doi:10.1093/brain/awt310 Brain 2014: 137; 172–182 | **172**



White matter integrity as a predictor of response to treatment in first episode psychosis

Tiago Reis Marques, ¹ Heather Taylor, ¹ Chris Chaddock, ¹ Flavio Dell'Acqua, ² Rowena Handley, ¹ A. A. T. Simone Reinders, ¹ Valeria Mondelli, ³ Stefania Bonaccorso, ¹ Marta DiForti, ¹ Andrew Simmons, ² Anthony S. David, ¹ Robin M. Murray, ¹ Carmine M. Pariante, ³ Shitij Kapur ¹ and Paola Dazzan ^{1,4}

- 1 Department of Psychosis Studies, Institute of Psychiatry, King's College London, UK
- 2 Department of Neuroimaging, Institute of Psychiatry, King's College London, UK
- 3 Department of Psychological Medicine, Institute of Psychiatry, King's College London, UK
- 4 National Institute for Health Research (NIHR) Mental Health Biomedical Research Centre at South London and Maudsley NHS Foundation Trust and King's College London, UK

Correspondence to: Tiago Reis Marques, MD, PO Box 063,
Department of Psychosis Studies,
Institute of Psychiatry;
De Crespigny Park,
London,
SE5 8AF,
UK

E-mail: tiago.marques@kcl.ac.uk

The integrity of brain white matter connections is central to a patient's ability to respond to pharmacological interventions. This study tested this hypothesis using a specific measure of white matter integrity, and examining its relationship to treatment response using a prospective design in patients within their first episode of psychosis. Diffusion tensor imaging data were acquired in 63 patients with first episode psychosis and 52 healthy control subjects (baseline). Response was assessed after 12 weeks and patients were classified as responders or non-responders according to treatment outcome. At this second time-point, they also underwent a second diffusion tensor imaging scan. Tract-based spatial statistics were used to assess fractional anisotropy as a marker of white matter integrity. At baseline, non-responders showed lower fractional anisotropy than both responders and healthy control subjects (P < 0.05; family-wise error-corrected), mainly in the uncinate, cingulum and corpus callosum, whereas responders were indistinguishable from healthy control subjects. After 12 weeks, there was an increase in fractional anisotropy in both responders and non-responders, positively correlated with antipsychotic exposure. This represents one of the largest, controlled investigations of white matter integrity and response to antipsychotic treatment early in psychosis. These data, together with earlier findings on cortical grey matter, suggest that grey and white matter integrity at the start of treatment is an important moderator of response to antipsychotics. These findings can inform patient stratification to anticipate care needs, and raise the possibility that antipsychotics may restore white matter integrity as part of the therapeutic response.

Keywords: antipsychotic drugs; psychosis; diffusion tensor imaging; white matter; schizophrenia

Abbreviation: PANSS = Positive and Negative Syndrome Scale

Introduction

Response to treatment in first episode of psychosis is heterogeneous, with approximately 55% of patients responding to antipsychotics in the first 12 months (Boter et al., 2009). The early discrimination between responders and non-responders is therefore of crucial importance, as it can reduce disability, healthcare costs, and eventually improve long-term outcome (Correll et al., 2003; Ascher-Svanum et al., 2008). With psychiatric neuroimaging moving towards approaches that may deliver clinically useful outcomes (Borgwardt and Fusar-Poli, 2012), attention has shifted to the identification of neuroimaging markers as potential early predictors of response. These could lead to a step forward in understanding not only the pathophysiology of the disorder, but most importantly, the neural basis of treatment response.

Most studies on neuroanatomical markers of response have investigated the role of grey matter (Bodnar et al., 2012), whereas little attention has been paid to white matter. This is surprising as loss of white matter integrity has been suggested as a key component of psychotic disorders (McGuire and Frith, 1996). In fact, reductions of fractional anisotropy, a marker of white matter microstructural integrity, are already apparent at the first psychotic episode (Pérez-Iglesias et al., 2010), before antipsychotic treatment (Cheung et al., 2008), and even in individuals at ultrahigh risk of developing psychosis (Carletti et al., 2012). Nevertheless, to date, only a handful of studies have examined the association between white matter and response to treatment, with inconsistent findings (Mitelman et al., 2006, 2009; Garver et al., 2008; Luck et al., 2011). Two cross-sectional studies, suggested an association between less extensive fractional anisotropy reductions and subsequent good clinical outcome, in the wholebrain and along tracts such as the uncinate and the superior longitudinal fasciculus (Mitelman et al., 2006; Luck et al., 2011). However, these studies did not perform a second evaluation of white matter at follow-up. Follow-up imaging studies can provide more information than cross-sectional studies, as they allow the identification of subtle within-subject anatomical changes and provide a true measure of the anatomical change over time that may relate to treatment outcome. The only two published longitudinal studies have been conducted in chronic patients, and they have reported conflicting results (Garver et al., 2008; Mitelman et al., 2009). The first study, in a small sample, showed that patients with chronic schizophrenia who responded to antipsychotics (n = 8), as assessed 28 days after the brain imaging, had reduced white matter microstructural integrity compared with nonresponders (n = 5), who were in turn no different from control subjects (Garver et al., 2008). In contrast, the second study, in a sample of chronic patients, assessed response 4 years after the brain imaging, and found that non-responders (n = 26) had lower frontal and parietal fractional anisotropy compared with responders (n = 23) (Mitelman et al., 2009). Therefore, possibly because of the small samples and the inclusion of chronic patients, it is difficult to disentangle which brain white matter changes may predict response to treatment. Importantly, recent animal studies indicate that antipsychotics may have a protective effect on white matter (Bartzokis, 2012; Zhang et al., 2012). Therefore, any

longitudinal change in fractional anisotropy may be, at least in part, attributable to antipsychotics. These questions can be best addressed by investigating patients with first episode psychosis, where treatment has occurred for only a short time, and the additional effect of illness duration on brain white matter is less likely to be present.

In this study, we evaluated a large sample (n = 63) of patients with first episode psychosis, at baseline and again at 12 weeks follow-up. To our knowledge, this is the first longitudinal diffusion tensor imaging study in first episode psychosis. To evaluate the relationship between white matter integrity and treatment response, we used a whole-brain skeleton-based technique (tractbased spatial statistics) (Smith et al., 2006). We hypothesized that: (i) patients would show a decrease in white matter microstructural integrity when compared with control subjects; (ii) patients' ability to respond to treatment would be associated with more marked alterations in white matter microstructural integrity at baseline; and (iii) antipsychotic treatment would be correlated with an improvement in white matter microstructural integrity at follow-up.

Materials and methods

Samples

Patients with first episode psychosis were recruited from the South London and Maudsley Foundation Trust. Healthy control subjects were recruited from the same catchment area. Healthy control subjects were administered the Psychosis Screening Questionnaire (Bebbington and Nayani, 1995), and excluded if they reported any psychotic symptom or a history of psychotic illness.

Exclusion criteria included: history of head trauma or injury with loss of consciousness >1 h; organic psychosis; learning disabilities; or lack of English fluency. Ethical permission was obtained from the Institute of Psychiatry Ethics Committee. After complete description of the study to the subjects, written informed consent was obtained.

Assessments

All patients underwent two clinical and MRI assessments: the first as soon as possible after their first contact with services, and the second 12 weeks after. This interval was chosen because of the clinical recommendation that antipsychotics should be continued for 6-8 weeks before switching to a different medication (Taylor et al., 2007); hence, a 12-week interval would provide information on treatment response after at least one drug taken for an appropriate period of time. Healthy control subjects had a single neuroimaging assessment. The mean interval between scans was 89 days (± 25.4 days). A total of 63 patients and 52 healthy control subjects underwent an MRI scan at baseline (Table 1); 53 patients were re-assessed at follow-up, as 10 patients could not be traced. Forty-eight patients agreed to undergo a second MRI scan: of these, six MRI scans could not be used because of movement artefacts, leaving 42 MRI scans for inclusion in the longitudinal neuroimaging analysis. To rule out potential systematic biases, we compared clinical and socio-demographic characteristics of patients who did and did not complete the second MRI, and found no significant differences. There were no differences between groups in the proportion of antipsychotic-naive subjects at baseline, nor in exposure to mood stabilizers, antidepressants or benzodiazepines (data not shown).

