Residual hydrophobic organic contaminants in soil: are they a barrier to risk-based approaches for managing contaminated land?

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# 1 List of Abbreviations

2	BR	Bound Residue
3	CSM	Conceptual Site Model
4	DCM	Dichloromethane
5	DCM/ACE	Dichloromethane/Acetone
6	ECETOC	European Centre for Ecotoxicology and Toxicology of Chemicals
7	EET	Exhaustive Extraction Technique
8	EHRA	Ecological Health Risk Assessment
9	ER	Extractable Residue
10	EVA	Ethylene Vinyl Acetate
11	GI	Gastrointestinal
12	HHRA	Human Health Risk Assessment
13	HOC	Hydrophobic Organic Contaminant
14	HPCD	Hydroxypropyl-β-Cyclodextrin
15	HPLC-FD	High-Performance Liquid Chromatography with Fluorescence Detection
16	MGP	Manufactured Gas Plant
17	NEET	Non-Exhaustive Extraction Technique
18	NEPM	National Environment Protection Measure
19	NER	Non-Extractable Residue
20	OC	Organic Carbon
21	PDMS	Polydimethylsiloxane
22	PED	Polyethylene devices
23	POM	Polyoxymethylene
24	RA	Risk Assessment
25	RBA	Relative Bioavailability
26	RBLM	Risk-Based Land Management
27	SARA	Soil-sediment Availability Ratio
28	SBE	Silicon-Based Extraction
29	SFE	Supercritical Fluid Extraction
30	SGV	Soil Guideline Value
31	SOP	Standard Operating Procedure
32	SPE	Solid Phase Extraction
33	SPMD	Semipermeable Membrane Devices
34	SPME	Solid Phase Micro-Extraction
35	TR	Total Residue
36	USEPA	United States Environmental Protection Agency

#### 37 Abstract

38 Risk-based approaches to managing contaminated land, rather than approaches based on 39 complete contaminant removal, have gained acceptance as they are likely to be more 40 feasible and cost effective. Risk-based approaches aim to minimise risks of exposure of a 41 specified contaminant to humans. However, adopting a risk-based approach over alternative 42 overly-conservative approaches requires that associated uncertainties in decision making 43 are understood and minimised. Irrespective of the nature of contaminants, a critical 44 uncertainty is whether there are potential risks associated with exposure to the residual 45 contaminant fractions in soil to humans and other ecological receptors, and how they should 46 be considered in the risk assessment process. This review focusing on hydrophobic organic 47 contaminants (HOCs), especially polycyclic aromatic hydrocarbons (PAHs), suggests that there is significant uncertainty on the residual fractions of contaminants from risk 48 49 perspectives. This is because very few studies have focused on understanding the 50 desorption behaviour of HOCs, with few or no studies considering the influence of exposure-51 specific factors. In particular, it is not clear whether the exposure of soil-associated HOCs to 52 gastrointestinal fluids and enzyme processes release bound residues. Although, in vitro 53 models have been used to predict PAH bioaccessibility, and chemical extractions have been 54 used to determine residual fractions in various soils, there are still doubts about what is 55 actually being measured. Therefore it is not certain which bioaccessibility method currently 56 represents the best choice, or provides the best estimate, of in vivo PAH bioavailability. It is 57 suggested that the fate and behaviour of HOCs in a wide range of soils, and that consider 58 exposure-specific scenarios, be investigated. Exposure-specific scenarios are important for 59 validation purposes, which may be useful for the development of standardised methods and procedures for HOC bioaccessibility determinations. Research is needed to propose the 60 61 most appropriate testing methods and for assessing potential risks posed by residual 62 fractions of HOCs. Such investigations may be useful for minimising uncertainties associated 63 with a risk-based approach, so that consideration may then be given to its adoption on a

global scale. This review critically appraises existing information on the bioavailability of
 HOC residues in soil to establish whether there may be risks from highly sequestered
 contaminant residues.

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Keywords: Risk-Based Approach, Residual HOC Fractions, Bioavailability and
Bioaccessibility, Sorption-Desorption, Standard Operating Procedures, soil-HOC-organism
interactions

72 1. Introduction

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"To forget how to tend soils is to forget ourselves" (Mahatma Gandhi).

Chemical contamination of soils is a global problem <sup>1</sup> and is arguably of similar 74 75 significance as other major environmental challenges such as climate change and 76 biodiversity loss<sup>2</sup>. Sustainable soil management has a vital role for addressing the challenge 77 of widespread contamination, especially with regard to attaining the United Nations 'Sustainable Development Goals' <sup>3</sup>. Among the anthropogenic contaminants of soils, 78 79 hydrophobic organic contaminants (HOCs) require attention as they are persistent, they bioaccumulate, and are toxic and potentially carcinogenic <sup>4-6</sup>. Examples of important HOCs 80 81 include PAHs, polybrominated diphenyl ethers (PBDEs), polychlorinated biphenyls (PCBs), 82 dichloro-diphenyl-trichloroethane (DDT) and other chlorinated pesticides. The risk of soil 83 contamination by HOCs is a matter of great concern for human and ecological health<sup>7</sup>. 84 Human exposure to HOCs in soils may be by oral ingestion, inhalation, and/or dermal routes <sup>7,8</sup>. Ingestion has been regarded as very important for HOCs, such as the PAHs <sup>7,9</sup>. 85

86 Assessment of HOC contamination in soils is currently based on the measurement of total concentrations and the assumption that the measured HOCs are 100% bioavailable <sup>10-</sup> 87 <sup>13</sup>. It has been reported that total contaminant concentrations in ingested soil or food do not 88 89 provide a good measure of the contaminant's absorption via the organism's gut membrane or that absorbed into its systemic circulation <sup>11, 14-18</sup>. This may be due to the interactions 90 91 between the contaminant and the soil. Wide variations in soil and HOC properties may significantly affect bioaccessibility (bioavailability + potential bioavailability) of HOCs in 92 humans <sup>14, 19-21</sup>. For example, when HOCs enter soil, sequestration processes such as 93 94 diffusion of molecules into inaccessible micro- and nano-pores, as well as sorption to soil components, are known to affect bioavailability and bioaccessibility<sup>19, 22-27</sup>. Consequently, 95 96 varying soil-organism-HOC interactions may influence the significance of potential harm that 97 may result from exposure. Hence, the bioaccessible contaminant concentration rather than 98 the total concentration in soil is more important for realistic risk assessment (RA) purposes.

99 Risk-based approaches to contaminant management and remediation offer feasible 100 remediation practices in that they recognise that complete removal of a contaminant is likely 101 to be technically very difficult, expensive, and sometimes unnecessary <sup>28-31</sup>. Risk-based 102 approaches need to be underpinned by a thorough understanding of the chemical behaviour 103 of HOCs in soils, and in particular definition of the fraction of the total concentration that is 104 relevant to biological or environmental impacts, i.e. bioaccessibility. Adoption and application 105 of risk-based approaches face a significant hurdle due to the lack of regulatory recognition of 106 the bioavailability concept which underpins the approach, although it is perceived as an important concept for RA within certain sections of the regulatory <sup>32</sup> and scientific <sup>11-13, 15, 33</sup> 107 108 communities. This is related to the lack of standard operating procedures (SOPs) for measuring the bioavailable fractions of HOCs in soil, as such measurements are still in their 109 110 infancy. Although validated SOPs for bioavailability of thoroughly studied inorganic 111 contaminants such as lead in soil have gained regulatory approval and are currently in use in the USA <sup>34</sup>, there is no generally accepted SOPs for determining HOC bioavailability in soil. 112 113 In addition, information on the bioavailability and risks associated with 'residual' HOCs in soil 114 is limited.

115 Although bioavailable fractions of HOCs are by definition of most concern in assessing 116 their environmental and health risks in soil, it is yet to be demonstrated conclusively whether residues that remain following removal of the bioavailable fractions pose ongoing risk from 117 long-term exposure, especially in historically contaminated soils <sup>29, 35-39</sup>. In the absence of 118 119 conclusive evidence regarding potential risk arising from long-term exposure to residual 120 fractions, the adoption of the risk-based approach for HOCs is often challenged. There have 121 been arguments regarding the significance of non-extractable residual (NER) fractions, defined in this review as highly sequestered residues, to ongoing risks in the long term <sup>35, 36,</sup> 122 <sup>39-45</sup>, because changes in environmental or exposure conditions may mobilise residual pools 123 in soil <sup>30, 43, 46-48</sup>. A number of papers have argued that long-term exposure to NER fractions 124 125 in soil poses little or no risk because of the strong interactions between HOCs and soil components especially organic matter (OM), resulting in very slow remobilisation HOCs, 126

mainly through desorption <sup>25, 26, 30, 31, 43, 49-51</sup>. Desorption rates of HOCs from historically 127 contaminated soils and sediments have been reported to be slow or very slow (10<sup>-2</sup> to 10<sup>-4</sup> h-128 1) <sup>26, 49</sup>, and with half-lives that could last decades <sup>50</sup>. For example, a study reported that the 129 130 slowly desorbing fraction of <sup>14</sup>C-phenanthrene slightly increased between 2 and 10 d in 4 different artificially-spiked soils (4.8 - 10.25 % OM), with the soil richest in OM showing more 131 pronounced effects with ageing <sup>52</sup>. Other studies have corroborated these findings, with 132 increases in the desorption-resistant (i.e. residual) fraction by up to 2 - 10 fold with ageing <sup>26,</sup> 133 134 <sup>53, 54</sup>. The implication is that with increase in the residual (or strongly-sorbed) fraction as 135 ageing progresses, desorption of contaminants becomes increasingly difficult. Even when 136 desorption does occur, it does so at a very slow rate such that the risk is acceptable, or that no risk is likely to be posed <sup>55</sup>. However, this argument is mainly derived from indirect 137 empirical relationships between contaminant sorption behaviour and soil OM structure <sup>56</sup>. 138 139 Although the afore-mentioned studies have been useful for understanding soil-HOC 140 interactions on a macroscopic scale, more convincing molecular-level based approaches 141 which directly depicts these interactions are lacking, especially given the current availability 142 of advanced analytical techniques such as the nuclear magnetic spectroscopy (NMR) among others <sup>56-58</sup>. While such advanced approaches may not be needed for routine assessment of 143 144 residual fractions, detailed investigations of residual HOCs in a wide range of long-term 145 contaminated soils will provide much needed data to regulators on potential long-term risks 146 arising from such fractions remaining in soils.

147 In contrast, it has been argued that long-term exposure to NER in soil may be harmful to 148 receptors. For instance, enhanced PAH mobilisation has been speculated in long-term aged 149 (50 years) soils from manufactured gas plant sites (MGP) under anoxic conditions and other changing environmental conditions such as temperature <sup>55, 59, 60</sup>. Some studies also reported 150 151 that higher organisms may have the capacity to access HOC residues, which were hitherto classified as inaccessible by microbial processes; this is due to differences in uptake 152 153 mechanisms and exposure conditions such as gut conditions of specific organisms, including humans <sup>38, 47, 61-65</sup>. However, none of these approaches would be a measure of the 154

bioaccessible fraction. Mayer et al. 65 also noted that it was important to investigate the 155 156 potential availability of strongly-sorbed contaminant fractions to non-microbial receptors such 157 as humans using *in vitro* gastrointestinal (GI) models, to further support the consideration of 158 bioavailability and the risk-based approach within regulatory structures. In addition, it has 159 been advised that regardless of whether risk-based land management (RBLM) approaches 160 are adopted, monitoring will be useful to check contaminant remobilisation should there be changes in land use or environmental conditions <sup>32</sup>. Such an approach is commonly used as 161 part of monitored natural attenuation of groundwater or contaminated soils <sup>66-73</sup>. 162

163 These contrasting arguments raise the questions:

164 (a) should RA be about the present, rather than the future?

165 (b) what are the residual and bioaccessible HOC fractions in soils?

166 (c) how they should be defined, classified, and measured? and

(d) whether remobilisation of residual HOCs in soil and its associated potential risks are
 exposure-specific, for example, upon contact of residues with the GI fluid.

169 To date, there have been only a few published studies that have reported potential 170 remobilisation of HOC residues in long-term aged or historically contaminated soils, and potential risks to receptors <sup>50, 55</sup>. These studies have mostly focused on ecological receptors, 171 172 especially microbes and earthworms. Other sensitive receptors, especially those that 173 simulate human physiology, are yet to be studied. The focus of much of the studies using 174 animal models has been the assessment of bioavailable fractions rather than the more 175 tightly bound residual fractions. While there have been studies that have attempted to 176 elucidate soil-contaminant relationships using advanced molecular based techniques, such 177 as the NMR, they have not focused on the dynamics of release or retention of residual 178 HOCs in soil with respect to the combined effects of ageing, varying soil properties, environmental conditions and realistic exposure scenarios, or the controlling mechanisms <sup>56,</sup> 179 58 180

181 To minimise uncertainties associated with risk-based approaches, understanding the 182 influence of soil properties, ageing and exposure-specific factors on the potential release and harmful effects of HOC residues in soil is necessary. Importantly, understanding the mechanisms controlling underlying processes, especially at a molecular scale, may complement previous empirical relationships describing the fate and behaviour of HOCs in soil. Within the context of RBLM approaches for soils contaminated with HOCs, the objective of this article is to critically review existing information on the bioaccessibility of HOC residues in an attempt to establish whether there is a need to further investigate potential, or the likelihood of harm being caused by highly sequestered residues in soil.

