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1 **Ecological drivers influence the distributions of two cryptic**
2 **lineages in an earthworm morphospecies**

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26 ABSTRACT

27 Substantial genetic diversity exists within earthworm morphotypes, such that traditional
28 species designations may be incomplete. It is, however, currently not known whether these
29 different genetic variants show ubiquity or specialty in their distribution across separated sites
30 subject to different climatic, biotic or soil physicochemical factors. Here we report on the results
31 of a survey in which individuals of the *Lumbricus rubellus* morphotype, a species known to
32 comprise two deeply divergent genetic lineages in England and Wales, were sampled from 26
33 plots. Sequences from the mitochondrial cytochrome oxidase I gene were used to distinguish
34 lineages for 787 individuals. In conjunction, a range of geographic, climatic, biotic and soil
35 physicochemical variables were also collected for each locality.

36

37 Genotyping indicated that Lineage A was more common than Lineage B, comprising 58% of
38 the collected *L. rubellus*. Six site populations comprised only Lineage A, while only a single
39 site comprised entirely Lineage B. The remaining 20 sites containing both lineages. A
40 multivariate ordination of site variables identified major difference between sites were
41 associated with low pH, organic-rich soils in Western wet upland areas and pollutant levels
42 associated with sites in the South. Earthworm genotype (as proportion of Lineage A) was not
43 correlated with either of these major environmental axes. When individual variables of soil pH
44 and the percentage of soil organic matter, which are known to be key driver of soil species
45 distributions, were investigated as single variables significant relationship with lineage
46 frequency were found. Soil organic matter content was significantly negatively correlated with
47 Lineage A proportion, while pH was significantly positively correlated. This lineage preference
48 may be related to lineage metabolism and/or behavioral differences.

49

50 Measurement of tissue metal concentrations in worms from 17 sites identified a significant site
51 effect in all cases, but a lineage effect only for arsenic (higher Lineage B). Tissue arsenic
52 concentrations varied between lineages, supporting previous observations that there are
53 differences in the way the two lineages have adapted to manage exposure to this metalloid.

54

55 Keywords: Biogeography, Earthworm, Cryptic species, pH, Soil organic matter

56 1. INTRODUCTION

57 Soils contain a wealth of invertebrate biodiversity recognised for their important contributions
58 to ecological processes (Bardgett and van der Putten, 2014; Fitter et al., 2005; Giller, 1996).
59 One key group of species are the “ecosystem engineers”: those organisms that modify the
60 physical state of the soil and resource availability for other species. Earthworms are known as
61 a key group of ecosystem engineers in many habitats. They perform a range of physical
62 (aeration, bioturbation, litter fragmentation) and biological (microbial interactions, exudate
63 production) roles in soil (Blouin et al., 2013; Lavelle et al., 1997; Sackett et al., 2013; Umarov
64 et al., 2008). Because of their functional importance, earthworms have emerged as a major
65 taxon for biomonitoring and biomarker assessments of human induced pressures on soil
66 communities (Cluzeau et al., 2012; Rutgers et al., 2009).

67

68 As soil invertebrate species, including earthworms, have been shown to be sensitive to a
69 range of land use change and pollution impacts (Bundy et al., 2007; Cluzeau et al., 2012),
70 different soil taxa have become a natural focus for research on the relationships between
71 environmental pressures, biodiversity and soil functioning (Bartlett et al., 2010; Leveque et al.,
72 2015; Rutgers et al., 2016). For community studies, a major constraint relates to current
73 uncertainties in earthworm taxonomy. Traditionally earthworm identification has relied on
74 morphology, but the paucity of suitable local keys and problems with application to juveniles
75 has also recently encouraged the use of molecular methods (Dominguez et al., 2015;
76 Emerson et al., 2011; Klarica et al., 2012). These genotyping studies have begun to challenge
77 current understanding of diversity through the identification of genetically distinct cryptic
78 lineages within previously established morphospecies.

79

80 Earthworm species in which cryptic lineage diversity has to date been identified include
81 *Eisenia fetida/andrei* (Römbke et al., 2016), *Lumbricus terrestris* (James et al., 2010),
82 *Aporrectodea caliginosa* (PerezLosada et al., 2009), *Allolobophora chlorotica* (King et al.,
83 2008), *Amyntas gracilis / Amyntas cortici* (Novo et al., 2015) and *Lumbricus rubellus*. For

84 *L. rubellus*, genotyping studies based on mitochondrial cytochrome oxidase I and II markers
85 have identified as many as 6 cryptic lineages across Europe (Giska et al., 2015), two of which
86 are found in the UK (Andre et al., 2010; Kille et al., 2013). The two UK lineages have 10-15%
87 divergence for the mitochondrial COI and COII sequences. While this implies they may
88 actually be cryptic species, recent analysis of multiple nuclear markers using RADseq has not
89 supported this interpretation, instead suggesting that different *L. rubellus* lineages may
90 actually correspond to a single highly polymorphic species (Giska et al., 2015). Comparative
91 studies of the two lineages in the UK have, nonetheless, identified physiological differences
92 between them, including variation in pheromone production (Jones et al., 2016), maturation
93 time (Anderson et al., 2013), metabolic profiles (Liebeke et al., 2014), mechanism of arsenic
94 adaptation (Kille et al., 2013), trace element metabolism (Andre et al., 2010), and microbiome
95 complement (Pass et al., 2015).

96
97 Despite known biological differences, the extent to which differences in distribution and
98 physiology are related to different geographical, climate and soil physicochemical preferences
99 between the two known UK lineages of *L. rubellus* is not established. The two lineages found
100 co-occur at some, but not all, sites meaning that they have some likely niche divergence that
101 facilitates coexistence (Andre et al., 2010; Giska et al., 2015; Kille et al., 2013). We aim to
102 better understand the nature of the spatial and geochemical drivers of lineage relative
103 abundance, and so here we test the hypothesis that the site distribution of the two cryptic *L.*
104 *rubellus* lineages is based on one or more geographical, climatic, physiochemical or biotic
105 drivers. We collected and genotyped morphotype *L. rubellus* at multiple well-characterized
106 sites that differed in their properties to investigate the relationships that determine lineage
107 distributions. Tissue metal concentrations were also measured to assess if trace metal levels
108 could also influence distributions, as could be the case if the two lineages had different
109 sensitivity to specific contaminants.

110 2. METHODS

111 2.1 Site selection

112 Twenty six sites located across England and Wales (Fig. 1) were visited between four times
113 (for Devon Great Consols Mine and Control, Shipham Mine and Control, Cwmystwyth Mine
114 and Control) and a single visit (for Porton Down, Parys Mountain, Castell, Clydach, Roman
115 Gravel, Didcot) over four separate sampling events from Spring 2011 to Spring 2014. The
116 chosen sites were selected to capture a range of the habitats and soil conditions under which
117 morphotype *L. rubellus* can be collected. Land-uses covered included arable systems,
118 broadleaf woodland, rough grassland and improved pasture habitats. Sites included both
119 mineral and organic soils, although not true peats.

120

121 To allow the role of soil geochemistry and pollution status on lineage distribution to be
122 addressed, sites of different known pollution history were sampled. Sites corresponded to
123 three groups with respect to past land-use and associated expected contamination level.
124 These were: 1) sites with no known pollution source (Unpolluted); 2) sites near to industrial
125 facilities expected to be characterised by moderate pollution (Industrial polluted); and 3) sites
126 at abandoned mining sites that can be expected to have high pollution (Mine polluted). For
127 expected polluted sites from categories 2 and 3, a local control site was also sampled. This
128 reference site was located outside of the area that was expected to be strongly influenced by
129 the main pollution source and so was on soil expected to contain regional background pollutant
130 concentrations.

131

132 2.2 Site geographical, biological and soil physiochemical characterisation

133 To allow the assessment of environmental drivers relating to lineage distribution, we used both
134 publically available resources as well as our own analyses to gather data on each sites. Site
135 geographical locations were collected as Easting and Northings from
136 www.gridreferencefinder.com and site altitudes from www.freemaptools.com/elevation-

137 [finder.htm](#). A series of site climate conditions were also assembled from
138 www.metoffice.gov.uk/. These were: annual average maximum temperature, annual average
139 minimum temperature, average January minimum temperature, average July minimum
140 temperature, average annual rainfall, average annual rain days and average annual frost
141 days. Initial visits to each site recorded main land-use (arable, broadleaf woodland, rough
142 grassland and improved pasture) and where present the average sward height of vegetation
143 at collection locations. The site was identified according to the level of shade (open, part
144 shaded, shaded) and the presence of livestock was noted.

145

146 An initial site survey identified points on the site where morphospecies *L. rubellus* could be
147 found. Thereafter all collections were focussed on these locations. For any one sampling event
148 at each site, between 6 and 25 fully clitellate *L. rubellus* were collected by digging and hand-
149 sorting from the soil to 20 cm depth. Generally the required number of worms could be
150 collected within a reasonable search period (approximately 2 h duration). There were,
151 however, some locations where this was not possible for particular sampling events. Climate
152 factors (notably dry soils), low frequency of adults in the population or the requirement to limit
153 site damage caused by digging were the major constraints. During collection, the presence of
154 other earthworm morphospecies was noted. Only common species were recorded (>5
155 individuals observed). In total 10 other species were found: *Aporrectodea caliginosa*,
156 *Aporrectodea rosea*, *Aporrectodea longa*, *Allolobophora chlorotica*, *Lumbricus castaneus*,
157 *Dendrobaena rubida*, *Lumbricus terrestris*, *Lumbricus festivus*, *Octolasion cyaneum*, and
158 *Octolasion tyrtaeum tyrtaeum*. At the end of sampling, the *L. rubellus* collected were washed
159 and blotted dry on-site and then snap frozen in liquid nitrogen before being transferred to the
160 laboratory under dry ice storage.

