

1 **Physical and biological characteristics of multi drug resistance (MDR): An integral**
2 **approach considering pH and drug resistance in cancer.**

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20 Abstract

21 The role of the Warburg effect in cancer remains to be elucidated with a resurgence in research
22 efforts over the past decade. Why a cancer cell would prefer to use energy inefficient
23 glycolysis, leading to an alteration of pH both inside and outside of the cell, remains to be
24 uncovered. The development of MDR represents a major challenge in the treatment of cancer
25 and it is explained, so far, by the over expression of drug transporters such as the well-known
26 and archetypal P-glycoprotein (Pgp). However, controversies exist regarding the function of
27 Pgp in multi-drug resistance. We suggest here that Pgp-mediated MDR relies fundamentally
28 on pH alterations mediated by the Warburg effect. Furthermore, we propose that the use of
29 proton pump and/or transporters inhibitors (PPIs/PTIs) in cancer are key to controlling both
30 MDR, i.e. sensitize tumors to antineoplastic agents, and drug-related adverse effects.

31

32 **A lost connection between research fields**

33 Over time, fields of scientific research gain autonomy in proportion to the extent to which they
34 have been freed from economic necessity [1]. They develop their own laws and logics which
35 become field-specific and very often run contrary to those in surrounding fields. They develop
36 increasingly specialized research programmes and these can lead to great achievements. As the
37 classical German social theorist Max Weber observed, ‘only by strict specialization can the
38 scientific worker become fully conscious, for once and perhaps never again in his lifetime, that
39 he has achieved something that will endure’ [2]. However, Weber also saw the melancholy
40 aspect of ultra-specialization: it leads to the development of research fields that are
41 incommensurable and between which communication is increasingly difficult. It also leads to
42 scientists ‘putting the blinkers on’ in relation to developments outside their areas of expertise.
43 For example, the somatic mutation theory of cancer together with the “war on cancer” have
44 paved the way to great achievements in molecular biology (e.g. genome project) but their
45 applications to medicine, i.e. oncology, remain minimal since the “magic bullet”, i.e. the one
46 gene mutated – one drug concept, that was initially promised is still missing. The constant
47 refining process that accompanies ultra-specialization in scientific fields is comparable to that
48 which occurred in the field of abstract art where, through a process of purification that gradually
49 isolated it from all reference to the wider social world, it became almost entirely propelled by
50 its own inner dialectic [3]. We see here that in its ‘purified’ state, a field becomes inward-
51 looking.

52 The results of specialization can be seen in the sub-field of research on MDR in cancer, which
53 suffers from an inherent fundamental paradox. As early as 1973 the drug efflux hypothesis was
54 suggested by Dano Keld [4], which was reinforced in 1976 when Juliano and Ling discovered
55 Pgp in multi drug resistant cells [5]. Since then many works have been carried out to understand
56 the function of Pgp in MDR. However the single use of Pgp to explain MDR in cancer is flawed

57 as Pgp violates the law of enzyme affinity/specificity on which the entire field of molecular
58 biology is built: '*MDR protein is a very unusual enzyme with extraordinarily broad substrate*
59 *recognition capabilities; that is, it violates the law of enzyme specificity*' [6]. What is staggering
60 is that even with the presence of a true scientific paradox in Pgp-mediated MDR in cancer, a
61 range of stakeholders, whether economic and market-oriented (Big Pharma), institutional
62 (academia, research organizations) or political (government, pressure groups), have shared (for
63 most are still sharing) many of the same presuppositions about the problem of MDR in cancer
64 and how it might be combatted, although rare attempts exist to suggest changes of strategy in
65 the field of Pgp-mediated MDR [7].

66 Why is this so? As scientists we know it, because specialist research fields tend to engender in
67 scientists who have been trained in the field, and are thus attuned to its logic, an implicit sense
68 of what is the correct way of doing science and this can inhibit them from gaining insights from
69 other fields [8].

