Pasireotide Long-Acting Release Treatment for Diabetic Cats with Underlying Hypersomatotropism

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Background: Long-term medical management of hypersomatotropism (HS) in cats has proved unrewarding. Pasireotide, a novel somatostatin analogue, decreases serum insulin-like growth factor 1 (IGF-1) and improves insulin sensitivity in cats with HS when administered as a short-acting preparation.

Objectives: Assess once-monthly administration of long-acting pasireotide (pasireotide LAR) for treatment of cats with HS.

Animals: Fourteen cats with HS, diagnosed based on diabetes mellitus, pituitary enlargement, and serum IGF-1 > 1000 ng/mL.

Methods: Uncontrolled, prospective cohort study. Cats received pasireotide LAR (6–8 mg/kg SC) once monthly for 6 months. Fructosamine and IGF-1 concentrations, and 12-hour blood glucose curves (BGCs) were assessed at baseline and then monthly. Product of fructosamine concentration and insulin dose was calculated as an indicator of insulin resistance (Insulin Resistance Index). Linear mixed-effects modeling assessed for significant change in fructosamine, IGF-1, mean blood glucose (MBG) of BGCs, insulin dose (U/kg) and Insulin Resistance Index.

Results: Eight cats completed the trial. Three cats entered diabetic remission. Median IGF-1 (baseline: 1962 ng/mL [range 1051–2000 ng/mL]; month 6: 1253 ng/mL [524–1987 ng/mL]; P < .001) and median Insulin Resistance Index (baseline: 812 µmolU/L kg [173–3565 µmolU/L kg]; month 6: 135 µmolU/L kg [0–443 µmolU/L kg]; P = .001) decreased significantly. No significant change was found in mean fructosamine (baseline: $494 \pm 127 \mu mol/L$; month 6: $319 \pm 113.3 \mu mol/L$; P = .07) or MBG (baseline: $347.7 \pm 111.0 mg/dL$; month 6: $319.5 \pm 113.3 mg/dL$; P = .11), despite a significant decrease in median insulin dose (baseline: 1.5 [0.4-5.2] U/kg; 6 months: 0.3 [0.0-1.4] U/kg; P < .001). Adverse events included diarrhea (n = 11), hypoglycemia (n = 5), and worsening polyphagia (n = 2).

Conclusions and Clinical Importance: Pasireotide LAR is the first drug to show potential as a long-term management option for cats with HS.

Key words: Acromegaly; Cat; SOM230; Somatostatin.

Hypersomatotropism (HS) is an increasingly recognized cause of diabetes mellitus (DM) in cats, with prevalence estimates among diabetic cats ranging from 17.8% in Switzerland and the Netherlands¹ to 24.8% in the United Kingdom.² Hypersomatotropism in cats usually results from excessive growth hormone (GH)

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Abbreviations:

ALT	alanine aminotransferase				
BGC	blood glucose curve				
CT	computed tomography				
DKA	diabetic ketoacidosis				
DM	diabetes mellitus				
GH	growth hormone				
HS	hypersomatotropism				
IGF-1	insulin-like growth factor 1				
LAR	long-acting release				
MBG	mean blood glucose				
RI	reference interval				
RVC	Royal Veterinary College				
SD	standard deviation				
SSTR	somatostatin receptor				
SST	somatostatin				

secretion from a functional somatotroph adenoma in the pituitary gland.³ Growth hormone's insulin-antagonistic effects cause most cats with HS to be diabetic, with many showing insulin resistance and persistent signs of poor glycemic control despite exogenous insulin administration.⁴ The mitogenic effects of GH excess, which are mediated by insulin-like growth factor-1 (IGF-1),⁵ can ultimately lead to body tissue proliferation and the clinical syndrome of acromegaly. In cats, clinical acromegaly is associated with physical changes, which can include prognathism inferior, broadening of the face, cardiomyopathy, pancreatopathy, and degenerative arthropathy.^{6,7}

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Treatment of HS in cats should aim to improve diabetic control and prevent progression of clinical acromegaly. Transsphenoidal hypophysectomy is the recommended first-line treatment for acromegalic people.⁸ Hypophysectomy has successfully been used to treat HS in cats,^{9,a} but availability of surgery is limited. In people, medical treatment is used to control HS when surgery has failed, been declined, or is contraindicated, and commonly used drugs include long-acting forms of the first-generation somatostatin (SST) analogues, lanreotide^b and octreotide.^{c,10} First-generation somatostatin analogue treatment leads to biochemical control of acromegaly in up to 54% of people, and pituitary adenoma shrinkage in 30-50% of people.^{10,11} A minority of cats show a short-term decrease in GH concentration when treated intravenously with octreotide,¹² but long-term medical management of HS in cats has otherwise proven unrewarding, and treatment with long-acting octreotide, or l-deprenyl, which is a monoamine oxidase-B inhibitor, has failed to provide either clinical improvement and biochemical improvement.13,d

