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Belowground connections underlying aboveground food production: a framework for optimising ecological connections in the rhizosphere

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- 2 framework for optimising ecological connections in the rhizosphere
- 3
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- 13
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- 15
- 16

17 Summary

- 18 1 Healthy soils that contain an active microbiome and food web are critical to
- 19 sustainably produce food for a growing global human population. Many studies have
- 20 focussed on the role of microbial species diversity and the presence of key functional
- 21 groups as important controls on the many functions that a sustainable food system
- 22 relies on.
- 23 3 Here, we synthesize recent ecological empirical evidence and theory to propose that
- the interactions between organisms in the soil food web are the critical determinant ofsoil function.
- 26 4 We propose the Rhizosphere Interactions for Sustainable Agriculture (RISA) Model, in
- 27 which crop roots recruit small, modular, highly connected soil rhizosphere networks
- 28 from large, static, relatively unconnected and diverse bulk soil networks. We argue that
- 29 conventional agricultural management disrupts the connections between rhizospere
- 30 and bulk soil networks.
- 31 *Synthesis* We identify future research directions for optimising ecological connections
- 32 between roots and rhizosphere microbial and faunal networks, and between
- 33 rhizosphere networks and bulk soil networks in agricultural production systems.
- 34 Knowledge on these connections can be applied in agricultural systems to sustainability
- 35 produce food for a growing global population.

36

- 37 Keywords: carbon, nitrogen, bacteria, fungi, microbial community, soil fauna, stability,
- 38 network, bulk soil, roots

39

40 Introduction

41 Healthy soils are critical to support food production as the human population grows 42 towards 9.7 billion and our planet warms (Paustian et al. 2016). But, what are the most 43 important characteristics of healthy soils that support sustainable agriculture? Most 44 definitions of soil health include not just physiochemical attributes like high organic 45 matter content, but also a rich biological component of microbes and soil fauna (Doran 46 & Zeiss 2000). Soils with active microbes and fauna in close interactions with plants 47 support efficient nutrient cycling, pathogen resistance and overall crop health (Bender, 48 Wagg & van der Heijden 2016). However, we lack a rigorous framework for directing 49 future research efforts towards quanitfying and optimising interactions between plants 50 and soil organisms to support healthy, productive crops while reducing our reliance on 51 agrochemicals. Here, we draw on recent ecological evidence and theory to develop a 52 conceptual framework that can support future research efforts to advance our 53 understanding of these interactions. Ultimately, an increased understanding of 54 interactions between plants and soil communities can accelerate new technological and 55 management innovations that harness belowground interactions to enhance 56 aboveground productivity. 57 When plants first started extending their roots into primordial soils, they encountered a 58 rich microbiome. Ever since, plants have co-evolved with soil microbes and fauna, and 59 traits that supported beneficial interactions with soil organisms likely incurred a fitness 60 advantage (Lambers et al. 2009). For example, plants that enable mycorrhizal 61 colonisation are able to enhance their nutrient acquisition. Plant root exudates also 62 support microbes that release enzymes to depolymerize organic matter and transform it 63 into plant-available forms through mineralisation. This long history of co-evolution 64 provides a roadmap for measuring and managing beneficial plant-microbe interactions 65 that are key components of healthy soils. Here, we suggest that rather than overall 66 microbial abundance or species diversity, the structure of the network of interactions 67 between plants and soil organisms and among soil organisms best predicts the 68 functionality of soil communities and provides a framework for assessing and managing 69 soil health. 70 Plants affect the structure and function of the soil microbiome and food webs directly

- through beneficial associations with mycorrhizal fungi and *Rhizobia*, and through
- 72 antagonistic associations with, for example, plant-parasitic nematodes. Indirect controls
- on soil communities are equally important and act primarily through the quantity and

