

Bioglass and bioactive glasses and their impact on healthcare

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Abstract

Glass caused a revolution in healthcare when Bioglass was discovered by Larry Hench. It was the first material to bond with bone, rather than be encapsulated by fibrous tissue, launching the field of bioactive ceramics. Bioglass is also biodegradable. Almost 50 years on from its discovery that revolution continues. Bioactive glasses stimulate more bone regeneration than other bioactive ceramics, which is attributed to their dissolution products stimulating cells at the genetic level. This second discovery has changed the way clinicians, scientists and regulatory bodies think about medical devices and the concept of bioactivity. The original 45S5 Bioglass has only recently found really widespread use in orthopaedics, having regenerated the bones of more than 1.5 million patients. Its full potential is still yet to be fulfilled. This article takes the reader from Hench's Bioglass 45S5 to its clinical uses and products, before giving examples of non-surgical products that now use Bioglass, from consumer products, such as toothpaste, to cosmetics. Other glasses have also found important healthcare applications, such as borate based glasses that heal chronic wounds. The revolution looks set to continue as new healthcare applications are being found for bioactive glasses, contributing to extending the glass age.

Keywords: Bioactive glass; Bioglass; wound healing; synthetic bone grafts; bone regeneration.

1. Introduction and scope

The discovery of Bioglass[®] was not quite an accident, but it was not far off. Prior to its discovery, all implants designed to repair body parts used materials were selected primarily for their corrosion resistance. The problem is that these implants stimulated a response from the immune system, which recognised them as foreign, isolating from the host tissue through fibrous encapsulation. In orthopaedics, this capsule of soft tissue meant that an implant would not integrate with the host bone and therefore would undergo micromotion and eventually cause the bone to fail. In some cases, the material interaction with the body caused mechanical failure of the implants. Bioglass was different, it was the first synthetic material found to form a chemical bond with bone. The results caused clinicians and medical device companies to change the way they thought about synthetic implant materials. Not only did it form a bond with bone, creating a stable implant, but it (and the bone defect site) was also remodelled over time, restoring healthy bone.

Bioglass was invented by Larry Hench at the University of Florida following a serendipitous bus ride conversation with a US Army Colonel ¹. The colonel challenged Hench to develop a material that could survive the aggressive environment of the human body. Hench made a degradable glass in the Na₂O-CaO-SiO₂-P₂O₅ system, with a composition close a ternary eutectic in the Na₂O-CaO-SiO₂ diagram ². The first composition he tried (46.1 mol% SiO₂, 24.4 mol% Na₂O, 26.9 mol% CaO and 2.6 mol% P₂O₅), which was later termed 45S5 Bioglass, formed such a strong bond with bone that it could not be removed without breaking the bone ³. This discovery launched the field of bioactive inorganic materials, and soon bioactive glass-ceramics ⁴ and calcium phosphate ceramics were also developed ⁵ as synthetic bone graft materials. Originally, the term “bioactive” referred to forming a bond with bone, and that

definition is still used in the context of synthetic bone grafts⁶. Applications for bioactive glass now stretch far wider than bone repair⁷ so a wider definition may be more appropriate, such as “stimulation of a beneficial biological response”.

Bioglass offers two modes of bioactivity in orthopaedics. Bone bonding is attributed to hydroxycarbonate apatite (HCA) layer on the glass that forms following dissolution of the glass surface and reprecipitation of the ions released from the glass surface³. HCA is similar to bone mineral and is thought to interact with collagen fibrils to integrate (bond) with the host bone. Comparative *in vivo* studies between 45S5 Bioglass and similar sized particles of synthetic hydroxyapatite (HA) and apatite/wollastonite (A/W) glass ceramics showed Bioglass could produce more rapid and higher quality bone regeneration⁸. After one week, there was 17 times more bone in defects filled with Bioglass, and twice as much bone after 24 weeks, compared to defects filled with HA (Figure 1a)⁸⁻¹¹. The superior osteogenic properties (later termed osteostimulation) of the glass is thought to be due to the dissolution products of the glass, i.e. soluble silica and calcium ions, that stimulate osteogenic cells to produce bone matrix.^{12, 13} Other studies have shown that a significant number of genes were up-regulated within 48 h which supported osteogenic behaviour¹⁴. Transcription of at least five extracellular matrix (the matrix that cells produce to form the basis of a tissue) components was also induced (Figure 1b). Extracellular matrix secretion increased, which mineralised without addition of supplements^{15, 16}. The gene expression was dose dependent, with the highest gene expression observed at $\sim 20 \mu\text{g ml}^{-1}$ of soluble silica, accompanied by $60\text{-}90 \mu\text{gml}^{-1}$ of calcium ions¹⁷. A similar dose-dependent response was observed to the mature osteoblasts with $15\text{-}20 \mu\text{gml}^{-1}$ of soluble silica promoting highest metabolic activity and enhanced formation of mineralised bone nodules¹⁸. Osteostimulation is

Bioglass' second mode of bioactivity and has led to bioactive glasses to be made to contain other cations with therapeutic benefits, but they are yet to reach clinical use ¹⁹.

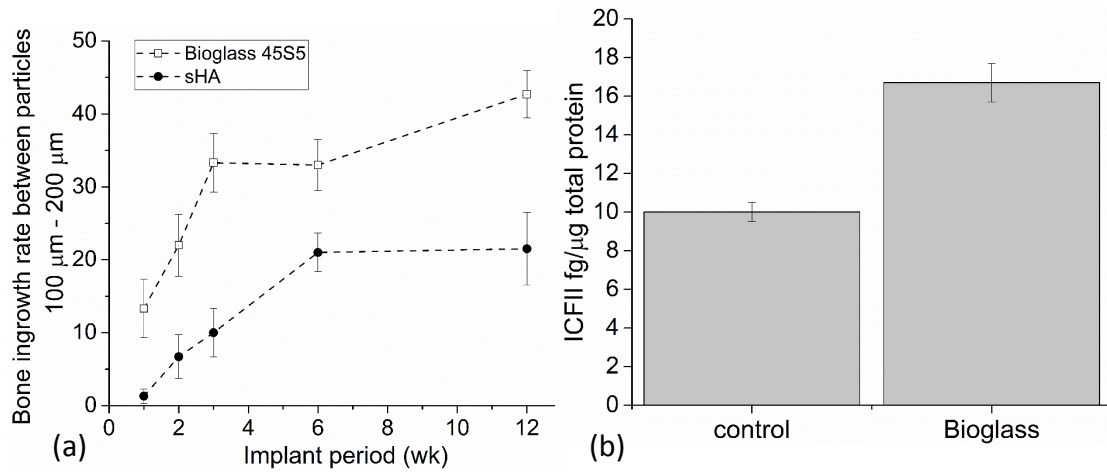


Figure 1. (a) Comparison of % bone ingrowth between particles of 45S5 Bioglass and synthetic HA (sHA) in a bone effect (rabbit femoral chondyle) from 1 to 12 weeks. Particle sizes were 100-300 μm. Data replotted from Oonishi *et al.*¹⁰; (b) Concentration of unbound insulin-like growth factor IGFII protein in culture media, produced from osteoblasts, comparing control medium to media containing Bioglass dissolution products. Data replotted from Xynos *et al.*¹⁵

Recently interest has increased in borate glasses ²⁰, largely due to very encouraging clinical results of healing of chronic wounds, such as diabetic ulcers that would not heal under conventional treatment ²¹. Phosphate glasses give the benefit of controllable total dissolution, but have not yet reached the clinic²². This article aims to summarise the currently available medical devices and products, for which the principle material is bioactive glass and then it discusses some applications that are likely to follow in the near future.

2. Synthetic bone graft granules

Synthetic bone grafts are designed to reduce use of autografts, where clinicians move bone from one part of a patient, usually the pelvis, to the defect site²³. Problems are that unnecessary bone is in limited supply and patients can experience pain and/or infection at the donor site, which then also needs to be repaired.