Somatostatin binds to 5 receptor subtypes, named somatostatin receptors (STTR) 1-5, which are variably expressed in human pituitary tissue.¹⁴ Octreotide and lanreotide predominantly bind to STTR 2^{15,16}, and low SSTR2 expression is a proposed reason for why some acromegalic people fail to respond to these treatments.¹⁴ Preliminary work on pituitary STTR expression in cats suggests that feline pituitaries of cats with HS show greater expression of STTRs 1 and 5 compared to STTR2.^e This finding could account for the poor response to octreotide treatment shown by cats with HS.^d

Recently, a multireceptor-binding STT analogue called pasireotide^f has been licensed as a long-acting, oncemonthly treatment for acromegalic people. Pasireotide binds to a wider range of STTR subtypes than firstgeneration STT analogues. Pasireotide can bind to STTRs 1, 2, 3, and 5, and shows a 30-, 5- and 39-times greater binding affinity for STTRs 1, 3, and 5, respectively, compared to octreotide, with a slightly lower affinity for STTR2.¹⁵ Pasireotide is effective in some acromegalic people who are resistant to first-generation STT analogues,¹⁷ and short-acting pasireotide has been shown to provide biochemical control, and improved insulin sensitivity, when used as a short-term treatment in acromegalic cats. The current study was performed to assess whether longacting pasireotide (pasireotide LAR) could offer effective, long-term treatment for diabetic cats with underlying HS.

Materials and Methods

This was an uncontrolled, 6-month, prospective, longitudinal cohort study. The study was approved by the Ethics Committee of The Royal Veterinary College (Royal Veterinary College Ethical Approval Number URN2011 1120), and informed consent was obtained from all participating owners. The study population was formed from a convenience sample of cats diagnosed with HS at The Royal Veterinary College (RVC) Acromegalic Cat Clinic. Trial participation was offered to cats diagnosed with HS at this clinic between December 2011 and January 2014. Diagnosis of HS was

based on the presence of DM, documented pituitary enlargement by computed tomography (CT), and a markedly increased serum IGF-1 concentration of >1000 ng/mL. Serum IGF-1 concentration was measured by a commercially available radioimmunoassay^g that had been previously validated in cats.¹⁹ This test has an upper limit of quantification of 2000 ng/mL, and values greater than this were assigned a value of 2000 ng/mL for the purpose of data analysis.

On the day of admission, all cats received a physical examination, echocardiography, abdominal ultrasonography, urinalysis, and measurement of CBC, serum biochemistry, fructosamine concentration, and IGF-1 concentration. Cats were excluded if these tests revealed relevant underlying disease that might affect successful trial completion. This included diseases with poor prognosis and diseases for which treatment was likely to interfere with trial commitments. Cats then received pituitary imaging under sedation by contrast-enhanced CT, as previously described,20 and a dorsoventral pituitary height of >4 mm was considered to be consistent with enlargement.²¹ Maximal pituitary height was recorded for each cat. Other recorded baseline data included signalment, bodyweight, and whether phenotypic signs of acromegaly were present. Cats then underwent a 12-hour blood glucose curve (BGC) while hospitalized using their usual insulin type and dose. Blood glucose curves were carried out a minimum of 12 hours after the above investigations and were performed by a handheld glucometer previously validated for cats.^h Curves consisted of a reading taken every 2 hours over a 12-hour period, starting at the time of morning food and insulin administration.

Cats received 8 mg/kg of pasireotide LAR by subcutaneous injection before discharge and once monthly thereafter for a planned 6-month follow-up period. Pasireotide LAR dose was based on manufacturer's recommendations. The majority of included cats received 3 days of short-acting pasireotide treatmentⁱ between collection of their baseline data and their first pasireotide LAR dose. This short-acting pasireotide was given as part of a separate, previously reported trial.¹⁸ Baseline data in these cats, including their baseline BGC, were recorded before any pasireotide, either short- or long-acting, was administered.

Monthly re-examination appointments and pasireotide LAR administration were carried out at the RVC Acromegalic Cat Clinic. Owners were asked to perform a 12-hour BGC on their cat at home, or at their primary veterinary practice, in the 5 days before each reexamination. At each re-examination, cats received a physical examination, and fructosamine and IGF-1 measurement in addition to pasireotide LAR administration. Adjustments in insulin dose were made based on the clinical opinion of the clinical trial team. Cats who remained normoglycemic without insulin or oral antihyperglycemic medications (with the exception of low-carbohydrate diet) for a minimum of 4 weeks were classified as being in diabetic remission. Cats received a repeated CBC, serum biochemistry measurement, and abdominal ultrasound examination at the month 6 time point, or sooner if clinically indicated, to monitor for abnormalities that might be related to pasireotide treatment. Suspected adverse events, the reason for any trial withdrawals, and any alterations made in pasireotide LAR dosage were recorded. Cats remaining in the study at the end of the 6-month period received repeated pituitary imaging by contrast-enhanced CT at the month 6 time point to assess for change in pituitary size. Change in phenotypic signs of acromegaly was not evaluated as part of this study. It was thought this would be challenging to assess because phenotypic signs of feline acromegaly are often only subtle when judged by CT.²⁰

Statistical Analysis

Average values for variables were reported as mean \pm standard deviation (SD) for normally distributed data and as median with range for non-normal data. Normality was judged by histograms, normal plots, and Shapiro-Wilk tests.