70 No one would contest the existence of drug efflux mediated by membrane pumps. The question
71 is simply that if membrane pumps exist in MDR cells, how can they work while, at the same
72 time, violating the law of enzyme specificity? Is it really drug transporters that are important
73 or have we overlooked essential components in multi drug resistant cancer cells?

74 When faced with an apparent paradox it is essential to step out from the discipline and research
75 around how similar issues are dealt with in other fields of enquiry. Understanding the
76 importance of pharmacokinetic / drug delivery is essential to uncover how a drug may or may
77 not cross the bilayer membrane of MDR cells.

78 To explore the existing connections between MDR in cancer and other fields one will start by
79 recalling concepts used in the field of pharmacokinetics that deals with similar barriers
80 constituted by ATP-ase drug transporters. We shall see that in this context, the Big Pharma
81 industry have focused on determining the optimal biophysical properties of drugs to cross those

82 barriers (irrespective of drug transporters). Next we shall investigate how those biophysical
83 properties emerge by a clearer understanding of membrane physics. This will allow one to
84 underline a number of studies that have emphasized the important role of the membrane in
85 MDR in cancer. We shall then explain how the notion of specificity or affinity is not required
86 as far as Pgp is involved. Finally, one will demonstrate how the Warburg effect and related
87 changes in pH are involved in changing the membrane in such a way to sustain Pgp activity
88 and MDR.

89 In conclusion we will discuss about the role of proton pump inhibitors (PPIs) and membrane-
90 bound proton transport inhibitors (PTIs) to circumvent MDR and improve drug efficacy in
91 cancer.

92

93 **The notion of pharmacokinetics and how it can help in understanding the MDR paradox**

94 The field of pharmacokinetics deals with how drug chemicals are dealt with by complex body
95 systems and as a result how drug chemicals reach their targets. Defining the drug transporters
96 that “cover” biological barriers has been essential for the success of the pharmaceutical
97 industry. The main difference between the field of molecular oncology and pharmacokinetics
98 is that the former works with simple systems (molecules and cells) whereas the later deals with
99 complex body systems. Looking at how the Big Pharma has dealt with biological barriers may
100 yield novel findings that could help to further define MDR.

101 The’ 90s were gloomy years for the pharmaceutical industry with productivity falling below
102 expectations and an average innovation deficit of ~1.3-1.8 for new chemical entities per year
103 [9]. During this period these companies adopted approaches that relied on retrieval of
104 information to determine if a chemical would make a ‘likely’ drug in advance of costly clinical
105 trials. To this end, Lipinski and collaborators [10] produced a set of rules that attempted to
106 identify the best statistical physico-chemical properties required for an oral compound to

107 achieve maximum bioavailability, i.e. to cross all biological barriers (where drug transporters
108 are present) before reaching its target. The first of Lipinski's rules is based on the lipophilic
109 index of the drug, the second on the drug's molecular weight (abbreviated "MW" in the
110 remaining text) and the third and fourth rules concern the drug's electrostatic charge properties.
111 These rules are now established drug discovery paradigms and have been largely embraced by
112 the pharmaceutical industry. Of the four rules, the second (MW<500) stands out by way of its
113 apparent simplicity, being unrelated to complex physico-chemical properties of a drug (as is
114 the charge or lipophilic index) but governed solely by a drug's size or volume. This simplicity
115 infers that basic mechanics apply when drugs cross membranes, cells, tissues and biological
116 barriers.

117 What is worth considering are the following points: (i) The Big Pharma did not focused on
118 drug transporters and Linpinski's rules do not mention drug transporter expression levels when
119 barriers to drugs are considered and; (ii) the drug volume and thus some mechanical properties
120 needs to be considered when drugs cross complex biological barriers.

