may act as a negative regulator for inflammatory responses in fish. The anti-inflammatory activities of DIGIRR have been investigated in zebrafish where DIGIRR is able to abrogate LPS-induced and IL-1 β -induced NF- κ B activation when injected into embryos.

When the DIGIRR expression is suppressed in fish by small interfering RNA molecules, expression of pro-inflammatory cytokines is elevated.

In addition to the receptors, the functions of IL-1 family cytokines are known to be controlled by multiple intracellular proteins. Upon activation of the receptors by the ligands, Myeloid differentiation factor 88 (MyD88) and Toll interacting protein (Tollip) are recruited to the IL-1R1/IL-1RAcP complex, resulting in phosphorylation of the IL-1R-associated kinases (IRAKs). The phosphorylated IRAKs interact with the TNF receptor associated factor (TRAF) 6 and NF- κ B is then activated, leading to expression of pro-inflammatory genes in the target cells. To date, all these signalling molecules have been found in fish, suggesting that the IL-1 signalling pathway is well conserved among vertebrates.

2.2. IL-18

IL-18 is an important cytokine found initially to induce IFN- γ production and promote Th1 immunity in vertebrates [82,83]. As a member of the IL-1 family, it also plays an important role in regulating inflammation in mucosal tissues such as skin and gut. Akin to IL-1 β , IL-18 is constitutively synthesised as an inactive precursor in healthy animals and is released after cleavage by caspase 1 following stimulation with PAMPs or DAMPs. Mature IL-18 binds to a heterodimeric receptor consisting of IL-18R1 and IL-18R2 (Figure 1). A soluble protein, termed IL-18 binding protein, is known to block binding of IL-18 to the receptors.

Compared with IL-1 β , fish IL-18 has been under-investigated. To date, IL-18 has been reported only in a few fish species including rainbow trout [18], pufferfish [16], seabream [84], turbot [85] and elephant shark [86]. Fish IL-18 is encoded by a single gene comprising six exons and five introns, a conserved organisation seen for all known IL-18 genes [16,18]. Unlike IL-1 β , which is temporally induced by inflammatory stimuli, IL-18 is constitutively expressed in fish immune or non-immune tissues [18]. Interestingly, IL-18 expression is not modulated by LPS, poly(I:C) or IL-1 β in freshly isolated trout leucocytes [18]. However, a shorter variant of the alternatively spliced transcript (trout IL-18B) is upregulated in RTG-2 cells (a trout fibroblast cell line) after stimulation with LPS or poly(I:C) but not IL-1 β . This work suggests that proteolytic processing of the inactive precursor may be crucial in mediating fish IL-18 functions which have yet to be characterised.

A partial IL-18 receptor (IL-18R1) has been sequenced in zebrafish [87]. A conserved TIR domain is predicted in the sequence. Constitutive expression is detected in healthy fish and is not affected by intraperitoneal injection with *Mycobacterium marinum* (*M. marinum*). However, an increase of IL-18R1 transcripts is observed in the intestine of seabream infected with *Enteromyxum leei*, a myxozoan parasite, suggesting that IL-18 may play a role in mucosal immunity [84]. The IL-18R2 has not been identified in fish.

2.3. Fish Specific Novel IL-1 Family Member

A novel IL-1 family member (nIL-1Fm) has recently been discovered and studied in a few fish species [20,88]. This gene(s) appears to be teleost specific and has ambiguous phylogenetic relationships with known IL-1 family members. Like most of the IL-1 family members, it lacks a signal peptide and has a pre-domain [17,20,22]. Sequence analysis indicates that conserved cut sites for caspase 1 and thrombin are present in the trout nIL-1Fm, suggesting proteolytic processing is required to generate the active protein. A recombinant trout nIL-1F protein comprising the last 169 aa of the C-terminal region has been produced in bacterial cells and is unable to trigger expression of inflammatory genes such as IL-1 β , TNF- α , COX-2 and CXCL8_L1/IL-8 but inhibits the IL-1 β -induced gene expression in RTS-11 cells, suggesting it acts as an antagonist of IL-1 β [17]. The antagonistic effect of nIL-1Fm is likely mediated by competition with IL-1 β for receptor binding, as shown in grass carp [88].

3. B-Jellyroll Cytokines

The jellyroll cytokine family, also referred to as the tumour necrosis factor superfamily, comprises 19 ligands (TNFSF) and 29 receptors (TNFSFR) in humans [89]. The three major members of

this cytokine family are TNF- α , lymphotoxin (LT) - α (also called TNF- β) and LT- β . The TNFSF members are usually type II membrane proteins that contain a short N-terminal intracellular domain, a transmembrane domain and a C-terminal extracellular TNF homology domain with a conserved TNF family signature motif. An exception is LT- α that has a signal peptide and can be secreted conventionally. However, following enzymatic cleavage many of the mature peptides of the TNFSF can be released from the cell surface as soluble forms. Both TNF- α and LT- α are released as homotrimers that interact with the receptors on target cells. In addition, LT- α is required for the transport of LT- β to the cell surface to form a membrane bound LT- α /LT- β complex [90], and increasing evidence suggests that membrane bound ligands play a critical role in regulating the inflammatory response [91]. A secreted LT- α /LT- β heterodimer has also been described [92].

TNF is an ancient cytokine and functional homologues can be traced back to invertebrates [4]. In bony fish, TNF genes have been reported in a number of species [93–105], and their phylogeny has been studied extensively [106,107]. Evidence gathered to date demonstrates that multiple TNF homologues/paralogues exist, which can be categorised into three phylogenetic groups, the type I and II TNF- α group and the TNF-N group [58,108,109]. The genes encoding type I TNF- α and TNF-N are tandemly linked in a single chromosomal locus across the teleosts, exhibiting a conserved synteny with that in humans. In contrast, the type II TNF- α genes appear to be teleost specific [58,108]. Within the subfamily, multiple copies of type I or II TNF- α genes have been found in some species such as Atlantic salmon, rainbow trout, goldfish and common carp [58,97,104,110–113]. The recombinant proteins of fish TNF- α paralogues display similar bioactivities, indicating redundant functions in immune regulation. However, they are differentially modulated in different cell types and the kinetics vary, suggesting they do play different roles in the immune response [58,104,111,114]. For example, trout TNF- α 3, a fish type II TNF- α , is implicated to be the major TNF- α isoform involved in T cell mediated immunity in addition to its roles in innate immunity [58,114]. In contrast to TNF- α , TNF-N is encoded by a single copy gene [58,100] and its functions have yet to be characterised.

Release of soluble TNF- α requires the removal of the precursor region, including the N-terminal intracellular domain and transmembrane region [91]. This process is facilitated by the TNF- α -converting enzyme (TACE, also referred to as ADAM17) which cleaves the proTNF- α at a specific site. Unlike fish IL-1 β s where the ICE cut site region differs significantly, a predicted cut site for TACE is present in all fish TNF- α sequences, suggesting a conserved mechanism for TNF- α processing and release in all vertebrates [58,96,97,105]. Recently, TACE activity has been described in rainbow trout macrophages after stimulation with LPS, where it promotes the processing of proTNF- α [115]. The TACE/ADAM17 gene has also been confirmed in ayu, supporting the involvement of TACE/ADAM17 in release of TNF- α in fish [116].

As a pro-inflammatory cytokine, TNF- α is one of the early immune genes expressed at an early stage of infection in fish and has a key role in regulating inflammation. Like its mammalian counterparts, fish TNF- α displays overlapping functions with IL-1 β . Many fish TNF- α s have been produced in bacteria as monomers, dimers and trimers and are able to activate macrophages/phagocytes and enhance their microbial killing activity [103,117,118]. *In vitro* treatment of primary trout HK leucocytes and monocytes/macrophages with TNF- α triggers expression of a number of immune genes associated with inflammation, including IL-1 β , IL-8, IL-17C, TNF- α and COX-2, and genes involved in antimicrobial responses [58,103,105,119]. The stimulatory effect of TNF- α involves the NF- κ B signalling pathway [58,103,119,120]. For example, incubation of grass carp leucocytes and EPC cells with TNF- α results in a rapid increase of NF- κ B activity in the treated cells [103,120].

The TNF- α protein enhances the phagocytic activity of fish leucocytes [105,110,117]. In *M. marinum*-infected zebrafish, TNF- α has been shown to promote macrophage survival and also restrict bacterial growth in infected macrophages [121]. The bacterial-killing activity appears to be accompanied by increased generation of reactive oxidative species (ROS) [116,122]. However, in the case of zebrafish infected with *M. marinum*, excessive production of TNF- α -activated ROS is

detrimental to the host cells, leading to programmed necrosis and increased release of mycobacteria from the infected macrophages [122].

Fish TNF- α s are suggested to be involved in the regulation of leucocyte homing, proliferation and migration. In trout thymus, constitutive expression of TNF- α is detected, suggesting it could play a role in promoting thymocyte growth [123]. Trout TNF- α s also increase migration of enriched primary HK macrophages in a dose-dependent manner [105,117], and an *in vivo* experiment shows that intraperitoneal injection of seabass with TNF- α results in rapid recruitment of phagocytic granulocytes to the peritoneal cavity [117]. In addition, TNF- α activates chemokine expression in local tissue cells such as endothelial cells in zebrafish and gilthead seabream [124]. Another study shows that the supernatants from TNF- α -treated endothelial cells can promote leucocyte migration and respiratory burst activity in carp [125]. In the zebrafish model, a cellular adaptor protein termed factor associated with neutral sphingomyelinase activity (FAN) has been shown to co-ordinate TNF- α activated chemotaxis in response to infection and wounds [126].

Fish TNF- α s exert pro-apoptotic activity, as seen with their mammalian homologues. High doses (400–4000 ng/mL) of Chinese perch TNF- α induce apoptosis of human Hela cells [102]. In tilapia TNF- α has been shown to upregulate granzyme expression in non-specific cytotoxic cells and provides protection of these cells from activation-induced cell death [127]. These results suggest that TNF- α triggered apoptosis is likely to be dose and target cell dependent.

In addition to its roles in acute infection, accumulating evidence suggests that TNF- α is associated with pathogenesis of several chronic diseases in fish. TNF- α , together with other pro-inflammatory genes, is induced in the heart during pathology of salmonid pancreas disease [128]. In turbot, during infection with *Enteromyxum scophthalmi*, the increased number of the TNF- α + cells leads to the infiltration of inflammatory cells in the intestine and is associated with the development of the lesions, epithelial shedding and intestinal barrier dysfunction [129]. Recently, Marjoram *et al.* [130] studied TNF- α production in the gut during infection with *M. marinum* using transgenic TNF/GFP reporter fish, and found strong up-regulation of TNF- α in granulomas surrounding the bacteria. The unregulated induction of TNF- α expression in intestinal epithelial cells (IECs) exacerbates gut inflammation, resulting in IEC shedding and apoptosis, immune cell recruitment, and barrier dysfunction of the gut in zebrafish [130]. Lastly, TNF- α is required for oocyte maturation in fish [131,132], and is essential for liver development in zebrafish [133], where knockdown leads to a reduction of liver size [133].

4. Cysteine Knot Cytokines

The cysteine knot cytokines get their name from a disulphide-rich, all-beta core domain protein secondary structure. They consist of two families of particular importance for immunity, namely the IL-17 family cytokines and the transforming growth factor- β (TGF- β) family cytokines. However, IL-17 family members contain only two of the three characteristic disulphide linkages that give the family its name [134]. They are ancient cytokines with TGF- β superfamily members and IL-17 homologues present in invertebrates. In addition, in invertebrates Spätzle, an endogenous Toll receptor ligand [135], has a cysteine knot structure and is critical for the initiation of innate immune responses in molluscs and arthropods.

4.1. Interleukin-17

Six IL-17 family members exist in mammals, IL-17A to IL-17F. However, in fish, only two clear homologues are known, IL-17B and IL-17D, with multiple isoforms of molecules termed IL-17A/F (IL-17A/F1-3, and IL-17N) with relatedness to both IL-17A and IL-17F, and IL-17C with relatedness to IL-17C and IL-17E (IL-25) being present [4,56,86,136]. Relatively few studies have reported the bioactivity of any of these IL-17 molecules in fish. In grass carp rIL-17A/F1 can increase IL-1 β , IL-6, IL-8 and TNF- α expression in HK leucocytes [137], and in trout rIL-17A/F2a increases the expression of IL-6, IL-8 and an antimicrobial peptide (β -defensin 3) in splenocytes [138]. In the case of grass carp rIL-17D, it can also increase IL-1 β , IL-8 and TNF- α expression in HK cells but does not

increase IL-6 expression [139], hinting at subtle differences in the ability of different isoforms to induce pro-inflammatory genes.

In relation to the possible sources of the IL-17 isoforms in fish, such as potential Th17 cells or group 3 innate lymphoid cells (ILC), it has been shown that enhanced IL-17A/F2 expression is seen in CD4-1+ lymphocytes isolated following specific antigen-restimulation of zebrafish three days earlier [140]. Th17 cell development is inhibited by Treg cells that are driven to differentiate by the transcription factor FoxP3, and also in zebrafish it has been shown that injection of zebrafish eggs with a plasmid encoding FoxP3 results in inhibition of IL-17 (isoform not given) expression in six-day-old embryos [141]. In contrast injection of eggs with morpholinos to inhibit FoxP3 translation results in up-regulation of IL-17 in five-day-old embryos.

4.2. Transforming Growth Factor- β

TGF- β also has multiple isoforms, TGF- β 1 to TGF- β 3, and there appear to be true homologues of these genes throughout vertebrates (note: Chicken TGF- β 4 = TGF- β 1; *Xenopus* TGF- β 5 = TGF- β 1). However, a fish specific TGF- β isoform (TGF- β 6) has been reported [142] although the role of this molecule remains to be determined. In the context of immune responses, most studies have focused on TGF- β 1 function as a molecule known to drive Th17 differentiation in combination with other cytokines (eg IL-6) but which is also a key immunosuppressive cytokine secreted by Treg cells. TGF- β 1 has been produced as a recombinant protein in several fish species, and shown to have an immunosuppressive function. For example, in goldfish rTGF- β 1 can inhibit the nitric oxide response of TNF- α activated macrophages [143], and in grass carp rTGF- β 1 inhibits the LPS induced up-regulation of TNF- α , IL-1 β , IL-8 and inducible nitric oxide synthase (iNOS) in monocytes/macrophages [144]. In the latter case incubation in the presence of an antibody to TGF- β 1 increases these transcript levels. Further studies have shown that grass carp rTGF- β 1 is able to inhibit the up-regulation of IL-1 β (transcript and protein) by rIL-1 β in HK leucocytes but has no effect on constitutive IL-1 β levels [63]. In addition, grass carp rTGF- β 1 decreases the basal level and IL-1 β induced up-regulation of the IL-1 agonistic receptor chains (IL-1R1 and IL-1RAcP) but enhances the expression of IL-1R2, the decoy receptor (Figure 1). These effects appear to be mediated by impaired I κ B phosphorylation but increased phosphorylation of Smad2. Co-incubation with the TGF- β RI kinase inhibitor VIII (ALK5 inhibitor) has also been shown to ablate rTGF- β 1 action in grass carp, indicating that TGF- β 1 has a self-regulatory function in fish [145].

rTGF- β 1 can also enhance the proliferation and viability of peripheral blood leucocytes (PBL) [145, 146] and the proliferation of fibroblasts [143] but reduces HK cell viability and blocks the proliferation of PBL induced by PHA or LPS [145,146]. It also suppresses erythropoiesis [147]. Indeed, there is an increased level of (active) TGF- β 1 protein and up-regulation of TGF- β signalling in the Antarctic notothenioid fish that lack hemoglobin/erythrocytes, compared to red-blooded notothenioids. TGF- β 1 protein has also been detected in tilapia, where it was reported to increase from 12 h to three days in the HK, and from one to three days in spleen post-injection with *Streptococcus agalactiae* [148].

5. Type I α Helical Cytokines

The type I helical cytokines include the bulk of the interleukins, and the colony stimulating factors. The former can be subdivided into the IL-2 subfamily, the IL-6 subfamily and the IL-12 subfamily, with the IL-2 subfamily considered short type 1 cytokines and the IL-6/12 subfamilies long chain type I cytokines. They signal via receptors of the type I cytokine receptor family [149]. Representatives of all these cytokine families are present in fish, although rather few of the proteins have had their bioactivity analysed.

5.1. IL-2 Subfamily

The IL-2 subfamily includes IL-2, IL-4/13, IL-7, IL-15 and IL-21 in fish. The IL-4/13 molecules have relatedness to both IL-4 and IL-13, and in teleost fish have diverged into IL-4/13A and IL-4/13B due to the duplication of the locus from the whole genome duplication (WGD) event at the base

of this fish lineage [150,151]. IL-2 has been made as a recombinant protein in rainbow trout, where it has been shown to increase the expression of STAT5 and Blimp-1, transcription factors involved in IL-2 signalling, IFN- γ , the chemokine γ IP, and itself in HK leucocytes [152]. Later, it was shown that rIL-2 can also up-regulate the expression of the fish type-2 cytokines (see below) IL-4/13B1 and IL-4/13B2 (in HK cells) but had no effect on IL-4/13A [153]. In addition, a small increase of IL-17A/F1b and IL-17A/F2a expression was seen after stimulation of HK cells [56]. A stable trout cell line has been made that over-expresses IL-2, and the conditioned media (CM) harvested for establishing IL-2 dependent cell lines [154]. When HK cells were incubated with these CM for 3–5 weeks a major shift to a dominance of CD4-1 transcripts (*vs.* CD8, IgMH and MCSFR) was seen in the cultures. However, if the cells were first treated with the T cell mitogen PHA, the CD8 transcripts were more highly represented (26%–51%) in addition to CD4-1 and MCSFR at these times.

In the case of IL-4/13 bioactivity, initial studies looked at the *in vivo* effects of injecting the protein into fish, and found that in zebrafish IL-4/13A could increase the number of CD209+ (DC-SIGN) cells in PBL five days post-injection [155], the number of IgZ-2+ B cells in PBL at two days post-injection [156], and the number of IgM+ B cells in PBL at three days post-injection [157]. rIL-4 administration also increased the transcript expression of mIgM, MHC II and CD80 in these cells at the latter timing. In addition, injection of rIL-4 ($\times 3$ at 12 h intervals) prior to immunisation gave higher specific serum antibody titres compared to fish given antigen (KLH) alone. The zebrafish recombinant IL-4R has also been shown to bind IL-4/13A in a pull-down assay, and *in vivo* administration of the rIL-4R soluble form or an anti-IL-4R α Ab (for receptor blockade) ablates the *in vivo* effects of rIL-4 described above [157]. Most recently the activity of rIL-4/13B has been studied. In carp, rIL-4/13B stimulates the *in vitro* proliferation of IgM+ B cells, in terms of colony formation and increased cell number [158]. In rainbow trout rIL-4/13B (IL-4/13B2, one of the two paralogues of this molecule present in trout) was studied alongside rIL-4/13A, and these cytokines were found to have shared but also distinct activities in terms of transcriptional modulation of other immune genes [153]. In addition, both cytokines increased the number of IgM secreting cells (likely by inducing the transcript for the secretory form of IgM rather than the membrane form) but had no effect on their proliferation, in contrast to the situation in carp. Taking into account the different expression patterns of these two genes in trout, it was proposed that IL-4/13A may be important for basal type-2 immunity, whilst IL-4/13B expression may aid adaptive responses post-infection/immunisation in this species.

Whilst IL-7 is known in fish [159] there have been no functional studies using the recombinant protein. However, in zebrafish it has been shown that fish with a mutation in the IL-7R, have impaired thymopoiesis [160]. Both *ikaros*-expressing and *rag1*-expressing cells are reduced in the thymus of such fish. The fish tend to die early but those that survive have a smaller and hypocellular thymus relative to that of wild type fish. However, the HK is unaffected, with numbers of IgMH chain expressing (B) cells being normal. The mutation arises from an insertion of an extra thymidine residue in exon 4 that induces a frameshift that impacts on the extracellular domain and the truncated protein, if produced, is predicted to be unable to interact with IL-7. These results support the paradigm that IL-7 signalling is required for T cell development throughout the vertebrates.