#### 190 2. Fate and behaviour of HOCs in soil

191 The fate and behaviour of HOCs, such as PAHs, in soil has been studied for more than two decades <sup>21, 29, 74, 75</sup>. Following their addition to soil, PAHs are subjected to various physio-192 193 chemical, biological, and environmental processes that are summarised in Fig. S1 in the supplementary material <sup>21, 29, 74, 76, 77</sup>. In the longer term, PAHs in soil are subject to the 194 195 'ageing effect' <sup>19, 22, 75, 78-81</sup> that results in significant partitioning of PAHs within the soil 196 leading to a reduction in mobility, extractability (bioaccessibility), and bioavailability. Over 197 time, HOC in soil becomes less available to exposed biological membranes (Fig. 1). The 198 ageing effect is due to the increasing HOC interactions with heterogeneous soil matrix 199 components, including mineral and organic matter and their associated pore structures and spaces <sup>27, 82, 83</sup>. Persistent HOCs diffuse into micro- and nano-interstitial spaces within the 200 201 soil matrix and interact through a number of potential physical and physiochemical processes with soil components <sup>25, 27, 54</sup>. The behaviour is termed 'sorption' and results in 202 203 weak or strong HOC retention in soil <sup>22, 82</sup>, depending on the key soil properties that influence the sorption at a particular time <sup>26, 27, 75, 84-89</sup>. Not only do soil-related properties influence 204 sorption, but the physicochemical properties of the HOCs (Table S1 in supplementary 205 material) may also influence the process as well 26, 89. In addition, the degree of 206 207 hydrophobicity or aqueous solubility may indicate whether HOCs associate with the organic 208 or mineral solid phases of the soil matrix, and could influence associated rates and extents 209 of sorption and desorption in soil.

2.1. Mechanisms controlling sequestration of HOCs in soils: Implications for mobility andbioaccessibility

The heterogeneity of soils at macro- (aggregate) and micro-(particle) scales may influence HOC accessibility and potential risk upon biological exposure <sup>19, 27, 82, 85, 87, 88, 90</sup>. However, modifying processes may influence risks due to exposure (Fig. S1 in supplementary material). Sorption and desorption play major roles in controlling HOC bioaccessibility in soil <sup>26, 27, 60, 82, 91, 92</sup>. Soil organic matter and mineral heterogeneity play significant roles in dictating mechanisms of sorption and desorption of contaminants, as they may control rates of diffusion of the contaminants in the soil matrix <sup>27, 93</sup>.

219 Isotherms fitted to sorption models (e.g. Freundlich and Langmuir, among other models) have shown that sorption is mostly non-linear as solution concentrations increase <sup>85</sup>. The 220 221 non-linearity index (n) is reported as less than unity, indicating heterogeneity of sorption sites in soil <sup>88, 94</sup>. Two sequestration phases are likely within soil, i.e. external (as influenced by 222 advection and diffusion) and internal (influenced by diffusion and adsorption)<sup>82</sup>. While the 223 former is unlikely to be rate-limiting in determining slow sorption and especially slow 224 desorption, the latter is likely <sup>27, 82, 84, 85, 95, 96</sup>. In a particle of soil, a HOC molecule could be 225 226 small enough that it fits into minute pore spaces, following diffusion through the dissolved 227 water phase. Such a tortuous journey occurs with increased ageing such that contaminant remobilisation is unlikely<sup>84</sup>, unless influenced by changing environmental conditions. Soil 228 conditions such as absence of oxygen, increased temperature, and soil mixing may 229 230 potentially influence the release of previously sorbed HOCs via a tortuous and reverse-231 phase journey (i.e. from the sorption sites within the soil matrix back to the dissolved water phase or soil surface) 59, 60, 97. 232

233 Pignatello and Xing <sup>82</sup> explained that to better understand mechanisms of slow sorption,
234 it was important to consider the following:

(a) That mass transfer processes including associated rate constants differ tortuous
 diffusion. This may be related to the diverse activation energies associated with
 different sorption sites within the soil matrix <sup>84, 85, 98</sup>;

(b) It appears that sorption becomes stronger with decreasing sorbate concentration.
 Sorption equilibrium of HOCs in soil is concentration-dependent <sup>99, 100</sup>; and

(c) The kinetic-hysterical nature of sorption-desorption processes, whereby the rate
 constants during slow sorption appear greater than slow desorption for the same HOC
 and soil. Hence, the rate and extent of formation of strong HOC-soil bonds becomes
 faster compared to breaking such bonds <sup>84, 85</sup>.

Models describing sorption and desorption mechanisms include diffusion in organic matter, sorption-retarded pore diffusion, intra-particle organic matter diffusion, pore deformation, and pore swelling, among others. These mechanisms may be stand-alone or synergistic in different situations; detailed reviews have been presented elsewhere <sup>27, 38, 39, 82, 101-104</sup>.

Many researchers have focused on sorption and associated mechanisms. Of more importance, however, is desorption which dictates contaminant mobility, accessibility and potential risks. Hence, understanding desorption of HOCs in soil, especially from residual pools, including associated mechanisms is important. Nonetheless, because of the interplay between both processes in defining rates and extents of HOC release from soil, and hence, potential risks, both processes are discussed in detail in subsequent sections.

254 2.1.1. Sorption of HOCs by soil and contaminant pool classification

Sorption plays an important role in influencing HOC bioavailability, and hence, associated mobility, transport, loss, and biological effects such as bioaccumulation, biomagnification and biotransformation <sup>82, 84, 105, 106</sup>. HOC pools in soil can be segregated into fractions ranging from that which is weakly sorbed to the strongly sorbed <sup>22, 27</sup>. The continuum may comprise dissolved, weakly sorbed and rapidly desorbed, the slowly desorbed, and the strongly bound fractions (Fig. 2). Although, this contaminant sorption continuum is a simplification considering the complex and dynamic nature of soil-contaminant interactions, it helps understanding of complex processes <sup>25, 54, 82</sup>. These pools presents different barriers for HOC
desorption, mobility, and availability. For example, PAHs in dissolved and weakly sorbed
(rapidly desorbing) pools are likely to be readily accessible to exposed organisms <sup>10, 12, 15, 107,</sup>
However, the strongly sorbed and bound (slowly desorbing) pool is presumed to be less
bioavailable <sup>43, 50, 109</sup>.

267 Earlier studies have mostly focused on using sorption isotherms (e.g. Freundlich and 268 Langmuir) and distribution coefficients ( $K_d$ ) in describing the mostly non-linear HOC sorption behaviour in soil<sup>85, 110</sup>. These have resulted in proposing empirical relationships between 269 distribution and partitioning coefficients (Log  $K_d$  or Log  $K_{oc}$ ) of different HOCs and soil 270 properties <sup>88, 94, 111-114</sup>. However, the sole use of Log  $K_{oc}$  or octanol-water partition coefficient 271 272 (Log  $K_{ow}$ ) to describe HOC sorption in soil should be used with caution, as other intrinsic soil 273 properties such as SOM-clay mineral interactions may play greater roles in influencing sorption processes <sup>113, 115</sup>. In addition, while these empirical relationships are valuable and 274 275 have formed the basis for emerging investigations, inferences regarding the relationship 276 between soil properties and HOC sorption behaviour have been based on indirect observations <sup>56</sup>. 277

278 Of the current advanced analytical techniques, NMR has been reported to provide 279 mechanistic information on molecular-scale soil-contaminant interactions at the solid, liquid, and soil-water (gel) phases <sup>56, 57</sup>. In many NMR investigations, the soil is physically or 280 281 chemically fractionated into operationally defined pools in an attempt to obtaining direct molecular information about SOM composition 57, 116-120 and understand precise HOC 282 sorption behaviours in soil <sup>99</sup>. Although, the actual nature of the SOM is still contentious <sup>121</sup>, 283 284 these investigations provide detailed understanding on the dynamics of SOM-HOC 285 interactions. However, physical or chemical fractionation is likely to change the behaviour of 286 HOCs. Some NMR investigations of soil-contaminant interactions have been reviewed elsewhere <sup>56, 58</sup>. However, while applications of NMR spectroscopy in soil science are 287 288 emerging, investigations towards understanding HOC sorption mechanisms especially for 289 historically contaminated soils are still lacking. Therefore, molecular level understanding of

the long-term stability of strongly sorbed HOC residues in historically contaminated soils would help minimise uncertainties relating to risk based approaches to managing contaminated soils. Such investigations should employ both empirical and advanced molecular-based techniques in a complementary and multi-tiered manner to provide robust evidence on the long-term stability of highly sequestered contaminant residues in soil.

295 2.1.1.1. HOC residual pools in soils

296 The total contaminant pool is made up of potentially labile and non-labile pools (Figs. 1 297 and 2). While the labile fraction consists of the dissolved and rapidly desorbable fractions, 298 the non-labile component is comprised of the non-desorbable and irreversibly sorbed or recalcitrant fractions <sup>10, 79, 107, 122, 123</sup>. There are no methods for the determination of non-labile 299 300 residual HOC fractions, especially relating to changing exposure scenarios and estimations 301 of harm. Furthermore, a residue which may not be accessible to microorganisms may be 302 accessible to other living organisms such as earthworms, plants and other higher forms 303 including humans due to differences in the mechanisms of contaminant uptake and release <sup>32, 62, 124</sup>. Specifically in relation to HHRA, that fraction of the mobilised or dissolved 304 305 contaminant in the gut, following ingestion, which is absorbed and may cause harm is the bioavailable fraction <sup>28</sup> and the un-mobilised fraction is referred to as the residue <sup>125</sup>. 306 307 However, there seems to be confusion regarding what the residual fraction in soil really is. It 308 could be the fraction remaining after the complete release of the labile fraction (measured by 309 an in vitro technique) or the residues from historically contaminated soils. The NER, 310 especially in EHRA, is mostly regarded as the fraction that is left behind after solvent extraction <sup>126</sup>. Tao et al. <sup>61, 127</sup> suggested that the difference between the concentration of 311 312 PAH and organochlorine pesticides mobilised by an in vitro digestion model (which was 313 greater) and that extracted by accelerated solvent extraction represents a part of the residual fraction. On the other hand, Juhasz et al. <sup>62</sup> suggested that the residual fraction was that 314 315 which remained after long-term ongoing bioremediation of a field-contaminated soil; this description appears similar to that in Thavamani et al. <sup>30</sup>. While the former description of the 316

residual fraction may be more relevant for HHRA purposes, the extracting medium shouldmimic the GI fluid.

A residual fraction from an EHRA perspective may differ from an HHRA perspective with respect to concentrations and specific receptors. Hence, while the use of highly sequestered fraction may be relevant for EHRA purposes <sup>107</sup>, its relevance for HHRA purposes must also be demonstrated <sup>19</sup>. To the regulator, a remediated soil is one in which risk from exposure to remaining or residual fraction is reduced to an acceptable level which may depend on ultimate end use.

325 2.1.2. Desorption of HOCs in soil

326 2.1.2.1. Slow desorption of HOCs in long-term contaminated soils

327 The bioavailability of HOCs in soil following exposure is limited by its desorption kinetics 328 <sup>97, 128</sup>. Desorption rates and extents may influence various contaminant loss processes such 329 as biodegradation, uptake and bioaccumulation among others, and are therefore key factors 330 for RA. Carmichael and others investigated whether microbial mineralisation of PAHs in soil is limited by their desorption rates <sup>91</sup>. Using <sup>14</sup>C-radiolabelling, they examined the desorption 331 332 kinetics and microbial mineralisation kinetics of freshly spiked <sup>14</sup>C- and aged <sup>12</sup>C-333 phenanthrene and chrysene in two soils from a tar and oil-recycling sites. The key findings 334 were that the rates of desorption were faster than mineralisation rates in <sup>14</sup>C-PAH spiked 335 contaminated soils, however, the mineralisation rates of native PAHs in the organic carbon-336 rich contaminated soil were faster than or equal to their desorption rates. Hence, the rates 337 and extents of PAH desorption in historically-contaminated soils may represent a limitation to 338 their biodegradation. This may be one reason why PAHs persist in microbially-active soils <sup>129</sup>. Based on their results, the sole use of <sup>14</sup>C-radiolabels in describing fate and behaviour of 339 340 PAHs in soil should be applied with caution <sup>91</sup>.

