1 CLEAN JACI-D-13-00451R3

2 TITLE PAGE

3	Inhaled long-acting β_2 -agonists enhance glucocorticoid receptor nuclear translocation and
4	efficacy in sputum macrophages in COPD
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19	FUNDING SOURCE
20	This work was funded by an unrestricted grant from GlaxoSmithKline (UK). This project was
21	supported by the National Institute for Health Research (NIHR) Respiratory Disease Biomedical
22	Research Unit at the Royal Brompton and Harefield NHS Foundation Trust and Imperial College
23	London. Dr Omar Usmani is a recipient of a UK NIHR Career Development Fellowship.

24 ABSTRACT

25 **Background:** Combination inhaled therapy with long-acting β_2 -agonist (LABA) and corticosteroid 26 is beneficial in treating asthma and chronic obstructive pulmonary disease (COPD).

27 **Objective:** In asthma, LABAs enhance glucocorticoid receptor (GR) nuclear translocation in the 28 presence of corticosteroids. Whether this biological mechanism occurs in COPD, a relatively 29 corticosteroid-resistant disease, is uncertain.

Methods: Eight patients with mild/moderate COPD participated in a double-blind, placebocontrolled, crossover study and inhaled single-doses of; fluticasone propionate (FP)-100µg, FP-500µg, salmeterol(SLM)-50µg and combination FP-100µg+SLM-50µg. One hour post-inhalation sputum was induced, nuclear proteins isolated from purified macrophages and levels of activated nuclear GR quantified using a GR-GRE ELISA-based assay.

35 **Results:** Nuclear GR significantly increased after FP-500µg (p<0.01), but not after FP-100µg or 36 SLM-50µg, compared to placebo. Interestingly, SLM in combination with FP-100µg increased 37 nuclear GR levels equivalent to those of FP-500µg alone. This was significantly greater than 38 either FP-100µg (p<0.05) or SLM-50µg (p<0.01) alone. In vitro in a human macrophage cell line, SLM (10⁻⁸M) enhanced FP (10⁻⁹M)-induced mitogen-activated protein kinase phosphatase-1 39 40 (MKP-1) mRNA (5.8±0.6 vs. 8.4±1.1 x 10⁻⁶ copies, p<0.05) and 2xGRE-luciferase reporter gene 41 activity (250.1±15.6 vs. 103.1±23.6 fold induction, p<0.001). Addition of SLM (10⁻⁹M) to FP (10⁻¹ 42 ¹¹M) significantly enhanced FP-mediated suppression of IL-1 β -induced CXCL8 (p<0.05).

43 Conclusion: Addition of SLM-50µg to FP-100µg, enhanced GR nuclear translocation equivalent
44 to that seen with a five-fold higher dose of FP in sputum macrophages from COPD patients. This
45 may account for the superior clinical effects observed in COPD of combination
46 LABA/corticosteroid treatment compared to either as monotherapy.

CLINICAL INFORMATION

49	Sputum macrophages from COPD patients are relatively corticosteroid insensitive. Inhaled	
50	salmeterol and fluticasone propionate greatly enhanced glucocorticoid receptor activation in	
51	sputum macrophages from COPD patients.	
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54	CAPSULE SUMMARY	
55	Combination therapy with inhaled long-acting $\beta_2\text{-}agonist$ and corticosteroid improves clinical	
56	outcomes in COPD and biologically, enhanced glucocorticoid receptor activation in sputum	
57	macrophages may partly explain this phenomenon.	
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60	KEYWORDS: long-acting β_2 -agonist, corticosteroid, transcription factor, glucocorticoid receptor,	
61	sputum, COPD, macrophage	
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ABBREVIATIONS

65	COPD	Chronic obstructive pulmonary disease
66	FCS	Fetal calf serum
67	FP	Fluticasone propionate
68	FEV_1	Forced expiratory volume in one-second
69	GR	Glucocorticoid receptor
70	GRE	Glucocorticoid response element
71	HBSS	Hanks' buffered saline solution
72	ICS	Inhaled corticosteroids
73	LABAs	Long-acting β_2 -agonists
74	MKP-1	Mitogen-activated protein kinase phosphatase-1
75	PBMCs	Peripheral blood mononuclear cells
76	SLM	Salmeterol xinafoate
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88 INTRODUCTION