IL-15 has been made as a recombinant protein in fish, in rainbow trout, and its bioactivity studied [161]. Initially it was shown to increase IFN- γ expression in primary cultures of splenocytes but had no effect on HK cells or the RTS-11 (macrophage) cell line. Later it was also shown to increase expression of two of the IL-12 family peptides (see below), p35 (two fold) and p40c (six fold), in HK cells but not p40b expression, the second p40 paralogue present in trout [162]. Studies in mammals suggest that IL-15 is pleiotropic with effects on many cell types. Thus, it remains to be determined if these relatively restricted responses are found in other fish species. A second IL-15 related gene that has also been identified in fish is located at a different locus to the IL-15 gene [163,164], and has been termed IL-15-like (IL-15L). It was subsequently found to be present in other vertebrate groups but is a pseudogene in man and mouse [165]. Functional studies have still to be performed with this molecule, which in mammals binds to the IL-15R α but not the IL-2R α , as seen with IL-15 itself [165].

In contrast to IL-15, rainbow trout rIL-21 has been made and shown to have a wide range of effects [166]. When added to HK cell cultures it was able to maintain the expression level of several T and B cell markers, suggesting it is likely an important lymphocyte survival factor. It also up-regulated the expression of several key cytokines and immune regulators, including IL-6, IL-10, IL-22, IFN- γ , γ IP, STAT3, STAT5, Suppressors of Cytokine Signalling (SOCS) 1, SOCS3 and itself. In addition, *in vivo* administration (intraperitoneal injection) of rIL-21 induced the expression of IL-10, IL-21 and IFN- γ in peritoneal adipose tissue, and IL-10 and IL-22 in the HK at Day 1 post-injection, and IgMH and CD8 in the HK at Day 3. Subsequent studies have shown that rIL-21 can also increase p40c expression (but not p35 or p40b) [162], IL-17A/F1a/b and IL-17A/F3 expression [56], and IL-4/13A (in contrast to IL-2)/IL-4/13B (albeit relatively late compared to IL-2) expression in HK cells [153]. The rIL-21 induced up-regulation of IFN- γ in HK cells was ablated by pre-treatment with Stattic, a STAT3-specific inhibitor, and was partially reduced by pre-treatment with Jak inhibitors [166]. Such findings suggest that these IL-21 induced effects are mediated via a Jak/STAT3 pathway.

These cytokines signal via receptors that contain the common gamma chain (γ C) CD132 as a shared subunit with at least one private chain. In some fish species two γ C genes are known, as in zebrafish where they are termed γ C.a and γ C.b. Gene knock-down of γ C.a with anti-sense morpholinos results in significantly lower embryonic lymphopoiesis and loss of mature T cells in the larvae, whilst mature B cells could still be detected similar to the situation with SCID mice [167]. This was not seen with knock-down of γ C.b, and its function remains to be determined. Receptor chains similar to the known private chains are present in fish [168] with the apparent exception of the IL-2R α . It has been suggested that in fish IL-2 and IL-15 may both bind to the IL-15R α (also termed CD25-like in some papers), although a 5-fold preference for IL-15 was seen in binding assays with the Tetraodon molecules [169]. Currently it is unknown whether IL-2R α evolved after the divergence of bony fish from the tetrapod lineage or whether it has been secondarily lost in these fish [165].

5.2. Beta Chain Cytokines

The beta chain (β c) cytokines include IL-3, IL-5 and GM-CSF, and signal via a receptor composed of the beta-chain (or CD131) and a cytokine specific private chain. The beta chain has been known in fish for some time [170] but until recently no obvious ligands had been detected. However, with the release of the elephant shark genome Dijkstra [171] reported the presence of two possible “IL-5 family” genes, and has subsequently found further possible IL-3/IL-5/GM-CSF related genes in spotted gar and in several cyprinid fish species [172]. The true nature of these genes remains to be determined and to date there are no functional studies.

5.3. IL-6 Subfamily

The IL-6 subfamily of cytokines are known to be major players in haematopoiesis, and have pro- and anti-inflammatory properties. In fish four members of the IL-6 family exist; IL-6 itself, IL-11, a molecule termed CNTF-like and M17 [173,174]. Phylogenetic analysis suggests that the fish CNTF-like and M17 genes may have arisen from ancestral genes that gave rise to mammalian CNTF/CLC/CT-1/CT-2 and LIF/OSM, respectively. IL-31, a further member of the IL-6 cytokine subfamily in mammals, is unknown in fish to date. As with other fish cytokines, the fish IL-6 subfamily genes can have multiple paralogues, as seen with IL-11 [175,176].

No bioactivity studies have been reported for fish IL-11 or CNTF-like to date. However, IL-6 and M17 have been made as recombinant proteins. The rIL-6 has been shown to have effects on macrophages and B cells. In rainbow trout rIL-6 promoted macrophage growth in culture, with numbers of (RTS-11) cells being significantly increased after 4–7 days of treatment [177]. In addition the trout rIL-6 induced transient expression (up to 4 h post-stimulation) of SOCS1-3 and IFN regulatory factor (IRF)-1 but more sustained up-regulation of antimicrobial peptide (AMP) gene expression in these cells, with similar results seen with primary (HK derived) macrophage cultures. Interestingly, whilst the AMP cathelicidin-2 was sensitive to rIL-6 stimulation, cathelicidin-1

was unresponsive. STAT3 was shown to be phosphorylated following rIL-6 stimulation of RTS-11 cells, and an inhibitor of Jak2 (WP1066) could abolish the phosphorylation and inhibit cathelicidin-2 up-regulation, suggesting the JAK2/STAT3 pathway plays an important role in IL-6 signalling. rIL-6 was subsequently shown to increase IL-17A/F3 in HK cell cultures, with no effects apparent on the other IL-17A/F isoforms including IL-17N [56].

In terms of effects on B cells, CM from a stable cell line (RTG-2 cells) over-expressing trout IL-6 were found to maintain cells expressing the IgMH transcript in HK primary cultures for up to five weeks [154]. This was verified by FACS analysis of the numbers of IgM+ (B) cells present one week after cell isolation, in comparison to cells not receiving the IL-6 CM. In orange-spotted grouper rIL-6 was injected into fish, at 50 ng/g fish, and shown to increase the expression of IL-6, a number of transcription factors involved in T cell differentiation (Tbet, GATA3, c-maf) as well as IgMH, with effects most pronounced in the HK but also apparent in the trunk kidney and spleen [178]. Interestingly, whilst Tbet was initially increased, unlike GATA3 and c-maf it was subsequently suppressed (until 360 h post-injection) relative to PBS injected fish, hinting at a preferential effect of IL-6 on Th2 pathways. In addition, serum total and specific IgM was found increased post-injection, with total IgM levels elevated from Weeks 1 to 6 (the end of the study) and specific IgM against BSA increased from Weeks 2 to 6, in a dose dependent fashion (50 ng/g to 200 ng/g rIL-6 injected). Lastly, in fugu it has been found that IgM+ B cells (from PBL) express the two IL-6 receptor chains (IL-6R and gp130), and that stimulation of these cells with rIL-6 increased the expression of the secretory form of IgMH [179]. As with trout macrophages, rIL-6 stimulation resulted in STAT3 phosphorylation in these IgM+ cells and in Ba/F3 cells (a murine IL-3 dependent pro-B cell line) co-transfected with fugu IL-6R and gp130. The potential for IL-6 trans-signalling in fish, as occurs in mammals, was also verified with the discovery of a soluble form of the IL-6R in this species, that may permit signalling in cells expressing only gp130.

Goldfish M17 also affects macrophage function. Cells treated with M17 release nitric oxide and this effect can be increased in the presence of LPS [180]. Whilst M17 does not affect the proliferation of macrophages/monocytes/progenitor cells by itself, in the presence of CM from kidney macrophage primary cultures an enhanced proliferation was seen in the progenitor cell population. M17 also induces the differentiation of monocytes into macrophages but has no effect on progenitor cells. Such results confirm M17 as an important cytokine for macrophage proliferation, differentiation and activation in fish. In zebrafish knockdown of M17 (LIF) *in vivo* delays functional recovery after optic nerve injury, showing it also has a role in the nervous system [181].

5.4. IL-12 Subfamily

The IL-12 subfamily of cytokines consist of heterodimers of an alpha chain (p19, p35, p28) with structural similarity to the IL-6 family ligands, and a beta chain (p40, EBI3) with similarity to the extracellular domain of IL-6 family receptors. To date four members are known, namely IL-12 (p35/p40), IL-23 (p19/p40), IL-27 (p28/EBI3) and IL-35 (p35/EBI3). Whilst engagement with their receptors is unknown it can be modelled based on the IL-6-IL-6R structure [182]. In addition, p40 can form homodimers, that can act as an IL-12 antagonist [183]. All of these chains are known in fish and in many cases multiple paralogues are present, as seen in salmonids where there are $\times 2$ p19, $\times 2$ p28, $\times 3$ p35, $\times 3$ p40 genes and a single EBI3 gene [4].

To date only IL-12 has been studied functionally in fish, and in one study two different paralogues of the p40 chain were used to make two proteins with a common p35 chain [162,184,185]. In grouper rIL-12 was able to increase the Con A induced proliferation of PBL, PBL migration and TNF- α gene expression [185]. p40 alone was also able to induce chemotaxis and TNF- α expression but to a lesser extent compared to IL-12, and when added to the rIL-12 reduced its activity. When delivered orally as chitosan nanoparticle encapsulated IL-12 (at a dose of 20 μ g/g fish) grouper survival was prolonged following challenge 20 days later with *Cryptocaryon irritans* theronts. In rock bream, rIL-12 pre-incubation (for 1 h) was shown to enhance the respiratory burst of PBL induced by

Vibrio alginolyticus [184]. *In vivo* administration of rIL-12 (intraperitoneal injection, 20 µg/fish) for 4 h resulted in increased expression of several immune genes in kidney, including IL-1β, IFN, CC chemokine 1 (CCK1), IRF4, IFN stimulated gene (ISG) 15 and C7. Following injection with *V. alginolyticus* at this time lower CFUs were recovered from the kidney at Days 1 and 2 post-infection compared to fish given saline and protein controls. In rainbow trout two rIL-12 proteins have been made and shown to have overlapping but distinct bioactivity on HK cell cultures. The p35/p40b protein induced IFN-γ1 and IFN-γ2 expression, and p40b and p40c expression. However, a protein consisting of p35/p40c had a faster effect on IFN-γ1/2 (*i.e.*, induced significant effects at 6 h *vs.* 24 h for p35/p40b) and p40c expression (6 h *vs.* 48 h for p35/p40b) and in addition to modulating the above genes could induce IL-10 expression. These effects may reflect direct and indirect effects on gene expression mediated by p35/p40c and p35/p40b, respectively. Whether these proteins signal via different receptors remains to be determined but the p40 paralogues share only 24% aa identity and p40c appears to be preferentially induced by viral (viral haemorrhagic septicaemia virus, VHSV) and parasitic (proliferative kidney disease, PKD) infection in this species.

5.5. Colony Stimulating Factors

Colony stimulating factors (CSF) drive the proliferation and differentiation of blood cells from haematopoietic stem cells (HSC). Three CSFs are important for phagocyte differentiation from the common myeloid progenitor (CMP) in mammals, termed CSF-1 to CSF-3. CSF-1, also called macrophage colony stimulating factor (M-CSF), is a regulator of the mononuclear phagocyte lineage. It elicits its effects on the myeloid lineage by binding to the CSF-1R (CD115), as does the related molecule IL-34 which can additionally form a heterodimer with M-CSF for receptor docking [186]. Multiple splice variants of M-CSF exist in mammals that can have different effects on target cells. M-CSF was first described in fish from goldfish [187]. The recombinant protein was found to form a homodimer and to bind to the soluble goldfish CSF-1R (sCSF-1R), previously characterised in this species [188]. The goldfish rM-CSF was also shown to induce the differentiation of monocytes into macrophages and the proliferation of monocyte-like cells, in line with having an important role in mononuclear phagocyte function in fish, and this could be inhibited by addition of the sCSF-1R or an antibody to the CSF-1R [189]. Continuous addition of rM-CSF to goldfish primary macrophage cultures extended their longevity and resulted in the long-term cultures of functional macrophages [190]. In addition, stimulation of goldfish macrophages with rCSF-1 elevated their respiratory burst and nitric oxide responses, phagocytic activity and locomotion; all of which were inhibited by addition of sCSF-1R. The rM-CSF treated macrophages showed increased expression of immune genes such as CXCL8, CCL-1, TNF-α, IL-1β, IL-10, p35, p40, type I IFN, and iNOS [189], indicating activation of the cells. That the goldfish M-CSF acts through the CSF-1R was demonstrated by loss-of-function studies, where knockdown of CSF-1R in monocytes using RNAi abrogated monocyte proliferation and the differentiation of monocytes into macrophages [190]. *In vivo* administration (25 ng) of the rM-CSF two days earlier resulted in increased numbers of circulating monocytes (34% to 47%), and using BrdU to analyse cells undergoing DNA synthesis it was found that the labelled cells increased from 1% to 13% post-injection [190]. Lastly, supernatants from the goldfish fibroblast cell line CCL-71 can also induce the proliferation of primary macrophage cultures, and this effect is inhibited by addition of sCSF-1R, suggesting that fibroblasts in fish secrete M-CSF as seen in mammals [190].

Subsequent analysis of several teleost fish species revealed that in fact two major forms of M-CSF exist, that were termed M-CSF1 and M-CSF2, that likely arose from the teleost WGD event [191]. The two forms differ in size (M-CSF1 being larger and M-CSF2 shorter) but share a similar domain structure. The difference in protein size is due to variation in the region between the CSF-1 domain and transmembrane domain. Similar to the results with the goldfish molecule, rainbow trout rM-CSF1 was shown to be a growth factor for macrophages, and when added to macrophages or newly adherent HK cells could increase CXCR3 expression, as well as CSF-1R expression in the latter case. However, in contrast to the results with goldfish rM-CSF, trout rM-CSF1 had no effect on the expression of

a variety of pro-inflammatory genes, including IL-1 β , IL-11, IL-15, IFN- γ and TNF- α . As with IL-2/IL-6 above, a stable cell line (RTG-2 cells) over-expressing trout M-CSF1 has been developed and CM from these cells increases MCSFR2 relative expression in HK cells, such that by five weeks of treatment MCSFR2 transcripts represent 70% of those analysed (CD4-1, CD8a, IgMH, MCSFR2) [154]. This suggests that mononuclear phagocytes were the dominant cells being maintained by these M-CSF1 containing CM. No bioactivity studies with the teleost M-CSF2 have been performed to date. Similarly, whilst IL-34 has been discovered in several teleost species, to date no bioactivity studies have been performed, although differential expression of the three possible CSF-1R ligands suggest functional differences are likely present [192,193]. In mammals, IL-34 specifically directs the differentiation of myeloid cells in the skin epidermis [194].

CSF-2, also called granulocyte-macrophage CSF (GM-CSF), is discussed above under the beta-chain cytokines and it is possible an ancestral gene with relatedness the β c cytokines exists in fish although bioactivity studies are lacking. However, CSF-3 or granulocyte-CSF (G-CSF) has been discovered and studied functionally in fish. Most studies have been performed in zebrafish, where knock-down of the G-CSFR with morpholinos and overexpression of G-CSF by injection of eggs with *in vitro* transcribed G-CSF mRNA has been used to determine G-CSF function [195]. Such experiments show that there is an increase in G-CSFR+ cells and lyz+ cells (in lyz::EGFP fish) when G-CSF is used, but that these numbers fall following injection with the G-CSFR morpholinos. Importantly no increases were seen when both were co-injected, showing a likely ligand-receptor dependence. Whilst no effects on myeloperoxidase (mpo)+ cells (monocytes/macrophages) were apparent in the posterior regions of the embryo following injection with G-CSF mRNA, the anterior region population did increase and injection of G-CSFR morpholinos reduced the total mpo+ population. These findings suggest that G-CSF acts more generally on myelopoiesis rather than just affecting granulopoiesis. Further studies in zebrafish, on thrombopoiesis, showed that when thrombopoietin (Tpo) and erythropoietin (Epo) are added together to haematopoietic stem and progenitor cells (HSPCs), mixed colony forming unit thrombocytic erythroid cell (CFU-TE) colonies are found, whilst Tpo alone gave rise to CFU-T colonies, suggesting there is a common bipotent thrombocytic/erythroid progenitor (TEP) [196]. However, when G-CSF is added together with Tpo and Epo to the HSPCs, colonies derived from blood progenitors upstream of the TEPs are found, such as CMP cells, multipotent progenitor cells (MPPs) and haematopoietic stem cells (HSCs).

It is now known that two paralogues of the G-CSF gene are present in teleost fish, where in zebrafish one is located on chromosome 12 (G-CSFa or Gcsf-Chr12) and one on chromosome 19 (G-CSFb or Gcsf-Chr19). Both have now been shown to signal via the G-CSFR using loss- and gain-of-function experiments in zebrafish similar to the above [197]. Both paralogues have been made as recombinant proteins and shown to stimulate granulocyte and monocyte/macrophage differentiation in whole kidney marrow cells cultured *in vitro*, and generate myeloid colonies. However, G-CSFa was more potent at higher concentrations *in vitro* and participates in emergency myelopoiesis *in vivo*, whilst G-CSFb is active at lower concentrations *in vitro* and affects neutrophil migration to wounds, with knock-down of G-CSFb decreasing the percentage of neutrophils that arrive at the wound site via blood vessels [197,198]. Interestingly it was the numbers arriving after 1 h that were reduced, in agreement with studies showing other factors such as hydrogen peroxide are more important in the early stages of the inflammatory process [199].

6. Type II α Helical Cytokines

The type II α -helical cytokines include the IL-10 subfamily (IL-10, IL-19, IL-20, IL-22, IL-24, IL-26 and IFN- γ) and the type I and III IFNs, that are considered to be derived from a common ancestor [200]. They signal via receptors of the type II cytokine receptor family [149]. Various members of the IL-10 subfamily and type I IFNs exist in fish as outlined below, with the IFNs being particularly well studied (Table 1).

6.1. Type I IFN

Type I IFNs have diverse immune functions, including activation of antiviral defences and anti-proliferative effects. Type I IFNs have now been identified in all the jawed vertebrate lineages [86,201–206]. As in mammals, teleost fish possess two or four cysteine-containing subgroups, namely group I and II IFNs, which can be further classified into six groups phylogenetically (IFN-a-f) [207–211]. Group I IFNs (IFN-a, -d and -e) are ubiquitously present whilst group II IFNs (IFN-b, -c and -f) appear to be restricted to certain species [211]. Both groups have a ternary protein structure of four α -helices, which are conserved for the type II α -helical cytokines [212]. A notable CAWE motif is present at the C-terminus of group II IFNs but seems less apparent for group I IFNs [210].

Fish IFN genes are clustered in the genome, at loci with conserved gene synteny [207,209,211]. In zebrafish two loci are found, on chromosomes 3 and 12, which harbour three and one IFN genes, respectively. The open reading frame of all teleost fish type I IFN genes is separated by four phase 0 introns [202,207,209,213–215], in common with amphibian genes and unlike amniotes where the genes are intronless. The number of IFN genes in fish varies significantly among species, with salmonids having the highest gene copy number in all the jawed vertebrates [207,209,211,216].

6.2. Group I IFNs

IFN-as were the first group of type I IFNs sequenced in fish [201,217,218], and are now known to be present in siluriformes [216,217], cyprinids [201,207], and salmonids [209–211,218]. Most recently, the IFN-a genes have also been identified in medaka and turbot, suggesting they are likely more widely distributed in teleosts than previously thought [219,220]. The IFN-as are ubiquitously induced in fish cells after viral infection or activation by viral PAMPs, to some degree mimicking the expression pattern of IFN- β in mammals, and have been shown to up-regulate a large panel of ISGs which are essential for host antiviral defence [201,207,210,214,218,219,221,222]. The common ISGs responsive to IFN-as include Mx [201,210,218,221], viperin [207], ISG15 [223], IRFs [224,225], and grass carp reovirus induced genes (Gigs) [226]. Furthermore, trout IFN-a2/IFN2 increases the expression of MDA-5 and LGP-2, which are important intracellular PRRs for recognition of viral PAMPs, leading to the magnification of the IFN response [227]. The positive regulation of the IFN response by IFN-a is also observed in RTG-2 cells, where IFN-a stimulation significantly increases the expression of IFN isoforms including itself [208]. Several studies have examined the global impact of IFN-as on gene expression at the transcript level in cell lines [228,229], and in general, the genes affected by IFN-a are similar to what is observed in mammals. Interestingly, a recent study indicates that the black carp IFN-a is glycosylated at Asn (N)38 when produced in EPC cells but that glycosylation does not contribute directly to the antiviral property [230].