One study combined both <sup>14</sup>C- and <sup>12</sup>C- techniques to monitor the long-term fate and behaviour of HOCs in soil <sup>50</sup>. Extractability was observed to be low and desorption very slow (calculated half-lives of up to 38 years for fluoranthene), which supports the notion that risks

from exposure will be minimal <sup>31, 55</sup>. However, it was pointed out that the results were only 344 345 applicable to contaminated agricultural soils used. They recommended that similar investigations be conducted on historically-contaminated soils from different sources <sup>50</sup>. It is 346 347 further suggested here that different exposure scenarios and other key risk endpoints, such 348 as earthworm bioaccumulation, plant uptake, and animal-based bioavailability and in vitro 349 bioaccessibility assays, should be considered in addition to microbial degradation and 350 chemical extractability for a robust conclusion. For instance, following maximum desorption 351 of labile HOCs in field-contaminated sediments, slow intercompartmental reorganisation and remobilisation of HOCs have been observed <sup>46</sup>. It is also known that gastric, and intestinal 352 353 fluids especially bile and other intestinal enzymes, can influence the solubility and mobilisation of soil-associated HOCs <sup>61, 125, 130</sup>. Such gastrointestinal effects may influence 354 355 HOC desorption behaviour from residual pools and their remobilisation. Hence, considering 356 other exposure scenarios, other than microbial-specific ones, may further demonstrate and 357 authenticate the fact that historically aged HOCs in field-contaminated soils are mostly 358 residual and slowly desorbing, and as a result, pose minimal risks. Again, application of 359 direct molecular-level tools in this regard may be useful.

360 2.1.2.2. Desorption phases of HOCs and associated kinetics in soil

Studies using artificially-spiked and historically-contaminated soils have described HOC desorption in two or three-phases <sup>26, 49, 52, 60, 83, 91, 131-133</sup>. While biphasic desorption comprise both rapid ( $F_{rapid}$ ) and slowly ( $F_{slow}$ ) desorbing fractions including associated rate constants ( $k_{rapid}$  and  $k_{slow}$  respectively), triphasic desorption includes a very slowly desorbing fraction and rate constant ( $F_{very slow}$  and  $k_{very slow}$ ). However, considering the heterogeneity and complexity of soil-HOC interactions in the field, it is likely biphasic and triphasic models are simplifications of multiple desorption phases in reality <sup>134</sup>.

To describe desorption kinetics of HOCs in different soils and humic materials, various models that assume different desorption phases have been used  $^{26, 49, 83, 134, 135}$ . The key observations from these studies were that the 3-parameter biphasic (i.e. one that assumes quick equilibration for  $F_{rapid}$ ) and 3-parameter biphasic polymer diffusion models were 372 recommended as preferable for predicting HOC desorption kinetics in contaminated soils 373 compared to others, although rate limiting site-specific conditions must be considered. 374 Importantly, these studies revealed that rates of slow desorption ( $k_{slow}$ ) from strongly sorbed pools could range from between 10<sup>-2</sup> to 10<sup>-4</sup> h<sup>-1 26, 49, 134</sup>, depending on stage of SOM 375 diagenesis and hydrophobicity of the HOCs of concern 37, 136, 137. Generally,  $k_{slow}$  was 376 reported to be up to 2 orders of magnitude lower than  $k_{rapid}$  <sup>50</sup>. Similarly, the highly 377 378 condensed soil or humic material (e.g. kerogen-like shale) tends to retain a greater 379 percentage (up to 70%) of the residual HOC fractions <sup>49, 134</sup>. This residual fraction may increase by up to 2 - 10 folds with ageing <sup>26, 53, 54</sup>. These observations support the argument 380 that long-term exposure to NER fraction may pose no risk <sup>31</sup>. However, a complementary 381 382 molecular-scale approach to providing such evidence for different exposure scenarios is still necessarv<sup>56, 57</sup>. 383

### 384 2.2. Biodegradation of HOCs in historically contaminated soils

Biodegradation influences the fate and behaviour of HOCs in soil <sup>12, 138</sup>. However, HOCs 385 386 sequestration happens along with the biodegradation process. Biodegradation is often 387 observed as a biphasic process which occurs in two phases, i.e. fast and slow loss phases. 388 Due to its biphasic nature, biodegradation may not necessarily lead to the total removal of 389 contaminants in soil, but may result in transformations that lead to reductions in initial 390 concentrations, mobility and toxicity of contaminants, thereby, reducing the level of harm to flora and fauna <sup>139-141</sup>. Bacteria, fungi and algae play important roles in the biotransformation 391 and biodegradation of contaminants in soil <sup>142-144</sup>. Generally, for biodegradation to occur, 392 393 microorganisms with sufficient capacity to degrade the contaminant of concern (i.e. 394 possesses relevant degradation enzymes) should be present in soil; the contaminant must 395 be bioavailable or become bioavailable, and environmental factors must permit the biodegradation process <sup>133, 142, 145, 146</sup>. Microbial mineralisation of HOCs is limited by HOCs' 396 397 desorption kinetics, especially in long-term contaminated soils, thereby supporting its 398 biphasic nature <sup>91</sup>. Therefore, bioremediation (cost-effective remediation technique relying on

optimising contaminant biodegradation) is sometimes constrained due to sorption limitations which limits microbial accessibility to HOCs in aged soils leaving behind residual contamination in soil or sediment <sup>29, 47</sup>. Different mechanisms through which microorganisms circumvent accessibility limitations include attachment to soil surfaces to allow HOCs partitioning from soil into microbial cell membrane, co-metabolism, and production of extracellular enzymes and biosurfactants to enhance HOCs mobilisation from sorbed phases to microbially-accessible spaces in soil <sup>145, 147</sup>.

406 The lingering question is whether the residual contaminant fraction (i.e. the fraction that is 407 not biodegradable or extractable/mobile/bioaccessible) in soil could potentially cause harm? With regards to biological uptake of contaminants, microbial inaccessibility of HOCs in 408 409 historically contaminated soil may not mean inaccessibility by other exposed higher organisms, as contaminant accessibility is species and matrix dependent <sup>62, 133</sup>. A 410 411 contaminant must desorb or be bioaccessible to allow biological uptake. Hence, like 412 sorption/desorption processes, different soil and organism properties may modify 413 contaminant uptake in soil. Other abiotic loss processes such as volatilisation, photolysis, 414 leaching, and chemical transformations may also affect fate and behaviour of HOCs in 415 historically contaminated soils (Fig. S1 in supplementary material). However, these effects 416 are mostly insignificant compared to losses by biodegradation, especially for very hydrophobic HOCs <sup>29</sup>. 417

418 3. Bioavailability and bioaccessibility of HOCs in soil

The concept of HOC availability has increasingly attracted diverse discussions within the scientific and regulatory community <sup>11, 107, 148</sup>. The concept has been used differently in specific fields such as toxicokinetics, pharmacokinetics, agriculture, and environmental science among others <sup>11, 22, 77, 149</sup>. The terms 'bioavailability' and 'bioaccessibility' have mostly been used interchangeably in the literature <sup>150, 151</sup>, within EHRA and HHRA. However, the terms are different but related measures <sup>15, 77</sup>. Specifically for HHRA, Hack and Selenka <sup>125</sup> and Ruby et al. <sup>151</sup> considered the bioaccessible contaminant fraction as the dissolved and

426 mobilisable fraction within simulated GI fluid, following ingestion of HOC-laden soil. This 427 bioaccessible fraction, therefore, represents the contaminant fraction maximally available for 428 intestinal absorption (Fig. 3). For EHRA, the different definitions proposed for bioavailability 429 and bioaccessibility has been compared and reviewed elsewhere <sup>77, 149</sup>.

A quick citation analysis of the definitions by Peijnenburg and Jager <sup>152</sup>, Semple et al. <sup>15</sup> 430 and Reichenberg and Mayer et al.<sup>153</sup> which are mostly used for EHRA is presented in Fig. 4. 431 432 The analysis was performed by comparing the number of times each definition was cited as 433 revealed from Web of science, Scopus, and Google Scholar on 02 December 2015. The 434 definitions provided by Semple et al. and Reichenberg and Mayer had been more cited. However, this analysis does not suggest actual uses of the bioavailability definitions within 435 the scientific and regulatory communities. The recent European Centre for Ecotoxicology 436 and Toxicology of Chemicals (ECETOC) workshops <sup>35, 36, 107</sup> adopted the bioavailability and 437 bioaccessibility concept by Semple and others <sup>15</sup>. Recently, the mostly used definitions of 438 contaminant availability in RA were articulated <sup>10</sup>, as shown in Fig. 5. Hence, bioavailability 439 440 and bioaccessibility in this article and with respect to EHRA follows definitions by Semple et 441 al. and Reichenberg and Mayer.

442 3.1. Measurement techniques for bioaccessible and residual HOC fractions in soil

Various techniques have been developed for HOC bioavailability or bioaccessibility measurement, and are presented in Table 1. However, there is no consensus on the choice technique. While it has been documented that a few methods are being standardised for ecological receptors such as microbes <sup>10</sup>, there is an urgent need to develop and validate SOPs for HOC bioaccessibility in soil, especially for HHRA.

Ultimately, a biological experiment is needed to study HOC bioavailability in soil. For example, if humans are the most sensitive receptors for a particular RA procedure, then a human or higher vertebrate model will be most appropriate test organism. However, this approach is unethical and in fact, there have been moves away from animal-based testing <sup>154, 155</sup>. Also, animal-based studies are expensive, time consuming, and may have challenges

with reproducibility <sup>33</sup>. Hence, over the past few decades, focus has shifted to development 453 454 and validation of simple and complex in vitro, including in silico, approaches for contaminant 455 bioavailability or bioaccessibility assessments. Ideally, the expectation for these in vitro 456 approaches are that they serve as uncomplicated bioavailability surrogates for the 457 environmental receptor of concern. This implies that for any realistic applications in RA, 458 these alternative approaches must be validated and standardised. However, while much 459 efforts has been focused on the development side, less efforts has been devoted to 460 validation of in vitro approaches for use in reality. It is therefore suggested that while 461 development of innovative bioavailability techniques are necessary, validation and 462 standardisation of existing promising in vitro methods are important for minimising 463 uncertainties in RA.

## 464 3.1.1. Brief classification of techniques

465 The approaches for determining bioavailability and bioaccessibility can either be classified as biological (in vivo), chemical extraction (in vitro) or computer modelling (in 466 467 silico). A clear distinction has to be made between the actual bioaccessible fraction and the 468 residual fraction as has been previously emphasised. Most currently developed in vitro 469 methods have focused on microbial degradation and earthworm bioaccumulation as ecological endpoints, hence, various biodegradation and ecotoxicity assays are available <sup>30</sup>. 470 471 However, significant efforts at developing *in vitro* bioaccessibility assays for HOCs, such as 472 PAHs, in soil especially for HHRA is only beginning to gain attention.

473 A detailed review of simple chemical extraction techniques for measuring bioaccessibility 474 for specified ecological endpoints can be found elsewhere <sup>12, 77, 133, 156, 157</sup>. Some of these 475 methods include Soxhlet extraction, solid-phase extraction (SPE), negligibly depleting solid-476 phase micro-extraction (nd-SPME), hydroxypropyl-β-cyclodextrin (HPCD) extraction, 477 supercritical fluid extraction (SFE), use of complexation or solubilisation agents and 478 surfactants, microwave extraction, persulphate oxidation, thermodesorption, soil-sediment 479 availability ratio (SARA), use of polyethylene devices (PEDs), semipermeable membrane

devices (SPMDs), polyoxymethylene (POM) sampler, and thin ethylene vinyl acetate (EVA).
While the *in silico* approaches for bioaccessibility calculations and predictions with computerbased programs could be potentially useful <sup>158</sup>, significant developments may be limited by
the quality and quantity of reliable bioavailability data from laboratory or field experiments.

484 None of the methods summarised above provide a clear measure of residual fraction. 485 Given this scenario, the most likely approach to assessing potential risks arising from such a 486 fraction may be via toxicological studies that expose sensitive receptors to such fractions. 487 Whether these highly sequestered fractions do get released in the gut or are bioaccumulated 488 in organisms including earth worms or plants may then be investigated.

