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Abstract: This study aims at investigating the occurrence, risk factors and production impacts on beef carcass parameters of three of the most important cattle helminth infections in England and Wales. Abomasa, reticulorumen and livers from healthy cattle were collected and examined post-mortem quarterly over a one year period in an abattoir in South-West England. Specific viscera from 974 cattle were collected, examined and scored for Ostertagia spp., adult rumen fluke and liver fluke lesions/presence. A total of 89%, 25% and 29% of the carcasses had lesions/presence of Ostertagia spp., rumen fluke and liver fluke, respectively, and 39% had presence of helminth co-infection. Animal demographic and carcass parameters associated with helminth infections were investigated using multilevel multinomial and multilevel linear mixed models respectively. After adjusting for other factors, significant differences in the distribution of helminth infections were observed among cattle by type of breed, animal category (cow, heifer, steer, young bull), age, season and concurrent helminth infections. Compared to carcasses free of helminths, carcasses presenting solely Ostertagia Spp. lesions or adult rumen fluke had significantly lower cold carcass weight (coef.: -30.58 [-50.92;-10.24] and -50.34 [-88.50;-12.18]) and fat coverage (coef.: -3.28 [-5.56;-1.00] and -5.49 [-10.28;-0.69]) and carcasses presenting solely liver fluke lesions had significantly lower conformation grade (coef.: -3.65 [-6.98;-0.32]). Presence of helminth poly-infections was negatively associated with cold carcass weight.

1	Ostertagia spp., rumen fluke and liver fluke single- and poly-infections in cattle: an
2	abattoir study of prevalence and production impacts in England and Wales
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28

#### 29 ABSTRACT

30 This study aims at investigating the occurrence, risk factors and production impacts on beef 31 carcass parameters of three of the most important cattle helminth infections in England and 32 Wales. Abomasa, reticulorumen and livers from healthy cattle were collected and examined 33 post-mortem quarterly over a one year period in an abattoir in South-West England. Specific 34 viscera from 974 cattle were collected, examined and scored for Ostertagia spp., adult rumen 35 fluke and liver fluke lesions/presence. A total of 89%, 25% and 29% of the carcasses had lesions/presence of Ostertagia spp., rumen fluke and liver fluke, respectively, and 39% had 36 presence of helminth co-infection. Animal demographic and carcass parameters associated 37 38 with helminth infections were investigated using multilevel multinomial and multilevel linear 39 mixed models respectively. After adjusting for other factors, significant differences in the 40 distribution of helminth infections were observed among cattle by type of breed, animal 41 category (cow, heifer, steer and young bull), age, season and concurrent helminth infections. 42 Compared to carcasses free of helminths, carcasses presenting solely Ostertagia Spp. lesions 43 or adult rumen fluke had significantly lower cold carcass weight (coef.: -30.58 [-50.92;-10.24] and -50.34 [-88.50;-12.18]) and fat coverage (coef.: -3.28 [-5.56;-1.00] and -5.49 [-44 10.28;-0.69]) and carcasses presenting solely liver fluke lesions had significantly lower 45

- 46 conformation grade (coef.: -3.65 [-6.98;-0.32]). Presence of helminth poly-infections was
  47 negatively associated with cold carcass weight.
- 48

Keywords: Ostertagia spp.; rumen fluke; F. hepatica; co-infection; beef production
impact; multilevel modelling.

51

#### 52 **1. Introduction**

53 Recent projections of the world population's growth have emphasized the urgent need to 54 increase worldwide food production, especially annual meat production (FAO, 2009), while 55 reducing environmental impacts and maintaining high levels of animal health and welfare. In 56 the United Kingdom (UK), parameters such as increased growth rate, higher carcass weight 57 and low-cost grazing systems will be key in enhancing production, given that animal numbers 58 are expected to decline (Thornton, 2010). In this context, production limiting diseases such as 59 helminth infections are of major concern. In temperate areas, helminth infections in grazing 60 livestock are not only an important cause of reduced productivity, but can also lead to poor welfare and contribute to increases in net greenhouse gas emissions (Sargison, 2014). 61 62 Evidence of increases in prevalence and spread of endemic helminths have already been reported in the UK (Sargison, 2014). Helminth infections are seasonal, ubiquitous on 63 64 livestock farms and responsible for major impacts on both animal production and 65 reproduction (Charlier et al., 2014). Beef cattle, are particularly susceptible to such chronic 66 and insidious production limiting diseases because the majority of UK production systems are 67 pasture-based (AHDB, 2009; Sargison, 2014). To date however, very few abattoir studies 68 have been published on the epidemiology and impact of helminth infection in beef cattle 69 (Charlier et al., 2009). In the UK especially, no published abattoir survey on prevalence of helminths in cattle were conducted since the eighties (Froyd, 1975; Bairden and Armour,1981).

In temperate areas such as the UK, two of the most economically important helminth parasites affecting cattle are the abomasal nematode, *Ostertagia ostertagi*, and the liver fluke, *Fasciola hepatica* (Charlier et al., 2014). The recent increasing number of rumen fluke cases in cattle that have been reported in Western Europe also raises concerns about the potential production impact this parasite could have. However, data remain scarce, especially in the UK, and the true prevalence of the rumen fluke in cattle is unknown (Gordon et al., 2013).

Although several diagnostic tools have been developed to detect host exposure to helminths, current methods often have poor specificity and a lack of correlation over time with the actual impact on the host (Charlier et al., 2014). Specific gross examinations of parasitized organs post-mortem is considered the 'gold standard' for assessing prevalence and pathology (Rapsch et al., 2006; Larraillet et al., 2012; Sanchez-Vazquez and Lewis, 2013; Toolan et al., 2015) and could aid in widening and refining our current knowledge on cattle helminth infections.

85 Very few studies have been published on poly-parasitism in adult cattle and none on the impact of such poly-parasitism on cattle production,, especially in the case of co-infections 86 87 with Ostertagia spp., F. hepatica and rumen fluke (Murphy et al., 2006). The aims of this 88 study were to: (1) estimate the prevalence and severity of helminth single and poly-infections 89 in cattle (beef and dairy) at slaughter in England and Wales, focussing on abomasal lesions 90 typical of Ostertagia spp., rumen fluke and liver fluke; 2) investigate if helminth prevalence 91 and severity differed between animal demography and (3) evaluate their production impacts 92 on prime beef carcass weight and classification.

#### 94 **2. Materials and Methods**

#### 95 2.1. Sample collection and viscera scoring

96 Abomasa, reticulorumens and livers from commercial cattle were collected and examined 97 post-mortem quarterly over a twelve month period from March 2014 to January 2015 in an 98 abattoir slaughtering up to 1500 cattle per week in South-West England. On each visit at 99 slaughter, specific viscera from all cattle were inspected on the slaughter line. Livers were 100 examined on-line with the meat inspectors at the abattoir. The liver was examined and scored 101 for the presence of typical cholangiohepatitis lesions ("pipe stem" appearance) and its surface 102 incised as deemed appropriate to detect the presence of liver fluke. Reticulorumens and 103 abomasa were examined in the "gut room", where they were excised and the contents 104 expelled. The internal surfaces of the reticulorumen were visually assessed for the presence 105 of adult rumen fluke and, if present, for their numbers. The abomasum was dissected from the 106 omasum, everted and rinsed to expose the mucosal surface and estimate the number of 107 characteristic lesions of Ostertagia spp. on the fundus and pylorus of each abomasum.

108 Abomasum gross lesions were classified into four categories (scores 0-3) based on the number of gastric gland lesions typical of Ostertagia spp. (Larraillet et al., 2012): 0- no 109 110 lesions; 1- less than 100 lesions; 2- between 100-1000 lesions; 3- more than 1000 lesions. 111 Each reticulum and rumen were thoroughly examined and classified on a numerical scale 112 according to the number of adult rumen fluke (scores 0-3): 0- no fluke; 1- between 1 and 10; 113 2- between 11 and 100; 3- between 101 and 200; 4- more than 200 fluke. The presence of 114 liver fluke (0- no fluke (i.e. neither fluke nor liver fluke lesions); 1- actual presence (i.e. 115 presence of fluke and liver fluke lesions); 3- historical presence (i.e. no fluke but presence of 116 liver fluke lesions)) and the severity of the liver lesions due to liver fluke (0- no lesions; 1-117 moderate lesions; 2- severe lesions) were also scored, based on gross-pathological scales used in previous studies (Sanchez-Vazquez and Lewis, 2013). The scoring of gross lesions was
conducted by the same group of operators at each visit, who were blinded to the identity of
the animal or farm.

Before the commencement of the study, the scoring system was pilot-tested in the same abattoir as a feasibility check. At the same time, a sample of adult rumen fluke specimens were collected from two animals and preserved in 70% methanol and were sent for speciation (Moredun Research Institute, UK), applying PCR amplification and DNA sequencing of the ITS-2 region using generic primers (Rinaldi et al., 2005) with subsequent sequencing of purified PCR amplicons (Gordon et al., 2013).