161

162 Triplicate soil samples from surface to 5 cm depth were collected from each site collection
163 location. All soil samples were oven dried at 80°C to constant weight and then sieved through

164 a 2 mm mesh to remove large roots and stones. Total concentrations of aluminium, arsenic,
165 barium, cadmium, cobalt, chromium, copper, iron, lead, manganese, mercury, molybdenum,
166 nickel, selenium, titanium, vanadium, zinc, calcium and total phosphorous were determined in
167 a 1 g sample of this processed soil following an aqua regia digestion protocol (Arnold et al.,
168 2008; Emmett et al., 2010; Spurgeon et al., 2008). Digests were subsequently analysed on a
169 Perkin Elmer Optima 7300 DV inductively coupled plasma optical emission spectrometry
170 instrument. For quality control, an in house reference traceable to BCR-143R (Commission of
171 the European Communities, Community Bureau of Reference) was included with each batch
172 of digestions. Measured concentrations were within 10% of certified values for all measured
173 elements with the exception of Al where the value was 55%. Organic matter content of each
174 soil sample was measured by proxy using loss on ignition following combustion at 500°C
175 (Rowell, 1994) and soil pH was quantified by electrode from a 1:2.5 volume soil:water mix (i.e.
176 1 volume soil with 2.5 volumes water added)(International Organisation for Standards, 2005).

177

178 *2.3 Lineage assignment by mitochondrial cytochrome oxidase I (COI) sequencing*

179 DNA was extracted from ~10 mg of frozen tissue (taken from the tail of each individual using
180 a scalpel) by automated DNA extraction using a Nucleplex Plants Tissues DNA Extraction
181 Kit (Nucleplex, Manchester, UK). After DNA quantification using Nanodrop (Thermo
182 Scientific, Willmington, DE), polymerase chain reaction amplification of the COI gene was
183 conducted using a set of established forward (GGTCAACAAATCATAAAGATATTGG) and
184 reverse (TAAACTTCAGGGTGACCAAAAAATCA) primers (Folmer et al., 1994) amplified after
185 5 minutes at 95°C over 40 cycles of 30 sec 95°C, 30 seconds 48°C and 60 seconds 48°C. A
186 sub-set of all PCR products were checked by gel electrophoresis to ensure successful
187 amplification and purified for sequencing using 0.25 U each of Exonuclease I and Shrimp
188 Alkaline Phosphatase (NEB, Hitchin, UK), incubated at 37°C for 45 minutes and 80 °C for 15
189 minutes. Purified PCR products were then sequenced as in Andre et al. (2010), using ABI
190 PRISM® BigDye v3.1 Terminator Sequencing technology (Applied Biosystems, USA).

191

192 Sequences were aligned and trimmed for tree construction using the Maximum Likelihood
193 method and General Time Reversible substitution model with a gamma distribution in Mega
194 v5.01. Sequences for *L. rubellus* associated with specific mitochondrial lineages already
195 documented in the UK were incorporated into the analysis as anchor sequences (Anderson
196 et al., 2013), with sequences for *L. terrestris*, *L. festivus* and *L. castaneus* included as an out-
197 group. Tree topology was supported by bootstrap analyses over 1000 iterations. Individuals
198 that showed a close relationship with one of the two previously identified UK *L. rubellus*
199 lineages were identified from the analysis. Any individuals showing intermediate status
200 resulting from probable sequencing errors were excluded from further analysis.

201

202 *2.4 Earthworm tissue trace element concentrations*

203 Earthworm tissues from 494 individuals taken from a sub-set of 17 sites (Alice Holt ECN
204 Control, Avonmouth Control, Avonmouth Incinerator, Avonmouth Savalco, Cwmystwyth
205 control, Cwmystwyth mine, Devon Great Consols Control, Devon Great Consols Mine,
206 Drayton ECN Control, Port Talbot Control, Port Talbot blast furnace, Porton Down ECN,
207 Scunthorpe blast furnace, Scunthorpe Control, Shipham control, Shipham mine, Snowdown
208 ECN control) were prepared for analysis (nb samples from remaining sites were lost due to
209 storage issues). These samples were analysed for tissue Al, As, Cd, Co, Cr, Cu, Fe, Mn, Mo,
210 Ni, Pb, Se, Sr and Zn concentrations. Whole earthworms, after tail removal for DNA extraction,
211 were initially ground to powder under liquid nitrogen in a cryogenic mill. The powder was
212 freeze-dried and a 100 mg sample digested with 10 ml of 70% HNO₃ (Ultrapure) at 200°C for
213 15 minutes within a microwave vessel. Samples were run as two batches on a Perkin Elmer
214 DRCII ICP-MS. Each batch included multiple certified reference material samples for TORT-
215 2 and DOLT-4 (National Research Council, Canada). Certified values for reference materials
216 corroborated well with measured values. Average recovery was 91% (range 85% for Se to
217 110% for Pb) in the first batch of samples and 94.8% (range 53.9 for Al to 129% for Se) in the
218 second batch. Recoveries of only two metals, Al and Se, were outside 80% of certified values

219 for any run, with 19 of 27 determinations within 10%. With systematic bias absent, acquired
220 data can be used for statistical processing without requirement for recovery correction.

221

222 *2.5 Data handling and statistical analysis*

223 The number of *L. rubellus* returning COI sequences that were closely related to reference
224 sequences from previously collected lineage A and Lineage B individuals were counted for
225 each study site. These were calculated as proportions before being logit transformed as the
226 most appropriate transformation for biological proportion data (Warton and Hui, 2011), with
227 value of zero and one modified by addition and subtraction of half of the lowest proportion
228 respectively. Environmental drivers were established as either categorical (e.g. site type, site
229 shading, livestock presence/absence, earthworm species presence/absence) or as
230 continuous measured variables. The values for soil metal concentrations were log transformed
231 to obtain a Gaussian distribution in accordance with established practice (Davies, 1989).

232

233 Relationships amongst site geographical, climate and soil variables (after appropriate
234 transformation) were initially investigated using principal component analysis in Minitab 14
235 (Minitab, PA, USA). This reduces the dimensionality of the original complex dataset in an
236 unsupervised fashion, by successively generating new axes (principal components), that are
237 the linear combinations of the original data that explain greatest overall variance. The principal
238 components (PCs) arising can be interpreted as ordinations representing a summary of
239 environmental factors. Pearson correlations between (logit transformed) proportions of
240 Lineage A individuals at each site and the individual principal component scores were then
241 calculated to investigate the relationships between ordinated site characteristics and
242 genotype. Based on this analysis and prior knowledge, possible individual primary driver
243 variables were identified that were in turn assessed for Pearson correlation with genotype.
244 Because of this key role, percentage organic matter content and soil pH were selected as
245 focus variables. Site and lineage effects on earthworm tissue log transformed trace element

246 concentrations were analysed using a mixed model based general linear model in Minitab 14.
247 Within the model, site and lineage were included as fixed variables, with sampling campaign
248 (1-4) as a random factor.

249 3. RESULTS

250 High quality *L. rubellus* COI sequences were obtained from DNA samples taken from 787
251 earthworms for assignment as either Lineage A or Lineage B individuals. The maximum
252 number of sequences from any one site was 73, from Cwmystwyth Mine, and the minimum 3,
253 from Avonmouth Control (Fig. 1). In total, 457 individuals were assigned as Lineage A, 58%
254 of the number collected. The remaining 330 (42%) were assigned as Lineage B. Eight sites
255 (Avonmouth Savalco, Avonmouth Incinerator, Clydach Smelter, Didcot Power Station, Dinas
256 Powys, Parys Mountain, Scunthorpe Control and Scunthorpe) had populations comprising
257 only Lineage A individuals. Seven sites (Avonmouth Control, Castell Mine, Cwmystwyth
258 Control, Cwmystwyth Mine, Drayton ECN Control, Shipham Control and Shipham Mine)
259 contained populations comprised largely Lineage B, although only Castell Mine was
260 exclusively B. All remaining sites had mixed lineage populations, although with more Lineage
261 A than B individuals.

262

263 The site geographical, physical and soil characteristics were analysed using principal
264 component analysis. The first PC explained 21.4% of total variance. A number of parameters
265 were positively correlated with this axis, including soil % loss on ignition (LOI); soil log Fe, log
266 Co and log Al concentrations; some earthworm species; and site altitude and climate variables
267 including average rainfall and number of rain days. Negatively correlated variables included
268 soil pH; average July max temperature and average temperature; log Ca and log P
269 concentrations; and Easting (Fig 2). This first PC axis could therefore be interpreted as
270 representing a set of variables characterised by the presence of high organic matter, low pH
271 soils, associated with wetter and colder upland regions located mainly in the West of England
272 and Wales. The second PC axis explained a further 17% of variation, and was positively
273 associated with Northing and the weather variable of average frost days and average rain
274 days. Variables negatively associated with this axis included pollutant metal concentrations
275 such as log Pb, log Zn and log Cd concentrations (Fig 2). This axis can be interpreted as

276 representing a gradient of metal contamination of sites located primarily in the South of
277 England and Wales.