489 3.1.2. Exhaustive extraction techniques (EETs) versus non-exhaustive extraction
490 techniques (NEETs)

491 The feasibility of using chemical extractions to predict the bioavailability or bioaccessibility 492 of HOCs and to determine associated residues in soil has been proposed and widely 493 discussed. Mostly, apolar and polar solvents of varying extraction strengths have been used. 494 Harsh extractions (or EETs) use strong solvents such as dichloromethane (DCM), hexane, acetone, and toluene <sup>159</sup>. Mild extractions (or NEETs) make use of weaker solvents (or mild 495 solvent - water mixture) such as methanol, *n*-butanol or propanol <sup>160</sup>. The extraction with 496 497 EETs and NEETs simply involves thorough mixing of a contaminated soil sample with 498 solvent for a given duration. Analysis of the extract is then performed by chromatography (HPLC or GC), mass spectrometry, spectrophotometry, or radio-isotopic analysis <sup>22, 133, 159,</sup> 499 161, 162 500

501 Exhaustive extractions generally overestimate the bioavailable contaminant fractions. As 502 a result, they are over-predictive of actual risks and often seen as too conservative and as a 503 consequence are likely to result in a technically difficult and expensive remediation of 504 contaminated sites <sup>75, 159, 161, 163, 164</sup>. The effect of ageing (120 d) on HOCs extractability was 505 unchanged using DCM, while it is well known that ageing results in a decrease in 506 bioavailability of HOCs in soil <sup>161</sup>. Kelsey et al. <sup>159</sup> showed that DCM extractions failed to

507 predict the availability of phenanthrene and atrazine to specialized bacteria and earthworms 508 over a 120 d period, while better prediction was achieved when mild extractions (*n*-butanol 509 and methanol-H<sub>2</sub>O) were used <sup>159</sup>. Similarly, a strong correlation was reported between 510 butanol extractability and microbial bioavailability for phenanthrene ( $R^2 = 0.971$ ; y = 0.952x + 0.943) and pyrene ( $R^2 = 0.994$ ; y = 0.998x - 0.990) in artificially spiked soil <sup>165</sup>. These studies 511 512 support the use of NEETs, rather than EETs, to predict HOCs bioavailability in soil. In 513 addition, the use of harsh and mild solvents for HOC bioaccessibility determination in RA 514 has been deemed unreliable, for example in Germany, since they do not possess a clear mechanistic basis <sup>108</sup>. 515

516 Advanced NEET methods exploit HOCs' hydrophobicity to allow their partitioning or 517 mobilisation towards a hydrophobic aqueous or solid phase within a soil or sediment 518 suspension. Advanced NEETs include an aqueous-based technique such as hydroxylpropyl-519 beta-cyclodextrin (HPCD) for extraction of HOCs in soil especially low molecular weight (LMW) PAHs<sup>164, 166, 167</sup>. They also include solid-phase adsorbents, such as SPME or Tenax, 520 521 for extraction of HOCs in sediments and soil-water suspensions <sup>26, 64, 168</sup>. Various studies 522 have shown that HOC extractability from a wide range of contaminated soils using these 523 techniques often correlates with different ecological endpoints such as microbial degradation, earthworm bioaccumulation, and plant uptake <sup>53, 64, 169, 170</sup>. Nonetheless, these 524 525 techniques have also shown poor HOCs bioaccessibility correlations especially when a nonmicrobial receptor is considered <sup>19, 64, 79, 171</sup>. Factors which may affect bioavailability such as 526 527 ageing, soil properties among others are presented in Table S2 in the supplementary material and have been reviewed in detail elsewhere <sup>75, 77, 133, 161, 172, 173</sup>. 528

529 3.1.3. Can the desorption-resistant fraction be measured or estimated?

530 Desorption techniques are presented in Table 1 and are further discussed below. These 531 techniques have different guiding principles and none has been standardised to date. They 532 have mostly been devoted to measuring the leachable, extractable, bioaccessible fractions 533 using microbes and earthworms among other ecological receptors, as sensitive endpoints. 534 However, measurements of the bioavailable and residual pools, as currently delineated, is

problematic <sup>43</sup>. The residual fraction may not simply be the total *minus* the bioavailable or 535 bioaccessible fraction. In a study by Cuypers et al. <sup>129</sup>, it was observed that persulphate 536 537 oxidation of soils (n = 7) and sediments (n = 7) under-predicted the biodegradable fractions 538 of 5 and 6 rings PAH. In contrast, the residual fractions after oxidation and biodegradation 539 agree with an almost 1:1 correlation. It is not clear if desorption and bioavailability which impact biodegradation are similar to those for a toxic endpoint. According to ISO 17402<sup>174</sup> 540 and as documented by Kordel et al. <sup>108</sup>, there are practical differences between 541 542 'environmental availability, environmental bioavailability, and toxicological bioavailability'. If 543 these do differ, it may imply that residual fractions may also vary for biodegradation and 544 toxicological endpoints, and between EHRA and HHRA. However, to predict the significance 545 of harm that may be caused from exposure, desorbable concentration from residual fractions 546 in soil must then be considered.

547 Recently, Mayer and others developed an isolation and quantification technique for the so called 'desorption-resistant' or residual fraction using a contaminant trap method <sup>65</sup>. The 548 549 contaminant trap was simply a PDMS-activated carbon sink and a cyclodextrin diffusive 550 carrier in a custom-made un-agitated reactor. In this study, 12 PAHs in two industrial-551 contaminated soils from different sources were studied; a contaminated material dumpsite in 552 Austria (4.6% organic carbon, OC, w/w) and a former historically-contaminated MGP site 553 (1.5% OC w/w). The soils were incubated in the trap for 1, 7, 31 and 92 d, and for 4, 7, 14, 554 28, 56, 77 and 92 d for desorption and biodegradation experiments respectively. After soil 555 slurry filtration following desorption and biodegradation, the filtrate and residual samples 556 were exhaustively extracted and analysed for PAH concentrations using HPLC-FLD and 557 reverse phase HPLC-FLD respectively, and then compared. The elimination kinetics and 558 trapping efficiency of the contaminant trap for the PAHs in soil were reportedly adequate, and improved if the traps were agitated <sup>65</sup>. Although, the residual concentrations of individual 559 and total PAHs quantified by the trap in the first soil were less than those analysed after 560 561 biodegradation, desorption limits by the trap and biodegradation limits plateaued similarly 562 above the Danish regulatory limit of 40 mg/kg dry weight of soil. However, PAH desorption

563 and biodegradation in the second soil was hardly noticed, as both processes were in good 564 agreement. The PAHs were strongly sorbed in both soils, however, the soil with lower 565 amounts of OC exhibited stronger sorption resulting in smaller biodegradative losses and 566 greater residual PAH concentration, compared to the soil richer in OC. The authors noted 567 that the quality (origin, type, and composition) of OC and pollution history of soil, rather than quantity of OC determines HOCs sorption behaviour <sup>65</sup>. This observation further revealed the 568 569 importance of using direct molecular-level techniques with the capacity for such information 570 to complement empirical approaches, for a robust description of HOC sorption-desorption 571 behaviour in soil.

572 An improved extraction method was developed which included a silicone rod as an adsorption sink to optimise mass transfer, and HPCD as diffusive carrier <sup>175</sup>. This silicone-573 574 based extraction (SBE) method was based on the principle that after desorption from the 575 solid to aqueous phase in soil, uptake by a microbial receptor creates a gradient which 576 facilitates mass transfer processes until equilibrium is reached. In the SBE method, the 577 silicone rod acted as a quasi-reservoir, rather than a trap, such that sorbed residues could 578 be back-extracted. Using the SBE method, the researchers evaluated desorption of PAHs 579 from wood soot (50 mg), as the sorbent matrix, with initial PAH concentration (mg/kg) 580 ranging from 7.02  $\pm$  0.22 - 108.54  $\pm$  3.36 (median and total PAHs concentration = 55.47  $\pm$ 581 1.42 mg/kg and 646.96 ± 19.95 mg/kg respectively). The mobilised and desorbed fraction 582 were compared in the presence and absence of sink (i.e. HPCD only). In comparison with 583 the HPCD only extraction, SBE extracts contained up to 3 - 25 times greater individual PAH 584 concentrations. Although, extractions using the SBE were compared with HPCD extractions, 585 direct comparison of SBE extractable and associated residue with the microbially degradable 586 fractions and residues may be more useful, as in Mayer et al. 65. However, the SBE 587 procedure seems complex and may be technically demanding, and does not demonstrate 588 the relevance of the derived residual fractions to HHRA.

589 It appears that only a few studies have focussed on understanding HOCs desorption or 590 extractability from residual pools in soil, especially with respect to HHRA <sup>61, 62, 127, 176</sup>. A

591 reliable measurement approach for the desorption-resistant fraction may be very useful in 592 RA. For example, such methods may be needed to test the effectiveness of techniques 593 which are based on extraction of the labile pools, in that the residue remaining after 594 extraction is investigated for further extractability. Such approach, may then reveal whether 595 the residual contaminants are actually bound in soil or could be potentially mobilised. Hence, 596 there is a need for further investigation into the development of a simple, reliable, robust, 597 adjustable, and exposure-relevant technique for delineating and quantifying residual HOC 598 pools in a wide range of soils.

599 3.1.3.1. Can chemical extractions be used to classify HOC residues in soil?

600 In a recent ECETOC report, extractions using mild extractions (including NEETs) and EETs were denoted as extractable and NER fractions <sup>35</sup>. The 'NER' term as used implies not 601 602 extractable by mild extractions, but by exhaustive extractions. However, this 'NER' term could be a misnomer and its use is confusing as a contaminant fraction is either extractable 603 604 or not. Hence, NER is referred to as a highly sequestered contaminant fraction in this article. 605 The residues left after exhaustive extractions which can only be assessed using <sup>14</sup>Cradiocombustion methods were referred to as bound residues <sup>35</sup>. Although the residue 606 607 classification scheme was only relevant for EHRA, it appears superficial and may not even 608 be appropriate for all ecological receptors. This is because the proposed scheme did not 609 fully consider issues such as relationships between reversibility of sorbed HOCs, physico-610 chemical properties of HOCs and the soil matrix, and physiological variations within complex 611 organisms such as plants and aquatic organisms, let alone humans. Therefore, the 612 development of an 'intelligent extraction strategy framework' which considers some of the aforementioned issues has been proposed <sup>107</sup>. Such an approach must enable risk 613 614 assessors including regulators to predict with confidence whether highly sequestered 615 fractions do pose ongoing risks from long-term exposure to historically contaminated soils. It 616 is therefore important to re-emphasise the need to investigate the differences in mechanisms 617 of desorption and uptake from residual HOC pools in soils by key organism classes, and to 618 conclude whether these differences can be measured and/or described using chemical

619 extractions. Such investigations may help in properly delineating residual and non-residual
620 HOC pools in soil, and help minimise uncertainties associated with RA.

3.2. General comments on the bioavailability or bioaccessibility of highly sequestered
 residues in soil and their implications for RA

623 There are obvious challenges associated with the incorporation of bioavailability or 624 bioaccessibility concepts within current regulatory structures including the lack of precision 625 and inconsistency in the use of the bioavailability and bioaccessibility terms; confusion 626 regarding what is actually being measured; inadequate systematic comparisons to reveal the 627 most reliable prediction methods for bioavailability or bioaccessibility after considering 628 contaminant-organism-matrix differences; contaminant availability prediction methods that 629 are not validated; and scarcity of clear guidance on how the contaminant availability concept 630 can be incorporated into current RA frameworks. While bioavailability underpins a risk based 631 approach, most regulators and risk assessors persist with total extractable concentration. 632 Unlike toxic metals for which the Australian National Environment Protection Measure 633 (NEPM) and the United States Environment Protection Agency (USEPA) includes standard operating procedures for assessment of the bioavailable fraction <sup>34, 177</sup>, there are still no 634 635 SOPs for HOCs. Comparison of studies may be difficult and caution must be applied when 636 interpreting HOCs bioavailability data generated from the different techniques. However, 637 both HPCD and TENAX extractions seem promising for standardised use for HOC bioavailability determinations in soil and/or sediments for EHRA purposes <sup>10</sup>. The former is 638 simpler and time-efficient compared to the latter <sup>53</sup>. On the other hand, only a few studies 639 640 have investigated PAH in vitro bioaccessibility for HHRA purposes and have been reviewed in detail elsewhere <sup>178-182</sup>. In addition, *in vitro* bioaccessibility-*in vivo* bioavailability 641 correlations for HOCs, such as PAHs, in soil have not been established. More so, 642 643 comparisons between PAH in vitro GI bioaccessibility and in vivo bioavailability are scarce, 644 revealing the paucity of data to allow such comparisons. However, these are likely to become subjects of future investigations <sup>19, 62, 183-186</sup>. 645

646 It may be worthwhile to subject a wide range of soils to single or sequential extractions 647 using aqueous, aqueous-based, solid-phase extraction, and sink-included in vitro GI 648 digestion techniques to differentiate between the labile and non-labile HOC fractions, as 649 validated using relevant receptors. Similarly, the non-labile/residual fractions obtained from 650 methods such as persulphate oxidation, SFE, and contaminant trap among other relevant 651 methods could be tested for remobilisation under weak to strong extraction regimes and 652 varying exposure conditions, for example under the influence of *in vitro* gut fluid. Further, the 653 labile and residual HOC fractions obtained using different techniques could be compared to 654 confirm if they had been obtained from the same (or similar) pools in soil. The use of 655 advanced analytical and empirical methods to understand the mechanisms controlling soil-656 HOC-organism interactions in these investigations should be key, and efforts should be 657 made to validate and standardise the choice methods. Such investigations may contribute to 658 understanding the kinetics, risks and associated significance of potentially desorbing HOC 659 fractions from non-residual and residual pools in soil, which may in turn be useful for RA.

## 660 4. Does exposure to residual HOC fractions in soil pose risks to receptors?

A recent study using a series of multi-species ecotoxicity assays revealed that exposure 661 662 to residual total petroleum hydrocarbons in field contaminated soils had no environmental risk <sup>30</sup>. Similarly, ecological risk from exposure to pesticide residues in soil is deemed 663 minimal <sup>43, 44</sup>. In addition, HOCs associated with diagenetically-advanced and recalcitrant 664 humic materials (e.g. black carbon and biochar) in soil are unlikely to be remobilised <sup>51</sup>. One 665 666 of the most important reasons advocated for such minimal or no risks is that aged HOCs are highly sequestered within the soil matrix <sup>21, 25, 31, 133</sup>. Further, it is interesting that in most 667 previous studies no significant relationship was reported between bioavailability and total 668 669 PAH concentrations (extractable) in soil, while a recent study that investigated the relative 670 bioavailability (RBA) of B[a]P in juvenile swine using 8 soils, and silica sand as reference 671 material, both spiked at the same B[a]P concentration found significant correlation between the RBA of B[a]P in spiked soils and DCM/ACE extractable fraction ( $R^2 = 0.67$ , p < 0.05) and 672

butanol extractable fraction ( $R^2 = 0.75$ , p < 0.01), but not HPCD or Milli-Q water extractable fractions <sup>19</sup>. Although, RBA of contaminants as measured by time-based blood concentrations following soil ingestion has been criticised <sup>178, 180</sup>, it may still be a reliable estimate of oral bioavailability compared to other bioavailability endpoints since systemic toxicity is mostly due to the effect of parent PAHs in systemic circulation <sup>183</sup>.