In salmon TO cells, the IFN responsive genes are significantly different from those modulated by Salmonid Alpha Virus (SAV), that has a positive single stranded genome. IFN-a1 enhances resistance of host cells against some but not all virus types. For example, IFN-a1 is effective in enhancing cell protection against SAV and Infectious Pancreatic Necrosis Virus (IPNV, a double-stranded RNA virus) but surprisingly is relatively ineffective against Infectious Salmon Anaemia Virus (ISAV, a single-stranded RNA segmented orthomyxovirus) in TO cells despite transient induction of antiviral genes [128,231–233]. The timing of IFN stimulation is critical to protection. When salmon TO cells are pre-incubated with IFN-a prior to infection (4–24 h) with SAV they are protected against virus-induced CPE but if administered at the time of infection and up to 24 h post-infection they are not [234]. In agreement with the *in vitro* studies, intramuscular injection of an expression plasmid encoding IFN-a1 into Atlantic salmon fails to increase survival against ISAV infection [223]. The mechanism(s) underlying the variable efficacies of IFN elicited protection against different viruses is currently not understood. However, it is known that viruses exploit diverse strategies to counteract host antiviral defences including the IFN-mediated pathway, and that strains of different virulence (as seen with virulent strain ISAV7 *vs.* avirulent strain ISAV4) vary in their activation of the IFN response [233].

Several studies suggest that IFN- α may play a role in the regulation of inflammation in fish. Zebrafish IFN- α induces expression of several pro-inflammatory genes *in vivo* when the recombinant protein (produced in HEK 293 cells) is injected into fish [235]. Turbot IFN2 (a member of the IFN- α group) has recently been characterised functionally. Surprisingly, when administered by intramuscular injection, it is unable to trigger an antiviral response and provide protection against VHSV [220]. However, unexpectedly, it has a stimulatory effect on the expression of pro-inflammatory genes such as IL-1 β . Based on the fact that turbot IFN2/IFN- α is up-regulated during infection with VHSV and the bacterial pathogen *Aeromonas salmonicida*, it is suggested that turbot IFN2/IFN- α may also serve as a mediator in regulating inflammation.

In addition to the secreted IFN- α s, transcript variants that translate into proteins that lack a signal peptide have been described in several teleost species [217,236,237]. Further analysis in trout reveals that such transcripts originate from mRNA alternative splicing of the intron(s) located at the 5' end of the IFN- α gene [237,238]. The two trout alternatively spliced IFN- α transcripts translate into intracellular proteins that display similar antiviral functions to the secreted protein form but via interaction with cytosolic IFN receptors to activate the Jak/STAT signalling pathway. These data suggest that an active IFN system operates within virus-infected cells in fish.

The antiviral effect of the IFN- δ subgroup is somewhat controversial. IFN- δ s are the most common IFNs found in teleosts and exist as a single copy in cyprinids and salmonids but as multiple copies in the Acanthopterygiiian species. Like the IFN- α group in cyprinids and salmonids, the IFN- δ s are the major type I IFNs that activate the antiviral responses in Acanthopterygiiians [214,219,239]. With the recent discovery of IFN- α in medaka, in addition to IFN- δ , their antiviral activities have now been studied comparatively. Both IFNs increase expression of ISG15 and Gig genes in the medaka DIT cell line when overexpressed and can enhance cell resistance to viruses [219].

In contrast to the Acanthopterygiiian IFN- δ , the functions of IFN- δ in cyprinids and salmonids are currently poorly studied. In Atlantic salmon, the IFN- δ gene is constitutively expressed in tissues but is not induced by viral mimics such as poly(I:C) or R848 [221]. However, in rainbow trout and zebrafish, it is activated during viral infection [207,208]. Salmon IFN- δ has been produced as a recombinant protein in bacteria and HEK 293 cells but no activities have been detected [221].

The IFN- ϵ s are the latest member of the group I IFN family. Seven IFN- ϵ genes have been found in rainbow trout [211] and are expected to be present in other salmonid species. Phylogenetically, trout IFN- ϵ s can be divided into two subgroups, one consisting of IFN- ϵ 1-4 and the other IFN- ϵ 5-7. They can be induced after stimulation with poly(I:C) and during viral infection but their functions remain to be characterised.

6.3. Group II IFNs

Group II IFNs have been identified in cyprinids, salmonids, and turbot, the latter belonging to the Pleuronectiforme family [207,209,210,220]. Akin to group I IFNs, group II IFNs also consist of three subgroups, IFN- β , - γ and - δ . The antiviral properties of IFN- β s and IFN- γ s have been studied but not as extensively as group I IFNs, and no functional data are available for IFN- δ . The IFN- β and IFN- γ group are viral inducible cytokines and expression is largely restricted to certain leucocyte populations [210,221], suggesting different roles to the group I IFNs. Recombinant trout IFN- β 1/IFN3 has been shown to weakly induce Mx expression in RTS-11 cells [210]. Later studies compared the antiviral activities of IFN- β and IFN- γ in Atlantic salmon, indicating that both activate antiviral genes but that IFN- γ is more potent than IFN- β [221]. Intramuscular injection with an IFN- γ encoding expression plasmid gives strong protection of fish against ISAV challenge. In contrast, an IFN- β encoding plasmid provides much weaker protection [223]. In zebrafish, recombinant IFN ϕ 2 (an IFN- β member) shows similar stimulatory effects to IFN ϕ 1 (an IFN- α member) in terms of modulating viperin expression in cultured cells and they confer comparable resistance of fish larvae against Infectious Haematopoietic Necrosis Virus (IHNV) [207]. Interestingly, zebrafish IFN ϕ 2 and IFN ϕ 3 (both IFN- β

members) and turbot IFN1 (a group II member) display weaker antibacterial effects and/or effects on pro-inflammatory gene expression than the group I IFNs [220,235].

Unlike mammalian type I IFNs, which bind to the same receptor complex consisting of IFNAR1 and IFNAR2, teleost group I and II IFNs activate different receptors to trigger cellular responses [207]. In zebrafish, group I (IFN ϕ 1/4, IFN-a/d) and II (IFN ϕ 2/3, IFN-c) IFNs share a common receptor chain (CRFB5) which is an orthologue of the mammalian IFNAR1 but each interacts with a ligand specific receptor; CRFB1 for group I and CRFB2 for group II IFNs [207]. The type I IFN receptors are even more complex in Atlantic salmon where eight copies of IFN receptor candidates have been found [240]. These include 3 \times CRFB5s, 2 \times CRFB1s, 1 \times CRFB2, 1 \times CRFB3 and 1 \times CRFBx. Consistent with studies in zebrafish, CRFB1a and CRFB2 mediate signal transduction of group I (IFN-a) and II (IFN-b/c) IFNs respectively. However, multiple CRFB5 paralogues seem to engage with different groups of IFN ligands. The IFN-a pairs with the CRFB5a, CRFB5b or CRFB5c, IFN-b with CRFBx and IFN-c with CRFB5a or CRFB5c respectively. In addition, intracellular forms of CRFB2 and CRFB5 exist in rainbow trout and are involved in activation of cellular responses by the intracellular IFN-a ligand [238].

The actions of type I IFNs are controlled by multiple transcription factors. Several studies have demonstrated that teleost fish type I IFNs signal via the conserved Jak-STAT pathway [203,205,206,241]. Many of these key transcription factors have now been characterised in fish. It has become apparent that zebrafish possess a complete set of Jak family members comprising Jak1, Jak2, Jak3 and Tyk2, with Jak2 further duplicated into two paralogues (Jak2a and Jak2b) [242,243]. Jak2a is suggested to be the functional orthologue of mammalian Jak2 [243]. The involvement of Tyk2 in IFN signalling has also been studied in Atlantic salmon. Structurally, salmon Tyk2 is organised into seven JAK-homology (JH) domains, with JH1 being the critical domain catalysing autophosphorylation [244]. In teleosts, multiple paralogues of STAT1 and STAT2 proteins (e.g., STAT1a, STAT2a and STAT2b) have been shown to control cellular signalling of type I IFNs [245].

The IFN responses can be negatively regulated by several transcription factors. The main negative regulators comprise the SOCS proteins and the protein inhibitor of activated STAT (PIAS) family. These transcription factors are highly conserved in sequence and protein structure and are expected to play similar roles to their mammalian counterparts in suppressing the IFN pathway in fish [246–248].

6.4. Type II IFNs

Type II IFN exerts regulatory roles in both innate and adaptive immunity, including activating macrophages, enhancing antigen presentation and promoting the Th1 T cell response. They are encoded by a single gene in mammals but in teleosts two members have been identified and are termed IFN- γ (IFN- γ 2 in zebrafish and other cyprinids) or IFN- γ related molecule (IFN- γ rel, or IFN- γ 1 in zebrafish and other cyprinids) [249–254]. An IFN- γ orthologue is also present in the elephant shark [86].

In general, teleost IFN- γ s display conserved functions with their mammalian orthologues (Table 1). The stimulatory effects of IFN- γ on fish monocytes/macrophages are well documented. IFN- γ is able to increase production of nitric oxide and reactive oxygen intermediates (ROI), and phagocyte activity in several species [254–257]. A recent report has demonstrated that carp IFN- γ 2 (IFN- γ) promotes formation and activation of extracellular traps in macrophages and neutrophils to enhance their antimicrobial activity [256]. In IFN- γ treated cells, genes associated with the inflammatory response are markedly activated, highlighted by the elevated expression of IL-20L [258], IL-34 [60], TNF- α [58], CXCL11_L1 (γ IP) [67,228,234,254], and iNOS [256]. However, IFN- γ has a weak stimulatory effect on the expression of IL-1 β [20,161,255] and CXCL8 paralogues [67,255], synergising with IFN- γ rel or LPS to increase their transcript levels [255,259]. Other genes shown to be induced by IFN- γ include the antiviral effector genes such as guanylate binding protein 1 (GBP1), Mx, and ISG15 [232,254], and those (e.g., gp91phox and p47phox) associated with NADPH oxidase activation [255].

Molecular evidence suggests that antigen presentation pathways are also modulated by IFN- γ . The transcripts of the MHC II gene are significantly increased in RTS-11 cells 24 h after treatment with IFN- γ [254]. In addition, global transcriptomic analysis has shown that several key genes involved in the MHC I antigen presentation pathway are also up-regulated by IFN- γ [57].

Cell-mediated T cell responses are also potentially modulated by IFN- γ in fish. Although Th cells have not been functionally characterised in fish, cytokines mainly involved in T cell immunity are now known to exist and have been shown to be altered by IFN- γ . For example, several γ -chain cytokines including IL-2 [56], IL-15 [161] and IL-21 [56], are induced by IFN- γ in trout or salmon leucocytes, confirming its role in promoting Th1 responses and memory T cell maturation. In contrast, the major cytokines (e.g., IL-17 isoforms) involved in Th17 responses are not affected [56].

Protective effects of IFN- γ against infection with intracellular pathogens have been examined in cultured cells and in fish. In trout RTS-11 cells, IFN- γ weakly induces Mx expression [254], whilst in salmon cells treated with IFN- γ , IPNV replication is inhibited and the CPE titres are substantially reduced. The protective effect is only partially dependent on IFN- α induction, since antibodies to IFN- α do not completely block the IFN- γ -mediated induction of Mx and ISG15 [232]. In addition, intraperitoneal injection of rIFN- γ activates immune responses against infection with *Edwardsiella tarda* in the olive flounder, resulting in significantly increased survival [260].

6.5. IFN- γ rel

The IFN- γ rel molecules share moderate homology with the teleost IFN- γ s and are considered as a second member of the type II IFN family. The IFN- γ rel proteins have recently been classified into two subgroups, namely IFN- γ rel1 and IFN- γ rel2, based on the presence of the nuclear localisation signal (NLS) at the C-terminus [261]. IFN- γ rel1 contains a functional NLS whilst IFN- γ rel2 does not. To date, most of the functional studies have been conducted in cyprinid species.

IFN- γ rel is thought to be involved in immune defence against bacterial infection. The Rohu carp IFN- γ rel gene is up-regulated after infection with *A. hydrophila*, *E. tarda* or *Shigella flexneri* [251]. The expression of IFN- γ rel is also detected in LPS-stimulated common carp leucocytes enriched for B-cells [262]. In zebrafish embryos, knockdown of the IFN- γ rel1 (IFN- γ 1) and its cognate receptor CRFB13 by morpholinos results in a significant reduction of embryo survival after infection with the bacterial pathogen *Yersinia ruckeri*, suggesting it plays an important role in innate defence against bacterial infection [263]. When injected intramuscularly into Fugu, IFN- γ rel increases expression of IL-1 β , IL-6, p35, p40, and IFN- γ , which are important mediators of immune responses against bacterial infection [264]. In Fugu, IFN- γ rel and IFN- γ appear to trigger similar responses, including phagocytic activity and lysozyme activity.

The activities of the IFN- γ rel2 (IFN- γ 1) protein have been studied extensively in goldfish monocytes/macrophages in parallel with IFN- γ [255]. Curiously, the goldfish IFN γ rel2 is more potent than IFN- γ in inducing expression of iNOS, TNF- α 2, CXCL8/IL-8 and ceruloplasmin, and in enhancing nitric oxide production and phagocytosis in monocytes/macrophages. Moreover, it has a stronger temporal priming effect on monocytes for ROI production but surprisingly down-regulates the priming effect of IFN- γ and TNF- α 2. As shown in goldfish monocytes/macrophages, IFN- γ rel signalling is mediated by the phosphorylation and nuclear transport of STAT1, a crucial transcription factor regulating IFN- γ signalling. However, in carp, no stimulatory effect of IFN- γ rel on iNOS expression has been detected in phagocytes [259].

The antiviral effect of the two IFN- γ rel subgroups have also been comparatively examined in the ginbuna crucian carp [252]. Both cytokines are able to protect fish against infection with crucian carp haematopoietic necrosis virus (CHNV), and monomers of both cytokines are active. Considering the fact that the IFN- γ rel2 lacks the NLS motif, it has been suggested that the two IFN- γ rels may exert overlapping antiviral functions but act in different compartments in fish cells.

6.6. IL-10

IL-10 is an anti-inflammatory cytokine and suppresses immune responses. The IL-10 gene has been identified in a number of teleost fish [200,265–272]. It exists as a single copy gene for most species but duplicated copies are present in rainbow trout. The herpesviruses of European eel and Koi carp also encode an IL-10 homologue [273–275].

The functions of IL-10 have been characterised in goldfish and carp [276] (Table 1). In general, the fish IL-10s, similar to the mammalian orthologues, act as a suppressor and exert a conserved role in dampening inflammatory responses. In goldfish monocytes activated with heat-killed *A. salmonicida*, incubation with IL-10 substantially decreases the expression of TNF- α 1, TNF- α 2, IL-1 β 1, IL-10, CXCL_8/IL-8, and the NADPH oxidase component p47(phox) [265]. On the other hand, pre-treatment of monocytes with 10–1000 ng/mL of IL-10 substantially abrogates *A. salmonicida* or IFN- γ activated ROI production. IL-10 also induces phosphorylation and nuclear translocation of STAT3 and rapid expression of SOCS3 [265]. Consistent with these observations, similar effects of carp IL-10 have been shown in PMA or LPS activated neutrophils and macrophages [276]. In addition, IL-10 inhibits expression of genes involved in MHC antigen presentation in carp macrophages.

Using an immunisation model, the roles of IL-10 have also been investigated in carp [276]. In HK cell cultures derived from fish immunised with *Trypanoplasma borreli* antigens, stimulation with IL-10 plus antigen significantly increases IgM antibody secretion. Interestingly, this stimulatory effect is not seen in PBL. When the HK cells from immunized fish were cultured with the *T. borreli* antigens alone or in combination with IL-10 for six days, the expression of cytokines and transcription factors associated with Th1 and Th2 responses was increased by antigen but decreased by the presence of IL-10, with a concomitant increase in CD8 α 2 and CD8 β 1 expression. These data suggest that carp IL-10 has differential effects on memory CD4+ and CD8+ T cells, and promotes B cell differentiation and IgM antibody secretion in an antigen specific manner.

A viral homologue of IL-10 has been identified recently in the Koi herpesvirus (Khv) genome and characterised functionally [273–275]. It has moderate homology with carp IL-10, and has a predicted signal peptide, but is not essential for viral replication *in vitro* or virulence *in vivo* [275]. In virus infected carp, the KhvIL-10 gene is highly expressed at the acute and reactivation phases. The KhvIL-10 increases the number of lyz+ cells when injected into zebrafish embryos, similar to the effect seen with zebrafish IL-10, and this activity can be inhibited by co-injection with IL-10R1 morpholinos [274]. A recent study has extended the functional analysis where the KhvIL-10 is shown to exert overlapping functions with the host IL-10, as an anti-inflammatory factor via the STAT3 signalling pathway [273]. These data demonstrate that fish herpesviruses produce a functional IL-10 analogue to suppress the host immune response to establish infections.

6.7. Other Members of the Type II α -Helical Cytokine Family

Other members of the type II α -helical cytokine family include IL-19, IL-20, IL-22, IL-24 and IL-26. The genes encoding IL-19, IL-20 and IL-24 are clustered with the IL-10 gene, whilst the IL-22 and IL-26 genes reside next to IFN- γ at a separate locus. In teleosts, a cytokine homologue related to the mammalian IL-19/IL-20/IL-24 genes, termed IL-20 like (IL-20L) in trout, has been identified [200,258]. Teleost fish also possess orthologues of IL-22 and IL-26 [242,250,277,278]. Among these cytokines, only the activity of the trout and So-iny mullet IL-22 has been described to date [277,278]. In trout splenocytes, IL-22 can up-regulate the expression of AMP genes including β -defensins 3 and 4, liver expressed antimicrobial peptide 2A and hepcidin [277] (Table 1). Furthermore, administration of the IL-22 into mullet significantly improves fish survival after challenge with the Gram positive bacterial pathogen *Streptococcus dysgalactiae* [278]. Like mammalian IL-22, fish IL-22s are expected to play a major role in co-ordinating immune defence against bacterial pathogens, and their expression has been shown to correlate with vaccine-induced protection [279].

Table 1. Biological activity of fish type II cytokines.