A linear mixed-effects model with first-order autoregressive covariance structure was used to assess for significant change in fructosamine concentration, IGF-1 concentration, insulin dose given at each twice-daily insulin administration (U/kg), and mean blood glucose (MBG) concentration during BGCs over the 6month trial. Similar to the insulin sensitivity product assessed in the previous short-acting pasireotide trial,¹⁸ for the purpose of this study, the product of fructosamine and twice-daily insulin dose (U/kg) was calculated as a surrogate indicator of insulin resistance, referred to as the Insulin Resistance Index. This enabled documentation of improvements in insulin sensitivity by integrating glycemic changes with exogenous insulin dose changes. Change in this variable over the duration of the trial was also assessed by a linear mixed-effects model with first-order autoregressive covariance structure, and values were logarithmically transformed before analysis to meet the model's assumptions. For variables that showed a significant change, Fisher's least significant difference post hoc comparison was used to assess pairwise differences between each time point and month 0.

The maximal pituitary height at month 0 and month 6 time points in cats completing the study was compared by a Wilcoxon signed-rank test. Significance for all tests was set at P < .05, and all analyses were performed by commercial statistical software packages.^j, ^k

Results

Thirty-three cats examined during the study period were eligible for inclusion. Of these, 8 owners declined trial participation and 11 owners decided to manage their cat's HS by hypophysectomy. Fourteen cats were therefore enrolled onto the trial. Twelve of these cats received 3 days of short-acting pasireotide before receiving their first dose of pasireotide LAR.¹⁸ The remaining 2 cats were enrolled after the short-acting pasireotide trial had ceased recruiting participants and so received only pasireotide LAR.

Cat Characteristics at Enrollment

The study population consisted of 11 domestic shorthairs, 1 domestic longhair, 1 Persian, and 1 Burmese. Ten cats were neutered males and 4 were neutered females. All cats had clinical signs of poor diabetic control, and 12 of the 14 cats had phenotypic changes consistent with acromegaly on physical examination. Mean age at enrollment was 9.9 ± 2.2 years, and mean bodyweight was 5.8 ± 1.1 kg. Complete blood cell counts revealed 1 cat to have a stress leukogram, 1 cat to have mild, nonregenerative anemia (PCV 21%; reference interval (RI) 24-45%), and 1 cat to have a moderate, mature neutrophilia, but revealed no other abnormalities. Serum biochemistry analyses at trial entry revealed no abnormalities, apart from hyperglycemia in all cats. Abdominal ultrasound findings noted at trial entry included bilateral adrenomegaly (n = 9), bilateral renomegaly (n = 8), hyperechoic hepatic parenchyma (n = 6), hepatomegaly (n = 5), and suspected pancreatopathy (n = 4), with most cats having more than 1 abnormality. For the purpose of this trial, adrenomegaly was defined as an adrenal width of >4.8 mm, as proposed by a previous study.²²

Cats had been receiving insulin treatment for a median of 231 (63-122) days before trial enrollment. All cats were receiving twice-daily (q12h) insulin with 10 cats receiving porcine lente insulin,¹ 2 cats receiving protamine zinc insulin^m, and 2 cats receiving insulin glargine.ⁿ No cats changed insulin type at the time of starting pasireotide LAR, but 6 cats changed insulin type later in the study period. Five cats were changed from porcine lente to glargine treatment, and 1 cat was changed from protamine zinc insulin to porcine lente insulin. These changes in insulin treatment occurred a median of 3 months (2–4 months) into the trial period.

Table 1 shows the number of cats remaining in the trial at each time point and average values for variables during the trial. Before starting pasireotide treatment, median q12h insulin dose was 1.5 U/kg (0.4–5.2 U/kg), median IGF-1 concentration was 1962 ng/mL (1051–2000 ng/mL), and mean fructosamine concentration was 494 \pm 127 µmol/L. Mean blood glucose (MBG) concentration during 12-hour BGCs before pasireotide treatment was 347.7 \pm 111.0 mg/dL (19.3 \pm 6.2 mmol/L). Median maximal pituitary height on CT at enrollment was 7.2 mm (5.5–12.0 mm).

Response to Treatment

Three cats (21%) entered diabetic remission. A fourth cat stopped insulin treatment for 15 days but then became hyperglycemic again so did not meet criteria for remission. In the 3 cats that achieved remission, insulin was discontinued 89 days (n = 2) and 91 days (n = 1) after their first pasireotide LAR dose (Fig 1). All cats that achieved remission continued to receive oncemonthly pasireotide LAR, and all completed the 6-month trial without a recurrence of DM. After trial completion, 2 cats remained in remission until death 479 and 580 days after starting pasireotide LAR, and the third cat is alive and in diabetic remission at the time of writing, 1420 days after starting pasireotide LAR treatment.