121 The next question is why and how biophysics is involved in drug efficacy?

122

123 **Why is the drug MW so important to cross barriers? An introduction to the biophysics**
124 **of drug-membrane interactions.**

125 To be (bio)available, drugs must traverse cellular barriers – usually the epithelium or
126 endothelium (e.g. of the gastrointestinal tract, renal tubules or the blood-brain and blood-
127 placenta barriers). To traverse cellular barriers, drugs must cross lipid membranes, and for this
128 Lipinski's 2nd rule postulates that drugs must have a MW<500. Therefore, in the sum of
129 energies making up the total activation energy required for a drug to cross cellular membranes
130 a term must exist to underline the role of the membrane. In this case, i.e. when the plasma
131 membrane is considered as a flat object, the physical parameter that best fits such an interaction

132 is the membrane leaflets' surface tension (σ and unit $[\sigma] = N/m$)¹. Of course the surface
133 tension parameter needs a proper definition especially in cells. All lipids are amphipathic
134 molecules and as a result optimize their individual surface area in membrane leaflet. This
135 optimization results from the energy balance between steric and/or electrostatic repulsion(s)
136 (related to lipids' head) and the lipid contact with water (related to the hydrophobic aliphatic
137 chain(s)). This balance defines the surface tension. Now, when a drug enters a membrane leaflet
138 it will have to "squeeze in" and compress the lipids of the leaflet, namely change the surface
139 tension. This impact on the energy balance of lipids composing the leaflet will have a tendency
140 to repulse, i.e. push out, the drug from the leaflet. However this process is not totally rigid as
141 otherwise chemicals would never cross membranes. In fact, lipids are not static as the thermal
142 agitation exists which allows for some flexibility. So if a small enough chemical incorporates
143 into the leaflet and perturbs it in such a way that the resulting membrane energy is below the
144 ambient thermal energy, then the lipids composing the leaflets will not "feel" any difference
145 between the thermal agitation and the incorporation of the drug. So a drug can incorporate a
146 membrane leaflet if it is small enough.

147 Dimensionally speaking, it follows that a critical cross section for the drug (a_c) can be defined
148 simply by: $a_c \sim k_B T / \sigma$, where $k_B T$ is the thermal energy (k_B is Boltzmann's constant and
149 T the absolute temperature). If the cross section of a drug is lower than the critical value it will
150 incorporate and cross the membrane leaflet, but if it is higher the drug will be blocked.

151 In bilayer membranes, two types of membrane surface tension can be distinguished, the mean
152 surface tension noted σ_0 , which corresponds to the sum of the individual leaflet's surface

¹ Thermodynamically speaking, the physical parameters that are related to spatial dimensions (namely, volume (V), cross section area (a) or line (r)) are the pressure "P": $\delta E = -P \cdot \delta V$, the surface tension " σ ": $\delta E = \sigma \cdot \delta a$, and the tension line " γ ": $\delta E = \gamma \cdot \delta r$. " δ " is the differential operator and "E" the energy. As far as a membrane is considered, it is the surface tension (and thus the cross section area of the drug) that best describes the mechanical (i.e. physical) interaction and is deduced by posing $\delta E \sim k_B T$.

153 tension, and the difference in surface tensions $\Delta\sigma$, which corresponds to the difference
 154 between individual leaflet's surface tension. Using optical techniques, M. Sheetz and his
 155 collaborators have demonstrated that cells have a large reservoir of membrane [11] and an
 156 average membrane tension that is remarkably low ($\sigma_0 \sim 0.003mN/m$) [12]. On the other hand,
 157 the difference in surface tensions between leaflets has been demonstrated to be much higher
 158 $\Delta\sigma \sim 0.9mN/m$ [13]. Accordingly, and given the magnitude of this parameter, it is more
 159 likely to be involved in impairing the transverse movement of chemicals. The previous
 160 equation can thus be refined as follows: $a_c \sim k_B T / \Delta\sigma$. Dealing with a parameter as $\Delta\sigma$ is
 161 not intuitive and the last equation needs to be resolved physiologically. A fundamental aspect
 162 of the difference in surface tension corresponds to its role in pinocytosis associated with the
 163 role of specific lipid flippases maintaining the membrane lipid asymmetry [14]. A direct
 164 consequence associated with this asymmetry is a more highly packed inner leaflet as it contains
 165 more phospholipids than the outer leaflet resulting in the difference in surface tensions (
 166 $\Delta\sigma = \sigma_{out} - \sigma_{int} \sim 0.9mN/m$) between the inner (cytosolic) and outer leaflets of the cell
 167 plasma membrane. Naturally, bilayer membranes are soft objects and as such, will attempt to
 168 release this stored energy. Accordingly, it has been demonstrated that lipid asymmetry
 169 corresponds to the physiological motor force that triggers membrane budding, leading to
 170 endocytosis (Figure 1) [13, 15, 16]. It is therefore possible to demonstrate that the vesicle radius
 171 is written as [13]: $R = 8k_c / h\Delta\sigma$; where k_c is the cell membrane bending modulus and h the
 172 membrane thickness. As for drugs small enough that their MW is proportional to their Van der
 173 Waals' volume (expressed in \AA^3), i.e. $MW \sim V \sim a^{3/2}$, a critical MW (MW_c) can be
 174 determined given by:

