



ADAM17/EGFR axis promotes transglutaminase-dependent skin barrier formation through phospholipase C 1 and protein kinase C pathways.

Wolf, C; Qian, Y; Brooke, MA; Kelsell, DP; Franzke, C-W

© The Author(s) 2016

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

For additional information about this publication click this link.

<http://qmro.qmul.ac.uk/xmlui/handle/123456789/19409>

Information about this research object was correct at the time of download; we occasionally make corrections to records, please therefore check the published record when citing. For more information contact scholarlycommunications@qmul.ac.uk

SCIENTIFIC REPORTS



OPEN

ADAM17/EGFR axis promotes transglutaminase-dependent skin barrier formation through phospholipase C γ 1 and protein kinase C pathways

Cristina Wolf^{1,*}, Yawen Qian^{1,*}, Matthew A. Brooke², David P. Kelsell²
& Claus-Werner Franzke¹

Received: 23 September 2016

Accepted: 28 November 2016

Published: 22 December 2016

The vitally important skin barrier is formed by extensive cross-linking activity of transglutaminases (TGs) during terminal epidermal differentiation. We have previously shown that epidermal deficiency of a disintegrin and metalloproteinase 17 (ADAM17), the principal EGFR ligand sheddase, results in postnatal skin barrier defects in mice due to impeded TG activity. However, the mechanism by which ADAM17/EGFR signalling maintains TG activity during epidermal differentiation remains elusive. Here we demonstrate that ADAM17-dependent EGFR signalling promotes TG activity in keratinocytes committed to terminal differentiation by direct induction of TG1 expression. Restored TG1 expression of EGF-stimulated differentiated *Adam17*^{-/-} keratinocytes was strongly repressed by inhibitors for PLC γ 1 or protein kinase C (PKC) pathways, while treatment with the PKC stimulator 12-*O*-tetradecanoylphorbol-13-acetate restored TG activity in the epidermis of keratinocyte-specific *Adam17*^{-/-} (*AD17* ^{Δ KC}) mice. Further investigations emphasized the expression of PKC η , a mediator of *TGM1* transcription, to be sensitive to EGFR activation. In agreement, topical skin application of cholesterol sulfate, an activator of PKC η , significantly improved TG activity in epidermis of *AD17* ^{Δ KC} mice. Our results suggest ADAM17/EGFR-driven PLC γ 1 and PKC pathways as important promoters of TG1 expression during terminal keratinocyte differentiation. These findings may help to identify new therapeutic targets for inflammatory skin diseases related to epidermal barrier defects.

The multilayered epidermis builds up a barrier that protects the body against transepidermal water loss, foreign substances and microbial invasion¹. This skin barrier is very important for the epidermal homeostasis and needs to be continuously renewed and enzymatically modified². After the basal keratinocytes detach from the underlying basement membrane, they stop to proliferate and become committed to terminal differentiation. During their passage to the skin surface the cells convert into a cornified envelope (CE) that forms the skin barrier. The CE represents an insoluble protein structure that is stabilized by the cross-linking activity of three epidermal transglutaminases (TGs), namely TG1, TG3 and TG5³. However, lack of either TG3 or TG5 activity only leads to minor alterations in CE formation and barrier stability^{4,5}, while lack of TG1 activity causes severe skin barrier defects. The crucial role of TG1 during CE formation is demonstrated in patients with nonsense or missense mutations in the *TGM1* gene, that led to impaired skin barrier formation, transepidermal water loss and skin inflammation in autosomal recessive lamellar ichthyosis or congenital ichthyosiform erythroderma^{3,6}. However the regulatory mechanisms that control TG activity during skin barrier maintenance remain elusive.

In the epidermis, epidermal growth factor receptor (EGFR) is expressed abundantly in the proliferative basal layer and to a lesser degree in the differentiating suprabasal layers⁷. It is thought that EGFR signalling in basal

¹Department of Dermatology, Medical Center - University of Freiburg, Freiburg, Germany. ²Blizard Institute, Barts and the London School of Medicine and Dentistry, Queen Mary University of London, London, United Kingdom. ³Present address: Luxembourg Centre for Systems Biomedicine, University of Luxembourg, Luxembourg. ⁴These authors contributed equally to this work. Correspondence and requests for materials should be addressed to C.W.-F. (email: claus-werner.franzke@uniklinik-freiburg.de)

keratinocytes mainly supports proliferation and survival but prevents differentiation. Moreover it delays apoptosis during early differentiation in suprabasal keratinocytes that have lost their interaction with the matrix^{8,9}. However, EGFR-ligands are also abundant in differentiated epidermis and there are several evidences that EGFR signalling contributes to terminal keratinocyte differentiation and skin barrier formation^{10–13}. EGFR deficiency causes defects in hair follicle development and immature epidermal differentiation with inflammatory skin reactions in both mice and humans^{14–17}. In addition, EGFR inhibitor therapy in cancer patients commonly induces dermatologic side effects including xerotic itchy skin¹⁸. Although these data corroborate the relevance of EGFR signalling in skin homeostasis, only little is known about the role of EGFR signalling in skin barrier formation and in suppressing chronic skin inflammation.

ADAM17 (a disintegrin and metalloproteinase 17) or tumor necrosis factor α -converting enzyme (TACE) is a membrane-anchored metalloproteinase that was originally identified to cleave membrane-bound tumor necrosis factor (TNF)- α by a process named as ectodomain shedding¹⁹. This protease is also known as crucial upstream regulator of EGFR signalling by shedding of the majority of EGFR ligands²⁰. Mice lacking ADAM17 die at birth due to defects in heart development and show epithelial abnormalities in several organs, such as intestine and skin²¹. Thereby, *Adam17*^{-/-} mice nearly phenocopy mice lacking EGFR, or mice lacking the EGFR ligands TGF- α , HB-EGF, or amphiregulin (AREG), indicating an *in vivo* relevance of ADAM17 in EGFR signalling²².

To investigate the role of ADAM17 and EGFR in skin homeostasis, mice with a conditional keratinocyte-specific deletion were generated. *Adam17* ^{Δ KC} (*AD17* ^{Δ KC}) mice phenocopy *Egfr* ^{Δ KC} mice in having an intact skin barrier at birth, but developing a pronounced defect in the skin barrier after the third postnatal week leading to more than 80% lethality. The surviving animals develop chronic dermatitis as adults^{13,16,23}. During the last five years several patients with germline loss of function mutations in *ADAM17* or *EGFR* have been described^{17,24,25}, which developed chronic dermatitis with iterated skin infections, very similar to the phenotype of *AD17* ^{Δ KC} mice, which suggests similar skin barrier defects^{13,23}. Our investigations on *AD17* ^{Δ KC} and *Egfr* ^{Δ KC} mice revealed that ADAM17/EGFR axis sustained the CE formation and postnatal skin barrier stability by tightly regulation of the expression and proteolytic processing of several CE components, especially by supporting TG activity^{13,26}. Accordingly, application of EGFR ligand TGF- α to *AD17* ^{Δ KC} mouse skin restored epidermal barrier integrity by stimulating skin TG activity¹³. However, the ADAM17-driven mechanisms that maintain epidermal TG activity during terminal differentiation are not well understood.

Here we demonstrate that ADAM17-dependent EGFR signalling directly induces TG1 expression in keratinocytes committed to terminal differentiation prominently through phospholipase C γ 1 (PLC γ 1) and protein kinase C (PKC) pathways. Further investigations identified PKC η expression to be responsive to EGFR activation. In agreement, topical skin application of cholesterol sulfate, an activator of PKC η , significantly improved TG activity in epidermis of *AD17* ^{Δ KC} mice. These findings will help to uncover novel therapeutic strategies for inflammatory skin diseases related to disrupted EGFR signalling.

Results

ADAM17-driven EGFR signalling induces expression and activity of TG1 in terminal differentiating keratinocytes. The TG activity in terminal differentiating keratinocyte is composed of activities derived from TG1, TG3 and TG5². We previously demonstrated that lack of *Adam17* in differentiated murine keratinocytes leads to significantly reduced TG activity and decreased expression of TG1 and TG3¹³ as well as reduced *Tgm5* transcription (data not shown). These findings suggest that ADAM17 controls TG activity by modulation of all epithelial TGs. However, lack of either TG3 or TG5 activity causes, if at all, only minor barrier defects^{4,5}, while lack or strongly reduced TG1 activity leads to severe barrier defects^{3,6} as seen in *AD17* ^{Δ KC} mice. Therefore, we focused our investigations on TG1 expression and activity.

For the determination of TG1 activity we analyzed skin cryo sections or culture cells on coverslips *in situ* using the biotinylated amine donor substrate monodansylcadaverine in a neutral pH 7.4 buffer system, in which TG3 and TG5 are not active^{5,27}. It has been previously demonstrated that this *in situ* fluorescence technique can discriminate among the activities of TG1 and TG3/TG5 when performed either at neutral pH 7.4 or at basic pH 8.4, respectively. This based on the fact that the catalytic optimum of TG1 lies between pH 7.4 and pH 8.4, while the catalytic optimum for TG3 and TG5 lies at basic pH 8.4^{27,28}. To investigate how the ADAM17-dependent EGFR signalling regulates TG activity in keratinocytes committed to differentiation, we used suspension cultures on polyhydroxyethylmethacrylate (polyHEMA)-coated plates¹⁰. This system mimicks terminal differentiation of suprabasal epidermal keratinocytes, since it disrupts the cell-extracellular matrix interaction, while maintaining cell-cell interactions. Time-resolved analysis of the TG activity in polyHEMA cultured wild type keratinocytes revealed strong activity after 6 h, which was retained for at least 48 h (Supplementary Fig. S1a). This correlated with a strong increase in transcription of the genes encoding for TG1 or involucrin (Supplementary Fig. S1b). Since permanent cell cycle exit and commitment to terminal differentiation of keratinocytes occurred within the first 24 h of suspension culture²⁹, we have used 24 h suspension culture in our following experiments.