#### 128 2.2. Animal demographic and carcass parameters

129 Data from the abattoir information management system were used to provide additional 130 information on each animal, using the kill number as the unique identifier. The following 131 demographic information was extracted: date of birth, date of slaughter, farm, breed, sex 132 (male/female), category (mature bull, cow, heifer, steer and young bull), cold carcass weight 133 (CCW) (kg), carcass conformation and fat classifications and liver condemnations (yes/no). 134 No additional information on the history of the animals in relation to previous grazing, 135 housing and anthelmintic treatments was available. To determine the geographic origin of the 136 farm the animals were submitted from, the postcodes of each farm were used and related 137 latitude, longitude and altitude extracted from "Google Maps" (Map data ©2016 Google). 138 The breed information was classified in four categories: pure-dairy, dairy-cross, pure-beef 139 and beef-cross, using the information provided on the passport and DEFRA (Department for 140 Environment, Food and Rural Affairs) breed classification list (DEFRA, 2014). The animal 141 slaughter-age in months was calculated from the date of birth to the kill date. Carcass

<sup>127</sup> 

142 conformation and fat classifications were evaluated referring to the EUROP scale (Pritchard143 et al., 2013).

144

145 2.3. Statistical analysis

Data were coded, checked and entered into a database (Microsoft Excel 2010). A
preliminary descriptive analysis was conducted using STATA 12.1 (STATA Inc., Texas,
USA) to summarize the data. Three sets of analysis were conducted, as described below:

150 2.3.1. Prevalence and severity of helminth infections

Descriptive statistics were conducted to summarise the prevalence of *Ostertagia spp.*, adult rumen fluke and liver fluke at farm level and at cattle level, based on abomasal lesions, presence of adult rumen fluke and *F. hepatica* presence and lesions respectively. For each helminth, the carcasses were summarised based on severity scores of the helminths, season and category of animal. Where scores were available for all three helminths, the percentage of co-infected animals was calculated.

157

### 158 2.3.2. Factors associated with presence and severity of helminth infections

Multinomial logistic regression was used to investigate the relationship between the carcass categorical severity scores for helminths and the general demographic and other collected variables (Dohoo et al., 2009). Three models (one for each helminth) were built. Since several carcasses originated from the same farm, observations could not be considered independent; hence a multilevel mixed-effect model was built accounting for the hierarchy in the data. The three models incorporated two hierarchical levels: level 1 (i), the cattle-level, level 2 (j) the herd-level. The outcome variable was: for model 1, the scores of *Ostertagia*  166 spp. lesions (0- no lesions, 1- less than 100 lesions; 2- between 100-1000 lesions; 3- more 167 than 1000 lesions), for model 2, the scores of adult rumen fluke (0- no fluke; 1- between 1 168 and 100 fluke; 2- more than 100 fluke) and for model 3, the scores of liver fluke lesions (0no lesions; 1- moderate lesions; 2- severe lesions). For all three models the reference category 169 170 for the outcome was score 0 and the predictor variables were: breed, category, age, month of 171 sampling, altitude and presence of co-infection. The model was built using a stepwise approach, combining both forward selection and backward elimination of predictor variables. 172 173 The evaluation of the effects of significant factors on the three outcomes was based on Wald 174 tests. P-value  $\leq 0.05$  was considered significant. Confounding variables also remained in the 175 final model. The multilevel multinomial models 1, 2 and 3 used a logit link function to express the ratio probability of a given helminth score to the probability of the reference 176 score, as shown in equation (1) (Rasbash et al., 2009): 177

178 
$$log\left(\frac{\pi_{ij}^{(s)}}{\pi_{ij}^{(0)}}\right) = \beta_0^{(s)} + \beta_1^{(s)} x_{ij} + u_{0j}^{(s)}$$
(1)

Where:  $\pi_{ij}^{(s)}$  was the probability of the *i*th carcass of the *j*th herd to have a score "s" (s=1, 2, 179 3, for model 1; s=1, 2, for model 2 and 3) compared to the score 0;  $\beta_0^{(s)}$  was the score-specific 180 intercept of the model;  $\beta_1^{(s)}$  represents the vector of coefficients;  $x_{ij}$  was the vector of 181 predictor variables and  $u_{0i}^{(s)}$  was the herd-level random effect, assumed to be normally 182 183 distributed. All statistical analyses were performed using MLwiN v2.30. All the calculations 184 were based on a Restricted Iterative Generalized Least Squares (RIGLS) procedure and a 185 second-order approximation by penalized quasi-likelihood (Rasbash et al., 2009). Models 186 were checked for any influential observations or outliers.

#### 188 2.3.3. Impact of helminths on carcass parameters

189 The impact of helminth past/current infections on beef production carcass parameters was 190 estimated using three multilevel mixed-effect linear regression models with outcomes: (1) the 191 cold carcass weight (CCW), (2) the carcass conformation and (3) the carcass fat 192 classification. Since several carcass originated from the same herd, the model had carcasses 193 nested within herds. Only steers, heifers and young bulls from 12 to less than 36 months were 194 included in this analysis, as these represent the population of cattle reared for prime beef in 195 the UK (AHDB, 2009). The predictor variables for the three models were: breed, category, 196 age, carcass parameters, month, altitude and an eight-level categorical variable for presence 197 of co-infection (i.e. no helminths; Ostertagia spp. lesions only; adult rumen fluke only; liver 198 fluke lesions only; Ostertagia spp. lesions and adult rumen fluke; Ostertagia spp. lesions and 199 liver fluke lesions; adult rumen fluke and liver fluke lesions; Ostertagia spp. lesions, adult 200 rumen fluke and liver fluke lesions). Models were developed using a Restricted Generalised 201 Iterative Least Squares (RIGLS) algorithm in MLwiN 2.30 (Rasbash et al., 2009). Both 202 conformation and fat classifications were converted into a 15-numerical scale (Pritchard et 203 al., 2013). The models were built following the stepwise approach and took the form of 204 equation (2) (Rasbash et al., 2009):

$$y_{ij} = \beta_0 + \beta_1 x_{ij} + u_{0j} + e_{ij}$$
(2)

Where:  $y_{ij}$  was the outcome (CCW/Carcass conformation/Carcass fat classification) of the ith carcass from the *j*th herd;  $\beta_0$  is the intercept;  $\beta_1$  was the coefficient for the effect of a unit increase of the predictor  $x_{ij}$  on the outcome  $y_{ij}$ ;  $u_{0j}$  is the herd-effect and  $e_{ij}$  was the bottom level residual, both assumed to be normally distributed. Model goodness-of-fit was assessed at each hierarchical level by the examination of the normal probability and the leverage plots of residuals (Dohoo et al., 2009; Rasbash et al., 2009).

## **3. Results**

### 214 *3.1. Description of animal and carcass parameters*

215 A total of 974 carcasses were sampled from March 2014 to January 2015: 298 (31%) in 216 March, 233 (24%) in June, 230 (24%) in October and 213 (22%) in January. The carcasses 217 originated from 156 UK farms, localised in 23 counties. A total of 134 (86%) farms could be 218 geo-localised, of which 82% (110/134) were from England and 18% (24/134) from Wales. The median [25<sup>th</sup> percentile (p25) - 75<sup>th</sup> percentile (p75)] number of carcasses per farm was 4 219 220 [2-8]. The sample included 64% males and 36% females, of which 53% (518/974) were 221 steers, 20% (193/974) cows, 16% (155/974) heifers, 11% (106/974) young bulls and less than 1% (2/974) mature bulls. Fifty percent (484/974) of the carcasses were from beef-cross 222 223 breeds, 36% (353/974) from pure-dairy breeds, 9% (83/974) from pure-beef breeds and 4% 224 (42/974) from dairy-cross breeds; the rest (12/974) belonging to either dual-purpose or other 225 breeds. Table 1 presents, by cattle category, the sample median [p25-p75] of age, CCW, 226 conformation and fat classifications, and percentage of liver condemnations.

227

228 3.2. Description of carcass parasites' presence/lesions

229 3.2.1. Prevalence and severity of helminth infections as defined by scores

Adult rumen fluke specimens isolated from the two carcasses sampled in the pilot study

231 were identified as *Calicophoron daubneyi*.

Out of 972 carcasses (mature bulls excluded), a total of 933 abomasa, 936 reticulorumen and 951 livers were scored for *Ostertagia spp.* lesions, presence of adult rumen fluke and liver fluke lesions, respectively; the others being either condemned or lost. There was a large variation in the prevalence of helminths with, at cattle-level, 89% (828/933), 25% (231/936) and 29% (272/951) of the carcasses and, at farm-level, 97% (149/154), 48% (73/153) and 64% (98/152) of the producers with at least one carcass with signs of ostertagiasis, adult rumen fluke and liver fluke lesions respectively. Distribution of carcasses by severity score for each category of animal is presented in Table 2.

240 Of the abomasa with lesions of ostertagiasis, 40% had scores of 3 (>1000 lesions). There 241 was a similar percentage of carcasses with  $\leq 100$  and >100 adult rumen fluke (51% and 49%) 242 respectively). Live F. hepatica were present in approximately 86% of the livers with liver 243 fluke lesions. A seasonal variation was present for the prevalence of helminth in carcasses, 244 with highest prevalence of Ostertagia spp. lesions observed in January (98%), compared with 245 84% in March, 85% in June and 89% in October. A similar pattern was observed for liver 246 fluke lesions and adult rumen fluke with the lowest relative prevalence in March (22% and 247 17% respectively) and highest prevalence in January (34% and 28% respectively) and 248 October (33% and 31% respectively). The prevalence of liver fluke and adult rumen fluke in 249 June was 28% and 25% respectively.