A significant decrease was detected in IGF-1 concentration (P=<.001), q12h insulin dose (P=<.001), and Insulin Resistance Index (P = .001) over the trial period. Post hoc analysis revealed a statistically significant difference between month 0 and all subsequent time points for all 3 variables (P-values shown in Table 1 and Figures 2, 3, and 4).

There was no significant change in serum fructosamine (P = .07) or MBG during BGCs (P = .11) over the study period (Table 1 and Figs 5 and 6). In the 8 cats that completed the 6-month trial, maximal pituitary height on CT examination changed by a median of -0.5 mm (-1.8 to +1.3 mm) (P = .16).

Adverse Events

Various adverse effects were recorded. Diarrhea, of varying severity, was the most common adverse effect and was experienced by 11 cats (76%). Ten of these cats developed diarrhea within 1 month of their first pasireotide LAR dose, and 1 cat developed diarrhea within 2 months of starting pasireotide LAR treatment. In 2 cats, diarrhea was transient and resolved within 2 months of starting treatment. Four cats had their

Time Point (month)	Number of Cats Remaining in Study	IGF-1 Concentration (ng/mL) ^a	Twice-Daily Insulin Dose (U/kg) ^a	Fructosamine Concentration (µmol/L) ^b	MBG During BGCs (mg/dL) ^b	Insulin Resistance Index (μmolU/L kg) ^a
0	14	1962 (1051–2000) n = 14	1.5 (0.4-5.2) n = 14	494 ± 127 n = 14	347.7 ± 111.0 n = 13	812 (173-3565) n = 14
1	12	909 (105–2000)*** n = 12	$1.3 (0.1-1.3)^{***}$ n = 12	437 ± 126 n = 12	302.2 ± 159.8 n = 12	155 (43-774)*** n = 12
2	10	$836 (415-1937)^{***}$ n = 10	$0.4 \ (0.0-1.5)^{***}$ n = 10	406 ± 116 n = 9	260.6 ± 128.8 n = 9	$99 (0-601)^{**}$ n = 9
3	9	1076 (386-2000)** n = 9	$0.2 \ (0.0-1.7)^{***}$ n = 9	483 ± 152 $n = 8$	283.7 ± 154.4 n = 9	$146 (0-850)^{**}$ n = 9
4	9	1437 (841-1613)* n = 9	$0.1 (0.0-1.2)^{***}$ n = 9	370 ± 143 n = 9	191.9 ± 115.1 n = 6	40 (0-830) *** n = 9
5	8	971 (520–1665)*** n = 8	n = 9 0.3 (0.0–1.0)*** n = 8	399 ± 132 n = 8	11 = 0 213.2 ± 96.3 n = 6	n = 9 29 (0-522)** n = 8
6	8	n = 8 1253 (524–1987)** n = 8	n = 8 0.3 (0.0–1.4)*** n = 8		n = 0 319.5 ± 113.3 n = 6	n = 8 135 (0-443)** n = 8

Table 1. Number of cats remaining in the study at each time point and average values for variables monitored during the trial.

^aIndicates variable described using median (range).

^bIndicates variable described using mean \pm standard deviation. n = number of patients with available measurement at that time point. IGF-1, insulin-like growth factor-1; MBG, mean blood glucose; BGC, blood glucose curve. *, ** and *** indicate a statistically significant difference compared to Month 0 value (* indicates P < .05, ** indicates P < .01, *** indicates P < .001).

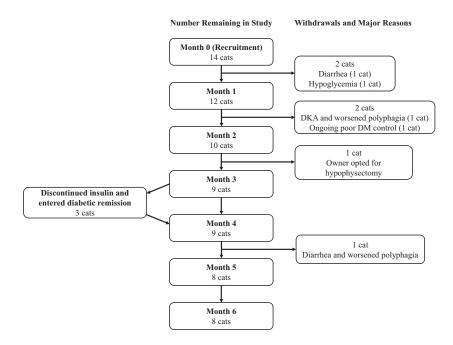


Fig 1. Flow diagram showing the number of cats remaining in the study at each time point, and the timing of study withdrawals and diabetic remission. Major reasons for study withdrawals are also shown. DKA, diabetic ketoacidosis; DM, diabetes mellitus.

diarrhea treated with one, or a combination, of exclusion diet treatment (n = 4), pancreatic enzyme supplementation (n = 3), parenteral cobalamin injections (n = 1), and soluble fiber supplementation (n = 1). Two cats had their monthly pasireotide LAR dose decreased to 6 mg/kg in the hope of improving their diarrhea. These management changes caused a mild-to-negligible improvement. One cat experienced 2 hypovolemic episodes that were suspected to be secondary to bouts of diarrhea. Severe diarrhea was a reason for trial withdrawal in 2 cats (Fig 1). Six hypoglycemic events were documented in the study population and occurred in 5 cats with 1 cat experiencing 2 episodes. Five of the 6 episodes were associated with clinical signs. Other adverse effects were worsening polyphagia in 2 cats, a transient injection site reaction in 1 cat, and delayed hair regrowth in 1 cat.