Cytokine	Human homologue	Receptors	Known immune functions	references
IL-10, viral IL-10	IL-10	IL-10R1/CRFB12, IL-10R2/CRFB4	Suppress immune responses, inhibit inflammation, promote T cell proliferation, memory B cells, and IgM production	[265,273,276]
IL-20L	IL19, IL-20, IL-24	Not characterised	Not characterised	[200,258]
IL-22	IL-22	Not characterised	Activate antimicrobial peptide genes and antibacterial immunity	[277,278]
IL-26	IL-26	Not characterised	Not characterised	[242,250]
Group I IFN (IFN-a, d, e)	Type I IFN	IFNAR1/CRFB5, IFNAR2/CRFB1	Induce expression of the antiviral effectors, promote apoptosis, regulate inflammation	[201,207,210,221,234]
Group II IFN (IFN-b, c, f)	Type I IFN	IFNAR1/CRFB5, IFNAR2/CRFB2	Induce expression of the antiviral effectors, promote apoptosis	[207,210,221,223]
IFN- γ	Type II IFN	IFN- γ R1/CRFB13, IFN- γ R2/CRFB6	Activate phagocytes, enhance antigen presentation, promote Th1 cytokine expression	[254–257]
IFN- γ rel	Type II IFN	CRFB17, IFN- γ R2/CRFB6	Regulate anti-bacterial and antiviral immunity	[255,261]

7. Conclusions

There is a clear conservation of the cytokine network throughout vertebrates, with many of the key cytokines important for haematopoiesis, inflammation and adaptive immunity present from fish to mammals. However, in many cases, the genes are not true homologues and may be related to ancestral genes that have diverged independently in different vertebrate lineages. Hence the importance of determining the protein function in each species studied. Nevertheless, the data obtained to date, and reviewed here, show that so far there have been relatively few surprises. Fish cytokines have a role in development and haematopoiesis, in line with knowledge from mammalian studies. They attract leucocytes to a site of infection and activate their antimicrobial mechanisms to kill any invaders. The cytokines present have the potential to influence the type of response elicited, with classical Type 1 and Type 2 immunity likely controlled by the release of appropriate cytokines from innate lymphoid cells or helper T cells. This information gives us an insight into what may have been the minimal cytokine network needed for an early adaptive immune system, to allow appropriate expansion of antigen-selected cells without the risk of excessive host damage. In addition, as we characterise the cell populations that release the different cytokines present, we will begin to realise the potential for manipulating such responses, with many possible applications. For example, they may be useful as markers of effective vaccination of fish in aquaculture, or for selective breeding of “robust” fish to improve fish welfare. The diversity of fish species can be a confounding factor in our efforts to characterise these genes and the functions of the encoded proteins but it is clear that a few key fish species are driving the field forwards, from economic necessity or the ability to perform genetic manipulations *in vivo*. So indeed it is true to say that we do now know a lot about the function of many fish cytokines, and this is aiding our ability to characterise the fish immune system and pathways leading to protective immunity.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

The following abbreviations are used in this manuscript:

CCK CC chemokine
 CD cluster of differentiation

CISH	cytokine inducible Src homology 2 (SH2)-containing protein
CNTF	ciliary neurotrophic factor
COX-2	cyclooxygenase-2
CRFB	class II cytokine receptor family member
CSF	colony stimulating factor
DAMP	danger associated molecular pattern
DC-SIGN	dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin
DIGIRR	double immunoglobulin IL-1R related molecule
γ IP	gamma interferon inducible protein
GBP	guanylate binding protein
G-CSF	granulocyte colony stimulating factor
GM-CSF	granulocyte-macrophage colony-stimulating factor
HK	head kidney
HSC	haematopoietic stem cell
IGFBP	insulin growth factor binding protein
ICE	IL-1 β converting enzyme
IEC	intestinal epithelial cell
IFN	interferon
IFN- γ rel	interferon gamma related molecule
Ig	immunoglobulin
IL	interleukin
IL-1F	interleukin-1 family member
IL-1Ra	interleukin 1 receptor antagonist
IL-1RAcP	interleukin-1 receptor accessory protein
IL-36Ra	interleukin-36 receptor antagonist
ILC	innate lymphoid cell
iNOS	inducible nitric oxide synthase
IRAK	interleukin 1 receptor associated kinase
IRF	interferon regulatory factor
ISG	interferon stimulated gene
JAK	Janus kinase
LPS	lipopolysaccharide
LT	lymphotoxin
MAPK	mitogen-activated protein kinase
M-CSFR	macrophage colony stimulating factor receptor
MyD88	myeloid differentiation factor 88
NF- κ B	nuclear factor kappa-light-chain-enhancer of activated B cells
nIL-1Fm	novel interleukin-1 family member
PBL	peripheral blood leucocyte
PAMP	pathogen associated molecular pattern
poly(I:C)	Polyinosinic-polycytidylic acid
PRR	pattern recognition receptor
ROS	reactive oxidative species
SIGIRR	single domain interleukin-1 receptor related molecule
SOCS	suppressors of cytokine signalling
STAT	signal transducers and activators of transcription
TACE	TNF- α -converting enzyme
TGF	transforming growth factor
TIR	Toll/IL-1 receptor
TNF	tumour necrosis factor
TNFSF	tumour necrosis factor superfamily member
TNFSFR	tumour necrosis factor superfamily member receptor
TRAF	tumour necrosis factor receptor associated factor
Treg	regulatory T cell
Tyk	tyrosine kinase
WGD	whole genome duplication

References

1. Secombes, C.J.; Wang, T.H.; Bird, S. The interleukins of fish. *Dev. Comp. Immunol.* **2011**, *35*, 1336–1345. [[CrossRef](#)] [[PubMed](#)]
2. Wang, T.; Secombes, C.J. The cytokine networks of adaptive immunity in fish. *Fish Shellfish Immunol.* **2013**, *35*, 1703–1718. [[CrossRef](#)] [[PubMed](#)]

3. Secombes, C.J.; Zou, J.; Bird, S. Cytokines of cartilaginous fish. In *Immunobiology of the Shark*; Smith, S.L., Sim, R.B., Flajnik, M.F., Eds.; CRC Press: Boca Raton, FL, USA, 2015; pp. 123–142.
4. Secombes, C.J.; Wang, T.; Bird, S. Vertebrate cytokines and their evolution. In *The Evolution of the Immune System: Conservation and Diversification*; Malagoli, D., Ed.; Academic Press Inc.: Cambridge, MA, USA, 2016; Chapter 5; pp. 87–151.
5. Bird, S.; Tafalla, C. Teleost chemokines and their receptors. *Biology* **2015**, *4*, 756–784. [[CrossRef](#)] [[PubMed](#)]
6. Schmitz, J.; Owyang, A.; Oldham, E.; Song, Y.; Murphy, E.; McClanahan, T.K.; Zurawski, G.; Moshrefi, M.; Qin, J.; Li, X.; *et al.* IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. *Immunity* **2005**, *23*, 479–490. [[CrossRef](#)] [[PubMed](#)]
7. Sims, J.E.; Nicklin, M.J.; Bazan, J.F.; Barton, J.L.; Busfield, S.J.; Ford, J.E.; Kastelein, R.A.; Kumar, S.; Lin, H.; Mulero, J.J.; *et al.* A new nomenclature for IL-1-family genes. *Trends Immunol.* **2001**, *22*, 536–537. [[CrossRef](#)]
8. Bird, S.; Wang, T.; Zou, J.; Cunningham, C.; Secombes, C.J. The first cytokine sequence within cartilaginous fish: IL-1 β in the small spotted catshark (*Scyliorhinus canicula*). *J. Immunol.* **2002**, *168*, 3329–3340. [[CrossRef](#)] [[PubMed](#)]
9. Engelsma, M.Y.; Stet, R.J.; Schipper, H.; Verburg-van Kemenade, B.M. Regulation of interleukin 1 beta RNA expression in the common carp, *Cyprinus carpio* L. *Dev. Comp. Immunol.* **2001**, *25*, 195–203. [[CrossRef](#)]
10. Fujiki, K.; Shin, D.H.; Nakao, M.; Yano, T. Molecular cloning and expression analysis of carp (*Cyprinus carpio*) interleukin-1 beta, high affinity immunoglobulin E Fc receptor gamma subunit and serum amyloid A. *Fish Shellfish Immunol.* **2000**, *10*, 229–242. [[CrossRef](#)] [[PubMed](#)]
11. Pleguezuelos, O.; Zou, J.; Cunningham, C.; Secombes, C.J. Cloning, sequencing, and analysis of expression of a second IL-1beta gene in rainbow trout (*Oncorhynchus mykiss*). *Immunogenetics* **2000**, *51*, 1002–1011. [[CrossRef](#)] [[PubMed](#)]
12. Scapigliati, G.; Buonocore, F.; Bird, S.; Zou, J.; Pelegrin, P.; Falasca, C.; Prugnoli, D.; Secombes, C.J. Phylogeny of cytokines: Molecular cloning and expression analysis of sea bass *Dicentrarchus labrax* interleukin-1beta. *Fish Shellfish Immunol.* **2001**, *11*, 711–726. [[CrossRef](#)] [[PubMed](#)]
13. Yin, Z.; Kwang, J. Carp interleukin-1 beta in the role of an immuno-adjuvant. *Fish Shellfish Immunol.* **2000**, *10*, 375–378. [[CrossRef](#)] [[PubMed](#)]
14. Zou, J.; Grabowski, P.S.; Cunningham, C.; Secombes, C.J. Molecular cloning of interleukin 1beta from rainbow trout *Oncorhynchus mykiss* reveals no evidence of an ice cut site. *Cytokine* **1999**, *11*, 552–560. [[CrossRef](#)] [[PubMed](#)]
15. Zou, J.; Cunningham, C.; Secombes, C.J. The rainbow trout *Oncorhynchus mykiss* interleukin-1 beta gene has a differ organization to mammals and undergoes incomplete splicing. *Eur. J. Biochem.* **1999**, *259*, 901–908. [[CrossRef](#)] [[PubMed](#)]
16. Huising, M.O.; Stet, R.J.; Savelkoul, H.F.; Verburg-van Kemenade, B.M. The molecular evolution of the interleukin-1 family of cytokines; IL-18 in teleost fish. *Dev. Comp. Immunol.* **2004**, *28*, 395–413. [[CrossRef](#)] [[PubMed](#)]
17. Wang, T.; Bird, S.; Koussounadis, A.; Holland, J.W.; Carrington, A.; Zou, J.; Secombes, C.J. Identification of a novel IL-1 cytokine family member in teleost fish. *J. Immunol.* **2009**, *183*, 962–974. [[CrossRef](#)] [[PubMed](#)]
18. Zou, J.; Bird, S.; Truckle, J.; Bols, N.; Horne, M.; Secombes, C. Identification and expression analysis of an IL-18 homologue and its alternatively spliced form in rainbow trout (*Oncorhynchus mykiss*). *Eur. J. Biochem.* **2004**, *271*, 1913–1923. [[CrossRef](#)] [[PubMed](#)]
19. Engelsma, M.Y.; Stet, R.J.; Saeij, J.P.; Verburg-van Kemenade, B.M. Differential expression and haplotypic variation of two interleukin-1beta genes in the common carp (*Cyprinus carpio* L.). *Cytokine* **2003**, *22*, 21–32. [[CrossRef](#)]
20. Husain, M.; Bird, S.; van Zwieten, R.; Secombes, C.J.; Wang, T. Cloning of the IL-1beta3 gene and IL-1beta4 pseudogene in salmonids uncovers a second type of IL-1beta gene in teleost fish. *Dev. Comp. Immunol.* **2012**, *38*, 431–446. [[CrossRef](#)] [[PubMed](#)]
21. Lee, D.S.; Hong, S.H.; Lee, H.J.; Jun, L.J.; Chung, J.K.; Kim, K.H.; Jeong, H.D. Molecular cDNA cloning and analysis of the organization and expression of the IL-1beta gene in the Nile tilapia, *Oreochromis niloticus*. *Comp. Biochem. Physiol. A. Mol. Integr. Physiol.* **2006**, *143*, 307–314. [[CrossRef](#)] [[PubMed](#)]
22. Ogryzko, N.V.; Renshaw, S.A.; Wilson, H.L. The IL-1 family in fish: Swimming through the muddy waters of inflammasome evolution. *Dev. Comp. Immunol.* **2014**, *46*, 53–62. [[CrossRef](#)] [[PubMed](#)]

23. Overgard, A.C.; Nepstad, I.; Nerland, A.H.; Patel, S. Characterisation and expression analysis of the Atlantic halibut (*Hippoglossus hippoglossus* L.) cytokines: IL-1 β , IL-6, IL-11, IL-12 β and IFN γ . *Mol. Biol. Rep.* **2012**, *39*, 2201–2213. [[CrossRef](#)] [[PubMed](#)]
24. Taechavasonyoo, A.; Kondo, H.; Nozaki, R.; Suzuki, Y.; Hirono, I. Identification of novel interleukin 1 beta family genes in Japanese flounder *Paralichthys olivaceus*. *Fish Shellfish Immunol.* **2013**, *34*, 393–396. [[CrossRef](#)] [[PubMed](#)]
25. Wang, Y.; Wang, Q.; Baoprasertkul, P.; Peatman, E.; Liu, Z. Genomic organization, gene duplication, and expression analysis of interleukin-1beta in channel catfish (*Ictalurus punctatus*). *Mol. Immunol.* **2006**, *43*, 1653–1664. [[CrossRef](#)] [[PubMed](#)]
26. Angosto, D.; Lopez-Castejon, G.; Lopez-Munoz, A.; Sepulcre, M.P.; Arizcun, M.; Meseguer, J.; Mulero, V. Evolution of inflammasome functions in vertebrates: Inflammasome and caspase-1 trigger fish macrophage cell death but are dispensable for the processing of IL-1 β . *Innate Immun.* **2012**, *18*, 815–824. [[CrossRef](#)] [[PubMed](#)]
27. Chaves-Pozo, E.; Pelegrin, P.; Garcia-Castillo, J.; Garcia-Ayala, A.; Mulero, V.; Meseguer, J. Acidophilic granulocytes of the marine fish gilthead seabream (*Sparus aurata* L.) produce interleukin-1beta following infection with *Vibrio anguillarum*. *Cell Tissue Res.* **2004**, *316*, 189–195. [[CrossRef](#)] [[PubMed](#)]
28. Pelegrin, P.; Chaves-Pozo, E.; Mulero, V.; Meseguer, J. Production and mechanism of secretion of interleukin-1beta from the marine fish gilthead seabream. *Dev. Comp. Immunol.* **2004**, *28*, 229–237. [[CrossRef](#)] [[PubMed](#)]
29. Vojtech, L.N.; Scharping, N.; Woodson, J.C.; Hansen, J.D. Roles of inflammatory caspases during processing of zebrafish interleukin-1beta in *Francisella noatunensis* infection. *Infect. Immun.* **2012**, *80*, 2878–2885. [[CrossRef](#)] [[PubMed](#)]
30. Garlanda, C.; Dinarello, C.A.; Mantovani, A. The interleukin-1 family: Back to the future. *Immunity* **2013**, *39*, 1003–1018. [[CrossRef](#)] [[PubMed](#)]
31. Afonina, I.S.; Muller, C.; Martin, S.J.; Beyaert, R. Proteolytic processing of interleukin-1 family cytokines: Variations on a common theme. *Immunity* **2015**, *42*, 991–1004. [[CrossRef](#)] [[PubMed](#)]
32. Bo, Y.X.; Song, X.H.; Wu, K.; Hu, B.; Sun, B.Y.; Liu, Z.J.; Fu, J.G. Characterization of interleukin-1beta as a proinflammatory cytokine in grass carp (*Ctenopharyngodon idella*). *Fish Shellfish Immunol.* **2015**, *46*, 584–595. [[CrossRef](#)] [[PubMed](#)]
33. Buonocore, F.; Mazzini, M.; Forlenza, M.; Randelli, E.; Secombes, C.J.; Zou, J.; Scapigliati, G. Expression in *Escherchia coli* and purification of sea bass (*Dicentrarchus labrax*) interleukin 1beta, a possible immunoadjuvant in aquaculture. *Mar. Biotechnol.* **2004**, *6*, 53–59. [[CrossRef](#)] [[PubMed](#)]
34. Hong, S.; Zou, J.; Crampe, M.; Peddie, S.; Scapigliati, G.; Bols, N.; Cunningham, C.; Secombes, C.J. The production and bioactivity of rainbow trout (*Oncorhynchus mykiss*) recombinant IL-1 beta. *Vet. Immunol. Immunopathol.* **2001**, *81*, 1–14. [[CrossRef](#)]
35. Yu, Y.; Zhong, Q.; Li, C.; Jiang, L.; Sun, Y.; Wang, X.; Wang, Z.; Zhang, Q. Molecular cloning and characterization of interleukin-1beta in half-smooth tongue sole *Cynoglossus semilaevis*. *Vet. Immunol. Immunopathol.* **2012**, *146*, 270–276. [[CrossRef](#)] [[PubMed](#)]
36. Latz, E.; Xiao, T.S.; Stutz, A. Activation and regulation of the inflammasomes. *Nat. Rev. Immunol.* **2013**, *13*, 397–411. [[CrossRef](#)] [[PubMed](#)]
37. Reis, M.I.; do Vale, A.; Pereira, P.J.; Azevedo, J.E.; Dos Santos, N.M. Caspase-1 and IL-1 β processing in a teleost fish. *PLoS ONE* **2012**, *7*, e50450. [[CrossRef](#)] [[PubMed](#)]
38. Reis, M.I.; Costa-Ramos, C.; do Vale, A.; dos Santos, N.M. Molecular cloning of sea bass (*Dicentrarchus labrax* L.) caspase-8 gene and its involvement in *Photobacterium damsela* ssp. piscicida triggered apoptosis. *Fish Shellfish Immunol.* **2010**, *29*, 58–65. [[CrossRef](#)] [[PubMed](#)]
39. Mathew, J.A.; Guo, Y.X.; Goh, K.P.; Chan, J.; Verburg-van Kemenade, B.M.; Kwang, J. Characterisation of a monoclonal antibody to carp IL-1beta and the development of a sensitive capture ELISA. *Fish Shellfish Immunol.* **2002**, *13*, 85–95. [[CrossRef](#)] [[PubMed](#)]
40. Hong, S.; Zou, J.; Collet, B.; Bols, N.C.; Secombes, C.J. Analysis and characterisation of IL-1beta processing in rainbow trout, *Oncorhynchus mykiss*. *Fish Shellfish Immunol.* **2004**, *16*, 453–459. [[CrossRef](#)] [[PubMed](#)]
41. Aachoui, Y.; Leaf, I.A.; Hagar, J.A.; Fontana, M.F.; Campos, C.G.; Zak, D.E.; Tan, M.H.; Cotter, P.A.; Vance, R.E.; Aderem, A.; et al. Caspase-11 protects against bacteria that escape the vacuole. *Science* **2013**, *339*, 975–978. [[CrossRef](#)] [[PubMed](#)]

42. Maelfait, J.; Vercammen, E.; Janssens, S.; Schotte, P.; Haegman, M.; Magez, S.; Beyaert, R. Stimulation of Toll-like receptor 3 and 4 induces interleukin-1beta maturation by caspase-8. *J. Exp. Med.* **2008**, *205*, 1967–1973. [[CrossRef](#)] [[PubMed](#)]
43. Rojas, V.; Camus-Guerra, H.; Guzman, F.; Mercado, L. Pro-inflammatory caspase-1 activation during the immune response in cells from rainbow trout *Oncorhynchus mykiss* (Walbaum 1792) challenged with pathogen-associated molecular patterns. *J. Fish Dis.* **2015**, *38*, 993–1003. [[CrossRef](#)] [[PubMed](#)]
44. Sakata, S.; Yan, Y.; Satou, Y.; Momoi, A.; Ngo-Hazelett, P.; Nozaki, M.; Furutani-Seiki, M.; Postlethwait, J.H.; Yonehara, S.; Sakamaki, K. Conserved function of caspase-8 in apoptosis during bony fish evolution. *Gene* **2007**, *396*, 134–148. [[CrossRef](#)] [[PubMed](#)]
45. Sun, S.; Ge, X.; Zhu, J.; Zhang, W.; Zhang, Q. Molecular cloning, immunohistochemical localization, characterization and expression analysis of caspase-8 from the blunt snout bream (*Megalobrama amblycephala*) exposed to ammonia. *Fish Shellfish Immunol.* **2015**, *47*, 645–654. [[CrossRef](#)] [[PubMed](#)]
46. Alfaidi, M.; Wilson, H.; Daigneault, M.; Burnett, A.; Ridger, V.; Chamberlain, J.; Francis, S. Neutrophil elastase promotes interleukin-1beta secretion from human coronary endothelium. *J. Biol. Chem.* **2015**, *290*, 24067–24078. [[CrossRef](#)] [[PubMed](#)]
47. Terada, K.; Yamada, J.; Hayashi, Y.; Wu, Z.; Uchiyama, Y.; Peters, C.; Nakanishi, H. Involvement of cathepsin B in the processing and secretion of interleukin-1beta in chromogranin A-stimulated microglia. *Glia* **2010**, *58*, 114–124. [[CrossRef](#)] [[PubMed](#)]
48. Hong, S.; Peddie, S.; Campos-Perez, J.J.; Zou, J.; Secombes, C.J. The effect of intraperitoneally administered recombinant IL-1beta on immune parameters and resistance to *Aeromonas salmonicida* in the rainbow trout (*Oncorhynchus mykiss*). *Dev. Comp. Immunol.* **2003**, *27*, 801–812. [[CrossRef](#)]
49. Jiang, S.; Zhang, D.; Li, J.; Liu, Z. Molecular characterization, recombinant expression and bioactivity analysis of the interleukin-1 beta from the yellowfin sea bream, *Acanthopagrus latus* (Houttuyn). *Fish Shellfish Immunol.* **2008**, *24*, 323–336. [[CrossRef](#)] [[PubMed](#)]
50. Kono, T.; Fujiki, K.; Nakao, M.; Yano, T.; Endo, M.; Sakai, M. The immune responses of common carp, *Cyprinus carpio* L., injected with carp interleukin-1beta gene. *J. Interferon Cytokine Res.* **2002**, *22*, 413–419. [[CrossRef](#)] [[PubMed](#)]
51. Lu, D.Q.; Bei, J.X.; Feng, L.N.; Zhang, Y.; Liu, X.C.; Wang, L.; Chen, J.L.; Lin, H.R. Interleukin-1beta gene in orange-spotted grouper, *Epinephelus coioides*: Molecular cloning, expression, biological activities and signal transduction. *Mol. Immunol.* **2008**, *45*, 857–867. [[CrossRef](#)] [[PubMed](#)]
52. Taechavasonyoo, A.; Hirono, I.; Kondo, H. The immune-adjuvant effect of Japanese flounder *Paralichthys olivaceus* IL-1 β . *Dev. Comp. Immunol.* **2013**, *41*, 564–568. [[CrossRef](#)] [[PubMed](#)]
53. Kono, T.; Korenaga, H.; Sakai, M. Genomics of fish IL-17 ligand and receptors: A review. *Fish Shellfish Immunol.* **2011**, *31*, 635–643. [[CrossRef](#)] [[PubMed](#)]
54. Wang, X.; Li, C.; Thongda, W.; Luo, Y.; Beck, B.; Peatman, E. Characterization and mucosal responses of interleukin 17 family ligand and receptor genes in channel catfish *Ictalurus punctatus*. *Fish Shellfish Immunol.* **2014**, *38*, 47–55. [[CrossRef](#)] [[PubMed](#)]
55. Wang, T.; Martin, S.A.; Secombes, C.J. Two interleukin-17C-like genes exist in rainbow trout *Oncorhynchus mykiss* that are differentially expressed and modulated. *Dev. Comp. Immunol.* **2010**, *34*, 491–500. [[CrossRef](#)] [[PubMed](#)]
56. Wang, T.; Jiang, Y.; Wang, A.; Husain, M.; Xu, Q.; Secombes, C.J. Identification of the salmonid IL-17A/F1a/b, IL-17A/F2b, IL-17A/F3 and IL-17N genes and analysis of their expression following *in vitro* stimulation and infection. *Immunogenetics* **2015**, *67*, 395–412. [[CrossRef](#)] [[PubMed](#)]
57. Martin, S.A.; Zou, J.; Houlihan, D.F.; Secombes, C.J. Directional responses following recombinant cytokine stimulation of rainbow trout (*Oncorhynchus mykiss*) RTS-11 macrophage cells as revealed by transcriptome profiling. *BMC Genomics* **2007**. [[CrossRef](#)] [[PubMed](#)]
58. Hong, S.; Li, R.; Xu, Q.; Secombes, C.J.; Wang, T. Two types of TNF- α exist in teleost fish: Phylogeny, expression, and bioactivity analysis of type-II TNF- α 3 in rainbow trout *Oncorhynchus mykiss*. *J. Immunol.* **2013**, *191*, 5959–5972. [[CrossRef](#)] [[PubMed](#)]
59. Castro, R.; Zou, J.; Secombes, C.J.; Martin, S.A. Cortisol modulates the induction of inflammatory gene expression in a rainbow trout macrophage cell line. *Fish Shellfish Immunol.* **2011**, *30*, 215–223. [[CrossRef](#)] [[PubMed](#)]