89 It is well-established in asthmatic patients that the addition of long-acting β_2 -agonist (LABA) to 90 inhaled corticosteroids (ICS) provides more effective disease control than monotherapy with ICS 91 [1-4]. Indeed, with recent concerns regarding the safety of LABA monotherapy in the treatment 92 of asthma, current guidance supports fixed dose combinations of LABA/ICS therapy from a 93 single inhaler [5-7]. Chronic obstructive pulmonary disease (COPD) is also characterized by 94 chronic airways inflammation, but unlike asthma, ICS treatment on its own has relatively little 95 effect on the accelerated decline in lung function seen in COPD patients [8,9]. However in 96 contrast, several large clinical trials of combination LABA/ICS therapy in patients with stable 97 COPD have shown better control of respiratory symptoms, lung function, quality of life, and 98 exacerbations with no greater risk of side-effects, compared to the use of either ICS alone or 99 LABA alone (10-13). Recent studies have demonstrated the effectiveness of LABA/ICS 100 combination therapy in reducing COPD morbidity and mortality compared with ICS treatment 101 alone (14, 15).

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103 The scientific rationale to explain the disproportionately superior clinical efficacy achieved by 104 the addition of LABAs to ICS in the treatment of asthma and COPD is being addressed, and 105 elucidating the underlying molecular mechanisms may help identify novel therapeutic targets 106 (16, 17). Combination therapy with SLM/FP reduces sputum differential cell counts, sputum 107 neutrophils and eosinophils, bronchial CD45+, CD8+ and CD4+ cells, and cells expressing genes 108 for tumor necrosis factor- α and interferon-y in comparison to placebo (18). These anti-109 inflammatory effects were also accompanied by improvements in the pre-bronchodilator forced 110 expiratory volume in one-second (FEV₁). Treatment with SLM/FP combination has also been shown to reduce tissue CD8+ and CD68+ cells compared with placebo, whereas no effect wasobserved with FP alone (19).

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114 It has been proposed LABAs may have anti-inflammatory properties similar to ICS (20, 21). 115 Studies show LABAs enhance corticosteroid-dependent anti-inflammatory effects, and the 116 biological mechanism for this may be through ligand-independent priming of the glucocorticoid 117 receptor (GR) (22). The cellular actions of corticosteroids are mediated by intracellular GRs, 118 which after binding to corticosteroid, translocate from the cell cytoplasm into the nucleus 119 where they mediate corticosteroid-dependent effects (23). Enhanced GR activation by LABAs 120 has been demonstrated in vitro in human lung fibroblasts and smooth muscle cells (22, 24), in 121 human neutrophils (25), and *ex vivo* in the sputum of patients with asthma (17). However, it is 122 unknown whether LABAs cause GR translocation after ICS therapy in COPD patients, as this 123 disease is much less responsive to the effects of corticosteroids compared to asthma (26). 124 Indeed, macrophages from COPD patients have been shown to exhibit a poor anti-inflammatory 125 response to corticosteroids in vitro compared to cells from healthy smokers and non-smokers 126 (21).

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In this study, we examined the effect of SLM/FP combination treatment on GR-glucocorticoid response element (GRE) binding in the nucleus of macrophages isolated from the induced sputum of patients with COPD. Importantly, we used inhaled drug doses used in the clinic. Our aim was to determine whether GR activation in response to corticosteroid therapy was enhanced by a LABA and whether this had any functional effects. As reduced nuclear translocation of GR contributes to corticosteroid insensitivity (16, 26), we hypothesized that the

134	LABA, salmeterol, could contribute to improve corticosteroid insensitivity in cells from COPD
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158 **METHODS**

159 Subjects

160 Eight patients diagnosed with moderate COPD (GOLD stages 2 and 3 (27)), <12% reversibility in 161 FEV₁ and a smoking history of >10 pack-years, participated in a randomized, double-blind, 162 placebo-controlled cross-over study. All patients were aged between 40 – 80 years and were not 163 taking either ICS or LABAs (Table 1). At each study-visit, patients' FEV₁ was required to be within 164 15% of their screening FEV₁ value to control for differences in airway function and inflammation 165 between visits. Patients were allowed to take the bronchodilators tiotropium bromide and/or 166 albuterol during the study, but none of the subjects were treated with ICS or LABAs for at least 167 one month preceding the start of the study.