The original Bioglass 45S5 has been used in more than 1.5 million patients²⁴ in the form of a particulate, marketed under the name NovaBone® (NovaBone Products LLC, Jacksonville, FL), to repair bone defects in orthopaedics and maxillofacial reconstruction²⁵. However other products exist, based on 45S5 and also on other compositions.

The first clinical use of Bioglass was actually in the form of a monolith, in the form of cones that were used to replace the small bones in the middle ear of a patient. Infection had caused the bones to degrade, causing deafness. The Bioglass implant restored the patient's hearing²⁶. The Bioglass middle ear prosthesis (MEP®) was cast into shape from the melt. Ten year follow up studies showed 17 out of 21 retained function (the other four fractured), improving on polymeric, metallic and ceramic implants²⁶. The product (DOUEK MED™, USBiomaterials, Alachua, FL, Figure 2) contained several glass cones of different sizes to enable the clinician to choose appropriately for the patient.

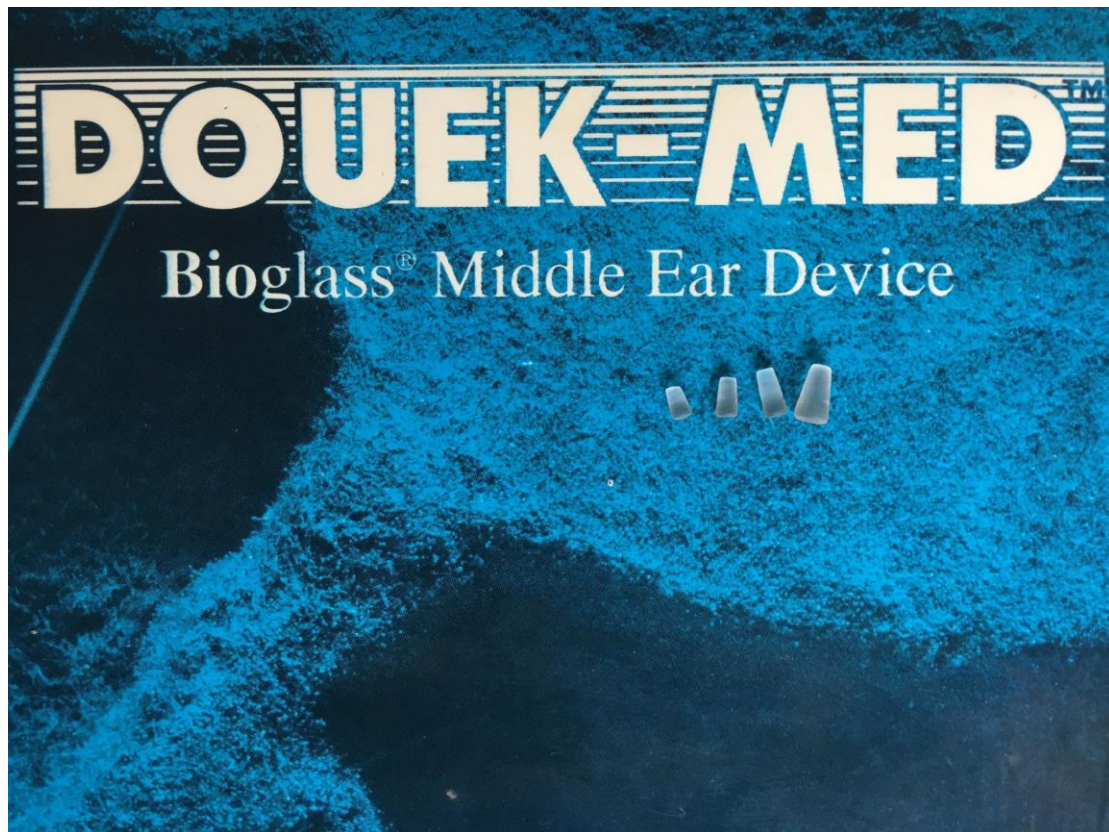


Figure 2. The four sizes of Bioglass cone shaped grafts, photographed (inset) on the packaging of the DOUEK-MED Bioglass Middle Ear device. Scale bar = 1 cm.

The second commercial Bioglass 45S5 device was also cones. The Endosseous Ridge Maintenance Implant [ERMI®] launched in 1988 were inserted into fresh tooth extraction sites (where the root would have been) to provide a stable platform for dentures. Five year follow up showed quantified improvements over synthetic HA implants²⁷.

These products are no longer in clinical use as surgeons want to be able to mould or cut bone grafts to shape. Providing the glass in the form of particles or granules meant that the glass could be pressed into a defect. Surgeons tend to mix the particles with blood from the patient to create a putty-like material (as the blood begins to clot), which is

pressed into the defect. The blood improves handling of the material and also contains natural growth factors and cells that can accelerate bone regeneration.

PerioGlas[®] was the first Bioglass particulate (90-710 μm), launched in 1993 by USBiomaterials, now sold by NovaBone Products LLC) as a synthetic bone graft for repair of bone defects in the jaw that resulted from periodontal disease, e.g. to regenerate bone around the root of a healthy tooth to save the tooth, or to repair bone in the jaw to allow the anchoring of titanium implants. Figure 3 shows its packaging and an SEM image of the particles. Clinical studies ²⁸⁻⁴⁰ showed that defects treated with PerioGlas were ~70% filled with new bone compared to ~35% for controls. The product has also been used with polymeric membranes, termed guided tissue regeneration ⁴¹.

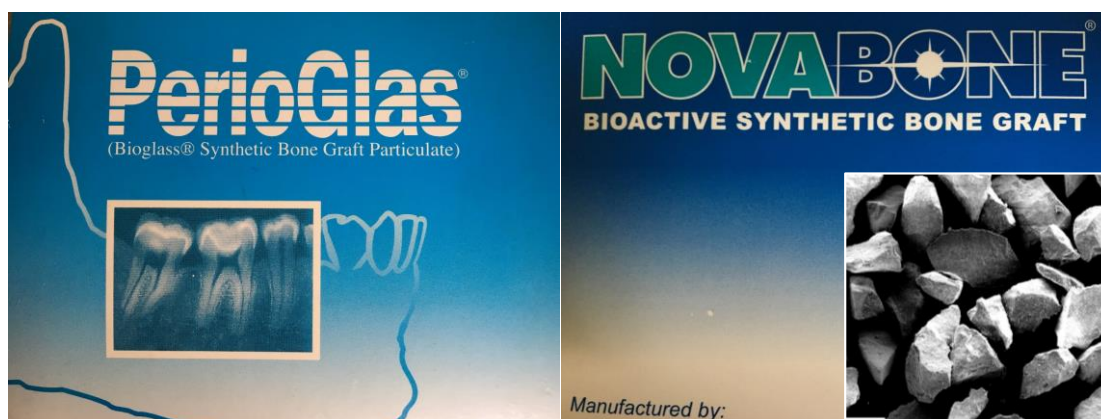


Figure 3. PerioGlas[®] and NovaBone[®] packaging. Inset is an SEM image of Bioglass particles of the equivalent particle size range of both products. Scale bar is 200 μm .

Due to the success of PerioGlas, a particulate for grafting of non-load bearing bone defects was released in 1999 named NovaBone (now distributed by NovaBone Products LLC). The particles have a similar distribution to PerioGlas (90-710 μm). FDA approval was granted for orthopaedic use in 2005. In clinical trials, NovaBone was

compared to autograft in posterior spinal fusion operations for treatment of adolescent idiopathic scoliosis (curvature of the spine). The NovaBone was mixed with the patient's blood and fixed in place by compressing the neighbouring vertebrae with metal screws and hooks⁴². NovaBone performed as well as autograft over the follow-up period of 4 years but with fewer infections (2% versus 5%) and fewer mechanical failures (2 versus 7.5%), with the main benefit that a donor site was not needed with NovaBone. The term "osteostimulation" was approved by the FDA in 2015.