174 | Brain 2014: 137; 172–182 T. R. Marques *et al.*

Table 1 Demographic and clinical characteristics of responders, non-responders and healthy control subjects

	Responders (n = 30)	Non-responders (n = 33)	Control subjects (n = 52)	Test statistic
Age at scan (mean years, SD)	27.7 (7.1)	27.7 (9.2)	25.1 (6.5)	df (113); $F = 0.6$; $P = \text{n.s.}$
Sex (male/female)	18/12	22/11	25/27	df (2); $\chi^2 = 2.9$; $P = \text{n.s.}$
Handedness: right-handed (%)	89	94	82	df (2); $\chi^2 = 1.9$; $P = \text{n.s.}$
Education (mean years, SD)	13.9 (2.9)	12.8 (2.5)	14.9 (3.1)	df (85); $F = 17$; $P = 0.09$
Full scale IQ (mean, SD)	95 (14)	91 (15)	103 (16)	df (87); $F = 0.9$; $P = n.s.$
Diagnosis (% of affective psychosis)	87	75	n/a	df (1); $\chi^2 = 0.34$; $P = \text{n.s.}$
Schizophrenia	80	70		
Schizoaffective disorder	7	5		
Bipolar disorder	10	17		
Depressive disorder	3	8		
Use of cannabis (%)	77	70	48	df (113); $F = 4.0$; $P = 0.19$
Duration of untreated illness (duration of untreated psychosis) (mean days, SD)	116 (182)	152 (200)	n/a	df (55); $t = 0.6$; $P = n.s.$
PANSS total score, baseline (mean, SD)	54.5 (13.0)	62.7 (13.5)	n/a	df (61); t = 2.3; P = 0.06
PANSS total score, follow-up (mean, SD)	41.3 (5.7)	58.7 (10.8)	n/a	df (40); t = 2.5; P = 0.001
Total antipsychotic dose at baseline (mean CPZ equivalents, SD)	9608 (14026)	9583 (11569)	n/a	df (61); $t = 0.5$; $P = n.s$.
Antipsychotics used (% of atypicals)	93	91	n/a	df (1); $\chi^2 = 0.15$; $P = \text{n.s.}$
Total antipsychotic dose at follow-up (mean CPZ equivalents, SD)	27572 (19859)	31608 (28999)	n/a	df (40); $t = 0.2$; $P = n.s.$
Inter-scan interval (mean days; SD)	89 (32)	90 (17)	n/a	df (40); t = 1.3; P = n.s.

CPZ = chlorpromazine; n.s. = P-value not significant; P-value presented if P < 0.1.

Clinical assessment

ICD-10 diagnoses were formulated by psychiatrists using OPCRIT+ (McGuffin *et al.*, 1991), with good inter-rater reliability (kappa = 0.9). Psychotic symptoms were evaluated, at baseline and follow-up on the day of MRI using the Positive and Negative Syndrome Scale (PANSS; Kay *et al.*, 1987). This scale was also used to evaluate treatment response as the primary outcome measure, as required by the Remission criteria of the Schizophrenia Working Group Consensus (Andreasen *et al.*, 2005).

These criteria identify an absolute threshold in severity of symptoms that should be reached for clinical improvement. We preferred this methodology to symptom change cut-offs, which are arbitrary, may be affected by variability in baseline symptom severity and are not intuitively understood by clinicians.

According to these criteria, patients were defined as 'responders' if, at 12-weeks, they had a final score of mild or less on eight defined PANSS items. For those 10 patients who could not be re-assessed at 12 weeks, information on treatment response was obtained using the Personal and Psychiatric History Schedule (PPHS; Jablensky et al., 1992), which showed substantial agreement with PANSS score criteria (kappa = 0.72). Therefore, all patients assessed at baseline were classified according to their subsequent response: 30 as responders, 33 as non-responders. Duration of untreated psychosis was quantified as the interval between first psychotic symptoms and first contact with psychiatric services. Antipsychotic doses were converted to chlorpromazine equivalents (Atkins et al., 1997; Woods, 2003), and a total cumulative antipsychotic dose was calculated by summing all daily doses from the first day of treatment up to each MRI scan. Fiftyfive patients were taking atypical antipsychotics, five typical and three were medication-naïve. Premorbid IQ and handedness were assessed using the Wechsler Abbreviated Scale for Intelligence (Wechsler, 1997) and the Annett Hand Preference Questionnaire, respectively (Annett 1970).

Neuroimaging evaluation

Image acquisition

Data were acquired on a 3.0 T, GE Signa-HDx system running software release 14M5 with actively shielded magnetic field gradients (maximum amplitude 40 mT/m). Body coil was used for radiofrequency transmission, with 8-channel head coil for signal reception, allowing a parallel imaging (ASSET) speed-up factor of two. Each volume was acquired using a multi-slice peripherally-gated doubly refocused spin echo-echo planar imaging sequence, optimized for precise measurement of the diffusion tensor in parenchyma, from 60 contiguous near-axial slice locations with isotropic ($2.4 \times 2.4 \times$ 2.4 mm) voxels. Echo time was 104.5 ms and repetition time varied between 12-20 R-R intervals. Maximum diffusion-weighting was 1300 s/mm², and at each slice location, four images were acquired with no diffusion gradients applied, together with 32 diffusionweighted images with gradient directions uniformly distributed in space. An inhouse automated analysis technique assessed the quality of echo planar imaging data (Simmons et al., 1999). Acquisition was gated to the cardiac cycle using a peripheral gating device placed on the subjects' forefingers.

Image processing

Diffusion data were processed using ExploreDTI. Data were first preprocessed correcting for eddy current distortions and head motion. For each subject the b-matrix was then re-oriented to provide a more accurate estimate of tensor orientations. The diffusion-tensor was estimated using a non-linear least square approach, with fractional anisotropy calculated from the diffusion-tensor. Voxel-wise statistical analysis of fractional anisotropy was carried out using TBSS v1.2 (tract-based spatial statistics) (http://www.fmrib.ox.ac.uk/fsl/tbss/) (Smith, 2002; Smith *et al.*, 2004, 2006) to compare groups of diffusion-weighted measures. First, fractional anisotropy images were registered to standard MNI space using the non-linear registration tool in FSL (FNIRT) to the JHU-ICMB-DTI-81 atlas. A voxel-wise average of all subjects was used to create a study-specific mean fractional anisotropy image, which was then 'skeletonized' to create a mean fractional anisotropy skeleton, representing the centres of all white matter tracts. To exclude low anisotropic regions in the skeleton, a fractional anisotropy threshold of 0.3 was applied. Each subjects aligned fractional anisotropy maps were then projected onto this skeleton and results fed into voxel-wise cross-subject statistics

Diffusion tensor imaging tractography

To investigate whether differences in white matter microstructural integrity were related to changes in white matter volume, a tractography analysis of the tracts identified as most affected in the whole-brain comparisons between responders and non-responders was performed using TrackVis, with placement of regions of interest described elsewhere (Catani et al., 2002; Catani and Thiebaut de Schotten, 2008). For each tract the number of streamlines was calculated to obtain proxy measurements of tract volume.