250

#### 251 *3.2.2 Presence of co-infection*

252 Out of the 972 carcasses, 909 (94%) had a score available for all three helminths. Of these, 253 92% (837/909) had at least one helminth presence/lesion. A total of 39% (351/909) of the 254 animals had co-infection, of which 15% (138/909) with Ostertagia spp. lesions and adult 255 rumen fluke, 12% (111/909) with Ostertagia spp. and liver fluke lesions, 11% (97/909) with 256 all the three helminths presence/lesions and 1% (5/909) with only adult rumen fluke and liver fluke lesions. Presence of adult rumen fluke and liver fluke lesions were mainly concurrent 257 258 with other infections, with only 3% (6/219) and 6 % (15/255) of infected animals having single-infection with adult rumen fluke and liver fluke respectively, compared to 57% 259

(465/811) with only *Ostertagia spp.* Out of 219 animals (24%) infected with adult rumen
fluke, 47% (102/219) also had signs of liver fluke lesions. The prevalence of co-infected
animals was highest in October with 50% (104/206) of the carcasses presenting signs of at
least two parasites, compared with 44% (83/189) in January, 35% (81/229) in June and 29%
(83/285) in March. The highest prevalence of co-infection was observed in cows with 51%
(83/162) of the carcasses infected with at least two helminths, compared with 42% (210/502)
for steers, 35% (51/145) for heifers and 7% (7/100) for young bulls.

267

268 3.3. Factors associated with helminth presence/lesions and carcass infection severity

The number of observations for predictor variables per model is presented in Table 3. The three final multilevel multinomial models are presented in Table 4. All significant variables and potential confounders were retained in the model to estimate the independent effect of variables (i.e. effect of variable presented is after adjusting for the effects of variables in the model).

274

### 275 *3.3.1. Model 1 (abomasal lesions due to Ostertagia spp.)*

276 Compared with pure-dairy breeds, dairy-cross breeds were significantly more likely to 277 have Ostertagia spp. lesions of all severities (Odds Ratios [OR]: 7.29; 8.63; 6.20). Whereas 278 beef-cross breeds were significantly less likely to have Ostertagia spp. lesions of higher 279 severity ( $\geq 100$  lesions) (OR: 0.49; 0.45). Compared to cows, heifers were significantly more 280 likely to have Ostertagia spp. lesions of all severities (OR: 2.16; 4.34; 7.11), steers were 281 more likely to have lesions of >100 (OR: 2.06; 2.54) and young bull between 100-1000 (OR: 282 3.15). There was a significant effect of age: compared to animals slaughtered at <24 months of age, animals slaughtered at >30 months were at significantly higher risk of having 283

*Ostertagia spp.* lesions with all severities (OR: 2.72; 2.27; 4.40) and animals slaughtered between 24-30 months more likely to have >1000 lesions (OR: 2.82). Compared to January, there were significant reduced numbers of *Ostertagia spp.* lesions of all severities in March (OR: 0.06 to 0.08), June (OR: 0.04 to 0.11) and October (OR: 0.06 to 0.20). The presence of adult rumen fluke was significantly associated with all severities (OR: 1.92 to 3.01) of abomasal lesions due to *Ostertagia spp.* There was no significant association between the presence of *Ostertagia spp.* lesions and the presence of liver fluke.

291

#### 292 *3.3.2. Model 2 (presence of adult rumen fluke)*

293 There was no significant association between the presence of adult rumen fluke and the 294 different breeds. Compared to cows, steers were significantly more likely to have adult rumen 295 fluke infestation of all severities (OR: 2.51 to 3.95) and heifers more likely to have 1 to 100 296 rumen fluke (OR: 2.55). Animals slaughtered older than 30 months were significantly more 297 likely to be heavily infected with adult rumen fluke (>100) than animals slaughtered younger 298 than 24 months (OR: 5.48). Compared with March, there were increased numbers of >100 299 adult rumen fluke infested animals in June (OR: 2.32), October (OR: 2.82) and January (OR: 300 4.45). Carcasses originating from higher altitude farms (>60m) were significantly less likely 301 to have adult rumen fluke compared to carcasses originating from lower altitude farms 302  $(\leq 60m)$  (OR: 0.44 to 0.58). Presence of liver fluke lesion was significantly associated with 303 adult rumen fluke infestation of all severities (OR: 1.79 to 5.34). There was no significant 304 association between the presence of abomasal lesions due to Ostertagia spp. and the 305 likelihood/severity of adult rumen fluke.

306

#### 307 *3.3.3. Model 3 (liver lesions due to liver fluke)*

308 Compared to pure-dairy breeds, beef-cross breeds were significantly more likely to have 309 both moderate and severe liver lesions due to liver fluke (OR: 2.30 to 3.18). Compared to 310 cows, heifers were significantly less likely to have liver fluke lesions (moderate and severe) 311 (OR: 0.08 to 0.43), steers less likely to have severe liver fluke lesions (OR: 0.13) and young 312 bulls less likely to have moderate liver fluke lesions (OR: 0.04). After controlling for the 313 other variables, there was no significant association between the age the animal was 314 slaughtered and the presence of liver fluke lesions. Compared with March, there were 315 significantly higher numbers of carcasses with liver fluke lesions of all severities in January 316 (OR: 1.75 to 3.20) and of moderate severity in October (OR: 2.06). Carcasses originating 317 from higher altitude farms (>60m) were significantly less likely to have moderate liver fluke 318 lesions compared to carcasses originating from lower altitude farms ( $\leq 60m$ ) (OR: 0.56). 319 Presence of adult rumen fluke was significantly associated with presence of liver fluke 320 lesions with all severities (OR: 2.71 to 4.08). There was no significant association between 321 the presence of liver fluke lesions and Ostertagia spp. lesions.

322

#### 323 *3.4. Impact of helminth presence/lesions on carcass parameters*

The final multilevel linear regression models are summarized Table 5. The total of variance explained by the different final models was: for Model 1 (CCW), 50%, for Model 2 (conformation), 33%, for Model 3 (fat classification), 64%.

After controlling for the effects of breed, category, age and season, animals with singleinfection of either ostertagiasis or adult rumen fluke had, on average, significantly lower CCW [Coef. (95% CI): -30.58 (-50.92;-10.24) and -50.34 (-88.50;-12.18)] and lower fat class [Coef. (95% CI): -3.28 (-5.56;-1.00) and -5.49 (-10.28;-0.69)] respectively than carcasses from helminth-free animals. The presence of liver fluke lesions had no significant impact on CCW except when present along with both abomasal lesions due to *Ostertagia spp.* and adult rumen fluke, leading to significantly lower CCW [Coef. (95% CI): -48.28 (-88.35;-8.21)] compared to carcasses free of the three helminths. Carcasses with both *Ostertagia spp.* lesions and adult rumen fluke had significantly lower CCW [Coef. (95% CI): -39.99 (-73.09;-6.88)] compared to carcasses free of the three helminths. The presence of liver fluke lesions on its own had a significant negative impact on carcass conformation by a 3.65 (-6.98;-0.32) point decrease in the class numerical scale compared to carcasses free of the 3 helminths.

339 Visual examinations of the three models final residuals at each hierarchical level340 suggested the model fits were good (data not shown).

341

#### 342 **4. Discussion**

To the authors' knowledge, this is not only the first abattoir study since the eighties on *Ostertagia spp.* and liver fluke prevalence in cattle in England and Wales (Froyd, 1975; Burrows et al., 1980; Bairden and Armour, 1981; Hong et al., 1981), but also the first abattoir survey on cattle helminths to include rumen fluke and co-infection in this region.

Although interpretation of these data should be cautious given the absence of information 347 on previous anthelmintic treatment and past grazing history, the prevalence of cattle 348 349 ostertagiasis reported in the current study was 89%, which is quite similar to that recorded in 350 previous European abattoir surveys (86% to 97%) (Agneessens et al., 2000; Borgsteede et al., 351 2000) and much higher than that observed in the current study for F. hepatica and adult 352 rumen fluke (29% and 25% respectively). Very few farms (3%) in the current study had cattle 353 with no evidence of abomasal lesions due to Ostertagia spp. compared with 52% and 36% of 354 farms without any presence of adult rumen fluke and liver fluke lesions, respectively. These 355 results confirm the predominance and ubiquity of Ostertagia spp. infection among cattle 356 farms in England and Wales (Hong et al., 1981), mainly related to the relatively simple direct 357 life-cycle of this parasite compared with the indirect life-cycles of the two trematodes 358 (McCann et al., 2010b; Gordon et al., 2013). The estimate of prevalence of adult rumen fluke 359 in the current study at 25% is quite similar to that previously recorded in cattle at slaughter in 360 mainland Europe (Szmidt-Adjide et al., 2000; Gonzalez-Warleta et al., 2013; Malrait et al., 361 2015) and confirms the establishment of this trematode in the UK (Gordon et al., 2013). A 362 higher prevalence (52%) of adult rumen fluke was recently recorded in a similar study in the 363 Republic of Ireland (ROI) (Toolan et al., 2015) and could be attributed to differences in 364 environment and cattle production systems (Murphy et al., 2006; Toolan et al., 2015). Overall, 29% of the cattle were infected with liver fluke. The only similar abattoir survey 365 366 conducted in Great Britain was more than forty years ago (Froyd, 1975). Given the expected huge variability in climate conditions and the important changes that occurred in UK 367 368 livestock farming since the eighties, comparison of both studies is difficult. However, there 369 has been evidence of a recent spread in the UK of liver fluke infection in cattle (Pritchard et 370 al., 2005).