278

279 To assess if the site characteristics summarised by the two first PCs potentially act as drivers
280 of lineage distribution, the PC1 and PC2 scores were correlated with the (logit transformed)
281 Lineage A proportion at each location. This did not identify significantly relationships between
282 Lineage A proportion with site PC1 or PC2 score ($p=0.25$ and $p=0.074$ respectively). As sites
283 PC scores were not significant, we next went on to investigate if individual variables measured
284 are related to the relative frequency of *L. rubellus* lineages. Specifically we selected soil pH
285 and LOI for initial assessment, as these are established as drivers of patterns of diversity
286 (Griffiths et al., 2011; Raty and Huhta, 2003). Both variables were significantly correlated with
287 logit Lineage A proportion (soil % OM -0.529 , $p=0.005$; pH -0.392 , $p=0.048$). The nature of
288 these two relationships were summarised by locally weighted scatterplot smoother model fits.
289 These indicate a decline in proportion of Lineage A (higher logit transformed values) as site
290 soil % LOI increases from 0-20%, thereafter remaining constant. The model fits for pH
291 indicated an initial decline in the proportion of Lineage A individuals (higher logit transformed
292 values) as pH increases from 4.5 to 5, with, thereafter, an increase in frequency (lower logit
293 transformed values) where site pH increases from 5 to 7.5 (Fig. 4 a,b). Amongst other
294 measured variables, only log soil Ca concentrations (-0.487 , $P=0.012$) and the average annual
295 number of rain days (0.433 , $p=0.027$) were also significantly correlated with Lineage A
296 proportion. Both of these variables are, however, also significantly correlated with soil pH
297 (Annual rain days: -0.584 , $p=0.002$; log soil Ca concentration: -0.751 , $p<0.001$) making precise
298 attribution of cause challenging.

299

300 Separate univariate models were generated to analyse tissue metal concentrations in relation
301 to collection site and lineage. The collection site had a highly significant ($p<0.001$) influence
302 on tissue concentrations for all analysed trace elements. This is only to be expected, given
303 that the sites include locations with no history of local pollution, to highly contaminated

304 industrial and mine sites. Lineage was also a significant factor in the model for As ($p < 0.02$).
305 This difference is, however, based on only a relatively small difference in average tissue
306 concentrations between lineages across all sites. Thus, average concentrations in Lineage A
307 of 10.69 ($n=300$) was slightly lower than the average tissue arsenic concentrations of 11.7
308 mg/kg ($n=195$) for Lineage B, Hence although statistically significant, the absolute magnitude
309 of difference in tissue As concentrations between lineages is small. For all other analysed
310 metals, there was no significant effect of lineage on tissue concentration ($p > 0.05$).

311 4. DISCUSSION

312 Species distributions can be affected by a range of environmental drivers, including
313 physiological tolerances, dispersal constraints, biotic interactions and anthropogenic
314 influences (Dennis and Hellberg, 2010; Gaston, 2003). Among earthworms, species show
315 preference for certain habitats, for example common compost earthworm species such as
316 *Eisenia fetida*, *Perionyx excavatus* and *Eudrilus eugeniae* preferentially occupy organic matter
317 rich habitats associated with animal manure or composting vegetation (Edwards, 2004).
318 Further, some species also have preference for different soil physiochemical properties. For
319 example, Jaensch et al (2013) found differences in morphospecies preference across different
320 soil pH classes, with species such as *Allolobophora chlorotica*, *Aporrectodea rosea*,
321 *Aporrectodea longa* and *Lumbricus terrestris* preferring soils with pH >5.6, and *Dendrodrilus*
322 *rubida*, *Dendrobaena octaedra* and *L. rubellus* soils with pH <5.6.

323

324 The UK earthworm fauna is notably denuded, comprising only around 20-25 native species,
325 compared to about 180 species that are found in neighboring France (Bouché, 1972; Sims
326 and Gerard, 1985). The reduced earthworm fauna of the UK can be linked to its recent history
327 of glaciation and the severing of the land bridge to Europe that restricted earthworm
328 colonization after glacial retreat. This influence of quaternary glaciation is consistent with what
329 is known about the current distribution and genetic structure of a range of species across
330 Europe and the UK (Hewitt, 2000). Among UK earthworm species, the majority show a
331 widespread and cosmopolitan distribution (Boag et al., 1997; Carpenter et al., 2012; Rutgers
332 et al., 2016; Sims and Gerard, 1985). Habitat preferences are known, such as those for pH
333 and for organic rich habitats as discussed previously, however the spatial heterogeneity of
334 terrestrial habitats means that at coarse recording scales (e.g. 10 km² or even 1 km²), a
335 significant proportion of UK earthworm species may be present in any given sampling area
336 (e.g. a mixed land-use area subject to comprehensive earthworm sampling within different
337 vegetation stands and habitats).

338

339 Genetic marker studies have identified deeply divergent cryptic lineages within many common
340 UK earthworm morphospecies based on mitochondrial or nuclear genetic marker analysis. An
341 active debate currently surrounds the question of whether these cryptic lineages correspond
342 to cryptic species or highly polymorphic species variants (Blakemore et al., 2010; Giska et al.,
343 2015; King et al., 2008). In the specific case of *L. rubellus*, the presence of cryptic lineages is
344 established from studies conducted from measurement of highly divergent (13-15%)
345 sequences for both of the cytochrome oxidase I and II mitochondrial genes (Andre et al., 2010;
346 Donnelly et al., 2014; Kille et al., 2013). Pan-European studies have shown that at continental
347 scale, morphotype *L. rubellus* may comprise of 5 or more such deeply divergent lineages
348 (Giska et al., 2015), two of which were here found across sites in England and Wales (Fig 1).
349 Recently RADseq analysis suggests that cryptic *L. rubellus* lineages may represent a case of
350 a highly polymorphic single species rather than true cryptic species (Giska et al., 2015).
351 Nonetheless, previous studies of the lineage physiology have identified a number of
352 differential responses between lineages (as previously outlined notably for the two UK
353 lineages). For example, Jones et al. (2016) found that the two lineage were favorably attracted
354 to soils that had previously been worked by earthworm of their own rather than the alternative
355 lineage. These results suggests that pheromone attractants may allow mate selection in mix
356 populations,. such as those that are found at the majority of our sampled site. Such selection
357 has the potential to underpin lineage differences in habitat preference and, as a consequence,
358 different spatial distributions at local scale.

359

360 Earthworms are key ecosystem engineers for the role that play an important role in the
361 creating of the spatial structure and chemistry of the soil habitat through bioturbation, litter
362 degradation and nutrient cycling (Edwards, 2004; Lavelle et al., 1997; Liebeke et al., 2015).
363 The extent to which the divergent lineages of common earthworm species overlap in respect
364 of habitat preference will be an important determinant of morphospecies contributions to
365 different ecosystem processes across space and time. The analysis here suggests that, in the
366 case of the two UK lineages of *L. rubellus*, there are ecological drivers of distribution.

367 Individually, soil pH and % OM were both significant correlated with the proportions of *L.*
368 *rubellus* Lineage A (and conversely Lineage B) collected across the 26 sample sites. These
369 two measurement parameters were selected for particular focus because they are recognized
370 as important environmental drivers of the distribution of a number of soil taxa (Cassagne et
371 al., 2003; Griffiths et al., 2011; Raty and Huhta, 2003). Additionally, there are also correlations
372 with other climate and soil variable that are themselves know to influence soil pH through soil
373 geochemistry and leaching.

374

375 Different soil pH preferences have direct effects on earthworm traits including reproduction,
376 growth and survival (Baker and Whitby, 2003; Spurgeon et al., 2006; Van Gestel et al., 1992).
377 *L. rubellus* is tolerant of relatively low pH, being commonly (and even preferentially) found in
378 moderately acidic soils (Jaensch et al., 2013). Results here suggest that this cosmopolitan
379 nature could partly arise from different lineage pH preferences, with Lineage B found in more
380 acid habitats from pH 4.5 to 5.5 and Lineage A preferentially in nearer neutral pHs of 5.5 and
381 above. Thus, within the current study, Lineage B was absent from 6 of 26 sampled sites, while
382 Lineage A was found at all except one of the sampled sites. The detailed genetics of the two
383 cryptic lineages may provide some clues to the basis of such differences. Studies of
384 mitochondrial and genetic marker genes have established that Lineage B has lower genetic
385 diversity of measured traits than Lineage A (Donnelly et al., 2014; Kille et al., 2013). This
386 suggests that Lineage B may have undergone a population bottleneck that restricted the
387 genetic diversity, and possibly, the colonization capacity of this lineage.

388

389 The strongest correlate of lineage frequencies was soil organic matter (% loss on ignition).
390 The fresh and partially degraded soil organic component provides earthworms with food. It is,
391 therefore, possible that this association is driven by different dietary requirements of the two
392 lineages, as has been recognized for different earthworm species (Pearce, 1978). However,
393 in addition to acting as food, soil organic matter also contributes to soil structure and moisture
394 retention. Earthworms are known to be sensitive to soil texture, with regional studies linking

395 species distributions to soil sand, clay and organic matter content (Joschko et al., 2006;
396 Salome et al., 2011). Soils lacking in organic matter are also vulnerable to prolonged periods
397 of high soil moisture deficit. This can be challenging for earthworms given their critical need
398 to retain water balance. The significant correlation with site average rain days also points to a
399 possible influence of soil hydrology on distributions. Metabolomic analyses have identified that
400 many earthworm species contain a high number of betaines which likely act as osmolytes that
401 help to retain soil water balance (Liebeke and Bundy, 2013). Any differences in the extent of
402 such protection between lineages may influence colonization ability for more drought
403 susceptible soils.

404

405 Although there is correlation of lineage frequency with both soil pH and soil organic matter,
406 the fact that these two soil variables are co-correlated to other environmental variables makes
407 it hard to unequivocally assign them as the major drivers of lineage distribution. For example,
408 high organic matter/low pH soils are more common in the West of England and Wales than in
409 the East. This geographic relationship could potentially be associated with different
410 recolonization histories for the two lineages, e.g. perhaps recolonization from different glacial
411 refugia (Hewitt, 2000). However, as there is no significant correlation of Easting to lineage
412 proportion, this seems less likely than direct effects of soil organic matter/pH. Ultimately, to
413 tease apart the drivers of lineage preference, higher resolution collection and mapping and
414 experimental manipulation of habitats would be required.