Biogran[®] (Biomet 3i, Palm Beach Gardens, FL) is another 45S5 glass product used in jaw bone defect regeneration. It has a more narrow (300-350 μm) particle size range, which was chosen based on *in vivo* studies that indicated particles with diameters in that particle size range hollowed out within four weeks of implantation. The HCA layer formed and the silica dissolved.⁴³ The silica dissolution is attributed to the action of phagocytes. Clinical trials on 87 patients, where Biogran was compared to synthetic HA for bone defects in the jaw (alveolar ridge) left by cystic defects, extraction sites, and defects left by surgery, showed that Biogran outperformed HA⁴⁴. After 6 months, little difference could be seen between glass particles and bone by X-ray.

A variation of the 45S5 composition is the S53P4 composition (53.8 mol% SiO₂, 21.8 mol% CaO, 22.7 mol% Na₂O, 1.7 mol% P₂O₅ but usually quoted as 53 wt% SiO₂, 23 wt% CaO, 20 wt% Na₂O, 4 wt% P₂O₅), which is now known as BonAlive[®] (BonAlive Biomaterials, Turku, Finland). BonAlive (Figure 4) received European approval as a bone graft substitute for orthopaedic use in 2006 and FDA 510k approval in 2008. The BonAlive brand name was introduced in 2007. Small enterprises in Turku, first Abmin Technologies (1996) and then Vivoxid Ltd. (2002) started to produce S53P4 for clinical trials in University Hospitals in Finland, such as the Turku University and Helsinki University. BonAlive Biomaterials began trading in 2010 and BonAlive products are

now available in more than 50 countries and it is estimated to be used in >6000 procedures in 2016.

The S53P4 composition evolved in 1990 from studies investigating the relationship between the incorporation of alumina and borate in silicate glasses and their bioactivity⁴⁵. A rabbit tibia model showed that alumina was detrimental to bioactivity (it increases network connectivity) but good bone bonding was seen for the alumina (and borate) free S53P4⁴⁵. In addition, *in vitro* studies demonstrated that this composition possessed anti-microbial properties⁴⁶. Preclinical studies for spinal fusion⁴⁷, grafting⁴⁸ and sinus obliteration⁴⁹ followed in the 1990's. Key orthopaedic clinical indications for which BonAlive are now used are for synthetic bone grafting following tumour removal, trauma and to treat chronic osteomyelitis (bone infection, usually caused by bacteria). The published data (in terms of journal articles) on clinical trials of S53P4 is extensive compared to that of the original 45S5 Bioglass.



Figure 4. BonAlive (S53P4 granules) packaging and application demonstration of BonAlive putty (biodegradable polymer containing S53P4 glass particles) into a simulated bone defect. Inset: photograph of BonAlive granules. Photographs obtained with permission from bonalive.com.

Removal of benign bone tumors: S53P4 granules (1-4 mm, 14 patients) were compared with autograft (11 patients), for bone defects (1-30 cm³) left by benign bone-tumour surgery in hands, tibia (shin) and humerus (arm), with 14 year follow up ⁵⁰. Following S53P4 treatment, the cortical bone thickness was twice as thick as it was when autograft was used. However, some of the glass remained in the bone, even after 14 years. The glass was observed to begin to decrease in size (degrade) between 12-36 months and stimulated remodelling of the bone ⁵¹ but remodelling was slower than for autograft (at 12 months) ⁵². The glass did not disturb the growth of bone in children (which is often problematic with synthetic bone grafts, as seen following a trial on a three year old child (two year follow up) that had a cyst removed ⁵³.

Bone defects from trauma: S53P4 particles (0.83–3.15 mm) were compared to autograft in tibial fractures ⁵⁴ that required joint realignment. The grafts were placed inside the subchondral bone defects (in the crushed porous bone), supported by metal condylar plates and casts. Full weight bearing was allowed when radiographs indicated healing had occurred, so the implants were loaded. Eleven year follow-up showed similar bone regeneration and no difference in articular depression. Some glass particles were still present, even at 11 years post-operation ⁵⁵. The lack of resorption of S53P4 may be due to glass composition, which has higher silica content, and therefore higher network connectivity than 45S5.

Osteomyelitis: 11 patients with chronic osteomyelitis in the spine, where quality of the vertebrae was reduced due to bacterial infection ⁵⁶, were treated with S53P4 by filling cavity bone defects, with metallic stabilisation of the vertebrae. The most common pathogen was *Staphylococcus aureus*. Follow up was 10–38 months. Nine patients healed without complications, while the other two had unrelated complications.

Spondylolisthesis (displacement of the vertebral column): S53P4 granules of 1-2 mm were compared to autograft and held in position between vertebrae by compression of the vertebrae using metal screws. After 11 years, the fusion rate for the glass was 88% compared to 100% for autograft ⁵⁷.

Dental/maxillofacial: While the mandible (lower jaw), consists mainly of compact cortical bone that can be easily grafted, the maxilla (upper jaw) consists of porous cancellous bone that resorbs rapidly in periodontitis and is therefore more difficult to graft. Treatment is usually maxillary sinus floor lifting, where bone grows partially into the sinus cavity. Implantation of a mixture of S53P4 granules in combination with autograft allowed the implantation of titanium roots in the porous maxilla and showed more rapid bone repair with thicker trabeculae compared to autograft alone ⁵⁸.

Craniofacial: Sinus obliteration is a procedure that eliminates the frontal sinuses in order to prevent chronic infection or in response to trauma or tumour removal. Traditionally, the defect is filled with fat, but this lead to up to 25% of patients experiencing complications. Trials with S53P4 (0.5-1 mm size range) showed improved bone repair, in terms of quantity and quality, compared to synthetic HA ⁵⁹.

BonAlive has also successfully been used in trials for filling cavities in the middle ear created by surgeons removing mastoid air cells and mucous membranes that were damaged by chronic infection ⁶⁰.

Clinical results are good for the BonAlive (S53P4) and Bioglass 45S5 particulates. The two compositions been compared in very few *in vivo* studies. Bioglass 45S5 reacts more rapidly than S53P4, so when cones were implanted in rat bone defects, the HCA layer was thicker for 45S5 than for S53P4 ⁶¹. The original Bioglass degrades more rapidly due to its lower silica content and therefore lower network connectivity.

Clinicians would prefer to use a bioactive glass that is in the form of a putty and/or that has the porous structure of autograft (cancellous bone). NovaBone have produced porous constructs that they term NovaBone OS-Si⁺ Morsels (Figure 5a). The morsels (scaffolds) have interconnected pores and are 1-5 mm in diameter. However these morsels are no longer totally amorphous, as sintering the Bioglass particles causes crystallisation⁶², so they are glass-ceramics. Partial crystallisation can lead to instability as the residual amorphous regions degrade preferentially⁶³ but the morsels still biodegrade within 12 months.

More recently, other 45S5 particulates have reached market, such as Unigraft® (Unicare Biomedical, Laguna Hills, CA), which is available in particle size ranges of 200-400 µm and 200-600 µm and used mainly for periodontal bone regeneration. GlassBone (Noraker, Lyon, France, Figure 5a) is a 45S5 particulate for orthopaedic and cranio-maxilo-facial surgeries (Figure 5b). GlassBone is available in particle size ranges of 90–500 µm, 500–1000 µm and 1000–3150 µm. Having been launched in 2008, GlassBone has sold in excess of 25 000 units and is available in the European Union (EU), Mexico, Turkey, Iran and Taiwan.