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169 Study design

Single actuations of fluticasone propionate (FP)-100µg, FP-500µg, salmeterol xinafoate (SLM)50µg, combination therapy FP/SLM 100µg/50µg (Seretide[™], GlaxoSmithKline, Stevenage, U.K.)
or placebo were delivered via a metered dose inhaler and spacer, separately at each study visit
Sputum was induced 60 minutes after drug inhalation and peripheral blood samples were also
taken at this time point. The study was approved by the Ethics Committee of the Royal
Brompton & Harefield Hospitals National Health Service Trust, and all subjects gave written
informed consent.

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There was a minimum washout period between treatments of seven days to prevent any crossover effects. All patients were randomised (pre-assigned by picking balls out of a bag) to determine the order of treatment. In this scenario, there were 24 possible crossover orders, and with 8 subjects, there would have been possible treatment orders that were not received.

182	However, the sample size of 8 subjects was ethically chosen based on our previous study (17),
183	where 7 subjects received 5 treatments, which was sufficient to detect statistical differences
184	between ICS alone vs. LABA/ICS combination in asthmatic subjects.

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186 Sputum induction and processing

Sputum induction was performed as previously described and processed within one hour of collection (17). Briefly, sputum was filtered (70µm filter) and centrifuged (1600rpm for 5 min) to obtain a cell pellet. Cells were resuspended in full RPMI-1640 media (Sigma-Aldrich), supplemented with 1% L-glutamine, 10% fetal calf serum (FCS; Invitrogen Ltd, Carlsbad, CA), 100 U/ml penicillin and 100µg/ml streptomycin (Sigma-Aldrich). Total cell count (Kimura stain) and viability (Trypan blue exclusion) were determined before cytospins were undertaken.

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194 Ex-vivo stimulation of sputum macrophages

Sputum macrophages were isolated from whole sputum using plastic adherence at a cell density of 0.5x10⁶ cells/ml for 4 h in a 5% CO₂ humidified atmosphere at 37°C. Non-adherent cells were removed and macrophages resuspended in charcoal-stripped RPMI-1640 minimal media (1% Lglutamine and 0.5% FCS) overnight, before being exposed to SLM (10⁻⁷M), FP (10⁻⁹M), or a combination of both, for 60 min. The same number of macrophage cells in culture were seeded for each treatment visit and utilised for each of the experiments, standardised to a cell density of 0.5x10⁶ cells/ml.

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203 Isolation of blood cells

Venous blood (80 ml) was diluted 1:1 with Hanks' buffered saline solution (HBSS, Invitrogen) and
 layered on Ficoll-Hypaque-Plus (Amersham plc, Amersham, UK). After centrifugation (30 min at

1,100 x g and 18°C), peripheral blood mononuclear cells (PBMCs) were collected, washed, and
centrifuged (250 x g for 10 min). PBMCs were resuspended in culture media and counted using
Kimura dye.

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210 Nuclear extraction

Nuclear and cytoplasmic fractions were extracted using a Nuclear Extraction kit (Active Motif, Carlsbad, CA). Briefly, cells were resuspended in hypotonic buffer, vortexed and incubated on ice for 15 min to extract the cytoplasmic fraction. Thereafter, the remaining nuclear pellets were resuspended in complete lysis buffer for 30 min on ice. The suspension was centrifuged (14000rpm, 10 min, 4°C) and the nuclear fraction obtained. The quality and purity of the nuclear extract was determined by Western Blotting (Figure 1A).

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GRE binding assay

219 GR activation was determined with a GR-GRE TransAM kit (Active Motif, Frixensart, Belgium) 220 according to the manufacturer's instructions and nuclear GR-GRE binding was determined. 221 Briefly, 5µg of nuclear extract of each sample was added to a well. Each well contained multiple 222 copies of a specific double-stranded oligonucleotide which activated GR binds to at its 223 consensus binding site. We confirmed using Western blotting in U937 cells that incubation with 224 the positive control, fluticasone propionate, led to an increase in nuclear GR, which correlated 225 to an induction in GR-GRE binding, relative to the negative control of untreated cells (Figure 1B 226 & C).