415

416 Differences in physiology that separate species in relation to habitat preference could also
417 affect the way that the two lineages handle and accumulate different trace elements. For the
418 site-level analysis, the soil concentrations of major pollutant metals were correlated with PC2,
419 which was not associated with lineage. For the individual analysis of tissue metals, arsenic
420 was the only one found to vary with lineage (significantly higher in Lineage B individuals).
421 Previous work has indicated that the two lineages differ in the genetic mechanisms underlying
422 the development of arsenic tolerance. Analysis of amplified fragment length polymorphisms

423 indicated that Lineage A showed differences in patterns of nuclear markers indicating genetic
424 tolerance, while Lineage B showed a difference in DNA methylation patterning, but not genetic
425 differences (Kille et al., 2013). The observed difference here in As accumulation between
426 lineages across sites suggests that these genetic differences lead to phenotypic differences
427 in the handling of As.

428

429 5. CONCLUSIONS

430 Earthworms represent 'super-sentinels' exploited for environmental monitoring and
431 ecotoxicology, as well as being keystone soil engineers essential for soil quality. The
432 identification of possible drivers of species and lineage distributions has potential implications
433 for their use in environmental assessment as well as in studies of ecosystem service delivery.
434 For example, when assessing biodiversity effects of pollution and land-use change it may be
435 valuable to consider the occurrence of different lineages to understand how populations may
436 adapt to change through changes in lineage frequency. This analysis may be required
437 because the two widespread cryptic lineages of *L. rubellus* differ in their habitat preferences
438 with frequencies changing as conditions change. Given that bacterial communities are also
439 known to differ in relation to soil pH, then difference in the nature and strengths of earthworm
440 and microbial interactions can be expected between lineages. These relationships between
441 soil macrofauna and microbes are key to soil carbon turnover, nutrient cycling and soil
442 structural characteristics and this aspect warrants further investigation. Earthworms are also
443 valuable for metal biomonitoring. Our results suggest that the lineages behave identically with
444 respect to metal bioaccumulation, with the exception of As. Thus, selection of morphotype *L.*
445 *rubellus* will provide a coherent picture of metal accumulation independent of lineage, unless
446 As is a specific focus of any assessment.

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451 Commission for Wales for allowing access to Environmental Change Network (ECN) sites and
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453 7. REFERENCES

- 454 Anderson, C.J., Kille, P., Lawlor, A.J., Spurgeon, D.J., 2013. Life-history effects of arsenic
455 toxicity in clades of the earthworm *Lumbricus rubellus*. *Environ Pollut* 172, 200-207.
- 456 Andre, J., King, R.A., Stürzenbaum, S.R., Kille, P., Hodson, M.E., Morgan, A.J., 2010.
457 Molecular genetic differentiation in earthworms inhabiting a heterogeneous Pb-polluted
458 landscape. *Environ Pollut* 158, 883-890.
- 459 Arnold, B.E., Hodson, M.E., Charnock, J., Peijnenburg, W., 2008. Comparison of subcellular
460 partitioning, distribution, and internal speciation of Cu between Cu-tolerant and naive
461 populations of *Dendrodrilus rubidus* Savigny. *Environ Sci Technol* 42, 3900-3905.
- 462 Baker, G.H., Whitby, W.A., 2003. Soil pH preferences and the influences of soil type and
463 temperature on the survival and growth of *Aporrectodea longa* (Lumbricidae). *Pedobiologia*
464 47, 745-753.
- 465 Bardgett, R.D., van der Putten, W.H., 2014. Belowground biodiversity and ecosystem
466 functioning. *Nature* 515, 505-511.
- 467 Bartlett, M.D., Briones, M.J.I., Neilson, R., Schmidt, O., Spurgeon, D., Creamer, R.E., 2010.
468 A critical review of current methods in earthworm ecology: From individuals to populations.
469 *European Journal of Soil Biology* 46, 67-73.
- 470 Blakemore, R.J., Kupriyanova, E.K., Grygier, M.J., 2010. Neotypification of *Drawida*
471 *hattamimizu* Hatai, 1930 (Annelida, Oligochaeta, Megadrili, Moniligastridae) as a model
472 linking mtDNA (COI) sequences to an earthworm type, with a response to the 'Can of Worms'
473 theory of cryptic species. *Zookeys*, 1-29.
- 474 Blouin, M., Hodson, M.E., Delgado, E.A., Baker, G., Brussaard, L., Butt, K.R., Dai, J.,
475 Dendooven, L., Peres, G., Tondoh, J.E., Cluzeau, D., Brun, J.J., 2013. A review of earthworm
476 impact on soil function and ecosystem services. *Eur. J. Soil Sci.* 64, 161-182.

477 Boag, B., Palmer, L.F., Neilson, R., Legg, R., Chambers, S.J., 1997. Distribution, prevalence
478 and intensity of earthworm populations in arable land and grassland in Scotland. *Ann. Appl.*
479 *Biol.* 130, 153-165.

480 Bouché, M.B., 1972. *Lombriciens de France. Ecologie et systématique.* Institut national de la
481 Recherche Agronomique, Paris.

482 Bundy, J.G., Keun, H., Sidhu, J.K., Spurgeon, D.J., Svendsen, C., Kille, P., Morgan, A.J., 2007.
483 Metabolic profile biomarkers of metal contamination in a sentinel terrestrial species are
484 applicable across multiple sites. *Environ Sci Technol* 41, 4458-4464.

485 Carpenter, D., Sherlock, E., Jones, D.T., Chiminoides, J., Writer, T., Neilson, R., Boag, B.,
486 Keith, A.M., Eggleton, P., 2012. Mapping of earthworm distribution for the British Isles and
487 Eire highlights the under-recording of an ecologically important group. *Biodivers. Conserv.*
488 21, 475-485.

489 Cassagne, N., Gers, C., Gauquelin, T., 2003. Relationships between Collembola, soil chemistry
490 and humus types in forest stands (France). *Biol. Fertil. Soils* 37, 355-361.

491 Cluzeau, D., Guernion, M., Chaussod, R., Martin-Laurent, F., Villenave, C., Cortet, J., Ruiz-
492 Camacho, N., Pernin, C., MATEILLE, T., Philippot, L., Bellido, A., Rouge, L., Arrouays, D.,
493 Bispo, A., Peres, G., 2012. Integration of biodiversity in soil quality monitoring: Baselines for
494 microbial and soil fauna parameters for different land-use types. *European Journal of Soil*
495 *Biology* 49, 63-72.

496 Davies, B.E., 1989. Data handling and pattern recognition for contaminated soils. *Environ.*
497 *Geochem. Health* 11, 137-143.

498 Dennis, A.B., Hellberg, M.E., 2010. Ecological partitioning among parapatric cryptic species.
499 *Mol. Ecol.* 19, 3206-3225.

500 Dominguez, J., Aira, M., Breinholt, J.W., Stojanovic, M., James, S.W., Perez-Losada, M.,
501 2015. Underground evolution: New roots for the old tree of lumbricid earthworms. Mol.
502 Phylogenet. Evol. 83, 7-19.

503 Donnelly, R.K., Harper, G.L., Morgan, A.J., Pinto-Juma, G.A., Bruford, M.W., 2014.
504 Mitochondrial DNA and morphological variation in the sentinel earthworm species *Lumbricus*
505 *rubellus*. European Journal of Soil Biology 64, 23-29.

506 Edwards, C.A., 2004. Earthworm Ecology, 2 ed. CRC Press, Boca Raton, Florida, USA, p.
507 441.

508 Emerson, B.C., Cicconardi, F., Fanciulli, P.P., Shaw, P.J.A., 2011. Phylogeny,
509 phylogeography, phylobetadiversity and the molecular analysis of biological communities.
510 Philosophical Transactions of the Royal Society B-Biological Sciences 366, 2391-2402.

511 Emmett, B.A., Reynolds, B., Chamberlain, P.M., Rowe, E., Spurgeon, D., Brittain, S.A.,
512 Frogbrook, Z., Hughes, S., Lawlor, A.J., Poskitt, J., Potter, E., Robinson, D.A., Scott, A.,
513 Wood, C., Woods, C., 2010. Countryside Survey: Soils Report from 2007. NERC Centre for
514 Ecology and Hydrology, Wallingford, UK.

515 Fitter, A.H., Gilligan, C.A., Hollingworth, K., Kleczkowski, A., Twyman, R.M., Pitchford,
516 J.W., Programme, N.S.B., 2005. Biodiversity and ecosystem function in soil. Funct. Ecol. 19,
517 369-377.

518 Folmer, O., Black, M., Hoeh, W., Lutz, R., Vrijenhoek, R., 1994. DNA primers for
519 amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan
520 invertebrates. Mol. Mar. Biol. Biotechnol. 3, 294-299.

521 Gaston, K.J., 2003. The Structure and Dynamics of Geographic Ranges. Oxford University
522 Presss, Oxford, UK.

523 Giller, P.S., 1996. The diversity of soil communities, the 'poor man's tropical rainforest'.
524 Biodivers. Conserv. 5, 135-168.

525 Giska, I., Sechi, P., Babik, W., 2015. Deeply divergent sympatric mitochondrial lineages of the
526 earthworm *Lumbricus rubellus* are not reproductively isolated. BMC Evol. Biol. 15.

527 Griffiths, R.I., Thomson, B.C., James, P., Bell, T., Bailey, M., Whiteley, A.S., 2011. The
528 bacterial biogeography of British soils. Environ. Microbiol. 13, 1642-1654.

