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1 **Reproductive biology of *Holepyris sylvanidis* (Hymenoptera: Bethyridae)**

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24 **ABSTRACT**

25 *Holepyris sylvanidis* (Hymenoptera: Bethyridae) is reported as a cosmopolitan parasitoid of
26 coleopteran including a major pest of stored products, the confused flour beetle *Tribolium*
27 *confusum*. The reproductive biology, and hence biocontrol potential of *H. sylvanidis* is
28 relatively little known. Here we describe and evaluate aspects of egg size, immature
29 development, adult mating behaviour, pre-oviposition time, fecundity, host attack and use,
30 and adult longevity. Our key findings are that $\approx 72\%$ of presented *T. confusum* immatures
31 presented to females were attacked and $\approx 39\%$ of these were fed on destructively with the
32 remainder oviposited onto. First instar host larvae and host pupae were only utilized for
33 feeding, second and third instar larvae were commonly used for either feeding or oviposition
34 while fourth, fifth and sixth instar hosts were almost exclusively used for oviposition. Females
35 usually laid single egg clutches and their average lifetime fecundity was ≈ 16.5 eggs. Females
36 regularly provisioned with hosts often ceased laying eggs up to several weeks before their
37 death but continued to kill and feed on hosts during this period. Adult longevity was greatly
38 enhanced by the provision of hosts (females) or by non-host food sources (males). We
39 conclude that the inoculative biocontrol potential of *H. sylvanidis* is likely to be limited by a
40 low realized fecundity and the combination with other control methods in an IPM-approach is
41 needed. Its ability to find and kill hosts may make it more suitable for inundative deployment.

42

43

44

45 **Keywords:** Stored product, *Tribolium confusum*, *Holepyris sylvanidis*, mating, fecundity,
46 longevity.

47

48 **1. Introduction**

49 There are over two thousand described species of wasps that belong to more than 100
50 genera within the hymenopteran family Bethyridae (Gordh and Móczár, 1990). Typically
51 these are parasitoids of coleopteran and lepidopteran larvae (Evans, 1978; Mayhew and
52 Hardy, 1998), although a minority of bethylid species attack hosts in the pupal stage, and
53 some can develop on hosts belonging to other insect orders (Pérez-Lachaud et al., 2004;
54 Zhang et al., 1984). Bethyrid species have been used in empirical investigations of factors
55 thought to influence the evolution of key behavioural and reproductive traits, such as clutch
56 size, sex allocation, aggression and sociality (Green et al., 1982; Griffiths and Godfray, 1988;
57 Hardy et al., 2013; Hu et al., 2012; Khidr et al., 2013; Lizé et al., 2012; Mayhew and Godfray,
58 1997; Mayhew and Hardy, 1998; Morgan and Cook, 1994; Suma et al., 2014; Tang et al.,
59 2014). As the hosts of many bethylid species are pests of field-crops, stored products and in
60 agro-forestry, bethylids have also been deployed as biological control agents in a wide range
61 of agro-ecosystems (Conlong and Graham, 1998; Damon and Valle, 2002; Gothilf and Mazor,
62 1987; Legner and Gordh, 1992; Jiang et al., 2015; Legner and Silveira-Guido, 1983; Pérez-
63 Lachaud et al., 2004; Sreenivas and Hardy, 2016; Venkatesan et al., 2007; Yang et al., 2014).
64 Most basic and applied studies of bethylids have, however, focused on species in a small sub-
65 set of genera, such as *Sclerodermus* Latreille and *Cephalonomia* Westwood in the sub-family
66 Epyrinae and *Goniozus* Förster in the sub-family Bethylinae. Here we contribute to
67 knowledge of the reproductive biology of *Holeptryris sylvanidis* (Brèthes) (= *Pareptryris*
68 *sylvanidis* Brèthes, *Rhabdepyris zae* Turner and Waterston); this a little studied species in a
69 relatively unstudied epyrine genus and a beneficial natural enemy of stored products pests.

70 *Holeptryris sylvanidis* is reported as a cosmopolitan parasitoid of coleopterans in the
71 families Cucujidae, Curculionidae and Tenebrionidae (Abdella et al., 1985; Gahan, 1930;
72 Evans, 1978; Gordh and Móczár, 1990; Eliopoulos et al., 2002; O'Connor and Ronayne,

73 2002). Among the Tenebrionidae the preferred host is the confused flour beetle *Tribolium*
74 *confusum* Jacquelin du Val (Ahmed and Islam, 1988; Hagstrum et al., 2012; Lorenz et al.,
75 2010), which feeds on many kinds of flour, grains, cereal products, dry fruits and nuts (Park,
76 1934; Sutton et al., 2011). *Tribolium confusum* infests in flour mills, feed mills, warehouses,
77 boxcars, semolina mills peanut shelling plants, bakeries, grocery stores, pet stores and human
78 residences (Eliopoulos et al., 2002; Hagstrum et al., 2012; Hagstrum and Subramanyam,
79 2009; Prozell and Schöller, 1998), particularly in areas with Mediterranean climate
80 (Trematerra et al., 2007). Contamination by *T. confusum* of flour mills, and other facilities,
81 can have significant economic consequences, including costs associated with treatment and
82 monitoring, rejection and return of contaminated products and loss of consumers' trust
83 (Campbell and Hagstrum, 2004). Application of synthetic pesticides may reduce *T. confusum*
84 populations but may have undesirable side effects, and is not applicable in organic bakeries
85 and mills (Lorenz et al., 2010; Prozell and Schöller, 1998). Knowledge of *T. confusum* natural
86 enemies could thus lead to economic benefits.

87 Previous studies of the biology of *H. sylvanidis* have described its mating behaviour,
88 foraging behaviour and host attack (Ahmed et al., 1997; Ahmed and Islam, 1988; Fürstenau et
89 al., 2016; Lorenz et al., 2010), while other studies have recorded its associations with pest
90 species and other natural enemy species in stored-product agroecosystems (Adler et al., 2012;
91 Eliopoulos et al., 2002). Foraging *H. sylvanidis* females are attracted by odours emanating
92 from the faeces of *T. confusum* larvae (Fürstenau et al., 2016). Lorenz et al. (2010) reported
93 that females are able to penetrate 8 cm into the flour substrate to find *T. confusum*, but that the
94 success in finding hosts declined with depth and also as particle size decreased. Hosts that are
95 attacked are typically transported to a crevice or hole prior to oviposition (Ahmed et al., 1997;
96 Lorenz et al., 2010). *Holepyris sylvanidis* females attack and feed on *T. confusum* larvae in the
97 early instars and oviposit on larger larvae, normally laying a single egg onto each hosts, but 2
98 and 3 eggs clutches are occasionally observed (Ahmed et al., 1997). Notwithstanding these

99 prior studies, obtaining empirical information on *H. sylvanidis* has been hampered by
100 difficulties associated with collecting and then rearing this species under laboratory
101 conditions. In this paper we report a viable laboratory-rearing protocol for *H. sylvanidis* along
102 with observations on its behaviour and reproduction. Specifically, we describe and evaluate
103 aspects of egg size, immature development, adult mating behaviour, pre-oviposition time,
104 fecundity, host attack and use, and adult longevity. We discuss these new findings in regard to
105 both the general biology of bethylid wasps and the potential for *H. sylvanidis* to suppress
106 populations of pests.

107

108 2. Materials and methods

109 2.1 Host and parasitoid cultures

110 The host *T. confusum*, was reared in 12 cm diameter Petri dishes containing 20 g of
111 *Triticum aestivum* flour and 5% of brewers' yeast as a substrate (Athanasidou and
112 Kavallieratos, 2014). The *H. sylvanidis* used were a strain collected from wheat infested by *T.*
113 *confusum* in a grain centre warehouse at Enna (Italy). Parasitoids were reared by keeping two
114 female and one male *H. sylvanidis* in a 12 cm diameter Petri dish with fourth instar host
115 larvae (Ahmed et al., 1997), with fresh fourth instar larvae added each week. Two squares (2
116 cm × 2 cm) of corrugated cardboard were placed into each Petri dish to provide places for
117 female parasitoids to hide paralysed hosts (Ahmed and Islam, 1988). Adult *H. sylvanidis* host
118 feed (Ahmed et al., 1997) but, in order to enhance egg production, were provided with a
119 supplementary diet composed of water (1 ml), honey (0.1 g), pollen (7 grains) and royal jelly
120 (0.05 g) absorbed onto a 1×1cm piece of filter paper. All cultures and experiments were
121 maintained at 27±1°C and 65±5 % relative humidity.