60. Wang, T.; Kono, T.; Monte, M.M.; Kuse, H.; Costa, M.M.; Korenaga, H.; Maehr, T.; Husain, M.; Sakai, M.; Secombes, C.J. Identification of IL-34 in teleost fish: Differential expression of rainbow trout IL-34, MCSF1 and MCSF2, ligands of the MCSF receptor. *Mol. Immunol.* **2013**, *53*, 398–409. [[CrossRef](#)] [[PubMed](#)]
61. Maehr, T.; Vecino, J.L.; Wadsworth, S.; Wang, T.; Secombes, C.J. Four CISH paralogues are present in rainbow trout *Oncorhynchus mykiss*: Differential expression and modulation during immune responses and development. *Mol. Immunol.* **2014**, *62*, 186–198. [[CrossRef](#)] [[PubMed](#)]
62. Maehr, T.; Costa, M.M.; Vecino, J.L.; Wadsworth, S.; Martin, S.A.; Wang, T.; Secombes, C.J. Transforming growth factor- β 1b: A second TGF- β 1 paralogue in the rainbow trout (*Oncorhynchus mykiss*) that has a lower constitutive expression but is more responsive to immune stimulation. *Fish Shellfish Immunol.* **2013**, *34*, 420–432. [[CrossRef](#)] [[PubMed](#)]
63. Yang, X.; Wei, H.; Qin, S.; Zhang, S.; Wang, X.; Zhang, A.; Du, L. Reciprocal interactions between fish TGF- β 1 and IL-1 β is responsible for restraining IL-1 β signalling activity in grass carp head kidney leukocytes. *Dev. Comp. Immunol.* **2014**, *47*, 197–204. [[CrossRef](#)] [[PubMed](#)]
64. Peddie, S.; Zou, J.; Cunningham, C.; Secombes, C.J. Rainbow trout (*Oncorhynchus mykiss*) recombinant IL-1 β and derived peptides induce migration of head-kidney leucocytes *in vitro*. *Fish Shellfish Immunol.* **2001**, *11*, 697–709. [[CrossRef](#)] [[PubMed](#)]
65. Benedetti, S.; Randelli, E.; Buonocore, F.; Zou, J.; Secombes, C.J.; Scapigliati, G. Evolution of cytokine responses: IL-1 β directly affects intracellular Ca²⁺ concentration of teleost fish leukocytes through a receptor-mediated mechanism. *Cytokine* **2006**, *34*, 9–16. [[CrossRef](#)] [[PubMed](#)]
66. Xu, Q.; Li, R.; Monte, M.M.; Jiang, Y.; Nie, P.; Holland, J.W.; Secombes, C.J.; Wang, T. Sequence and expression analysis of rainbow trout CXCR2, CXCR3a and CXCR3b aids interpretation of lineage-specific conversion, loss and expansion of these receptors during vertebrate evolution. *Dev. Comp. Immunol.* **2014**, *45*, 201–213. [[CrossRef](#)] [[PubMed](#)]
67. Chen, J.; Xu, Q.; Wang, T.; Collet, B.; Corripio-Miyar, Y.; Bird, S.; Xie, P.; Nie, P.; Secombes, C.J.; Zou, J. Phylogenetic analysis of vertebrate CXC chemokines reveals novel lineage specific groups in teleost fish. *Dev. Comp. Immunol.* **2013**, *41*, 137–152. [[CrossRef](#)] [[PubMed](#)]
68. Pooley, N.J.; Tacchi, L.; Secombes, C.J.; Martin, S.A. Inflammatory responses in primary muscle cell cultures in Atlantic salmon (*Salmo salar*). *BMC Genomics* **2013**. [[CrossRef](#)] [[PubMed](#)]
69. Heidari, Z.; Bickerdike, R.; Tinsley, J.; Zou, J.; Wang, T.Y.; Chen, T.Y.; Martin, S.A. Regulatory factors controlling muscle mass: Competition between innate immune function and anabolic signals in regulation of atrogen-1 in Atlantic salmon. *Mol. Immunol.* **2015**, *67*, 341–349. [[CrossRef](#)] [[PubMed](#)]
70. Costa, I.A.; Hein, T.W.; Secombes, C.J.; Gamperl, A.K. Recombinant interleukin-1 β dilates steelhead trout coronary microvessels: Effect of temperature and role of the endothelium, nitric oxide and prostaglandins. *J. Exp. Biol.* **2015**, *218*, 2269–2278. [[CrossRef](#)] [[PubMed](#)]
71. Palomo, J.; Dietrich, D.; Martin, P.; Palmer, G.; Gabay, C. The interleukin (IL)-1 cytokine family—Balance between agonists and antagonists in inflammatory diseases. *Cytokine* **2015**, *76*, 25–37. [[CrossRef](#)] [[PubMed](#)]
72. Subramaniam, S.; Stansberg, C.; Olsen, L.; Zou, J.; Secombes, C.J.; Cunningham, C. Cloning of a *Salmo salar* interleukin-1 receptor-like cDNA. *Dev. Comp. Immunol.* **2002**, *26*, 415–431. [[CrossRef](#)]
73. Stansberg, C.; Subramaniam, S.; Collet, B.; Secombes, C.J.; Cunningham, C. Cloning of the Atlantic salmon (*Salmo salar*) IL-1 receptor associated protein. *Fish Shellfish Immunol.* **2005**, *19*, 53–65. [[CrossRef](#)] [[PubMed](#)]
74. Lu, D.Q.; Yao, M.; Yi, S.B.; Li, Y.W.; Liu, X.C.; Zhang, Y.; Lin, H.R. Soluble interleukin-1 receptor, a potential negative regulator of orange-spotted grouper *Epinephelus coioides* interleukin-1 system. *J. Fish Biol.* **2013**, *83*, 642–658. [[CrossRef](#)] [[PubMed](#)]
75. Lopez-Castejon, G.; Sepulcre, M.P.; Roca, F.J.; Castellana, B.; Planas, J.V.; Meseguer, J.; Mulero, V. The type II interleukin-1 receptor (IL-1RII) of the bony fish gilthead seabream *Sparus aurata* is strongly induced after infection and tightly regulated at transcriptional and post-transcriptional levels. *Mol. Immunol.* **2007**, *44*, 2772–2780. [[CrossRef](#)] [[PubMed](#)]
76. Morrison, R.N.; Young, N.D.; Nowak, B.F. Description of an Atlantic salmon (*Salmo salar* L.) type II interleukin-1 receptor cDNA and analysis of interleukin-1 receptor expression in amoebic gill disease-affected fish. *Fish Shellfish Immunol.* **2012**, *32*, 1185–1190. [[CrossRef](#)] [[PubMed](#)]
77. Sangrador-Vegas, A.; Martin, S.A.; O’Dea, P.G.; Smith, T.J. Cloning and characterization of the rainbow trout (*Oncorhynchus mykiss*) type II interleukin-1 receptor cDNA. *Eur. J. Biochem.* **2000**, *267*, 7031–7037. [[CrossRef](#)] [[PubMed](#)]

78. Yang, X.; Wang, S.; Du, L.; Yang, K.; Wang, X.; Zhang, A.; Zhou, H. Molecular and functional characterization of IL-1 receptor type 2 in grass carp: A potent inhibitor of IL-1 β signaling in head kidney leukocytes. *Dev. Comp. Immunol.* **2013**, *41*, 738–745. [[CrossRef](#)] [[PubMed](#)]
79. Fan, Y.; Li, S.; Qi, J.; Zeng, L.; Zhong, Q.; Zhang, Q. Cloning and characterization of type II interleukin-1 receptor cDNA from Japanese flounder (*Paralichthys olivaceus*). *Comp. Biochem. Physiol. B. Biochem. Mol. Biol.* **2010**, *157*, 59–65. [[CrossRef](#)] [[PubMed](#)]
80. Sigh, J.; Lindstrom, T.; Buchmann, K. Expression of pro-inflammatory cytokines in rainbow trout (*Oncorhynchus mykiss*) during an infection with *Ichthyophthirius multifiliis*. *Fish Shellfish Immunol.* **2004**, *17*, 75–86. [[CrossRef](#)] [[PubMed](#)]
81. Gu, Y.F.; Fang, Y.; Jin, Y.; Dong, W.R.; Xiang, L.X.; Shao, J.Z. Discovery of the DIGIRR gene from teleost fish: A novel Toll-IL-1 receptor family member serving as a negative regulator of IL-1 signaling. *J. Immunol.* **2011**, *187*, 2514–2530. [[CrossRef](#)] [[PubMed](#)]
82. Dinarello, C.A. IL-18: A TH1-inducing, proinflammatory cytokine and new member of the IL-1 family. *J. Allergy Clin. Immunol.* **1999**, *103*, 11–24. [[CrossRef](#)]
83. Okamura, H.; Kashiwamura, S.; Tsutsui, H.; Yoshimoto, T.; Nakanishi, K. Regulation of interferon-gamma production by IL-12 and IL-18. *Curr. Opin. Immunol.* **1998**, *10*, 259–264. [[CrossRef](#)]
84. Perez-Cordon, G.; Estensoro, I.; Benedito-Palos, L.; Calduch-Giner, J.A.; Sitja-Bobadilla, A.; Perez-Sanchez, J. Interleukin gene expression is strongly modulated at the local level in a fish-parasite model. *Fish Shellfish Immunol.* **2014**, *37*, 201–208. [[CrossRef](#)] [[PubMed](#)]
85. Pereiro, P.; Balseiro, P.; Romero, A.; Dios, S.; Forn-Cuni, G.; Fuste, B.; Planas, J.V.; Beltran, S.; Novoa, B.; Figueras, A. High-throughput sequence analysis of turbot (*Scophthalmus maximus*) transcriptome using 454-pyrosequencing for the discovery of antiviral immune genes. *PLoS ONE* **2012**, *7*, e35369. [[CrossRef](#)] [[PubMed](#)]
86. Venkatesh, B.; Lee, A.P.; Ravi, V.; Maurya, A.K.; Lian, M.M.; Swann, J.B.; Ohta, Y.; Flajnik, M.F.; Sutoh, Y.; Kasahara, M.; *et al.* Elephant shark genome provides unique insights into gnathostome evolution. *Nature* **2014**, *505*, 174–179. [[CrossRef](#)] [[PubMed](#)]
87. Meijer, A.H.; Gabby Krens, S.F.; Medina Rodriguez, I.A.; He, S.; Bitter, W.; Ewa Snaar-Jagalska, B.; Spaink, H.P. Expression analysis of the Toll-like receptor and TIR domain adaptor families of zebrafish. *Mol. Immunol.* **2004**, *40*, 773–783. [[CrossRef](#)] [[PubMed](#)]
88. Yao, F.; Yang, X.; Wang, X.; Wei, H.; Zhang, A.; Zhou, H. Molecular and functional characterization of an IL-1 β receptor antagonist in grass carp (*Ctenopharyngodon idella*). *Dev. Comp. Immunol.* **2015**, *49*, 207–216. [[CrossRef](#)] [[PubMed](#)]
89. Aggarwal, B.B. Signalling pathways of the TNF superfamily: A double-edged sword. *Nat. Rev. Immunol.* **2003**, *3*, 745–756. [[CrossRef](#)] [[PubMed](#)]
90. Androlewicz, M.J.; Browning, J.L.; Ware, C.F. Lymphotoxin is expressed as a heteromeric complex with a distinct 33-kDa glycoprotein on the surface of an activated human T cell hybridoma. *J. Biol. Chem.* **1992**, *267*, 2542–2547. [[PubMed](#)]
91. Horiuchi, T.; Mitoma, H.; Harashima, S.; Tsukamoto, H.; Shimoda, T. Transmembrane TNF- α : Structure, function and interaction with anti-TNF agents. *Rheumatology* **2010**, *49*, 1215–1228. [[CrossRef](#)] [[PubMed](#)]
92. Young, J.; Yu, X.; Wolslegel, K.; Nguyen, A.; Kung, C.; Chiang, E.; Kolumam, G.; Wei, N.; Wong, W.L.; DeForge, L.; *et al.* Lymphotoxin- α heterotrimers are cleaved by metalloproteinases and contribute to synovitis in rheumatoid arthritis. *Cytokine* **2010**, *51*, 78–86. [[CrossRef](#)] [[PubMed](#)]
93. Bobe, J.; Goetz, F.W. Molecular cloning and expression of a TNF receptor and two TNF ligands in the fish ovary. *Comp. Biochem. Physiol. B. Biochem. Mol. Biol.* **2001**, *129*, 475–481. [[CrossRef](#)]
94. Covello, J.M.; Bird, S.; Morrison, R.N.; Battaglene, S.C.; Secombes, C.J.; Nowak, B.F. Cloning and expression analysis of three striped trumpeter (*Latris lineata*) pro-inflammatory cytokines, TNF- α , IL-1 β and IL-8, in response to infection by the ectoparasitic, *Chondracanthus goldsmidi*. *Fish Shellfish Immunol.* **2009**, *26*, 773–786. [[CrossRef](#)] [[PubMed](#)]
95. Glenney, G.W.; Wiens, G.D. Early diversification of the TNF superfamily in teleosts: Genomic characterization and expression analysis. *J. Immunol.* **2007**, *178*, 7955–7973. [[CrossRef](#)] [[PubMed](#)]
96. Hirono, I.; Nam, B.H.; Kurobe, T.; Aoki, T. Molecular cloning, characterization, and expression of TNF cDNA and gene from Japanese flounder *Paralichthys olivaceus*. *J. Immunol.* **2000**, *165*, 4423–4427. [[CrossRef](#)] [[PubMed](#)]

97. Laing, K.J.; Wang, T.; Zou, J.; Holland, J.; Hong, S.; Bols, N.; Hirono, I.; Aoki, T.; Secombes, C.J. Cloning and expression analysis of rainbow trout *Oncorhynchus mykiss* tumour necrosis factor-alpha. *Eur. J. Biochem.* **2001**, *268*, 1315–1322. [[CrossRef](#)] [[PubMed](#)]
98. Nascimento, D.S.; Pereira, P.J.; Reis, M.I.; do Vale, A.; Zou, J.; Silva, M.T.; Secombes, C.J.; dos Santos, N.M. Molecular cloning and expression analysis of sea bass (*Dicentrarchus labrax* L.) tumor necrosis factor-alpha (TNF-alpha). *Fish Shellfish Immunol.* **2007**, *23*, 701–710. [[CrossRef](#)] [[PubMed](#)]
99. Pleic, I.L.; Secombes, C.J.; Bird, S.; Mladineo, I. Characterization of three pro-inflammatory cytokines, TNFalpha1, TNFalpha2 and IL-1beta, in cage-reared Atlantic bluefin tuna *Thunnus thynnus*. *Fish Shellfish Immunol.* **2014**, *36*, 98–112. [[CrossRef](#)] [[PubMed](#)]
100. Savan, R.; Kono, T.; Igawa, D.; Sakai, M. A novel tumor necrosis factor (TNF) gene present in tandem with the TNF-alpha gene on the same chromosome in teleosts. *Immunogenetics* **2005**, *57*, 140–150. [[CrossRef](#)] [[PubMed](#)]
101. Uenobe, M.; Kohchi, C.; Yoshioka, N.; Yuasa, A.; Inagawa, H.; Morii, K.; Nishizawa, T.; Takahashi, Y.; Soma, G. Cloning and characterization of a TNF-like protein of *Plecoglossus altivelis* (ayu fish). *Mol. Immunol.* **2007**, *44*, 1115–1122. [[CrossRef](#)] [[PubMed](#)]
102. Xiao, J.; Zhou, Z.C.; Chen, C.; Huo, W.L.; Yin, Z.X.; Weng, S.P.; Chan, S.M.; Yu, X.Q.; He, J.G. Tumor necrosis factor-alpha gene from mandarin fish, *Siniperca chuatsi*: Molecular cloning, cytotoxicity analysis and expression profile. *Mol. Immunol.* **2007**, *44*, 3615–3622. [[CrossRef](#)] [[PubMed](#)]
103. Zhang, A.; Chen, D.; Wei, H.; Du, L.; Zhao, T.; Wang, X.; Zhou, H. Functional characterization of TNF- α in grass carp head kidney leukocytes: Induction and involvement in the regulation of NF- κ B signaling. *Fish Shellfish Immunol.* **2012**, *33*, 1123–1132. [[CrossRef](#)] [[PubMed](#)]
104. Zou, J.; Wang, T.; Hirono, I.; Aoki, T.; Inagawa, H.; Honda, T.; Soma, G.I.; Ototake, M.; Nakanishi, T.; Ellis, A.E.; *et al.* Differential expression of two tumor necrosis factor genes in rainbow trout, *Oncorhynchus mykiss*. *Dev. Comp. Immunol.* **2002**, *26*, 161–172. [[CrossRef](#)]
105. Zou, J.; Peddie, S.; Scapigliati, G.; Zhang, Y.; Bols, N.C.; Ellis, A.E.; Secombes, C.J. Functional characterisation of the recombinant tumor necrosis factors in rainbow trout, *Oncorhynchus mykiss*. *Dev. Comp. Immunol.* **2003**, *27*, 813–822. [[CrossRef](#)]
106. Goetz, F.W.; Planas, J.V.; MacKenzie, S. Tumor necrosis factors. *Dev. Comp. Immunol.* **2004**, *28*, 487–497. [[CrossRef](#)] [[PubMed](#)]
107. Wiens, G.D.; Glenney, G.W. Origin and evolution of TNF and TNF receptor superfamilies. *Dev. Comp. Immunol.* **2011**, *35*, 1324–1335. [[CrossRef](#)] [[PubMed](#)]
108. Kinoshita, S.; Biswas, G.; Kono, T.; Hikima, J.; Sakai, M. Presence of two tumor necrosis factor (tnf)-alpha homologs on different chromosomes of zebrafish (*Danio rerio*) and medaka (*Oryzias latipes*). *Mar. Genomics* **2014**, *13*, 1–9. [[CrossRef](#)] [[PubMed](#)]
109. Kono, T.; Zou, J.; Bird, S.; Savan, R.; Sakai, M.; Secombes, C.J. Identification and expression analysis of lymphotoxin-beta like homologues in rainbow trout *Oncorhynchus mykiss*. *Mol. Immunol.* **2006**, *43*, 1390–1401. [[CrossRef](#)] [[PubMed](#)]
110. Grayfer, L.; Walsh, J.G.; Belosevic, M. Characterization and functional analysis of goldfish (*Carassius auratus* L.) tumor necrosis factor-alpha. *Dev. Comp. Immunol.* **2008**, *32*, 532–543. [[CrossRef](#)] [[PubMed](#)]
111. Morrison, R.N.; Zou, J.; Secombes, C.J.; Scapigliati, G.; Adams, M.B.; Nowak, B.F. Molecular cloning and expression analysis of tumour necrosis factor-alpha in amoebic gill disease (AGD)-affected Atlantic salmon (*Salmo salar* L.). *Fish Shellfish Immunol.* **2007**, *23*, 1015–1031. [[CrossRef](#)] [[PubMed](#)]
112. Saeij, J.P.; Stet, R.J.; de Vries, B.J.; van Muiswinkel, W.B.; Wiegertjes, G.F. Molecular and functional characterization of carp TNF: A link between TNF polymorphism and trypanotolerance? *Dev. Comp. Immunol.* **2003**, *27*, 29–41. [[CrossRef](#)]
113. Savan, R.; Sakai, M. Presence of multiple isoforms of TNF alpha in carp (*Cyprinus carpio* L.): Genomic and expression analysis. *Fish Shellfish Immunol.* **2004**, *17*, 87–94. [[CrossRef](#)] [[PubMed](#)]
114. Benedicenti, O.; Collins, C.; Wang, T.; McCarthy, U.; Secombes, C.J. Which Th pathway is involved during late stage amoebic gill disease? *Fish Shellfish Immunol.* **2015**, *46*, 417–425. [[CrossRef](#)] [[PubMed](#)]
115. Roher, N.; Callol, A.; Planas, J.V.; Goetz, F.W.; MacKenzie, S.A. Endotoxin recognition in fish results in inflammatory cytokine secretion not gene expression. *Innate Immun.* **2011**, *17*, 16–28. [[CrossRef](#)] [[PubMed](#)]
116. Lu, X.J.; Chen, Q.; Yang, G.J.; Chen, J. The TNF α converting enzyme (TACE) from ayu (*Plecoglossus altivelis*) exhibits TNF α shedding activity. *Mol. Immunol.* **2015**, *63*, 497–504. [[CrossRef](#)] [[PubMed](#)]