529 Hewitt, G., 2000. The genetic legacy of the Quaternary ice ages. Nature 405, 907-913.

530 International Organisation for Standards, 2005. Determination of pH., Geneva, Switzerland.

531 Jaensch, S., Steffens, L., Hoefler, H., Horak, F., Ross-Nickoll, M., Russell, D., Toschki, A.,
532 Roembke, J., 2013. State of knowledge of earthworm communities in German soils as a basis
533 for biological soil quality assessment. Soil Organisms 85, 215-233.

534 James, S.W., Porco, D., Decaens, T., Richard, B., Rougerie, R., Erseus, C.A.F.N.J.S.W., Porco,
535 D., Decaens, T., Richard, B., Rougerie, R., Erseus, C., 2010. DNA Barcoding Reveals Cryptic
536 Diversity in *Lumbricus terrestris* L., 1758 (Clitellata): Resurrection of *L. herculeus* (Savigny,
537 1826). PLoS One 5, e15629.

538 Jones, G.L., Wills, A., Morgan, A.J., Thiomias, R.J., Kille, P., Novo, M., 2016. The worm has
539 turned: Behavioural drivers of reproductive isolation between cryptic lineages. Soil Biol.
540 Biochem. 98, 11-17.

541 Joschko, M., Fox, C.A., Lentzsch, P., Kiesel, J., Hierold, W., Kruck, S., Timmer, J., 2006.
542 Spatial analysis of earthworm biodiversity at the regional scale. Agric. Ecosys. Environ. 112,
543 367-380.

544 Kille, P., Andre, J., Anderson, C., Ang, H.N., Bruford, M.W., Bundy, J.G., Donnelly, R.,
545 Hodson, M.E., Juma, G., Lahive, E., Morgan, A.J., Sturzenbaum, S.R., Spurgeon, D.J., 2013.
546 DNA sequence variation and methylation in an arsenic tolerant earthworm population. Soil
547 Biol. Biochem. 57, 524-532.

548 King, R.A., Tibble, A.L., Symondson, W.O.C., 2008. Opening a can of worms: unprecedented
549 sympatric cryptic diversity within British lumbricid earthworms. Mol. Ecol. 17, 4684-4698.

550 Klarica, J., Kloss-Brandstatter, A., Traugott, M., Juen, A., 2012. Comparing four mitochondrial
551 genes in earthworms - Implications for identification, phylogenetics, and discovery of cryptic
552 species. *Soil Biol. Biochem.* 45, 23-30.

553 Lavelle, P., Bignell, D., Lepage, M., Wolters, V., Roger, P., Ineson, P., Heal, O.W., Dhillon,
554 S., 1997. Soil function in a changing world: the role of invertebrate ecosystem engineers.
555 *European Journal of Soil Biology* 33, 159-193.

556 Leveque, T., Capowiez, Y., Schreck, E., Mombo, S., Mazzia, C., Foucault, Y., Dumat, C.,
557 2015. Effects of historic metal(loid) pollution on earthworm communities. *Sci Total Environ*
558 511, 738-746.

559 Liebeke, M., Bruford, M.W., Donnelly, R.K., Ebbels, T.M.D., Hao, J., Kille, P., Lahive, E.,
560 Madison, R.M., Morgan, A.J., Pinto-Juma, G.A., Spurgeon, D.J., Svendsen, C., Bundy, J.G.,
561 2014. Identifying biochemical phenotypic differences between cryptic species. *Biol. Lett.* 10.

562 Liebeke, M., Bundy, J.G., 2013. Biochemical diversity of betaines in earthworms. *Biochem.*
563 *Biophys. Res. Commun.* 430, 1306-1311.

564 Liebeke, M., Strittmatter, N., Fearn, S., Morgan, A.J., Kille, P., Fuchser, J., Wallis, D.,
565 Palchykov, V., Robertson, J., Lahive, E., Spurgeon, D.J., McPhail, D., Takats, Z., Bundy, J.G.,
566 2015. Unique metabolites protect earthworms against plant polyphenols. *Nature Comms.* 6.

567 Novo, M., Cunha, L., Maceda-Veiga, A., Talavera, J.A., Hodson, M.E., Spurgeon, D., Bruford,
568 M.W., Morgan, A.J., Kille, P., 2015. Multiple introductions and environmental factors
569 affecting the establishment of invasive species on a volcanic island. *Soil Biol. Biochem.* 85,
570 89-100.

571 Pass, D.A., Morgan, A.J., Read, D.S., Field, D., Weightman, A.J., Kille, P., 2015. The effect
572 of anthropogenic arsenic contamination on the earthworm microbiome. *Environ. Microbiol.*
573 17, 1884-1896.

574 PerezLosada, M., Ricoy, M., Marshall, J.C., Dominguez, J., 2009. Phylogenetic assessment of
575 the earthworm *Aporrectodea caliginosa* species complex (Oligochaeta: Lumbricidae) based on
576 mitochondrial and nuclear DNA sequences. *Mol. Phylogenet. Evol.* 52, 293-302.

577 Pearce, T.G., 1978. Gut contents of some Lumbricid earthworms. *Pedobiologia* 18, 153-157.

578 Raty, M., Huhta, V., 2003. Earthworms and pH affect communities of nematodes and
579 enchytraeids in forest soil. *Biol. Fertil. Soils* 38, 52-58.

580 Römbke, J., Aira, M., Backeljau, T., Breugelmans, K., Domínguez, J., Funke, E., Graf, N.,
581 Hajibabaei, M., Pérez-Losada, M., Porto, P.G., Schmelz, R.M., Vierna, J., Vizcaíno, A.,
582 Pfenninger, M., 2016. DNA barcoding of earthworms (*Eisenia fetida/andrei* complex) from 28
583 ecotoxicological test laboratories. *Appl. Soil Ecol.* doi:10.1016/j.apsoil.2015.02.010.

584 Rowell, D.L., 1994. *Soil Science: Methods and Applications*. Longman Scientific and
585 Technical, Harlow, UK.

586 Rutgers, M., Orgiazzi, A., Gardi, C., Roembke, J., Jaensch, S., Keith, A.M., Neilson, R., Boag,
587 B., Schmidt, O., Murchie, A.K., Blackshaw, R.P., Peres, G., Cluzeau, D., Guernion, M.,
588 Briones, M.J.I., Rodeiro, J., Pineiro, R., Diaz Cosin, D.J., Paulo Sousa, J., Suhadolc, M., Kos,
589 I., Krogh, P.-H., Faber, J.H., Mulder, C., Bogte, J.J., van Wijnen, H.J., Schouten, A.J., de
590 Zwart, D., 2016. Mapping earthworm communities in Europe. *Appl. Soil Ecol.* 97, 98-111.

591 Rutgers, M., Schouten, A.J., Bloem, J., vanEekeren, N., deGoede, R.G.M., Akkerhuis, G.,
592 vanderWal, A., Mulder, C., Brussaard, L., Breure, A.M., 2009. Biological measurements in a
593 nationwide soil monitoring network. *Eur. J. Soil Sci.* 60, 820-832.

594 Sackett, T.E., Smith, S.M., Basiliko, N., 2013. Indirect and direct effects of exotic earthworms
595 on soil nutrient and carbon pools in North American temperate forests. *Soil Biol. Biochem.* 57,
596 459-467.

597 Salome, C., Guenat, C., Bullinger-Weber, G., Gobat, J.M., Le Bayon, R.C., 2011. Earthworm
598 communities in alluvial forests: Influence of altitude, vegetation stages and soil parameters.
599 *Pedobiologia* 54, S89-S98.

600 Sims, R.W., Gerard, B.M., 1985. Earthworms. Linnean Society and the Estuarine and
601 Brackish-Water Sciences Association, London.

602 Spurgeon, D.J., Lofts, S., Hankard, P.K., Toal, M., McLellan, D., Fishwick, S., Svendsen, C.,
603 2006. Effect of pH on metal speciation and resulting metal uptake and toxicity for earthworms.
604 *Environ Toxicol Chem* 25, 788-796.

605 Spurgeon, D.J., Rowland, P., Ainsworth, G., Rothery, P., Long, S., Black, H.I.J., 2008.
606 Geographical and pedological drivers of distribution and risks to soil fauna of seven metals
607 (Cd, Cu, Cr, Ni, Pb, V and Zn) in British soils. *Environ Pollut* 153, 273-283.

608 Umarov, M.M., Striganova, B.R., Kostin, N.V., 2008. Specific features of nitrogen
609 transformation in the gut and coprolites of earthworms. *Biol. Bulletin* 35, 643-652.

610 Van Gestel, C.A.M., Dirven-Van Breemen, E.M., Baerselman, R., 1992. Influence of
611 environmental conditions on the growth and reproduction of the earthworm *Eisenia andrei* in
612 an artificial soil substrate. *Pedobiologia* 36, 109-120.

613 Warton, D.I., Hui, F.K.C., 2011. The arcsine is asinine: the analysis of proportions in ecology.
614 *Ecology* 92, 3-10.

615

616 LEGENDS TO FIGURES

617

618 Figure 1. Location of collection sites and the proportion of Lineage A (dark blue shading) and
619 Lineage B (light yellow shading) *L. rubellus* based on the total number of collected and
620 assigned genotyped individual (given in brackets) for the 26 sites visit over four separate
621 collection campaigns

622

623 Figure 2. Principal component analysis results show the ordination of site geographical,
624 climatic, biotic and soil chemical variables of sample sites showing the major related site
625 characteristic variables.

626

627 Figure 3. Boxplots showing median (centre line), upper and lower quartile (box limits) and
628 upper and 95% confidence intervals (whiskers) of trace metal concentrations measured
629 across 17 samples site for assigned Lineage A and Lineage B *L. rubellus*.