All the specimens of adult rumen fluke isolated were identified as C. daubneyi and not P. 371 372 cervi, which was previously assumed to be the predominant rumen fluke species in the 373 British Isles (Gordon et al., 2013). Despite this, the possibility of other species being present 374 in England and Wales cannot be excluded, given that only two carcasses were sampled for 375 adult rumen fluke speciation. However, this result complements previous work conducted in 376 Scotland and Ireland (Gordon et al., 2013; Zintl et al., 2014) and emphasizes the importance, 377 if not predominance, of C. daubneyi in the UK, as it is in mainland Europe (Szmidt-Adjide et 378 al., 2000; Gonzalez-Warleta et al., 2013; Gordon et al., 2013).

379 In the current study, 39% of the carcasses had signs of co-infection. The similar environmental requirements and common microclimate and microhabitat shared by the three 380 381 helminths and their intermediate hosts may explain some of the animals' co-infection, but 382 not entirely (Viney and Graham, 2013). As for instance, cattle anthelmintic or management 383 practices on farms may generate different patterns of co-infection (Gordon et al., 2013). 384 However, this information was not currently available to explore any patterns. The presence 385 of adult rumen fluke was significantly associated with the presence of liver fluke lesions. 386 Because both helminths have very similar life cycles and both F. hepatica and C. daubneyi 387 can share the same intermediary host Galba truncatula (Zintl et al., 2014), it has been 388 suggested that cattle infected with one fluke could simultaneously be infected with the other 389 (Gordon et al., 2013). Although the presence of both fluke species was associated, only half 390 of the animals (102/219) infected in the current study with adult rumen fluke had signs of 391 liver fluke lesions. As reported previously, different lymnaeid communities can act as 392 intermediate hosts for the two helminths and in the UK snails other than Galba truncatula 393 may play an important role as intermediate host (Dreyfuss et al., 2014). Under these circumstances, competition between either the parasites or the intermediate hosts, especially 394 395 for food in colonized habitat, could explain the predominance of such fluke single-infections 396 (Dreyfuss et al., 2014). These results raise questions on the current dynamic of helminth 397 infections in cattle in the UK and the need to fully understand host-helminths interactions and 398 co-evolution, especially in the context of specific helminth poly-infections (Gasbarre, 1997; 399 Viney and Graham, 2013).

400 As previously reported in the literature (Myers and Taylor, 1989; McCann et al., 2010a), 401 there was a significantly higher risk of carcass helminth infection/lesions in October-January, 402 compared to March-June, which could be related to the specific life cycles of the three 403 helminths. It is also possible that exposure of animals slaughtered in March-June was 404 reduced, given, in the UK, animals are often housed in the winter and beef cattle often 405 undergo a two-month fattening period while housed before slaughter (AHDB, 2009). Unlike 406 this study, the seasonality of *Ostertagia spp*. was not reported in a similar beef study 407 (Charlier et al., 2009), which we could be attributable to its study design and lack of test 408 specificity of the diagnostic ELISA test used.

After controlling for breed, cows were less likely to present *Ostertagia spp.* lesions and adult rumen fluke, but more likely to present liver fluke lesions compared to heifers and steers. In both cases, this is likely to be related to the development of some host immunity that, for both *Ostertagia spp.* (Gasbarre, 1997) and rumen fluke (Diaz et al., 2006), would reduce the worm burden and for liver fluke would cause liver fibrosis, enabling the maintenance of the infection (Mendes et al., 2013).

415 Presence of liver fluke lesion solely compared with no lesion was only significantly 416 associated with lower conformation, but neither CCW nor fat classification as reported in 417 previous similar study (Sanchez-Vazquez and Lewis, 2013). There are several studies that 418 have failed to demonstrate effect of liver fluke infection on cattle growth rate and there is a 419 possibility that *F. hepatica* may alter host performance through mechanisms other than body weight (Loyacano et al., 2002; Charlier et al., 2009). The study by Sanchez-Vazquez and 420 421 Lewis (2013) reported small significant negative effects of liver fluke on CCW and fat 422 classification. There is possibility that this effect observed in their study could be attributed to 423 the impact of presence of other co-infections that were not investigated, especially, given in 424 the current study, liver fluke in combination with Ostertagia spp. and rumen fluke did have 425 an impact on CCW. The current results on Ostertagia spp. single effect on CCW and fat 426 classification agree with previous intervention studies on beef cattle (Suarez et al., 1991;

427 Loyacano et al., 2002) but contradict a recent abattoir survey in which no similar association was reported, though there was an effect on conformation (Charlier et al., 2009). It is likely 428 429 in this case that the lower specificity of O. ostertagi ELISA used in the latter study, combined 430 with the inclusion of only adult cows and the non-control of other helminth infections in the 431 model may explain such differences. Our model results suggest that compared to no lesion 432 negative impact of Ostertagia spp on CCW was higher on average (coefficient values) when 433 present along with the other two parasites. It is possible, as reported in a previous study that 434 gastro-intestinal nematodes and liver fluke impact on host performance through different 435 mechanisms and that if present simultaneously the resulting effect might be additive on the 436 CCW (Loyacano et al., 2002). Further research would need to be conducted to confirm this 437 hypothesis.

438 To our knowledge, there has not been any study on the effect of adult rumen fluke on 439 carcass weight and classification. In the current study, there was significant negative 440 association between rumen fluke and CCW and fat classification. Compared to carcasses with 441 no lesion this effect was seen when rumen fluke was present on its own or along with both 442 Ostertagia spp and liver fluke. These results bring into question the widely held view in 443 Europe that adult rumen fluke are relatively benign and well tolerated by their host, contrary 444 to tropical regions where its high pathogenicity was confirmed (Zintl et al., 2014; Fuertes et 445 al., 2015). Given in the current study there were only few animals solely infected by rumen 446 fluke, there is a need of further investigations into pathogenicity of adult rumen fluke in 447 cattle. In addition, what cannot be ascertained in the current study is whether any of the 448 animals that were positive for adult rumen fluke may also have been infected with juvenile 449 fluke in the duodenum; these stages are known to be highly pathogenic when present in large 450 numbers (Millar et al., 2012).

451 Although highly specific, meat inspection is considered as a poorly sensitive diagnostic 452 tool (Rapsch et al., 2006; Sanchez-Vazquez and Lewis, 2013), which is likely to underestimate the prevalence estimates. Moreover, only the presence/lesions of adult 453 454 parasites but not juveniles were screened in the current study, which also may have led to 455 underestimation of prevalence. However, this underestimation is less likely to effect the 456 observed associations and co-infection patterns. This cross-sectional study provides us with 457 associations between various factors and presence of helminths but does not infer causality. 458 During this study, steps were taken to minimise bias by validating the feasibility and 459 reliability of the scoring system in a pilot study and by maintaining throughout the study the 460 same group of operators for scoring. Though the study was only conducted on one abattoir 461 limiting its generalisability, this abattoir is one of the largest abattoir in England with a 462 relatively high throughput. The farms were localised in 23 counties and given the study 463 sampling occurred throughout the year, it was possible to include different types of cattle 464 production systems. Finally, the study sample demographic agreed with a recent survey on 465 the general characteristics of the British beef production cattle (Pritchard et al., 2013). As a conclusion, the current study provided a good picture of Ostertagia spp., rumen fluke and 466 467 liver fluke prevalence/intensity, associated factors and production impacts on cattle in 468 England and Wales.

469

#### 470 **Conflict of interest**

471 The authors declare that they have no competing interests.

472

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#### 477 **References**

- 478 Agneessens, J., Claerebout, E., Dorny, P., Borgsteede, F.H., Vercruysse, J., 2000. Nematode
  479 parasitism in adult dairy cows in Belgium. Vet. Parasitol. 90, 83-92.
- 480 AHDB, 2009. In the balance: the future of the English beef industry. AHDB Beef and Lamb
  481 Report. 24 pp.
- Bairden, K., Armour, J., 1981. A survey of abomasal parasitism in dairy and beef cows in
  south-west Scotland. Vet. Rec. 109, 153-155.
- Borgsteede, F.H., Tibben, J., Cornelissen, J.B., Agneessens, J., Gaasenbeek, C.P., 2000.
  Nematode parasites of adult dairy cattle in the Netherlands. Vet. Parasitol. 89, 287296.
- Burrows, R.O., Davison, C.C., Best, P.J., 1980. Survey of abomasal parasitism of culled dairy
  cows in southern Britain. Vet. Rec. 107, 289-290.
- Charlier, J., De Cat, A., Forbes, A., Vercruysse, J., 2009. Measurement of antibodies to
  gastrointestinal nematodes and liver fluke in meat juice of beef cattle and associations
  with carcass parameters. Vet. Parasitol. 166, 235-240.
- Charlier, J., van der Voort, M., Kenyon, F., Skuce, P., Vercruysse, J., 2014. Chasing
  helminths and their economic impact on farmed ruminants. Trends Parasitol. 30, 361367.
- 495 DEFRA, 2014. CTS updated breed code list. Guidance on keeping cattle, bison and buffalo in
  496 Great Britain. 68 pp.
- 497 Diaz, P., Lomba, C., Pedreira, J., Arias, M., Sanchez-Andrade, R., Suarez, J.L., Diez-Banos,
  498 P., Morrondo, P., Paz-Silva, A., 2006. Analysis of the IgG antibody response against