$$175 \quad MW_c = (4/3\sqrt{\pi})(hRk_B T / 8k_c)^{3/2} \quad (\text{Eq.1})$$

176 The later relation provides a law with regard to the drugs size (or MW) selectivity on their
177 permeation across cellular membranes: $MW_c \cong 240 - 250$ at 37°C [17]. As the MW cut off
178 defined by Lipinski's 2nd rule, i.e. $MW_c = 500$, describes the 90th percentile; the former value
179 (i.e. $MW_c \cong 240 - 250$) is an average in line with Lipinski's rule. Two other important results
180 follow. The first one is that it is also possible to demonstrate that the kinetics of membrane
181 endocytosis is inversely proportional to the vesicle radius [18], i.e.:

$$182 \quad k_{endo} \sim 1/R \quad (\text{Eq.2})$$

183 And that the kinetics of transverse movement across the membrane is [17]:

$$184 \quad k_{Drug} \sim \exp\left(A \times MW^{\frac{2}{3}} \times k_{Endo}\right) \quad (\text{Eq.3})$$

185 Where, A, is a constant. It does seem that Lipinski's 2nd rule can be explained by considering
186 simple biophysical arguments and that the membrane plays a key role in this process. But what
187 about drug resistant cancer cells?

188

189 **Are alterations in the cell membrane observed in MDR and is the drug MW important in**
190 **multi drug resistant cells?**

191 From what was seen above, if the drug MW is important it is because the membrane is also
192 involved. So changes in the lipid membrane composition and membrane recycling should be
193 expected in drug resistant cells and this seems to clearly be the case. Different studies have
194 reported changes in membrane composition including neutral lipids, phospholipids, cholesterol
195 and fatty acids [19-24], in some cases related to a change in the lipid metabolism of drug
196 resistant cells [22, 25, 26]. This point has been particularly well underlined when the lipid
197 profile of released exosomes was analysed [27]. Also, ultrastructure studies have revealed an
198 increased density of small and large membrane organelles [22, 28-32] and an increase in the
199 kinetics of membrane endocytosis or membrane recycling [29-31, 33, 34] in drug resistant

200 cells. It is noteworthy that the release of exosomes is also involved in MDR [35]. What is
201 perhaps more important is that the MW of a drug itself was also underlined very early (in 1970)
202 in MDR studies in line with the role that the membrane has in delaying a chemical's influx [36,
203 37]. It is worth noting here that the role of a drug's MW was underlined prior to the discovery
204 of Pgp by Juliano and Ling in 1976 [5]. The connection between membrane endocytosis and
205 the size of a drug chemical with passive influx/uptake of drugs into cells is given by the set of
206 equations described above.

207 The data points clearly to the membrane as a strong effector of drug resistance but why would
208 the membrane be so central when drug transporting is involved in MDR?