122

123 2.2 Life history evaluations

124 We evaluated aspects of egg size, immature development, adult mating behaviour, pre-
125 oviposition time, fecundity, host use and adult longevity.

126

127 *2.3 Development of offspring*

128 Parasitoid cultures were inspected daily and any fourth instar host larvae bearing a *H.*
129 *sylvanidis* egg were collected kept individually in 4 cm diameter Petri dishes. The length and
130 width of each egg was measured under a binocular dissecting microscope with image analysis
131 software. The development of each *H. sylvanidis* offspring was observed daily until adult
132 eclosion. Replicates in which the egg did not mature to adulthood were excluded. We
133 recorded the number of days before the egg hatched (when head of the larva emerged from
134 the chorion, according to Powel 1938), the durations of the larval and pupal stages and the sex
135 of the emerging adult. We also recorded the time between each egg hatching and the
136 penetration of the body of the host by the larva, the time taken for the host to be consumed
137 and the time it took for larvae to spin a silken cocoon and pupate.

138

139 *2.4 Mating behaviour*

140 In each replicate, one male and one female *H. sylvanidis*, with no prior mating experience
141 and within one day of eclosion as adults, were placed into a 4 cm diameter Petri dish and
142 observed. In order to ensure that the adults were unmated, their cocoons had been isolated
143 from the cultures for at least 4 days before adult emergence. Courtship and mating
144 interactions were observed and the time until mating (pre-copulatory time) and the duration of
145 copulation, from when the male mounted the female until the male dismounted, was recorded.
146 After each replication, the Petri dish was cleaned with ethanol to remove any chemical
147 stimuli.

148 2.5 Pre-oviposition period

149 In each replicate, a newly emerged (<1 day old) and unmated female was placed into a 12
150 cm diameter Petri dish containing a square of corrugated cardboard and 5 fourth instar *T.*
151 *confusum* host larvae, which are the preferred host developmental stage (Ahmed et al., 1997).
152 In some replicates a newly emerged male *H. sylvanidis* was also present, leading to the female
153 being mated. The hosts were observed daily until the first *H. sylvanidis* egg was observed.
154 Host larvae that were fed upon or had died and were no longer suitable for oviposition were
155 replaced at each observation. The pre-oviposition period was calculated as the number of days
156 between the emergence of the female and the first egg laid (Finlayson, 1950). There were 50
157 replicates using unmated and 50 using mated females.

158

159 2.6 Host attack and use

160 In each replicate ($n = 10$) one ≤ 5 day-old female was placed into a 12 cm diameter Petri
161 dish containing 35 hosts, comprising 5 hosts of each of the 6 larval instars and 5 pupae. Host
162 attack behaviour was observed. Each replicate was inspected daily for 10 days noting the
163 number of hosts that had been fed upon or paralysed by the female. We note that it is difficult
164 to detect the paralysis of host pupae but feeding damage was straightforward to observe.

165

166 2.7 Lifetime fecundity

167 In each replicate, a newly emerged female was placed into a 12 cm diameter Petri dish
168 containing a square of corrugated cardboard and supplied with fourth instar host larvae twice
169 per week: on Mondays, 5 fresh hosts were added to the Petri dish and on Fridays all larvae
170 were removed and 10 fresh hosts provided. Hosts with eggs laid on them were collected on
171 both Mondays and Fridays. The procedure was repeated until each female died. In some

172 replicates a newly emerged male *H. sylvanidis* was also present from the start such that the
173 female was mated.

174

175 2.8 Longevity

176 We evaluated the longevity of females ($n = 92$), with and without the provision of hosts
177 and of males, with and without the provision of food (while adult males may not interact with
178 hosts they could potentially feed on nectar and pollen). Mated females (<1 day old) were
179 placed individually into plastic vials (2.6 cm \times 0.6 cm) stoppered with cotton gauze. In some
180 replicates, 5 fresh host larvae were provided twice per week and any previously provided
181 hosts removed. In other replicates the females were starved. Male longevity was similarly
182 evaluated ($n = 144$); males were either starved or provided with food on the gauze stopper
183 twice per week. Some fed males received sucrose solution (0.5 g per 2.5 ml water) and others
184 were provided with sucrose solution with 4 grains of pollen added. For both male and
185 females, longevity was evaluated by inspecting the vial each day until the wasp died.

186

187 3. Statistical analysis

188 Where possible data were explored using generalized linear modelling (log-linear
189 modelling, and parametric survival analysis; Crawley, 1993) but non-parametric Mann-
190 Whitney U tests were employed when error distributions did not conform well to parametric
191 assumptions. All tests were two-tailed and used the 0.05 significance level. Data were
192 analysed using GenStat (version 17, VSN International, Hemel Hempsted) and StatPlus
193 (version 6.0.3, AnalystSoft Inc.).

194

195 4. Results

196 4.1 Development of offspring

197 Eggs were on average $0.32 (\pm 0.003 \text{ SE})$ mm long and 0.13 ± 0.010 mm wide ($n = 50$). Eggs
198 typically hatched after 2 days and there was no significant difference in time to hatching
199 between eggs that eventually developed into males and eggs that developed into females
200 (Table 1). Hatched larvae inserted the mouthparts into the host body to feed and initially fed
201 on the host completely ectoparasitically but after around 3.5 days each larva penetrated the
202 host such that its anterior half was embedded into the host. Each larva fed on its host until
203 only the empty exoskeleton remained, around 2-4 days after penetrating the host. For both
204 sexes, the larval stage typically lasted nearly 9 days (Table 1). It was not possible to document
205 the number of larval instars because no exuviae were observed (as in other epyrines,
206 Finlayson, 1950; Mertins, 1980). Once hosts were consumed, each larva spun a white cocoon
207 over a 1-2 day period before pupating. In some cases a proper cocoon was not formed but
208 pupae that were not fully enclosed in cocoons were able to develop to adulthood. *H.*
209 *sylvanidis* pupae are initially white but become dark. The large compound eyes start to
210 become brownish by the end of the first day of pupation and become darkly pigmented by the
211 second day. The thorax becomes dark before the abdomen and the head. The intersegmental
212 membrane of the abdomen is the last part to change colour, becoming completely dark one
213 day after the adult wasp emerged. The pupal stage of males was more than a day shorter than
214 females (protandry) (Table 1) and on emergence a typical meconium (Rilett, 1949) was
215 observed. Adult males are smaller than adult females.

216
217 4.2 Mating behaviour

218 We identified multiple components of courtship and mating behaviour (Table 2) and the
219 sequence in which they typically occurred (Fig. 2). Courtship started when the male verified
220 the opportunity to mate by approaching the female and touching her with his antennae.

221 Unreceptive females moved away from the male rapidly but receptive females vibrated their
222 antennae then turned their bodies by 180°. Then the wasps touched antennae from one to
223 seven times, in separate bouts, prior to mating. Males then suddenly jumped onto the female
224 and copulation occurs. On occasion, when a female was receptive but the male did not initiate
225 courtship, the female touched the male's abdomen from behind with the antennae, apparently
226 to stimulate the male to enter courtship.

227 Copulation began after a mean courtship period of 208.6 ± 30.2 seconds ($n = 30$) and the
228 mean duration of copulation was 119.5 ± 8.2 s. During copulation the male gripped the female
229 with his body at 90° to hers, and the antennae of both sexes were motionless. Usually the
230 female did not move during copulation but in some instances she ran around. Male assumed
231 an elevated position and displayed rhythmic movements of their legs during copulation, but
232 only when the female was stationary. At the end of the copulation, the males dismounted from
233 the female.

234

235 4.3 Pre-oviposition period

236 Mated females began laying eggs after 3.68 ± 0.28 days. Unmated females had a
237 significantly longer pre-oviposition period (5.36 ± 0.51 days; Mann-Whitney U-test, $U =$
238 927.5 , $P = 0.024$, Fig. 2).

239

240 4.4 Host attack and use

241 When a female *H. sylvanidis* encountered a potential host she inspected it by touching its
242 surface with her labial and maxillary palps. On recognizing a larva as a suitable host, the
243 female immediately attacked it by grasping it with her legs and bending her abdomen around
244 the larva and rotating her body by 360 degrees for three or four times. The attack is very rapid
245 and it is difficult to observe exactly when the female stings the host with venom. The host was

246 typically paralysed immediately and permanently (on occasion the host larva later regained
247 activity and began to crawl, with a parasitoid egg adhering to its venter). Females continued to
248 touch the host with their antennae for some seconds after stinging. While paralysed hosts were
249 almost motionless they showed rhythmic movements of their legs. Sometimes the female
250 attacked the legs of the host larva, biting the intersegmental membrane. Although each host
251 larva was entirely explored, females paid most attention to the segments close to the legs. The
252 females then typically grasped their host with their mandibles and moved it to another locality
253 prior to oviposition, but some hosts were instead fed upon.