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228 Confocal microscopy

229 Sputum macrophages were seeded onto glass coverslips in 6-well plates and allowed to attach 230 at 37°C. The cells were fixed with 4% paraformaldehye in phosphate buffered saline (PBS) for 10 231 min at 37°C and washed three times with PBS. Cells were then permeabilized with 0.2% Triton 232 X-100 (Sigma-Aldrich, Poole, UK) for 5min and then incubated with primary anti-GR antibody (H-233 300, Santa Cruz Biotechnology, CA, USA) for 3 hours followed by 30 min with secondary 234 antibody Alexa 488 (Invitrogen). Coverslips were washed three times with PBS, rinsed in distilled 235 water and immediately dried. Prolong Antifade 4', 6-diamidino-2-phenylindole dihydrochloride 236 (DAPI) solution (Invitrogen) was added to delineate the nuclear boundary in each cell and help 237 define the distribution of GR protein (green) within the respective subcellular compartments 238 (Figure 2). Coverslips were mounted onto slides and allowed to dry for 4 hours before analysis 239 using confocal microscopy with imaging software (Leica Confocal Software Lite, Heidelberg, 240 Germany).

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242 Cell Culture

243 U937 cells (ATCC, Rockville, MD) were maintained in RPMI-40 medium (Invitrogen), 244 supplemented with L-glutamine (1%, Invitrogen) and fetal calf serum (FCS, 10%, Invitrogen), and 245 were differentiated as previously described (28). After washing with HBSS, cells were allowed to 246 recover in medium supplemented with 1% L-glutamine and 0.1% charcoal-stripped FCS for 48 247 hours before experiments.

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249 MKP-1 mRNA expression

Differentiated U937 cells were cultured in RPMI supplemented with 0.1% charcoal-stripped-FCS and L-glutamine (1%) for 24 h before treatment. Total RNA was isolated from cells using an RNeasy Kit (Qiagen, Hilden, Germany). cDNA was synthesised from 1µg of total RNA with the 253 Qiagen Quantitect Kit (Qiagen) according to the manufacturer's protocol. Expression of 254 mitogen-activated protein kinase phosphatase-1 (MKP-1) mRNA was quantified by RT-qPCR with 255 SYBR Green PCR Master Mix (Applied Biosystems, Foster City, CA, US). Primers are shown in 256 Table 2. MKP-1 mRNA levels were normalized to an endogenous reference GAPDH and absolute 257 quantification was determined by using known standards of MKP-1. MKP-1 standards were 258 produced from PCR products containing the target sequence from U937 monocytes. The copy 259 number of the standards was determined by measuring the concentration using a 260 spectrophotometer.

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262 **GR reporter gene assay**

263 Differentiated U937 cells were transfected with GRE-luciferase and β -galactosidase plasmids as 264 previously described and GR transactivation determined by luciferase assay as previously 265 described (17).

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267 CXCL8 enzyme-linked immunosorbent assay (ELISA)

Serum-starved U937 cells were incubated with IL-1β for 30 min, followed by FP and SLM alone and in combination for 16 hours. Subsequently, extracellular concentrations of CXCL8 were measured with human CXCL8 Duoset ELISA kits (R&D Systems Europe, Abingdon, UK) according to the manufacturer's instructions.

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273 Statistical analysis

274 In the *in vivo* study, as sputum cells are not normally distributed, a non-Gaussian distribution 275 was adopted and data were analysed using non-parametric statistical analyses using the PC analysis package Graph Pad Prism (Graph Pad Prism, San Diego, CA). Results were analyzed using Friedman analysis of variance (ANOVA) including factors for period, treatment, and multiple comparisons, and applying Bonferroni's post-test correction. Comparisons between treatments were made using the Wilcoxon matched-pairs signed rank sum test. A p value of less than 0.05 was considered statistically significant. For the difference between treatment groups in the in vitro data, analyses were undertaken using the Mann-Whitney test for comparison of two unpaired groups with Graph Pad Prism software. A p value of less than 0.05 was considered statistically significant.