209

210 **Drug-membrane biophysical interactions to resolve the multi specificity of drug** 211 **transporters**

212 It is very often suggested that drug transporters work similarly to enzymes in line with the
213 notion of affinity, namely that a drug needs to interact with a transporter to activate the
214 transporter and be expelled. However this view does not work for at least three reasons when
215 focusing on Pgp: (i) the ATP concentration in cells is usually 3-5mM that always exceeds the
216 affinity of Pgp for ATP ($K_{mATP} \sim 0.3-1\text{mM}$) [38, 39], suggesting that the transporter is always
217 "active". (ii) Pgp ATPase activity is relatively independent of the presence of drugs [40], and
218 the affinity of drugs toward transporters is chiefly dependent on their affinity toward the
219 membrane [41]. Finally (iii), the apparent stoichiometry of the hypothesized ATP-coupled
220 active drug transport, i.e. the number of ATP molecules hydrolyzed per drug transported, can
221 be enormous (calculated to be up to $\sim 36000\text{ATP}/\text{drug}$ in reconstituted proteo-liposomes) [6,
222 38]. This suggests that while consuming ATP Pgp does not necessarily lead to drug extrusion.
223 Due to the fact that similar conclusions cannot be drawn for drug transporters other than Pgp
224 due to lack of experimental observations, Pgp remains the archetypal transporter involved in

225 MDR and it is believed that Pgp is very likely continuously recycling between “open” and
226 “closed” states by over-consuming ATP. This may explain why Pgp and drug resistance are so
227 sensitive to cellular metabolism [42]. It is interesting to note that Pgp activity leads to a parallel
228 acidification of the extracellular medium [43] that, in turn, is thought to be related to initial
229 metastatic steps [44]. Given that the vast majority of metastatic tumours are also multi drug
230 resistant [45], the recycling between open and closed conformations is likely to be essential to
231 explain the *multi* of drug resistance [46].

232 Here comes an essential point. If Pgp switches between open and closed conformations
233 independently of drugs, what is essential in MDR is that for drugs to be expelled they must
234 remain in the membrane long enough to encounter (or collide with) Pgp. From Eq.3 the kinetics
235 of drug transverse movement is modulated exponentially by two physical parameters related
236 to the biophysical state of the membrane involving the size of the drug (see above) and the
237 kinetics of endocytosis (see below). An increase in the kinetics of membrane endocytosis
238 supporting Pgp function is possible if the Warburg effect and relatively high cytosolic pH are
239 considered.

240

241 **Cytosolic pH, endocytosis and MDR**

242 Regardless of their origin and genetic background cancer cells and tissues have been found to
243 display an abnormality called “proton reversal” which describes the state by which a cell
244 consists of an interstitial acidic microenvironment secondary to an initial, specific and
245 etiopathogenic intracellular alkalosis [47-53]. A failure to induce intracellular acidification and
246 reverse this phenomenon in cancer tissues has been proposed to be the main factor underlying
247 drug resistance including resistance to the induction of therapeutic apoptosis [54-58]. Also,
248 because inner leaflet lipids bear protonable polar heads, pH changes will modify their net
249 charge. In turn this will impact on the sum of electrostatic repulsions and modify membrane

250 difference in surface tension (i.e. decrease the size of pinocytic vesicles and as a result increase
251 the kinetics of endocytosis) [59].

252 To consider any effect of the cytosolic pH on lipid packing it is central to understand the notion
253 of packing from a physics standpoint. At a constant membrane surface area, the lipid packing
254 is given by the optimal area per lipid in the cell membrane. The latter is deduced from the
255 balance between repulsions that occur mostly through electrostatic effects on the polar heads,
256 and attractions, which concern more the hydrophobic and geometric effects that take place
257 between the aliphatic chain(s). Any changes in this balance are expected to affect the optimal
258 area per lipid (i.e. their packing) and membrane shape. As a non-negligible fraction of the inner
259 leaflet consists of negatively charged lipids, such as phosphatidylserine or PIP₂, for example
260 [60] a slight increase in proton concentration around neutrality (e.g. decrease in cytosolic pH)
261 will eliminate or shield these negative charges and decrease the electrostatic repulsion between
262 polar groups. Although such an electrostatic counterion effect might in principle be generalized
263 to intracellular cations, it is obvious that exchangeable protons will have a more pronounced
264 effect on negatively charged lipids. As a final result, a low cytosolic pH is more likely to be
265 central in abolishing the physical repulsion between lipids, and thus decreases the surface
266 tension (i.e. the lipid packing of the cytosolic leaflet - note that both lipid packing and surface
267 tension are proportional to each other). Such a relationship between free electrolytes and the
268 cross section area per lipid in model biomembranes is well known experimentally [61-63]. A
269 similar result was also obtained on living cells [64]. Conversely, when the cytosolic pH
270 increases (i.e. when cells become reliant on the Warburg effect), fewer positive charges will be
271 available to mask the lipids charge, which in turn is expected to increase their repulsions and
272 thus their packing. Thus, this higher lipid packing would increase the surface tension of the
273 leaflet in contact with the milieu of elevated cellular pH in the case of drug resistant cells. So,
274 if the pH affects the packing of lipids, and the packing of lipids affects the intracellular