254 Of all hosts presented, 72.3% were utilized for feeding or were parasitized by *H. sylvanidis*
255 females; of these, 39.1% were fed upon. Host acceptance and usage differed according to host
256 developmental stage. Host acceptance increased as larvae had developed until the fifth instar
257 and then declined (Fig. 4). In terms of usage, first instar larvae and pupae were only utilized
258 for feeding, second and third instar larvae were commonly used for either feeding or
259 oviposition while fourth, fifth and sixth instar hosts were almost exclusively use for
260 oviposition (Fig. 3). As far as we could discern, each host was either fed upon or oviposited
261 upon, but not both.

262 Ovipositing females usually laid a single egg onto the host, but clutches of two eggs were
263 occasionally laid. Eggs were normally placed longitudinally between the first and second
264 abdominal sternite, as figured in Reichmuth et al. (2007), but sometimes the egg was laid on
265 the second and third sternite, or between the third and fourth abdominal sternites.

266 267 4.5 Lifetime fecundity

268 Females laid on average 16.42 (+SE = 0.871, -SE = 0.828) eggs during their lifetimes ($n =$
269 68). There was no significant difference in the overall fecundity realized by mated and
270 unmated females (log-linear ANOVA corrected for overdispersion by assuming a quasi-

271 Poisson distribution of errors using an empirically estimated scale parameter: $F_{1,66} = 2.42$, $P =$
272 0.124 , deviance explained = 3.5%). Egg laying activity was initially higher among mated
273 females than unmated females and declined as females aged, with a more gradual decline
274 among unmated females (Fig. 4a). The risk of a female dying increased between successive
275 observation periods (fitting a Weibull model of cohort survival reduced the deviance
276 significantly compared to an exponential model, $G_1 = 91.1$, $P < 0.001$) and unmated females
277 lived significantly longer than mated females ($G_1 = 25.6$, $P < 0.001$, Fig. 4b). In many
278 replicates females ceased laying eggs before all of the provided hosts had been oviposited on
279 and several weeks before their time of death. The period of active reproduction was
280 significantly shorter among mated females than among unmated females ($G_1 = 14.98$,
281 $P < 0.001$) but we observed that females continued to kill and eat hosts after ceasing to
282 oviposit. The functions describing the models for mated and unmated females are
283 respectively: = initial number \times (Exp(-0.0001219 \times Observation period^{3.684})) and = initial
284 number \times (Exp(-0.00003438 \times Observation period^{3.684})).

285

286 4.6 Longevity

287 Female longevity was significantly greater when females were provided with fourth instar
288 *T. confusum* larvae (48.6 ± 2.7 days) then when they were not (8.1 ± 0.5 days) (Weibull
289 cohort survival model: $G_1 = 355.16$, $P < 0.001$): all unprovisioned females died within 20
290 days while females with hosts lived up to 86 days (Fig. 5a). Similarly, male longevity was
291 affected by feeding treatment ($G_2 = 83.40$, $P < 0.001$, Fig. 5b): when starved males lived
292 around 7 (± 0.2) days, males provided with sucrose lived on average for 18.18 (± 1.64) days
293 and the males nourished with the sucrose solution and pollen lived on average for 35.48 (\pm
294 4.97) days. Although the survival of males fed with sucrose only and sucrose and pollen was
295 initially similar (Fig. 5b), males fed with sucrose and pollen had significantly greater

296 longevities ($G_2 = 21.10$, $P < 0.001$). The functions describing the models for the longevity of
297 females and fed females are respectively: $= 100 \times (\text{Exp}(-(0,01574 \times (\text{Observation}$
298 $\text{period}^{1,928}))))$ and $= 100 \times (\text{Exp}(-(0,0005029 \times (\text{Observation period}^{1,928}))))$. The functions
299 describing the models for the longevity of males, males and sugar, males sugar and protein are
300 the following: $= 100 \times (\text{Exp}(-(0,02237 \times (\text{Observation period}^{1,928}))))$, $= 100 \times (\text{Exp}(-$
301 $(0,0027559 \times (\text{Observation period}^{1,928}))))$, $= 100 \times (\text{Exp}(-(0,00058318 \times (\text{Observation}$
302 $\text{period}^{1,928}))))$.

303

304 5. Discussion

305 *Holepyris sylvanidis* is a natural enemy of immature beetles, acting either as a destructive
306 host feeder or as a parasitoid of each suitable host encountered. As such its observed biology
307 conforms broadly to expectations derived from knowledge of other species within the bethylid
308 sub-family Epyrinae (e.g. Lauzière et al., 1999; Mayhew and Hardy, 1998; Pérez-Lachaud
309 and Hardy, 1999). Further, host hiding prior to oviposition is exhibited in common with
310 several other Epyrinae (Eliopoulos et al., in press; Griffiths and Godfray, 1988) as is laying
311 single-egg, or otherwise, small clutches (Ahmed et al., 1997; Finlayson, 1950; Mayhew and
312 Godfray, 1997; Mayhew and Hardy, 1998). The observed positions in which eggs were
313 observed to be laid onto the hosts agree with patterns reported by Ahmed and Islam (1988).

314 *Holepyris sylvanidis* is protandrous, with males emerging in advance of females and
315 mating occurs at the natal site. This observation agrees with Abdella et al. (1985). We
316 observed that mating occurred soon after emergence and that males jump suddenly onto the
317 female just prior to copulation, both observations tally with prior reports (Ahmed et al., 1997;
318 Ahmed and Islam, 1988). Ahmed et al. (1997) further reported that females remain stationary
319 during copulation, while we observed some females running around whilst *in copula*. The

320 mating behaviour of *H. sylvanidis* is not complex and would not present obstacles to this
321 species being mass reared for release as a biocontrol agent.

322 *Holepyris sylvanidis* is synovigenic (Ahmed et al., 1997) and host feeding likely serves to
323 supply nutrients for egg maturation, as in other epyrine species (Lauzière et al., 1999; Pérez-
324 Lachaud and Hardy, 1999). We observed that female *H. sylvanidis* fed primarily on host
325 haemolymph but were also able to eat the tissues of the host larvae. Ahmed et al. (1997) have
326 previously reported that *H. sylvanidis* females attack and feed on the first to the fourth larval
327 instars but not the fifth and sixth, or the pupal stage. In contrast, we observed some feeding on
328 all host developmental stages, although it was rare on fourth, fifth and sixth instar larvae
329 which are primarily used for oviposition. The major discrepancy between our results and
330 those of Ahmed et al. (1997) is that they reported an absence of host feeding on pupal *T.*
331 *confusum* while we observed this commonly.

332 While host feeding leads to more hosts being killed by a current generation of adult
333 parasitoids, it reduces the number of offspring produced per host found and as such may not
334 be a desirable characteristic of beneficials as destructive host-feeders are predicted to suppress
335 host population equilibria as less strongly than other parasitoids (Emerick and Singh, 2016;
336 Jervis et al., 1996a, b; Jervis and Kidd, 1986). Host feeding could influence both the pre-
337 oviposition period and the time required for laying successive eggs once a female has started
338 to oviposit. For example, in the epyrine *Cephalonomia stephanoderis* Betrem the pre-
339 oviposition period was shorter when there were more hosts available (Lauzière et al., 1999).
340 As in several other epyrine species (Mayhew and Heitmans, 2000; Yamasaki, 1982) unmated
341 female *H. sylvanidis* had longer pre-oviposition periods than mated females, possibly because
342 there are fitness advantages to being able to produce both male and female offspring and thus
343 virgin females may tend to delay the production of progeny to increase the probability of
344 being mated before they oviposit (Godfray, 1990; Guertin et al., 1996).

345 We observed that fourth, fifth and sixth instar host larvae were usually used for
346 oviposition, whereas Ahmed et al. (1997) found that only fourth instar larvae were oviposited
347 on. It is likely that early instar hosts are too small to provide sufficient nutrients to support
348 parasitoid development and are thus fed on (see above) but why no oviposition was observed
349 on larger, fifth and sixth instar, hosts by Ahmed et al. (1997) is unclear. The fact that
350 oviposition onto pupal *T. confusum* was observed by neither study suggests that biochemical
351 changes during pupation make pupae nutritionally unsuitable for parasitoid development. Key
352 nutritional differences between the developmental stages of *T. confusum* could potentially be
353 identified using a metabolomics approach tailored to low biomass organisms (Kapranas et al.,
354 submitted; Snart et al., 2015) and the same approach could also be employed to understand
355 better the range of host species it attacks (Eliopoulos et al., 2002).