Statistical analysis

Whole-brain statistical analyses were performed using Randomise v2.1 (FSL). Longitudinal changes in fractional anisotropy were assessed by subtracting the follow-up skeletonized fractional anisotropy image from each individual's baseline image. Statistical comparisons used the threshold-free cluster enhancement (TFCE), using a non-parametric permutation test, in which group membership was permuted 5000 times to generate a null distribution for each contrast. Significant clusters were identified where voxel-wise P-values were corrected for multiple comparisons [P < 0.05, family-wise error (FWE) corrected]. Age, gender and handedness were de-meaned before analysis and used as covariates of no-interest within the voxel-based analysis. Because of differences in the duration of untreated psychosis between groups, duration of untreated psychosis was calculated and added as a covariate after logarithmic transformation. To localize significant voxel effects, contrast maps were subdivided according to the 48 regions of the JHU-ICBM-DTI-81 white matter atlas. This allowed quantification of the number of significant voxels within each regional mask, and spatial identification of the peak voxel. We then extracted the individual's mean fractional anisotropy value for clusters identified as significantly different between responders and non-responders at baseline, and for the same clusters at follow-up, to quantify progressive fractional anisotropy change in these regions. For these clusters we also estimated the values of other white matter diffusion tensor imaging parameters: mean diffusivity (a composite measure as fractional anisotropy, which increases with the loss of structural barriers that normally restrict water diffusion); axial diffusivity (which is more specific to axonal integrity and can be considered an index of axonal damage); and perpendicular diffusivity, suggested to be a marker of reduction in the myelin content, hence representing an index of axonal demyelination) (Song et al., 2002, 2003).

Socio-demographic and clinical characteristics were analysed using one-way ANOVA for continuous variables, Mann-Whitney for skewed distributions and chi-square for categorical variables. Repeated measures ANOVA were used for the analysis of extracted fractional anisotropy values and tractography volumes. Analyses were conducted using SPSS v.18.0.

Results

Baseline differences in white matter microstructural integrity

Patients versus healthy control subjects

A total of 63 patients and 52 healthy control subjects underwent an MRI scan at baseline, and their demographic and clinical characteristics are presented in Table 1.

We first compared the white matter maps of the whole first episode psychosis patient group with the maps of the healthy controls at baseline. This comparison revealed areas of reduced fractional anisotropy values across the brain in first episode psychotic patients (P < 0.05; FWE-corrected), with the most affected tracts including corpus callosum, superior longitudinal fasciculus, corona radiata and left cingulum and thalamic radiation (Fig. 1) (for details of regions see Supplementary Table 1).

Responders versus non-responders

We then examined differences in white matter microstructural integrity at baseline between responders and non-responders, and found that non-responders had significantly lower fractional anisotropy across multiple brain regions when compared to responders (P < 0.05; FWE-corrected) (Fig. 2A). The most affected tracts comprised associative fibres, including the uncinate, stria terminalis and superior frontal-occipital tract; commissural fibres such as the corpus callosum; and several projections fibres, such as the internal and external capsule and corona radiata. The tracts with highest percentage of significant voxels were those interconnecting frontal and temporal cortices (uncinate and fornix) (Table 2). We re-ran this analysis including total baseline PANSS scores as a covariate, together with follow-up length, and the results remained significant, with an overlap in the affected regions. There were no regions in which nonresponders showed significantly higher fractional anisotropy than responders.

Non-responders versus healthy control subjects

Similarly, non-responders also had a significantly lower fractional anisotropy across multiple white matter regions when compared to healthy controls (P < 0.05; FWE-corrected). The most affected tracts included associative tracts, such as the left uncinate, cingulum and superior longitudinal fasciculus, and commissural tracts such as the corpus callosum (for details of the regions see Supplementary Fig. 1 and Supplementary Table 2) There were no white matter regions where controls had significantly lower fractional anisotropy values than non-responders.

Responders versus healthy control subjects

Interestingly, when we compared the baseline white matter maps of the responders to those of the healthy controls, we found no differences in fractional anisotropy values across the brain between these two groups.

176 | Brain 2014: 137; 172–182 T. R. Marques *et al.*

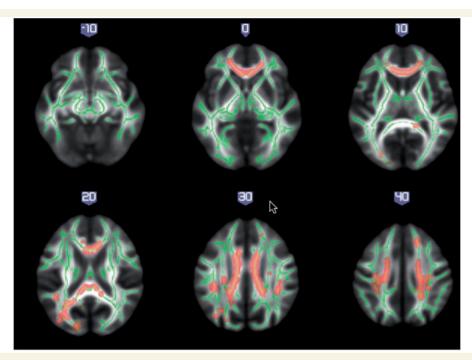


Figure 1 White matter maps showing significantly decreased fractional anisotropy in all patients when compared to healthy control subjects, at baseline (P < 0.05, FWE-corrected). Background image corresponds to the mean fractional anisotropy image in standard MNI152 brain space (radiological view). Fractional anisotropy white matter skeleton is represented by green voxels. Red–yellow voxels represent regions in which the fractional anisotropy was significantly lower in the patient group relative to the healthy control group.

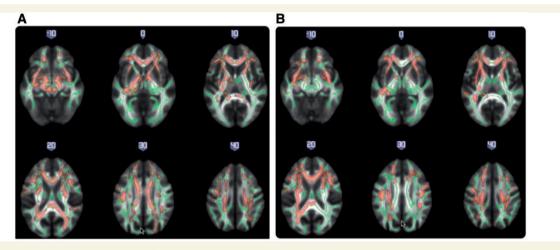


Figure 2 White matter maps showing significantly decreased fractional anisotropy in non-responders when compared with responders, at baseline (P < 0.05, FWE-corrected) (A); and at 12-weeks follow-up (P < 0.05, FWE-corrected) (B). Background image corresponds to the mean fractional anisotropy image in standard MNI152 brain space (radiological view). Fractional anisotropy white matter skeleton is represented by green voxels. Red-yellow voxels represent regions in which the fractional anisotropy was significantly lower in the non-responder group relative to the responder group.

Differences in white matter microstructural integrity at 12-weeks follow-up and longitudinal changes

We compared cross-sectionally the white matter microstructural integrity at follow-up, in those 42 subjects with two usable scans: 20 responders and 22 non-responders. Non-responders continued to

display lower fractional anisotropy compared with responders, mostly in the same brain tracts where they differed at baseline, including the uncinate, corona radiata, fornix, external and internal capsule and corpus callosum, although the differences appeared less widespread (P < 0.05; FWE-corrected) (Fig. 2B and Table 3). In addition, the cingulum and the superior and inferior longitudinal fasciculus were also affected (P < 0.05; FWE-corrected).

Table 2 White matter regions of fractional anisotropy reduction between non-responders and responders at baseline