- 499 *Paramphistomidae trematoda* in naturally infected cattle. Application to serological
  500 surveys. Vet. Parasitol. 140, 281-288.
- 501 Dohoo, I., Martin, W., Stryhn, H., 2009. Veterinary epidemiologic research VER Inc., 865
  502 pp.
- 503 Dreyfuss, G., Vignoles, P., Rondelaud, D., 2014. *Fasciola hepatica* and *Paramphistomum*504 *daubneyi*: decrease in prevalence of natural infection in habitats colonized by *Galba*505 *truncatula* and *Lymnaea glabra*. Rev. Med. Vet 165, 160-166.
- 506 FAO, 2009. How to feed the world in 2050. 35 pp.
- Froyd, G., 1975. Liver fluke in Great Britain: a survey of affected livers. Vet. Rec. 97, 492495.
- Fuertes, M., Perez, V., Benavides, J., Gonzalez-Lanza, M.C., Mezo, M., Gonzalez-Warleta,
  M., Giraldez, F.J., Fernandez, M., Manga-Gonzalez, M.Y., Ferreras, M.C., 2015.
  Pathological changes in cattle naturally infected by *Calicophoron daubneyi* adult
- 512 flukes. Vet. Parasitol. 209, 188-196.
- Gasbarre, L.C., 1997. Effects of gastrointestinal nematode infection on the ruminant immune
  system. Vet. Parasitol. 72, 327-337; discussion 337-343.
- 515 Gonzalez-Warleta, M., Lladosa, S., Castro-Hermida, J.A., Martinez-Ibeas, A.M., Conesa, D.,
- 516 Munoz, F., Lopez-Quilez, A., Manga-Gonzalez, Y., Mezo, M., 2013. Bovine
- 517 paramphistomosis in Galicia (Spain): prevalence, intensity, aetiology and geospatial
- 518 distribution of the infection. Vet. Parasitol. 191, 252-263.

519	Gordon, D.K., Roberts, L.C., Lean, N., Zadoks, R.N., Sargison, N.D., Skuce, P.J., 2013.
520	Identification of the rumen fluke, Calicophoron daubneyi, in GB livestock: possible
521	implications for liver fluke diagnosis. Vet. Parasitol. 195, 65-71.

- Hong, C., Lancaster, M.B., Michel, J.F., 1981. Worm burdens of dairy heifers in England and
  Wales. Vet. Rec. 109, 12-14.
- Larraillet, L., Forbes, A.B., Pravieux, J.J., 2012. Abattoir survey of abomasal lesions
  associated with ostertagiosis in adult cattle. Vet. Rec. 171, 299.
- Loyacano, A.F., Williams, J.C., Gurie, J., DeRosa, A.A., 2002. Effect of gastrointestinal
  nematode and liver fluke infections on weight gain and reproductive performance of
  beef heifers. Vet. Parasitol. 107, 227-234.
- Malrait, K., Verschave, S., Skuce, P., Van Loo, H., Vercruysse, J., Charlier, J., 2015. Novel
  insights into the pathogenic importance, diagnosis and treatment of the rumen fluke
  (*Calicophoron daubneyi*) in cattle. Vet. Parasitol. 207, 134-139.
- McCann, C.M., Baylis, M., Williams, D.J., 2010a. The development of linear regression
   models using environmental variables to explain the spatial distribution of *Fasciola hepatica* infection in dairy herds in England and Wales. Int. J. Parasitol. 40, 1021 1028.
- McCann, C.M., Baylis, M., Williams, D.J., 2010b. Seroprevalence and spatial distribution of
   *Fasciola hepatica*-infected dairy herds in England and Wales. Vet. Rec. 166, 612-617.
- 538 Mendes, E.A., Mendes, T.A., dos Santos, S.L., Menezes-Souza, D., Bartholomeu, D.C.,
- 539 Martins, I.V., Silva, L.M., Lima Wdos, S., 2013. Expression of IL-4, IL-10 and IFN-

- 540 gamma in the liver tissue of cattle that are naturally infected with *Fasciola hepatica*.
  541 Vet. Parasitol. 195, 177-182.
- 542 Millar, M., Colloff, A., Scholes, S., 2012. Disease associated with immature paramphistome
  543 infection. Vet. Rec. 171, 509-510.
- Murphy, T.M., Fahy, K.N., McAuliffe, A., Forbes, A.B., Clegg, T.A., O'Brien, D.J., 2006. A
  study of helminth parasites in culled cows from Ireland. Prev. Vet. Med. 76, 1-10.
- 546 Myers, G.H., Taylor, R.F., 1989. Ostertagiasis in cattle. J. Vet. Diagn. Invest. : official
  547 publication of the American Association of Veterinary Laboratory Diagnosticians, Inc
  548 1, 195-200.
- 549 Pritchard, G.C., Forbes, A.B., Williams, D.J., Salimi-Bejestani, M.R., Daniel, R.G., 2005.
  550 Emergence of fasciolosis in cattle in East Anglia. Vet. Rec. 157, 578-582.
- 551 Pritchard, T., Wall, E., Moore, K., Coffey, M., 2013. Feasibility of using abattoir generated
  552 data and BCMS records for carcass trait evaluations (Carcass Trait Evaluations).
  553 SRUC Project.
- Rapsch, C., Schweizer, G., Grimm, F., Kohler, L., Bauer, C., Deplazes, P., Braun, U.,
  Torgerson, P.R., 2006. Estimating the true prevalence of *Fasciola hepatica* in cattle
  slaughtered in Switzerland in the absence of an absolute diagnostic test. Int. J.
  Parasitol. 36, 1153-1158.
- Rasbash, J., Steele, F., Browne, W.J., Goldstein, H., 2009. A User's Guide to MLwiN.
  Manual, Centre for Multilevel Modelling, University of Bristol. 306 pp.
- 560 Rinaldi, L., Perugini, A.G., Capuano, F., Fenizia, D., Musella, V., Veneziano, V., Cringoli,
  561 G., 2005. Characterization of the second internal transcribed spacer of ribosomal

- 562 DNA of *Calicophoron daubneyi* from various hosts and locations in southern Italy.
  563 Vet. Parasitol. 131, 247-253.
- Sanchez-Vazquez, M.J., Lewis, F.I., 2013. Investigating the impact of fasciolosis on cattle
  carcase performance. Vet. Parasitol. 193, 307-311.
- Sargison, N.D., 2014. Sustainable helminth control practices in the United Kingdom. Small
  Rumin. Res. 118, 35-40.
- Suarez, V.H., Bedotti, D.O., Larrea, S., Busetti, M.R., Garriz, C.A., 1991. Effects of an
  integrated control programme with ivermectin on growth, carcase composition and
  nematode infection of beef cattle in Argentina's western pampas. Res. Vet. Sci. 50,
  195-199.
- 572 Szmidt-Adjide, V., Abrous, M., Adjide, C.C., Dreyfuss, G., Lecompte, A., Cabaret, J.,
  573 Rondelaud, D., 2000. Prevalence of *Paramphistomum daubneyi* infection in cattle in
  574 central France. Vet. Parasitol. 87, 133-138.
- 575 Thornton, P.K., 2010. Livestock production: recent trends, future prospects. Philosophical
  576 transactions of the Royal Society of London. Series B, Biolog. Sci. 365, 2853-2867.
- Toolan, D.P., Mitchell, G., Searle, K., Sheehan, M., Skuce, P.J., Zadoks, R.N., 2015. Bovine
  and ovine rumen fluke in Ireland-Prevalence, risk factors and species identity based
  on passive veterinary surveillance and abattoir findings. Vet. Parasitol. 212, 168-174.
- 580 Viney, M.E., Graham, A.L., 2013. Patterns and processes in parasite co-infection. Adv.
  581 Parasitol. 82, 321-369.

582	Zintl, A., Garcia-Campos, A., Trudgett, A., Chryssafidis, A.L., Talavera-Arce, S., Fu, Y.,
583	Egan, S., Lawlor, A., Negredo, C., Brennan, G., Hanna, R.E., De Waal, T., Mulcahy,
584	G., 2014. Bovine paramphistomes in Ireland. Vet. Parasitol. 204, 199-208.
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586	

# **Table 1.**

- 589 Cattle median [p25-p75] age, cold carcass weight (CCW), carcass conformation and fat
- 590 classifications and percentage of liver condemnations by category (N=972).

Variables (N)	<b>Cows (193)</b>	Heifers (155)	Steers (518)	Young Bulls (106)
Age (Months)	79 [56-113]	29 [26-31]	29 [26-31]	14 [14-15]
CCW (Kg)	323 [283-346]	314 [290-334]	344 [307-384]	294 [267-334]
Conformation	$P^{+}[P^{+}-O^{+}]$	R [O <sup>+</sup> -R]	$O^{+}[O^{+}-R]$	$O^{+}[O^{+}-R]$
Fat classification	3 [2-4L]	4L [3-4L]	3 [3-4L]	2 [2-3]
Liver condemnation	31.6	12.9	14.1	9.4

# **593 Table 2.**

- 594 Stratification of abomasa, reticulorumen and livers scoring percentages by cattle category
- 595 (N=972).