356 The realized fecundity (a mean of 16.4 eggs laid by each female) of *H. sylvanidis* female is
357 lower than estimates for other epyrines which are natural enemies of stored products pests:
358 *Cephalonomia waterstoni* Gahan, 102 eggs (Finlayson, 1950) and *C. tarsalis*, 85 eggs
359 (Powell, 1938) or 50-200 eggs (Lukáš, 2007). Unlike *C. tarsalis* (Cheng et al., 2003), mating
360 status did not affect fecundity but abiotic environmental conditions could have influenced our
361 estimate of fecundity, given that the epyrine *Prorops nasuta* Waterston is recorded as laying
362 322 eggs at 27°C and only 33 at 18°C (Infante, 2000). However, Abdella et al. (1985) found
363 in a laboratory study optimum conditions for *H. sylvanidis* of 27°C and 57.7% relative
364 humidity. This is close to the conditions in our study. We further found that *H. sylvanidis*
365 females become egg limited as they age and thus do not oviposit on all suitable hosts
366 provided. Some mated females ceased laying as much as 1-2 weeks before dying while in
367 some unmated females oviposition ceased as many as 4 weeks before death.

368 The provision of hosts clearly affected female *H. sylvanidis* longevity, probably because
369 provisioned females were able to host-feed to obtain nutrients for somatic maintenance, as

370 well as egg production, extending their adult life expectancies from around 8 to around 48
371 days. Similarly, estimates of longevity of females of species in the epyrine genus
372 *Cephalonomia* provided with hosts range between 30 and 94 days (Finlayson, 1950; Pérez-
373 Lachaud and Hardy, 1999; Powell, 1938; van Emden, 1931; Yamasaki, 1982), depending on
374 environmental conditions, while without hosts longevity is typically between 10-20 days
375 (Yamasaki, 1982). We also found that unmated *H. sylvanidis* lived longer than mated females,
376 in contrast to *C. tarsalis* in which both mated and unmated females lived an average of 35
377 days (Powell, 1938). Longevity can play an important role in the impact of parasitoids on pest
378 populations (Hausmann et al., 2005).

379 Male parasitoids are typically shorter-lived than females; for instance, Powell (1938)
380 reported that *C. tarsalis* males live for 6 days and our estimate of male *H. sylvanidis* longevity
381 is 7 days. However, male *H. sylvanidis* longevity was greatly enhanced by the provision of
382 nutrients. Male *H. sylvanidis* do not attack or feed on host larvae but may feed on non-host
383 sources of sugar and proteins such as nectar and pollen. We found that providing sucrose
384 alone approximately doubled male life-expectancy; similar enhancements have been reported
385 in other parasitoid species (Fadamiro and Heimpel, 2001). Further providing both sucrose and
386 pollen increased the average male *H. sylvanidis* longevity to 35 days, with some males living
387 over 150 days, possibly because sugars could be used to aid the digestion of pollen (Sigsgaard
388 et al., 2013). Further work will be required to establish how diet components influence
389 longevity and thus how parasitoids should be fed most efficiently to enhance their life-
390 histories (Hausmann et al., 2005). Fed males may also have enhanced courting, mating and
391 dispersal abilities (Kaspi and Yuval, 2000) but the relative mating abilities of fed and unfed
392 male *H. sylvanidis* have not yet been evaluated. Given that females must mate before being
393 able to produce daughters and that only females attack populations of hosts (Ode and Hardy,
394 2008), enhancing the longevity of males and their abilities to find and mate with females may
395 be beneficial in terms of improving the suppression of pest populations.

396 Although data from an industrial bakery and a mill suggest that *T. confusum* populations
397 may be regulated by *H. sylvanidis* (Adler et al., 2012; Prozell and Schöller, 1998), there is
398 limited evidence for the population dynamic interactions between *H. sylvanidis* and its hosts.
399 The need to host-feed and a relatively low fecundity are unlikely to promote *H. sylvanidis*
400 population growth and, if high parasitoid densities are reached, intra-specific mutual
401 interference interactions may further limit its efficacy for biocontrol (Eliopoulos et al., in
402 press).

403

404 6. Conclusions

405 *Holepyris sylvanidis* is a natural enemy of important stored product pests, most notably the
406 flour beetle *Tribolium confusum*, and has the potential to be beneficially deployed for
407 biological control, thus helping to decrease reliance on synthetic pesticides and maintain the
408 organic status of some bakeries (Lorenz et al., 2010; Prozell and Schöller, 1998). While its
409 ability to find hosts within the substrate is good (Lorenz et al., 2010), our data suggest that its
410 realized fecundity is low and on that basis it seems unlikely that this species will have the
411 capability of suppressing pest populations below an economic threshold after inoculative
412 biocontrol release, as may be the case for other epyrine species that attack stored products
413 pests (Eliopoulos et al., in press; Powell, 1938) or field crop pests (Batchelor et al., 2006).
414 Given that many encountered hosts are killed by adult females feeding upon them, and that
415 host-feeding enhances female longevity, *H. sylvanidis* may have better potential for
416 inundative or augmentative deployment, largely as a predator, but whether this is
417 economically feasible will depend on the efficiency of mass rearing methods. As shown here,
418 the food supplemented during rearing can increase the number of progeny per female, but
419 more research on mass-rearing is needed. Indeed, data from a flour mill suggest that *T.*
420 *confusum* population suppression was achieved when several hundred *H. sylvanidis* were

421 released every two weeks but ceased once parasitoid release stopped (Adler et al., 2012).
422 However, the last mentioned study also proved the potential for establishment of *H. sylvanidis*
423 after release. The longevity of this parasitoid makes it a good candidate for an inoculative
424 release strategy implemented in an integrated control strategy, e.g. with effective sanitation
425 programs and heat treatments (Schöller et al., 2006).

426

427 **References**

428 Abdella, M.M.H., Tawfic, M.F.S., Awadallah, K.T., 1985. Biological studies on the bethylid
429 parasitoid *Holepyris sylvanidis* Bréthes. Ann. Agric. Sc. 23, 1355–1363.

430 Adler, C., Schöller, M., Beier, S., 2012. Development of insects in a flour mill treated with
431 *Holepyris sylvanidis* (Hym., Bethylidae) for biological control of the confused flour
432 beetle *Tribolium confusum* (Col., Tenebrionidae). Integrated Protection of Stored
433 Products IOBC-WPRS Bull. 81, 169–170.

434 Ahmed, K.N., Islam, W., 1988. A new record of the parasite *Rhabdepyris zae* Waterston
435 (Hymenoptera: Bethylidae) from Bangladesh and some aspects of its biology. Bangl. J.
436 of Zool. 16, 137–141.

437 Ahmed, K.N., Khatun, M., Nargis, A., Dey, N.C., 1997. Mating, egg laying and host feeding
438 behaviour of *Rhabdepyris zae* Waterston (Hymenoptera: Bethylidae) parasitizing
439 *Tribolium confusum* larvae. Bangl. J. Sci. Ind. Res. 32, 633–637.

440 Athanassiou, C.G., Kavallieratos, N.G., 2014. Evaluation of spinetoram and spinosad for
441 control of *Prostephanus truncatus*, *Rhyzopertha dominica*, *Sitophilus oryzae*, and
442 *Tribolium confusum* on stored grains under laboratory tests. J. Pest. Sci. 87, 469–483.