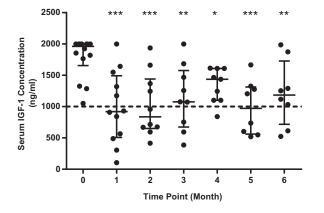


Fig 2. Scatterplot showing individual serum IGF-1 concentrations at each study time point. Central horizontal bar indicates median value with error bars indicating interquartile range. Dashed line shows recommended cutoff for screening for HS in cats (1000 ng/mL). *, ** and *** indicate a statistically significant difference compared to Month 0 value (* indicates P < .05, ** indicates P < .01, *** indicates P < .001).

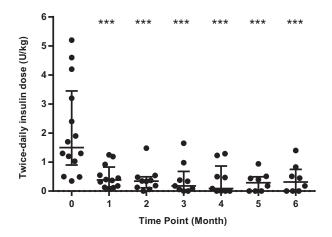


Fig 3. Scatterplot showing individual twice-daily insulin doses at each study time point. Central horizontal bar indicates median value with error bars indicating interquartile range. *** indicate a statistically significant difference compared to Month 0 value with P < .001.

Repeated CBC results at month 6 revealed no abnormalities, apart from mild eosinophilia in 1 cat. Repeated serum biochemical analyses at month 6 also revealed no abnormalities, apart from moderate hypercholesterolemia in 1 cat, and a mild-to-moderate increase in serum alanine aminotransferase (ALT) activity in 2 cats (126 U/L and 274 U/L; RI 20-100 U/L). Both cats with ALT elevation remained clinically well, apart from diarrhea in 1 cat, and neither showed other biochemical indicators of hepatic dysfunction. One cat with severe diarrhea underwent repeated serum biochemistry measurement at the month 2 time point, but this revealed no abnormalities. The only notable changes on repeated abdominal ultrasonography at month 6, compared to trial entry, were intestinal muscularis layer thickening in 2 cats, and gall bladder sediment in 1 cat. Both cats

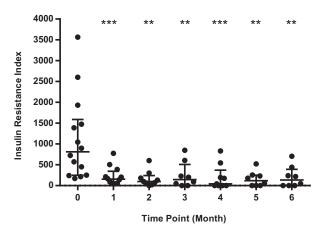


Fig 4. Scatterplot showing individual Insulin Resistance Index values (fructosamine \times q12h insulin dose in U/kg) at each study time point. Central horizontal bar indicates median value with error bars indicating interquartile range. ** and *** indicate a statistically significant difference compared to Month 0 value (** indicates P < .01, *** indicates P < .001).

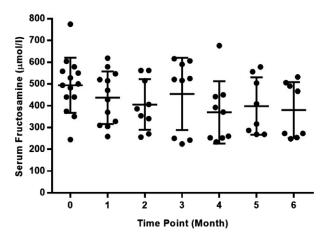


Fig 5. Scatterplot showing individual serum fructosamine concentrations at each study time point. Central horizontal bar indicates mean value with error bars indicating 1 standard deviation.

with muscularis layer thickening experienced diarrhea as an adverse effect of pasireotide LAR treatment. The cat with gall bladder sediment was 1 of 2 cats that developed an increased serum ALT activity.

Trial Completion and Withdrawals

Eight cats (57%) completed the trial. Trial withdrawals occurred after a median of 2 months (1–5 months). Figure 1 shows the timing of each withdrawal and the major reasons for which cats stopped trial treatment. Withdrawals occurred for a variety of reasons with some having more than 1 contributing factor. Reasons included severe diarrhea (n = 2), worsened polyphagia (n = 2), failure of diabetic control to improve (n = 1), and euthanasia after a hypoglycemic event (n = 1) or episode of diabetic ketoacidosis

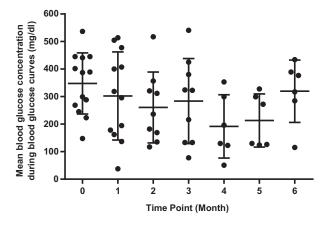


Fig 6. Scatterplot showing individual mean blood glucose values during blood glucose curves at each study time point. Central horizontal bar indicates mean value with error bars indicating 1 standard deviation.

(n = 1). One cat was withdrawn from the study after the owner decided to manage its HS by hypophysectomy rather than pasireotide LAR.

Discussion

This study found that once-monthly pasireotide LAR treatment was associated with a significant reduction in IGF-1 concentration in diabetic cats with HS, suggesting that pasireotide LAR significantly improves excessive GH production from feline pituitary somatotrophinomas. Several pasireotide-treated cats achieved long-standing diabetic remission, which rarely occurs when diabetic cats with HS are treated with routine diabetic management alone.²³ Cats receiving pasireotide LAR experienced a significant decrease in insulin requirement and estimated insulin resistance, although serum fructosamine concentration and MBG did not significantly change over the study period.