Analysis of EGFR signalling in *Adam17*^{-/-} keratinocytes by Western blot (WB) revealed about 80% reduced EGFR activation (Fig. 1a). To validate whether the reduced TG activity is caused by lack of EGFR signalling, we analyzed 24 h suspension cultures of *Egfr*^{-/-} keratinocytes. Indeed, *Egfr*^{-/-} keratinocytes showed strongly reduced TG activity (Fig. 1b). In addition, the reduced TG1 expression in ECM-disrupted *Adam17*^{-/-} keratinocytes was nearly restored by supplementation with the EGFR-ligand TGF- α (40 ng/ml) on protein (Fig. 1c) and on RNA level (Supplementary Fig. S2), suggesting that the lack of EGFR activation is the major cause of reduced TG1 expression in *Adam17*^{-/-} keratinocytes. Since reduced EGFR signalling in the suprabasal *Adam17*^{-/-} keratinocytes can lead to decreased survival signals and apoptosis³⁰, we analyzed skin sections of 10-day-old *AD17* ^{Δ KC} mice with skin barrier defects and their littermates by TUNEL labelling. No difference in the proportion of TUNEL-positive keratinocytes was detectable (Supplementary Fig. S3), indicating no increase of apoptosis in the epidermis of *AD17* ^{Δ KC} mice. Moreover, supplementation of the EGFR-ligands EGF, TGF- α , and Epiregulin

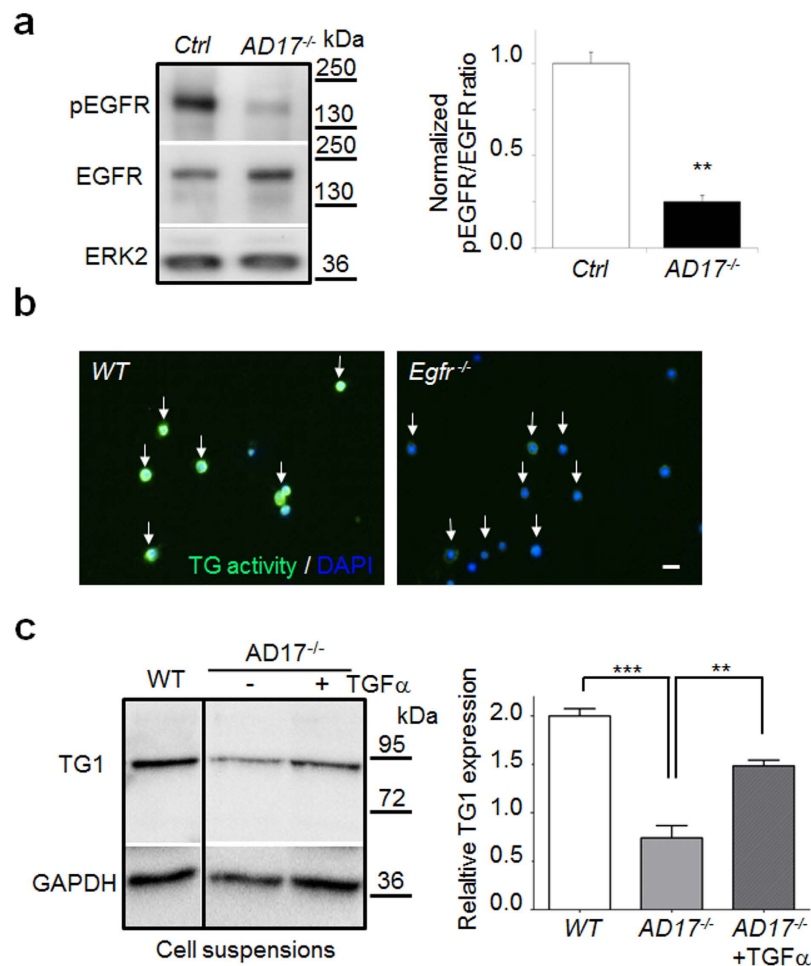


Figure 1. Transglutaminase activity in differentiating mouse keratinocytes is regulated by EGFR signalling. (a) WB of primary keratinocyte lysates derived from *AD17^{AKC}* mice or control littermates (Ctrl) revealed significantly reduced level of activated EGFR (normalized ratio of pEGFR/total EGFR), while ERK 2 protein as control was equal. Graph shows band intensities as mean \pm SD, $n = 3$, $**p < 0.01$. (b) Suspension cultured *Egfr^{-/-}* keratinocytes analyzed for TG activity at pH 7.4 by fluorescence microscopy revealed strongly reduced signals (white arrows, scale bar, 20 μ m) (representative staining from three independent experiments). (c) WB analysis of mouse *AD17^{-/-}* keratinocytes with or without stimulation with 40 ng/ml TGF- α after 24 h suspension culture with antibodies directed against TG1. ($n = 3$ per group). Data as mean \pm SEM, $**p < 0.01$, $***p < 0.001$.

(40 ng/ml) to 24 h *Adam17^{-/-}* keratinocyte suspension cultures resulted in very similar recovery of TG activity (Supplementary Fig. S4), suggesting coinciding functions among these EGFR-ligands. These results suggest that EGFR signalling accelerates TG activity in differentiating keratinocytes via transcription.

PKC regulates TG1 expression downstream of EGFR. The terminal differentiation of keratinocytes is partly regulated by PKC³¹. These kinases are thought to play an important role in the modulation of transglutaminase activity³²⁻³⁴. However, only little is known about the interactions of EGFR and PKC pathways in keratinocytes committed to terminal differentiation. To analyse the interactions of both pathways, we used TGF α -stimulated differentiated *Adam17^{-/-}* keratinocytes as a model for EGFR-dependent TG activity. Addition of the general PKC inhibitor bisindolylmaleimide (1 μ M) led to strongly reduced TG activity and TG1 expression in TGF α -stimulated differentiated *Adam17^{-/-}* keratinocytes (Fig. 2a,b), suggesting a connection of both pathways in the regulation of TG expression and that PKCs are localized downstream of ADAM17/EGFR signalling. These results were confirmed by stimulation experiments with the phorbol ester 12-O-tetradecanoylphorbol-13-acetate (TPA), a strong PKC activator. Stimulation of 24 h suspension cultures with 1 μ M TPA completely restored the TG activity in differentiating *Adam17^{-/-}* keratinocytes (Supplementary Fig. S5a). WB analysis revealed significantly increased TG1 expression in these cells (Supplementary Fig. S5b), suggesting a transcriptional regulation. To further validate the *in vivo* relevance of our results, we used 10 days old keratinocyte-specific *Adam17^{-/-}* mice (*A17^{AKC}*) with strongly reduced epidermal TG activity¹³ and topically applied a single dose of 1.5 μ g TPA in acetone or acetone vehicle alone (controls) on their back skin. TPA treatment for 6 h resulted