678 For clarity, it is important to carefully consider the following, bearing in mind previous discussions in this article. By ECETOC's classification, the total HOC residue (TR) 679 comprises the extractable (ER) + non-extractable residue (NER) + bound residue (BR), 680 which is based on Zarfl et al. delineation of TR in soil <sup>35, 122</sup>. According to the bioavailability 681 682 framework suggested by Ortega-Calvo and others for use in regulation and modified from an earlier study <sup>187</sup>, the total extractable residue (TER) = ER + NER <sup>10</sup>. Further, the TER 683 comprise the slowly and very slowly desorbing + rapidly desorbing + dissolved fractions <sup>188</sup>. 684 Also note that a single HPCD or Tenax extraction of  $\leq 20$  h measures the rapidly desorbing 685 fraction <sup>49, 108</sup>, whereas sequential extractions can also access the slowly desorbing pool <sup>49,</sup> 686 <sup>83</sup>. However, the TER has been defined as the fraction measurable by EETs, while an 687 apparent earthworm- or microbial-bioavailable fraction is measurable by NEET <sup>79</sup>; and it has 688 689 been frequently reported that RA which assumes that TER is bioavailable overestimate risks to humans, plants, and animals <sup>11, 15, 43</sup>. Hence, the implications of the reported correlation 690 between RBA and EETs, but not NEETs <sup>19</sup> are 3-fold: 691

(a) That the so called 'highly sequestered residue' may still be accessible upon human oranimal exposure to aged HOC-laden soil;

(b) That chemical stability of residues must be complemented by biological stability.
There may be a need to support the wide use of simple chemical extractions for
classifying bioavailable and residual HOC pools in soil, to EHRA and HHRA. Perhaps,
through validation with biological experiments and the complementary use of
advanced molecular techniques, while simple chemical extractions may be restricted
to uses in the EHRA, rather than the HHRA; and that

(c) There is a need for clearer characterisations of the residual and non-residual HOC
 pools in soil, and further investigations into the potential risks associated with
 consequent exposure to these residues.

703 5. Risk assessment framework for contaminated sites

704 The general RA framework for contaminated sites consist of a series of preliminary and 705 detailed phases of investigation which must be completed to determine the certainty of risk from exposure to a contaminated site <sup>108, 189, 190</sup>. The preliminary investigation phase (Tier 1) 706 707 includes site reconnaissance; design of a conceptual site model (CSM); and confirmation of 708 potential contamination by sampling soil and analysing its total extractable contaminant 709 concentrations (TER) for comparison with soil guideline values (SGVs). The detailed 710 investigation phases (Tiers 2 and 3) usually proceeds when TER exceeds the SGV 711 considered as safe. A detailed investigation of the sources, pathways, and receptor 712 relationships (or pollutant linkage) is then conducted and the CSM is subsequently modified 713 to reflect realistic exposure scenarios on the site. Establishing a significant pollutant linkage 714 implies that there is a risk, hence, such sites must be managed or remediated to minimise 715 the risks, protect receptors of concern, or permit proposed changes to land uses.

716 While such pollutant linkages could be established, associated human or ecological 717 health risks are frequently overestimated. This is because the total contaminant load 718 quantified on a site, at every point in time, is traditionally equated to potential risks that may 719 result following exposure <sup>10, 187</sup>. In fact, the criteria for developing SGVs do not consider contaminant-matrix-organism dynamics<sup>191</sup> which may modify biological exposure conditions 720 721 and consequent risks. This traditional approach for risk estimation by measuring total 722 contaminant load in soil (including overly-conservative regulatory guidance) has resulted in 723 high costs and slow pace of remediation, including dereliction of potentially reusable lands <sup>32</sup>. These challenges are constraints to efficient contaminated land management <sup>30</sup>. 724

Risk-based approaches can ensure cost- and time-effectiveness in contaminated land management and may have positive socio-economic implications such as rapid

727 redevelopment of derelict sites, but its regulatory adoption is largely constrained by 728 uncertainties as described previously. Perhaps, the most confounding problem may be the 729 unclear fate associated with the residual contaminant fraction in soil.

730 Bioavailability research has gained increasing attention, and the determination of 731 bioavailability and its consideration for incorporation into risk assessment policies is considered essential <sup>192</sup>. Clear framework guidance documents to support wide regulatory 732 adoption of the bioavailability concept have recently been published <sup>10, 32, 108, 187, 189, 190</sup>. It still 733 734 appears unlikely that regulators are ready to make the change from the traditional to the risk-735 based approach for contaminated site land management. Some researchers have recently 736 argued that while current available evidence may not be sufficient for such departure from 737 the status quo, the bioavailability concept is sufficiently developed for inclusion into existing 738 RA framework, as part of higher tier steps (i.e. Tiers 2 and 3) to modify exposure scenarios and risk estimations <sup>10, 187, 189</sup>. Perhaps, the ideal but unlikely situation would be to consider 739 740 contaminant bioavailability at the initial stages of developing SGVs, and throughout the 741 preliminary and detailed RA phases. While the proposed frameworks may ensure a realistic 742 RA process, uncertainties with the risk-based approach need to be considerably minimised.

5.1. Could the residual fraction be the defining point for risk based land management?

744 The importance of the risk-based approach to contaminated land management cannot be 745 over-emphasised. There has been moves to include bioaccessibility or bioavailability into RA 746 to reduce the overprotective nature of RA caused by the use of total extractable 747 concentrations. By including an extractable fraction (bioaccessible) which is less than the total extractable concentration as part of site-specific RA, and thus reducing remedial 748 749 concentration endpoints, site management becomes more achievable and less expensive. 750 Further, by considering bioaccessibility in RA rather than bioavailability, there is still a degree 751 of overprotection, although it is not just as conservative as using total concentrations. 752 However, should the associated uncertainties especially those related to the fate of residual 753 contaminant fractions be overlooked, or resolved to allow seamless incorporation of the risk-

754 based approach into existing regulatory structures? Exposure to the rapidly desorbing 755 contaminant fractions (i.e. the bioavailable) in soil defines realistic risks, therefore 756 understanding the dynamic processes that control bioavailable fractions may be more critical than the residual fractions, as suggested in the pesticides literature <sup>43</sup>. Whether pesticide 757 758 residual fractions should be included in RA is still being debated, including clear guidance on how they should be considered <sup>40, 126</sup>. Arguably, bringing 'residual HOC fractions' into the 759 760 RA debate, may negatively impact on RA making it more expensive and time consuming to 761 remediate contaminated land, thereby, making pragmatic remediation to an acceptable level 762 almost impossible. Nonetheless, regulatory confidence in adopting the risk-based approach 763 may be significantly increased when convincing evidence shows that contaminant residues 764 in soil are not taken up by a wide range of receptors, and in the event they are taken up, consequent biological effects are non-existent or negligible <sup>48</sup>. In other words, the so called 765 'contaminant residues' must be demonstrated to pose minimal or no risks, from exposure, in 766 767 the short- and long-term. It is suggested that future investigations should focus on clearly 768 answering the key question: how bioavailable are soil-associated HOC residues to key 769 receptors and are there potential risks from exposure? Finally, whether the need for 770 investigations into the likely role played by residual fractions in potentially defining risk based 771 land management, is merely a routine academic endeavour or one which is necessary for 772 pragmatic and realistic RA would be beyond the scope of this review and is thus subject to 773 discussions in the future.

774 6. Conclusion and future research prospects

The overall purpose of the risk-based approach to managing contaminated lands is to minimise risks from exposure. However, adopting the risk-based approach within current regulatory structures over the overly-conservative traditional approach, requires that associated uncertainties are understood and minimised. One such uncertainty is whether there are potential risks associated with exposure to the residual contaminant fractions in soil and how they should be considered in the risk assessment process. There have been

781 arguments for and against the certainty of risks in this regard. However, evidence supporting 782 these arguments is based on empirical observations which may not be completely 783 convincing. The review of the literature suggests that the exclusion of the residual fractions 784 appears inappropriate since very few studies have focused on systematically understanding 785 desorption behaviour of residual hydrophobic organic contaminants in soil, with few studies 786 considering the influence of exposure-specific factors on residual contaminant mobilisation. 787 For example, the exposure of soil-associated hydrophobic organic contaminants to 788 gastrointestinal fluids and enzyme processes may elicit dynamic contaminant release 789 responses, which may affect remobilisation of highly sequestered residues. Although, in vitro 790 models have been used to predict PAH bioaccessibility and chemical extractions used to determine different residual fractions in soils, there are still doubts as to what is actually 791 792 being measured given variations in results generated. Therefore, it is not certain which 793 bioaccessibility model currently represents the best choice, or provides the best fit for in vivo 794 bioavailability determinations. Neither, has the relevance of current definitions of the residual 795 contaminant fractions to ecological and human health risk assessments been convincingly 796 demonstrated. How should the residual and bioaccessible contaminant pools be defined? 797 Would it be prudent to define the residual contaminant fractions in soil based on exposure-798 specific scenarios? How should these highly sequestered contaminant residues in soil be 799 isolated and quantified? Is it possible to predict potential risks from exposure to these 800 residual fractions? And in fact, could there be any scientific, regulatory and/or pragmatic 801 basis for these investigations? These remain questions of concern which may hamper the 802 adoption of the risk based approach. To provide convincing evidence, it is suggested that 803 desorption behaviour of hydrophobic organic contaminants in a wide range of aged 804 historically contaminated soils should be investigated, considering exposure-specific 805 scenarios and also bearing in mind the importance of mechanistic molecular-level data to 806 supporting empirically-derived data. Investigations to develop and validate standard 807 operating procedures for hydrophobic organic contaminants bioavailability determinations 808 are required, and inter-laboratory research efforts may be needed to propose the most appropriate testing method(s). Such investigations may be useful for minimising
uncertainties associated with the risk-based approach, so that due consideration may then
be given to its regulatory adoption on a wider scale.

812 7. Acknowledgments

The authors acknowledge the Cooperative Research Centre for Contamination Assessment and Remediation of the Environment (CRC CARE) and Global Centre for Environmental Remediation (GCER), University of Newcastle (UoN) Australia for providing financial and infrastructural support. AU is grateful to UoN and CRC CARE for awarding him the International Postgraduate Research and CRC CARE scholarships. FAO; ITPS, *Status of the world's soil resources (SWSR)* - Main Report. Natural
 Resources and Environment Department, Food and Agriculture Organization (FAO) of the
 United Nations and Intergovernmental Technical Panel on Soils (ITPS): Rome, Italy, **2015**;
 648 p.