	Cows	Heifers	Steers	Young	Bulls	TOTAL					
Ostertagia spp. lesion (N=9	33)										
0- No lesion	16 (9)	12 (8)	65 (13)	12 (12)		105 (11)					
<b>1-</b> ≤ 100	48 (28)	36 (25)	136 (26)	33 (32)		253 (27)					
2-101-1000	43 (25)	34 (23)	126 (25)	37 (36)		240 (26)					
3- > 1000	65 (38)	64 (44)	186 (36)	20 (20)		335 (36)					
Adult rumen fluke presence (N=936)											
0- No fluke	135 (77)	112 (76)	361 (70)	97 (95)		705 (75)					
1- ≤100	17 (10)	23 (16)	75 (15)	4 (4)		119 (13)					
2->100	23 (13)	12 (8)	76 (15)	1 (1)		112 (12)					
Liver fluke lesion (N=951)											
0- No lesion	94 (51)	116 (75)	367 (72)	102 (98)		679 (72)					
1- Moderate	62 (34)	32 (21)	128 (25)	1 (1)		223 (23)					
2- Severe	28 (15)	6 (4)	14 (3)	1 (1)		49 (5)					
F. hepatica presence (N=95	50)										
0- No fluke	115 (63)	119 (77)	380 (75)	103 (99)		717 (76)					
1- Actual presence	22 (12)	13 (9)	82 (16)	1 (1)		118 (12)					
2- Historical presence	47 (25)	22 (14)	46 (9)	0 (0)		115 (12)					

596

598 **Table 3.** 

599 Cattle level variables in multilevel multinomial models predicting cattle carcasses intensity of

600 Ostertagia spp. lesions (Model 1), adult rumen fluke presence (Model 2) and liver fluke

601 lesions (Model 3).

		Model 1	: Ostertag	<i>ia spp</i> . lesi	ons	Model 2 :	adult rum	en fluke	Model 3 : liver fluke lesions				
		(933 Cat	ttle)			presence	(936 Cattle	)	(951 Cattle)				
		None	<100	100-	>1000	None	<b>≤100</b>	> 100	None	Moderate	Severe		
Variables	Categories	N (%)	N (%)	1000 N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)		
Breed	Pure dairy	30 (29)	76 (30)	97 (41)	135 (40)	263 (37)	38 (32)	41 (37)	255 (38)	64 (29)	20 (41)		
	Pure beef	9 (9)	22 (9)	21 (9)	26 (8)	54 (8)	14 (12)	10 (9)	55 (8)	25 (11)	3 (6)		
	Beef X	65 (62)	145 (58)	106 (44)	152 (46)	354 (51)	59 (50)	54 (49)	331 (49)	123 (56)	24 (49)		
	Dairy X	0 (0)	8 (3)	14 (6)	19 (6)	29 (4)	7 (6)	5 (5)	30 (5)	9 (4)	2 (4)		
Category*	Cow	16 (15)	48 (19)	43 (18)	65 (19)	135 (19)	17 (14)	23 (20)	94 (14)	62 (28)	28 (57)		
	Heifer	12 (11)	36 (14)	34 (14)	64 (19)	112 (16)	23 (19)	12 (11)	116 (17)	32 (14)	6 (12)		
	Steer	65 (62)	136 (54)	126 (52)	186 (55)	361 (51)	75 (63)	76 (68)	367 (54)	128 (57)	14 (29)		
	Young Bull	12 (11)	33 (13)	37 (15)	20 (6)	97 (14)	4 (3)	1(1)	102 (15)	1(1)	1 (2)		
Age (Month)	<24	29 (28)	53(21)	59 (25)	41 (12)	165 (23)	13 (11)	4(1)	166 (25)	17 (8)	2 (4)		
-	24-30	46 (44)	91 (36)	82 (34)	137 (41)	264 (38)	58 (48)	34 (11)	273 (40)	79 (35)	7 (14)		
	>30	30 (28)	110 (43)	99 (41)	158 (47)	277 (39)	49 (41)	274 (88)	240 (35)	129 (57)	40 (82)		
Month	March	46 (44)	62 (25)	72 (30)	111 (33)	238 (34)	35 (29)	16 (14)	232 (34)	57 (25)	8 (16)		
	June	33 (31)	74 (29)	49 (20)	74 (22)	173 (24)	31 (26)	27 (24)	166 (24)	58 (26)	8 (16)		
	January	23 (22)	72 (28)	60 (25)	59 (18)	148 (21)	26 (22)	40 (36)	148 (22)	53 (24)	21 (43)		
	October	3 (3)	46 (18)	59 (25)	92 (27)	147 (21)	28 (23)	29 (26)	133 (20)	57 (25)	12 (25)		
Altitude (m)	≤60	-	-	-	-	194 (31)	49 (45)	43 (48)	187 (30)	79 (43)	18 (46)		
	>60	-	-	-	-	438 (69)	60 (55)	46 (52)	432 (70)	104 (57)	21 (54)		
O <sup>(#)</sup>	None	-	-	-	-	89 (13)	5 (4)	6 (5)	79 (12)	23 (11)	1 (2)		
	Present	-	-	-	-	613 (87)	111 (96)	106 (95)	576 (88)	193 (89)	44 (98)		
RF <sup>(#)</sup>	None	89 (89)	180 (71)	185 (77)	248 (74)	-	-	-	540 (82)	128(60)	27 (59)		
	Present	11 (11)	74 (39)	55 (33)	88 (26)	-	-	-	120 (18)	85 (40)	19 (41)		
F. hepatica <sup>(#)</sup>	None	79 (77)	170 (69)	158 (68)	248 (75)	540(78)	73 (62)	47 (44)	-	-	-		
	Present	24 (23)	77 (31)	76 (32)	84 (25)	155 (22)	44 (38)	60 (56)	-	-	-		

Mature bull excluded; \* O = Ostertagia spp. lesions; RF= presence of adult rumen fluke; F. hepatica =
 presence of liver fluke

- 605 **Table 4.**
- 606 Final multilevel multinomial models predicting cattle carcasses intensity of *Ostertagia spp.* lesions (Model 1), adult rumen fluke presence
- 607 (Model 2) and liver fluke lesions (Model 3), containing cow and herd as random effects and general demographic and carcass parameters
- as fixed effects with respectively no pathology (Model 1 and 3) and no worm (Model 2) as a reference [CCW = Cold Carcass Weight; X
- 609 = Cross].

Variables		Mode	l 1: Ostertagi	lesions		Model	2: Adult ru	men fluk	e presence	Mode	el 3: liver flu	ıke lesio	ons		
		(154 Herds, 933 cattle, 2697 Obs.) <sup>a,b,c</sup>							lerds, 936 ca	ttle, 158	84 Obs.) <sup>a,b,c</sup>	(153 Herds, 951 cows, 1584 Obs.) <sup>a,b,c</sup>			
Categories		<100		100-1	000	>100	0	≤100		>100		Mode	erate	Sever	·e
		O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.
Breed	Pure	Baseli	ne					Baselir	ie			Basel	ine		
	Pure beef	1.40	0.86-2.27	0.79	0.46-1.34	0.69	0.44-1.09	1.87	0.87-4.00	1.73	0.69-4.35	1.99	1.00-3.96	0.92	0.20-4.32
	Beef X	1.14	0.83-1.55	0.49*	0.35-0.69	0.45*	0.34-0.61	0.91	0.53-1.56	1.13	0.64-2.02	2.30*	1.46-3.64	3.18*	1.42-7.11
	Dairy X	7.29*	4.48-11.88	8.63*	5.16-14.42	6.20*	3.92-9.78	2.03	0.80-5.11	1.01	0.29-3.51	1.03	0.36-2.96	0.79	0.09-7.33
Category	Cow	Baseli	ne					Baselir	ie			Basel	ine		
	Heifer	2.16*	1.35-3.45	4.34*	2.52-7.46	7.11*	4.38-11.53	2.55*	1.07-6.12	2.15	0.81-5.70	0.43*	0.21-0.86	0.08*	0.01-0.41
	Steer	1.24	0.84-1.85	2.06*	1.32-3.20	2.54*	1.72-3.75	2.51*	1.20-5.28	3.95*	1.91-8.18	0.64	0.38-1.10	0.13*	0.05-0.33
	Young	2.08	0.99-4.37	3.15*	1.48-6.66	2.01	0.95-4.22	0.92	0.21-3.93	1.21	0.11-13.94	0.04*	0.01-0.38	0.14	0.01-2.80
Age (months	) <24	Baseli	ne					Baselin	ie			Basel	ine		
	24-30	1.59	0.93-2.71	1.60	0.95-2.70	2.82*	1.70-4.67	1.50	0.68-3.31	3.08	0.88-10.72	1.07	0.54-2.10	0.66	0.06-6.78
	>30	2.72*	1.56-4.75	2.27*	1.31-3.94	4.40*	2.59-7.46	1.35	0.57-3.16	5.48*	1.56-19.21	1.87	0.92-3.80	3.75	0.43-32.94
Month	March	0.08*	0.05-0.12	0.07*	0.04-0.10	0.06*	0.04-0.08	Baselir	ie			Basel	ine		
	June	0.11*	0.07-0.16	0.05*	0.03-0.08	0.04*	0.03-0.05	1.24	0.69-2.23	2.32*	1.13-4.75	1.09	0.66-1.82	1.25	0.36-4.34
	January	Baseli	ne					1.11	0.58-2.12	2.82*	1.34-5.92	1.75*	1.04-2.94	3.20*	1.16-8.86
	October	0.20*	0.13-0.29	0.09*	0.06-0.14	0.06*	0.04-0.09	2.01*	1.08-3.72	4.45*	2.12-9.38	2.06*	1.21-3.50	1.81	0.63-5.17
Altitude (m)	≤60	-	-	-	-	-	-	Baselir	ne			Basel	ine		