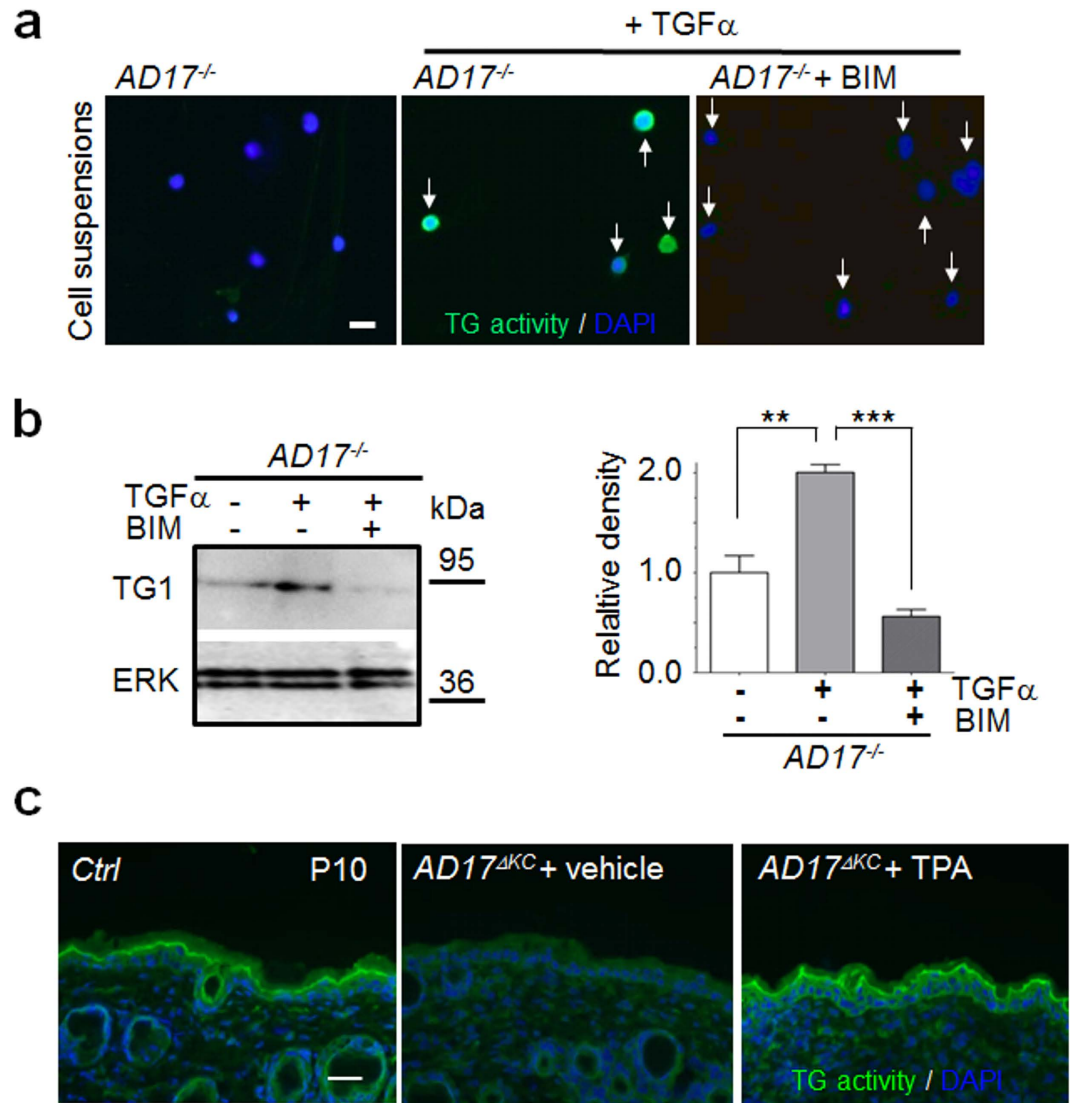


Figure 2. Protein kinase C (PKC) in murine keratinocytes regulates TG1 expression & activity downstream of EGFR *in vitro* and *in vivo*. (a,b) Mouse wild type and *Adam17^{-/-}* keratinocytes were cultured in suspension for 24 h. *Adam17^{-/-}* keratinocytes were either treated with 40 ng/ml TGF- α or 40 ng/ml TGF- α and 1 μ M bisindolylmaleimide (BIM) and then analyzed by (a) *in situ* TG activity detection at pH 7.4 by fluorescence microscopy or (b) WB with antibodies against TG1 and ERK as control. The improved TG1 expression and activity of TGF- α stimulated *Adam17^{-/-}* keratinocytes was strongly inhibited by 1 μ M bisindolylmaleimide. Scale bar, 20 μ m. Graph shows band intensities as mean \pm SEM, n = 3, **p < 0.01, ***p < 0.001. (c) The phorbol ester TPA was topically applied on the shaved skin surface of 10 days old *AD17 Δ KC* mice (P10) and the skin was analyzed for *in situ* TG activity at pH 7.4 after 6 h. Scale bar, 50 μ m. TPA treatment resulted in strongly improved TG activity (representative staining from three independent experiments).

in strongly improved TG activity in the epidermis (Fig. 2c). In contrast, TPA treatment for 1 h did not led to improved TG activity (data not shown). In summary, our results demonstrate that the ADAM17/EGFR axis regulates TG1 expression during terminal keratinocyte differentiation via PKCs.

ADAM17/EGFR axis induces TG activity mainly through PLC γ 1 and PKC pathways. To identify the pathways by which the ADAM17/EGFR axis regulates TG activity in keratinocytes during terminal differentiation we have analyzed the MAPK, PI3K/Akt and PLC/PKC signalling as well known EGFR downstream pathways^{9,35}. WB analysis of lysates of epidermis splits derived from 10 day old *AD17 Δ KC* mice with strongly decreased TG activity¹³ revealed significant reduced activation of the PLC γ 1 pathway, but no changes in ERK1/2 and Akt activation (Fig. 3a). The reduction in PLC γ 1 signalling was verified as a cell-autonomous mechanism, since it was also detected in differentiation-committed *Adam17^{-/-}* keratinocytes (Fig. 3b). In addition, we also saw reduced ERK1/2 signalling in these cells (Fig. 3b), suggesting that the ERK pathway might be involved as well. The PLC γ 1 and the ERK1/2 signalling was strongly induced by EGF stimulation of 24 h *Adam17^{-/-}* keratinocyte suspension cultures, validating the responsiveness of both pathways in differentiation-committed keratinocytes (Fig. 3c).

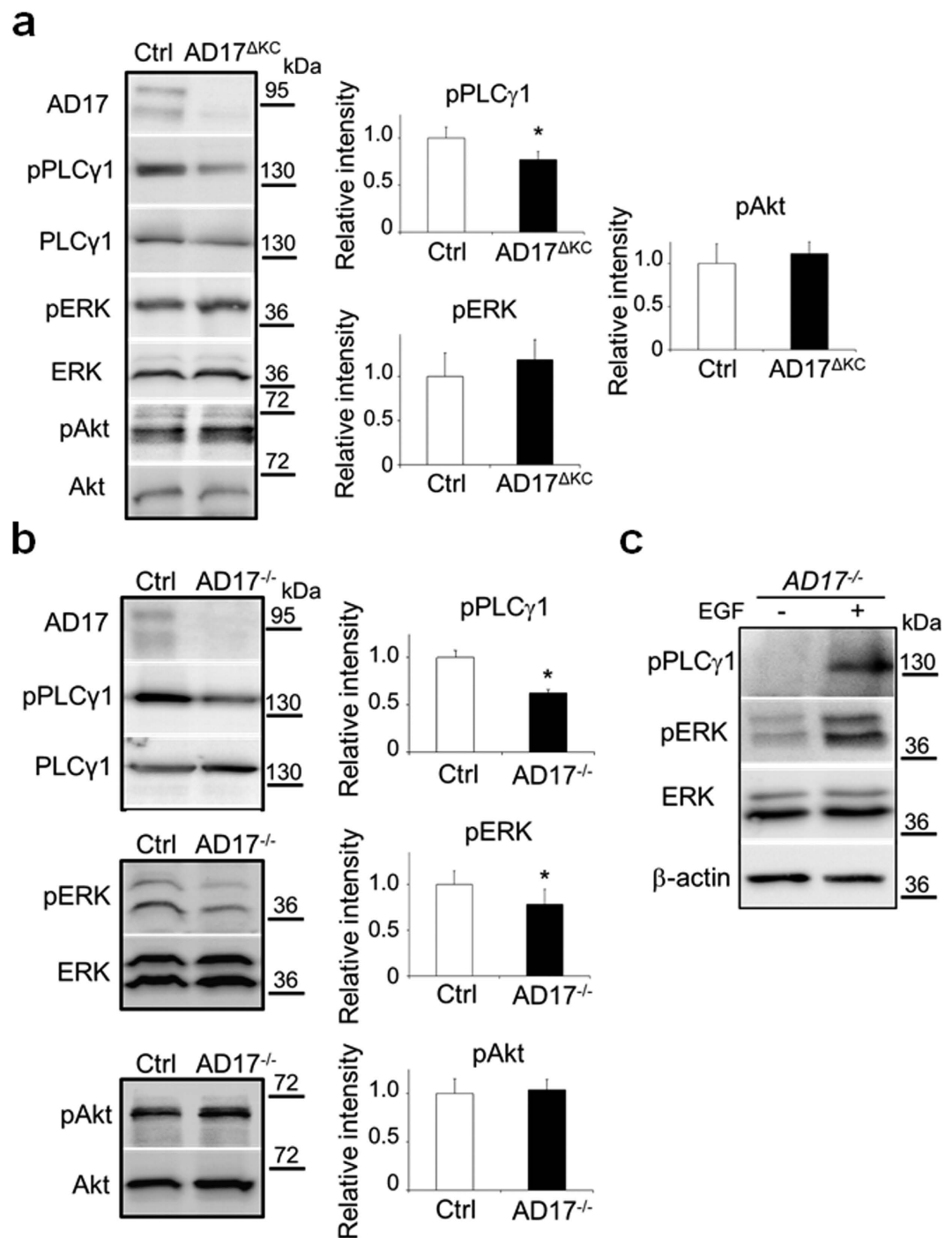
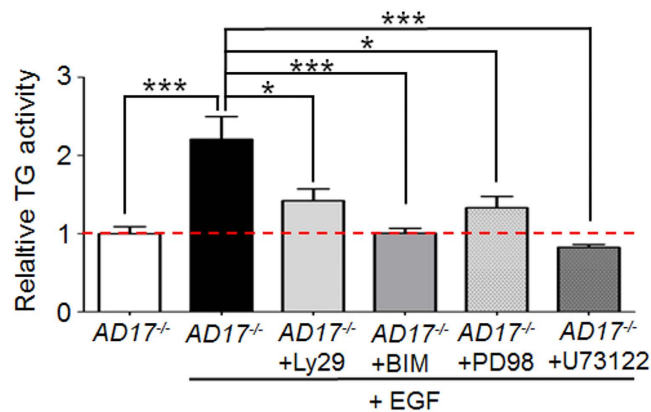
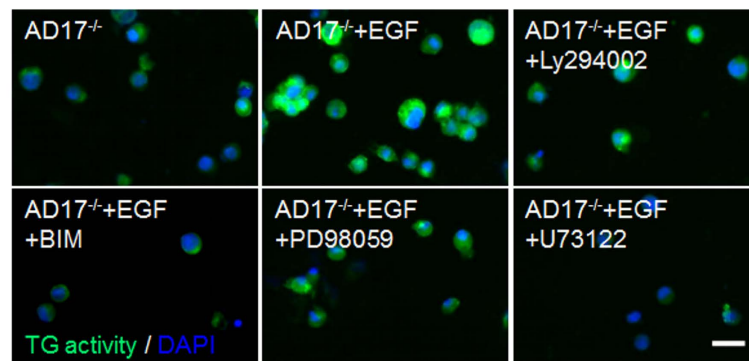