- Rockstrom, J.; Steffen, W.; Noone, K.; Persson, A.; Chapin, F. S.; Lambin, E. F.;
   Lenton, T. M.; Scheffer, M.; Folke, C.; Schellnhuber, H. J.; Nykvist, B.; de Wit, C. A.;
   Hughes, T.; van der Leeuw, S.; Rodhe, H.; Sorlin, S.; Snyder, P. K.; Costanza, R.; Svedin,
   U.; Falkenmark, M.; Karlberg, L.; Corell, R. W.; Fabry, V. J.; Hansen, J.; Walker, B.;
   Liverman, D.; Richardson, K.; Crutzen, P.; Foley, J. A., A safe operating space for humanity.
   *Nature* 2009, *461*, (7263), 472-475.
- 829 3. UN, United Nations Sustainable Development Goals (SDGs). **2016.** 830 <u>http://www.un.org/sustainabledevelopment/sustainable-development-goals/</u>
- 4. IARC, Benzo[a]pyrene, polynuclear aromatic compounds, part 1, chemical, environmental and experimental data. In International Agency for Research on Cancer (IARC) Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans
  1983, 32, 211-224.
- 835 5. USEPA, *Priority Pollutants*. 2008.
- 836 <u>http://water.epa.gov/scitech/methods/cwa/pollutants.cfm</u>
- 6. Jones, K.; de Voogt, P., Persistent organic pollutants (POPs): State of the science. *Environmental Pollution* **1999**, *100*, (1-3), 209-221.
- 839 7. enHealth, Australian Exposure Factor Guidance Handbook. enHealth Council,
  840 Environmental Health Committee: Canberra, 2012, 153 p.
- 841 8. Khan, S.; Cao, Q.; Lin, A. J.; Zhu, Y. G., Concentrations and bioaccessibility of 842 polycyclic aromatic hydrocarbons in wastewater-irrigated soil using in vitro gastrointestinal 843 test. *Environmental Science & Pollution Research* **2008**, *15*, (4), 344-353.
- 844 9. Ramesh, A.; Walker, S. A.; Hood, D. B.; Guillen, M. D.; Schneider, K.; Weyand, E.
  845 H., Bioavailability and risk assessment of orally ingested polycyclic aromatic hydrocarbons.
  846 *International Journal of Toxicology* **2004**, *23*, (5), 301-333.
- Nortega-Calvo, J.-J.; Harmsen, J.; Parsons, J. R.; Semple, K. T.; Aitken, M. D.; Ajao,
  C.; Eadsforth, C.; Galay-Burgos, M.; Naidu, R.; Oliver, R.; Peijnenburg, W. J. G. M.;
  Römbke, J.; Streck, G.; Versonnen, B., From bioavailability science to regulation of organic
  chemicals. *Environmental Science & Technology* **2015**, *49*, (17), 10255-10264.
- 11. NRC, *Bioavailability of contaminants in soils and sediments: Processes, tools, and applications*. National Academies Press: Washington, D.C., **2003**, 420 p.
- 853 12. Semple, K.; Morriss, A.; Paton, G., Bioavailability of hydrophobic organic
  854 contaminants in soils: Fundamental concepts and techniques for analysis. *European Journal*855 of Soil Science 2003, 54, (4), 809-818.
- Naidu, R., *Chemical bioavailability in terrestrial environments*. Elsevier: Amsterdam,
  Boston, **2008**, 809 p.
- Lu, M.; Yuan, D. X.; Lin, Q. M.; Ouyang, T., Assessment of the bioaccessibility of polycyclic aromatic hydrocarbons in topsoils from different urban functional areas using an in vitro gastrointestinal test. *Environmental monitoring & assessment* **2010**, *166*, (1-4), 29-39.
- 15. Semple, K. T.; Doick, K. J.; Jones, K. C.; Burauel, P.; Craven, A.; Harms, H., Peer
  reviewed: Defining bioavailability and bioaccessibility of contaminated soil and sediment is
  complicated. *Environmental Science & Technology* **2004**, *38*, (12), 228A-231A.
- 16. Siciliano, S. D.; Laird, B. D.; Lemieux, C. L., Polycyclic aromatic hydrocarbons are enriched but bioaccessibility reduced in brownfield soils adhered to human hands. *Chemosphere* **2010**, *80*, (9), 1101-1108.
- 17. Versantvoort, C. H. M.; Oomen, A. G.; Van de Kamp, E.; Rompelberg, C. J. M.; Sips,
  A. J. A. M., Applicability of an in vitro digestion model in assessing the bioaccessibility of
  mycotoxins from food. *Food & Chemical Toxicology* **2005**, *43*, (1), 31-40.

- 18. Cave, M. R.; Wragg, J.; Harrison, I.; Vane, C. H.; Van de Wiele, T.; De Groeve, E.;
  Nathanail, C. P.; Ashmore, M.; Thomas, R.; Robinson, J.; Daly, P., Comparison of batch
  mode and dynamic physiologically based bioaccessibility tests for PAHs in soil samples. *Environmental Science & Technology* **2010**, *44*, (7), 2654-2660.
- 19. Duan, L.; Palanisami, T.; Liu, Y. J.; Dong, Z. M.; Mallavarapu, M.; Kuchel, T.; Semple, K. T.; Naidu, R., Effects of ageing and soil properties on the oral bioavailability of benzo[a]pyrene using a swine model. *Environment International* **2014**, *70*, 192-202.
- Nathanail, C. P.; Ogden, R. C., Derivation of a site-specific assessment criterion for
  benzo[a]pyrene in red shale at a former coking works. *Journal of Environmental Science and Health, Part A: Toxic/Hazardous Substances & Environmental Engineering* 2013, 48, (6),
  594-603.
- Semple, K. T.; Riding, M. J.; McAllister, L. E.; Sopena-Vazquez, F.; Bending, G. D.,
  Impact of black carbon on the bioaccessibility of organic contaminants in soil. *Journal of Hazardous Materials* 2013, *261*, 808-816.
- 884 22. Alexander, M., Aging, bioavailability, and overestimation of risk from environmental 885 pollutants. *Environmental Science & Technology* **2000**, *34*, (20), 4259-4265.
- 23. Luo, L.; Lin, S.; Huang, H. L.; Zhang, S. Z., Relationships between aging of PAHs and soil properties. *Environmental Pollution* **2012**, *170*, 177-182.
- 888 24. Ma, L.; Zhang, J.; Han, L.; Li, W.; Xu, L.; Hu, F.; Li, H., The effects of aging time on 889 the fraction distribution and bioavailability of PAH. *Chemosphere* **2012**, *86*, (10), 1072-1078.
- 890 25. Northcott, G. L.; Jones, K. C., Partitioning, extractability, and formation of
  891 nonextractable PAH residues in soil. 1. Compound differences in aging and sequestration.
  892 *Environmental Science & Technology* **2001**, *35*, (6), 1103-1110.
- 26. Cornelissen, G.; vanNoort, P. C. M.; Govers, H. A. J., Desorption kinetics of
  chlorobenzenes, polycyclic aromatic hydrocarbons, and polychlorinated biphenyls: Sediment
  extraction with Tenax(R) and effects of contact time and solute hydrophobicity. *Environmental Toxicology & Chemistry* 1997, *16*, (7), 1351-1357.
- Luthy, R.; Aiken, G.; Brusseau, M.; Cunningham, S.; Gschwend, P.; Pignatello, J.;
  Reinhard, M.; Traina, S.; Weber, W.; Westall, J., Sequestration of hydrophobic organic
  contaminants by geosorbents. *Environmental Science & Technology* **1997**, *31*, (12), 33413347.
- 901 28. Naidu, R.; Pollard, S.; Bolan, N.; Owens, G.; Pruszinski, A., *Bioavailability: The*902 *underlying basis for risk based land management.* In Chemical bioavailability in terrestrial
  903 environment, Naidu, R. (Ed.) Elsevier: Amsterdam, **2008**, 53 72.
- 904 29. Duan, L.; Naidu, R.; Thavamani, P.; Meaklim, J.; Megharaj, M., Managing long-term
  905 polycyclic aromatic hydrocarbon contaminated soils: A risk-based approach. *Environmental*906 Science and Pollution Research 2015, 22, (12), 8927-8941.
- 30. Thavamani, P.; Smith, E.; Kavitha, R.; Mathieson, G.; Megharaj, M.; Srivastava, P.;
  Naidu, R., Risk based land management requires focus beyond the target contaminants A
  case study involving weathered hydrocarbon contaminated soils. *Environmental Technology & Innovation* 2015, *4*, 98-109.
- 911 31. Das, P.; Arias, V. A.; Kambala, V.; Mallavarapu, M.; Naidu, R., Remediation of 912 perfluorooctane sulfonate in contaminated soils by modified clay adsorbent-a risk-based 913 approach. *Water Air & Soil Pollution* **2013**, *224*, 1714.
- 32. Latawiec, A.; Swindell, A.; Simmons, P.; Reid, B., Bringing bioavailability into
  contaminated land decision making: the way forward? *Critical Reviews in Environmental Science and Technology* **2011**, *41*, (1), 52-77.
- 33. Ng, J. C.; Juhasz, A. L.; Smith, E.; Naidu, R. Contaminant bioavailability and
  bioaccessibility. Part 1: A scientific and technical review. Cooperative Research Centre for
  Contamination Assessment and Remediation of the Environment (CRC CARE) Technical
  Report 14: Adelaide, Australia, **2010**, 67 p.
- 921 34. USEPA, Standard Operating Procedure for an In Vitro Bioaccessibility Assay for 922 Lead in Soil, **2012**.
- 923 <u>http://nepis.epa.gov/EPA/html/DLwait.htm?url=/Exe/ZyPDF.cgi/P100GESL.PDF?Dockey=P1</u>
   924 00GESL.PDF

- 35. ECETOC, Significance of bound residues in environmental risk assessment.
  European Centre for Ecotoxicology and Toxicology of Chemicals (ECETOC) Workshop
  Report No. 17: Brussels, Belgium, **2010**, 76 p.
- 36. ECETOC, Development of interim guidance for the inclusion of non-extractable
  residues in the risk assessment of chemicals. Workshop Report No. 118: Brussels, Belgium,
  2013b; 110 p.
- 37. Rhodes, A.; Carlin, A.; Semple, K., Impact of black carbon in the extraction and
  mineralization of phenanthrene in soil. *Environmental Science & Technology* 2008, 42, (3),
  740-745.
- 38. Kastner, M.; Nowak, K. M.; Miltner, A.; Trapp, S.; Schaffer, A., Classification and
  modelling of nonextractable residue (NER) formation of xenobiotics in soil A synthesis. *Critical Reviews in Environmental Science & Technology* **2014**, *44*, (19), 2107-2171.
- 937 39. Barraclough, D.; Kearney, T.; Croxford, A., Bound residues: Environmental solution 938 or future problem? *Environmental Pollution* **2005**, *133*, (1), 85-90.
- 939 40. Barriuso, E.; Benoit, P.; Dubus, I. G., Formation of pesticide nonextractable (bound)
  940 residues in soil: Magnitude, controlling factors and reversibility. *Environmental Science* &
  941 *Technology* 2008, *42*, (6), 1845-1854.
- 942 41. Craven, A., Bound residues of organic compounds in the soil: the significance of
  943 pesticide persistence in soil and water: a European regulatory view. *Environmental Pollution*944 **2000**, *108*, (1), 15-18.
- 945 42. Craven, A.; Hoy, S., Pesticide persistence and bound residues in soil regulatory
  946 significance. *Environmental Pollution* **2005**, *133*, (1), 5-9.
- 947 43. Gevao, B.; Jones, K.; Semple, K.; Craven, A.; Burauel, P., Nonextractable pesticide
  948 residues in soil. *Environmental Science & Technology* 2003, 37, (7), 138a-144a.
- 949 44. Gevao, B.; Semple, K.; Jones, K., Bound pesticide residues in soils: a review.
  950 *Environmental Pollution* 2000, *108*, (1), 3-14.
- 45. Richnow, H. H.; Eschenbach, A.; Mahro, B.; Kastner, M.; Annweiler, E.; Seifert, R.;
  Michaelis, W., Formation of nonextractable soil residues: A stable isotope approach. *Environmental Science & Technology* **1999**, *33*, (21), 3761-3767.
- Birdwell, J. E.; Thibodeaux, L. J., PAH repartitioning in field-contaminated sediment
  following removal of the labile chemical fraction. *Environmental Science & Technology* 2009,
  43, (21), 8092-8097.
- 47. Juhasz, A. L.; Smith, E.; Waller, N.; Stewart, R.; Weber, J., Bioavailability of residual polycyclic aromatic hydrocarbons following enhanced natural attenuation of creosotecontaminated soil. *Environmental Pollution* **2010**, *158*, (2), 585-591.
- 48. Calderbank, A., The occurrence and significance of bound pesticide-residues in soil.
   *Reviews of Environmental Contamination and Toxicology* **1989**, *108*, 71-103.
- 962 49. Rhodes, A. H.; McAllister, L. E.; Semple, K. T., Linking desorption kinetics to 963 phenanthrene biodegradation in soil. *Environmental Pollution* **2010**, *158*, (5), 1348-1353.
- 50. Doick, K. J.; Klingelmann, E.; Burauel, P.; Jones, K. C.; Semple, K. T., Long-term
  fate of polychlorinated biphenyls and polycyclic aromatic hydrocarbons in an agricultural soil. *Environmental Science & Technology* **2005**, *39*, (10), 3663-3670.
- Mayer, P.; Hilber, I.; Gouliarmou, V.; Hale, S. E.; Cornelissen, G.; Bucheli, T. D., How
  to determine the environmental exposure of PAHs originating from biochar. *Environmental Science & Technology* **2016**, *50*, (4), 1941-1948.
- 52. Rhodes, A. H.; Dew, N. M.; Semple, K. T., Relationship between cyclodextrin extraction and biodegradation of phenanthrene in soil. *Environmental Toxicology and Chemistry* **2008**, *27*, (7), 1488-1495.
- 53. Bernhardt, C.; Derz, K.; Kordel, W.; Terytze, K., Applicability of non-exhaustive
  extraction procedures with Tenax and HPCD. *Journal of Hazardous Materials* 2013, 261,
  711-717.
- 976 54. Northcott, G. L.; Jones, K. C., Partitioning, extractability, and formation of
  977 nonextractable PAH residues in soil. 2. Effects on compound dissolution behavior.
  978 Environmental Science & Technology 2001, 35, (6), 1111-1117.

55. Eschenbach, A.; Wienberg, R.; Mahro, B., Fate and stability of nonextractable
residues of [(14)C]PAH in contaminated soils under environmental stress conditions. *Environmental Science & Technology* **1998**, *32*, (17), 2585-2590.