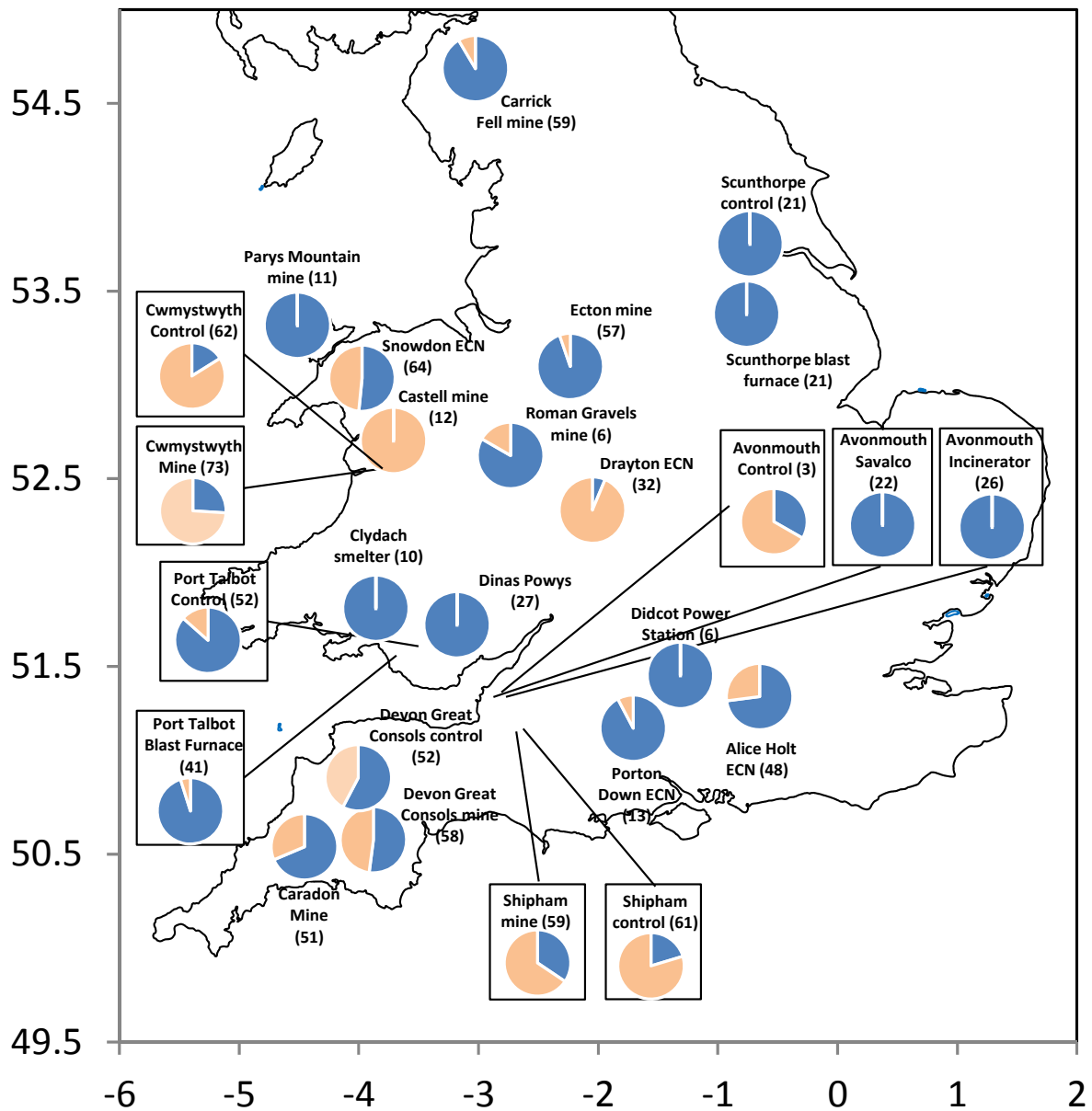
630

631 Figure 4. Scatterplots with fitted locally weighted scatterplot smoother line of proportion of
632 Lineage A *L. rubellus* in relation to (a) Soil % OM and (b) soil pH.