Figure 3. The PLC γ 1 pathway is downregulated in murine *Adam17* $^{-/-}$ keratinocytes. (a,b) Representative WBs of (a) epidermal splits derived from P10 *AD17* Δ KC or wild type mice or (b) 24 h suspension cultured wild type and *Adam17* $^{-/-}$ keratinocytes for activation of PLC γ 1, ERK and Akt pathways. The graphs on the right show the mean \pm SD of the relative intensities of pPLC γ 1/PLC γ 1, pERK/ERK and pAkt/Akt (n = 3), *p < 0.05. The activation of the PLC γ 1 pathway was significantly reduced in *AD17* Δ KC epidermis as well as *Adam17* $^{-/-}$ keratinocytes. (c) After suspension culture for 24 h, *Adam17* $^{-/-}$ keratinocytes were treated with 40 ng/ml EGF or vehicle for additional 30 min and analyzed by WB for activation of PLC γ 1 and ERK. Both, pPLC γ 1 and pERK were strongly increased by EGF stimulation (representative blots of three independent experiments).

Using EGF-stimulated differentiated *Adam17* $^{-/-}$ keratinocytes as a model for EGFR dependent TG activity, the addition of either the PLC inhibitor U73122 or the general PKC inhibitors bisindolylmaleimide or Gö6983 (data not shown) strongly repressed TG activity. However, the addition of the selective PI3K inhibitor Ly294002 or

a



b

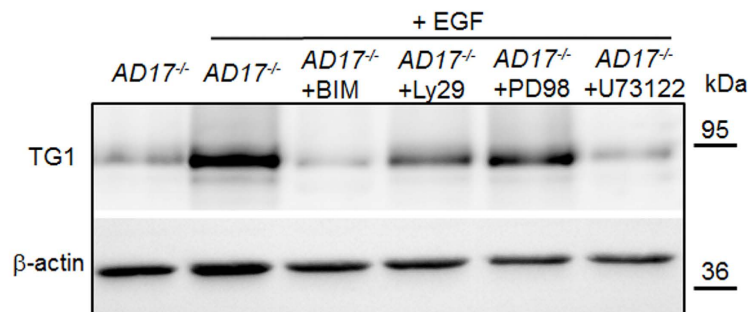


Figure 4. EGFR induced TG activity in murine *Adam17*^{-/-} keratinocytes is mainly driven by PLC γ 1 and PKC signalling. (a) Murine *Adam17*^{-/-} keratinocytes were cultured in suspension on poly-HEMA for 24 h in the presence of 40 ng/ml EGF and either DMSO (vehicle control), 50 μ M Ly294002, 1 μ M bisindolylmaleimide (BIM), 50 μ M PD98059, or 5 μ M U73122. Afterwards, the keratinocytes were analyzed for TG activity at pH 7.4 by fluorescence microscopy. The addition of either the PLC inhibitor U73122 or the general PKC inhibitor bisindolylmaleimide completely decreased EGF-induced TG activity in *Adam17*^{-/-} keratinocytes, while addition of the selective PI3K inhibitor Ly294002 or the selective MEK1 and MEK2 inhibitor PD98059 caused partial inhibition. (n = 4). Scale bar, 20 μ m. Graph shows quantitative analysis of fluorescence intensities: mean \pm SEM, *p < 0.05, ***p < 0.001. (b) WB analysis of cell lysates from the in (a) treated *Adam17*^{-/-} keratinocytes with antibodies against TG1. β -actin was used as loading control (representative blots of two experiments).

the selective MEK1 and MEK2 inhibitor PD98059 led to partial inhibition of TG activity in these cells (Fig. 4a). Further analysis of TG1 expression of those cells by WB revealed very similar results (Fig. 4b). In conclusion, these results suggest that especially the PLC γ 1 and PKC pathways are involved in the regulation of TG activity in differentiation-committed keratinocytes.

Expression of PKC η , but not PKC δ is responsive to EGFR/PLC γ 1 signalling. The expression of three protein kinase C members, namely PKC α , PKC δ and PKC η , has been linked to epidermal terminal differentiation^{36,37}. However, only PKC δ and PKC η seem to be involved in the induction of TG1 activity and expression³⁷. To further understand which of the epidermal PKC isoforms are directly affected by absence of ADAM17/EGFR signalling, we investigated their expression in *AD17* ^{Δ KC} versus wild type mice by WB analysis. Interestingly, we found significantly reduced expression of PKC η in *Adam17*^{-/-} epidermal splits as well as in ECM-disrupted

differentiated *Adam17*^{-/-} keratinocytes. In contrast, no changes were observed for PKC δ (Fig. 5a,b) or PKC α (data not shown). The reduced PKC η expression in differentiation-committed *Adam17*^{-/-} keratinocytes was significantly induced by stimulation with EGF, while PKC δ expression was not affected (Fig. 5c). This result was also seen in wild type keratinocytes (Supplementary Fig. S6), indicating that PKC η expression is modulated by EGFR signalling. The linkage of PLC γ 1 signalling and PKC η expression was assessed by treatment of wild type or *Adam17*^{-/-} keratinocyte suspensions with the PLC inhibitor U73122. Supplementation of 5 μ M U73122 led to dramatically decreased protein expression of PKC η and TG1 in wild type and *Adam17*^{-/-} keratinocytes, while the expression PKC δ was not altered (Fig. 5d). To further confirm the role of PKC η in the modulation of TGs, wild type ECM-disrupted keratinocytes were cultivated for 24 h in the presents of selective inhibitors for either PKC δ or PKC η . The selective myristoylated pseudosubstrate inhibitor (Myr-TRKRQRAMRRRVHQING-OH) for PKC η caused a significant reduction in TG activity. In contrast, rottlerin, a selective PKC δ inhibitor, had no effect on TG activity (Supplementary Fig. S7). Taken together, these results suggest that EGFR/PLC γ 1 signalling is involved in the induction of PKC η expression, which in turn leads to increased TG1 expression and activity.

Cholesterol-sulfate supplementation restores TG activity in *Adam17*^{-/-} keratinocytes *in vitro* and *in vivo*.

The membrane lipid cholesterol sulfate is thought to play an important role in the physiological induction of epidermal terminal differentiation and skin barrier formation. Cholesterol sulfate is constitutively produced in the suprabasal epidermal layers where it progressively accumulates during terminal differentiation. It has previously been shown *in vitro* that stimulation of adhesive keratinocytes with cholesterol sulfate leads to increased expression of terminal differentiation markers, including TG1. This effect is mediated via PKC activity, in which cholesterol sulfate acts as a more specific activator of novel PKCs, especially PKC η ^{38–40}.

To investigate whether cholesterol sulfate can increase TG activity in *Adam17*^{-/-} keratinocytes, we analyzed either the addition of 5 or 20 μ M cholesterol-3-sulfate to suspension cultures. Stimulation of 24 h suspension cultures completely restored the TG activity in the *Adam17*^{-/-} keratinocytes in a dose dependent manner (Fig. 6a). However, WB analysis revealed no increase in PKC η expression in these cells (data not shown), suggesting an induction in kinase activity. To further validate the *in vivo* relevance of our results, we treated 10 days old *AD17* ^{Δ KC} mice by a single topically application of 100 μ g cholesterol-3-sulfate or acetone vehicle (controls) on their back skin. Cholesterol sulfate treatment for 6 h was sufficient to restore the TG activity in the epidermis of *AD17* ^{Δ KC} mice (Fig. 6b), indicating a therapeutic value of local cholesterol sulfate administration.

Reduced TG activity in ADAM17-deficient human skin. After the discovery of the first two *ADAM17* deficient patients with inflammatory skin and bowel disease in 2011²⁴, additional pediatric patients with deficiencies in *ADAM17* or *EGFR* were described^{17,25}. All of these patients suffered of skin inflammation and iterated skin infections, very similar to the phenotype of keratinocyte-specific *Adam17*^{-/-} mice, which suggests similar skin barrier defects^{13,23}. However, the TG activity in the skin of these patients was not evaluated. Thus, we analyzed skin cryo sections derived from an *ADAM17*-deficient patient or healthy donors by *in situ* immunofluorescence microscopy using biotinylated monodansylcadaverine at neutral pH 7.4. As shown in Fig. 7, the TG activity in the stratum granulosum and stratum corneum of the epidermis was strongly reduced in *ADAM17* deficient skin compared to control skin. Therefore, we conclude that the mechanisms of the loss of transglutaminase activity due to *ADAM17*-deficiency are very similar in human and mouse skin.