Uncinate Left 70 91% 3.38 -33 -6 -14 Right 59 86% 2.24 35 -4 -11 Fornik Left \$70 76% 2.98 -34 -11 -15 Right stria terminalis 270 76% 2.98 -34 -11 -15 Right stria terminalis 11 4% 2.64 34 -11 -15 Right stria terminalis 11 4% 2.64 34 -11 -15 Right stria terminalis 13 5 51% 3.02 10 -27 -27 Left 203 75% 1.92 9 -29 -23 Right 135 51% 3.02 10 -27 -27 Cerebral peduncle Left 436 72% 2.35 -9 -29 -29 -20 Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 372 47% 2.44 -24 -19 0 Right anterior limb 372 47% 2.44 -24 -19 0 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left posterior 578 42% 3.30 2.20 -20 -20 -20 -20 -20 -20 -20 -20 -20 -	JHU white matter atlas region	Significant cluster size (n voxels)	% Region significant	t-statistic	MNI coordinates of peak voxel (mm)		
Left 70 91% 3.38 -33 -6 -14 Right 59 86% 2.24 35 -4 -14 -14 Right 59 86% 2.24 35 -4 -14 -14 -15 Fornix Left stria terminalis 270 76% 2.98 -34 -11 -15 Right stria terminalis 11 4% 2.64 34 -11 -15 Corticospinal Left 203 75% 1.92 9 -29 -23 Right 135 51% 3.02 10 -27 -27 Cerebral peduncle Left 436 72% 2.35 -9 -29 -20 Right 360 59% 3.40 10 -28 -19 External capsule Right 790 61% 4.36 -35 -8 -13 Left 790 61% 4.36 -35 -8 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left 906 85% 3.24 10 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1					x	у	z
Right 59 86% 2.24 35 —4 —14 Fornix Left stria terminalis 270 76% 2.98 —34 —11 —15 Right stria terminalis 11 4% 2.64 34 —11 —15 Corlicospinal Left 203 75% 1.92 9 —29 —29 —23 Right 135 —51% 3.02 10 —27 —27 Cerebral peduncle Left 436 72% 2.35 —9 —29 —29 —20 Right 360 59% 3.40 10 —28 —19 External capsule Right 781 67% 2.48 36 —7 —13 Left 790 66% 2.64 —20 —14 —0 Right anterior limb 575 66% 3.24 10 1 1 1 1 Left posterior limb 372 47% 2.44 —24 —19 0 Right anterior limb 361 43% 3.00 —13 —1 4 Left retrolenticular limb 372 47% 2.44 —24 —19 0 Left anterior limb 372 47% 2.44 —24 —19 0 Left anterior limb 361 43% 3.00 —13 —1 4 Corpus callosum Body 1985 63% 2.70 —13 17 24 Genu 1039 60% 3.55 —15 32 14 Splenium 1145 46% 2.13 1 —31 —31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 —15 33 15 Left posterior 364 48% 1.27 —21 —44 35 Left superior 578 42% 2.36 —22 —25 36 Right superior 578 42% 2.36 —22 —25 36 Right superior 99 48% 2.16 —5 —30 —60 Posting representations Left superior 99 48% 2.16 —5 —30 —21 Right superior 90 36% 2.46 7 —31 —46 Left inferior 66 33% 3.19 —23 —46 —36 Pontine cross tract 98 25% 2.35 —2 —30 —30 Middle 753 —33 —34 —5 —32 Right 30 —124 —4 —38 —37 Right 50 —5 —50 —50 —50 —50 Superior frontal occipital	Uncinate						
Formix Left stria terminalis 270 76% 2.98 -34 -11 -15 Right stria terminalis 11 4% 2.64 34 -111 -15 Corticospinal Left 203 75% 1.92 9 -29 -23 Right 135 51% 3.02 10 -27 -27 Cerebral peduncle Left 436 72% 2.35 -9 -29 -29 Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left 950 66% 2.64 -20 -14 0 Right atherior limb 480 58% 3.24 10 1 1 1 Left tertorlonticular limb 372 47% 2.44 -24 -19 0 Left atherior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right atherior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left anterior 942 56% 3.75 -15 33 15 Left superior 591 42% 3.21 21 -16 36 Right superior 591 42% 3.21 21 -16 36 Right superior 591 42% 3.21 21 -16 36 Right superior 99 48% 2.16 -5 -30 -21 Right superior 99 -36% 2.46 -7 -31 -16 Left superior 99 -36% 2.46 -7 -31 -45 Right superior 99 -36% 2.46 -7 -31 -46 Right superior 99 -36% 2.46 -7 -31 -46 Right superior 90 -36% 2.46 -7 -31 -		70	91%	3.38	-33	-6	-14
Left stria terminalis 270 76% 2.98 -34 -11 -15 Right stria terminalis 11 4% 2.64 34 -11 -15 Corpus calloum 1039 60% 3.55 -15 32 14 Corpus calloum 1039 60% 3.55 -15 32 14 Corpus calloum 1039 60% 3.55 -15 32 14 Corpus calloum 104 48% 1.27 -21 -24 Corpus calloum 104 48% 1.27 -21 -24 -38 Right anterior 1005 63% 3.26 27 28 10 Corpus calloum 139 60% 3.55 -15 33 15 Corpus calloum 144 60% 2.13 1 -31 -31 21 Corpus calloum 150 60% 3.55 -15 33 15 Corpus cal	Right	59	86%	2.24	35	-4	-14
Right stria terminalis 11 4% 2.64 34 -11 -15 Corticospinal Left 203 75% 1.92 9 -29 -23 Right 135 51% 3.02 10 -27 -27 -27 Right 135 51% 3.02 10 -27 -27 -27 Cerebral peduncle Left 436 72% 2.35 -9 -29 -29 Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -31 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left 790 61% 4.36 -35 -8 -13 Internal capsule Left 790 66% 2.64 -20 -14 0 1 1 1 Left retrolenticular limb 480 58% 3.24 10 1 1 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 1 1 Left attenior limb 361 43% 3.00 -13 -1 4 4 Corpus callosum 800 9 88 2.70 -13 17 24 Corpus callosum 1145 46% 2.13 1 -31 -31 21 Corpus caldista Right anterior 9 42 56% 3.75 -15 32 14 Splenium 1145 48% 1.27 -21 -44 35 Left posterior 364 48% 1.27 -21 -44 35 Right superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right superior 591 42% 3.21 21 -16 36 Right superior 99 48% 2.16 -5 -30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 99 48% 2.16 -5 -30 -30 -60 Portine cross tract 98 25% 2.35 -2 -30 -30 -30 Medial lermiscus Left 98 25% 2.35 -2 -30 -30 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 30 -20 -20 -30 Left 30 -20 -30 Left 30 -20 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 30 -20 -30 Left 50 -30 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 30 -20 -30 Left 50 -30 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 30 -20 -30 Left 50 -30 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 30 -20 -30 Left 50 -30 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 30 -20 -30 Left 50 -30 -30 Medial lermiscus Left 61 24% 2.13 -4 -38 -37 Right 50 -30 -30 Left 50 -3	Fornix						
Corticospinal Left 203 75% 1.92 9 -29 -23 Right 135 51% 3.02 10 -27 -27 Cerebral peduncle Left 436 72% 2.35 -9 -29 -20 Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Left corpus callosus Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 -31 -21 Corona tradiata Right anterior 1 005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left anterior 942 56% 3.75 -15 33 15 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 2.36 -22 -25 36 Right superior 591 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right superior 593 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 161 24% 2.13 -4 -38 -37 Right 30 12% 2.13 -	Left stria terminalis	270	76%	2.98	-34	-11	-15
Left 203 75% 1,92 9 -29 -23 Right 135 51% 3,02 10 -27 -27 Cerebral peduncle Left 436 72% 2,35 -9 -29 -20 Right 360 59% 3,40 10 -28 -19 External capsule Right 781 67% 2,48 36 -7 -13 Left 790 61% 4,36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2,64 -20 -14 0 Right anterior limb 480 58% 3,24 10 1	Right stria terminalis	11	4%	2.64	34	-11	-15
Right 135 51% 3.02 10 -27 -27 Cerebral peduncle Left 436 72% 2.35 -9 -29 -29 Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 10 Left anterior limb 361 43% 3.00 -13 -1 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 942 56% 3.75 -15 33 15 Left superior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right superior 99 48% 2.16 -5 -30 -21 Right superior 99 48% 2.16 -5 -30 -21 Right superior 99 48% 2.16 -5 -30 -21 Right superior 99 36% 2.56 2.35 -2 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 66 -34 -25 Superior frontal occipital	Corticospinal						
Cerebral peduncle Left 436 72% 2.35 -9 -29 -20 Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 1 Left tertorleticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 90 36% 2.46 7 -31 -16 Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	Left	203	75%	1.92	9	-29	-23
Left 436 72% 2.35 —9 —29 —20 Right 360 59% 3.40 10 —28 —19 External capsule Right 781 67% 2.48 36 —7 —13 Left 790 61% 4.36 —35 —8 —13 Internal capsule Left posterior limb 575 66% 2.64 —20 —14 0 1 1 1 Left retrolenticular limb 372 47% 2.44 —24 —19 0 Left action of the control limb 361 43% 3.00 —13 ——1 4 4 1 1 Left sterior limb 361 43% 3.00 —13 ——1 4 4 1 4 1 1 1 1 1 1 1 1 1 1 1 1 1 1	Right	135	51%	3.02	10	-27	-27
Right 360 59% 3.40 10 -28 -19 External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right superior 591 42% 3.21 21 -16 36 Right superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -46 Left superior 99 48% 2.16 -5 -30 -21 Right superior 99 48% 2.16 -5 -30 -21 Right superior 99 36% 2.46 7 -31 -46 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 1 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	Cerebral peduncle						
External capsule Right 781 67% 2.48 36 -7 -73 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left atterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left atterior 942 56% 3.75 -15 33 15 Left superior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -46 Left inferior 66 33% 3.19 -23 -46 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	Left	436	72%	2.35	-9	-29	-20
External capsule Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left atterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left superior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -46 Left superior 99 48% 2.16 -5 -30 -21 Right superior 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -38 Superior frontal occipital	Right	360	59%	3.40	10	-28	-19
Right 781 67% 2.48 36 -7 -13 Left 790 61% 4.36 -35 -8 -13 Internal capsule Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left superior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 99 36% 2.46 7 -31 -66 Left inferior 66 33% 3.3% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital							
Left 790 61% 4.36 —35 —8 —13 Internal capsule Left posterior limb 575 66% 2.64 —20 —14 0 Right anterior limb 480 58% 3.24 10 1 1 Left retrolenticular limb 372 47% 2.44 —24 —19 0 Left anterior limb 361 43% 3.00 —13 —1 4 Corpus callosum Body 1985 63% 2.70 —13 17 24 Genu 1039 60% 3.55 —15 32 14 Splenium 1145 46% 2.13 1 —31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left splenior 942 56% 3.75 —15 33 15 Left superior 978 42% 2.36 —22 —25 36 </td <td></td> <td>781</td> <td>67%</td> <td>2.48</td> <td>36</td> <td>-7</td> <td>-13</td>		781	67%	2.48	36	-7	-13
Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum 80dy 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left superior 364 48% 1.27 -21 -44 35 Left superior 591 42% 2.36 -22 -25 36 Right superior	_	790	61%	4.36	-35	-8	-13
Left posterior limb 575 66% 2.64 -20 -14 0 Right anterior limb 480 58% 3.24 10 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum 80dy 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left superior 364 48% 1.27 -21 -44 35 Left superior 591 42% 2.36 -22 -25 36 Right superior	Internal capsule						
Right anterior limb 480 58% 3.24 10 1 1 Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right superior 99 48% 2.16 -5 -	·	575	66%	2.64	-20	-14	0
Left retrolenticular limb 372 47% 2.44 -24 -19 0 Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left soperior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebel	•	480	58%	3.24	10	1	1
Left anterior limb 361 43% 3.00 -13 -1 4 Corpus callosum Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33%	O .	372	47%	2.44	-24	–19	0
Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corna radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16	Left anterior limb	361	43%	3.00		-1	4
Body 1985 63% 2.70 -13 17 24 Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16	Corpus callosum						
Genu 1039 60% 3.55 -15 32 14 Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle <t< td=""><td></td><td>1985</td><td>63%</td><td>2.70</td><td>–13</td><td>17</td><td>24</td></t<>		1985	63%	2.70	–13	17	24
Splenium 1145 46% 2.13 1 -31 21 Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle 2.67 30 -60 19 Cerebellar peduncle 2.67 30 -50 -21 Right superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -	,				– 15	32	14
Corona radiata Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus 2 2 2 -30 -34 <td></td> <td>1145</td> <td></td> <td></td> <td>1</td> <td>-31</td> <td>21</td>		1145			1	-31	21
Right anterior 1005 63% 3.26 27 28 10 Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle	•						
Left anterior 942 56% 3.75 -15 33 15 Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle 2.67 30 -60 19 Cerebellar peduncle 2.16 -5 -30 -21 Right superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus 2.16 -34 -35 -34 -25		1005	63%	3.26	27	28	10
Left posterior 364 48% 1.27 -21 -44 35 Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	· ·	942		3.75	– 15	33	15
Left superior 578 42% 2.36 -22 -25 36 Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital							35
Right superior 591 42% 3.21 21 -16 36 Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital 25 25 2.20 6 -34 -25							36
Right posterior 153 20% 2.67 30 -60 19 Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	•						36
Cerebellar peduncle Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	- ,						19
Left superior 99 48% 2.16 -5 -30 -21 Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	<u> </u>						
Right superior 90 36% 2.46 7 -31 -16 Left inferior 66 33% 1.97 -13 -45 -32 Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital	•	99	48%	2.16	-5	-30	-21
Left inferior 66 33 % 1.97 -13 -45 -32 Middle 753 33 % 3.19 -23 -46 -36 Pontine cross tract 98 25 % 2.35 -2 -30 -30 Medial lemniscus Left 61 24 % 2.13 -4 -38 -37 Right 30 12 % 2.10 6 -34 -25 Superior frontal occipital	•						-16
Middle 753 33% 3.19 -23 -46 -36 Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital							-32
Pontine cross tract 98 25% 2.35 -2 -30 -30 Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital							
Medial lemniscus Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital							
Left 61 24% 2.13 -4 -38 -37 Right 30 12% 2.10 6 -34 -25 Superior frontal occipital		20	25 / 5	2.55	_		
Right 30 12% 2.10 6 -34 -25 Superior frontal occipital		61	24%	2.13	_4	-38	-37
Superior frontal occipital							
	0	30	12 /0	2.10	9	5-7	23
10 25/0 1.32 25 2 19		18	23%	1 52	23	2	19
Left 9 10% 1.45 -21 5 19							19