275 accumulation of drugs, it follows that the cytosolic pH should affect the intracellular
276 accumulation of those drugs. As a result, the changes in cytosolic pH observed when cells
277 switch their state of resistance is an important clue for understanding the observed alterations
278 of intracellular accumulation of drugs as a function of their size. This way of thinking has
279 permitted the theoretical corroboration of the connection between the cytosolic pH (linked to
280 Warburg effect), the membrane biophysical properties and the MDR levels in several cell types
281 [59] (see figure 2). The interaction between the membrane and the cytosolic pH can explain
282 why PPIs overcome the Pgp-mediated MDR [65].

283

284 **Beyond the cell membrane**

285 Using arguments and results developed by us and others the general view is that drug sensitivity
286 or drug resistance can only be understood if one steps outside of a Pgp-centred view to engage
287 with a holistic approach of cancer. This true and fundamental scientific approach is equivalent
288 of saying that what has been exposed in this review needs to be duly criticized as well to push
289 the boundary that it creates under the form of a new research field. In the context of drug
290 sensitivity (or drug resistance or drug refractoriness) in cancer it is essential to underline the
291 fact that many interactions between the various cellular compartments exist that underlines the
292 complexity of the disease that, in turn, may provide fundamental clues as to how MDR
293 progresses. An illuminating study performed in resinless ultrathin EM sections has shown that
294 a staggering network of interconnected cytoskeletal filaments does exist between
295 polyribosomes, mitochondria and a myriad of unidentified small structures attached to the
296 cytoskeleton [66]. Using the same technique, the nuclear space appears as a complex network
297 of core filaments connecting with the nuclear lamina, and the chromosomes appear attached to
298 spindle fibers, which are in turn interconnected through several thin filaments. None of these
299 structures are visible using conventional resin embedding technique. This introduces the

300 concept that the cell has to be considered as a whole, and that this whole is not entirely known
301 also because of the compartmentalization of the research approaches; and this is true for MDR
302 as well. In general the membrane to cytoskeleton connection is entirely deranged in cancer
303 cells, determining an aberrant cell polarization in turn related to the metastatic behaviour [67].
304 Research has been carried out showing that Pgp is linked to actin through ERM and that this
305 connection is key for MDR in human tumor cells [68, 69]. How such interaction can be
306 understood in the framework provided by the membrane is unclear but it underlines that fact
307 that cells should be considered in a holistic way, also because cancer cells are independent and
308 behave as an unicellular microorganism committed to survive in a very hostile environment
309 [70].

310

311 **Conclusion: From bench to bedside**

312 While MDR remains linked to drug transporters, alterations in pH gradient resulting from the
313 Warburg effect across the cell membrane or organelles is well known to impact on the
314 biophysical properties of the cancer cell membrane sustaining drug transporter activity.
315 Therefore it is in theory possible to improve drug uptake by cells by normalizing the pH using
316 PPIs. This point was demonstrated recently in tumor spheroids [71]. Furthermore the same
317 study demonstrated that tumor spheroids were becoming more sensitive to lower drug doses
318 of anticancer agents raising hope that adverse effects linked to the administration of
319 chemotherapy could, one day, be reduced or controlled in patients [71]. PPIs are amongst the
320 most commonly prescribed drugs in human medicine and have gone through the process of
321 rigorous safety testing and monitoring. Very few clinical side effects are seen even at higher
322 doses and as such it seems easy to justify the continued investigation into the use of this class
323 of drug for the treatment of cancer in companion animals [71-74] and humans [75-78]. They
324 may provide an alternative or additional source of therapy to animals and humans which could