	>60	-	-	-	-	-	-	0.58*	0.36-0.92	0.44*	0.26-0.72	0.56*	0.38-0.82	0.63	0.29-1.33
O <sup>(*)</sup>	None	-	-	-	-	-	-	Baselin	e			Basel	ine		
	Present	-	-	-	-	-	-	2.40	0.93-6.18	1.51	0.58-43.94	0.90	0.49-1.65	3.42	0.41-28.22
<b>RF</b> <sup>(*)</sup>	None	Baseli	ne					-	-	-	-	Basel	ine		
	Present	3.01*	2.27-4.00	1.92*	1.38-2.67	2.27*	1.70-3.03	-	-	-	-	2.71*	1.83-4.02	4.08*	1.95-8.50
F. hepatica <sup>(*)</sup>	None	Baseli	ne					Baselin	e			-	-	-	-
	Present	1.06	0.80-1.41	1.57*	1.13-2.19	0.92	0.68-1.25	1.79*	1.08-2.96	3.21*	1.93-5.34	-	-	-	-

610 OR - Odds Ratio; 95% CI - 95% Confidence Interval; \* O = Ostertagia spp. lesions; RF= presence adult rumen fluke; F. hepatica = presence of liver fluke

Table 5.

Final multilevel linear regression models predicting impacts on carcass parameters, respectively Cold Carcass Weight (Model 1), Conformation (Model 2) and Fat classification (Model 3), containing cow and herd as random effects and cattle parameters and helminths scoring as fixed effects [CCW = Cold Carcass Weight; Obs. = Observations; \* =

Significant].

		Mod	el 1: CCW		Мо	del 2: Co	onformation	Moo	Model 3: Fat classification				
		(115	Herds, 756	cattle, 618 Obs.)	(11	5 Herds,	756 cattle, 709 Obs.	) (115	Herds, 7	56 cattle, 630 O			
Fixed effects													
Variables	Categories	Ν	β	95% C.I.	Ν	β	95% C.I.	Ν	β	95% C.I.			
Intercept (SE)			295.35 (12	2.49)		14.15 (2	2.25)		28.30(1.6	53)			
Helminth Inf. (*#	<sup>b</sup> None	64	Baseline		64	Baseline	e	64	Baseline				
	O only	401	-30.58*	-50.92;-10.24	401	1.13	-0.53;2.78	401	-3.28*	-5.56;-1.00			
	<b>RF</b> only	6	-50.34*	-88.50;-12.18	6	2.41	-1.27;6.09	6	-5.49*	-10.28;-0.69			
	LF only	11	-20.39	-50.76;9.98	11	-3.65*	-6.98;-0.32	11	-1.41	-5.71;2.89			
	O-RF	102	-39.99*	-73.09;-6.88	102	-1.69	-4.36;0.98	102	-1.72	-5.57;2.14			
	O-LF	80	-22.94	-52.89;7.01	80	-1.26	-3.65;1.12	80	-0.35	-3.91;3.21			
	RF-LF	4	-32.41	-73.06;8.24	4	3.48	-0.66;7.64	4	-4.85	-10.19;0.49			
	<b>O-RF-LF</b>	57	-48.28*	-88.35;-8.21	57	-1.27	-4.68;2.14	57	-3.81	-8.61;0.99			
<b>Random effects</b>													
	Level		Variance	SE		Varianc	e SE		Variance	SE			
	Herd		561.42	101.81		2.31	0.68		4.45	1.26			
	Cattle		844.80	56.10		13.34	0.803		20.98	1.34			

<sup>\*</sup> Breed, category, age, CCW, conformation, fat, month and altitude were included in model as confounders, and results presented adjusted for these variables; <sup>#</sup> O = Ostertagia spp. lesions; RF= presence of adult rumen 

fluke; LF= liver fluke lesions

1	Tables
2	
3	Table 1.
4	Cattle median [p25-p75] age, cold carcass weight (CCW), carcass conformation and fat
5	classifications and percentage of liver condemnations by category (N=972).
6	
7	Table 2.
8	Stratification of abomasa, reticulorumen and livers scoring percentages by cattle category
9	(N=972).
10	
11	Table 3.
12	Cattle level variables in multilevel multinomial models predicting cattle carcasses intensity of
13	Ostertagia spp. lesions (Model 1), adult rumen fluke presence (Model 2) and liver fluke
14	lesions (Model 3).
15	
16	Table 4.
17	Final multilevel multinomial models predicting cattle carcasses intensity of Ostertagia spp.
18	lesions (Model 1), adult rumen fluke presence (Model 2) and liver fluke lesions (Model 3),
19	containing cow and herd as random effects and general demographic and carcass parameters
20	as fixed effects with respectively no pathology (Model 1 and 3) and no worm (Model 2) as a
21	reference [CCW = Cold Carcass Weight; X = Cross].
22	
23	Table 5.

Final multilevel linear regression models predicting impacts on carcass parameters, respectively Cold Carcass Weight (Model 1), Conformation (Model 2) and Fat classification (Model 3), containing cow and herd as random effects and cattle parameters and helminths scoring as fixed effects [CCW = Cold Carcass Weight; Obs. = Observations; \* = Significant].

# **Table 1.**

Variables (N)	<b>Cows (193)</b>	Heifers (155)	Steers (518)	Young Bulls (106)
Age (Months)	79 [56-113]	29 [26-31]	29 [26-31]	14 [14-15]
CCW (Kg)	323 [283-346]	314 [290-334]	344 [307-384]	294 [267-334]
Conformation	$P^+ \left[ P^+ \text{-} O^+ \right]$	R [O <sup>+</sup> -R]	$O^{+}[O^{+}-R]$	$O^{+}[O^{+}-R]$
Fat classification	3 [2-4L]	4L [3-4L]	3 [3-4L]	2 [2-3]
Liver condemnation	31.6	12.9	14.1	9.4

# **Table 2.**

	Cows	Heifers	Steers	Young	Bulls	TOTAL
Ostertagia spp. lesion (N=9.	33)					
0- No lesion	16 (9)	12 (8)	65 (13)	12 (12)		105 (11)
<b>1-</b> ≤ 100	48 (28)	36 (25)	136 (26)	33 (32)		253 (27)
2-101-1000	43 (25)	34 (23)	126 (25)	37 (36)		240 (26)
3->1000	65 (38)	64 (44)	186 (36)	20 (20)		335 (36)
Adult rumen fluke presenc	e (N=936)					
0- No fluke	135 (77)	112 (76)	361 (70)	97 (95)		705 (75)
1- ≤100	17 (10)	23 (16)	75 (15)	4 (4)		119 (13)
2- > 100	23 (13)	12 (8)	76 (15)	1 (1)		112 (12)
Liver fluke lesion (N=951)						
0- No lesion	94 (51)	116 (75)	367 (72)	102 (98)		679 (72)
1- Moderate	62 (34)	32 (21)	128 (25)	1 (1)		223 (23)
2- Severe	28 (15)	6 (4)	14 (3)	1 (1)		49 (5)
F. hepatica presence (N=95	(0)					
0- No fluke	115 (63)	119 (77)	380 (75)	103 (99)		717 (76)
1- Actual presence	22 (12)	13 (9)	82 (16)	1 (1)		118 (12)
2- Historical presence	47 (25)	22 (14)	46 (9)	0 (0)		115 (12)