117. Garcia-Castillo, J.; Chaves-Pozo, E.; Olivares, P.; Pelegrin, P.; Meseguer, J.; Mulero, V. The tumor necrosis factor alpha of the bony fish seabream exhibits the *in vivo* proinflammatory and proliferative activities of its mammalian counterparts, yet it functions in a species-specific manner. *Cell Mol. Life Sci.* **2004**, *61*, 1331–1340. [[CrossRef](#)] [[PubMed](#)]
118. Zou, J.; Secombes, C.J.; Long, S.; Miller, N.; Clem, L.W.; Chinchar, V.G. Molecular identification and expression analysis of tumor necrosis factor in channel catfish (*Ictalurus punctatus*). *Dev. Comp. Immunol.* **2003**, *27*, 845–858. [[CrossRef](#)]
119. Kim, M.S.; Hwang, Y.J.; Yoon, K.J.; Zenke, K.; Nam, Y.K.; Kim, S.K.; Kim, K.H. Molecular cloning of rock bream (*Oplegnathus fasciatus*) tumor necrosis factor-alpha and its effect on the respiratory burst activity of phagocytes. *Fish Shellfish Immunol.* **2009**, *27*, 618–624. [[CrossRef](#)] [[PubMed](#)]
120. Kim, W.L.; Kim, M.S.; Kim, K.H. Molecular cloning of rock bream's (*Oplegnathus fasciatus*) tumor necrosis factor receptor-associated factor 2 and its role in NF- κ B activation. *Fish Shellfish Immunol.* **2011**, *30*, 1178–1183. [[CrossRef](#)] [[PubMed](#)]
121. Clay, H.; Volkman, H.E.; Ramakrishnan, L. Tumor necrosis factor signaling mediates resistance to mycobacteria by inhibiting bacterial growth and macrophage death. *Immunity* **2008**, *29*, 283–294. [[CrossRef](#)] [[PubMed](#)]
122. Roca, F.J.; Ramakrishnan, L. TNF dually mediates resistance and susceptibility to mycobacteria via mitochondrial reactive oxygen species. *Cell* **2013**, *153*, 521–534. [[CrossRef](#)] [[PubMed](#)]
123. Hino, K.; Nakamura, O.; Yoshiura, Y.; Suetake, H.; Suzuki, Y.; Watanabe, T. TNF induces the growth of thymocytes in rainbow trout. *Dev. Comp. Immunol.* **2006**, *30*, 639–647. [[CrossRef](#)] [[PubMed](#)]
124. Roca, F.J.; Mulero, I.; Lopez-Munoz, A.; Sepulcre, M.P.; Renshaw, S.A.; Meseguer, J.; Mulero, V. Evolution of the inflammatory response in vertebrates: Fish TNF-alpha is a powerful activator of endothelial cells but hardly activates phagocytes. *J. Immunol.* **2008**, *181*, 5071–5081. [[CrossRef](#)] [[PubMed](#)]
125. Forlenza, M.; Magez, S.; Scharfack, J.P.; Westphal, A.; Savelkoul, H.F.; Wiegertjes, G.F. Receptor-mediated and lectin-like activities of carp (*Cyprinus carpio*) TNF-alpha. *J. Immunol.* **2009**, *183*, 5319–5332. [[CrossRef](#)] [[PubMed](#)]
126. Boecke, A.; Sieger, D.; Neacsu, C.D.; Kashkar, H.; Kronke, M. Factor associated with neutral sphingomyelinase activity mediates navigational capacity of leukocytes responding to wounds and infection: Live imaging studies in zebrafish larvae. *J. Immunol.* **2012**, *189*, 1559–1566. [[CrossRef](#)] [[PubMed](#)]
127. Praveen, K.; Evans, D.L.; Jaso-Friedmann, L. Constitutive expression of tumor necrosis factor-alpha in cytotoxic cells of teleosts and its role in regulation of cell-mediated cytotoxicity. *Mol. Immunol.* **2006**, *43*, 279–291. [[CrossRef](#)] [[PubMed](#)]
128. Xu, C.; Guo, T.C.; Mutoloki, S.; Haugland, O.; Evensen, O. Gene expression studies of host response to Salmonid alphavirus subtype 3 experimental infections in Atlantic salmon. *Vet. Res.* **2012**. [[CrossRef](#)] [[PubMed](#)]
129. Ronza, P.; Bermudez, R.; Losada, A.P.; Sitja-Bobadilla, A.; Pardo, B.G.; Quiroga, M.I. Immunohistochemical detection and gene expression of TNFalpha in turbot (*Scophthalmus maximus*) enteromyxosis. *Fish Shellfish Immunol.* **2015**, *47*, 368–376. [[CrossRef](#)] [[PubMed](#)]
130. Marjoram, L.; Alvers, A.; Deerkake, M.E.; Bagwell, J.; Mankiewicz, J.; Cocchiari, J.L.; Beerman, R.W.; Willer, J.; Sumigray, K.D.; Katsanis, N.; *et al.* Epigenetic control of intestinal barrier function and inflammation in zebrafish. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 2770–2775. [[CrossRef](#)] [[PubMed](#)]
131. Crespo, D.; Bonnet, E.; Roher, N.; MacKenzie, S.A.; Krasnov, A.; Goetz, F.W.; Bobe, J.; Planas, J.V. Cellular and molecular evidence for a role of tumor necrosis factor alpha in the ovulatory mechanism of trout. *Reprod. Biol. Endocrinol.* **2010**. [[CrossRef](#)] [[PubMed](#)]
132. Crespo, D.; Mananos, E.L.; Roher, N.; MacKenzie, S.A.; Planas, J.V. Tumor necrosis factor alpha may act as an intraovarian mediator of luteinizing hormone-induced oocyte maturation in trout. *Biol. Reprod.* **2012**, *86*, 1–12. [[CrossRef](#)] [[PubMed](#)]
133. Qi, F.; Song, J.; Yang, H.; Gao, W.; Liu, N.A.; Zhang, B.; Lin, S. Mmp23b promotes liver development and hepatocyte proliferation through the tumor necrosis factor pathway in zebrafish. *Hepatology* **2010**, *52*, 2158–2166. [[CrossRef](#)] [[PubMed](#)]
134. Hymowitz, S.G.; Filvaroff, E.H.; Yin, J.P.; Lee, J.; Cai, L.; Risser, P.; Maruoka, M.; Mao, W.; Foster, J.; Kelley, R.F.; *et al.* IL-17s adopt a cystine knot fold: Structure and activity of a novel cytokine, IL-17F, and implications for receptor binding. *EMBO J.* **2001**, *20*, 5332–5341. [[CrossRef](#)] [[PubMed](#)]

135. Parthier, C.; Stelter, M.; Ursel, C.; Fandrich, U.; Lilie, H.; Breithaupt, C.; Stubbs, M.T. Structure of the Toll-Spatzle complex, a molecular hub in *Drosophila* development and innate immunity. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 6281–6286. [[CrossRef](#)] [[PubMed](#)]
136. Gunimaladevi, I.; Savan, R.; Sakai, M. Identification, cloning and characterization of interleukin-17 and its family from zebrafish. *Fish Shellfish Immunol.* **2006**, *21*, 393–403. [[CrossRef](#)] [[PubMed](#)]
137. Du, L.; Feng, S.; Yin, L.; Wang, X.; Zhang, A.; Yang, K.; Zhou, H. Identification and functional characterization of grass carp IL-17A/F1: An evaluation of the immunoregulatory role of teleost IL-17A/F1. *Dev. Comp. Immunol.* **2015**, *51*, 202–211. [[CrossRef](#)] [[PubMed](#)]
138. Monte, M.M.; Wang, T.; Holland, J.W.; Zou, J.; Secombes, C.J. Cloning and characterisation of rainbow trout interleukin-17A/F2 (IL-17A/F2) and IL-17 receptor A: Expression during infection and bioactivity of recombinant IL-17A/F2. *Infect. Immun.* **2013**, *81*, 340–353. [[CrossRef](#)] [[PubMed](#)]
139. Du, L.; Qin, L.; Wang, X.; Zhang, A.; Wei, H.; Zhou, H. Characterization of grass carp (*Ctenopharyngodon idella*) IL-17D: Molecular cloning, functional implication and signal transduction. *Dev. Comp. Immunol.* **2014**, *42*, 220–228. [[CrossRef](#)] [[PubMed](#)]
140. Yoon, S.; Mitra, S.; Wyse, C.; Alnabulsi, A.; Zou, J.; Weerdenburg, E.M.; van der Sar, A.M.; Wang, D.; Secombes, C.J.; Bird, S. First demonstration of antigen induced cytokine expression by CD4-1+ lymphocytes in a poikilotherm: Studies in zebrafish (*Danio rerio*). *PLoS ONE* **2015**, *10*, e0126378. [[CrossRef](#)] [[PubMed](#)]
141. Quintana, F.J.; Iglesias, A.H.; Farez, M.F.; Caccamo, M.; Burns, E.J.; Kassam, N.; Oukka, M.; Weiner, H.L. Adaptive autoimmunity and Foxp3-based immunoregulation in zebrafish. *PLoS ONE* **2010**, *5*, e9478. [[CrossRef](#)] [[PubMed](#)]
142. Funkenstein, B.; Olekh, E.; Jakowlew, S.B. Identification of a novel transforming growth factor-beta (TGF-beta6) gene in fish: Regulation in skeletal muscle by nutritional state. *BMC Mol. Biol.* **2010**. [[CrossRef](#)] [[PubMed](#)]
143. Haddad, G.; Hanington, P.C.; Wilson, E.C.; Grayfer, L.; Belosevic, M. Molecular and functional characterization of goldfish (*Carassius auratus* L.) transforming growth factor beta. *Dev. Comp. Immunol.* **2008**, *32*, 654–663. [[CrossRef](#)] [[PubMed](#)]
144. Wei, H.; Yin, L.; Feng, S.; Wang, X.; Yang, K.; Zhang, A.; Zhou, H. Dual-parallel inhibition of IL-10 and TGF-β1 controls LPS-induced inflammatory response via NF-κB signalling in grass carp monocytes/macrophages. *Fish Shellfish Immunol.* **2015**, *44*, 445–452. [[CrossRef](#)] [[PubMed](#)]
145. Yang, M.; Wang, X.; Chen, D.; Wang, Y.; Zhang, A.; Zhou, H. TGF-β1 exerts opposing effects on grass carp leukocytes: Implications in teleost immunity, receptor signaling and potential self-regulatory mechanisms. *PLoS ONE* **2012**, *7*, e35011. [[CrossRef](#)] [[PubMed](#)]
146. Yang, M.; Zhou, H. Grass carp transforming growth factor-β1 (TGF-β1): Molecular cloning, tissue distribution and immunobiological activity in teleost peripheral blood lymphocytes. *Mol. Immunol.* **2008**, *45*, 1792–1798. [[CrossRef](#)] [[PubMed](#)]
147. Xu, Q.; Cai, C.; Hu, X.; Liu, Y.; Guo, Y.; Hu, P.; Chen, Z.; Peng, S.; Zhang, D.; Jiang, S.; *et al.* Evolutionary suppression of erythropoiesis via the modulation of TGF-β signalling in an Antarctic icefish. *Mol. Ecol.* **2015**, *24*, 4664–4678. [[CrossRef](#)] [[PubMed](#)]
148. Zhan, X.L.; Ma, T.Y.; Wu, J.Y.; Yi, L.Y.; Wang, J.Y.; Gao, X.K. Cloning and primary immunological study of TGF-β1 and its receptors TβR I/TβR II in tilapia (*Oreochromis niloticus*). *Dev. Comp. Immunol.* **2015**, *51*, 134–140. [[CrossRef](#)] [[PubMed](#)]
149. Vosshenrich, C.A.J.; Di Santo, J.P. Interleukin signalling. *Curr. Biol.* **2002**, *12*, 760–763. [[CrossRef](#)]
150. Ohtani, M.; Hayashi, N.; Hashimoto, K.; Nakanishi, T.; Dijkstra, J.M. Comprehensive clarification of two paralogous interleukin 4/13 loci in teleost fish. *Immunogenetics* **2008**, *60*, 383–397. [[CrossRef](#)] [[PubMed](#)]
151. Wang, T.; Secombes, C.J. The evolution of IL-4 and IL-13 and their receptor subunits. *Cytokine* **2015**, *75*, 8–13. [[CrossRef](#)] [[PubMed](#)]
152. Díaz-Rosales, P.; Bird, S.; Wang, T.H.; Fujiki, K.; Davidson, W.S.; Zou, J.; Secombes, C.J. Rainbow trout interleukin-2: Cloning, expression and bioactivity analysis. *Fish Shellfish Immunol.* **2009**, *27*, 414–422. [[CrossRef](#)] [[PubMed](#)]
153. Wang, T.; Johansson, P.; Abós, B.; Holt, A.; Tafalla, C.; Jiang, Y.; Wang, A.; Xu, Q.; Qi, Z.; Huang, W.; *et al.* First in-depth analysis of the novel Th2-type cytokines in salmonid fish reveals distinct patterns of expression and modulation but overlapping bioactivities. *Oncotarget* **2016**, *7*, 10917–10946. [[PubMed](#)]

154. Corripio-Miyar, Y.; Secombes, C.J.; Zou, J. Long-term stimulation of trout head kidney cells with the cytokines MCSF, IL-2 and IL-6: Gene expression dynamics. *Fish Shellfish Immunol.* **2012**, *32*, 35–44. [[CrossRef](#)] [[PubMed](#)]
155. Lin, A.F.; Xiang, L.X.; Wang, Q.L.; Dong, W.R.; Gong, Y.F.; Shao, J.Z. The DC-SIGN of zebrafish: Insights into the existence of a CD209 homologue in a lower vertebrate and its involvement in adaptive immunity. *J. Immunol.* **2009**, *183*, 7398–7410. [[CrossRef](#)] [[PubMed](#)]
156. Hu, Y.L.; Xiang, L.X.; Shao, J.Z. Identification and characterization of a novel immunoglobulin Z isotype in zebrafish: Implications for a distinct B cell receptor in lower vertebrates. *Mol. Immunol.* **2010**, *47*, 738–746. [[CrossRef](#)] [[PubMed](#)]
157. Zhu, L.Y.; Pan, P.P.; Fang, W.; Shao, J.Z.; Xiang, L.X. Essential role of IL-4 and IL-4R α in interaction in adaptive immunity of zebrafish: Insight into the origin of Th2-like regulatory mechanisms in ancient vertebrates. *J. Immunol.* **2012**, *188*, 5571–5584. [[CrossRef](#)] [[PubMed](#)]
158. Yamaguchi, T.; Miyata, S.; Katakura, F.; Nagasawa, T.; Shibasaki, Y.; Yabu, T.; Fischer, U.; Nakayasu, C.; Nakanishi, T.; Moritomo, T. Recombinant carp IL-4/13B stimulates *in vitro* proliferation of carp IgM+ B cells. *Fish Shellfish Immunol.* **2016**, *49*, 225–229. [[CrossRef](#)] [[PubMed](#)]
159. Kono, T.; Bird, S.; Sonoda, K.; Savan, R.; Secombes, C.J.; Sakai, M. Characterization and expression analysis of an interleukin-7 homologue in the Japanese pufferfish, *Takifugu rubripes*. *FEBS J.* **2008**, *275*, 1213–1226. [[CrossRef](#)] [[PubMed](#)]
160. Iwanami, N.; Mateos, F.; Hess, I.; Riffel, N.; Soza-Ried, C.; Schorpp, M.; Boehm, T. Genetic evidence for an evolutionarily conserved role of IL-7 signaling in T cell development in zebrafish. *J. Immunol.* **2011**, *186*, 7060–7066. [[CrossRef](#)] [[PubMed](#)]
161. Wang, T.; Holland, J.W.; Carrington, A.; Zou, J.; Secombes, C.J. Molecular and functional characterisation of interleukin-15 in rainbow trout *Oncorhynchus mykiss*: A potent inducer of interferon-gamma expression in spleen leucocytes. *J. Immunol.* **2007**, *179*, 1475–1488. [[CrossRef](#)] [[PubMed](#)]
162. Wang, T.H.; Husain, M.; Hong, S.; Holland, J.W. Differential expression, modulation and bioactivity of distinct fish IL-12 isoforms: Implication towards the evolution of Th1-like immune responses. *Eur. J. Immunol.* **2014**, *44*, 1541–1551. [[CrossRef](#)] [[PubMed](#)]
163. Bei, J.X.; Suetake, H.; Araki, K.; Kikuchi, K.; Yoshiura, Y.; Lin, H.R.; Suzuki, Y. Two interleukin (IL)-15 homologues in fish from two distinct origins. *Mol. Immunol.* **2006**, *43*, 860–869. [[CrossRef](#)] [[PubMed](#)]
164. Fang, W.; Xiang, L.X.; Shao, J.Z.; Wen, Y.; Chen, S.Y. Identification and characterization of an interleukin-15 homologue from *Tetraodon nigroviridis*. *Comp. Biochem. Physiol. B. Biochem. Mol. Biol.* **2006**, *143*, 335–343. [[CrossRef](#)] [[PubMed](#)]
165. Dijkstra, J.M.; Takizawa, F.; Fischer, U.; Friedrich, M.; Soto-Lampe, V.; Lefèvre, C.; Lenk, M.; Karger, A.; Matsui, T.; Hashimoto, K. Identification of a gene for an ancient cytokine, interleukin 15-like, in mammals; interleukins 2 and 15 co-evolved with this third family member, all sharing binding motifs for IL-15R α . *Immunogenetics* **2014**, *66*, 93–103. [[CrossRef](#)] [[PubMed](#)]
166. Wang, T.; Diaz-Rosales, P.; Costa, M.M.; Campbell, S.; Snow, M.; Collet, B.; Martin, S.A.M.; Secombes, C.J. Functional characterisation of a nonmammalian IL-21: Rainbow trout *Oncorhynchus mykiss* IL-21 upregulates the expression of the Th cell signature cytokines IFN-g, IL-10 and IL-22. *J. Immunol.* **2011**, *186*, 708–721. [[CrossRef](#)] [[PubMed](#)]
167. Sertori, R.; Liongue, C.; Basheer, F.; Lewis, K.L.; Rasighaemi, P.; de Coninck, D.; Traver, D.; Ward, A.C. Conserved IL-2R γ signaling mediates lymphopoiesis in zebrafish. *J. Immunol.* **2016**, *196*, 135–143. [[CrossRef](#)] [[PubMed](#)]
168. Wang, T.; Huang, W.; Costa, M.M.; Secombes, C.J. The gamma-chain cytokine/receptor system in fish: More ligands and receptors. *Fish Shellfish Immunol.* **2011**, *31*, 673–687. [[CrossRef](#)] [[PubMed](#)]
169. Wen, Y.; Fang, W.; Xiang, L.X.; Pan, R.L.; Shao, J.Z. Identification of Treg-like cells in *Tetraodon*: Insight into the origin of regulatory T subsets during early vertebrate evolution. *Cell. Mol. Life Sci.* **2011**, *68*, 2615–2626. [[CrossRef](#)] [[PubMed](#)]
170. Liongue, C.; Ward, A.C. Evolution of class I cytokine receptors. *BMC Evol. Biol.* **2007**. [[CrossRef](#)] [[PubMed](#)]
171. Dijkstra, J.M. Th2 and Treg candidate genes in elephant shark. *Nature* **2014**, *511*, 7–9. [[CrossRef](#)] [[PubMed](#)]
172. Yamaguchi, T.; Takizawa, F.; Fischer, U.; Dijkstra, J.M. Along the axis between type I and type 2 immunity: Principles conserved in evolution from fish to mammals. *Biology* **2015**, *4*, 814–859. [[CrossRef](#)] [[PubMed](#)]