- 56. Simpson, M. J., Nuclear magnetic resonance based investigations of contaminant interactions with soil organic matter. *Soil Science Society of America Journal* **2006**, *70*, (3), 984 995-1004.
- 57. Masoom, H.; Courtier-Murias, D.; Farooq, H.; Soong, R.; Kelleher, B. P.; Zhang, C.;
  Maas, W. E.; Fey, M.; Kumar, R.; Monette, M.; Stronks, H. J.; Simpson, M. J.; Simpson, A.
  J., Soil organic matter in its native state. Unravelling the most complex biomaterial on earth. *Environmental Science & Technology* **2016**, *50*, (4), 1670-1680
- 58. Simpson, A. J.; McNally, D. J.; Simpson, M. J., NMR spectroscopy in environmental
  research: from molecular interactions to global processes. *Progress in Nuclear Magnetic Resonance Spectroscopy* 2011, 58, (3), 97-175.
- 59. Kim, H. S.; Lindsay, K. S.; Pfaender, F. K., Enhanced mobilization of field
  contaminated soil-bound PAHs to the aqueous phase under anaerobic conditions. *Water Air & Soil Pollution* 2008, *189*, (1-4), 135-147.
- 995 60. Enell, A.; Reichenberg, F.; Ewald, G.; Warfvinge, P., Desorption kinetics studies on 996 PAH-contaminated soil under varying temperatures. *Chemosphere* **2005**, *61*, (10), 1529-997 1538.
- 61. Tao, S.; Li, L.; Ding, J. N.; Zhong, J. J.; Zhang, D. Y.; Lu, Y.; Yang, Y. F.; Wang, X.
  L.; Li, X. Q.; Cao, J.; Lu, X. X.; Liu, W. X., Mobilization of soil-bound residue of organochlorine pesticides and polycyclic aromatic hydrocarbons in an in vitro gastrointestinal model. *Environmental Science & Technology* **2011**, *45*, (3), 1127-1132.
- 1002 62. Juhasz, A. L.; Weber, J.; Stevenson, G.; Slee, D.; Gancarz, D.; Rofe, A.; Smith, E., *In* 1003 *vivo* measurement, *in vitro* estimation and fugacity prediction of PAH bioavailability in post-1004 remediated creosote-contaminated soil. *Science of the Total Environment* **2014**, *473*, 147-1005 154.
- 1006 63. Gomez-Eyles, J. L.; Collins, C. D.; Hodson, M. E., Relative proportions of polycyclic 1007 aromatic hydrocarbons differ between accumulation bioassays and chemical methods to 1008 predict bioavailability. *Environmental Pollution* **2010**, *158*, (1), 278-284.
- 1009 64. Gomez-Eyles, J. L.; Jonker, M. T. O.; Hodson, M. E.; Collins, C. D., Passive
  1010 samplers provide a better prediction of pah bioaccumulation in earthworms and plant roots
  1011 than exhaustive, mild solvent, and cyclodextrin extractions. *Environmental Science* &
  1012 *Technology* 2012, *46*, (2), 962-969.
- 1013 65. Mayer, P.; Olsen, J. L.; Gouliarmou, V.; Hasinger, M.; Kendler, R.; Loibner, A. P., A 1014 contaminant trap as a tool for isolating and measuring the desorption resistant fraction of soil 1015 pollutants. *Environmental Science & Technology* **2011**, *45*, (7), 2932-2937.
- 1016 66. Beck, P.; Mann, B. *A technical guide for demonstrating monitored natural attenuation* 1017 of petroleum hydrocarbons in groundwater, CRC CARE Technical Report: 2010.
- 1018 67. Bekins, B.; Rittmann, B. E.; MacDonald, J. A., Natural attenuation strategy for 1019 groundwater cleanup focuses on demonstrating cause and effect. *Eos, Transactions* 1020 *American Geophysical Union* **2001**, *8*2, (5), 53-58.
- 1021 68. Declercq, I.; Cappuyns, V.; Duclos, Y., Monitored natural attenuation (MNA) of 1022 contaminated soils: State of the art in Europe-A critical evaluation. *Science of the Total* 1023 *Environment* **2012**, *426*, 393-405.
- 1024 69. Mulligan, C. N.; Yong, R. N., Natural attenuation of contaminated soils. *Environment* 1025 *International* **2004**, *30*, (4), 587-601.
- 1026
  1027
  1027
  1027
  1028
  1028
  2005, 39, (22), 458a-464a.
- 1029 71. Rugner, H.; Finkel, M.; Kaschl, A.; Bittens, M., Application of monitored natural 1030 attenuation in contaminated land management - A review and recommended approach for 1031 Europe. *Environmental Science & Policy* **2006**, *9*, (6), 568-576.
  - 36

- 1032 72. Schirmer, M.; Dahmke, A.; Dietrich, P.; Dietze, M.; Godeke, S.; Richnow, H. H.; 1033 Schirmer, K.; Weiss, H.; Teutsch, G., Natural attenuation research at the contaminated 1034 megasite Zeitz. *Journal of Hydrology* **2006**, *328*, (3-4), 393-407.
- 103573.USEPAA citizen's guide to monitored natural attenuation.https://clu-1036in.org/download/Citizens/a citizens guide to monitored natural attenuation.pdf
- 1037 74. Sims, R.; Overcash, M., Fate of polynuclear aromatic compounds (PNAS) on soil-1038 plant systems. *Residue Reviews* **1983**, *88*, 1-68.
- 1039 75. Reid, B.; Jones, K.; Semple, K., Bioavailability of persistent organic pollutants in soils
  1040 and sediments A perspective on mechanisms, consequences and assessment.
  1041 *Environmental Pollution* 2000, *108*, (1), 103-112.
- 1042 76. Rombola, A. G.; Meredith, W.; Snape, C. E.; Baronti, S.; Genesio, L.; Vaccari, F. P.;
  1043 Miglietta, F.; Fabbri, D., Fate of soil organic carbon and polycyclic aromatic hydrocarbons in
  1044 a vineyard soil treated with biochar. *Environmental Science & Technology* 2015, 49, (18),
  1045 11037-11044.
- 1046 77. Riding, M. J.; Doick, K. J.; Martin, F. L.; Jones, K. C.; Semple, K. T., Chemical
  1047 measures of bioavailability/bioaccessibility of PAHs in soil: Fundamentals to application.
  1048 *Journal of Hazardous Materials* 2013, *261*, (0), 687-700.
- 1049 78. Semple, K.; Stokes, J.; Reid, B.; Leeson, A.; Alleman, B., Temporal chances in the 1050 bioavailability of PAHs in soils. *Bioremediation Technologies For Polycyclic Aromatic* 1051 *Hydrocarbon Compounds* **1999**, 247-252.
- 1052 79. Duan, L.; Naidu, R.; Liu, Y. J.; Palanisami, T.; Dong, Z. M.; Mallavarapu, M.; Semple,
  1053 K. T., Effect of ageing on benzo[a]pyrene extractability in contrasting soils. *Journal of*1054 *Hazardous Materials* 2015, 296, 175-184.
- 1055 80. Macleod, C.; Semple, K., Influence of contact time on extractability and degradation 1056 of pyrene in soils. *Environmental Science & Technology* **2000**, *34*, (23), 4952-4957.
- 1057 81. Macleod, C. J. A.; Semple, K. T., Sequential extraction of low concentrations of
  1058 pyrene and formation of non-extractable residues in sterile and non-sterile soils. *Soil Biology*1059 & *Biochemistry* 2003, 35, (11), 1443-1450.
- 1060 82. Pignatello, J. J.; Xing, B. S., Mechanisms of slow sorption of organic chemicals to 1061 natural particles. *Environmental Science & Technology* **1996**, *30*, (1), 1-11.
- 1062 83. Cornelissen, G.; Van Noort, P.; Govers, H., Mechanism of slow desorption of organic
  1063 compounds from sediments: A study using model sorbents. *Environmental Science* &
  1064 *Technology* 1998, *32*, (20), 3124-3131.
- 1065 84. Weber, W. J.; Huang, W. L.; Yu, H., Hysteresis in the sorption and desorption of
  1066 hydrophobic organic contaminants by soils and sediments 2. Effects of soil organic matter
  1067 heterogeneity. *Journal of Contaminant hydrology* **1998**, *31*, (1), 149-165.
- 1068 85. Huang, W. L.; Ping, P. A.; Yu, Z. Q.; Fu, H. M., Effects of organic matter
  1069 heterogeneity on sorption and desorption of organic contaminants by soils and sediments.
  1070 Applied Geochemistry 2003, 18, (7), 955-972.
- 1071 86. Chung, N.; Alexander, M., Differences in sequestration and bioavailability of organic 1072 compounds aged in dissimilar soils. *Environmental Science & Technology* **1998**, *32*, (7), 1073 855-860.
- 1074 87. Chung, N.; Alexander, M., Effect of soil properties on bioavailability and extractability 1075 of phenanthrene and atrazine sequestered in soil. *Chemosphere* **2002**, *48*, (1), 109-115.
- 1076 88. Duan, L.; Naidu, R., Effect of ionic strength and index cation on the sorption of 1077 phenanthrene. *Water Air & Soil Pollution* **2013**, *224*, 1700.
- 1078 89. Hickman, Z. A.; Swindell, A. L.; Allan, I. J.; Rhodes, A. H.; Hare, R.; Semple, K. T.;
  1079 Reid, B. J., Assessing biodegradation potential of PAHs in complex multi-contaminant
  1080 matrices. *Environmental Pollution* **2008**, *156*, (3), 1041-1045.
- 1081 90. Macleod, C.; Semple, K., The adaptation of two similar soils to pyrene catabolism. 1082 *Environmental Pollution* **2002**, *119*, (3), 357-364.
- 1083 91. Carmichael, L. M.; Christman, R. F.; Pfaender, F. K., Desorption and mineralization 1084 kinetics of phenanthrene and chrysene in contaminated soils. *Environmental Science* & 1085 *Technology* **1997**, *31*, (1), 126-132.

- Yang, W. C.; Duan, L.; Zhang, N.; Zhang, C. D.; Shipley, H. J.; Kan, A. T.; Tomson,
  M. B.; Chen, W., Resistant desorption of hydrophobic organic contaminants in typical
  Chinese soils: Implications for long-term fate and soil quality standards. *Environmental Toxicology and Chemistry* 2008, 27, (1), 235-242.
- 1090 93. Abu, A.; Smith, S., Mechanistic characterization of adsorption and slow desorption of 1091 phenanthrene aged in soils. *Environmental Science & Technology* **2006**, *40*, (17), 5409-1092 5414.
- 1093 94. Cornelissen, G.; Gustafsson, O., Sorption of phenanthrene to environmental black 1094 carbon in sediment with and without organic matter and native sorbates. *Environmental* 1095 *Science & Technology* **2004**, *38*, (1), 148-155.
- 1096 95. Ran, Y.; Sun, K.; Yang, Y.; Xing, B. S.; Zeng, E., Strong sorption of phenanthrene by 1097 condensed organic matter in soils and sediments. *Environmental Science & Technology* 1098 **2007**, *41*, (11), 3952-3958.
- 1099 96. Smith, K. E. C.; Thullner, M.; Wick, L. Y.; Harms, H., Sorption to humic acids 1100 enhances polycyclic aromatic hydrocarbon biodegradation. *Environmental Science* & 1101 *Technology* **2009**, *43*, (19), 7205-7211.
- 1102 97. Hawthorne, S. B.; Poppendieck, D. G.; Grabanski, C. B.; Loehr, R. C., PAH release
  1103 during water desorption, supercritical carbon dioxide extraction, and field bioremediation.
  1104 *Environmental Science & Technology* 2001, *35*, (22), 4577-4583.
- 1105 98. Cornelissen, G.; Gustafsson, O.; Bucheli, T.; Jonker, M.; Koelmans, A.; Van Noort, 1106 P., Extensive sorption of organic compounds to black carbon, coal, and kerogen in 1107 sediments and soils: Mechanisms and consequences for distribution, bioaccumulation, and 1108 biodegradation. *Environmental Science & Technology* **2005**, *39*, (18), 6881-6895.
- 99. Hwang, S. C.; Cutright, T. J., Evidence of underestimation in PAH
  sorption/desorption due to system nonequilibrium and interaction with soil constituents. *Journal of Environmental Science and Health Part a-Toxic/Hazardous Substances* & *Environmental Engineering* 2004, 39, (5), 1147-1162.
- 1113 100. Zhao, D. Y.; Pignatello, J. J.; White, J. C.; Braida, W.; Ferrandino, F., Dual-mode 1114 modeling of competitive and concentration-dependent sorption and desorption kinetics of 1115 polycyclic aromatic hydrocarbons in soils. *Water Resources Research* **2001**, *37*, (8), 2205-1116 2212.
- 1117 101. Braida, W. J.; Pignatello, J. J.; Lu, Y. F.; Ravikovitch, P. I.; Neimark, A. V.; Xing, B.
  1118 S., Sorption hysteresis of benzene in charcoal particles. *Environmental Science* &
  1119 *Technology* 2003, *37*, (2), 409-417.
- 1120 102. Jonker, M. T. O.; Koelmans, A. A., Extraction of polycyclic aromatic hydrocarbons 1121 from soot and sediment: Solvent evaluation and implications for sorption mechanism. 1122 *Environmental Science & Technology* **2002**, *36*, (19), 4107-4113.
- 1123 103. Lu, Y. F.; Pignatello, J. J., Demonstration of the "Conditioning effect" in soil organic 1124 matter in support of a pore deformation mechanism for sorption hysteresis. *Environmental* 1125 *Science & Technology* **2002**, *36*, (21), 4553-4561.
- 1126 104. Sander, M.; Pignatello, J. J., On the reversibility of sorption to black carbon: 1127 Distinguishing true hysteresis from artificial hysteresis caused by dilution of a competing 1128 adsorbate. *Environmental Science & Technology* **2007**, *41*, (3), 843-849.
- 1129 105. Chen, S.; Xu, Y. P.; Wang, Z. J., Assessing desorption resistance of PAHs in 1130 dissolved humic substances by membrane-based passive samplers. *Journal of colloid &* 1131 *interface science* **2010**, *350*, (1), 348-354.
- 1132 106. Wang, Z. J.; Chen, S.; Xu, Y. P.; Tang, J. F., Aging effects on sorption-desorption 1133 behaviors of PAHs in different natural organic matters. *Journal of colloid & interface science* 1134 **2012**, 382, (1), 117-122.
- 1135 107. ECETOC, Understanding the Relationship between Extraction Technique and 1136 Bioavailability. Workshop Report No. 117: Brussels, Belgium, **2013a**, 90 p.
- 1137 108. Kordel, W.; Bernhardt, C.; Derz, K.; Hund-Rinke, K.; Harmsen, J.; Peijnenburg, W.; 1138 Comans, R.; Terytze, K., Incorporating availability/bioavailability in risk assessment and 1139 decision making of polluted sites, using Germany as an example. *Journal of Hazardous* 1140 Materials **2012**, 261, 854, 862