- 443 Batchelor, T.P., Hardy I.C.W., Barrera J.F., 2006. Interactions among bethylid parasitoid
444 species attacking the coffee berry borer, *Hypothenemus hampei* (Coleoptera:
445 Scolytidae). Biol. Control 36, 106–118.
- 446 Campbell, J.F., Hagstrum, D.W., 2004. Stored product insects in a flour mill: population
447 dynamics and response to fumigant treatments. Entomol. Exp. Appl. 112, 217–225.
- 448 Cheng, L.I., Howard, R.W., Campbell, J.F., Charlton, R.E., Nechols, J.R., Ramaswamy, S.,
449 2003. Behavioral interaction between males of *Cephalonomia tarsalis* (Ashmead)
450 (Hymenoptera: Bethyridae) competing for females. J. Insect Behav. 16(5), 625–645.
- 451 Conlong, D.E., Graham, D.Y., 1988. Notes on the natural host surveys and laboratory rearing
452 of *Goniozus natalensis* Gordh (Hymenoptera, Bethyridae), a parasitoid of *Eldana*
453 *saccharina* Walker (Lepidoptera, Pyralidae) larvae from *Cyperus papyrus* L in southern
454 Africa. J. Entomol. Soc. South. Afr. 51, 115–127.
- 455 Crawley, M.J., 1993. GLIM for ecologists. Oxford: Blackwell Scientific Publications.
- 456 Damon, A., Valle, J., 2002. Comparison of two release techniques for the use of
457 *Cephalonomia stephanoderis* (Hymenoptera: Bethyridae), to control the coffee berry
458 borer *Hypothenemus hampei* (Coleoptera: Scolytidae) in Soconusco, southeastern
459 Mexico. Biol. Control 24, 117–127.
- 460 Eliopoulos, P.A., Athanasiou, C.G., Buchelos, H.C., 2002. Occurrence of Hymenopterous
461 parasitoids of stored product pests in Greece. Integrated Protection of Stored Products
462 IOBC-WPRS Bull. 25, 127–139.
- 463 Eliopoulos, P.A., Kapranas, A., Givropoulou, E.G., Hardy I.C.W. Reproductive efficiency of
464 the bethylid wasp *Cephalonomia tarsalis*: the influences of spatial structure and host
465 density. Bull. Entomol. Res. in press.

- 466 Emerick, B., Singh, A. 2016. The effects of host-feeding on stability of discrete-time host-
467 parasitoid population dynamic models. *Math. Biosci.* 272, 54–63.
- 468 Evans, H.E., 1978. The Bethylidae of America North of Mexico. *Mem. Am. Entomol. Inst.*
469 27, 1–332.
- 470 Fadamiro, H.Y., Heimpel G.E., 2001. Effects of partial sugar deprivation on lifespan and
471 carbohydrate mobilization in the parasitoid *Macrocentrus grandii* (Hymenoptera:
472 Braconidae). *Ann. Entomol. Soc. Am.* 94(6), 909–916.
- 473 Finlayson, L.H., 1950. The biology of *Cephalonomia waterstoni* Gahan (Hymenoptera:
474 Bethylidae), a parasite of *Laemophloeus* (Coleoptera: Cucujidae). *Bull. Entomol. Res.*
475 41(1), 79–97.
- 476 Fürstenau, B., Adler, C., Schultz, H., Hilker, M., 2016. Host habitat volatiles enhance
477 olfactory response of the larval parasitoid *Holepyris sylvanidis* to specifically host-
478 associated cues. *Chem. Senses* (doi: 10.1093/chemse/bjw065).
- 479 Gahan, A.B., 1930. Synomical and descriptive notes on parasitic Hymenoptera. United States
480 Natl. Mus. Proc. 77, 1–12.
- 481 Godfray, H.C.J., 1990. The causes and consequences of constrained sex allocation in
482 haplodiploid animals. *J. Evol. Biol.* 3, 3–17.
- 483 Gordh, G., Móczár, L., 1990. A catalog of the world Bethylidae (Hymenoptera: Aculeata).
484 *Mem. Am. Entomol. Inst.* 46, 1–364.
- 485 Gothilf, S., Mazor, M., 1987. Release and recovery of imported parasites of the carob moth,
486 *Spectorbates ceratonia* (Lepidoptera: Pyralidae) in Israel. *Isr. J. Entomol.* 21, 19–23.
- 487 Green, R.F., Gordh, G., Hawkins, B.A., 1982. Precise sex ratios in highly inbred parasitic
488 wasps. *Am. Nat.* 120(5), 653–665.
- 489 Griffiths, N.T., Godfray, H.C.J., 1988. Local mate competition, sex ratio and clutch size in

- 490 bethylid wasps. *Behav. Ecol. Sociobiol.* 22, 211–217.
- 491 Guertin, D.S., Ode, P.J., Strand, M.R., Antolin, M.F. 1996. Host-searching and mating in an
492 outbreeding parasitoid species. *Ecol. Entomol.* 21, 27–33.
- 493 Hagstrum, D.W., Phillips, T.W., Cuperus G., (Eds) 2012. *Stored product protection*. Kansas
494 State University Agricultural Experiment Station and cooperative Extension Service,
495 Kansas State University, 350 pp.
- 496 Hagstrum, D.W., Subramanyam, B.H., 2009. *Stored-Product Insect Resource*, AACC
497 International INC., St. Paul, MN.
- 498 Hardy, I.C.W., Goubault, M., Batchelor, T.P., 2013. Hymenopteran contests and agonistic
499 behaviour. In: (Hardy ICW & Briffa M, Eds): *Animal Contests*, Cambridge University
500 Press, pp. 147–177.
- 501 Hausmann, C., Wäckers, F.L., Dorn, S., 2005. Sugar convertibility in the parasitoid *Cotesia*
502 *glomerata* (Hymenoptera: Braconidae). *Arch. Insect Biochem. Physiol.* 60, 223–229.
- 503 Hu, Z., Zhao, X., Li, Y., Liu, X., Zhang, Q., 2012. Maternal care in the parasitoid
504 *Sclerodermus harmandi* (Hymenoptera: Bethylinidae). *Plos One* 7(12), 1–7.
- 505 Infante, F., 2000. Development and population growth rates of *Prorops nasuta* (Hymenoptera:
506 Bethylinidae) at constant temperatures. *J. Appl. Ent.* 124, 343–348.
- 507 Jervis, M.A., Kidd, N.A.C., 1986. Host-feeding strategies in Hymenopteran parasitoids. *Biol.*
508 *Rev.* 61, 395–434.
- 509 Jervis, M.A., Hawkins, B.A., Kidd, N.A.C., 1996a. The usefulness of destructive host feeding
510 parasitoids in classic biological control: Theory and observation conflict. *Ecol.*
511 *Entomol.* 21, 41–46.
- 512 Jervis, M.A., Kidd, N.A.C., Heimpel, G.E. 1996b. Parasitoids adult feeding behaviour and
513 biocontrol—a review. *Biocontrol News Inform.* 17, 11–22.

- 514 Jiang, Y., Yang, Z., Wang, X., Hou, Y., (2015) Molecular identification of sibling species of
515 *Sclerodermus* (Hymenoptera: Bethyridae) that parasitize buprestid and cerambycid
516 beetles by using partial sequences of mitochondrial DNA cytochrome oxidase subunit I
517 and 28S ribosomal RNA gene. PLoS ONE 10(3), 1–15.
- 518 Kapranas, A., Snart, C.J.P., Williams, H., Hardy, I.C.W., Barrett, D.A. Untargeted
519 metabolomics of low-biomass samples: metabolome profiles of individual parasitoid
520 wasps differ with age. Scientific Reports, submitted.
- 521 Kaspi, R., Yuval, B., 2000. Post-teneral protein feeding improves sexual competitiveness but
522 reduces longevity of mass reared sterile male Mediterranean fruit flies. Ann. Entomol.
523 Soc. Am. 93, 949–955.
- 524 Khidr, S.K., Mayes, S., Hardy I.C.W., 2013. Primary and secondary sex ratios in a gregarious
525 parasitoid with local mate competition. Behav. Ecol. 24, 435–443.
- 526 Lauzière, I., Pérez-Lachaud, G., Brodeur, J., 1999. Influence of host density on the
527 reproductive strategy of *Cephalonomia stephanoderis*, a parasitoid of the coffee berry
528 borer. Entomol. Exp. Appl. 92, 21–28.
- 529 Legner, E.F., Gordh, G., 1992. Lower navel orangeworm (Lepidoptera, Phycitidae) population
530 densities following establishment of *Goniozus legneri* (Hymenoptera, Bethyridae) in
531 California. J. Econ. Entomol. 85, 2153–2160.
- 532 Legner, E.F., Silveira-Guido A., 1983. Establishment of *Goniozus emigratus* and *Goniozus*
533 *legneri* (Hym: Bethyridae) on navel orangeworm, *Amyelois transitella* (Lep: Phycitidae)
534 in California and biological control potential. Entomophaga. 28, 97–106.
- 535 Lizé, A., Khidr, S.K., Hardy I.C.W., 2012. Two components of kin recognition influence
536 parasitoid aggression in resource competition. Anim. Behav. 83, 793–799.