74 quality of plant aboveground and belowground litter inputs (Orwin *et al.* 2010; De Vries 75 et al. 2012b; Baxendale et al. 2014). In the rhizosphere, plant N uptake is a dominant 76 control on bacterial and fungal community composition (Bell et al. 2015; Moreau et al. 77 2015; Giagnoni et al. 2016; Thion et al. 2016). In addition, rhizodeposits and root litter 78 have been shown to strongly influence the composition of soil microbial and faunal 79 communities and soil food webs (Scheunemann et al. 2015; Sauvadet et al. 2016), and 80 feed back to the performance of neighbouring plant species by altering soil N availability 81 (Meier, Keyserling & Bowman 2009). Root exudates have been hypothesised as an 82 important mechanism driving microbial community and soil food web composition and 83 activity (Bardgett, Mommer & De Vries 2014; De Vries & Caruso 2016), though this has 84 only been experimentally demonstrated for Arabadopsis (Shi et al. 2011; Badri et al. 85 2013). Finally, antimicrobial compounds isolated from crop plant root exudates can 86 affect the growth and activity of pathogenic soil bacteria (Liu et al. 2015; Fang et al. 87 2016). These mechanisms form a tight link between plants, microbes, and the soil food 88 web in the rhizosphere.

89 As a result of these interactions, the rhizosphere and bulk soil host contrasting microbial 90 and faunal communities. The rhizosphere is typically enriched in bacterial taxa that can 91 be classified as copiotrophic, such as Alphaproteobacteria, Betaproteobacteria, and 92 Bacteroidetes (Shi et al. 2015; Uksa et al. 2015), as well as in specialist decomposers and 93 plant-growth promoting bacteria, compared to the bulk soil (Hargreaves, Williams & 94 Hofmockel 2015; Chen et al. 2016). Soil animal communities generally have higher 95 biomass and abundance in the rhizosphere, and similar to bacterial communities, the 96 rhizosphere is enriched in species that are able to rapidly use root-derived C (Fujii, 97 Saitoh & Takeda 2014; Scharroba et al. 2016). Turner et al. (2013) found that the 98 rhizosphere communities of the major crop plants wheat, pea, and oat were clearly 99 differentiated despite some overlap. In particular, grain rhizospheres were enriched in 100 cellulolytic bacteria, while peas recruited more plant growth promoting bacteria. The 101 wheat rhizosphere was strongly enriched in the bacterial-feeding nematode 102 Acrobeloides, while the pea rhizosphere was highly enriched in fungi. Recent evidence 103 also shows that bacterial alpha-diversity tends to be lower in the rhizosphere 104 (Hargreaves, Williams & Hofmockel 2015; Chen et al. 2016). And while alpha diversity is 105 often proposed as an indicator of robust, healthy soil communities, there is little 106 experimental evidence to support this notion.

107 Rather than species numbers, interactions between organisms appear to be crucial for

108 the functioning of soil communities. Networks of interactions can predict whole

109 ecosystem functioning (Kuiper *et al.* 2015) and drive multiple processes and ecosystem 110 services that underlie this functioning (Hines et al. 2015). Soil food webs with a more 111 prominent fungal energy channel (Box 1) are more resistant to, and continue to function 112 better under drought (De Vries *et al.* 2012a). In addition, the ratio between the bacterial 113 and fungal energy channel has been correlated to the rates of soil C and N cycling 114 processes (De Vries et al. 2013). The success of pathogen invasion in the rhizosphere 115 has been shown to depend on the network structure of resident bacterial communities 116 (Wei *et al.* 2015). Specifically, pathogen success was lower when resident networks had 117 low nestedness (or modularity, see Box 1) and high connectance, presumably through 118 more efficient consumption of resources. Other recent studies found that associations 119 between methane oxidising bacteria and other microbes were central in modulating 120 methane-oxidation (Ho et al. 2016), and that fungal co-occurrence network structure 121 was strongly associated with different stages of litter decomposition (Purahong et al. 122 2016). Thus, new ecological knowledge of the structure of interactive networks among 123 organisms and their environment might be used as an indicator of their functional 124 attributes.

125

126 Rhizosphere networks are recruited from bulk soil networks

127 In the rhizosphere, bacterial networks typically have more nodes, more hubs, and 128 stronger connections (see Box 1 for definitions) than those in bulk soil (Mendes et al. 129 2014; Shi et al. 2016). In a recent study, Shi et al. (2016) found that while bulk soil 130 bacterial communities contained more OTUs, rhizosphere networks of bacterial taxa 131 were larger and more connected. Moreover, rhizosphere networks grew larger and 132 gained more connections and hubs during plant growth, while networks in the bulk soil 133 remained relatively static. Similarly, rhizosphere decomposer soil food webs tend to be 134 dominated by the bacterial energy channel (Thakur & Eisenhauer 2015; Muller et al. 135 2016), which typically have lower diversity than fungal-dominated soil food webs (De 136 Vries et al. 2012a, 2013). These patterns likely occur because the high-resource 137 rhizosphere 'recruits' a subset of species from the bulk soil community. The selected 138 species then share niche space in the rhizosphere and as a result show strong positive 139 associations in networks (Mendes et al. 2014; Edwards et al. 2015). Importantly, the 140 presence of plant roots has been shown to be as important as land use and soil type for 141 shaping bacterial community composition (Edwards et al. 2015; Hargreaves, Williams & 142 Hofmockel 2015).