Discussion

Using epidermis-specific conditional *Adam17* or *Egfr* knockout mice, we have previously shown that lack of *ADAM17*-dependent EGFR ligand shedding in keratinocytes leads to postnatal skin barrier defects due to decreased TG activity^{13,26}. Here, we provide new insights into the downstream mechanisms and demonstrate that *ADAM17*-driven EGFR signalling directly promotes TG1 expression and activity in keratinocytes committed to differentiation mainly via PLC γ 1 and PKC pathways. At first glance our results seem to be in direct conflict with previously published data, which showed EGFR-dependent repression of *TGM1* transcription in keratinocytes due to induced production of the transcription factor homeobox protein A7 (HOXA7)⁴¹. However, these results derived from adherent, proliferative cells that strongly express HOXA7, while its expression is lost in terminal differentiated keratinocytes due to enhanced PKC signalling⁴¹. Thus, our results emphasize the difference in the biological effect of EGFR signalling in delaminated terminal differentiating keratinocytes, where it promotes terminal differentiation via induction of TG1 expression and activity. In agreement, a constitutive increase of *ADAM17*-driven EGFR signalling was observed in tylosis (TOC) keratinocytes⁴². TOC is an autosomal dominant syndrome with focal palmoplantar keratoderma and esophageal cancer due to autosomal dominant *iRHOM2* mutations⁴³. TOC skin also shows epidermal hyperproliferation and hyperkeratosis with elevated epidermal TG activity⁴².

EGFR signalling in basal membrane anchored keratinocytes promotes proliferation and survival via the ERK and Akt pathways, but not via PLC γ 1⁹. In contrast, PLC γ 1 signalling is proposed to lead to terminal keratinocyte differentiation through activation of PKCs^{44,45}. Our data provides the novel finding that the *ADAM17*/EGFR axis in terminal differentiating keratinocytes mainly promotes PLC γ 1 signalling. Although PLC γ 1 activation by direct membrane recruitment to EGFR has been shown in several cell systems^{46,47}, no such activation has been demonstrated so far in keratinocytes⁴⁸. The increased cholesterol production and membrane integration during terminal keratinocyte differentiation seems to have an important impact on the morphological cell changes and the lipid raft-mediated signalling⁴⁹. Therefore it is very likely that this increase in lipid microdomains leads to enriched EGFR cell surface localization in differentiation-committed keratinocytes, which in turn facilitates cell surface recruitment of PLC γ 1 and EGFR/PLC γ 1 signalling⁵⁰. Upon membrane recruitment and activation, PLC γ 1 catalyses the hydrolysis of phosphatidylinositol 4,5-bisphosphate (PIP₂) to inositol 1,4,5-trisphosphate (IP₃) and diacylglycerol (DAG), which both act as second messengers in the activation of PKCs in terminal differentiation⁴⁴.

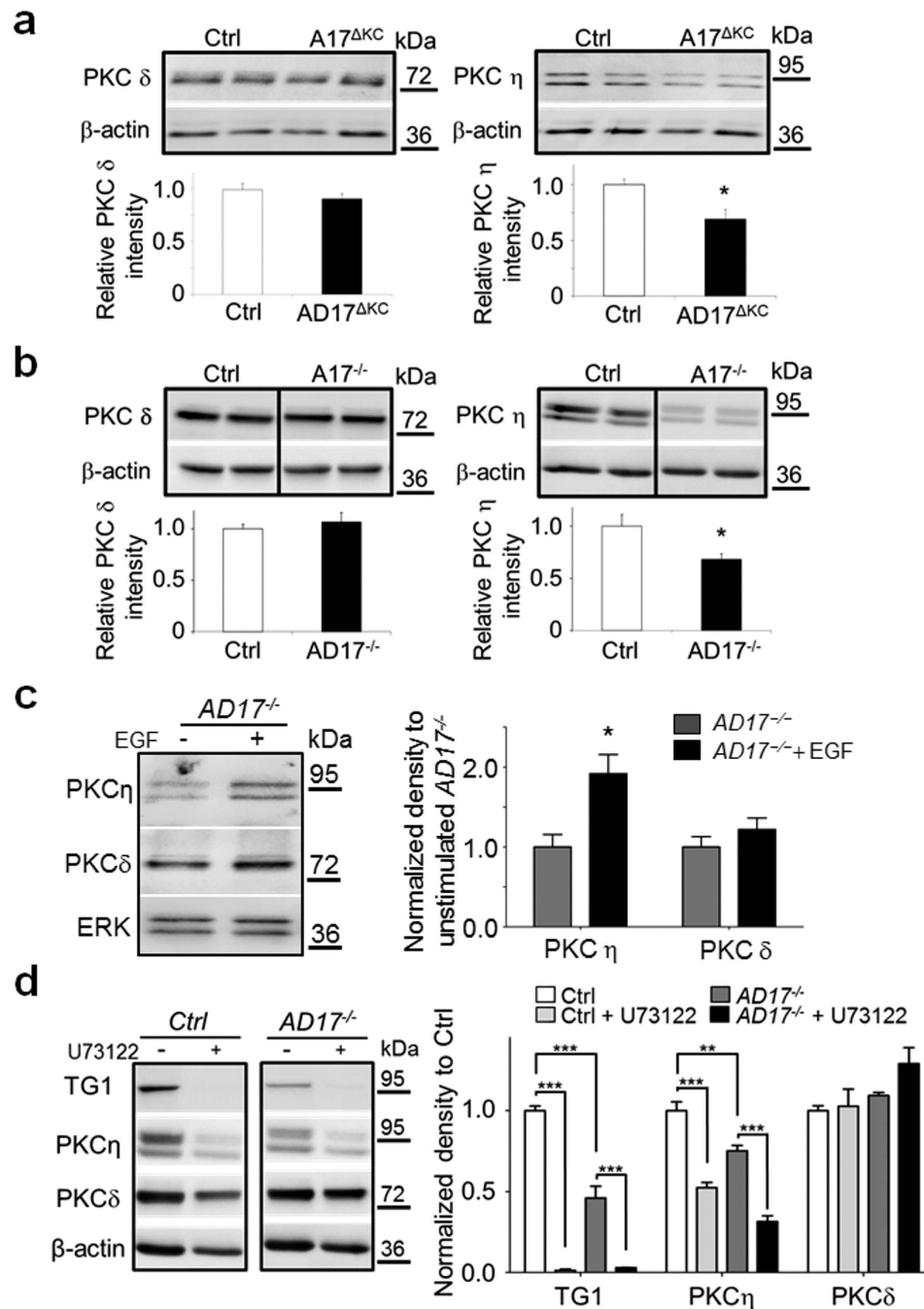
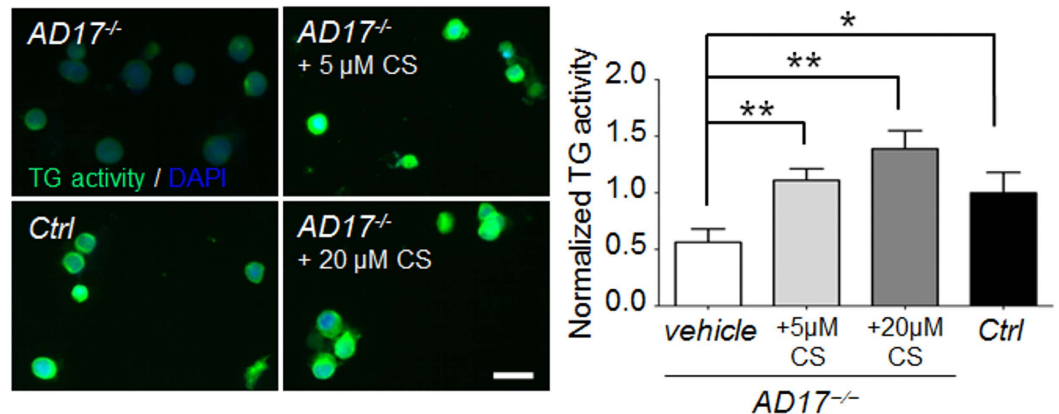


Figure 5. Expression of PKC η but not PKC δ is enhanced by EGFR-PLC γ 1 signalling in murine epidermis. (a,b) WB analysis of (a) epidermal splits derived from P10 AD17 Δ KC or wild type mice or (b) lysates derived from 24 h suspension cultured mouse wild type and *Adam17* $^{-/-}$ keratinocytes with antibodies against PKC isoforms eta or delta, and β -actin as loading control. The graphs below show the mean \pm SD of the signal intensities (n = 5), *p < 0.05; **p < 0.01. The expression of PKC η was significantly reduced in *Adam17* $^{-/-}$ keratinocytes *in vivo* and *in vitro*. (c) *Adam17* $^{-/-}$ keratinocytes were suspension-cultured on poly-HEMA with or without 40 ng/ml EGF for 24 h and further analyzed by WB for PKC isoforms eta or delta and ERK as loading control. Graph on the right shows quantitative analysis of band intensities as mean \pm SEM, *p < 0.05. PKC η expression in *Adam17* $^{-/-}$ keratinocytes was significantly induced by stimulation with EGF, while PKC δ expression was unchanged. (d) Wild type or *Adam17* $^{-/-}$ keratinocytes were suspension-cultured for 24 h with or without addition of 5 μ M U73122 and further analyzed by WB for TG1, PKC isoforms eta or delta. Antibodies against β -actin were used as loading control. Graph on the right shows quantitative analysis of band intensities normalized to controls (n = 3): mean \pm SEM, **p < 0.01; ***p < 0.001. The supplementation of 5 μ M U73122 led to dramatically decreased protein expression of PKC η and TG1 in wild type and *Adam17* $^{-/-}$ keratinocytes, while the expression PKC δ was not affected. Full-length blots of (b) and (d) are presented in Supplementary Fig. S8.