173. Fujiki, K.; Nakao, M.; Dixon, B. Molecular cloning and characterisation of a carp (*Cyprinus carpio*) cytokine-like cDNA that shares sequence similarity with IL-6 subfamily cytokines CNTF, OSM and LIF. *Dev. Comp. Immunol.* **2003**, *27*, 127–136. [[CrossRef](#)]
174. Wang, T.; Secombes, C.J. Identification and expression analysis of two fish-specific IL-6 cytokine family members, the ciliary neurotrophic factor (CNTF)-like and M17 genes, in rainbow trout *Oncorhynchus mykiss*. *Mol. Immunol.* **2009**, *46*, 2290–2298. [[CrossRef](#)] [[PubMed](#)]
175. Huising, M.O.; Kruiswijk, C.P.; van Schijndel, J.E.; Savelkoul, H.F.; Flik, G.; Verburg-van Kemenade, B.M. Multiple and highly divergent IL-11 genes in teleost fish. *Immunogenetics* **2005**, *57*, 432–443. [[CrossRef](#)] [[PubMed](#)]
176. Santos, M.D.; Yasuike, M.; Kondo, H.; Hirono, I.; Aoki, T. Teleostean IL11b exhibits complementing function to IL11a and expansive involvement in antibacterial and antiviral responses. *Mol. Immunol.* **2008**, *45*, 3494–3501. [[CrossRef](#)] [[PubMed](#)]
177. Costa, M.M.; Maehr, T.; Diaz-Rosales, P.; Secombes, C.J.; Wang, T. Bioactivity studies of rainbow trout (*Oncorhynchus mykiss*) interleukin-6: Effects on macrophage growth and antimicrobial peptide gene expression. *Mol. Immunol.* **2011**, *48*, 1903–1916. [[CrossRef](#)] [[PubMed](#)]
178. Chen, H.H.; Lin, H.T.; Fong, Y.F.; Lin, J.H. The bioactivity of teleost IL-6: IL-6 protein in orange-spotted grouper (*Epinephelus coioides*) induces Th2 cell differentiation and antibody production. *Dev. Comp. Immunol.* **2012**, *38*, 285–294. [[CrossRef](#)] [[PubMed](#)]
179. Kaneda, M.; Odaka, T.; Suetake, H.; Tahara, D.; Miyadai, T. Teleost IL-6 promotes antibody production through STAT3 signaling via IL-6R and gp130. *Dev. Comp. Immunol.* **2012**, *38*, 224–231. [[CrossRef](#)] [[PubMed](#)]
180. Hanington, P.C.; Belosevic, M. Interleukin-6 family cytokine M17 induces differentiation and nitric oxide response of goldfish (*Carassius auratus* L.) macrophages. *Dev. Comp. Immunol.* **2007**, *31*, 817–829. [[CrossRef](#)] [[PubMed](#)]
181. Ogai, K.; Kuwana, A.; Hisano, S.; Nagashima, M.; Koriyama, Y.; Sugitani, K.; Mawatari, K.; Nakashima, H.; Kato, S. Upregulation of leukemia inhibitory factor (LIF) during the early stage of optic nerve regeneration in zebrafish. *PLoS ONE* **2014**, *9*, e106010. [[CrossRef](#)] [[PubMed](#)]
182. Collison, L.W.; Delgoffe, G.M.; Guy, C.S.; Vignali, K.M.; Chaturvedi, V.; Fairweather, D.; Satoskar, A.R.; Garcia, K.C.; Hunter, C.A.; Drake, C.G.; *et al.* The composition and signalling of the IL-35 receptor are unconventional. *Nat. Immunol.* **2012**, *13*, 290–299. [[CrossRef](#)] [[PubMed](#)]
183. Heinzl, F.P.; Hujer, A.M.; Ahmed, F.N.; Kerko, R.M. *Vivo* production and function of IL-12 p40 homodimers. *J. Immunol.* **1997**, *158*, 4381–4388. [[PubMed](#)]
184. Zhang, L.; Zhang, B.C.; Hu, Y.H. Rock bream (*Oplegnathus fasciatus*) IL-12p40: Identification, expression, and effect on bacterial infection. *Fish Shellfish Immunol.* **2014**, *39*, 312–320. [[CrossRef](#)] [[PubMed](#)]
185. Tsai, J.L.; Priya, T.A.J.; Hu, K.Y.; Yan, H.Y.; Shen, S.T.; Song, Y.L. Grouper interleukin-12, linked by an ancient disulphide-bond architecture, exhibits cytokine and chemokine activities. *Fish Shellfish Immunol.* **2014**, *36*, 27–37. [[CrossRef](#)] [[PubMed](#)]
186. Segaliny, A.I.; Brion, R.; Brulin, B.; Maillason, M.; Charrier, C.; Teletchea, S.; Heymann, D. IL-34 and M-CSF form a novel heteromeric cytokine and regulate M-CSF receptor activation and localization. *Cytokine* **2015**, *76*, 170–181. [[CrossRef](#)] [[PubMed](#)]
187. Hanington, P.C.; Wang, T.; Secombes, C.J.; Belosevic, M. Growth factors of lower vertebrates: Characterisation of goldfish (*Carassius auratus* L.) macrophage colony stimulating factor-1. *J. Biol. Chem.* **2007**, *282*, 31865–31872. [[CrossRef](#)] [[PubMed](#)]
188. Barreda, D.R.; Hanington, P.C.; Stafford, J.L.; Belosevic, M. A novel soluble form of the CSF-1 receptor inhibits proliferation of self-renewing macrophages of goldfish (*Carassius auratus* L.). *Dev. Comp. Immunol.* **2005**, *29*, 879–894. [[CrossRef](#)] [[PubMed](#)]
189. Grayfer, L.; Hanington, P.C.; Belosevic, M. Macrophage colony-stimulating factor (CSF-1) induces pro-inflammatory gene expression and enhances antimicrobial responses of goldfish (*Carassius auratus* L.) macrophages. *Fish Shellfish Immunol.* **2009**, *26*, 406–413. [[CrossRef](#)] [[PubMed](#)]
190. Hanington, P.C.; Hitchen, S.J.; Beamish, L.A.; Belosevic, M. Macrophage colony stimulating factor (CSF-1) is a central growth factor of goldfish macrophages. *Fish Shellfish Immunol.* **2009**, *26*, 1–9. [[CrossRef](#)] [[PubMed](#)]
191. Wang, T.; Hanington, P.C.; Belosevic, M.; Secombes, C.J. Two macrophage colony-stimulating factor genes exist in fish that differ in gene organisation and are differentially expressed. *J. Immunol.* **2008**, *181*, 3310–3322. [[CrossRef](#)] [[PubMed](#)]

192. Wang, T.; Kono, T.; Monte, M.M.; Kuse, H.; Costa, M.M.; Korenaga, H.; Maehr, T.; Husain, M.; Sakai, M.; Secombes, C.J. Identification of IL-34 in teleost fish: Differential expression of rainbow trout IL-34, MCSF1 and MCSF2, ligands of the MCSF receptor. *Mol. Immunol.* **2013**, *53*, 398–409. [[CrossRef](#)] [[PubMed](#)]
193. Mo, Z.Q.; Li, Y.W.; Zhou, L.; Li, A.X.; Luo, X.C.; Dan, X.M. Grouper (*Epinephelus coioides*) IL-34/MCSF2 and MSCFR1/MCSFR2 were involved in mononuclear phagocytes activation against *Cryptocaryon irritans* infection. *Fish Shellfish Immunol.* **2015**, *43*, 142–149. [[CrossRef](#)] [[PubMed](#)]
194. Wang, Y.; Szretter, K.J.; Vermi, W.; Gilfillan, S.; Rossini, C.; Cella, M.; Barrow, A.D.; Diamond, M.S.; Colonna, M. IL-34 is a tissue-restricted ligand of CSF1R required for the development of Langerhans cells and microglia. *Nat. Immunol.* **2012**, *13*, 753–760. [[CrossRef](#)] [[PubMed](#)]
195. Liongue, C.; Hall, C.J.; O’Connell, B.A.; Crosier, P.; Ward, A.C. Zebrafish granulocyte colony-stimulating factor receptor signalling promotes myelopoiesis and myeloid cell migration. *Blood* **2009**, *113*, 2535–2546. [[CrossRef](#)] [[PubMed](#)]
196. Svoboda, O.; Stachura, D.L.; Machoňová, O.; Pajer, P.; Brynda, J.; Zon, L.I.; Traver, D.; Bartůněk, P. Dissection of vertebrate hematopoiesis using zebrafish thrombopoietin. *Blood* **2014**, *124*, 220–228. [[CrossRef](#)] [[PubMed](#)]
197. Stachura, D.L.; Svoboda, O.; Campbell, C.A.; Espin-Palazón, R.; Lau, R.P.; Zon, L.I.; Bartůněk, P.; Traver, D. The zebrafish granulocyte colony-stimulating factors (Gcsfs): 2 paralogous cytokines and their roles in hematopoietic development and maintenance. *Blood* **2013**, *122*, 3918–3928. [[CrossRef](#)] [[PubMed](#)]
198. Galdames, J.A.; Zuñiga-Traslaviña, C.; Reyes, A.E.; Feijóo, C.G. Gcsf-Chr19 promotes neutrophil migration to damaged tissue through blood vessels in zebrafish. *J. Immunol.* **2014**, *193*, 372–378. [[CrossRef](#)] [[PubMed](#)]
199. Niethammer, P.; Grabher, C.; Look, A.T.; Mitchison, T.J. A tissue-scale gradient of hydrogen peroxide mediates rapid wound detection in zebrafish. *Nature* **2009**, *459*, 996–999. [[CrossRef](#)] [[PubMed](#)]
200. Lutfalla, G.; Roest Crolius, H.; Stange-Thomann, N.; Jaillon, O.; Mogensen, K.; Monneron, D. Comparative genomic analysis reveals independent expansion of a lineage-specific gene family in vertebrates: The class II cytokine receptors and their ligands in mammals and fish. *BMC Genomics* **2003**. [[CrossRef](#)] [[PubMed](#)]
201. Altmann, S.M.; Mellon, M.T.; Distel, D.L.; Kim, C.H. Molecular and functional analysis of an interferon gene from the zebrafish, *Danio rerio*. *J. Virol.* **2003**, *77*, 1992–2002. [[CrossRef](#)] [[PubMed](#)]
202. Boudinot, P.; Zou, J.; Ota, T.; Buonocore, F.; Scapigliati, G.; Canapa, A.; Cannon, J.; Litman, G.; Hansen, J.D. A tetrapod-like repertoire of innate immune receptors and effectors for coelacanth. *J. Exp. Zool. B. Mol. Dev. Evol.* **2014**, *322*, 415–437. [[CrossRef](#)] [[PubMed](#)]
203. Langevin, C.; Aleksejeva, E.; Passoni, G.; Palha, N.; Levraud, J.P.; Boudinot, P. The antiviral innate immune response in fish: Evolution and conservation of the IFN system. *J. Mol. Biol.* **2013**, *425*, 4904–4920. [[CrossRef](#)] [[PubMed](#)]
204. Robertsen, B. The interferon system of teleost fish. *Fish Shellfish Immunol.* **2006**, *20*, 172–191. [[CrossRef](#)] [[PubMed](#)]
205. Zhang, Y.B.; Gui, J.F. Molecular regulation of interferon antiviral response in fish. *Dev. Comp. Immunol.* **2012**, *38*, 193–202. [[CrossRef](#)] [[PubMed](#)]
206. Zou, J.; Secombes, C.J. Teleost fish interferons and their role in immunity. *Dev. Comp. Immunol.* **2011**, *35*, 1376–1387. [[CrossRef](#)] [[PubMed](#)]
207. Aggad, D.; Mazel, M.; Boudinot, P.; Mogensen, K.E.; Hamming, O.J.; Hartmann, R.; Kotenko, S.; Herbomel, P.; Lutfalla, G.; Levraud, J.P. The two groups of zebrafish virus-induced interferons signal via distinct receptors with specific and shared chains. *J. Immunol.* **2009**, *183*, 3924–3931. [[CrossRef](#)] [[PubMed](#)]
208. Chang, M.; Nie, P.; Collet, B.; Secombes, C.J.; Zou, J. Identification of an additional two-cysteine containing type I interferon in rainbow trout *Oncorhynchus mykiss* provides evidence of a major gene duplication event within this gene family in teleosts. *Immunogenetics* **2009**, *61*, 315–325. [[CrossRef](#)] [[PubMed](#)]
209. Sun, B.; Robertsen, B.; Wang, Z.; Liu, B. Identification of an Atlantic salmon IFN multigene cluster encoding three IFN subtypes with very different expression properties. *Dev. Comp. Immunol.* **2009**, *33*, 547–558. [[CrossRef](#)] [[PubMed](#)]
210. Zou, J.; Tafalla, C.; Truckle, J.; Secombes, C.J. Identification of a second group of type I IFNs in fish sheds light on IFN evolution in vertebrates. *J. Immunol.* **2007**, *179*, 3859–3871. [[CrossRef](#)] [[PubMed](#)]
211. Zou, J.; Gorgoglione, B.; Taylor, N.G.; Summated, T.; Lee, P.T.; Panigrahi, A.; Genet, C.; Chen, Y.M.; Chen, T.Y.; Ul Hassan, M.; *et al.* Salmonids have an extraordinary complex type I IFN system: Characterization of the IFN locus in rainbow trout *Oncorhynchus mykiss* reveals two novel IFN subgroups. *J. Immunol.* **2014**, *193*, 2273–2286. [[CrossRef](#)] [[PubMed](#)]

212. Hamming, O.J.; Lutfalla, G.; Levraud, J.P.; Hartmann, R. Crystal structure of zebrafish interferons I and II reveals conservation of type I interferon structure in vertebrates. *J. Virol.* **2011**, *85*, 8181–8187. [[CrossRef](#)] [[PubMed](#)]
213. Casani, D.; Randelli, E.; Costantini, S.; Facchiano, A.M.; Zou, J.; Martin, S.; Secombes, C.J.; Scapigliati, G.; Buonocore, F. Molecular characterisation and structural analysis of an interferon homologue in sea bass (*Dicentrarchus labrax* L.). *Mol. Immunol.* **2009**, *46*, 943–952. [[CrossRef](#)] [[PubMed](#)]
214. Chen, Y.M.; Kuo, C.E.; Chen, G.R.; Kao, Y.T.; Zou, J.; Secombes, C.J.; Chen, T.Y. Functional analysis of an orange-spotted grouper (*Epinephelus coioides*) interferon gene and characterisation of its expression in response to nodavirus infection. *Dev. Comp. Immunol.* **2014**, *46*, 117–128. [[CrossRef](#)] [[PubMed](#)]
215. Ohtani, M.; Hikima, J.; Hwang, S.D.; Morita, T.; Suzuki, Y.; Kato, G.; Kondo, H.; Hirono, I.; Jung, T.S.; Aoki, T. Transcriptional regulation of type I interferon gene expression by interferon regulatory factor-3 in Japanese flounder, *Paralichthys olivaceus*. *Dev. Comp. Immunol.* **2012**, *36*, 697–706. [[CrossRef](#)] [[PubMed](#)]
216. Long, S.; Milev-Milovanovic, I.; Wilson, M.; Bengten, E.; Clem, L.W.; Miller, N.W.; Chinchar, V.G. Identification and expression analysis of cDNAs encoding channel catfish type I interferons. *Fish Shellfish Immunol.* **2006**, *21*, 42–59. [[CrossRef](#)] [[PubMed](#)]
217. Long, S.; Wilson, M.; Bengten, E.; Bryan, L.; Clem, L.W.; Miller, N.W.; Chinchar, V.G. Identification of a cDNA encoding channel catfish interferon. *Dev. Comp. Immunol.* **2004**, *28*, 97–111. [[CrossRef](#)]
218. Robertsen, B.; Bergan, V.; Rokenes, T.; Larsen, R.; Albuquerque, A. Atlantic salmon interferon genes: Cloning, sequence analysis, expression, and biological activity. *J. Interferon Cytokine Res.* **2003**, *23*, 601–612. [[CrossRef](#)] [[PubMed](#)]
219. Maekawa, S.; Chiang, Y.A.; Hikima, J.; Sakai, M.; Lo, C.F.; Wang, H.C.; Aoki, T. Expression and biological activity of two types of interferon genes in medaka (*Oryzias latipes*). *Fish Shellfish Immunol.* **2016**, *48*, 20–29. [[CrossRef](#)] [[PubMed](#)]
220. Pereira, P.; Costa, M.M.; Diaz-Rosales, P.; Dios, S.; Figueras, A.; Novoa, B. The first characterization of two type I interferons in turbot (*Scophthalmus maximus*) reveals their differential role, expression pattern and gene induction. *Dev. Comp. Immunol.* **2014**, *45*, 233–244. [[CrossRef](#)] [[PubMed](#)]
221. Svingerud, T.; Solstad, T.; Sun, B.; Nyrud, M.L.; Kileng, O.; Greiner-Tollersrud, L.; Robertsen, B. Atlantic salmon type I IFN subtypes show differences in antiviral activity and cell-dependent expression: Evidence for high IFNb/IFNc-producing cells in fish lymphoid tissues. *J. Immunol.* **2012**, *189*, 5912–5923. [[CrossRef](#)] [[PubMed](#)]
222. Zhang, Y.B.; Li, Q.; Gui, J.F. Differential expression of two *Carassius auratus* Mx genes in cultured CAB cells induced by grass carp hemorrhage virus and interferon. *Immunogenetics* **2004**, *56*, 68–75. [[CrossRef](#)] [[PubMed](#)]
223. Chang, C.J.; Robertsen, C.; Sun, B.; Robertsen, B. Protection of Atlantic salmon against virus infection by intramuscular injection of IFNc expression plasmid. *Vaccine* **2014**, *32*, 4695–4702. [[CrossRef](#)] [[PubMed](#)]
224. Li, S.; Lu, L.F.; Feng, H.; Wu, N.; Chen, D.D.; Zhang, Y.B.; Gui, J.F.; Nie, P.; Zhang, Y.A. IFN regulatory factor 10 is a negative regulator of the IFN responses in fish. *J. Immunol.* **2014**, *193*, 1100–1109. [[CrossRef](#)] [[PubMed](#)]
225. Sun, F.; Zhang, Y.B.; Liu, T.K.; Gan, L.; Yu, F.F.; Liu, Y.; Gui, J.F. Characterization of fish IRF3 as an IFN-inducible protein reveals evolving regulation of IFN response in vertebrates. *J. Immunol.* **2010**, *185*, 7573–7582. [[CrossRef](#)] [[PubMed](#)]
226. Sun, F.; Zhang, Y.B.; Jiang, J.; Wang, B.; Chen, C.; Zhang, J.; Gui, J.F. Gig1, a novel antiviral effector involved in fish interferon response. *Virology* **2014**, *448*, 322–332. [[CrossRef](#)] [[PubMed](#)]
227. Chang, M.; Collet, B.; Nie, P.; Lester, K.; Campbell, S.; Secombes, C.J.; Zou, J. Expression and functional characterization of the RIG-I-like receptors MDA5 and LGP2 in rainbow trout (*Oncorhynchus mykiss*). *J. Virol.* **2011**, *85*, 8403–8412. [[CrossRef](#)] [[PubMed](#)]
228. Martin, S.A.; Taggart, J.B.; Seear, P.; Bron, J.E.; Talbot, R.; Teale, A.J.; Sweeney, G.E.; Hoyheim, B.; Houlihan, D.F.; Tocher, D.R.; *et al.* Interferon type I and type II responses in an Atlantic salmon (*Salmo salar*) SHK-1 cell line by the salmon TRAILS/SGP microarray. *Physiol. Genomics* **2007**, *32*, 33–44. [[CrossRef](#)] [[PubMed](#)]
229. Xu, C.; Evensen, O.; Munang'andu, H.M. De novo assembly and transcriptome analysis of Atlantic salmon macrophage/dendritic-like TO cells following type I IFN treatment and Salmonid alphavirus subtype-3 infection. *BMC Genomics* **2015**. [[CrossRef](#)] [[PubMed](#)]