143 As a result of these different network structures, rhizosphere networks are likely to be 144 less stable under changing environmental conditions than bulk soil networks. 145 Rhizosphere microbial networks are characterised by overwhelmingly strong, positive 146 interactions, which have been shown to compromise network stability (Box 1). In 147 addition, bacterial-dominated rhizosphere soil food webs consist of strong interactions, 148 and a reduction of the fungal energy channel effectively reduces network 149 compartmentalisation, thereby reducing network stability (Box 1). These properties 150 allow rhizosphere networks to respond quickly to changes in resource availability, for 151 example as a result of plant growth stages or agricultural management. 152 In contrast to the rhizosphere, the bulk soil harbours a large, diverse network of 153 organisms with low abundances that consists of weak interactions. The majority of 154 these organisms are dormant or inactive and are thus highly resistant to environmental 155 stresses (Lennon & Jones 2011) and can persist in soil for long periods of time (Troxler 156 et al. 2012). This network of bulk soil organisms is mostly unaffected by the dynamic 157 processes that affect rhizosphere networks and remains relatively static during the 158 growing season (Shi et al. 2016), but would be highly stable under changing 159 environmental conditions because of its properties. We propose that this bulk soil 160 network provides the 'seed bank' from which rhizosphere networks are recruited, and 161 we argue that crops will be able to recruit a functioning rhizosphere network as long as 162 this seed bank is intact (Fig. 1). The composition and structure of the recruited 163 rhizosphere network depends on the traits, and in particular root traits, of the crop 164 grown, as well as on the abundance and composition of the bulk soil community. 165 However, we argue that conventional agricultural management reduces the ability of the 166 rhizosphere to recruit from the bulk soil.

167

168 Agricultural management affects network structure of, and connections between,

169 rhizosphere and bulk soil networks

- 170 Agricultural management directly affects both microbial network and soil food web
- 171 structure in the rhizosphere. It is well known that agricultural practices such as
- 172 fertilisation and tillage reduce the diversity of soil organisms (Tsiafouli *et al.* 2015) and
- 173 shift soil food webs towards being bacterial-dominated rather than fungal-dominated
- 174 (Beare *et al.* 1997; De Vries *et al.* 2006). Recent evidence also indicates that microbial
- 175 networks are affected by agricultural management. For example, in a field sampling
- 176 across China , chemical fertiliser application reduced the size, modularity, and number

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177 of connections of bacterial networks compared to organic amendments (Ling et al. 178 2016). Similarly, in a field experiment, organic manure amendment increased the 179 modularity of arbuscular mycorrhizal fungal networks (Zhu et al. 2016). But 180 agricultural management also disproportionately reduces the biomass of groups of 181 organisms that form a connection between rhizosphere and bulk soil, and thus facilitate 182 rhizosphere recruitment. For example, fungal hyphae can form a connection between 183 rhizosphere and bulk soil microbial communities and food webs by transporting C and 184 facilitating bacterial movement (Fransson & Rosling 2014; Gahan & Schmalenberger 185 2015; Pausch et al. 2016). Similarly, soil fauna can form a connection between 186 rhizosphere and bulk soil networks. Earthworms incorporate and distribute organic 187 residues into different soil layers and facilitate microbial dispersal (Tao *et al.* 2009). 188 Both fungal and bacterial grazers can carry bacterial cells and fungal spores in their gut 189 and on their surface (Ingham 1999; Renker et al. 2005), and facilitate their movement 190 by migrating between soil layers (Lindberg & Bengtsson 2005). In addition, 191 conventional agricultural management can also reduce the movement of soil organisms 192 between rhizosphere and bulk soil by compacting and homogenising the soil (Tao et al. 193 2009; Warmink et al. 2011; Ebrahimi & Or 2014). Moreover, the composition and 194 persistence of the bulk soil 'seed bank' can be affected by agricultural amendments 195 seeping to deeper soil layers, such as fertiliser, pesticides, manure, and the antibiotics 196 within manure.