Previous attempts at long-term medical management of HS in cats have proven unsuccessful,^{13,d} making pasireotide LAR the first drug to show potential as a long-term management option for feline HS. Octreotide is the only other long-acting STT analogue to have been assessed as a treatment for feline HS, but was found to be ineffective.^d Together, these findings are similar to those of recent human trials, which have shown pasireotide LAR to be superior to long-acting octreotide treatment.^{17,24} This superiority has been attributed to pasireotide's wide receptor range and binding affinity, which allow it to accommodate variations in STTR expression in patients' somatotrophinomas. Octreotide predominantly binds to STTR2, with lesser affinity for STTR3 and 5, whereas pasireotide binds to STTRs 1. 2. 3, and 5 and also shows a substantially greater binding affinity for STTRs 1, 3, and 5 compared to octreotide.¹⁵ The biochemical improvement shown by cats in this study, and the poor previous experience with octreotide treatment, supports that the pituitaries of cats with HS have greater expression of STTRs 1 and 5 compared to STTR2.^e

That several cats in the current study achieved diabetic remission supports that pasireotide LAR offers a viable medical treatment option for HS in cats. Diabetic cats with HS are most likely to achieve remission when treated with a therapy that successfully addresses their excess GH production.³ Diabetic remission can occur in cats whose HS is treated by radiotherapy²⁵ or hypophysectomy,⁹ with a recent study reporting a 78% remission rate among cats who survived more than 17 days after hypophysectomy surgery.^a In contrast, treatment that addresses a cat's DM alone, such as insulin and low-carbohydrate diet, typically provides inadequate glycemic control and little chance of remission.²³ The presence of remission among cats in the current trial therefore supports that pasireotide LAR effectively reduces GH secretion from somatotrophinomas in cats and improves insulin sensitivity in treated cats. Pituitary radiotherapy for HS in cats requires frequent anesthesia and treated cats can have an unpredictable response to treatment.²⁵ Hypophysectomy seems more predictably successful and requires only a single anesthetic, but is offered at reasonably few veterinary centers, is associated with a postoperative fatality rate of approximately 14% and requires lifelong, postoperative hormone supplementation.^a Pasireotide LAR could therefore provide a viable alternative option for control of HS in cats that are poor surgical or anesthetic candidates, or have no access to hypophysectomy. However, the common occurrence of diarrhea, need for regular injections, and lack of information regarding ideal dosing frequency still pose challenges to its use.

This study did not identify a significant difference in MBG, or fructosamine concentration, over the duration of the trial. Lack of change in MBG could be secondary to how BGCs, from which MBG was calculated, were performed just before each cat's next monthly pasireotide LAR injection. As the LAR is designed to last 1 month, it is likely that each cat's previous pasireotide LAR treatment was waning by this time leading to worsening diabetic control at the end of each monthly treatment period and no apparent improvement in MBG. The once-monthly dosing frequency used in this trial was based on recommendations from pasireotide LAR treatment in people, and further work would be necessary to establish the optimum dosing frequency in cats. Mean blood glucose results were also likely to be artificially increased toward the end of the study because 2 owners whose cats went into diabetic remission stopped performing monthly blood glucose curves, opting instead to monitor their cat with intermittent blood glucose measurements. Serum fructosamine concentration is generally accepted to reflect cats' glycemic control over the preceding 1–2 weeks²⁶ and so sampling at the end of each monthly treatment period is less likely to account for the lack of change in serum fructosamine among recruited cats. This may instead have been caused by many cats already having a fructosamine concentration consistent with good-tomoderate glycemic control at study entry, though only when receiving relatively high dosages of exogenous insulin.

The fact that fructosamine and MBG remained unchanged, despite a significant decrease in insulin dose, suggests that pasireotide LAR is associated with improved insulin sensitivity in treated cats. A simple Insulin Resistance Index was used to illustrate this in the current trial and showed a significant decrease at all time points compared to time of enrollment. This index has not been previously evaluated in cats, but the use of reference methods for estimating insulin resistance, such as the hyperinsulinemic euglycemic clamp or frequently sampled intravenous glucose tolerance test, was thought to be too invasive to justify their use on client-owned cats. Simpler methods for evaluating insulin sensitivity, such as the Homeostasis Model Assessment (HOMA) and Quantitative Insulin Sensitivity Check Index (QUICKI), have been used in diabetic cats.²⁷ However, the fasting insulin and glucose values required for these calculations were not available for cats in this study.