325 result in lower treatment costs, greater availability and safer handling compared to current
326 cytotoxic protocols. PPIs and PTIs could potentially form part of a universal treatment which
327 may have direct benefits in treating a number of different cancer types while combating
328 problems associated with chemotherapy such as drug resistance, severe side-effects and even
329 death secondary to present day chemotherapy.

330

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553 Legends

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555 Figure 1: (A) Lipid asymmetry at the vesicular scale: Given the small size of vesicles, the radius
556 and membrane thickness are relatively close together ($R/h \sim 10$). Thus, the outer leaflet of a
557 vesicle (S_{out}) has significantly more lipids than the inner leaflet (S_{in}). As the vesicle is
558 spherical, noting $S_0 = 4\pi R^2$ the neutral surface area namely the surface area between the outer
559 and inner leaflets, it follows at the first order that $S_{out} = 4\pi(R+h/2)^2 \sim S_0(1+h/R)$ and
560 $S_{in} = 4\pi(R-h/2)^2 \sim S_0(1-h/R)$. Thus $S_{out} - S_{in} \sim S_0 \cdot h/R$. (B) Sketch representing the
561 current model linking fluid phase endocytosis to the membrane phospholipid number
562 asymmetry [14]. In the left panel, the translocation of dark-headed lipids into the inner leaflet
563 induces a differential packing of lipids between leaflets leading to membrane bending and
564 vesiculation [13, 15]. Note the membrane recycling that occurs in cells (right panel), i.e. the
565 exocytosis of vesicles with a size similar to endocytic vesicles, allows the maintenance of lipid
566 asymmetry and thus the maintenance of the differential packing of leaflets at the level of the
567 plasmalemma. Accordingly, the lipid number asymmetry has been experimentally deduced
568 from studies on drug sensitive cells (K562) with a value $\Delta N/N_0 = 4\%$ providing a $\sim 35\text{nm}$
569 vesicle radius [13]. (C) Representation of the different energy barriers (noted together $U(x)$)
570 and involved when a drug traverses the bilayer cellular membrane. Two leaflets have been
571 represented with an inner leaflet containing more phospholipids related to the increase in the
572 difference in surface tensions (upper graph). Energy profiles of lipid packing in both leaflet
573 (plain curve-middle graph) and hydrophobic core of membrane (dashed curve-middle graph)
574 are both involved in providing penalty energies with regard to the transbilayer movement of
575 drugs. As the inner leaflet is packed, drugs crossing the membrane will be trapped in this leaflet
576 which will delay and impair their flow into the cytosol [79]. The latter effect will be dependent