633

634 FIG. 1

635



636

FIG. 2

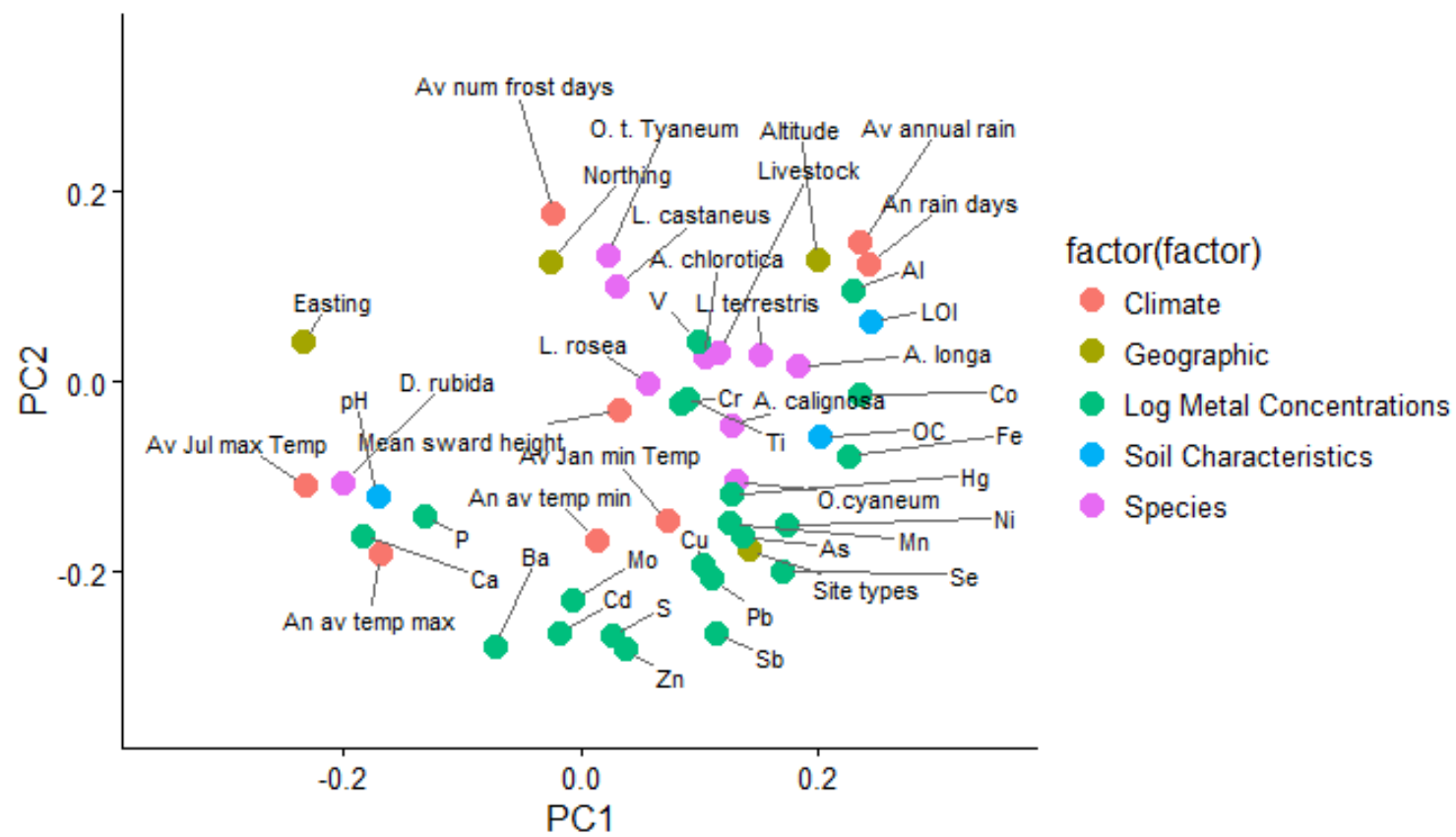


FIG. 3

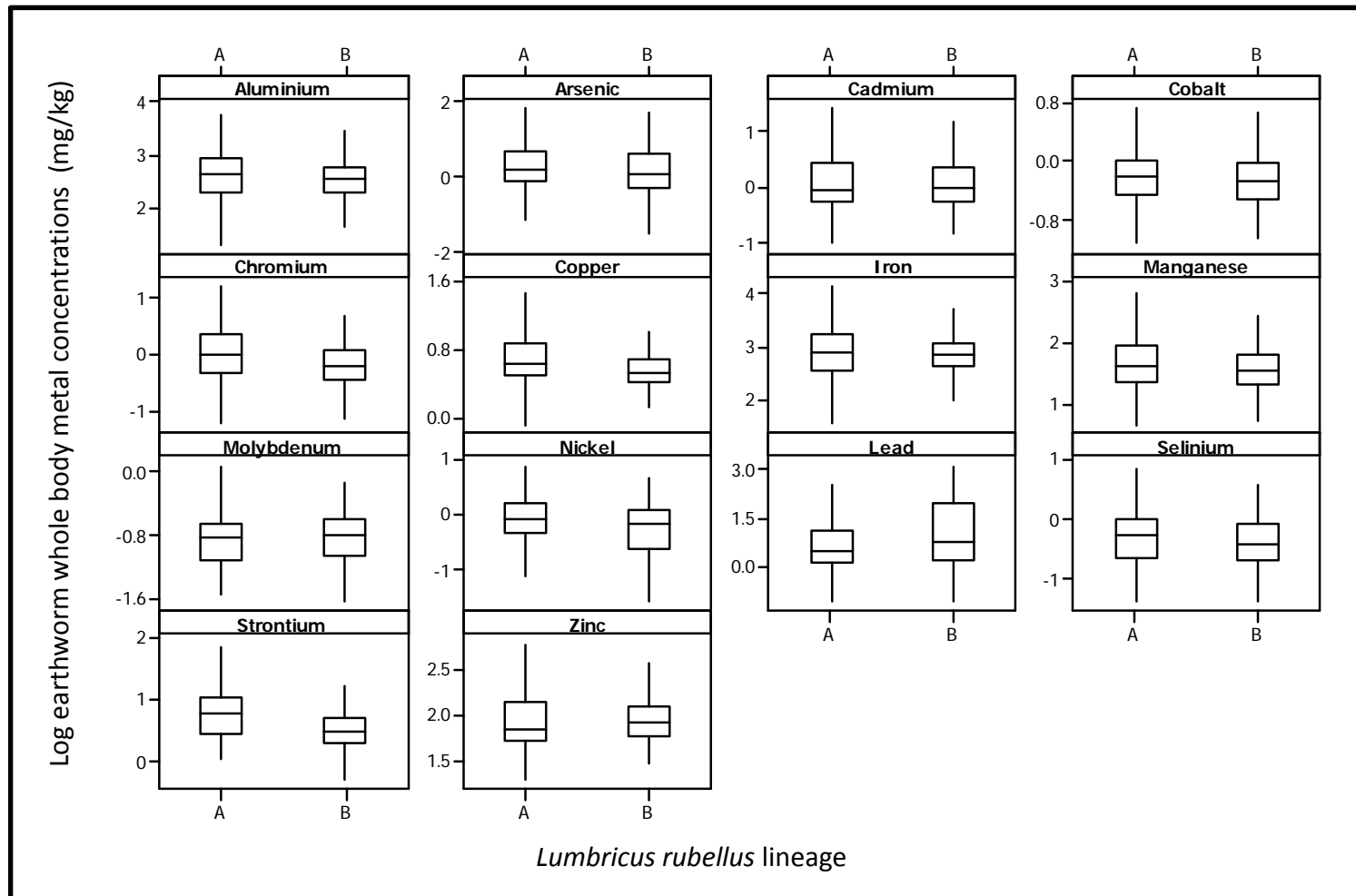
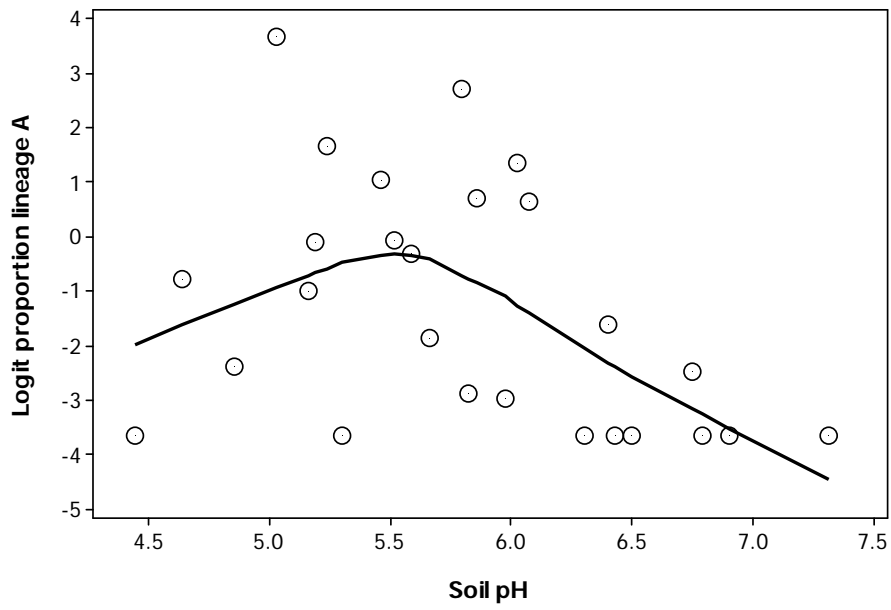
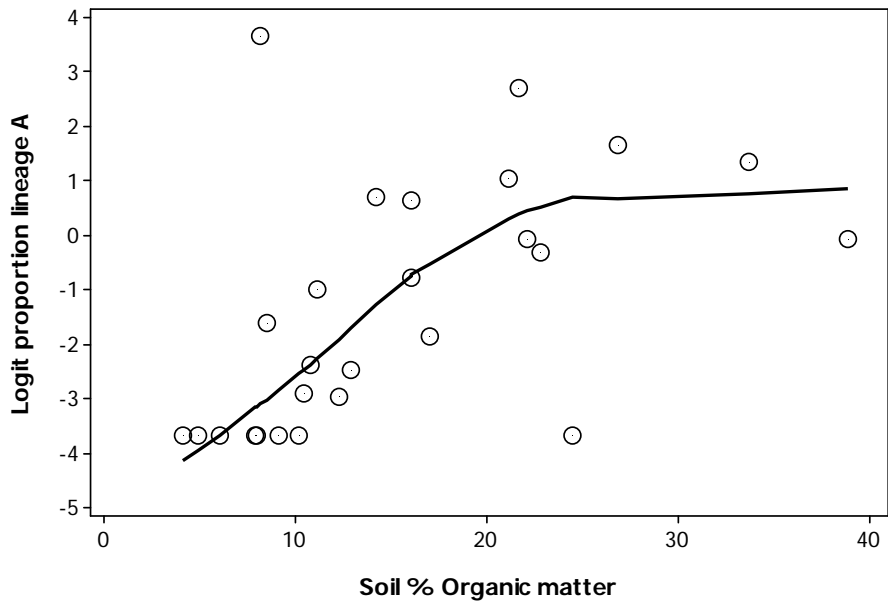


FIG. 4



SUPPLEMENTARY TABLES

Supplementary Table 1. Geographical locations and reported climatic conditions of the 23 sites used for the collection of morphotype *L. rubellus*.

Site name	Site types	Land use	Ordnance Survey Grid reference	Easting	Northing	Altitude	Annual Average temp Max	Annual average temp min	Average Jan Min Temp	Average Jul Max Temp	Average Annual Rain	Annual rain days	Average No Forst Days
Alice Holt ECN Control	Unpolluted	Broadleaf woodland	SU 80060 39821	480060	139821	88.6	14.1	6.4	1.6	21.9	755	121	45.9
Avonmouth Control	Unpolluted	Improved pasture	ST 57006 82149	357006	182149	7	14.2	7	2.2	21.5	802	126	34.9
Avonmouth Incinerator	Industrial polluted	Rough grassland	ST 54099 81659	354099	181652	6	14.2	7	2.2	21.5	802	126	34.9
Avonmouth Savalco	Industrial polluted	Rough grassland	ST 53859 79411	353859	179411	6	14.2	7	2.2	21.5	802	126	34.9
Caradon Mine	Mining polluted	Rough grassland	SX 25624 69792	225624	69792	226	13.2	7	3	19.1	1385	172	30.6
Carrick Fell Mine	Mining polluted	Rough grassland	NY 32211 32982	332211	532982	661	13	5.8	1.6	19.7	1521	176	56.5
Castell Mine	Mining polluted	Rough grassland	SN 77415 81254	277415	281254	297	11.9	5.2	1	18.2	186	191	58.4
Clydach Smelter	Industrial polluted	Broadleaf woodland	SN 69587 01409	269587	201409	25	13.5	8.5	4	19.6	999	148	9.7
Cwmystwyth control	Unpolluted	Rough grassland	SN 79598 74222	279598	274222	198	11.9	5.2	1	18.2	1856	191	58.4
Cwmystwyth mine	Mining polluted	Rough grassland	SN 80852 75166	280852	275166	177	11.9	5.2	1	18.2	1856	191	58.4
Devon Great Consouls Control	Unpolluted	Improved pasture	SX 42560 74019	242560	74019	133	14	8.1	4	19.9	1007.4	142	16.3
Devon Great Consouls Mine	Mining polluted	Broadleaf woodland	SX 42385 73152	242385	73152	133	14	8.1	4	19.9	1007.4	142	16.3
Didcot Power Station	Industrial polluted	Broadleaf woodland	SU 51645 91402	451645	191402	53	14.4	5.9	1.2	22.6	661	112	57.7
Drayton ECN Control	Unpolluted	Improved pasture	SP 16391 55061	416391	255061	66	14.5	5.9	1.3	22.8	614	114	52.2
Dinas Powys	Unpolluted	Broadleaf woodland	ST 15868 70431	315868	170431	57	14.7	7	2.3	21.7	1151.9	149	35.7
Ecton Mine	Mining polluted	Broadleaf woodland	SK 09698 58263	409698	358263	103	13.9	6	1.2	22.1	598	112	49.1
Parys Mountain	Mining polluted	Rough grassland	SH 43829 89971	243829	389971	117	13.2	7.7	3.6	18.8	841	143	20.3
Port Talbot Control	Unpolluted	Rough grassland	SS 83690 84574	283690	184574	150	13.5	8.5	4	19.6	999	148	9.7
Port Talbot Blast Furnace	Industrial polluted	Rough grassland	SS 79001 85463	279001	185463	6.1	13.5	8.5	4	19.6	999	148	9.7
Porton Down ECN	Unpolluted	Improved pasture	SU 19575 37692	419575	137692	102	14.1	6.2	1.4	21.9	749	122	47.6
Roman Gravels Mine	Mining polluted	Rough grassland	SJ 33592 00339	333592	300339	368	14.1	5.6	1.3	21.6	668	126	51.8
Scunthorpe blast furnace	Industrial polluted	Arable	SE 94800 15840	494800	415835	19.7	13.4	5.7	0.9	21.3	613	115	49.8
Scunthorpe Control	Unpolluted	Arable	SE 93156 12000	493156	412000	10.7	13.4	5.7	0.9	21.3	613	115	49.8
Shipham control	Unpolluted	Improved pasture	ST 46312 59409	346312	159409	54	14.6	7	2.6	21.7	899	134	28.9
Shipham mine	Mining polluted	Improved pasture	ST 44799 57273	344799	157273	169	14.6	7	2.6	21.7	899	134	28.9
Snowdon ECN control	Unpolluted	Rough grassland	SH 63674 55116	263674	355116	748	12	5.9	1.8	18.1	2612	199	50.1

Supplementary Table 2. Vegetation and presence (shaded) or absence (unshaded) for earthworm species at the 23 sites used for the collection of morphotype *L. rubellus*.