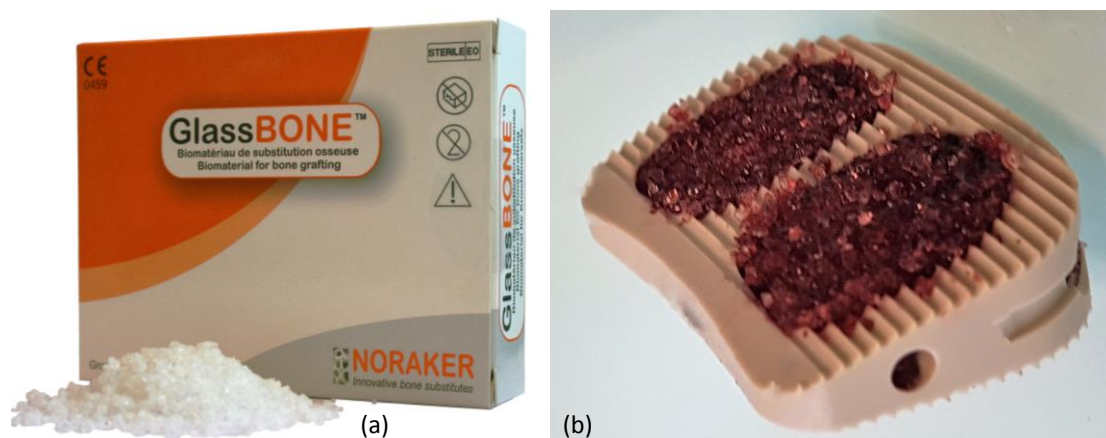


Figure 5. (a) GlassBone® particles and packaging and (b) demonstration of use of

GlassBone, mixed with blood and inserted in a spinal fusion cage. Photographs courtesy of Noraker.

3. Composites and putties for bone repair

Both NovaBone (45S5) and BonAlive (S53P4) glasses are available in bioresorbable putties (Figure 4 and Figure 6b).⁶⁴ NovaBone putties consist of a carrier matrix of polyethylene glycol (PEG) and glycerine, with 69 wt% Bioglass 45S5 (32 μm -710 μm , NovaBone Putty[®]) or a combination of 25 wt % macroporous morsels (500-700 μm) and 44 wt% Bioglass 45S5 (32 μm -710 μm), termed NovaBone Macroporous Putty (Figure 6b).

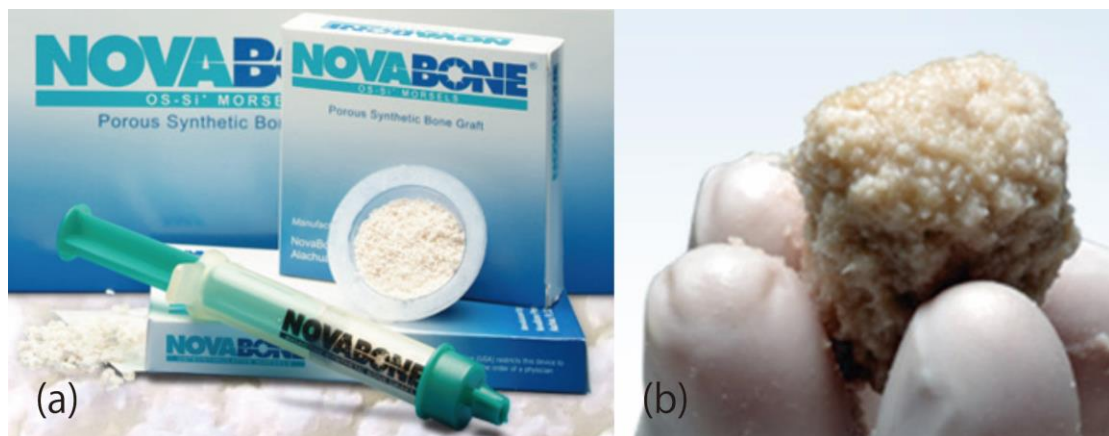


Figure 6. (a) NovaBone macroporous morsels; (b) NovaBone Putty[®] of PEG and glycerine containing Bioglass 45S5 particles; c) MacroFORM[™] composite of bovine collagen and Bioglass 45S5 particles. Photographs courtesy of NovaBone LLC.

Comparing putty to particulate in an ovine model, of 10 mm diameter critical sized defects in spine, bone defects filled with the putty filled with 42% bone compared to 20% in the defect filled with NovaBone particulate and 5% bone in the empty control⁶⁵. The putty matrix may separate the particles to allow more new bone to grow between

them than the tightly packed particles allowed. Signafuse[®] (Biostructures LLC, Newport Beach, CA) is a similar formulation with different particle size ranges. Synergy Biomedical (Collegeville, PA) also have a putty containing 45S5 Bioglass, but the glass is in the form of spheres (BioSphere[®], modal diameter ~400 µm), which they hypothesise leave interstices between the spheres during packing, which leaves space for bone ingrowth. Fibergraft[®] Putty (Prosidyan, Warren, NJ) also contains Bioglass spheres (Fibergraft BG), but the spheres have a unique architecture of a porous shell around a bundle of microfibres. The aim of the microfibres is to increase the surface area the glass and encourage bone ingrowth once the outer shell has degraded.

Bone graft products often combine the bioactive glass with a natural polymeric matrix, in an attempt to mimic natural bone, e.g. demineralised bone matrix (DBM), collagen, such as bovine collagen. An example is a mouldable composite of 90 wt% Bioglass particles (either NovaBone particles or macroporous morsels) in bovine hide collagen (Figure 4c). Other examples of similar products are: Vitoss[®] Bioactive Bone Graft (Stryker, NJ) and NanoFuse[®] (Amend Surgical, Alachua, FL). Clinical studies show NanoFuse (approved for orthopaedic and spinal use) improving bone ingrowth into a bone defect compared to the DBM alone⁶⁷. Vitoss Bioactive and BA2X, launched in 2011, are for filling bone defects and have 90% porosity in the DBM matrix. Vitoss BBTrauma (2012) is exclusively sold for trauma surgery and has a greater specific surface area of bioactive glass. Kinex[®] putty (Globus Medical Inc., Audubon, PA) combines higher concentrations of Bioglass (compared to with collagen with hyaluronic acid. All of these DBM based devices are a mixture of components rather than true composites.

Structural composites are needed that can take cyclic loads that contain bioactive glasses. Cortoss[®] (Stryker) is a polymethyl methacrylate (non-biodegradable) bone

cement that contains Bioglass particles, that is used to stabilise weakened vertebrae by filling the porous bone with cement. As it is not degradable, it is for bone augmentation, rather than regeneration. Noraker have developed a screw for anterior cruciate ligament reconstruction called LockActiv™, which is a poly(L-lactide)-co-poly(D,L)lactide (PLLA-co-PDLLA) polymer matrix containing 15wt% 45S5 Bioglass. LockActiv™ received its CE in 2015 and clinical trials are in progress. The screw must fix a detached ligament into a hole that the surgeon has drilled into the host bone. Screws that have a combination of the required mechanical properties to hold a ligament in place and that bond with the host bone and then biodegrade, successfully leaving the ligament integrated in the bone would fulfil an unmet clinical need. A future strategy is to produce inorganic/organic hybrids that have molecular level interactions between the glass and the polymer, but their journey to the clinic is likely to take 10 years⁶⁶.

4. Wound healing

Bioglass does not only have applications in orthopaedics, but also in soft tissue repair⁶. Exciting clinical results have been reported in human trials and in veterinary practices for treating chronic wounds with a cotton candy like scaffold made of biodegradable borate glasses²⁰. Studies included healing diabetic ulcers in human patients, which were not healing under conventional treatment²¹. There are no reports on the mechanism of action for why the scaffolds work so well, but the results are impressive. A product, RediHeal, is available to veterinary practices and FDA approval for the human product is pending.

5. Bioactive glass in toothpaste

Bioactive glass has also had an impact in consumer healthcare. The largest commercial use of bioactive glass, and perhaps any bioactive biomaterial, is in

toothpaste. Enamel and dentine of the tooth are very similar to bone, in that they contain HCA mineral and collagen. Up to 35% of the adult population are affected by dentine hypersensitivity, which is pain associated with chemical (e.g. acid) or thermal (e.g. hot or cold beverages) stimuli. The pain is usually explained by the dentine of the tooth, which is usually covered by enamel, becoming exposed. Dentine contains tubules that lead to the pulp cavity and nerves⁶⁸. Early toothpaste for hypersensitivity delivered anaesthetics during brushing. Recently, occlusion of the dentine tubules has become standard treatment⁶⁹ and toothpastes have been developed that can occlude tubules during brushing. Occlusion can be by physical occlusion by particles, stimulation of natural mineral (HCA) formation over and in the tubules, or a combination of the two.