295 **RESULTS**

296 **Patient characteristics**

297 The mean age of the eight patients (6 female) was 65 ± 8 years and the mean post-298 bronchodilator FEV₁ was 72 \pm 12% of predicted (Table 1). No patient showed >12% reversibility 299 to albuterol, was atopic, or had a previous history of asthma. Median % sputum cell counts are 300 shown in Table 1 averaged from different study visits across all patients, and most patients had 301 high levels of neutrophilia. The percentage of sputum macrophages obtained varied between 302 visits within each patient (range -6.7% to +12.2%) compared to the sample taken at visit 1. 303 However, the amount of protein extracted from the isolated macrophage nuclear extracts for 304 the GR-GRE assay was normalised to be the same for all samples from all patients for all visits, 305 irrespective of the variability in the sputum macrophages yielded following sputum induction, 306 and this was 5µg.

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308 Effect of LABA and corticosteroid on GR-GRE binding in sputum cells *in vivo*

309 GR-GRE binding levels in the nuclei of macrophages obtained from induced sputum were not 310 significantly altered at 60 minutes following inhalation of FP-100µg or SLM-50µg compared to 311 placebo (Figure 3A). In contrast, FP-500µg significantly increased GR-GRE binding levels 2.2-fold 312 compared to placebo (0.607 ± 0.089 vs. 0.274 ± 0.038 absorbance units, p<0.01) and induction 313 of GR-GRE binding was significantly greater with FP-500µg compared to that observed with FP-314 100µg (0.607 ± 0.089 vs. 0.310 ± 0.028 absorbance units, p<0.05). Combination treatment of FP-315 100µg with SLM-50µg increased nuclear GR-GRE levels to similar levels to those seen with FP-316 500µg (respectively 0.598 ± 0.061 vs. 0.607 ± 0.089 , p = NS). This effect observed with the 317 combination treatment was significantly greater than that seen with either placebo (p<0.01), FP-318 100µg alone (p<0.05) or SLM-50µg alone (p<0.01). The addition of SLM-50µg to FP-100µg as

combination treatment, achieved an enhancement of GR activation in sputum macrophages from patients with COPD that was equivalent to that observed with a five-fold increase in the dose of FP. That is, combination therapy allowed a five-fold reduction in the inhaled corticosteroid dose, yet achieved the same biological effects. Confocal microscopy documented the correlation between the glucocorticoid receptor GRE binding assay and glucocorticoid receptor nuclear translocation in that, treatment with corticosteroid (FP-500µg) led to GR nuclear translocation in sputum macrophage cells (Figure 2).

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327 Effect of LABA and corticosteroid on GR-GRE binding in PBMCs in vivo

The levels of GR-GRE binding in PBMCs were measured 60 minutes after inhalation of the drugs (Figure 3B). None of the active treatments had an effect on GR-GRE binding levels and these levels did not differ significantly from those obtained with placebo.

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332 Effect of LABA and corticosteroid on GR-GRE binding in sputum cells ex vivo

Sputum macrophages were isolated by adherence to plastic, allowed to recover for 24 h before stimulation with SLM (10^{-7} M), FP (10^{-9} M) or a combination of both for 60 minutes. FP significantly enhanced nuclear GR-GRE binding compared to control unstimulated cells ($0.506 \pm$ 0.042 vs. 0.252 ± 0.038 absorbance units, p<0.01). Salmeterol alone had no significant effect on GR-GRE binding levels compared to control (Figure 4). However, the addition of SLM to FP significantly enhanced the ability of FP alone (10^{-9} M) to stimulate GR-GRE binding (0.710 ± 0.073 vs. 0.506 ± 0.042 absorbance units, p<0.05).

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341 Induction of MKP-1 mRNA in macrophage-like U937 cells

FP (10^{-9} M) alone significantly induced MKP-1 mRNA expression after 2 h compared to the levels seen in control unstimulated cells ($5.8 \pm 0.6 \times 10^{-6}$ vs. $6.4\pm2.5 \times 10^{-8}$ copies, p<0.001), whilst SLM (10^{-8} M) alone had no effect ($3.2 \pm 0.6 \times 10^{-8}$ copies, p=NS) (Figure 5A). However, upon addition to FP, SLM significantly enhanced FP-induced MKP-1 mRNA (5.8 ± 0.6 vs. $8.4 \pm 1.1 \times 10^{-6}$ copies, p<0.05).