a



b

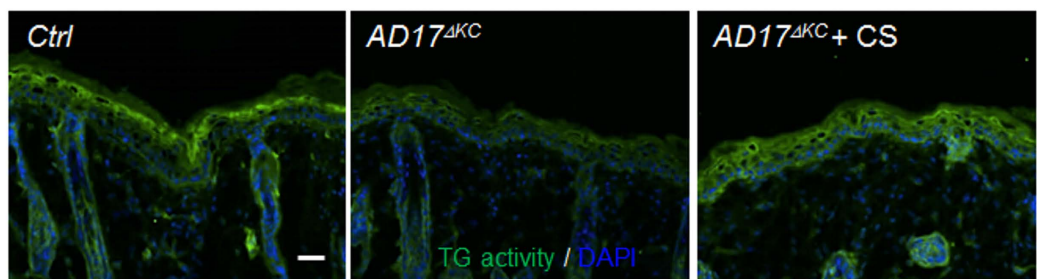


Figure 6. Cholesterol sulfate supplementation increased TG activity in murine keratinocytes *in vivo* and *in vitro*. (a) Mouse *Adam17*^{-/-} keratinocytes were cultured in suspension and stimulated with either 5 μM or 20 μM cholesterol-3-sulfate for 24 h. Analysis of TG activity at pH 7.4 by fluorescence microscopy. TG activity of wild type keratinocytes is shown as control. n = 3. Scale bar, 20 μm. Graph shows fluorescence intensities as mean ± SEM, **p < 0.01, ***p < 0.001. Cholesterol sulfate stimulation resulted in a dose-dependent increase of TG activity in differentiating *Adam17*^{-/-} keratinocytes. (b) Cholesterol sulfate (100 μg/20 μl) or 20 μl acetone-vehicle (control) was topically applied on the shaved skin surface of 10 days old *AD17*^{ΔKC} mice (P10) and the skin was analyzed for *in situ* TG activity at pH 7.4 after 6 h. Untreated skin of wild type mice was used as positive control. Fluorescence microscopy analysis revealed strongly improved TG activity by cholesterol sulfate treatment (representative images of three independent experiments). Scale bar, 50 μm.

Thus, our data suggest that EGFR signalling stabilizes terminal differentiation by maintenance of PKC activity in differentiation-committed keratinocytes via PLCγ1 activation.

In vitro studies revealed that lack of EGFR/ERK signalling in suspension cultured keratinocytes for more than 24 h can lead to apoptosis due to the loss of basement membrane interactions³⁰, which could be the reason for decreased activity and expression of TG1 in *Adam17*^{-/-} keratinocytes. Although we used a different suspension culture system than the authors and cultured the cells for a maximum of 24 h under EGF supplementation, we saw reduced ERK activation in *Adam17*^{-/-} keratinocytes. However, we neither detected decreased ERK signalling nor increased apoptosis as determined by TUNEL labelling in the epidermis of *AD17*^{ΔKC} mice, which questions the *in vivo* relevance of the above findings. Furthermore, it has been demonstrated that PKCs can be involved in the activation of ERK signalling^{51,52}. Thus, reduced PKC signalling might cause decreased ERK activation in *Adam17*^{-/-} keratinocytes.

Our results suggest that ADAM17-driven EGFR signalling in differentiated keratinocytes acts in two different ways to induce PKC signalling and promote TG1 activity. Firstly, it activates PLCγ1 that produces second messengers for the activation of keratinocyte-derived PKC isoforms, namely the classical calcium- and diacylglycerol-dependent PKCα and the novel calcium-independent but diacylglycerol-dependent PKCδ, PKCε and PKCη³¹. Secondly, it selectively induces the expression of PKCη. This result is of particular interest, since only little is known about the transcriptional control of the novel PKCs during terminal differentiation. The induction of PKCη expression is probably driven via EGFR/PLCγ1 signalling, since PKCη expression was highly sensitive to PLCγ1 inhibition. However, it can not be ruled out that other signalling pathways are involved as well. PKCη is strongly linked to terminal keratinocyte differentiation, since it is selectively expressed in suprabasal epidermal layers and its viral overexpression in adherent human keratinocytes caused growth inhibition and induction of TG1 expression and activity^{37,39,53}. However, PKCη activity seems not to be crucial for epidermal differentiation

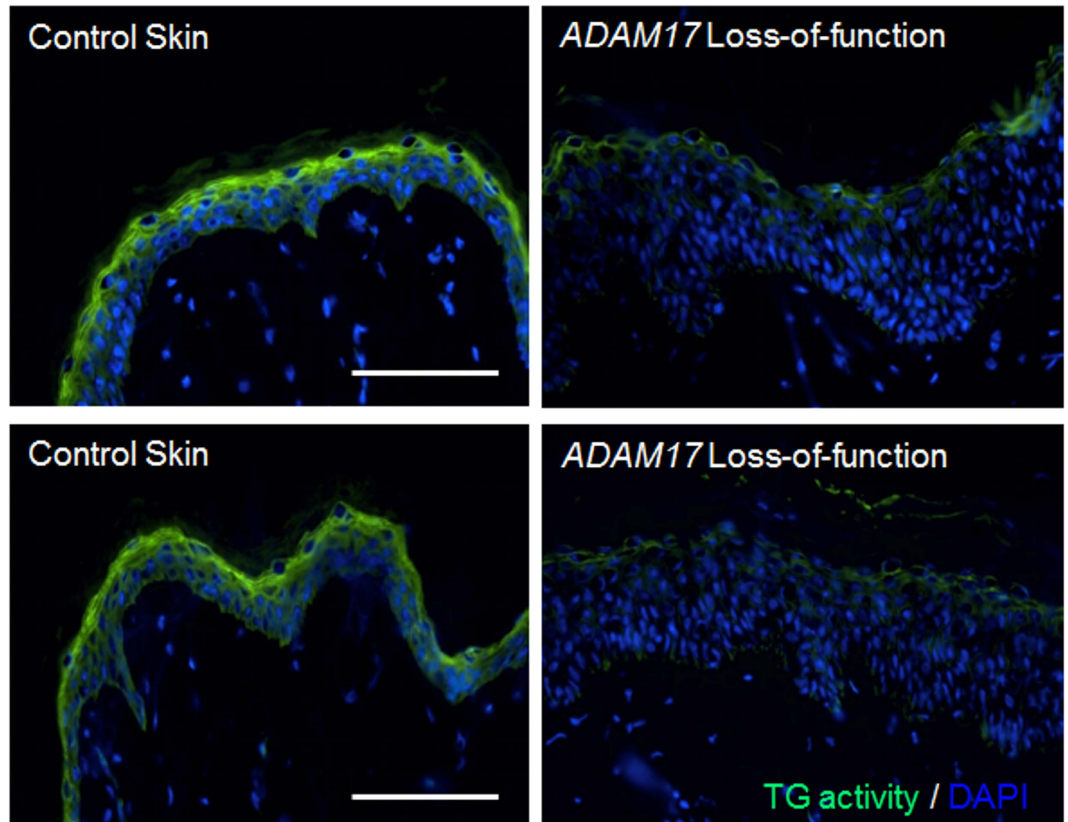


Figure 7. Lack of ADAM17 in humans is associated with decreased epidermal TG activity. *In situ* detection of TG activity in skin sections at pH 7.4 derived from an *ADAM17* deficient patient²⁴ and healthy volunteers by fluorescence microscopy. The TG activity in the stratum granulosum and stratum corneum of the epidermis was strongly reduced in *ADAM17* deficient skin compared to control skin (representative images of three independent experiments). Scale bars, 100 μ m.

in normal skin and is most likely replaceable by other PKC isoforms, since PKC η null mice showed no defects in skin architecture and development⁵⁴.

Taken together, our data suggest ADAM17/EGFR-driven PLC γ 1 and PKC pathways as important promoters of TG1 expression during terminal keratinocyte differentiation and skin barrier formation. It further provides evidence that topical skin application of TPA or cholesterol sulfate can restore the TG activity in the epidermis of *AD17 Δ KC* mice, which should have a beneficial effect on the skin barrier function. In contrast to the tumor-promoting phorbol ester TPA, that activates almost all PKCs, cholesterol sulfate is a more specific activator of novel PKCs, especially PKC η , and rather acts as an anticarcinogenic component^{38–40}. Thus, cholesterol sulfate seems to have therapeutic value for the local treatment of disrupted EGFR signalling caused skin barrier defects in atopic dermatitis⁵⁵ or EGFR inhibitor-treated cancer patients¹⁸.

Methods

Material. The PI3K inhibitor Ly294002 was purchased from Merck Millipore (Germany) and the ERK1/2 inhibitor PD98059 derived from Cell signalling technology (Germany). The general PKC inhibitors Bisindolylmaleimide and Gö6983 were obtained from Sigma-Aldrich (Germany) and Merck Millipore (Germany), respectively. Rottlerin, a selective inhibitor of PKC δ was obtained from Sigma-Aldrich (Germany) and the PKC η pseudosubstrate inhibitor, myristoylated (Myr-TRKRQRAMRRRVHQING-OH) was bought from Merck Millipore. Recombinant murine EGF, human TGF- α and human Epiregulin were obtained from Peprotech (Germany).