JHU = Johns Hopkins University; % Region significant was obtained by dividing the number of significant voxels within each regional mask by the total number of voxels in that mask.

We then evaluated longitudinal within-group changes over the 12 weeks. A whole-brain analysis of within-group, and Group \times Time interaction for fractional anisotropy change did not identify any significant difference between responders and nonresponders. We then explored whether the regions identified at baseline as being significantly different between responders and non-responders at baseline had changed over time, and we found an overall increase in fractional anisotropy over the 12-weeks [F(1,42) = 16.7; P < 0.001]. Interestingly, there was no Group

(responders versus non-responders) × Time (baseline versus follow-up) interaction for these fractional anisotropy changes [F(1,42) = 2.53, P = 0.12], suggesting that changes were broadly in the same direction in both groups.

To explore if changes in white matter microstructural integrity were driven by volumetric changes, we explored whether there had been longitudinal changes in the volume of the most affected tracts. However, there was no significant longitudinal change in volume in any of the tracts analysed [right uncinate, F(1,41) = 0.367,

178 | Brain 2014: 137; 172–182 T. R. Marques *et al.*

Table 3 White matter regions of fractional anisotropy reduction between non-responders and responders at 12-weeks follow-up

JHU white matter atlas region	Significant cluster size (n voxels)	% Region significant	t-statistic	MNI coordinates of peak voxel (mm)		
				x	у	Z
Uncinate						
Left	58	81%	2.48	-34	-6	-14
Right	65	96%	1.88	36	-3	-16
External capsule						
Left	831	64%	4.61	-33	-6	-13
Right	341	29%	2.55	36	-7	-13
Corona radiata						
Left anterior	1014	59%	2.70	– 15	36	-1
Right anterior	978	59%	3.49	23	19	18
Left superior	760	54%	3.56	-22	-25	35
Right superior	292	21%	3.47	23	-19	38
Left posterior	406	53%	2.19	-26	-30	27
Right posterior	21	3%	1.76	19	-26	35
Fornix						
Left stria terminalis	202	58%	2.55	-28	-26	-8
Right stria terminalis	10	3%	2.63	34	-11	-15
Internal capsule						
Left anterior limb	446	53%	2.51	– 17	6	9
Right anterior limb	327	39%	3.13	21	4	15
Left retrolenticular limb	286	36%	2.01	-26	-24	8
Left posterior limb	160	18%	1.42	-22	_9	11
Right posterior limb	88	10%	2.25	20	-4	10
Corpus callosum						
Genu	806	46%	2.91	–13	33	-3
Body	725	23%	3.01	15	17	25
Splenium	377	15%	2.10	-23	-54	17
Superior frontal occipital	· · ·	.5 ,5	25		J.	
Left	34	35%	2.25	-21	14	19
Right	26	32%	1.27	23	2	19
Superior longitudinal fasci		32 /0	1.27	23	_	.,
Left	331	24%	2.36	-35	-25	35
Right	367	24%	1.89	38	-39	28
Thalamic radiation		, ,		55	3,5	20
Left posterior	207	19%	1.73	-28	-53	18
Inferior longitudinal fascio		10 /0	1.75	20	55	10
Left sagittal stratum	38	8%	2.54	-35	-11	-14
Right sagittal stratum	7	1%	2.54	-35 36	-11 -11	- 14 - 14
Cingulum	/	1 /0	2.19	30	-11	- 14
_	25	7%	2.30	10	17	26
Right	20	/ /0	2.30	10	17	20

JHU = Johns Hopkins University.