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Induction of GRE-luciferase activity by LABA and corticosteroid in macrophage-like U937 cells Salmeterol (10^{-8} M) alone had no significant effect on a 2xGRE-luciferase reporter gene activity compared to control unstimulated cells (1.0 ± 0.6 vs. 1.5 ± 0.4 fold induction, p=NS) (Figure 5B). In contrast, FP (10^{-9} M) significantly enhanced GRE-luciferase activity compared to control (103.1 ± 23.6 fold induction, p<0.05). Yet, the addition of SLM to FP resulted in a highly significant increase in GRE-luciferase activity compared to that seen with FP alone (250.1 ± 15.6 vs. 103.1 ± 23.6 fold induction, p<0.001).

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356 Effect of LABA and corticosteroid on IL-1β-induced CXCL8 in macrophage-like U937 cells

FP (10⁻¹¹M) and (10⁻⁹M) significantly suppressed IL-1β-induced CXCL8 (28.3 ± 3.2 and 71.2 ± 3.1 % respectively, p<0.01), whilst SLM (10⁻⁹M) alone had no considerable effect (Figure 6). However, the addition of SLM (10⁻⁹M) to FP (10⁻¹¹M) significantly enhanced FP-mediated suppression of IL-1β-induced CXCL8 (46.7 ± 4.8 vs. 28.3 ± 3.2 %, p<0.05).

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366 **DISCUSSION**

367 Our study shows that nuclear levels of activated GR in macrophages obtained from the induced 368 sputum of patients with COPD increased following single-dose inhalation treatment with high-369 dose fluticasone propionate (FP)-500µg, but not after low-dose FP-100µg alone or salmeterol 370 (SLM)-50µg alone. Most importantly, the addition of SLM-50µg to low-dose FP-100µg increased 371 activated GR levels equivalent to those seen with high-dose FP-500µg. That is, in COPD patients, 372 addition of salmeterol to a low-dose of FP achieved an enhancement of GR nuclear 373 translocation equivalent to that observed with a five-fold higher corticosteroid dose. Consistent 374 with our *in vivo* patient data, we were able to demonstrate that salmeterol significantly 375 enhanced the ability of FP to induce GR activation in sputum macrophages ex vivo and enhanced 376 FP-induction of MKP-1 mRNA and 2xGRE reporter gene activity in a human cultured 377 macrophage cell line *in vitro*. Additionally, we were able to show a functional anti-inflammatory 378 effect of combination therapy in that salmeterol significantly enhanced FP-mediated 379 suppression of IL-1 β -induced CXCL8 in macrophage-like U937 cells.

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381 When LABAs were introduced as an add-on therapy to inhaled corticosteroids, the magnitude of 382 their clinical beneficial effect in the control of asthma, and in COPD, was not predicted. The role 383 of LABAs was seen purely as bronchodilatation without any appreciable effect on bronchial 384 inflammation. In asthma, LABAs combined with corticosteroids produced a better effect on 385 symptoms (1, 3), lung function (29), exacerbation rates (30) and health status (31) than 386 expected. Adding LABA to low-dose ICS was observed to be more effective than increasing the 387 dose of corticosteroids (1, 29, 30, 32) and asthma guidelines were updated to reflect these 388 findings (33).

390 The beneficial effects of the LABA/ICS combination have also been observed in COPD patients, 391 where they are now an established treatment. The TORCH (TOwards a Revolution in COPD 392 Health) study demonstrated reductions in exacerbation rates, improvements in health status 393 and lung function with SLM/FP combination compared to placebo in COPD patients (15). The 394 study observed a relative reduction in mortality of 17% over three years for patients receiving 395 SLM/FP, although this just failed to reach statistical significance. A subsequent Cochrane review 396 observed survival benefits with combination therapy compared both to placebo (34) and to ICS 397 alone (14). In vivo studies using bronchial biopsies have generated further supporting evidence 398 for complementary interactions between LABAs and corticosteroids on airway inflammation and 399 pathology in COPD patients (18, 19).