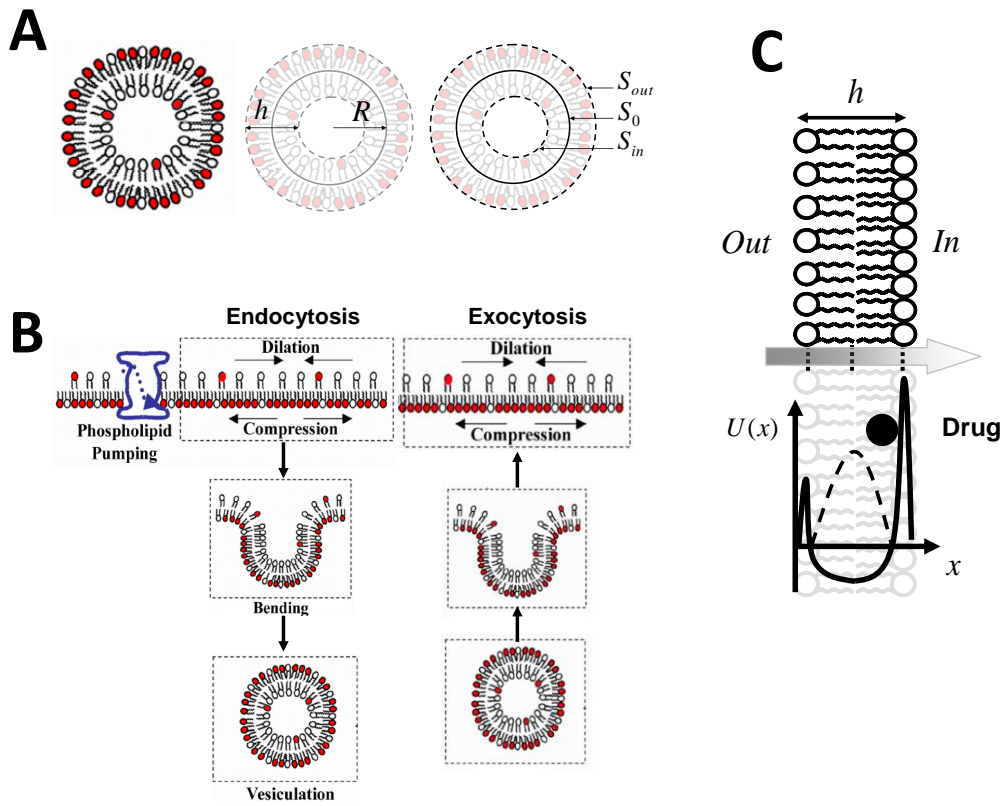
577 on the size of drugs as bigger drugs will “feel more strongly” this mechanical barrier. In the
578 present paper, this effect is supposed to be central for the high levels of cross resistance to
579 drugs.

580

581 Figure 2: (A) Comparison between experimentally measured doxorubicin resistance levels
582 obtained in cells (blanked circles) and the theory (filled circles). The open circles
583 corresponding to SW1573 (lung derived cancer cells), K562R (leukemic cancer cells) and
584 MCF-7R (breast derived cancer cells) are indicated with arrows and labels. Finally the straight
585 line is the linear regression of experimental data which agrees very well with the theory.

586 Figure 1

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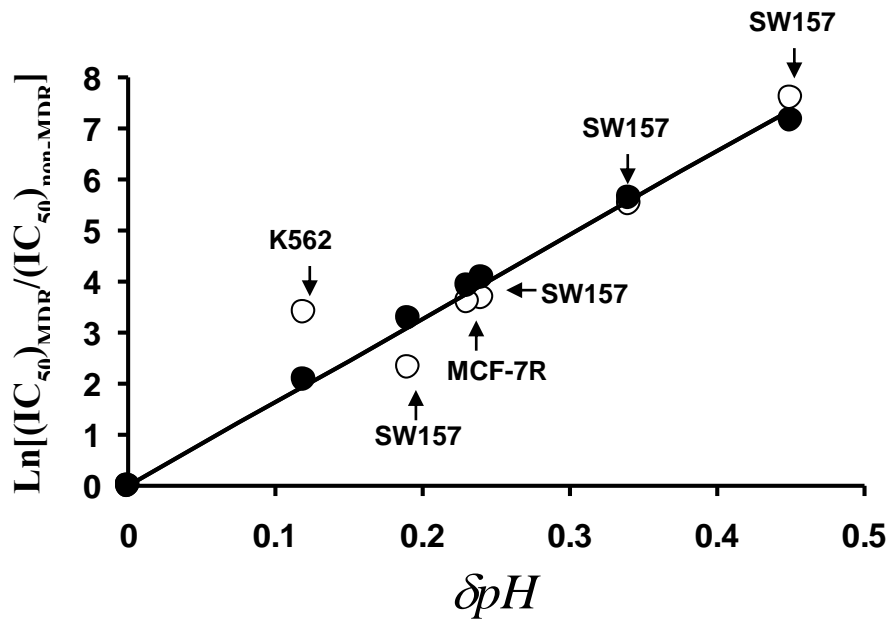


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590 Figure 2

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