Animals. The generation of *Adam17^{fllox/fllox} Krt14-Cre (AD17 Δ KC)* and *Egfr^{fllox/fllox} Krt14-Cre (Egfr Δ KC)* mice has been described previously¹³. All mice were of mixed genetic background (129 Sv, C57BL/6), and all comparisons were between littermates. In experiments, mice received a single 100 μ l application of either 1.5 μ g 12-O-tetradecanoylphorbol-13-acetate (TPA, Sigma-Aldrich) or 100 μ g cholesterol 3-sulfate (Sigma-Aldrich) in acetone or acetone vehicle on the shaved back skin. The mice were maintained in the Center for Experimental Models and Transgenic Service (CEMT) of the Medical Center University of Freiburg, and all experiments were performed according to the guidelines of the German Animal Welfare association and approved by the Regierungspräsidium Freiburg (G-11/98 and G-15/145).

Primary keratinocyte preparation and cultivation. Keratinocytes were isolated from the skin of neonatal *AD17^{ΔKC}* mice and their wild type littermates as described previously⁵⁶. The primary cells were cultured in defined serum-free keratinocyte growth medium (CellnTec, Switzerland) supplemented with 100 U/ml penicillin, 100 μg/ml streptomycin (Invitrogen). The cells were maintained at 37 °C, 5% CO₂ and 95% humidity. Keratinocytes derived from passages 1–2 were used for the experiments.

Keratinocyte suspension culture. Suspension culture on poly (2-hydroxyethyl methacrylate (HEMA))–coated plates was performed as previously described¹⁰. 6-well or 12-well plates were coated with either 10 mg or 4 mg poly-HEMA (Sigma-Aldrich) respectively, followed by extensive PBS washes. 1×10^6 or 0.5×10^6 cells in keratinocyte growth medium (CellnTec, Switzerland) were added to each coated 6-well or 12-well respectively and incubated in a humidified incubator with 5% CO₂ in air at 37 °C for 24 h. EGFR stimulations were performed with either 30 ng/ml EGF, TGFα or Epiregulin. In some experiments, the cell suspensions were supplied with 500 nM GÖ 6983, 1 μM bisindolylmaleimide, 50 μM Ly294002, 50 μM PD98059, or 5 μM U73122 for 24 h. Cell suspensions were harvested by centrifugation and either used for preparation of cell lysates or detection of TG activity. For *in vitro* TG activity detection, 24-h suspension-cultured keratinocytes were attached on gelatin-coated coverslips for 1 h and TG activity was determined as described below.

Quantitative RT-PCR analysis. Total RNA from suspension-cultured *Adam17^{-/-}* and wild type keratinocytes was extracted using RNeasy kit (Qiagen). 1 μg of total RNA was reverse transcribed using a First Strand cDNA Synthesis kit (Fermentas). Relative quantification of gene expression was performed by real-time quantitative PCR using iQ SYBR-Green Supermix on the CFX96TM C1000TM Thermal Cycler (Bio-Rad, Germany) following the manufacturer's protocols. The used primer sequences for *Tgm1*, *Adam17*, *Egfr*, *Inv*, *Krt14* and *Gapdh* were described previously¹³. Relative expression was normalized for levels of *Gapdh*.

Transglutaminase activity assay. For *in situ* detection of TG activity in skin sections or cells, the biotinylated amine donor substrate monodansylcadaverine (MDC) was used as described earlier¹³. 5 μm thick cryostat sections were air dried and preincubated with 1% BSA in 0.1 M Tris-HCl, pH 7.4 for 30 min at room temperature. The sections were then incubated for 45 to 60 min with 100 μM MDC, 5 mM CaCl₂, 0.1 M Tris-HCl, pH 7.4. After stopping the reaction with 10 mM EDTA and extensive PBS washing, the sections were stained with Streptavidin-conjugated Alexa Fluor 488 (Invitrogen) and DAPI-supplemented mounting medium.

Immunoblotting. For WB analysis dispase-separated epidermis splits or cells were homogenized in 50 mM Tris-HCl, pH 8.0, 0.15 M NaCl, 1% Nonidet P-40, 0.5% sodium deoxycholate supplemented with 2 mM EDTA, 5 mM 1,10-ortho-phenanthroline (Sigma-Aldrich) and protease inhibitor cocktail set III (Calbiochem) as described previously¹³. Total protein content was determined using the BCATM protein assay kit (Invitrogen). 30 μg of protein was separated and transferred either onto nitrocellulose or PVDF membranes. Protein detection on membranes was performed with subsequent primary antibodies: rabbit anti-PKCη, rabbit anti-PKCδ, rabbit anti-transglutaminase 1 (TG1), (Santa Cruz Biotechnology), rabbit anti-PLCγ1, rabbit anti-phospho PLC-γ1, rabbit anti-EGFR (Cell Signaling), rabbit anti-phospho EGFR (pY1068, clone EP774Y; Epitomics Inc.), rabbit anti-ERK1/2, mouse anti-β-actin (Sigma-Aldrich) and mouse anti-GAPDH (Invitrogen). Visualization was performed with secondary horseradish peroxidase labeled goat anti-mouse IgG (Merck) or horseradish peroxidase labeled goat anti-rabbit IgG (KPL) and Amersham ECL prime western blotting detection reagent as described by the manufacturer (GE Healthcare).

TUNEL assay. The fluorometric DeadEndTMTUNEL System (Promega) was used to analyze apoptosis in paraffin skin sections (5 μm) of *AD17^{ΔKC}* mice and their wild type littermates according the manufacturer's recommendations.

Patient material. Skin samples were obtained from a 17-year-old male *ADAM17*-deficient patient and healthy volunteers undergoing cosmetic surgery after informed consent and in adherence to the declaration of Helsinki principles. Clinical details of the *ADAM17*-deficient patient were described elsewhere²⁴. The study was approved by the ethics committee of the Queen Mary University of London.

Statistics. The data are present as mean ± SEM. Data of two groups were analyzed for significance using the unpaired Student's t test and differences are considered to be statistically significant at $P < 0.05$.

References

1. Proksch, E., Brandner, J. M. & Jensen, J. M. The skin: an indispensable barrier. *Exp Dermatol* **17**, 1063–72 (2008).
2. Eckhart, L., Lippens, S., Tschachler, E. & Declercq, W. Cell death by cornification. *Biochim Biophys Acta* **1833**, 3471–80 (2013).
3. Candi, E., Schmidt, R. & Melino, G. The cornified envelope: a model of cell death in the skin. *Nat Rev Mol Cell Biol* **6**, 328–340 (2005).
4. John, S. *et al.* Epidermal transglutaminase (TGase 3) is required for proper hair development, but not the formation of the epidermal barrier. *PLoS One* **7**, e34252 (2012).
5. Pigors, M. *et al.* TGM5 mutations impact epidermal differentiation in acral peeling skin syndrome. *J Invest Dermatol* **132**, 2422–9 (2012).
6. Russell, L. J. *et al.* Mutations in the gene for transglutaminase 1 in autosomal recessive lamellar ichthyosis. *Nat Genet* **9**, 279–83 (1995).
7. Piepkorn, M., Predd, H., Underwood, R. & Cook, P. Proliferation-differentiation relationships in the expression of heparin-binding epidermal growth factor-related factors and erbB receptors by normal and psoriatic human keratinocytes. *Arch Dermatol Res* **295**, 93–101 (2003).
8. Pastore, S. & Mascia, F. Novel acquisitions on the immunoprotective roles of the EGF receptor in the skin. *Expert Rev Dermatol* **3**, 525–527 (2008).
9. Schneider, M. R., Werner, S., Paus, R. & Wolf, E. Beyond wavy hairs: the epidermal growth factor receptor and its ligands in skin biology and pathology. *Am J Pathol* **173**, 14–24 (2008).