- 537 Lorenz, S., Adler, C., Reichmuth, C., 2010. Penetration ability of *Holepyris sylvanidis* into the
538 feeding substrate of its host *Tribolium confusum*. *Julius-Kühn-Archiv*. 425, 721–725.
- 539 Lukáš, J., 2007. Age specific fecundity and survivorship of *Cephalonomia tarsalis* (Ashmead)
540 (Hymenoptera: Bethyridae) at different temperatures. *Integrated Protection of Stored*
541 *Products IOBC/WPRS Bull.* 30, 45–51.
- 542 Mayhew, P.J., Godfray, H.C.J., 1997. Mixed sex allocation strategies in a parasitoid wasp.
543 *Oecologia* 110, 218–221.
- 544 Mayhew, P.J., Heitmans, W.R.B., 2000. Life history correlates and reproductive biology of
545 *Laelius pedatus* (Hymenoptera: Bethyridae) in the Netherlands. *Eur. J. Entomol.* 97,
546 313–322.
- 547 Mayhew, P.J., Hardy, I.C.W., 1998. Nonsiblicidal behavior and the evolution of clutch size in
548 bethylid wasps. *Am. Nat.* 151, 409–424.
- 549 Mertins, J.W., 1980. Life history and behaviour of *Laelius pedatus*, a gregarious Bethyrid
550 ectoparasitoid of *Anthrenus verbasci*. *Ann. Entomol. Soc. Am.* 73, 686–693.
- 551 Morgan, D.J.W., Cook, J.M., 1994. Extremely precise sex ratios in small clutches of a
552 bethylid wasp. *Oikos*. 71, 423–430.
- 553 O'Connor, J.P., Ronayne, C., 2002. Irish records of *Holepyris sylvanidis* (Brèthes) and
554 *Plastanoxus munroi* Richards (Hym., Bethyridae). *Entomol. Monthly Mag.* 138,
555 1652–1655.
- 556 Ode, P.J., Hardy, I.C.W., 2008. Parasitoid sex ratios and biological control. In: Wajnberg E,
557 Bernstein C & van Alphen JJM (Eds) *Behavioral ecology of insect parasitoids: from*
558 *theoretical approaches to field applications*. Blackwell Publishing, Oxford pp 253-291.
- 559 Park, T., 1934. Observations on the general biology of the flour beetle *Tribolium confusum*.
560 *Q. Rev. Biol.* 9(1), 36–54.

- 561 Pérez-Lachaud, G., Batchelor, T.P., Hardy, I.C.W., 2004. Wasp eat wasp: Facultative
562 hyperparasitism and intra-guild predation by bethylid wasps. *Biol. Control* 30, 149–155.
- 563 Pérez-Lachaud, G., Hardy, I.C.W., 1999. Reproductive biology of *Cephalonomia*
564 *hyalinipennis* (Hymenoptera: Bethyridae), a native parasitoid of the coffee berry borer,
565 *Hypothenemus hampei* (Coleoptera: Scolytidae), in Chaipas, Mexico. *Biol. Control* 14,
566 152–158.
- 567 Powell, D., 1938. The biology of *Cephalonomia tarsalis* (Ash.), a vespid wasp (Bethyridae:
568 Hymenoptera) parasitic on the saw-toothed grain beetle. *Ann. Entomol. Soc. Am.* 31,
569 44–48.
- 570 Prozell, S; Schöller, M., 1998. Insect fauna of a bakery, processing organic grain and applying
571 *Trichogramma evanescens* Westwood. *Integrated Protection of Stored Products IOBC*
572 *Bull.* 21, 39–44.
- 573 Reichmuth, C., Schöller, M., Ulrichs, C., 2007. *Stored product pests in grain: Morphology –*
574 *Biology – Damage – Control*, AgroConcept Verlagsgesellschaft, Bonn.
- 575 Rilett, R.O., 1949. The biology of *Cephalonomia waterstoni* Gahan. *Can. J. Res.* 27(3),
576 93–111.
- 577 Schöller, M., Flinn, P.W., Grieshop, M.J., Ždárková, E., 2006. Biological control of stored
578 product pests. Heaps, J.W. (Ed), *Insect Management for Food Storage and Processing*
579 *Second Edition*. American Association of Cereal Chemistry International, St Paul, MN,
580 USA, pp. 67–87.
- 581 Sigsgaard, L., Betzer, C., Naulin, C., Eilenberg, J., Enkegaard, A., Kristensen, K., 2013. The
582 effect of floral resources on parasitoid and host longevity: Prospects for conservation
583 biological control on strawberries. *J. Insect Sci.* 13(13), 1–17.

- 584 Snart, C.J.P., Hardy, I.C.W., Barratt, D.A., 2015. Entometabolomics: applications of modern
585 analytical techniques to insect studies. *Entomologia Experimentalis et Applicata* 155,
586 1–17.
- 587 Sreenivas, A.G., Hardy, I.C.W., 2016. Mutual interference reduces offspring production in a
588 brood-guarding bethylid wasp. *Entomol. Exp. Appl.* 159, 260–269.
- 589 Suma, P., Amante, M., Bella, S., La Pergola A., Russo A., 2014. Stored-product insect natural
590 enemies in wheat industry in Sicily. *Integrated Protection of Stored Products IOBC-
591 WPRS Bull.* 98, 227–233.
- 592 Sutton, A.E., Arthur, F.H., Zhu, K.Y., Campbell, J.F., Murray, L.W., 2011. Residual efficacy
593 of synergized pyrethrin + methoprene aerosol against larvae of *Tribolium castaneum*
594 and *Tribolium confusum* (Coleoptera: Tenebrionidae). *J. Stored Prod. Res.* 47, 399–406.
- 595 Tang, X., Meng, L., Kapranas, A., Xu, F., Hardy, I.C.W., Li, B. 2014. Mutually beneficial
596 host exploitation and ultra-biased sex ratios in quasisocial parasitoids. *Nat. Commun.* 5,
597 1–7.
- 598 Trematerra, P., Gentile, P., Brunetti, A., Collins, L.E., Chambers J., 2007. Spatio-temporal
599 analysis of trap catches of *Tribolium confusum* du Val in a semolina-mill, with a
600 comparison of female and male distributions. *J. Stored Prod. Res.* 43, 315–322.
- 601 van Emden, F., 1931. Zur Kenntnis der Morphologie und Ökologie des Brotkäfer-Parasiten
602 *Cephalonomia quadridentata* Duchaussoy. *Z. Morph. Oekol. Tiere*, 23, 425–574.
- 603 Venkatesan, T., Jalali, S.K., SrinivasaMurthy, K., Rabindra, R.J., Bharathi Dasan, C., 2007.
604 Economics of production of *Goniozus nephantidis* (Muesebeck), an important parasitoid
605 of coconut black-headed caterpillar, *Opisina arenosella* (Walker) for biofactories. *J.*
606 *Biol. Control* 21, 53–58.

- 607 Yamasaki, M., 1982. Biology of a sanitary injurious Bethyloid wasp, *Cephalonomia gallicola*
608 (Ashmead) (Hymenoptera: Bethyloidea). Med. Entomol. Zool. 33(3), 221–226.
- 609 Yang, Z.Q., Wang, X.Y., Zhang, Y.N., 2014. Recent advances in biological control of
610 important native and invasive forest pests in China. Biol. Control 68, 117–128.
- 611 Zhang, L.G., Song, S.H., Fan, J.X., 1984. Multiplication of *Scleroderma guani* by male pupae
612 of honeybee. Nat. Enemies Insects (Kunchong Tiandi) 6, 244–247.

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616 **Figure captions.**

617

618 **Table 1** Development time (days \pm SE) of *Holepyris sylvanidis* eggs, larvae and pupae.
619 Means within rows followed by different letters are significantly different (Mann-Whitney-U
620 test, $P < 0.05$).

621 **Table 2.** Components of *Holepyris sylvanidis* mating behaviour

622 **Fig. 1.** Schematic representation of the course of events in a typical sequence of mating
623 behaviour. Each of the main behaviours are reported in upper case, with successive
624 component behaviours reported in lower case.

625 **Fig. 2.** Pre-oviposition period for unmated and mated *H. sylvanidis* females. * = mean
626 significantly different, Mann-Whitney U-test, $P < 0.05$

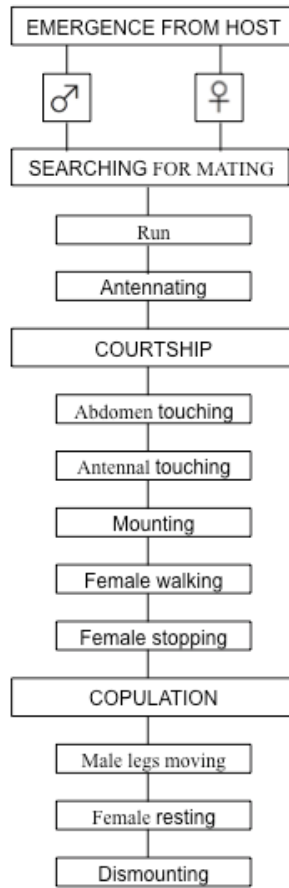
627 **Fig. 3.** The use of presented hosts over ten across ten replicates, with 5 hosts of each of the 7
628 stages present per replicate. The maximum number of hosts of a given stage that could be
629 attacked was 50. Bars are shown stacked.