197

198 The Rhizosphere Interactions for Sustainable Agriculture (RISA) Model

199 Drawing on recent advances in understanding of the structure and functioning of 200 ecological networks, we introduce a new conceptual framework to guide future research 201 efforts as well as technological and management innovations that optimise the abilities 202 of soils to support sustainable crop production. Specifically, this framework aims to 203 optimise connections between roots and rhizosphere microbial and faunal networks, 204 and connections between rhizosphere networks and bulk soil networks. Based on the 205 emerging evidence reviewed above, we propose that agricultural bulk soils should 206 contain diverse organismal communities characterized by weak interactions, whereas 207 rhizosphere communities should be less diverse and characterized by a high degree of 208 nodes and strongly positive connections, both within organismal networks and between 209 these network and rhizosphere properties. Most importantly, connections between the 210 rhizosphere and bulk soils should be supported.

211 Most evidence that organismal network structure underlies soil functioning originates 212 from studies on soil food webs, in which feeding interactions between organisms have 213 empirically been quantified through decades of research (Bradford 2016). While the 214 nature of soil food web interactions are currently under debate (Ballhausen & de Boer 215 2016; Geisen 2016), existing food web models have successfully predicted C and N 216 fluxes in natural and agricultural systems (De Ruiter et al. 1993; Holtkamp et al. 2011). 217 In soil microbial networks, correlations between microbial taxa can result from a variety 218 of interaction types (Box 1). To model process rates from data on interactions within 219 microbial networks, and to predict functioning based on microbial network structure, 220 we need to first elucidate the exact nature of dynamic microbe-microbe interactions 221 (Gottstein *et al.* 2016). Therefore, an important challenge is to identify interaction types 222 between microbial groups or species and how these determine network structure. Here, 223 we can learn from gut microbiologists, who have successfully linked gene transcripts to 224 species-species interactions (Plichta et al. 2016). Several covarying transcripts were 225 down-regulated in interacting species pairs, indicating that the functional overlap 226 between species was reduced and that species activities were context specific. A 227 creative alternative to experimentally elucidating the nature of the millions possible 228 specific microbe-microbe interactions might be text-mining of existing literature (Lim et 229 al. 2016).

230 Once we know the functionalities of key nodes and network modules, we can identify 231 nodes and hubs associated with desirable functions such as nutrient mineralisation or 232 plant protection from disease. Taxa that often appear as nodes in structured 233 rhizosphere networks may be promising targets for inoculants, since the addition of just 234 a few species may lead to a structured network. There is a long history of inoculating 235 beneficial microbes either directly into soils or as seed coatings (Calvo, Nelson & 236 Kloepper 2014). Mycorrhizae are often inoculated on tree seedlings and crops to 237 improve establishment and nutrient uptake (du Jardin 2015). Rhizobia are inoculated on 238 legumes to ensure nodule formation (Catroux, Hartmann & Revellin 2001; du Jardin 239 2015). However, along with many other types of plant-growth promoting bacteria, 240 inoculants have had only limited success to date in field agriculture (Calvo, Nelson & 241 Kloepper 2014). There is little evidence that most inoculants are able to persist and 242 compete with other microbes in most soils (Verbruggen et al. 2013; Berruti, Lumini & 243 Bianciotto 2016), although one study found that the plant-growth promoting bacterium 244 Pseudomonas fluorescens persisted in the bulk soil for months after inoculation (Troxler 245 et al. 2012).