P=0.549; left uncinate, F(1,41)=0.681, P=0.415; or fornix, F(1,41)=0.808, P=0.374]. There was also no Group \times Time interaction for white matter tracts volume (P=0.44 to P=0.78) and no significant correlations were observed between longitudinal changes in volumes and longitudinal changes in fractional anisotropy or exposure to antipsychotics.

White matter microstructural integrity, exposure to antipsychotics and psychopathology

We further explored whether the fractional anisotropy differences we observed between responders and non-responders were accompanied by a change in other diffusion tensor imaging measures in these brain regions. Results showed that at baseline responders had significant lower mean and perpendicular diffusivity than non-responders in the white matter regions that differentiated these two groups $[F(1,63)=7.50,\ P<0.001;\$ and $F(1,63)=17.17,\ P<0.001,\$ respectively]. In contrast, there were no differences in axial diffusivity $[F(1,63)=2.374;\ P=0.1].$ Furthermore, we found an overall increase in axial diffusivity $[F(1,42)=0.97;\ P=0.32]$ and a decrease in mean diffusivity and perpendicular diffusivity $[F(1,42)=0.1.17,\ P=0.29;\$ and $F(1,42)=2.60,\ P=0.11,\$ respectively].

We then explored whether differences in fractional anisotropy could be explained as an effect of antipsychotic medication. We found that, at baseline, the cumulative dose of antipsychotic was not correlated with fractional anisotropy values in the regions that discriminated responders from non-responders. However, the longitudinal change in fractional anisotropy (between baseline and follow-up scans) within these brain regions was positively correlated with the cumulative antipsychotic dose at follow-up [r(40) = 0.37, P = 0.02], indicating that the higher the exposure to antipsychotic medication over the 12-week period, the larger the increase in fractional anisotropy (and therefore the improvement in white matter microstructural integrity) (Fig. 3). Additionally, for those same regions we also found a significant negative correlation between exposure to antipsychotic medication and perpendicular diffusivity [r(40) = -0.273, P = 0.046], but not with mean and axial diffusivity.

Finally, in the regions that discriminated responders from nonresponders at baseline, a significant negative correlation was observed between fractional anisotropy and baseline PANSS total score [r(63) = -0.273, P = 0.046], as well as between PANSS total scores and follow-up fractional anisotropy values in these regions [r(42) = -0.581, P < 0.001]. However, there was no correlation between change in psychotic symptoms and change in fractional anisotropy.

Discussion

To our knowledge, this is the first longitudinal study investigating white matter microstructural changes and treatment response in first episode psychosis. In this large sample, we found that nonresponders already have significantly impaired white matter microstructure at baseline, when compared with both healthy control subjects and future responders. In contrast, responders are not distinguishable from controls. Furthermore, fractional anisotropy differences between responders and non-responders remain present, albeit to a lower extent, even at follow-up. Interestingly, both patient groups show an increase in white matter microstructural integrity over time in these regions, which seems associated with the extent of exposure to antipsychotics.

Our main finding is that at the time of the first episode of illness, subsequent non-responders already show altered white matter microstructural integrity than responders. So far, results from the few existing studies assessing white matter microstructural integrity and clinical outcome have been inconsistent, possibly because of small samples, different follow-up periods and longer duration of illness. However, consistent with our findings, a cross-sectional tractography study showed that first episode

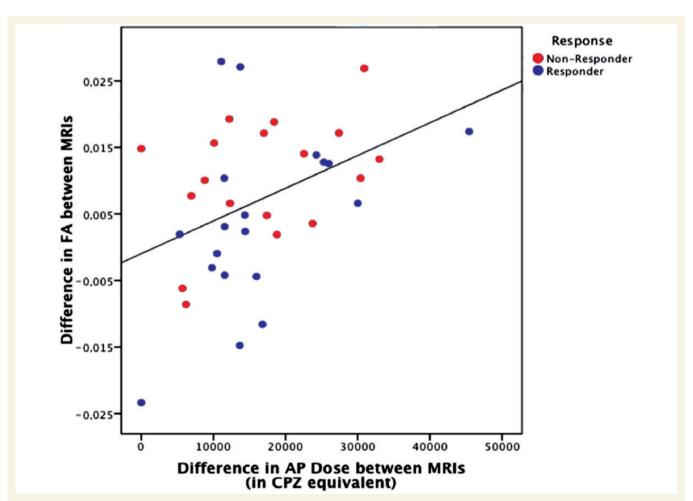


Figure 3 Relationship between total exposure to antipsychotic (AP) medications over the follow-up period [chlorpromazine (CPZ) equivalents] and longitudinal change in fractional anisotropy, in responders (blue) and non-responders (red).

180 Brain 2014: 137; 172–182 T. R. Marques *et al.*

patients who do not respond to 6 months of treatment have lower fractional anisotropy than responders in the superior longitudinal fasciculus and the uncinate (Luck *et al.*, 2011). The uncinate has been suggested to play an important role in the pathophysiology of psychosis and response to treatment (Van Veelen *et al.*, 2011) and indeed it was the most significantly affected tract in our study. In addition, we found substantial impairments in other tracts, such as the stria terminalis. This tract is part of the extended amygdala, and acts as a relay site within the main stress response system, the hypothalamic–pituitary–adrenal (HPA) axis. Alterations in the stress response have been linked to the pathophysiology of psychosis, as suggested by evidence of an enlarged pituitary gland and raised salivary cortisol in early stages of illness (Garner *et al.*, 2009; Mondelli *et al.*, 2010).

Interestingly, the cross-sectional comparison of the white matter maps obtained at the 12-week follow-up showed that the differences between responders and non-responders were still present, although less widespread. We found that higher PANSS scores were correlated with lower fractional anisotropy in the regions that differentiated non-responders from responders. This supports previous evidence of a negative correlation between fractional anisotropy and PANSS positive symptoms, mainly in the left uncinate and superior longitudinal fasciculus (Skelly et al., 2008). The presence of this negative correlation at both baseline and follow-up suggests that this association may represent a 'trait' marker, related to the underlying pathophysiology, which changes very little with treatment. However, we cannot exclude that the altered white matter integrity was already present in non-responders long before illness onset, reflecting a different, possibly neurodevelopmental origin. Further support for a heterogeneous pathophysiology across patients with psychosis comes from our finding that responders show no difference in white matter integrity from controls. This finding might also help to explain inconsistencies in previous diffusion tensor imaging studies of psychosis. Although studies in chronic patients suggest a reduced fractional anisotropy when compared with control subjects (Kubicki et al., 2005), studies in first episode are less consistent (White et al., 2011).

Finally, these findings extend and confirm our previous data on grey matter, in a different first episode psychosis sample, showing that patients who subsequently do not experience symptom remission are significantly different from both healthy controls and those who have an episodic illness course, whereas this latter group is indistinguishable from healthy control subjects (Mourao-Miranda et al., 2012). Consistently with those data, we have also found, in a sample partially overlapping with the one used in this study, that non-responders have prominent hypogyria at insular, frontal and temporal regions when compared with responders and healthy control subjects, whereas responders are not distinguishable from healthy control subjects (Palaniyappan et al., 2013). Taken together, this evidence supports the combined use of white and grey matter measures as potential early neuroimaging markers for short and long-term clinical outcome, which could potentially guide treatment decisions.