This study revealed pasireotide LAR treatment to be associated with various adverse effects, the most common being diarrhea. Diarrhea is a common adverse effect of all long-acting STT analogue therapies in people^{17,24} and is caused by STT's widespread actions in the gastrointestinal tract, which include suppression of gastrointestinal exocrine secretions, inhibition of gastric and gallbladder emptying, and alterations in intestinal motility.¹⁶ Alopecia is a recognized adverse effect of pasireotide LAR treatment in people²⁴, and 1 cat in the current study showed delayed fur regrowth. This might have been secondary to pasireotide treatment. Alternatively, this cat could have been affected by concurrent hyperadrenocorticism, which is associated with DM, insulin resistance and the presence of a pituitary adenoma, and has been reported to occur alongside hypersomatotropism in cats.²⁸ This cat did not receive pituitary-adrenal axis testing and did not have adrenomegaly detected on abdominal ultrasound examination. Hypoglycemia was the second most common adverse event in the present study, whereas hyperglycemia and development of DM are frequent adverse effects in people receiving pasireotide LAR.²⁹ Pasireotide-associated hyperglycemia is thought to result from decreases in insulin and incretin secretion.³⁰ In people, STT decreases insulin secretion by binding to SSTR5 on pancreatic β -cells,³¹ and pasireotide's high affinity for STTR5 therefore reduces insulin secretion in treated patients. The mechanism by which pasireotide decreases incretin secretion is currently unknown. The frequency of hypoglycemia in the current study and the fact that 3 cats entered diabetic remission suggest that STT's effect on insulin and incretin secretion are mediated differently in cats compared to people, or that improved insulin sensitivity likely outweighs any decrease in insulin and incretin secretion in treated cats.

Two cats in the current study developed a mildto-moderate increase in ALT activity while receiving pasireotide LAR, and a third cat developed gall bladder sediment. Increased serum ALT activity occurs in up to 5% of acromegalic people treated with pasireotide LAR,³² whereas cholelithiasis and gall bladder abnormalities are more common adverse effects of long-acting

STT analogue treatment in people. Gall bladder stones or sediment develop in up to 45% of acromegalic people treated with long-acting STT analogues, and often occur in the absence of liver enzyme elevations.^{24,33} Several factors contribute to the predisposition for cholelithiasis among acromegalic people receiving longacting STT analogue treatment. Somatostatin and its analogues reduce meal-stimulated cholecystokinin secretion from the small intestine, which decreases gall bladder emptying.³⁴ Also, acromegalic people receiving octreotide are shown to have changes in bile composition, which predispose to cholelithiasis, including increased biliary cholesterol content and alterations in bile acid concentrations.³⁵ All cats who developed increased serum ALT activity, or ultrasonographic biliary tract abnormalities, in this study remained clinically well. However, it would nevertheless be advisable to monitor cats receiving pasireotide LAR by intermittent bloodwork and abdominal ultrasonography until the long-term tolerability of pasireotide LAR in cats has been better established.

Pasireotide LAR causes a significant reduction in tumor volume in up to 80% of treated people.²⁴ This is thought to be caused by STT, and STT analogues, having both direct and indirect effects on tumor growth. Somatostatin analogues induce cell cycle arrest and apoptosis through direct interaction with STTRs on tumor cells.^{36,37} Indirect effects include decreasing secretion of growth-promoting hormones and factors, such as GH and IGF-1, and by decreasing angiogenesis in the local tumor environment.³⁸ Octreotide decreased tumor vascularity in a small series of people with acromegaly,³⁹ which might be caused by its ability to decrease vascular endothelial growth factor expression.40 The failure to detect a significant change in pituitary height in the current study might be due to only a small number of cats completing the 6-month treatment period. Furthermore, pituitary tumor shrinkage is shown to increase with time in people who receive STT treatment over several years⁴¹ and so the relatively short duration of this trial may have been insufficient to observe a difference. A larger sample size and longer cat follow-up could therefore be required to demonstrate whether pasireotide LAR treatment is associated with tumor shrinkage in acromegalic cats.

This study has a number of limitations. People with HS are often monitored using both GH and serum IGF-1 concentration, whereas only serum IGF-1 concentration was measured in this study. This was firstly due to lack of a commercially available assay for feline GH. Secondly, GH release is pulsatile with a serum half-life of less than 20 minutes in humans,⁴² whereas IGF-1 is more constantly produced and has a half-life of approximately 15 hours.⁸ IGF-1 concentration is therefore the recommended screening test for HS in people, rather than single GH measurement.⁴³ IGF-1 is the main mediator for GH's mitogenic effects,⁵ so an IGF-1 concentration within normal limits is a treatment goal in people with HS because this represents clinical control of acromegaly.⁸ Interestingly, many of the cats in the current study failed to achieve an IGF-1

concentration below the recommended screening value for HS (<1000 ng/mL)⁴ during the 6-month trial (Fig 2), so might still have subclinical HS, despite pasireotide LAR treatment. Increasing pasireotide LAR dose, or dosing frequency, might result in more cats achieving a normalized IGF-1 value, but it is possible that adverse effects might become dose-limiting in many cats. Another limitation is that most included cats took part in a 3-day trial examining the effect of short-acting pasireotide treatment before their first injection of pasireotide LAR.¹⁸ Short-acting pasireotide's pharmacodynamics have not been studied in cats, but it has a half-life of approximately 12 hours in healthy people.⁴⁴ If the rate of elimination is similar in cats, previous short-acting pasireotide treatment is unlikely to have affected any patient values during this study. A proportion of cats also changed insulin type during the study, and it is possible this may have contributed to minor changes in these cats' diabetic control over the 6-month period. Nevertheless, insulin treatment alone is rarely successful in providing adequate glycemic control in diabetic cats with HS, regardless of the insulin type,²³ and it is therefore unlikely that any change in insulin type was responsible for the changes in diabetic control observed in the study population.