#### Table 3. 35

		Model 1: (933 Cat	0	<i>ia spp</i> . lesi	ons		adult rume (936 Cattle)		Model 3 : liver fluke lesions (951 Cattle)			
		None	<100	100- 1000	>1000	None	<b>≤100</b>	> 100	None	Moderate	Severe	
Variables	Categories	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	N (%)	
Breed	Pure dairy	30 (29)	76 (30)	97 (41)	135 (40)	263 (37)	38 (32)	41 (37)	255 (38)	64 (29)	20 (41)	
	Pure beef	9 (9)	22 (9)	21 (9)	26 (8)	54 (8)	14 (12)	10 (9)	55 (8)	25 (11)	3 (6)	
	Beef X	65 (62)	145 (58)	106 (44)	152 (46)	354 (51)	59 (50)	54 (49)	331 (49)	123 (56)	24 (49)	
	Dairy X	0 (0)	8 (3)	14 (6)	19 (6)	29 (4)	7 (6)	5 (5)	30 (5)	9 (4)	2 (4)	
Category*	Cow	16 (15)	48 (19)	43 (18)	65 (19)	135 (19)	17 (14)	23 (20)	94 (14)	62 (28)	28 (57)	
	Heifer	12 (11)	36 (14)	34 (14)	64 (19)	112 (16)	23 (19)	12 (11)	116 (17)	32 (14)	6 (12)	
	Steer	65 (62)	136 (54)	126 (52)	186 (55)	361 (51)	75 (63)	76 (68)	367 (54)	128 (57)	14 (29)	
	Young Bull	12 (11)	33 (13)	37 (15)	20 (6)	97 (14)	4 (3)	1 (1)	102 (15)	1 (1)	1 (2)	
	0	29 (28)	53(21)	59 (25)	41 (12)	165 (23)	13 (11)	4 (1)	166 (25)	17 (8)	2 (4)	
2	24-30	46 (44)	91 (36)	82 (34)		264 (38)	58 (48)	34 (11)	273 (40)	79 (35)	7 (14)	
	>30	30 (28)	110 (43)	99 (41)	158 (47)	277 (39)	49 (41)	274 (88)	240 (35)	129 (57)	40 (82)	
Month	March	46 (44)	62 (25)	72 (30)		238 (34)	35 (29)	16 (14)	232 (34)	57 (25)	8 (16)	
	June	33 (31)	74 (29)	49 (20)	74 (22)	173 (24)	31 (26)	27 (24)	166 (24)	58 (26)	8 (16)	
	January	23 (22)	72 (28)	60 (25)	59 (18)	148 (21)	26 (22)	40 (36)	148 (22)	53 (24)	21 (43)	
	October	3 (3)	46 (18)	59 (25)	92 (27)	147 (21)	28 (23)	29 (26)	133 (20)	57 (25)	12 (25)	
Altitude (m)	≤60	-	-	-	-	194 (31)	49 (45)	43 (48)	187 (30)	79 (43)	18 (46)	
-	>60	-	-	-	-	438 (69)	60 (55)	46 (52)	432 (70)	104 (57)	21 (54)	
O <sup>(#)</sup>	None	-	-	-	-	89 (13)	5 (4)	6 (5)	79 (12)	23 (11)	1 (2)	
	Present	-	-	-	-	613 (87)	111 (96)	106 (95)	576 (88)	193 (89)	44 (98)	
<b>RF</b> <sup>(#)</sup>	None	89 (89)	180 (71)	185 (77)	248 (74)	-	-	-	540 (82)	128(60)	27 (59)	
	Present	11 (11)	74 (39)	55 (33)	88 (26)	-	-	-	120 (18)	85 (40)	19 (41)	
F. hepatica (#)	None	79 (77)		. ,	· · ·	540(78)	73 (62)	47 (44)	-	-	-	
-	Present	24 (23)	77 (31)	76 (32)	84 (25)	155 (22)	44 (38)	60 (56)	-	-	-	

Mature bull excluded; " O = Ostertagia spp. lesions; RF= presence of adult rumen fluke; F. hepatica =

36 37 presence of liver fluke

# **Table 4.**

Variables Categories								Model 2: Adult rumen fluke presence (153 Herds, 936 cattle, 1584 Obs.) <sup>a,b,c</sup>				Model 3: liver fluke lesions (153 Herds, 951 cows, 1584 Obs.) <sup>a,b,c</sup>			
Categories		<100		100-1	.000	>100	0	<b>≤100</b>		>100		Mode	erate	Sever	e
		O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.	O.R	95% C.I.
Breed	Pure	Baseli	ne					Baselir	ne			Basel	ine		
	Pure beef	1.40	0.86-2.27	0.79	0.46-1.34	0.69	0.44-1.09	1.87	0.87-4.00	1.73	0.69-4.35	1.99	1.00-3.96	0.92	0.20-4.32
	Beef X	1.14	0.83-1.55	0.49*	0.35-0.69	0.45*	0.34-0.61	0.91	0.53-1.56	1.13	0.64-2.02	2.30*	1.46-3.64	3.18*	1.42-7.11
	Dairy X	7.29*	4.48-11.88	8.63*	5.16-14.42	6.20*	3.92-9.78	2.03	0.80-5.11	1.01	0.29-3.51	1.03	0.36-2.96	0.79	0.09-7.33
Category	Cow	Baseli	ne					Baselir	ie			Basel	ine		
	Heifer	2.16*	1.35-3.45	4.34*	2.52-7.46	7.11*	4.38-11.53	2.55*	1.07-6.12	2.15	0.81-5.70	0.43*	0.21-0.86	0.08*	0.01-0.41
	Steer	1.24	0.84-1.85	2.06*	1.32-3.20	2.54*	1.72-3.75	2.51*	1.20-5.28	3.95*	1.91-8.18	0.64	0.38-1.10	0.13*	0.05-0.33
	Young	2.08	0.99-4.37	3.15*	1.48-6.66	2.01	0.95-4.22	0.92	0.21-3.93	1.21	0.11-13.94	0.04*	0.01-0.38	0.14	0.01-2.80
Age (months)	<24	Baseli	ne					Baselin	ie			Basel	ine		
	24-30	1.59	0.93-2.71	1.60	0.95-2.70	2.82*	1.70-4.67	1.50	0.68-3.31	3.08	0.88-10.72	1.07	0.54-2.10	0.66	0.06-6.78
	>30	2.72*	1.56-4.75	2.27*	1.31-3.94	4.40*	2.59-7.46	1.35	0.57-3.16	5.48*	1.56-19.21	1.87	0.92-3.80	3.75	0.43-32.94
Month	March	0.08*	0.05-0.12	0.07*	0.04-0.10	0.06*	0.04-0.08	Baselin	ie			Basel	ine		
	June	0.11*	0.07-0.16	0.05*	0.03-0.08	0.04*	0.03-0.05	1.24	0.69-2.23	2.32*	1.13-4.75	1.09	0.66-1.82	1.25	0.36-4.34
	January	Baseli	ne					1.11	0.58-2.12	2.82*	1.34-5.92	1.75*	1.04-2.94	3.20*	1.16-8.86
	October	0.20*	0.13-0.29	0.09*	0.06-0.14	0.06*	0.04-0.09	2.01*	1.08-3.72	4.45*	2.12-9.38	2.06*	1.21-3.50	1.81	0.63-5.17
Altitude (m)	≤60	-	-	-	-	-	-	Baselin	ie			Basel	ine		
	>60	-	-	-	-	-	-	0.58*	0.36-0.92	0.44*	0.26-0.72	0.56*	0.38-0.82	0.63	0.29-1.33
O <sup>(*)</sup>	None	-	-	-	-	-	-	Baselir	ie			Basel	ine		
	Present	-	-	-	-	-	-	2.40	0.93-6.18	1.51	0.58-43.94	0.90	0.49-1.65	3.42	0.41-28.22
<b>RF</b> <sup>(*)</sup>	None	Baseli	ne									Baseline			
	Present	3.01*	2.27-4.00	1.92*	1.38-2.67	2.27*	1.70-3.03	-	-	-	-	2.71*	1.83-4.02	4.08*	1.95-8.50
F. hepatica <sup>(*)</sup>	None	Baseli	ne					Baselir	ie			-	-	-	-
•	Present	1.06	0.80-1.41	1.57*	1.13-2.19	0.92	0.68-1.25	1.79*	1.08-2.96	3.21*	1.93-5.34	-	-	-	-

40 OR - Odds Ratio; 95% CI - 95% Confidence Interval; \* O = Ostertagia spp. lesions; RF= presence adult rumen fluke; F. hepatica = presence of liver fluke

## Table 5.

	Model 1: CCW (115 Herds, 756 cattle, 618 Obs.)					el 2: Confo Herds, 756	ormation 5 cattle, 709	Model 3: Fat classification (115 Herds, 756 cattle, 630 Obs.)			
Fixed effects											
Variables	Categories	Ν	β	95% C.I.	Ν	β	95% C.I.	Ν	β	95% C.I.	
Intercept (SE)			295.35(12	2.49)		14.15(2.25)			28.30(1.6	(3)	
Helminth Inf.	None	64	Baseline		64	Baseline		64	Baseline		
726 IK	O only	401	-30.58*	-50.92;-10.24	401	1.13	-0.53;2.78	401	-3.28*	-5.56;-1.00	
	RF only	6	-50.34*	-88.50;-12.18	6	2.41	-1.27;6.09	6	-5.49*	-10.28;-0.69	
	LF only	11	-20.39	-50.76;9.98	11	-3.65*	-6.98;-0.32	11	-1.41	-5.71;2.89	
	O-RF	102	-39.99*	-73.09;-6.88	102	-1.69	-4.36;0.98	102	-1.72	-5.57;2.14	
	O-LF	80	-22.94	-52.89;7.01	80	-1.26	-3.65;1.12	80	-0.35	-3.91;3.21	
	RF-LF	4	-32.41	-73.06;8.24	4	3.48	-0.66;7.64	4	-4.85	-10.19;0.49	
	<b>O-RF-LF</b>	57	-48.28*	-88.35;-8.21	57	-1.27	-4.68;2.14	57	-3.81	-8.61;0.99	
Random effects	6										
	Level		Variance	SE		Variance	SE		Variance	SE	
	Herd		561.42	101.81		2.31	0.68		4.45	1.26	
	Cattle		844.80	56.10		13.34	0.803		20.98	1.34	

\* Breed, category, age, CCW, conformation, fat, month and altitude were included in model as confounders, and results presented adjusted for these variables; <sup>#</sup> O = *Ostertagia spp*. lesions; RF= presence of adult rumen fluke; LF= liver fluke lesions