Since 2004, a fine Bioglass 45S5 particulate, named NovaMin[®] (NovaMin Technology, FL, owned by GlaxoSmithKline, UK since 2010), has been used in certain toothpastes. NovaMin has a particle size (D₅₀ value) of ~18 µm and releases calcium and phosphate species during glass dissolution, which then form HCA on the dentine⁷⁰. NovaMin was first available in the USA in fluoride free toothpastes, but since the 2010 acquisition by GSK, it has been used in Sensodyne Repair and Protect[®] formulations (Figure 7a) that are “powered by NovaMin”, which are available in more than 20 countries. Clinical studies (>100 volunteers) showed improved pain relief when brushing with a NovaMin containing toothpaste compared to a toothpaste containing the anaesthetic potassium nitrate⁷¹. Figure 7b shows exposed dentine tubules following a brief acid etch. Figure 7c shows the NovaMin immediately after it was brushed onto the dentine in artificial saliva (AS). The particles attached and, within 24 h, the surface was almost completely covered by an HCA (Figure 7c) mineralisation.

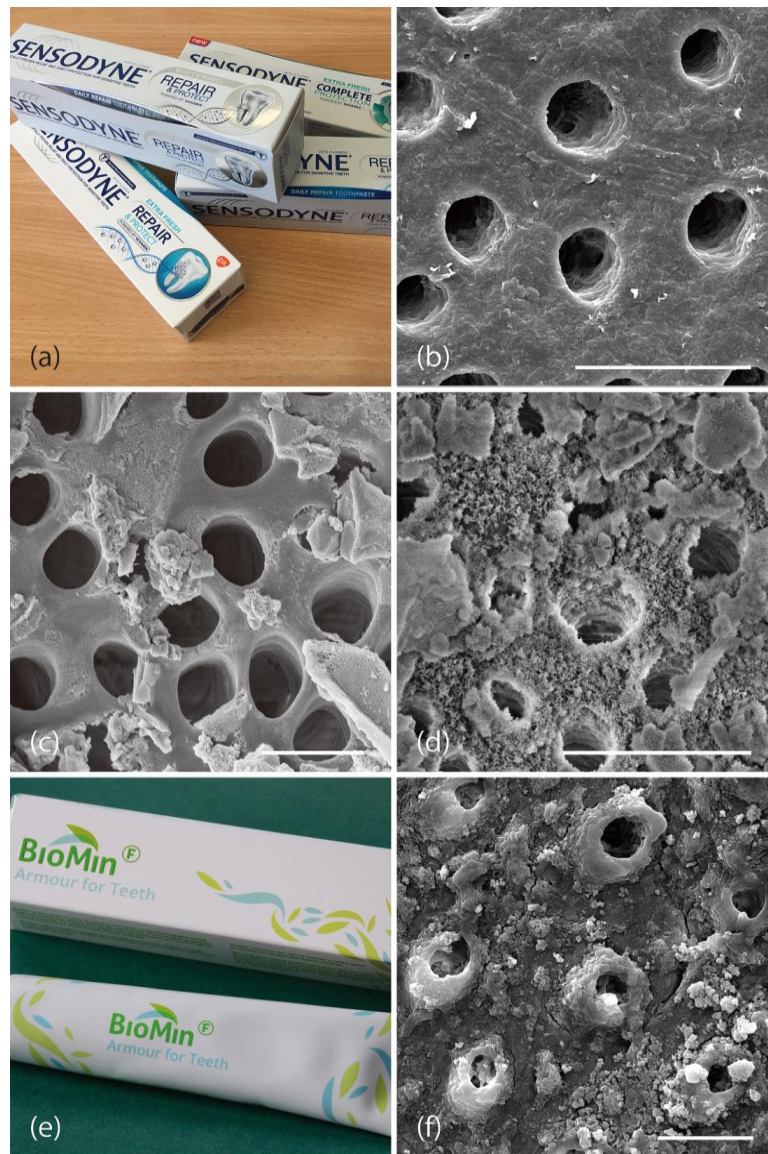


Figure 7. (a) Photograph of Sensodyne Repair and Protect[®] toothpastes that contain NovaMin (45S5) particles; (b-d) SEMs of human dentin: (b) untreated; (c) immediately after application of NovaMin in artificial saliva (AS); (d) 24 h after application of NovaMin in AS; (e) photograph of BioMinF toothpaste; (f) SEM image of dentine after 2 min brushing with BioMinF toothpaste (c,d) modified with permission from Earl *et al.* ⁶⁶; (b, f) provided by Prof. Robert Hill, Queen Mary, University of London/BioMin Technologies Ltd, UK). Scale = 5 μ m.

The success of NovaMin led to the development of more complex glass compositions, such as those designed to stimulate the formation of fluorapatite on the dentine, which is more resistant to acid attack than HCA ^{72,73}. Fluoride has long been identified as the key agent in preventing caries, as it inhibits tooth demineralisation. It also inhibits the metabolism of bacteria associated with caries, by preventing their metabolic acid production ⁷⁴. An example composition that incorporates CaF₂ in the composition is 36.41 mol% SiO₂, 28.28 mol% Na₂O, 24.74 CaO, 6.04 mol% P₂O₅, 4.53 mol% CaF₂ ⁷⁵. Fluoride-containing bioactive glasses released fluoride ions during dissolution ^{76,77}, which resulted in the formation of fluorapatite ^{75,78}. Studies showed that substituting CaF₂ at the expense of CaO increased glass dissolution ⁷⁹. The ability to form apatite increases with phosphate content, as long as the phosphate remains predominantly as orthophosphate in the glass⁸⁰. 6 mol% P₂O₅ seemed to favour fluoroapatite formation over fluorite ^{75,76}.

Based on this knowledge, a fluoride-releasing bioactive glass, BioMinF® (BioMin Technologies Ltd, London, UK) for use in toothpaste was developed ⁸¹. It differs from NovaMin by its higher phosphate content, the presence of CaF₂ in the glass and the smaller average particle size (D₅₀ of 6 μm). After two minutes of brushing dentine samples with BioMinF toothpaste (Figure 7e), small particles were seen to instantly occlude some of the tubules (Figure 7f) and the tubules remained occluded even after washing with 6% citric acid for 30 s. The BioMinF toothpaste was launched in 2016 in the UK (online only) and in pharmacies in Germany and India. Its efficacy in preventing or treating tooth decay, acid erosion or dentine hypersensitivity still needs to be demonstrated in clinical studies.

Dental care with bioactive glass is not limited to toothpaste. Bleaching teeth, which usually uses hydrogen peroxide, can demineralize enamel. *In vitro* studies indicate that NovaMin can repair the enamel through remineralisation to pre-bleaching levels (5 minute exposure and brushing) ⁸². Dentists can use air polishing to whiten teeth, which uses particles (traditionally sodium bicarbonate) as abrasives to remove stains, but the procedure is too painful for patients with hypersensitivity. Replacing the sodium bicarbonate with Bioglass 45S5 powder (Sylc, Denfotex, Ltd, UK) in the polishing procedure can stimulate mineralisation of dentine tubules in a similar mechanism to that of Novamin containing toothpaste ⁸³. Patients reported the Bioglass 45S5 polishing resulted in a 44% reduction in tooth sensitivity, and enhanced whiteness, according to their subjective scoring.

6. Cosmetics

Bioglass has recently been used in a number of cosmetic creams, particularly as Vitryxx® (Schott AG), a finely ground particulate (D₅₀ of 4 µm). Vitryxx is thought to have anti-ageing benefits, such as reducing redness and wrinkles. An example is Visible Youth Professional, a hyaluronic acid based gel that contains Vitryxx.