246 In addition to inoculation, mechanistic knowledge on links between plant traits and soil 247 organismal networks could enable another promising approach for promoting soil 248 organismal networks that deliver desirable functions (Kumar et al. 2016). While there is 249 accumulating evidence that plant traits, and in particular root traits, can predict soil 250 faunal and microbial community structure and functioning (De Vries *et al.* 2012b; 251 Grigulis et al. 2013; De Vries & Bardgett 2016; Legay et al. 2016), there is only limited 252 evidence that links plant traits to soil microbial networks. For example, it has been 253 shown that specific compounds in root exudates can promote the formation of bacterial 254 clusters (Thomas & Cebron 2016). In addition, in a mesocosm study with eucalyptus 255 seedlings, networks of ammonia-oxidising archaea were more connected and had more 256 modules in soil under elevated CO_2 than those under control conditions, presumably 257 through changes in belowground plant C inputs (Hu et al. 2016). As future research 258 further elucidates possible linkages between plant traits and microbial functions, plant 259 breeding and engineering could be directed to enhance these interactions. 260 Many research challenges need to be addressed before we can use the analysis of soil 261 networks to design management interventions that optimise connections between plant 262 roots and soil networks and between rhizosphere and bulk soil. However, the 263 effectiveness of many new agricultural management approaches for improving soil 264 health and agricultural sustainability (Tilman et al. 2002) can be explained by our 265 conceptual model. For example, it is well known that no-till or reduced tillage supports 266 active decomposer communities (Arshad et al. 1990; Ogle, Swan & Paustian 2012), but 267 they also promote linkages between rhizosphere and bulk soil networks by reducing soil 268 disturbance. Cover crops displace fallow periods with plants that fix N, increase C 269 inputs (Olson, Ebelhar & Lang 2010), and also likely enhance soil network structure. 270 Along with other forms of organic inputs such as manure and compost, cover crops may 271 support diverse bulk soils with weak interactions (Ling et al. 2016). Finally, crop 272 rotations that prevent the build-up of diseases have been shown to differ in their fungal 273 network structure from diseased soils under continuous monoculture, with rhizosphere 274 networks in healthy soils showing higher modularity and meta-modularity, and more 275 highly-connected generalists (Lu et al. 2013). However, it is important to acknowledge 276 that soil organismal networks and their connections with plants are likely to depend on 277 edaphic soil characteristics, such as soil texture and cation exchange capacity (Ma et al. 278 2016a; Ma et al. 2016b). The design of novel management approaches to promote 279 networks will have to take into account context-specificity and be tailored for different 280 crops, soil types, and climatic conditions.

281 Over the years, various indices have been proposed as indicators of healthy soils 282 including soil organic matter content (Reeves 1997), microbial diversity (Lehman et al. 283 2015), fungal:bacterial ratios (De Vries *et al.* 2006), enzyme activities (Bandick & Dick 284 1999), metabolic quotients (Anderson 2003), or soil respiration burst tests (Morrow et 285 al. 2016). These indices do not recognize the importance of interactions among these 286 components, but we expect many of these metrics to be correlated to the structure of 287 soil organismal networks, as has been shown already for certain soil food web 288 characteristics (De Vries et al. 2013). While much research is needed to establish links 289 between soil properties, plant traits, network structure and soil functioning, novel 290 understanding of ecological interactions within and between soil communities and 291 plants has tremendous potential to be applied in agricultural systems to sustainability 292 produce food for a growing global population.

293

Box 1. Networks and food webs – interaction types, configuration, and stability

295 Ecological networks are webs of connections between organisms. Connections in a 296 network can consist of a range different interaction types, from predator-prev 297 interactions to mutualisms, competition, and coexistence. Most studies of microbial 298 networks focus on bacteria and only include significant, positive interactions that might 299 consist of mutualisms, cooperation, or niche sharing. In contrast, classical soil food webs 300 only consist of feeding interactions, which can be positive (bottom-up) or negative (top-301 down) (but see Buchkowski (2016) and Hawlena & Zaguri (2016) for arguments for 302 including non-feeding interactions in soil food webs). However, networks can include 303 any functional group and type of interaction, and hybrids between microbial networks 304 and traditional soil food webs, including both positive and negative interactions, might 305 be more informative than separate networks.

306 Seminal work by May (1973) focussed on the number of organisms or species present in 307 a network and the number of connections between these. Counterintuitively, May found 308 that more diverse networks were less stable under perturbations than simpler 309 networks. However, these early networks were randomly created, and since then, 310 multiple studies have shown that the configuration and interaction types within 311 networks determine their stability. For example, networks that consist of many weak 312 links are more stable than those consisting of few strong links (Neutel, Heesterbeek & de 313 Ruiter 2002). Compartmentalisation, as well as the presence of weak connectors 314 between compartments, has also been shown to increase the stability of networks