Site name	Mean sward height	Livestock	Exposure	<i>Aporrectodea caliginosa</i>	<i>Aporrectodea rosea</i>	<i>Lumbricus castaneus</i>	<i>Dendrobaena rubida</i>	<i>Aporrectodea longa</i>	<i>Lumbricus terrestris</i>	<i>Lumbricus festivus</i>	<i>Octolasion cyaneum</i> ,	<i>Allobophora chlorotica</i>	<i>Octolasion tyrtaeum tyaneum</i>
Alice Holt ECN Control	-	Deer	shaded	shaded				shaded			shaded		
Avonmouth Control	20 cm	None	part-shaded	shaded				shaded				shaded	
Avonmouth Incinerator	10 cm	None	open	shaded		shaded		shaded					shaded
Avonmouth Savalco	35 cm	None	open			shaded			shaded				
Caradon Mine	5 cm	horses	open										
Carrick Fell Mine	10 cm	sheep	open						shaded				
Castell Mine	5 cm	None	open			shaded				shaded			
Clydach Smelter	5 cm	None	shaded	shaded			shaded				shaded		
Cwmystwyth control	10 cm	None	open				shaded						
Cwmystwyth mine	10 cm	sheep	open										
Devon Great Consouls Control	10 cm	None	open	shaded	shaded			shaded				shaded	
Devon Great Consouls Mine	10 cm	None	part-shaded	shaded			shaded						
Didcot Power Station	-	None	Shaded	shaded	shaded			shaded			shaded	shaded	
Drayton ECN Control	15 cm	None	open	shaded				shaded					
Dinas Powys	-	None	shaded	shaded	shaded			shaded				shaded	
Ecton Mine	-	None	part-shaded						shaded				
Parys Mountain	5 cm	None	part-shaded	shaded									
Port Talbot Control	10 cm	None	part-shaded	shaded	shaded			shaded					shaded
Port Talbot Blast Furnace	10 cm	horses	open	shaded	shaded			shaded					shaded
Porton Down ECN	8 cm	None	part_shaded										
Roman Gravels Mine	3 cm	horses	open										
Scunthorpe blast furnace	5 cm	None	open	shaded	shaded			shaded				shaded	
Scunthorpe Control	6 cm	None	part-shaded	shaded	shaded			shaded			shaded		
Shipham control	10 cm	None	open	shaded				shaded			shaded		
Shipham mine	10 cm	cattle	part-shaded			shaded			shaded				
Snowdown ECN control	15 cm	sheep	open	shaded		shaded	shaded						

Supplementary Table 3. Arithmetic mean of measured soil chemical properties for pH, loss on ignition and concentrations of a suite of trace elements based on analysis of three samples collected from sites the 23 sites used for the collection of morphotype *L. rubellus*.

Site name	Soil pH	% Soil loss on ignition	Al (mg/kg)	As (mg/kg)	Ba (mg/kg)	Cd (mg/kg)	Co (mg/kg)	Cr (mg/kg)	Cu (mg/kg)	Fe (mg/kg)	Hg (mg/kg)	Mn (mg/kg)	Mo (mg/kg)	Ni (mg/kg)	Pb (mg/kg)	Sb (mg/kg)	Se (mg/kg)	Tl (mg/kg)	V (mg/kg)	Zn (mg/kg)	Ca (mg/kg)	P (mg/kg)	S (mg/kg)
Alice Holt ECN Control	5.16	11.15	8737	13.2	19.3	0.1	6	12.6	12.4	16500	0.17	78	0.3	9.5	27.1	0.4	0.5	7	24.6	43	1640	424	202
Avonmouth Control	5.86	14.23	11130	20.9	356	2.3	10	46.3	75.1	26133	0.76	539	1.2	25.1	207	1.3	1.7	32.9	26.9	697	68633	524	1029
Avonmouth Incinerator	6.5	7.94	11000	36.9	439	50.6	12.3	23.6	204.3	26133	1.07	1010	2	25.7	1943	14.9	5.6	34.4	31.8	4640	39267	1600	3190
Avonmouth Savaleo	6.3	24.51	14733	7.8	75.1	2.6	8.1	25.1	29.8	19233	0.25	663	0.6	18.4	99.6	1.2	0.6	95.5	29.6	299	4777	1233	734
Caradon Mine	4.64	16	5147	407	33.7	1.17	2.03	3.9	609	20633	0.33	221	2.57	4.27	69.1	3.17	1.49	27.4	13.8	43.5	342	776	884
Carrick Fell Mine	4.85	10.8	16300	737	33.5	3.07	8.18	16.4	59	33300	2.06	837	18.10	11.4	173	7.01	3.85	165	92.6	282	1607	961	1457
Castell Mine	5.03	8.15	14333	14.7	15.7	9.03	12.5	16.4	60	46267	0.5	844	1.16	16.5	210	2.28	1.35	8.1	26.8	1792	321	506	312
Clydach Smelter	5.3	10.2	6780	44.1	142	1.61	90.1	32.3	465	25067	0.34	1369	2.68	1799	278	3.57	14.43	70.9	47.2	370	39277	945	1230
Cwmystwyth control	5.23	26.89	16167	19.8	14.6	0.1	6.7	18.7	19.7	38033	0.12	467	0.9	14.3	626	1.6	1.2	6.6	24.6	116	209	688	709
Cwmystwyth mine	5.46	21.13	20033	49	29.8	0.2	41.2	24.1	28.8	50433	1.12	2597	1.5	23.3	657	2.5	1.4	22	33.3	127	551	689	534
Devon Great Consols Control	5.58	22.79	21533	310	45.5	0.4	14.8	31.7	107.3	45800	0.05	585	1	27.5	68	1.4	2.3	41.8	38.7	140	2213	1177	919
Devon Great Consols Mine	5.19	38.9	17300	6270	45.9	0.2	25.7	17.8	2647	79600	0.64	630	1.3	22.3	225	13.9	2.6	38.4	38.5	277	3340	286	1503
Didcot Power Station	7.31	4.17	10570	14.5	139	0.53	8.89	14.7	41	22900	0.34	309	0.93	24.1	102	2.42	1.40	15.1	26.9	181.00	37600	901	1250
Drayton ECN Control	5.79	21.65	12833	25.1	48.4	0.4	4.9	15.8	46	28533	0.1	805	2.3	10.7	30.2	0.6	0.5	34.3	32	176	35000	2000	1183
Dinas Powys	6.9	8	17166	23	85.4	1.2	16.1	39	42.6	24933	0.1	1120	1.7	45.1	109	*	1.2	41.5	45.7	770	12766	2070	*
Ecton Mine	5.82	10.4	1403	136	537	61.4	33.2	5.4	5787	12267	0.35	965	101	72.4	1553	92.1	5.42	20.1	20.2	6047	116200	203	9280
Parys Mountain	4.44	4.95	681	1480	96	8.77	7.78	5	3673	175667	3.34	35.7	37.5	3.17	29033	210	42	54.4	19	2333	1749	26.7	22467
Port Talbot Control	5.66	16.99	15300	14.5	127	0.6	11.3	37.7	33.8	31467	0.27	1287	0.9	17.4	39.1	0.8	1.2	15.7	62.9	480	13903	1427	1083
Port Talbot Blast Furnace	5.98	12.3	10323	14.5	287	0.8	4.7	129	35.5	34967	0.33	3550	1.7	16.8	117	2.2	1.1	230.7	140	341	89133	1227	1680
Porton Down ECN	6.75	12.93	6153	21	88.3	0.7	8	20	39.2	13567	0.37	611	0.9	17.3	109	1.4	0.5	46.3	17.2	184	148333	3100	1423
Roman Gravels Mine	6.4	8.48	10633	13.7	123	14.2	14.3	13.3	99	30133	0.33	582	1.17	24.8	1125	2.48	1.33	15.7	18.4	1788	6367	455	878
Scunthorpe blast furnace	6.43	9.14	10803	40.6	90.3	0.3	13.1	42	25.4	59900	0.33	1227	1.5	29.5	124	1.8	1	68.3	128.7	183	13500	1497	453
Scunthorpe Control	6.79	6.04	7197	24.6	47.4	0.4	4.9	15.4	20.2	21767	0.12	535	2.3	11.2	30.0	0.6	0.5	33.6	30.7	69.1	130667	1253	1167
Shipham control	6.07	16.1	12967	37.1	302	2.1	7.5	23.1	19.1	26500	0.03	465	0.7	16.4	163	0.9	0.4	53.9	34.8	328	3520	972	584
Shipham mine	6.03	33.7	9907	867	1526	404	12.5	21	85.1	85100	12.3	2277	5.1	31.6	7260	48.8	3.8	51.4	28.5	31833	16117	2370	3547
Snowdown ECN control	5.52	22.07	38600	17.5	10.1	0.3	30.6	78.2	19.5	70733	0.19	1350	0.5	30.4	37	0.5	1.4	1530	239	114	1111	524	658