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401 Our data reveal a complementary interaction between the LABA (salmeterol) and corticosteroid 402 (fluticasone propionate) in that the combination therapy has a greater anti-inflammatory effect 403 than either drug alone and was also steroid sparing; allowing a five-fold reduction in the inhaled 404 corticosteroid dose, yet achieving the same biological effects. Indeed our data support the 405 postulate that changes in the anti-inflammatory effects with combination therapy may account 406 for the improvements in the rate of decline in lung function seen in the TORCH study (35).

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We confirm that sputum macrophages from COPD patients are less sensitive to corticosteroid action compared to those obtained from asthmatic subjects (36, 37). In our previous study examining the effect of corticosteroid (FP) on GR activation in sputum macrophages from mild/moderate asthmatic subjects we demonstrated that 60 minutes post-treatment, FP-100µg significantly enhanced GR nuclear translocation (17). Here we show that only the higher dose of FP-500µg had an effect on GR-GRE binding. These data suggest that GR-GRE binding in sputum

414 macrophages may be a good biomarker for corticosteroid actions. As in our previous study (17), 415 we were able to detect a quantifiable degree of basal GR-GRE binding in sputum macrophages 416 in the placebo-treated patients (Figure 3) and this level of GR activation was maintained for at 417 least 24 hours in culture. Since none of the patients in this study were exposed to exogenous 418 inhaled or oral corticosteroids this activation must reflect the action of endogenous 419 corticosteroids.

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421 Consistent with our in vivo patient data, our in vitro experiments, showed salmeterol enhanced 422 the ability of low concentrations of FP to induce GRE-luciferase activity and switch on MKP-1 423 expression in macrophage-like U937 cells. Similar data for enhanced induction of glucocorticoid-424 responsive genes in the presence of a LABA such as MKP-1 and glucocorticoid inducible leucine 425 zipper (GILZ) has been reported in airway epithelial cells and smooth muscle cells (38). Our 426 results are consistent with literature that suggest the enhanced effects may result from the 427 ability of LABAs to promote GR nuclear translocation in human lung fibroblasts (22), airway 428 smooth muscle cells (24, 39, 40) and in sputum cells from asthmatic patients (17). This 429 enhancement of GR translocation achieved by LABAs when added to corticosteroids is likely to 430 underlie some of the clinical benefits seen with combination therapy. Additionally, we 431 demonstrated that the enhancement of GR translocation with combination therapy was 432 associated with a functional anti-inflammatory effect; in that, salmeterol significantly enhanced 433 FP-mediated suppression of IL-1 β -induced CXCL8 in U937 cells.

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We were unable to assess the anti-inflammatory properties of the combination treatment in sputum macrophages *in vivo* due to limitations in cells numbers and ethical considerations regarding repeated sputum induction in patients. Future experiments examining gene

438 expression profiles in sputum macrophages 4 hours after inhaled drugs will provide evidence for 439 additional genes that are regulated in a similar manner to those described here. A role for other 440 aspects of GR activation including GR post-translational modifications and the activity of GR 441 chaperone proteins is difficult to rule out as potential modulators of GR-GRE binding. An 442 experimental clinical patient study specifically designed to address these points would be 443 needed. In contrast to our observations in sputum macrophages, the levels of activated GR in 444 the nuclei of PBMCs did not change significantly after any treatment. This is likely to be due to 445 peripheral blood levels of the study medications being too low after a single-dose inhalation to 446 exert a measurable effect on the biological machinery of GR translocation. It would be 447 interesting to see if a measurable effect is achieved after a more prolonged course of treatment.

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The marked effect of LABAs on GR translocation *in vitro* and *in vivo*, in asthmatics and in COPD patients warrants further investigation, and elucidation of the precise molecular mechanisms of how LABAs (which acting extracellularly via the β_2 receptor) are able to influence translocation of cytoplasmic GRs. Altered phosphorylation, 'priming' of the GR increasing its tendency to translocate without the need for its natural ligand has been proposed (41, 42). Recently we have shown that LABA increase GR nuclear translocation through the activation of a phosphatase PPA2, which dephosphorylates hyperphosphorylated GR (43).