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315 (Moore et al. 2003; Rooney et al. 2006; Stouffer & Bascompte 2011). Finally, trophic 316 coherence and the presence of negative interactions, such as top-down controls, can 317 increase network stability (Johnson et al. 2014; Coyte, Schluter & Foster 2015). 318 We currently lack enough examples to determine whether there are general 319 configurations in microbial networks, how these networks are affected by changes in 320 agricultural management, and what the implications are for their stability and 321 functioning. In contrast, the effects of agricultural management on soil food webs and 322 their functioning are relatively clear. Traditionally, food webs have been divided into 323 clear energy channels, or compartments: the root energy channel, which is fuelled by 324 live roots through the activities of root-feeding nematodes and mycorrhizal fungi, and 325 two detritus-fuelled compartments: the fungal and the bacterial energy channel. While 326 these compartmentalisations are currently hotly debated and under revision 327 (Ballhausen & de Boer 2016; Geisen 2016, see Figure), agricultural intensification 328 reduces the biomass of the root and fungal energy channels, and thereby increases the 329 relative importance of the bacterial energy channel. Experimental and modelling studies 330 have shown that the fungal energy channel, which consists of slow growing organisms 331 and weak interactions, is more stable under disturbance than the bacterial energy 332 channel and continues to function better (Rooney *et al.* 2006; De Vries *et al.* 2012a). It is 333 assumed that the root energy channel and the fungal energy channel are controlled by 334 bottom-up interactions, while the bacterial energy channel is top-down controlled 335 (Moore et al. 2003). Organisms that feed on multiple functional groups, such as 336 predatory nematodes and mites, can connect these compartments, and thereby increase 337 food web stability by dampening fluctuations in their prey (Rooney *et al.* 2006).

338

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643 Figure captions

644 <u>Figure in Box</u>

645 An example of a network (left) and a soil food web (right). A network consists of nodes 646 (circles) that are connected by edges (lines) through weak and strong, positive and 647 negative interactions; here, strong connections have a heavier line weight than weak 648 connections, positive interactions are grey and negative interactions are black. Modules 649 are indicated by different colours, and connectors between modules are in red. Soil food 650 webs are traditionally represented by three compartments, as indicated by arrow 651 colours. The fungal and bacterial energy channels (yellow and orange arrows 652 respectively) are fuelled by aboveground litter, root litter, and root exudates (grey 653 ellipse); the root energy channel (purple arrows) is fuelled by live roots. However, 654 recent evidence suggests that bacteria can feed on fungal hyphae, and protozoa are part 655 of both the fungal and the bacterial energy channel (as indicated by grey dashed 656 arrows), thus increasing the number of connections within the soil food web and 657 challenging the concept of distinct energy channels.

658

659 <u>Figure 1</u>

660 Rhizosphere networks consist of relatively few but highly abundant and connected 661 species that are recruited from the much more diverse but weakly connected bulk soil 662 network. Connections between rhizosphere and bulk soil networks (indicated by red 663 dotted lines) are crucial for the recruitment of functional rhizosphere networks. 664 Understanding and manipulating network structure of both rhizosphere and bulk soil 665 networks in agricultural soils, and the connections between them, is a promising avenue for optimising healthy soils and the benefits they provide for sustainable food 666 667 production.



Figure in Box. # + An example of a network (left) and a soil food web (right). A network consists of nodes (circles) that are connected by edges (lines) through weak and strong, positive and negative interactions; here, strong connections have a heavier line weight than weak connections, positive interactions are grey and negative interactions are black. Modules are indicated by different colours, and connectors between modules are in red. Soil food webs are traditionally represented by three compartments, as indicated by arrow colours. The fungal and bacterial energy channels (yellow and orange arrows respectively) are fuelled by aboveground litter, root litter, and root exudates (grey ellipse); the root energy channel (purple arrows) is fuelled by live roots. However, recent evidence suggests that bacteria can feed on fungal hyphae, and protozoa are part of both the fungal and the bacterial energy channel (as indicated by grey dashed arrows), thus increasing the number of connections within the soil food web and challenging the concept of distinct energy channels.!! +

144x82mm (300 x 300 DPI)



Figure 1. # + Rhizosphere networks consist of relatively few but highly abundant and connected species that are recruited from the much more diverse but weakly connected bulk soil network. Connections between rhizosphere and bulk soil networks (indicated by red dotted lines) are crucial for the recruitment of functional rhizosphere networks. Understanding and manipulating network structure of both rhizosphere and bulk soil networks in agricultural soils, and the connections between them, is a promising avenue for optimising healthy soils and the benefits they provide for sustainable food production. # +

258x310mm (300 x 300 DPI)