630 **Fig. 4.** Egg production and longevity of females supplied with hosts twice per week until
631 death. There were two observation periods per week, such that 30 periods represents ≈ 105
632 days. Panel (a) shows the egg production by females alive at a given period. Panel (b) shows
633 the survival of the same cohorts of females: Data are shown as dashed lines and the models
634 fitted using Weibull survival analysis are shown as smooth curves.

635 **Fig. 5.** Cohort survival of females (a) and of males (b). Data are shown as dashed lines and
636 the models fitted using Weibull survival analysis are shown as smooth curves.

637 **Graphical abstract** Development of the parasitoid *H. sylvanidis*: the female laid and egg
638 longitudinally between the first and second abdominal sternite of the host larvae (a), the
639 parasitoid larvae developed as semi-ectoparasitoid (b), and spun a cocoon some centimetres
640 away after the host body was devoured (c,d). The overall development time changed
641 significantly between the two sexes of *H. sylvanidis*.

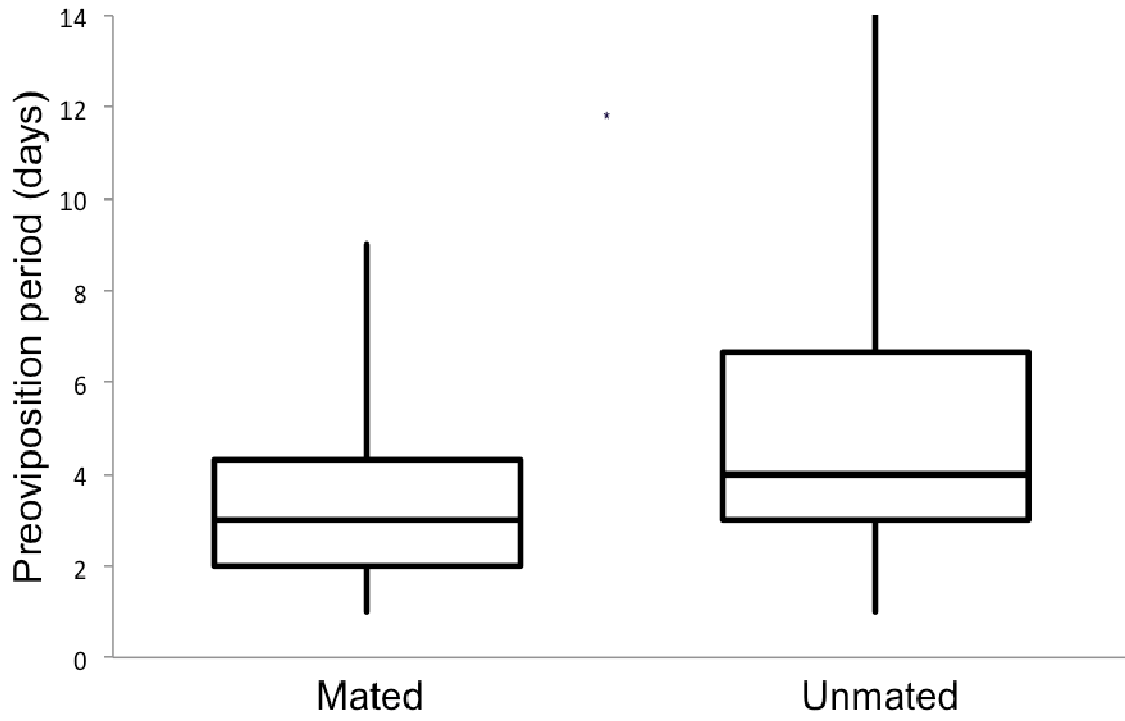
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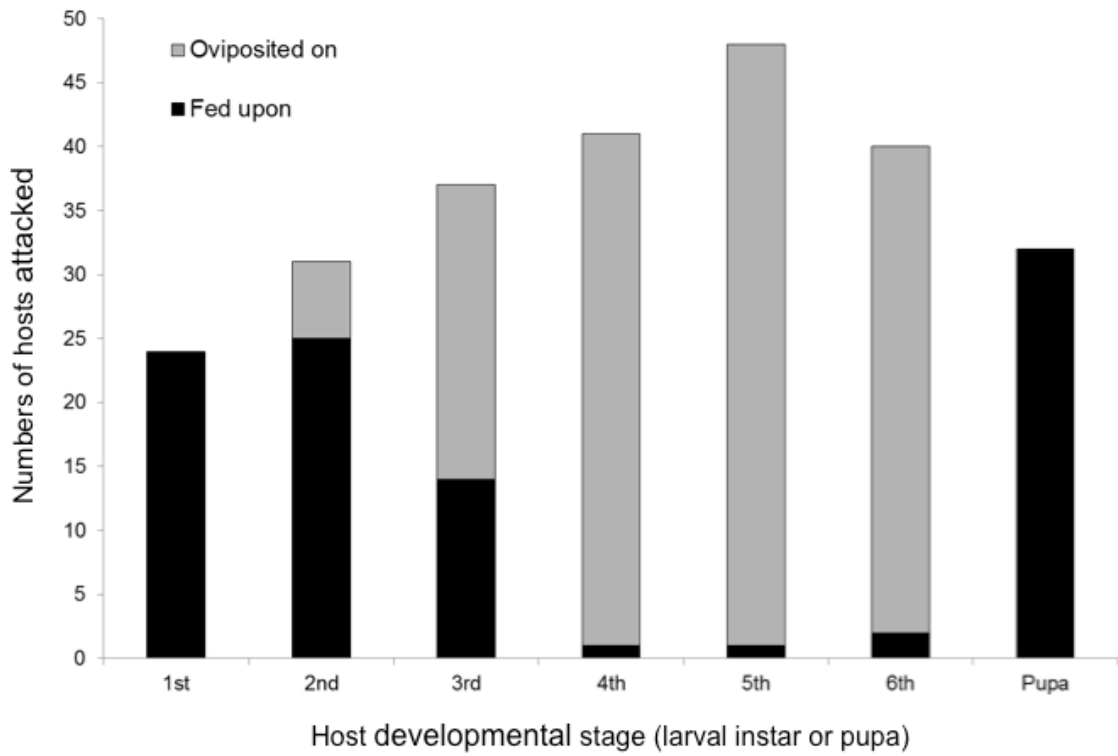
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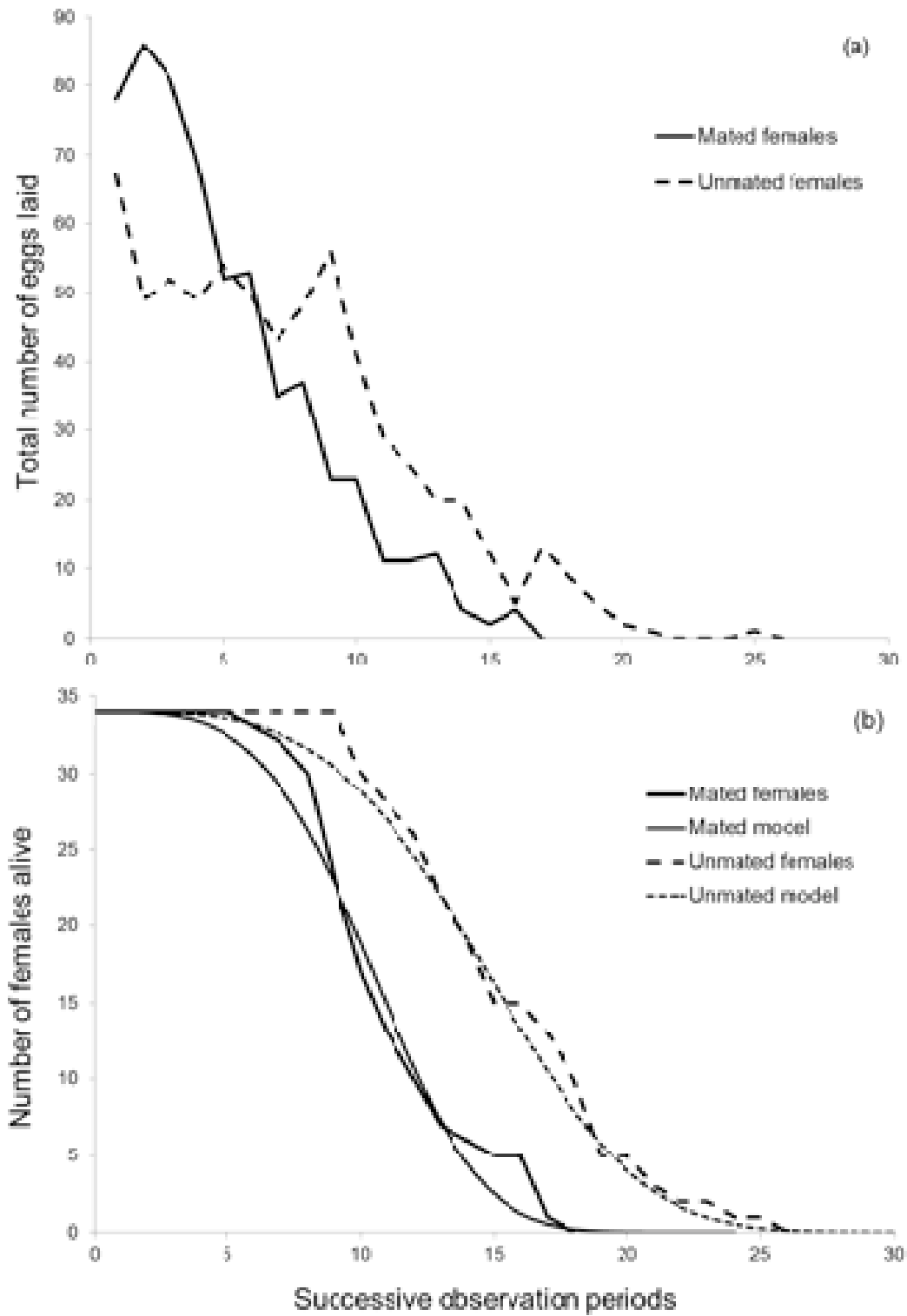
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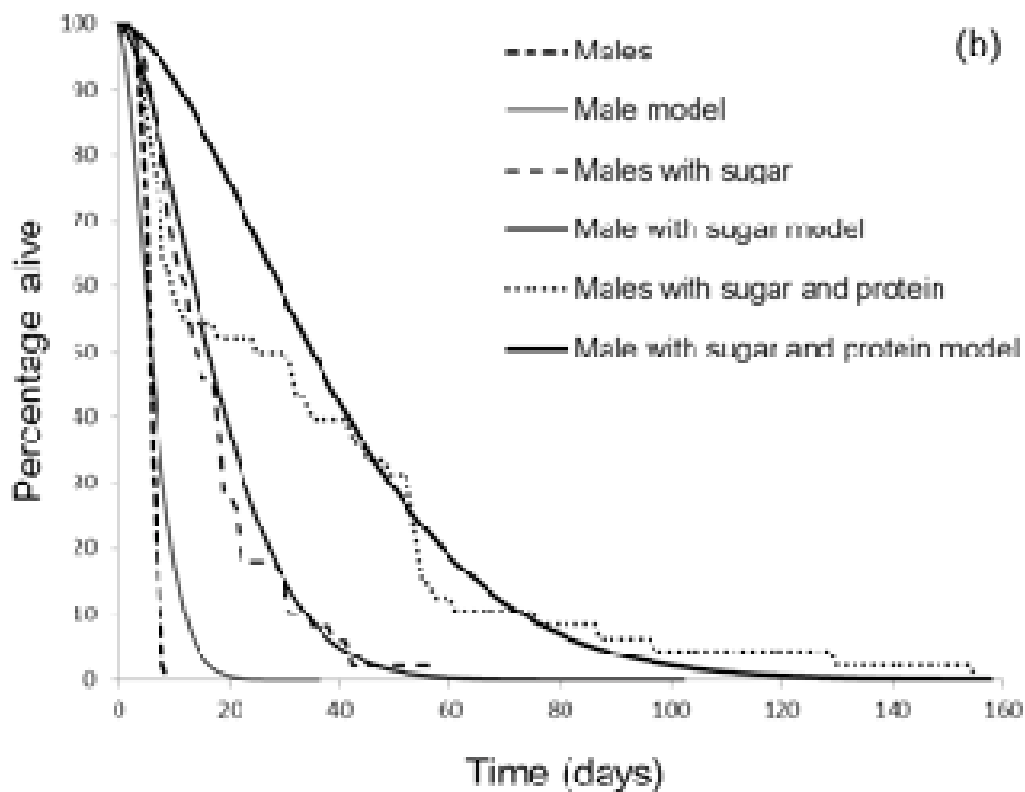
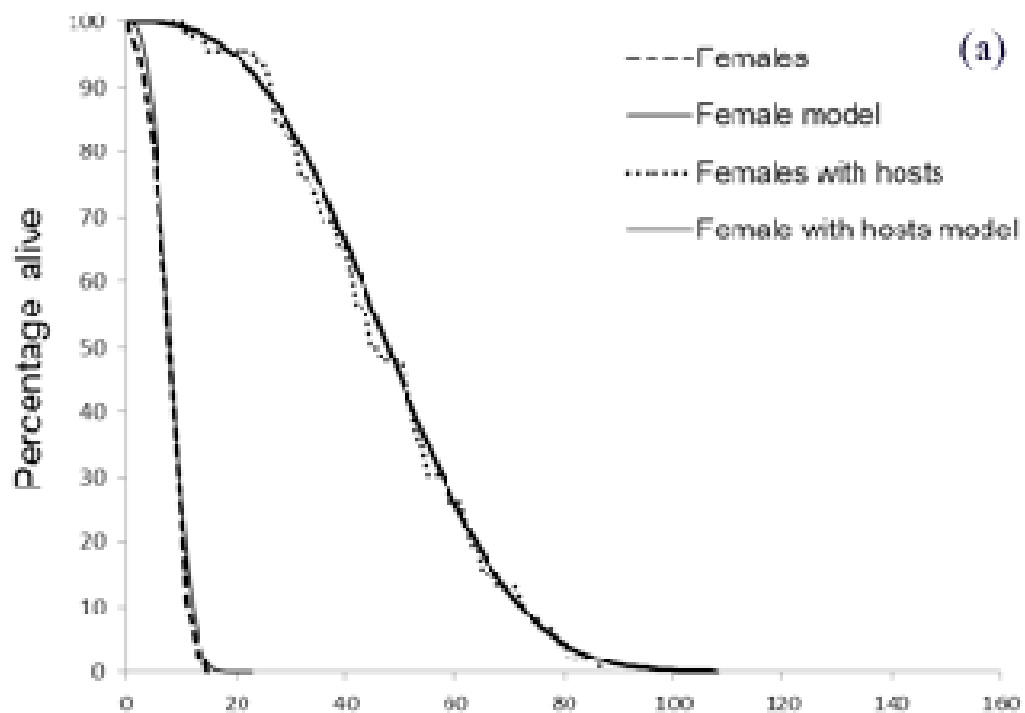
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ACCEPTED MANUSCRIPT