7. Summary

Bioactive glass is a key contributor to the Glass Age. It has improved the quality of life for millions of patients, regenerating bone faster and in some cases healing defects that would not heal by other means. The near future will see a glass revolution in other tissues, such as wound care, particularly reducing amputations arising from diabetic ulcers and sports injuries, including cruciate ligament damage and cartilage

tears. Applications will then extend to other soft tissues and delivery of therapeutic ions to treat a variety of conditions.

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References

1. L. L. Hench. Opening paper 2015- Some comments on Bioglass: Four Eras of Discovery and Development, *Biomedical Glasses* **1**(1) 1-11 (2015).
2. L. L. Hench. The story of Bioglass®, *Journal of Materials Science-Materials in Medicine* **17**(11) 967-978 (2006).
3. L. L. Hench, R. J. Splinter, W. C. Allen, T. K. Greenlee. Bonding mechanisms at the interface of ceramic prosthetic materials, *Journal of Biomedical Materials Research Biomedical Materials Symposium* **2**(1) 117-141 (1971).
4. T. Kokubo. Bioactive Glass-Ceramics - Properties and Applications, *Biomaterials* **12**(2) 155-163 (1991).
5. R. Z. LeGeros. Properties of osteoconductive biomaterials: Calcium phosphates, *Clinical Orthopaedics and Related Research* (395) 81-98 (2002).
6. ASTM. F1538-03 Glass and Glass Ceramic Biomaterials for Implantation. 2009.
7. V. Miguez-Pacheco, L. L. Hench, A. R. Boccaccini. Bioactive glasses beyond bone and teeth: Emerging applications in contact with soft tissues, *Acta Biomaterialia* **13** 1-15 (2015).
8. H. Oonishi, L. L. Hench, J. Wilson, F. Sugihara, E. Tsuji et al. Comparative bone growth behavior in granules of bioceramic materials of various sizes, *Journal of Biomedical Materials Research* **44**(1) 31-43 (1999).

9. H. Oonishi, S. Kushitani, E. Yasukawa, H. Iwaki, L. L. Hench et al. Particulate bioglass compared with hydroxyapatite as a bone graft substitute, *Clinical Orthopaedics and Related Research* (334) 316-325 (1997).
10. H. Oonishi, L. L. Hench, J. Wilson, F. Sugihara, E. Tsuji et al. Quantitative comparison of bone growth behavior in granules of Bioglass (R), A-W glass-ceramic, and hydroxyapatite, *Journal of Biomedical Materials Research* **51**(1) 37-46 (2000).
11. D. L. Wheeler, E. J. Eschbach, R. G. Hoellrich, M. J. Montfort, D. L. Chamberland. Assessment of resorbable bioactive material for grafting of critical-size cancellous defects, *Journal of Orthopaedic Research* **18**(1) 140-148 (2000).
12. L. L. Hench, J. M. Polak. Third-generation biomedical materials, *Science* **295**(5557) 1014-1017 (2002).
13. I. A. Silver, J. Deas, M. Erecinska. Interactions of bioactive glasses with osteoblasts in vitro: effects of 45S5 Bioglass (R), and 58S and 77S bioactive glasses on metabolism, intracellular ion concentrations and cell viability, *Biomaterials* **22**(2) 175-185 (2001).
14. I. D. Xynos, A. J. Edgar, L. D. K. Buttery, L. L. Hench, J. M. Polak. Gene-expression profiling of human osteoblasts following treatment with the ionic products of Bioglass (R) 45S5 dissolution, *Journal of Biomedical Materials Research* **55**(2) 151-157 (2001).
15. I. D. Xynos, A. J. Edgar, L. D. K. Buttery, L. L. Hench, J. M. Polak. Ionic products of bioactive glass dissolution increase proliferation of human osteoblasts and induce insulin-like growth factor II mRNA expression and protein synthesis, *Biochemical and Biophysical Research Communications* **276**(2) 461-465 (2000).
16. I. D. Xynos, M. V. J. Hukkanen, J. J. Batten, L. D. Buttery, L. L. Hench et al. Bioglass 45S5 stimulates osteoblast turnover and enhances bone formation in vitro: Implications and applications for bone tissue engineering, *Calcified Tissue International* **67**(4) 321-329 (2000).
17. L. L. Hench. Genetic design of bioactive glass, *Journal of the European Ceramic Society* **29**(7) 1257-1265 (2009).
18. O. Tsigkou, J. R. Jones, J. M. Polak, M. M. Stevens. Differentiation of fetal osteoblasts and formation of mineralized bone nodules by 45S5 Bioglass (R) conditioned medium in the absence of osteogenic supplements, *Biomaterials* **30**(21) 3542-3550 (2009).
19. A. Hoppe, N. S. Gueldal, A. R. Boccaccini. A review of the biological response to ionic dissolution products from bioactive glasses and glass-ceramics, *Biomaterials* **32**(11) 2757-2774 (2011).
20. M. N. Rahaman, D. E. Day, B. S. Bal, Q. Fu, S. B. Jung et al. Bioactive glass in tissue engineering, *Acta Biomaterialia* **7**(6) 2355-2373 (2011).
21. S. B. Jung, D. E. Day, T. Day, W. Stoecker, P. Taylor. Treatment of non-healing diabetic venous stasis ulcers with bioactive glass nanofibers, *Wound Repair and Regeneration* **19**(2) A30-A30 (2011).
22. E. A. Abou Neel, D. M. Pickup, S. P. Valappil, R. J. Newport, J. C. Knowles. Bioactive functional materials: a perspective on phosphate-based glasses, *Journal of Materials Chemistry* **19**(6) 690-701 (2009).
23. T. Kucera, K. Urban, S. Ragkou. Healing of cavitary bone defects, *European Journal of Orthopaedic Surgery and Traumatology* **22**(2) 123-128 (2012).

24. G. Pomrink. Personal Communication, *NovaBone Products LLC* (2016).
25. L. L. Hench. Bioactive materials for gene control. In: L. L. Hench, J. R. Jones, M. B. Fenn, editors. *New Materials and Technologies for Healthcare*. Singapore: World Scientific; 2011. p. 25-48.
26. K. R. Rust, G. T. Singleton, J. Wilson, P. J. Antonelli. Bioglass middle ear prosthesis: Long-term results, *American Journal of Otology* **17**(3) 371-374 (1996).
27. H. R. Stanley, M. B. Hall, A. E. Clark, C. J. King, L. L. Hench et al. Using 45S5 bioglass cones as endosseous ridge maintenance implants to prevent alveolar ridge resorption: A 5-year evaluation, *International Journal of Oral & Maxillofacial Implants* **12**(1) 95-105 (1997).
28. S. B. Low, C. J. King, J. Krieger. An evaluation of bioactive ceramic in the treatment of periodontal osseous defects, *International Journal of Periodontics & Restorative Dentistry* **17**(4) 359-367 (1997).
29. T. B. Lovelace, J. T. Mellonig, R. M. Meffert, A. A. Jones, P. V. Nummikoski et al. Clinical evaluation of bioactive glass in the treatment of periodontal osseous defects in humans, *Journal of Periodontology* **69**(9) 1027-1035 (1998).
30. E. S. Rosenberg, S. C. Cho, N. Elian, Z. N. Jalbout, S. Froum et al. A comparison of characteristics of implant failure and survival in periodontally compromised and periodontally healthy patients: A clinical report, *International Journal of Oral & Maxillofacial Implants* **19**(6) 873-879 (2004).
31. C. R. Anderegg, D. C. Alexander, M. Freidman. A bioactive glass particulate in the treatment of molar furcation invasions, *Journal of Periodontology* **70**(4) 384-387 (1999).
32. R. A. Yukna, G. H. Evans, M. B. Aichelmann-Reidy, E. T. Mayer. Clinical comparison of bioactive glass bone replacement graft material and expanded polytetrafluoroethylene barrier membrane in treating human mandibular molar Class II furcations, *Journal of Periodontology* **72**(2) 125-133 (2001).
33. J. S. Park, J. J. Suh, S. H. Choi, I. S. Moon, K. S. Cho et al. Effects of pretreatment clinical parameters on bioactive glass implantation in intrabony periodontal defects, *Journal of Periodontology* **72**(6) 730-740 (2001).
34. M. R. Norton, J. Wilson. Dental implants placed in extraction sites implanted with bioactive glass: Human histology and clinical outcome, *International Journal of Oral & Maxillofacial Implants* **17**(2) 249-257 (2002).
35. A. Sculean, G. Barbe, G. C. Chiantella, N. B. Arweiler, M. Berakdar et al. Clinical evaluation of an enamel matrix protein derivative combined with a bioactive glass for the treatment of intrabony periodontal defects in humans, *Journal of Periodontology* **73**(4) 401-408 (2002).
36. R. Mengel, M. Soffner, L. Flores-De-Jacoby. Bioabsorbable membrane and bioactive glass in the treatment of intrabony defects in patients with generalized aggressive periodontitis: Results of a 12-month clinical and radiological study, *Journal of Periodontology* **74**(6) 899-908 (2003).
37. S. J. Froum, M. A. Weinberg, D. Tarnow. Comparison of bioactive glass synthetic bone graft particles and open debridement in the treatment of human periodontal defects. A clinical study, *Journal of Periodontology* **69**(6) 698-709 (1998).
38. C. A. Shapoff, D. C. Alexander, A. E. Clark. Clinical use of a bioactive glass particulate in the treatment of human osseous defects, *Compendium of continuing education in dentistry (Jamesburg, NJ : 1995)* **18**(4) 352-358 (1997).