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In conclusion, we report here for the first-time that a LABA in combination with a low-dose of ICS enhances GR-GRE binding *in vivo* in COPD patients as effectively as a five-fold higher dose of ICS alone. This suggests LABAs may play an important role in the benefits clinically seen with combination therapy. Our data has clinical relevance to the use of combination inhalers in view of the reduced sensitivity to ICS in COPD. Further studies are needed to address whether these

- 462 differences are maintained following chronic treatment of LABAs with ICS in COPD patients, and463 whether this results in an alteration in sputum cytokine profiles *in vivo*.
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584	

TABLES

- **TABLE 1.**
- **Patient characteristics**

Age (years)	65 ± 8
Gender	2M/6F
GOLD stage	2 stage 2, 6 stage 3
FEV ₁ (L)	1.8 ± 0.89
Post-bronchodilator FEV ₁ (% predicted)	72.1 ± 11.8
Reversibility (%)	5.6 ± 4.2
FEV ₁ /FVC ratio	60.2±5.2
Smoking status	2 current, 6 ex
Smoking history (pack-years)	35.3 ± 16.2

Sputum Cell Counts

Macrophages (% total) median % (range)	7 (4.8 – 33.4)
Neutrophils (% total) median % (range)	91.9 (39.7 – 93.1)
Lymphocytes (% total) median % (range)	1 (0 – 1.7)
Eosinophils (% total) median % (range)	0.6 (0 – 25.7)

TABLE 2.

PCR primers for MKP-1 mRNA analysis

	Forward	Reverse
MKP-1	5'-GTACATCAAGTCCATCTGAC	5' GGTTCTTCTAGGAGTAGACA
GAPDH	5'- GAAGGTGAAGGTCGGAGTC	5'- GAAGATGGTGATGGGATTTC

598 **FIGURE LEGENDS**

599 Figure 1. Validation of the quality of nuclear extract and GR-GRE binding.

The quality of nuclear extracts (panel A) was determined by Western Blotting. Fluticasone
propionate (FP)-induced GR-GRE binding (panel B) and FP-induced GR nuclear localisation (panel
C) were assessed in U937 monocytes. Results represent the mean ± SEM of 3 independent
experiments. *p<0.05 denotes statistical significance *vs.* untreated cells.

604

Figure 2. Confocal laser images of GR subcellular localisation in sputum macrophages
following corticosteroid (FP 500μg) treatment. (panels A, D) 4', 6-diamidino-2-phenylindole
dihydrochloride (DAPI) staining (blue channel); (panels B, E) cytoplasmic and nuclear GR
immunostaining (green channel); (panels C, F) overlay of panels A and B.

609

Figure 3. Effect of inhaled salmeterol (SLM) and fluticasone propionate (FP) on GR-GRE binding in nuclear extracts from induced sputum macrophages (panel A) and PBMCs (panel B). Mean ± SEM absorbance values of GR-GRE binding are shown 60 minutes post-inhalation of treatment, where numeric values are inhaled drug doses in micrograms. n = 8 subjects.

614 *p<0.05 and **p<0.01 denote statistical significance between treatments.

615

Figure 4. Fffect of fluticasone propionate (FP, 10^{-9} M) and salmeterol (SLM, 10^{-7} M) on GR-GRE DNA binding in sputum macrophages from COPD patients *ex vivo*. Mean ± SEM absorbance values of GR-GRE binding are shown. n = 5subjects. *p<0.05 denotes statistical significance between treatments.

Figure 5. Effect of fluticasone (FP, 10^{-9} M) and salmeterol (SLM, 10^{-8} M) on MKP-1 mRNA induction and GRE-luciferase activity in macrophage-like PMA-treated U937 cells in vitro. (A) MKP-1 mRNA expression normalized with GAPDH (panel A) and the fold-increase in GR of stimulated vs. non stimulated cells (panel B) are shown. Results represent the mean ± SEM of three independent experiments. * p<0.05 and *** p<0.001 denote statistical significance between treatments.

627

Figure 6. Effect of fluticasone propionate (FP) and salmeterol (SLM) on IL-1β-induced CXCL8 in macrophage-like U937 cells. Results represent the mean ± SEM of 8 independent experiments. *p<0.05 denotes statistical significance vs. FP (10^{-11} M) and #p<0.01 denotes statistical significance vs. IL-1β stimulated.