This study used a convenience sample formed from diabetic cats diagnosed with HS at the RVC Acromegalic Cat Clinic. All owners were offered hypophysectomy for management of their cat's HS, as an alternative to trial enrollment, at the initial examination. It is possible that cats whose owners opted for trial inclusion systematically differed from those who underwent hypophysectomy and that this resulted in selection bias when forming the study population. Treatment with pasireotide LAR was provided at no cost to participating cats, and this financial incentive is likely to have had a major influence on many owners' decision to partake in the trial. However, it is possible that other factors, such as large pituitary tumor size, might have caused clinicians to less strongly recommend hypophysectomy in certain cats. In support of this theory, a study of cats that underwent hypophysectomy at the same hospital over the same time period revealed them to have a smaller median pituitary height (6.0 mm; range 4.0-10.6 mm) than cats in the current study.^a Pituitary mass volume is unrelated to IGF-1 concentration in cats with HS.45 However, cats with larger tumors could be predisposed to deterioration and early trial withdrawal because of neurological compromise from their mass.⁴ Although this potentially could have negatively biased this study's observed response to pasireotide, no cats in the current trial were excluded due to neurological deterioration.

Inclusion of a control group would have helped to support that the observed changes in recruited cats were directly due to pasireotide treatment, rather than another explanatory factor. Inclusion of a control group was considered ethically controversial because cats with HS typically experience persistent signs of poor diabetic control when treated with routine DM management alone.²³ The fact that diabetic cats with HS predictably show a poor

response to routine diabetic treatment supports that the changes observed in the current trial are most likely to be related to pasireotide treatment. It is also possible that some suspected adverse events recorded in this study were actually unrelated to pasireotide treatment. However, diarrhea was the most common adverse event in this study and is also a common adverse effect of long-acting STT analogue treatment in people.^{17,24} This similarity and the fact that several other adverse effects observed in this trial are also reported in people receiving pasireotide LAR suggest that most adverse effects in this study were caused by pasireotide treatment. Finally, the pharmacokinetics of pasireotide LAR in cats are currently unknown and require future investigation in order to establish the optimum dosing protocol for pasireotide LAR as a treatment for HS in cats.

In conclusion, this study shows pasireotide LAR to be the first drug to show potential as a long-term management option for diabetic cats with HS, with treated cats showing a significantly decreased IGF-1, improved insulin sensitivity, and some cats achieving diabetic remission. Diarrhea is a significant adverse effect and further work is necessary to establish the optimum dosing strategy to provide effective diabetic control while limiting adverse effects.

Footnotes

- ^a Kenny P, Scudder C, Keyte S, et al. Treatment of Feline Hypersomatotropism – efficacy, morbidity and mortality of hypophysectomy [abstract]. J Vet Int Med. 2015; 29(4):1271
- ^b Somatuline[®] Depot, Ipsen Biopharmaceuticals Inc., Basking Ridge, NJ
- ^c Sandostatin[®] LAR[®], Novartis International AG, Basel, Switzerland
- ^d Timian J and Lunn KF. Evaluation of a long-acting somatostatin receptor ligand for the treatment of feline acromegaly. 2012 ACVIM Forum Research Abstracts Program. J Vet Int Med 2012; 26(3): 757
- ^e Scudder C, Mirczuk SM, Richardson K, et al. Feline hypersomatotropism as a spontaneous animal model of acromegaly: molecular analyses of somatostatin receptor expression in GHsecreting adenomas in the cat (Felis catus) [abstract]. ENDO2016, Boston, MA
- f Signifor® LAR, Novartis International AG, Basel, Switzerland
- ^g IGF-1 RIA-CT, Mediagnost[®], Reutlingen, Germany
- ^h Alphatrak2[®], Abbott Animal Health, Chicago, IL
- ⁱ Signifor[®], Novartis International AG, Basel, Switzerland
- ^j SPSSTM, IBM Corporation, Armonk, New York
- ^k GraphPad Prism[®], GraphPad Software Inc, California
- ¹ Caninsulin[®], Intervet UK Ltd, Milton Keynes, UK
- ^m Hypurin[®] Bovine PZI, Wockhardt UK Ltd, Wrexham, UK
- ⁿ Lantus[®], Sanofi, Paris, France

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Off-label Antimicrobial Declaration: Authors declare no off-label use of antimicrobials.

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