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Stage	Male (n= 35)	Female (n=35)
Egg	2.37 ± 0.11 ^a	2.31 ± 0.09 ^a
Larval	8.80 ± 0.17 ^a	8.86 ± 0.16 ^a
Pupal	11.94 ± 0.15 ^a	13.26 ± 0.23 ^b
Overall egg-to-adulthood	23.11 ± 0.21 ^a	24.43 ± 0.28 ^b

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Behaviour	Description
Run	Male and female run around
Antennating	Male and female waving the antennae kept straight while walking around
Abdomen touching	The female touches the male's abdomen with the antennae
Antennal touching	Male and female wave, and make contact with, their antennae
Mounting	The male jumps suddenly upon the female and establishes genital contact
Female walking	The female moves some centimetres, waving her antennae, while the male is still stationary and upon the female
Female stopping	The female stops walking
Copulation	Male establishes genitalia contact
Male legs moving	The male waves the legs rhythmically
Female resting	The female remains stationary during copulation
Dismounting	The male dismounts from the female and the mating ends

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- 661 • The mating status significantly influences the pre-oviposition period of *H. sylvanidis*
- 662 • The mating status does not influence the fecundity of *H. sylvanidis*
- 663 • A food source significantly influenced the longevity of female and male
- 664 • The life cycle lasts for 23 days in male and 24 days in female at 27 °C
- 665

ACCEPTED MANUSCRIPT