Intriguingly, although there were no whole-brain significant longitudinal differences between groups, there was an increase in fractional anisotropy values in the regions that differentiated responders and non-responders at baseline, which was positively

correlated with higher cumulative antipsychotic dose. Furthermore, the increase in fractional anisotropy and axonal diffusivity was accompanied by a decrease in mean and perpendicular diffusivity. This finding is difficult to interpret, because of the paucity of previous longitudinal studies. Cross-sectional studies suggest that chronic patients show more marked fractional anisotropy reductions than first episode patients, and the existing longitudinal studies also provide evidence of progressive changes in fractional anisotropy after illness onset (Mori et al., 2007; Garver et al., 2008; Mitelman et al., 2009; Wang et al., 2013). This was also evident in a recent longitudinal study in first episode schizophrenia, which identified a progressive reduction in fractional anisotropy at 6 weeks follow-up in the white matter microstructure around the bilateral anterior cingulate gyrus and the right anterior corona radiata (Wang et al., 2013). It is therefore intriguing to speculate whether the increase we observed reflects a restoration of white matter microstructural integrity, promoted by some 'protective' factors activated by antipsychotic medications. The impact of antipsychotics on white matter remains controversial. Recently, it has been suggested that first episode patients who receive more antipsychotics show a greater reduction in white matter volume (Ho et al., 2011), whereas other studies report no relationship between fractional anisotropy and dose of antipsychotic medication (White et al., 2011; Wang et al., 2013). In contrast, some studies suggested that antipsychotics have a pro-myelinating effect, promoting myelin repair and oligodendrocyte differentiation (Xiao et al., 2008), thus improving white matter microstructural integrity (Bartzokis, 2012). Similarly, a diffusion tensor imaging animal study showed that mice that were concomitantly treated with an antipsychotic and cuprizone, a copper chelator that induces oligodendrocyte loss and demyelination, showed an attenuation of the cuprizone-induced white matter changes compared with mice receiving cuprizone alone (Chandran et al., 2012). In patients, antipsychotics have been shown to increase white matter volume and intracortical myelin (Bartzokis et al., 2009). However, only the aforementioned study by Garver et al. (2008) has used diffusion tensor imaging to evaluate patients before and after antipsychotics. This study showed a reduction in mean diffusivity over time in responders, interpreted as reflecting a partial restoration of myelin integrity. Our results extend these findings to first episode patients, using fractional anisotropy as well as other diffusion tensor measures. In our study, there was a positive correlation between fractional anisotropy and exposure to antipsychotics, together with a negative correlation between perpendicular diffusivity and exposure to these drugs. These data are important, because although fractional anisotropy can be regarded as a weighted average of the different eigenvalues, perpendicular diffusivity may be modulated by myelin quantity and has been suggested to be a marker of myelin content (Song et al., 2002, 2003). Therefore, the longitudinal diffusion tensor imaging changes we observed could represent an effect of antipsychotics on white matter, potentially resulting from changes in myelin content. Finally, it is unlikely that the longitudinal fractional anisotropy change we observed was related to volumetric changes, as we found no significant difference in white matter volume over time in the most affected tracts. This suggests that

a reduction in white matter volume is unlikely to have driven our findings.

Our study has a number of strengths. This is the first diffusion tensor imaging study in first episode psychosis to use a longitudinal design to explore the relationship to treatment outcome. Second, this is the largest diffusion tensor imaging study conducted to date in first episode psychosis, as previous studies used an average sample size of 30 patients. Third, our study highlights the potential relationship between white matter and antipsychotics, supporting a role for these drugs in restoring white matter microstructural integrity, while informing the neurobiological basis of treatment response. The effect of antipsychotics in white matter has been a matter of strong debate, and our study supports a potential beneficial role of these drugs on this brain functional component. Finally, this is one of the few applications of tract-based spatial statistics to first episode psychosis and the first time tract-based spatial statistics has been used to evaluate treatment response in this particular patient population. Tractbased spatial statistics offers several advantages over standard voxel-based analytic techniques, particularly relevant to longitudinal analyses, as it removes the need for spatial smoothing and minimizes the methodological pitfalls caused by misalignment and misregistration, consequently increasing the sensitivity and interpretability of findings. Also, compared to standard tractography techniques, tract-based spatial statistics is a fully automated process, which allows the whole brain to be investigated, whereas tractography, as a traditional region of interest approach, only provides information on pre-selected regions, and suffers from intrinsic operator variability.

In terms of limitation, 10 of our patients could not be contacted at follow-up, and we cannot exclude that they may have represented a different group in terms of their white matter microstructural integrity. However, their demographic and clinical characteristics were similar to the remaining sample, and this bias is therefore unlikely.

In conclusion, this work provides evidence that white matter microstructural integrity can be helpful in the early identification of patients less likely to respond to antipsychotic drugs. In combination with other neuroimaging and clinical measures, these findings could considerably help patient stratification in psychiatry, ultimately allowing individualized patient management from the time of the first presentation to services. Furthermore, by identifying underlying brain structure and connectivity as potential factors moderating antipsychotic response, it raises the question as to whether treatments that enhance and restore brain connectivity may elicit a better response to antipsychotics in those who currently do not respond to them.

Funding

This work was supported by the National Institute for Health Research (NIHR) Mental Health Biomedical Research Centre at South London and Maudsley NHS Foundation Trust and King's College London. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health. The study was also partially supported by

a King's College London Translational Research Grant to P. Dazzan. P. Dazzan's research is also supported by the Psychiatry Research Trust. Tiago Reis Marques and P. Dazzan's research are supported by NARSAD. A.A.T. Simone Reinders is supported by the Netherlands Organization for Scientific Research (NWO-VENI grant no. 451-07-009).

Supplementary material

Supplementary material is available at Brain online.

References

- Andreasen NC, Carpenter WT Jr, Kane JM, Lasser RA, Marder SR, Weinberger DR. Remission in schizophrenia: proposed criteria and rationale for consensus [Review]. Am J Psychiatry 2005: 162: 441-9.
- Annett M. A classification of hand preference by association analysis. Br J Psychol 1970; 61: 303-21.
- Ascher-Svanum H, Nyhuis AW, Faries DE, Kinon BJ, Baker RW, Shekhar A. Clinical, functional, and economic ramifications of early nonresponse to antipsychotics in the naturalistic treatment of schizophrenia. Schizophr Bull 2008; 34: 1163-71.
- Atkins M, Burgess A, Bottomley C, Riccio M. Chlorpromazine equivalents: a consensus of opinion for both clinical and research implications. Psychiatr Bull 1997; 21: 224-6.
- Bartzokis G, Lu PH, Stewart SB, Oluwadara B, Lucas AJ, Pantages J, et al. In vivo evidence of differential impact of typical and atypical antipsychotics on intracortical myelin in adults with schizophrenia. Schizophr Res 2009; 113: 322-31.
- Bartzokis G. Neuroglialpharmacology: myelination as a shared mechanism of action of psychotropic treatments. Neuropharmacology 2012; 62 2137-53
- Bebbington PE, Nayani T. The psychosis screening questionnaire. Int J Methods Psychiatr Res 1995; 5: 11-20.
- Bodnar M, Achim AM, Malla AK, Joober R, Benoit A, Lepage M. Functional magnetic resonance imaging correlates of memory encoding in relation to achieving remission in first episode schizophrenia. Br J Psychiatry 2012; 200: 300-7.
- Borgwardt S, Fusar-Poli P. Third-generation neuroimaging in early schizophrenia: translating research evidence into clinical utility. Br J Psychiatry 2012; 200: 270-2.
- Boter H, Peuskens J, Libiger J, Fleischhacker WW, Davidson M, Galderisi S, et al. Effectiveness of antipsychotics in first episode schizophrenia and schizophreniform disorder on response and remission: an open randomized clinical trial (EUFEST). Schizophr Res 2009; 115: 97-103.
- Carletti F, Woolley JB, Bhattacharyya S, Perez-Iglesias R, Fusar Poli P, Valmaggia L, et al. Alterations in white matter evident before the onset of psychosis. Schizophr Bull 2012; 38: 1170-9.
- Catani M, Howard RJ, Pajevic S, Jones DK. Virtual in vivo interactive dissection of white matter fasciculi in the human brain. Neuroimage 2002; 17: 77-94.
- Catani M, Thiebaut de Schotten M. A diffusion tensor imaging tractography atlas for virtual in vivo dissections. Cortex 2008; 44: 1105-32.
- Chandran P, Upadhyay J, Markosyan S, Lisowski A, Buck W, Chin CL, et al. Magnetic resonance imaging and histological evidence for the blockade of cuprizone-induced demyelination in C57BL/6 mice. Neuroscience 2012; 202: 446-53.
- Cheung V, Cheung C, McAlonan GM, Deng Y, Wong JG, Yip L, et al. A diffusion tensor imaging study of structural dysconnectivity in nevermedicated, first episode schizophrenia. Psychol Med 2008; 38: 877-85.