39. J. S. Zomet, U. R. Darbar, G. S. Griffiths, J. S. Bulman, U. Bragger et al. Particulate bioglass(R) as a grafting material in the treatment of periodontal intrabony defects, *Journal of Clinical Periodontology* **24**(6) 410-418 (1997).
40. J. S. Zomet, U. R. Darbar, G. S. Griffiths, W. Burgin, H. N. Newman. Particulate bioglass (Perioglas(R)) in the treatment of periodontal intrabony defects, *Journal of Dental Research* **76** 2219-2219 (1997).
41. V. S. Yadav, S. C. Narula, R. K. Sharma, S. Tewari, R. Yadav. Clinical evaluation of guided tissue regeneration combined with autogenous bone or autogenous bone mixed with bioactive glass in intrabony defects, *Journal of oral science* **53**(4) 481-488 (2011).
42. B. Ilharreborde, E. Morel, F. Fitoussi, A. Presedo, P. Souchet et al. Bioactive glass as a bone substitute for spinal fusion in adolescent idiopathic scoliosis a comparative study with iliac crest autograft, *Journal of Pediatric Orthopaedics* **28**(3) 347-351 (2008).
43. E. J. G. Schepers, P. Ducheyne. Bioactive glass particles of narrow size range for the treatment of oral bone defects: A 1-24 month experiment with several materials and particle sizes and size ranges, *Journal of Oral Rehabilitation* **24**(3) 171-181 (1997).
44. E. Schepers, P. Ducheyne, L. Barbier, S. Schepers. Bioactive glass particles of narrow size range: a new material for the repair of bone defects., *Implant Dentistry* **2**(3) 151-156 (1993).
45. O. H. Andersson, G. Z. Liu, K. H. Karlsson, L. Niemi, J. Miettinen et al. In vivo Behavior of Glasses in the SiO₂-Na₂O-CaO-P₂O₅-Al₂O₃-B₂O₃ System, *Journal of Materials Science-Materials in Medicine* **1**(4) 219-227 (1990).
46. P. Stoor, E. Soderling, J. I. Salonen. Antibacterial effects of a bioactive glass paste on oral microorganisms, *Acta Odontologica Scandinavica* **56**(3) 161-165 (1998).
47. N. C. Lindfors, K. Tallroth, A. J. Aho. Bioactive glass as bone-graft substitute for posterior spinal fusion in rabbit, *Journal of Biomedical Materials Research* **63**(2) 237-244 (2002).
48. J. T. Heikkila, H. J. Aho, A. Yliurpo, R. P. Happonen, A. J. Aho. Bone-formation in rabbit cancellous bone defects filled with bioactive glass granules, *Acta Orthopaedica Scandinavica* **66**(5) 463-467 (1995).
49. M. J. Peltola, K. M. J. Aitasalo, J. T. K. Suonpaa, A. Yli-Urpo, P. J. Laippala et al. Frontal sinus and skull bone defect obliteration with three synthetic bioactive materials. A comparative study, *Journal of Biomedical Materials Research Part B-Applied Biomaterials* **66B**(1) 364-372 (2003).
50. N. C. Lindfors, I. Koski, J. T. Heikkila, K. Mattila, A. J. Aho. A prospective randomized 14-year follow-up study of bioactive glass and autogenous bone as bone graft substitutes in benign bone tumors, *Journal of Biomedical Materials Research Part B-Applied Biomaterials* **94B**(1) 157-164 (2010).
51. N. C. Lindfors, J. T. Heikkila, I. Koski, K. Mattila, A. J. Aho. Bioactive Glass and Autogenous Bone as Bone Graft Substitutes in Benign Bone Tumors, *Journal of Biomedical Materials Research Part B-Applied Biomaterials* **90B**(1) 131-136 (2009).
52. N. C. Lindfors, J. T. Heikkila, A. J. Aho. Long-term evaluation of blood silicon and osteocalcin in operatively treated patients with benign bone tumors using bioactive glass and autogenous bone, *Journal of Biomedical Materials Research Part B-Applied Biomaterials* **87B**(1) 73-76 (2008).

53. N. C. Lindfors. Treatment of a recurrent aneurysmal bone cyst with bioactive glass in a child allows for good bone remodelling and growth, *Bone* **45**(2) 398-400 (2009).
54. K. Perna, I. Koski, K. Mattila, E. Gullichsen, J. Heikkila et al. Bioactive glass S53P4 and autograft bone in treatment of depressed tibial plateau fractures - A prospective randomized 11-year follow-up, *Journal of long-term effects of medical implants* **21**(2) 139-148 (2011).
55. J. T. Heikkila, J. Kukkonen, A. J. Aho, S. Moisander, T. Kyyronen et al. Bioactive glass granules: a suitable bone substitute material in the operative treatment of depressed lateral tibial plateau fractures: a prospective, randomized 1 year follow-up study, *Journal of Materials Science-Materials in Medicine* **22**(4) 1073-1080 (2011).
56. N. C. Lindfors, P. Hyvonen, M. Nyysönen, M. Kirjavainen, J. Kankare et al. Bioactive glass S53P4 as bone graft substitute in treatment of osteomyelitis, *Bone* **47**(2) 212-218 (2010).
57. J. Frantzen, J. Rantakokko, H. T. Aro, J. Heinanen, S. Kajander et al. Instrumented spondylodesis in degenerative spondylolisthesis with bioactive glass and autologous bone a prospective 11-year follow-up, *Journal of Spinal Disorders & Techniques* **24**(7) 455-461 (2011).
58. T. Turunen, J. Peltola, A. Yli-Urpo, R. P. Happonen. Bioactive glass granules as a bone adjunctive material in maxillary sinus floor augmentation, *Clinical Oral Implants Research* **15**(2) 135-141 (2004).
59. M. Peltola, K. Aitasalo, J. Suonpaa, M. Varpula, A. Yli-Urpo. Bioactive glass S53P4 in frontal sinus obliteration: A long-term clinical experience, *Head and Neck-Journal for the Sciences and Specialties of the Head and Neck* **28**(9) 834-841 (2006).
60. P. Stoor, J. Pulkkinen, R. Grenman. Bioactive Glass S53P4 in the Filling of Cavities in the Mastoid Cell Area in Surgery for Chronic Otitis Media, *Annals of Otolaryngology and Laryngology* **119**(6) 377-382 (2010).
61. L. Hupa, K. H. Karlsson, M. Hupa, H. T. Aro. Comparison of bioactive glasses in vitro and in vivo, *Glass Technology-European Journal of Glass Science and Technology Part A* **51**(2) 89-92 (2010).
62. Q. Z. Z. Chen, I. D. Thompson, A. R. Boccaccini. 45S5 Bioglass-derived glass-ceramic scaffolds for bone tissue engineering, *Biomaterials* **27**(11) 2414-2425 (2006).
63. O. Peitl, G. P. LaTorre, L. L. Hench. Effect of crystallization on apatite-layer formation of bioactive glass 45S5, *Journal of Biomedical Materials Research* **30**(4) 509-514 (1996).
64. M. A. Schallenberger, K. Rossmeier, H. M. Lovick, T. R. Meyer, H. M. Aberman et al. Comparison of the Osteogenic Potential of OsteoSelect Demineralized Bone Matrix Putty to NovaBone Calcium- Phosphosilicate Synthetic Putty in a Cranial Defect Model, *Journal of Craniofacial Surgery* **25**(2) 657-661 (2014).
65. Z. Wang, B. Lu, L. Chen, J. A. Chang. Evaluation of an osteostimulative putty in the sheep spine, *Journal of Materials Science-Materials in Medicine* **22**(1) 185-191 (2011).
66. J. R. Jones. Review of bioactive glass: From Hench to hybrids, *Acta Biomaterialia* **9**(1) 4457-4486 (2013).