10. Wakita, H. & Takigawa, M. Activation of epidermal growth factor receptor promotes late terminal differentiation of cell-matrix interaction-disrupted keratinocytes. *J Biol Chem* **274**, 37285–91 (1999).
11. Denning, M. F. *et al.* Cross-talk between epidermal growth factor receptor and protein kinase C during calcium-induced differentiation of keratinocytes. *Exp Dermatol* **9**, 192–9 (2000).
12. Cheng, X. *et al.* TRP channel regulates EGFR signaling in hair morphogenesis and skin barrier formation. *Cell* **141**, 331–43 (2010).
13. Franzke, C. W. *et al.* Epidermal ADAM17 maintains the skin barrier by regulating EGFR ligand-dependent terminal keratinocyte differentiation. *J Exp Med* **209**, 1105–19 (2012).
14. Miettinen, P. J. *et al.* Epithelial immaturity and multiorgan failure in mice lacking epidermal growth factor receptor. *Nature* **376**, 337–341 (1995).
15. Sibilia, M. *et al.* Mice humanised for the EGF receptor display hypomorphic phenotypes in skin, bone and heart. *Development* **130**, 4515–25 (2003).
16. Lichtenberger, B. M. *et al.* Epidermal EGFR controls cutaneous host defense and prevents inflammation. *Sci Transl Med* **5**, 199ra111 (2013).
17. Campbell, P. *et al.* Epithelial inflammation resulting from an inherited loss-of-function mutation in EGFR. *J Invest Derm* **134**, 2570–2578, (2014).
18. Chanprapaph, K., Vachiramon, V. & Rattanakaemakorn, P. Epidermal growth factor receptor inhibitors: a review of cutaneous adverse events and management. *Dermatology research and practice* **2014**, 734249 (2014).
19. Black, R. A. *et al.* A metalloproteinase disintegrin that releases tumour-necrosis factor- α from cells. *Nature* **385**, 729–33 (1997).
20. Sahin, U. *et al.* Distinct roles for ADAM10 and ADAM17 in ectodomain shedding of six EGFR ligands. *J Cell Biol* **164**, 769–79 (2004).
21. Peschon, J. J. *et al.* An essential role for ectodomain shedding in mammalian development. *Science* **282**, 1281–4 (1998).
22. Blobel, C. P., ADAMs: key components in EGFR signalling and development. *Nature reviews. Molecular cell biology* **6**, 32–43 (2005).
23. Murthy, A. *et al.* Notch activation by the metalloproteinase ADAM17 regulates myelopoiesis and atopic barrier immunity by suppressing epithelial cytokine synthesis. *Immunity* **36**, 105–19 (2012).
24. Blaydon, D. C. *et al.* Inflammatory skin and bowel disease linked to ADAM17 deletion. *N Engl J Med* **365**, 1502–8 (2011).
25. Tsukerman, P. *et al.* Cytokine secretion and NK cell activity in human ADAM17 deficiency. *Oncotarget* **6**, 44151–60 (2015).
26. Tholen, S. *et al.* Skin Barrier Defects Caused by Keratinocyte-Specific Deletion of ADAM17 or EGFR Are Based on Highly Similar Proteome and Degradome Alterations. *J Proteome Res* **15**, 1402–17 (2016).
27. Raghunath, M. *et al.* A novel *in situ* method for the detection of deficient transglutaminase activity in the skin. *Arch Dermatol Res* **290**, 621–7 (1998).
28. Candi, E. *et al.* Transglutaminase 5 cross-links loricrin, involucrin, and small proline-rich proteins *in vitro*. *J Biol Chem* **276**, 35014–23 (2001).
29. Nishi, K., Inoue, H., Schnier, J. B. & Rice, R. H. Cyclin D1 downregulation is important for permanent cell cycle exit and initiation of differentiation induced by anchorage-deprivation in human keratinocytes. *J Cell Biochem* **106**, 63–72 (2009).
30. Jost, M., Huggett, T. M., Kari, C. & Rodeck, U. Matrix-independent survival of human keratinocytes through an EGF receptor/ MAPK-kinase-dependent pathway. *Mol Biol Cell* **12**, 1519–27 (2001).
31. Denning, M. F. Epidermal keratinocytes: regulation of multiple cell phenotypes by multiple protein kinase C isoforms. *Int J Biochem Cell Biol* **36**, 1141–6 (2004).
32. Dlugosz, A. A. & Yuspa, S. H. Protein kinase C regulates keratinocyte transglutaminase (TGK) gene expression in cultured primary mouse epidermal keratinocytes induced to terminally differentiate by calcium. *J Invest Dermatol* **102**, 409–14 (1994).
33. Tibudan, S. S., Wang, Y. & Denning, M. F. Activation of protein kinase C triggers irreversible cell cycle withdrawal in human keratinocytes. *J Invest Dermatol* **119**, 1282–9 (2002).
34. Adhikary, G., Chew, Y. C., Reece, E. A. & Eckert, R. L. PKC- δ and - ϵ , MEK-1, MEK-6, MEK-3, and p38- δ are essential mediators of the response of normal human epidermal keratinocytes to differentiating agents. *J Invest Dermatol* **130**, 2017–30 (2010).
35. Wieduwilt, M. J. & Moasser, M. M. The epidermal growth factor receptor family: Biology driving targeted therapeutics. *Cellular and molecular life sciences : CMLS* **65**, 1566–1584 (2008).
36. Seo, H. R. *et al.* PKC α induces differentiation through ERK1/2 phosphorylation in mouse keratinocytes. *Exp Mol Med* **36**, 292–9 (2004).
37. Ohba, M. *et al.* Induction of differentiation in normal human keratinocytes by adenovirus-mediated introduction of the ϵ and δ isoforms of protein kinase C. *Mol Cell Biol* **18**, 5199–207 (1998).
38. Strott, C. A. & Higashi, Y. Cholesterol sulfate in human physiology: what's it all about? *J Lipid Res* **44**, 1268–78 (2003).
39. Kashiwagi, M., Ohba, M., Chida, K. & Kuroki, T. Protein kinase C ϵ (PKC ϵ): its involvement in keratinocyte differentiation. *J Biochem* **132**, 853–7 (2002).
40. Kawabe, S. *et al.* Cholesterol sulfate activates transcription of transglutaminase 1 gene in normal human keratinocytes. *J Invest Dermatol* **111**, 1098–102 (1998).
41. La Celle, P. T. & Polakowska, R. R. Human homeobox HOXA7 regulates keratinocyte transglutaminase type 1 and inhibits differentiation. *J Biol Chem* **276**, 32844–53 (2001).
42. Brooke, M. A. *et al.* iRHOM2-dependent regulation of ADAM17 in cutaneous disease and epidermal barrier function. *Hum Mol Genet* **23**, 4064–76 (2014).
43. Blaydon, D. C. *et al.* RHBDF2 mutations are associated with tylosis, a familial esophageal cancer syndrome. *Am J Hum Genet* **90**, 340–6 (2012).
44. Kim, M. J., Kim, E., Ryu, S. H. & Suh, P. G. The mechanism of phospholipase C- γ 1 regulation. *Exp Mol Med* **32**, 101–109 (2000).
45. Xie, Z., Singleton, P. A., Bourguignon, L. Y. & Bikle, D. D. Calcium-induced human keratinocyte differentiation requires src- and fyn-mediated phosphatidylinositol 3-kinase-dependent activation of phospholipase C- γ 1. *Mol Biol Cell* **16**, 3236–46 (2005).
46. Haugh, J. M., Schooler, K., Wells A., Wiley, H. S. & Lauffenburger, D. A. Effect of epidermal growth factor receptor internalization on regulation of the phospholipase C- γ 1 signaling pathway. *J Biol Chem* **274**, 8958–65 (1999).
47. Gierschik, P., Buehler, A. & Walliser, C. Activated PLC γ Breaking Loose. *Structure* **20**, 1989–90 (2012).
48. Reynolds, N. J. *et al.* Differential induction of phosphatidylcholine hydrolysis, diacylglycerol formation and protein kinase C activation by epidermal growth factor and transforming growth factor- α in normal human skin fibroblasts and keratinocytes. *Biochem J* **294**, 535–44. Erratum in: *Biochem J* **295**, 903 (1993).
49. Spörl, F. *et al.* Real-time monitoring of membrane cholesterol reveals new insights into epidermal differentiation. *J Invest Dermatol* **130**, 1268–78 (2010).
50. Hur, E. M. *et al.* Sensitization of epidermal growth factor-induced signaling by bradykinin is mediated by c-Src. Implications for a role of lipid microdomains. *J Biol Chem* **279**, 5852–60 (2004).
51. Toker, A. Signaling through protein kinase C. *Front Biosci* **3**, D1134–47 (1998).
52. Mitev, V., Le Panse, R., Coulomb, B., Miteva, L. & Houdebine, L. M. Epidermal growth factor stimulates mitogen-activated protein kinase by a PKC-dependent pathway in human keratinocytes. *Biochem Biophys Res Commun* **208**, 245–52 (1995).
53. Ueda, E. *et al.* The ϵ isoform of protein kinase C mediates transcriptional activation of the human transglutaminase 1 gene. *J Biol Chem* **271**, 9790–4 (1996).

54. Chida, K. *et al.* Disruption of protein kinase Ceta results in impairment of wound healing and enhancement of tumor formation in mouse skin carcinogenesis. *Cancer Res* **63**, 2404–8 (2003).
55. Säaf, A. *et al.* Characterization of EGFR and ErbB2 expression in atopic dermatitis patients. *Arch Dermatol Res* **304**, 773–80 (2012).
56. Franzke, C. W., Bruckner-Tuderman, L. & Blobel, C. P. Shedding of collagen XVII/BP180 in skin depends on both ADAM10 and ADAM9. *J Biol Chem* **284**, 23386–96 (2009).

Acknowledgements

This work was supported by the German Research Foundation DFG (SFB 850/B6) and by the Fritz-Thyssen foundation (Az.10.14.2.150) to C.-W.F and the Medical Research Council (MR/L010402/1) to D.P.K. We thank M. Hartmann for technical assistance.

Author Contributions

C.-W.F. conceived the study. Y.Q., C.W., M.B., D.K. and C.-W.F. performed experiments and/or analyzed data. C.W., Y.Q., D.K. and C.-W.F. wrote the manuscript. All authors have read and approved the final manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Wolf, C. *et al.* ADAM17/EGFR axis promotes transglutaminase-dependent skin barrier formation through phospholipase C γ 1 and protein kinase C pathways. *Sci. Rep.* **6**, 39780; doi: 10.1038/srep39780 (2016).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016