230. Huang, Z.; Chen, S.; Liu, J.; Xiao, J.; Yan, J.; Feng, H. IFN α of black carp is an antiviral cytokine modified with N-linked glycosylation. *Fish Shellfish Immunol.* **2015**, *46*, 477–485. [[CrossRef](#)] [[PubMed](#)]
231. Kileng, O.; Brundtland, M.I.; Robertsen, B. Infectious salmon anemia virus is a powerful inducer of key genes of the type I interferon system of Atlantic salmon, but is not inhibited by interferon. *Fish Shellfish Immunol.* **2007**, *23*, 378–389. [[CrossRef](#)] [[PubMed](#)]
232. Sun, B.; Skjaveland, I.; Svingerud, T.; Zou, J.; Jorgensen, J.; Robertsen, B. Antiviral activity of salmonid gamma interferon against infectious pancreatic necrosis virus and salmonid alphavirus and its dependency on type I interferon. *J. Virol.* **2011**, *85*, 9188–9198. [[CrossRef](#)] [[PubMed](#)]
233. Svingerud, T.; Holand, J.K.; Robertsen, B. Infectious salmon anemia virus (ISAV) replication is transiently inhibited by Atlantic salmon type I interferon in cell culture. *Virus Res.* **2013**, *177*, 163–170. [[CrossRef](#)] [[PubMed](#)]
234. Xu, C.; Guo, T.C.; Mutoloki, S.; Haugland, O.; Marjara, I.S.; Evensen, O. Alpha interferon and not gamma interferon inhibits salmonid alphavirus subtype 3 replication *in vitro*. *J. Virol.* **2010**, *84*, 8903–8912. [[CrossRef](#)] [[PubMed](#)]
235. Lopez-Munoz, A.; Roca, F.J.; Meseguer, J.; Mulero, V. New insights into the evolution of IFNs: Zebrafish group II IFNs induce a rapid and transient expression of IFN-dependent genes and display powerful antiviral activities. *J. Immunol.* **2009**, *182*, 3440–3449. [[CrossRef](#)] [[PubMed](#)]
236. Bergan, V.; Steinsvik, S.; Xu, H.; Kileng, O.; Robertsen, B. Promoters of type I interferon genes from Atlantic salmon contain two main regulatory regions. *FEBS J.* **2006**, *273*, 3893–3906. [[CrossRef](#)] [[PubMed](#)]
237. Purcell, M.K.; Laing, K.J.; Woodson, J.C.; Thorgaard, G.H.; Hansen, J.D. Characterization of the interferon genes in homozygous rainbow trout reveals two novel genes, alternate splicing and differential regulation of duplicated genes. *Fish Shellfish Immunol.* **2009**, *26*, 293–304. [[CrossRef](#)] [[PubMed](#)]
238. Chang, M.X.; Zou, J.; Nie, P.; Huang, B.; Yu, Z.; Collet, B.; Secombes, C.J. Intracellular interferons in fish: A unique means to combat viral infection. *PLoS Pathog.* **2013**. [[CrossRef](#)] [[PubMed](#)]
239. Kuo, H.P.; Chung, C.L.; Hung, Y.F.; Lai, Y.S.; Chiou, P.P.; Lu, M.W.; Kong, Z.L. Comparison of the responses of different recombinant fish type I interferons against betanodavirus infection in grouper. *Fish Shellfish Immunol.* **2016**, *49*, 143–153. [[CrossRef](#)] [[PubMed](#)]
240. Sun, B.; Greiner-Tollersrud, L.; Koop, B.F.; Robertsen, B. Atlantic salmon possesses two clusters of type I interferon receptor genes on different chromosomes, which allows for a larger repertoire of interferon receptors than in zebrafish and mammals. *Dev. Comp. Immunol.* **2014**, *47*, 275–286. [[CrossRef](#)] [[PubMed](#)]
241. Hou, S.X.; Zheng, Z.; Chen, X.; Perrimon, N. The Jak/STAT pathway in model organisms: Emerging roles in cell movement. *Dev. Cell* **2002**, *3*, 765–778. [[CrossRef](#)]
242. Stein, C.; Caccamo, M.; Laird, G.; Leptin, M. Conservation and divergence of gene families encoding components of innate immune response systems in zebrafish. *Genome Biol.* **2007**. [[CrossRef](#)] [[PubMed](#)]
243. Ma, A.C.; Ward, A.C.; Liang, R.; Leung, A.Y. The role of jak2a in zebrafish hematopoiesis. *Blood* **2007**, *110*, 1824–1830. [[CrossRef](#)] [[PubMed](#)]
244. Sobhkhez, M.; Hansen, T.; Iliev, D.B.; Skjesol, A.; Jorgensen, J.B. The Atlantic salmon protein tyrosine kinase Tyk2: Molecular cloning, modulation of expression and function. *Dev. Comp. Immunol.* **2013**, *41*, 553–563. [[CrossRef](#)] [[PubMed](#)]
245. Sobhkhez, M.; Skjesol, A.; Thomassen, E.; Tollersrud, L.G.; Iliev, D.B.; Sun, B.; Robertsen, B.; Jorgensen, J.B. Structural and functional characterization of salmon STAT1, STAT2 and IRF9 homologs sheds light on interferon signaling in teleosts. *FEBS Open Bio.* **2014**, *4*, 858–871. [[CrossRef](#)] [[PubMed](#)]
246. Nie, L.; Xiong, R.; Zhang, Y.S.; Zhu, L.Y.; Shao, J.Z.; Xiang, L.X. Conserved inhibitory role of teleost SOCS-1s in IFN signaling pathways. *Dev. Comp. Immunol.* **2014**, *43*, 23–29. [[CrossRef](#)] [[PubMed](#)]
247. Skjesol, A.; Liebe, T.; Iliev, D.B.; Thomassen, E.I.; Tollersrud, L.G.; Sobhkhez, M.; Lindenskov Joensen, L.; Secombes, C.J.; Jorgensen, J.B. Functional conservation of suppressors of cytokine signaling proteins between teleosts and mammals: Atlantic salmon SOCS1 binds to JAK/STAT family members and suppresses type I and II IFN signaling. *Dev. Comp. Immunol.* **2014**, *45*, 177–189. [[CrossRef](#)] [[PubMed](#)]
248. Xiong, R.; Nie, L.; Xiang, L.X.; Shao, J.Z. Characterization of a PIAS4 homologue from zebrafish: Insights into its conserved negative regulatory mechanism in the TRIF, MAVS, and IFN signaling pathways during vertebrate evolution. *J. Immunol.* **2012**, *188*, 2653–2668. [[CrossRef](#)] [[PubMed](#)]

249. Chen, W.Q.; Xu, Q.Q.; Chang, M.X.; Zou, J.; Secombes, C.J.; Peng, K.M.; Nie, P. Molecular characterization and expression analysis of the IFN-gamma related gene (IFN-gammarel) in grass carp *Ctenopharyngodon idella*. *Vet. Immunol. Immunopathol.* **2010**, *134*, 199–207. [[CrossRef](#)] [[PubMed](#)]
250. Igawa, D.; Sakai, M.; Savan, R. An unexpected discovery of two interferon gamma-like genes along with interleukin (IL)-22 and -26 from teleost: IL-22 and -26 genes have been described for the first time outside mammals. *Mol. Immunol.* **2006**, *43*, 999–1009. [[CrossRef](#)] [[PubMed](#)]
251. Swain, B.; Basu, M.; Lenka, S.S.; Das, S.; Jayasankar, P.; Samanta, M. Characterization and inductive expression analysis of interferon gamma-related gene in the Indian major carp, rohu (*Labeo rohita*). *DNA Cell Biol.* **2015**, *34*, 367–378. [[CrossRef](#)] [[PubMed](#)]
252. Yabu, T.; Toda, H.; Shibasaki, Y.; Araki, K.; Yamashita, M.; Anzai, H.; Mano, N.; Masuhiro, Y.; Hanazawa, S.; Shiba, H.; *et al.* Antiviral protection mechanisms mediated by ginbuna crucian carp interferon gamma isoforms 1 and 2 through two distinct interferon gamma-receptors. *J. Biochem.* **2011**, *150*, 635–648. [[CrossRef](#)] [[PubMed](#)]
253. Zou, J.; Yoshiura, Y.; Dijkstra, J.M.; Sakai, M.; Ototake, M.; Secombes, C. Identification of an interferon gamma homologue in Fugu, *Takifugu rubripes*. *Fish Shellfish Immunol.* **2004**, *17*, 403–409. [[CrossRef](#)] [[PubMed](#)]
254. Zou, J.; Carrington, A.; Collet, B.; Dijkstra, J.M.; Yoshiura, Y.; Bols, N.; Secombes, C. Identification and bioactivities of IFN-gamma in rainbow trout *Oncorhynchus mykiss*: The first Th1-type cytokine characterized functionally in fish. *J. Immunol.* **2005**, *175*, 2484–2494. [[CrossRef](#)] [[PubMed](#)]
255. Grayfer, L.; Garcia, E.G.; Belosevic, M. Comparison of macrophage antimicrobial responses induced by type II interferons of the goldfish (*Carassius auratus* L.). *J. Biol. Chem.* **2010**, *285*, 23537–23547. [[CrossRef](#)] [[PubMed](#)]
256. Pijanowski, L.; Scheer, M.; Verburg-van Kemenade, B.M.; Chadzinska, M. Production of inflammatory mediators and extracellular traps by carp macrophages and neutrophils in response to lipopolysaccharide and/or interferon-gamma2. *Fish Shellfish Immunol.* **2015**, *42*, 473–482. [[CrossRef](#)] [[PubMed](#)]
257. Yang, K.; Zhang, S.; Chen, D.; Zhang, A.; Wang, X.; Zhou, H. IFN-gamma-activated lymphocytes boost nitric oxide production in grass carp monocytes/macrophages. *Fish Shellfish Immunol.* **2013**, *35*, 1635–1641. [[CrossRef](#)] [[PubMed](#)]
258. Wang, T.; Diaz-Rosales, P.; Martin, S.A.; Secombes, C.J. Cloning of a novel interleukin (IL)-20-like gene in rainbow trout *Oncorhynchus mykiss* gives an insight into the evolution of the IL-10 family. *Dev. Comp. Immunol.* **2010**, *34*, 158–167. [[CrossRef](#)] [[PubMed](#)]
259. Arts, J.A.; Tijhaar, E.J.; Chadzinska, M.; Savelkoul, H.F.; Verburg-van Kemenade, B.M. Functional analysis of carp interferon-gamma: Evolutionary conservation of classical phagocyte activation. *Fish Shellfish Immunol.* **2010**, *29*, 793–802. [[CrossRef](#)] [[PubMed](#)]
260. Jung, C.Y.; Hikima, J.; Ohtani, M.; Jang, H.B.; del Castillo, C.S.; Nho, S.W.; Cha, I.S.; Park, S.B.; Aoki, T.; Jung, T.S. Recombinant interferon-gamma activates immune responses against *Edwardsiella tarda* infection in the olive flounder, *Paralichthys olivaceus*. *Fish Shellfish Immunol.* **2012**, *33*, 197–203. [[CrossRef](#)] [[PubMed](#)]
261. Shibasaki, Y.; Yabu, T.; Araki, K.; Mano, N.; Shiba, H.; Moritomo, T.; Nakanishi, T. Peculiar monomeric interferon gammas, IFN γ rel 1 and IFN γ rel 2, in ginbuna crucian carp. *FEBS J.* **2014**, *281*, 1046–1056. [[CrossRef](#)] [[PubMed](#)]
262. Stolte, E.H.; Savelkoul, H.F.; Wiegertjes, G.; Flik, G.; Lidy Verburg-van Kemenade, B.M. Differential expression of two interferon-gamma genes in common carp (*Cyprinus carpio* L.). *Dev. Comp. Immunol.* **2008**, *32*, 1467–1481. [[CrossRef](#)] [[PubMed](#)]
263. Aggad, D.; Stein, C.; Sieger, D.; Mazel, M.; Boudinot, P.; Herbomel, P.; Levraud, J.P.; Lutfalla, G.; Leptin, M. *In vivo* analysis of Ifn-gamma1 and Ifn-gamma2 signaling in zebrafish. *J. Immunol.* **2010**, *185*, 6774–6782. [[CrossRef](#)] [[PubMed](#)]
264. Biswas, G.; Nagamine, R.; Hikima, J.; Sakai, M.; Kono, T. Inductive immune responses in the Japanese pufferfish (*Takifugu rubripes*) treated with recombinant IFN-gamma, IFN-gammarel, IL-4/13A and IL-4/13B. *Int. Immunopharmacol.* **2016**, *31*, 50–56. [[CrossRef](#)] [[PubMed](#)]
265. Grayfer, L.; Hodgkinson, J.W.; Hitchen, S.J.; Belosevic, M. Characterization and functional analysis of goldfish (*Carassius auratus* L.) interleukin-10. *Mol. Immunol.* **2011**, *48*, 563–571. [[CrossRef](#)] [[PubMed](#)]
266. Harun, N.O.; Costa, M.M.; Secombes, C.J.; Wang, T. Sequencing of a second interleukin-10 gene in rainbow trout *Oncorhynchus mykiss* and comparative investigation of the expression and modulation of the paralogues *in vitro* and *in vivo*. *Fish Shellfish Immunol.* **2011**, *31*, 107–117. [[CrossRef](#)] [[PubMed](#)]

267. Inoue, Y.; Kamota, S.; Ito, K.; Yoshiura, Y.; Ototake, M.; Moritomo, T.; Nakanishi, T. Molecular cloning and expression analysis of rainbow trout (*Oncorhynchus mykiss*) interleukin-10 cDNAs. *Fish Shellfish Immunol.* **2005**, *18*, 335–344. [[CrossRef](#)] [[PubMed](#)]
268. Pinto, R.D.; Nascimento, D.S.; Reis, M.I.; do Vale, A.; Dos Santos, N.M. Molecular characterization, 3D modelling and expression analysis of sea bass (*Dicentrarchus labrax* L.) interleukin-10. *Mol. Immunol.* **2007**, *44*, 2056–2065. [[CrossRef](#)] [[PubMed](#)]
269. Savan, R.; Igawa, D.; Sakai, M. Cloning, characterization and expression analysis of interleukin-10 from the common carp, *Cyprinus carpio* L. *Eur. J. Biochem.* **2003**, *270*, 4647–4654. [[CrossRef](#)] [[PubMed](#)]
270. Seppola, M.; Larsen, A.N.; Steiro, K.; Robertsen, B.; Jensen, I. Characterisation and expression analysis of the interleukin genes, IL-1beta, IL-8 and IL-10, in Atlantic cod (*Gadus morhua* L.). *Mol. Immunol.* **2008**, *45*, 887–897. [[CrossRef](#)] [[PubMed](#)]
271. Wei, H.; Yang, M.; Zhao, T.; Wang, X.; Zhou, H. Functional expression and characterization of grass carp IL-10: An essential mediator of TGF-beta1 immune regulation in peripheral blood lymphocytes. *Mol. Immunol.* **2013**, *53*, 313–320. [[CrossRef](#)] [[PubMed](#)]
272. Zou, J.; Clark, M.S.; Secombes, C.J. Characterisation, expression and promoter analysis of an interleukin 10 homologue in the puffer fish, *Fugu rubripes*. *Immunogenetics* **2003**, *55*, 325–335. [[CrossRef](#)] [[PubMed](#)]
273. Piazzon, M.C.; Wentzel, A.S.; Tijhaar, E.J.; Rakus, K.L.; Vanderplasschen, A.; Wiegertjes, G.F.; Forlenza, M. Cyprinid herpesvirus 3 IL10 inhibits inflammatory activities of carp macrophages and promotes proliferation of IgM+ B cells and memory T cells in a manner similar to carp IL10. *J. Immunol.* **2015**, *195*, 3694–3704. [[CrossRef](#)] [[PubMed](#)]
274. Sunarto, A.; Liongue, C.; McColl, K.A.; Adams, M.M.; Bulach, D.; Crane, M.S.; Schat, K.A.; Slobedman, B.; Barnes, A.C.; Ward, A.C.; *et al.* Koi herpesvirus encodes and expresses a functional interleukin-10. *J. Virol.* **2012**, *86*, 11512–11520. [[CrossRef](#)] [[PubMed](#)]
275. Van Beurden, S.J.; Forlenza, M.; Westphal, A.H.; Wiegertjes, G.F.; Haenen, O.L.; Engelsma, M.Y. The alloherpesviral counterparts of interleukin 10 in European eel and common carp. *Fish Shellfish Immunol.* **2011**, *31*, 1211–1217. [[CrossRef](#)] [[PubMed](#)]
276. Piazzon, M.C.; Savelkoul, H.S.; Pietretti, D.; Wiegertjes, G.F.; Forlenza, M. Carp IL10 has anti-inflammatory activities on phagocytes, promotes proliferation of memory T cells, and regulates B cell differentiation and antibody secretion. *J. Immunol.* **2015**, *194*, 187–199. [[CrossRef](#)] [[PubMed](#)]
277. Monte, M.M.; Zou, J.; Wang, T.; Carrington, A.; Secombes, C.J. Cloning, expression analysis and bioactivity studies of rainbow trout (*Oncorhynchus mykiss*) interleukin-22. *Cytokine* **2011**, *55*, 62–73. [[CrossRef](#)] [[PubMed](#)]
278. Qi, Z.; Zhang, Q.; Wang, Z.; Zhao, W.; Chen, S.; Gao, Q. Molecular cloning, expression analysis and functional characterization of interleukin-22 in So-iny mullet, *Liza haematocheila*. *Mol. Immunol.* **2015**, *63*, 245–252. [[CrossRef](#)] [[PubMed](#)]
279. Corripio-Miyar, Y.; Zou, J.; Richmond, H.; Secombes, C.J. Identification of interleukin-22 in gadoids and examination of its expression level in vaccinated fish. *Mol. Immunol.* **2009**, *46*, 2098–2106. [[CrossRef](#)] [[PubMed](#)]