67. J. F. Kirk, G. Ritter, C. Waters, S. Narisawa, J. L. Millan et al. Osteoconductivity and osteoinductivity of NanoFUSE (R) DBM, *Cell and Tissue Banking* **14**(1) 33-44 (2013).
68. J. S. Rees, M. Addy. A cross-sectional study of dentine hypersensitivity, *Journal of Clinical Periodontology* **29**(11) 997-1003 (2002).
69. R. Orchardson, D. G. Gillam. Managing dentin hypersensitivity, *Journal of the American Dental Association* **137**(7) 990-998 (2006).
70. D. G. Gillam, J. Y. Tang, N. J. Mordan, H. N. Newman. The effects of a novel Bioglass (R) dentifrice on dentine sensitivity: a scanning electron microscopy investigation, *Journal of Oral Rehabilitation* **29**(4) 305-313 (2002).
71. A. R. Pradeep, A. Sharma. Comparison of Clinical Efficacy of a Dentifrice Containing Calcium Sodium Phosphosilicate to a Dentifrice Containing Potassium Nitrate and to a Placebo on Dentinal Hypersensitivity: A Randomized Clinical Trial, *Journal of Periodontology* **81**(8) 1167-1173 (2010).
72. P. C. Lammers, I. M. P. M. Borggreven, F. C. M. Driessens, I. W. E. van Dijk. Influence of fluoride and carbonate on in vitro remineralization of bovine enamel, *Journal of Dental Research* **70** 970-974 (1991).
73. E. C. Moreno, M. Kresak, R. T. Zahradnik. Fluoridated hydroxyapatite solubility and caries formation, *Nature* **247** 64-65 (1974).
74. J. D. B. Featherstone. The science and practice of caries prevention, *Journal of the American Dental Association* **131**(7) 887-899 (2000).
75. M. Mneimne, R. G. Hill, A. J. Bushby, D. S. Brauer. High phosphate content significantly increases apatite formation of fluoride-containing bioactive glasses, *Acta Biomaterialia* **7**(4) 1827-1834 (2011).
76. D. S. Brauer, M. Mneimne, R. G. Hill. Fluoride-containing bioactive glasses: Fluoride loss during melting and ion release in tris buffer solution, *Journal of Non-Crystalline Solids* **357**(18) 3328-3333 (2011).
77. G. Lusvardi, G. Malavasi, L. Menabue, V. Aina, C. Morterra. Fluoride-containing bioactive glasses: Surface reactivity in simulated body fluids solutions, *Acta Biomaterialia* **5** 3548-3562 (2009).
78. D. S. Brauer, N. Karpukhina, M. D. O'Donnell, R. V. Law, R. G. Hill. Fluoride-containing bioactive glasses: Effect of glass design and structure on degradation, pH and apatite formation in simulated body fluid, *Acta Biomaterialia* **6** 3275-3282 (2010).
79. G. Lusvardi, G. Malavasi, L. Menabue, V. Aina, C. Morterra. Fluoride-containing bioactive glasses: Surface reactivity in simulated body fluids solutions, *Acta Biomaterialia* **5**(9) 3548-3562 (2009).
80. M. Eden. The split network analysis for exploring composition-structure correlations in multi-component glasses: I. Rationalizing bioactivity-composition trends of bioglasses, *Journal of Non-Crystalline Solids* **357**(6-7) 1595-1602 (2011).
81. R. Hill, D. Brauer, D. G. Gillam, N. Karpukhina, A. Bushby et al., inventors; Queen Mary and Westfield College, UK, assignee. Bioactive glass composition. WO patent WO 2011/161422 A1. 2011 29 December 2011.
82. J. S. Earl, R. K. Leary, K. H. Muller, R. M. Langford, D. C. Greenspan. Physical and chemical characterization of dentin surface, following treatment with NovaMin® technology, *Journal of Clinical Dentistry* **22** 2-67 (2011).

83. A. Banerjee, M. Hajatdoost-Sani, S. Farrell, I. Thompson. A clinical evaluation and comparison of bioactive glass and sodium bicarbonate air-polishing powders, *Journal of Dentistry* **38**(6) 475-479 (2010).

Captions

Figure 1. (a) Comparison of % bone ingrowth between particles of 45S5 Bioglass and synthetic HA (sHA) in a bone defect (rabbit femoral condyle) from 1 to 12 weeks.

Particle sizes were 100-300 μm . Data replotted from Oonishi *et al.*¹⁰; (b)

Concentration of unbound insulin-like growth factor IGFII protein in culture media, produced from osteoblasts, comparing control medium to media containing Bioglass dissolution products. Data replotted from Xynos *et al.*¹⁵

Figure 2. The four sizes of Bioglass cone shaped grafts, photographed (inset) on the packaging of the DOUEK-MED Bioglass Middle Ear device. Scale bar = 1 cm.

Figure 3. PerioGlas[®] and NovaBone[®] packaging. Inset is an SEM image of Bioglass particles of the equivalent particle size range of both products. Scale bar is 200 μm .

Figure 4. BonAlive (S53P4 granules) packaging and application demonstration of BonAlive putty (biodegradable polymer containing S53P4 glass particles) into a simulated bone defect. Inset: photograph of BonAlive granules. Photographs obtained with permission from bonalive.com.

Figure 5. (a) GlassBone[®] particles and packaging and (b) demonstration of use of GlassBone, mixed with blood and inserted in a spinal fusion cage. Photographs courtesy of Noraker.

Figure 6. (a) NovaBone macroporous morsels; (b) NovaBone Putty[®] of PEG and glycerine containing Bioglass 45S5 particles; (c) MacroFORM[™] composite of bovine collagen and Bioglass 45S5 particles. Photographs courtesy of NovaBone LLC.

Figure 7. (a) Photograph of Sensodyne Repair and Protect[®] toothpastes that contain NovaMin (45S5) particles; (b-d) SEMs of human dentin: (b) untreated; (c) immediately after application of NovaMin in artificial saliva (AS); (d) 24 h after application of NovaMin in AS; (e) photograph of BioMinF toothpaste; (f) SEM image of dentine after 2 min brushing with BioMinF toothpaste (c,d) modified with permission from Earl *et al.*⁶⁶; (b, f) provided by Prof. Robert Hill, Queen Mary, University of London/BioMin Technologies Ltd, UK). Scale = 5 μm .