182 Brain 2014: 137; 172–182 T. R. Marques *et al.*

Correll CU, Malhotra AK, Kaushik S, McMeniman M, Kane JM. Early prediction of antipsychotic response in schizophrenia. Am J Psychiatry 2003; 160: 2063–5.

- Garner B, Berger GE, Nicolo JP, Mackinnon A, Wood SJ, Pariante CM, et al. Pituitary volume and early treatment response in drug-naïve first episode psychosis patients. Schizophr Res 2009: 113: 65–71.
- Garver DL, Holcomb JA, Christensen JD. Compromised myelin integrity during psychosis with repair during remission in drug-responding schizophrenia. Int J Neuropsychopharmacol 2008; 11: 49–61.
- Ho BC, Andreasen NC, Ziebell S, Pierson R, Magnotta V. Long-term antipsychotic treatment and brain volumes: a longitudinal study of first episode schizophrenia. Arch Gen Psychiatry 2011; 68: 128–37.
- Jablensky A, Sartorius N, Ernberg G, Anker A, Cooper JE, Day R, et al. Schizophrenia: manifestations, incidence and course in different cultures. A World Health Organization ten-country study. Psychol Med Monogr Suppl 1992; 20: 1–97.
- Kay SR, Fiszbein A, Opler LA. The positive and negative syndrome scale (PANSS) for schizophrenia. Schizophr Bull 1987; 13: 261–76.
- Kubicki M, McCarley RW, Shenton ME. Evidence for white matter abnormalities in schizophrenia. Curr Opin Psychiatry 2005; 18: 121–34.
- Luck D, Buchy L, Czechowska Y, Bodnar M, Pike GB, Campbell JS, et al. Fronto-temporal disconnectivity and clinical short-term outcome in first episode psychosis: a DTI-tractography study. J Psychiatr Res 2011; 45: 369–77.
- McGuffin P, Farmer A, Harvey I. A polydiagnostic application of operational criteria in studies of psychotic illness. Development and reliability of the OPCRIT system. Arch Gen Psychiatry 1991; 48: 764–70.
- McGuire PK, Frith CD. Disordered functional connectivity in schizophrenia. Psychol Med 1996; 26: 663–7.
- Mitelman SA, Newmark RE, Torosjan Y, Chu KW, Brickman AM, Haznedar MM, et al. White matter fractional anisotropy and outcome in schizophrenia. Schizophr Res 2006; 87: 138–59.
- Mitelman SA, Canfield EL, Newmark RE, Brickman AM, Torosjan Y, Chu KW, et al. Longitudinal assessment of gray and white matter in chronic schizophrenia: a combined diffusion-tensor and structural magnetic resonance imaging study open. Neuroimag J 2009; 3: 31–47.
- Mondelli V, Dazzan P, Hepgul N, Di Forti M, Aas M, D'Albenzio A, et al. Abnormal cortisol levels during the day and cortisol awakening response in first episode psychosis: the role of stress and of antipsychotic treatment. Schizophr Res 2010; 116: 234–42.
- Mori T, Ohnishi T, Hashimoto R, Nemoto K, Moriguchi Y, Noguchi H, et al. Progressive changes of white matter integrity in schizophrenia revealed by diffusion tensor imaging. Psychiatry Res 2007; 154: 133–45.
- Mourao-Miranda J, Reinders AA, Rocha-Rego V, Lappin J, Rondina J, Morgan C, et al. Individualized prediction of illness course at the first psychotic episode: a support vector machine MRI study. Psychol Med 2012; 42: 1037–47.
- Palaniyappan L, Reis Marques T, Taylor H, Handley R, Mondelli V, Bonaccorso S, et al. Cortical folding defects as markers of poor

- treatment response in first episode psychosis. JAMA Psychiatry 2013; 70: 1031–40.
- Pérez-Iglesias R, Tordesillas-Gutiérrez D, Barker GJ, McGuire PK, Roiz-Santiañez R, Mata I, et al. White matter defects in first episode psychosis patients: a voxelwise analysis of diffusion tensor imaging. Neuroimage 2010; 49: 199–204.
- Simmons A, Moore E, Williams SC. Quality control for functional magnetic resonance imaging using automated data analysis and Shewhart charting. Magn Reson Med 1999; 41: 1274–8.
- Skelly LR, Calhoun V, Meda SA, Kim J, Mathalon DH, Pearlson GD. Diffusion tensor imaging in schizophrenia: relationship to symptoms. Schizophr Res 2008; 98: 157–62.
- Smith SM. Fast robust automated brain extraction. Hum Brain Mapp 2002: 17: 143–55.
- Smith SM, Jenkinson M, Woolrich MW, Beckmann CF, Behrens TE, Johansen-Berg H, et al. Advances in functional and structural MR image analysis and implementation as FSL [Review]. Neuroimage 2004; 23 (Suppl 1): S208–19.
- Smith SM, Jenkinson M, Johansen-Berg H, Rueckert D, Nichols TE, Mackay CE, et al. Tract-based spatial statistics: voxelwise analysis of multi-subject diffusion data. Neuroimage 2006; 31: 1487–505.
- Song SK, Sun SW, Ramsbottom MJ, Chang C, Russell J, Cross AH. Dysmyelination revealed through MRI as increased radial (but unchanged axial) diffusion of water. Neuroimage 2002; 17: 1429–36.
- Song SK, Sun SW, Ju WK, Lin SJ, Cross AH, Neufeld AH. Diffusion tensor imaging detects and differentiates axon and myelin degeneration in mouse optic nerve after retinal ischemia. Neuroimage 2003; 20: 1714–22.
- Taylor D, Paton C, Kerwin RW. The Maudsley Prescribing Guidelines. 9th edn. London: Informa Health Care; 2007.
- Van Veelen NM, Vink M, Ramsey NF, van Buuren M, Hoogendam JM, Kahn RS. Prefrontal lobe dysfunction predicts treatment response in medication-naive first episode schizophrenia. Schizophr Res 2011; 129: 156–62
- Wang Q, Cheung C, Deng W, Li M, Huang C, Ma X, et al. White-matter microstructure in previously drug-naive patients with schizo-phrenia after 6 weeks of treatment. Psychol Med 2013; 43: 2301–9.
- Wechsler D. WAIS—III administration and scoring manual. 3rd edn. San Antonio, TX: The Psychological Corporation; 1997.
- White T, Magnotta VA, Bockholt HJ, Williams S, Wallace S, Ehrlich S, et al. Global white matter abnormalities in schizophrenia: a multisite diffusion tensor imaging study. Schizophr Bull 2011; 37: 222–32.
- Woods SW. Chlorpromazine equivalent doses for the newer atypical antipsychotics [Review]. J Clin Psychiatry 2003; 64: 663–7.
- Xiao L, Xu H, Zhang Y, Wei Z, He J, Jiang W, et al. Quetiapine facilitates oligodendrocyte development and prevents mice from myelin breakdown and behavioral changes. Mol Psychiatry 2008; 13: 697–708.
- Zhang Y, Zhang H, Wang L, Jiang W, Xu H, Xiao L, et al. Quetiapine enhances oligodendrocyte regeneration and myelin repair after cuprizone-induced demyelination. Schizophr Res 2012; 138: 8–17.