- 1141 109. Wanner, U.; Burauel, P.; Fuhr, F., Characterisation of soil-bound residue fractions of 1142 the fungicide dithianon by gel permeation chromatography and polyacrylamide gel 1143 electrophoresis. Environmental Pollution 2000, 108, (1), 53-59.
- Karickhoff, S. W.; Brown, D. S.; Scott, T. A., Sorption of Hydrophobic Pollutants on 1144 110. 1145 Natural Sediments. Water Research 1979, 13, (3), 241-248.
- Pan, B.; Tao, S.; Wu, D.; Zhang, D.; Peng, H. B.; Xing, B. S., Phenanthrene 1146 111. 1147 sorption/desorption sequences provide new insight to explain high sorption coefficients in 1148 field studies. Chemosphere 2011, 84, (11), 1578-1583.
- Ramos, E.; Meijer, S.; Vaes, W.; Verhaar, H.; Hermens, J., Using solid-phase 1149 112. 1150 microextraction to determine partition coefficients to humic acids and bioavailable 1151 concentrations of hydrophobic chemicals. Environmental Science & Technology 1998, 32, 1152 (21), 3430-3435.
- 1153 Seth, R.; Mackay, D.; Muncke, J., Estimating the organic carbon partition coefficient 113. 1154 and its variability for hydrophobic chemicals. Environmental Science & Technology 1999, 33, 1155 (14), 2390-2394.
- 1156 Hwang, S.; Cutright, T., Effect of expandable clays and cometabolism on PAH 114. 1157 biodegradability. Environmental Science and Pollution Research 2003, 10, (5), 277-280.
- 1158 Luo, L.; Zhang, S. Z.; Ma, Y. B., Evaluation of impacts of soil fractions on 115. phenanthrene sorption. Chemosphere 2008, 72, (6), 891-896. 1159
- 1160 Rabbi, S. M. F.; Linser, R.; Hook, J. M.; Wilson, B. R.; Lockwood, P. V.; Daniel, H.; 116. 1161 Young, I. M., Characterization of soil organic matter in aggregates and size-density fractions 1162 by solid state C-13 CPMAS NMR spectroscopy. Communications in soil science & plant
- 1163 analysis 2014, 45, (11), 1523-1537.
- 1164 Oades, J. M.; Vassallo, A. M.; Waters, A. G.; Wilson, M. A., Characterization of 117. 1165 organic-matter in particle-size and density fractions from a red-brown earth by solid-state C-13 NMR. Soil Research 1987, 25, (1), 71-82. 1166
- 1167 Chen, J. S.; Chiu, C. Y., Characterization of soil organic matter in different particle-118. 1168 size fractions in humid subalpine soils by CP/MAS C-13 NMR. Geoderma 2003, 117, (1-2), 1169 129-141.
- 1170 Clemente, J. S.; Gregorich, E. G.; Simpson, A. J.; Kumar, R.; Courtier-Murias, D.; 119. 1171 Simpson, M. J., Comparison of nuclear magnetic resonance methods for the analysis of 1172 organic matter composition from soil density and particle fractions. Environmental Chemistry 1173 **2012**, *9*, (1), 97-107.
- 1174 Salati, S.; Adam, F.; Cosentino, C.; Torri, G., Studying soil organic matter using C-13 120. 1175 CP-MAS NMR: The effect of soil chemical pre-treatments on spectra quality and representativity. Chemosphere 2008, 70, (11), 2092-2098. 1176
- 1177 Lehmann, J.; Kleber, M., The contentious nature of soil organic matter. Nature 2015, 121. 1178 528, (7580), 60-68.
- 1179 Zarfl, C.; Klasmeier, J.; M, M., Non-extractable residues are not necessarily bound 122. 1180 residues. In poster presented at SETAC Europe 19th Annual Meeting: Goteborg, Sweden, 1181 2009.
- 1182 Gao, Y. Z.; Zeng, Y. C.; Shen, Q.; Ling, W. T.; Han, J., Fractionation of polycyclic 123. 1183 aromatic hydrocarbon residues in soils. Journal of Hazardous Materials 2009, 172, (2-3), 1184 897-903.
- 1185 Northcott, G. L.; Jones, K. C., Experimental approaches and analytical techniques for 124. 1186 determining organic compound bound residues in soil and sediment. Environmental Pollution 1187 **2000**, *108*, (1), 19-43.
- 1188 Hack, A.; Selenka, F., Mobilization of PAH and PCB from contaminated soil using a 125. 1189 digestive tract model. Toxicol Lett 1996, 88, (1-3), 199-210.
- 1190 USEPA Guidance for addressing unextracted pesticide residues in laboratory 126. https://www.epa.gov/pesticide-science-and-assessing-pesticide-risks/guidance-1191 studies. 1192 addressing-unextracted-pesticide-residues#foot11
- 1193 Tao, S.; Lu, Y.; Zhang, D. Y.; Yang, Y. F.; Yang, Y.; Lu, X. X.; Sai, D. J., Assessment 127. 1194 of oral bioaccessibility of organochlorine pesticides in soil using an in vitro gastrointestinal 1195

- 1196 128. Johnson, M. D.; Weber, W. J., Rapid prediction of long-term rates of contaminant 1197 desorption from soils and sediments. *Environmental Science & Technology* **2001**, *35*, (2), 1198 427-433.
- 1199 129. Cuypers, C.; Grotenhuis, T.; Joziasse, J.; Rulkens, W., Rapid persulfate oxidation
  1200 predicts PAH bioavailability in soils and sediments. *Environmental Science & Technology*1201 **2000**, *34*, (10), 2057-2063.
- 1202 130. Gron, C.; Oomen, A.; Weyand, E.; Wittsiepe, J., Bioaccessibility of PAH from Danish
  1203 soils. Journal of Environmental Science and Health Part a-Toxic/Hazardous Substances &
  1204 Environmental Engineering 2007, 42, (9), 1233-1239.
- 1205 131. Barnier, C.; Ouvrard, S.; Robin, C.; Morel, J. L., Desorption kinetics of PAHs from
  1206 aged industrial soils for availability assessment. *Science of the Total Environment* 2014, 470,
  1207 639-645.
- 1208 132. Errett, D. H.; Chin, Y. P.; Xu, Y. P.; Yan, Y. J., The sorption and desorption kinetics of 1209 polycyclic aromatic hydrocarbons in methanol/water mixtures. *Hazardous waste* & 1210 *hazardous materials* **1996**, *13*, (2), 177-195.
- 1211 133. Semple, K.; Doick, K.; Wick, L.; Harms, H., Microbial interactions with organic 1212 contaminants in soil: Definitions, processes and measurement. *Environmental Pollution* 1213 **2007**, *150*, (1), 166-176.
- 1214 134. Johnson, M. D.; Keinath, T. M.; Weber, W. J., A Distributed reactivity model for 1215 sorption by soils and sediments. 14. Characterization and modeling of phenanthrene 1216 desorption rates. *Environmental Science & Technology* **2001**, *35*, (8), 1688-1695.
- 1217 135. Cornelissen, G.; VanNoort, P. C. M.; Parsons, J. R.; Govers, H. A. J., Temperature 1218 dependence of slow adsorption and desorption kinetics of organic compounds in sediments. 1219 *Environmental Science & Technology* **1997**, *31*, (2), 454-460.
- 1220 136. Rhodes, A.; Hofman, J.; Semple, K., Development of phenanthrene catabolism in natural and artificial soils. *Environmental Pollution* **2008**, *152*, (2), 424-430.
- 1222 137. Rhodes, A.; Riding, M.; McAllister, L.; Lee, K.; Semple, K., Influence of activated 1223 charcoal on desorption kinetics and biodegradation of phenanthrene in soil. *Environmental* 1224 *Science & Technology* **2012**, *46*, (22), 12445-12451.
- 1225 138. Haritash, A.; Kaushik, C., Biodegradation aspects of polycyclic aromatic 1226 hydrocarbons (PAHs): A review. *Journal of Hazardous Materials* **2009**, *169*, (1-3), 1-15.
- 1227 139. Semple, K.; Dew, N.; Doick, K.; Rhodes, A., Can microbial mineralization be used to
  1228 estimate microbial availability of organic contaminants in soil? *Environmental Pollution* 2006,
  1229 140, (1), 164-172.
- 1230 140. Juhasz, A. L.; Waller, N.; Lease, C.; Bentham, R.; Stewart, R., Pilot scale 1231 bioremediation of creosote-contaminated soil—efficacy of enhanced natural attenuation and 1232 bioaugmentation strategies. *Bioremediation journal* **2005**, *9*, (3-4), 139-154.
- 1233 141. Juhasz, A. L.; Waller, N.; Stewart, R., Predicting the efficacy of polycyclic aromatic
  1234 hydrocarbon bioremediation in creosote-contaminated soil using bioavailability assays.
  1235 Bioremediation journal 2005, 9, (2), 99-114.
- 1236 142. Atlas, R. M.; Bartha, R., *Microbial ecology: Fundamentals and applications*. 4th ed. 1237 Benjamin/Cummings: Menlo Park, California, **1998**.
- 1238 143. Juhasz, A.; Naidu, R., Bioremediation of high molecular weight polycyclic aromatic 1239 hydrocarbons: a review of the microbial degradation of benzo[a]pyrene. *International* 1240 *Biodeterioration & Biodegradation* **2000**, *45*, (1-2), 57-88.
- 1241 144. Kanaly, R.; Harayama, S., Biodegradation of high-molecular-weight polycyclic 1242 aromatic hydrocarbons by bacteria. *Journal of Bacteriology* **2000**, *182*, (8), 2059-2067.
- 1243 145. Alexander, M., *Biodegradation and bioremediation*. 2nd ed. Academic Press: San 1244 Diego, **1999**.
- 1245 146. Biswas, B.; Sarkar, B.; Rusmin, R.; Naidu, R., Bioremediation of PAHs and VOCs: 1246 advances in clay mineral-microbial interaction. *Environment International* **2015**, *85*, 168-81.
- 1247 147. Ekschmitt, K.; Liu, M. Q.; Vetter, S.; Fox, O.; Wolters, V., Strategies used by soil 1248 biota to overcome soil organic matter stability - why is dead organic matter left over in the 1249 soil? *Geoderma* **2005**, *128*, (1-2), 167-176.

- 1250 148. ECETOC, Scientific Principles for Soil Hazard Assessment of Substances. Technical
  1251 Report No. 084: Brussels, Belgium, **2002**, 47 p.
- 1252 149. SERDP and ESTCP, Expert panel workshop on research and development needs for
  1253 understanding and assessing the bioavailability of contaminants in soils and sediments.
  1254 Strategic Environmental Research and Development Program (SERDP) and Environmental
  1255 Security Technology Certification Program (ESTCP), **2008**.
- 1256 http://www.serdp.org/Featured-Initiatives/Cleanup-Initiatives/Bioavailability
- 1257 150. Kramer, B.; Ryan, P., Changes in the efficiency of Soxhlet and microwave extraction 1258 of pesticides from soil and model solids over time. *National Meeting-American Chemical* 1259 Society Division Of Environmental Chemistry **2000**, *40*, (2), 13-14.
- 1260 151. Ruby, M.; Davis, A.; Schoof, R.; Eberle, S.; Sellstone, C., Estimation of lead and 1261 arsenic bioavailability using a physiologically based extraction test. *Environmental Science* & 1262 *Technology* **1996**, *30*, (2), 422-430.
- 1263 152. Peijnenburg, W.; Jager, T., Monitoring approaches to assess bioaccessibility and 1264 bioavailability of metals: Matrix issues. *Ecotoxicology and Environmental Safety* **2003**, *56*, 1265 (1), 63-77.
- 1266 153. Reichenberg, F.; Mayer, P., Two complementary sides of bioavailability: Accessibility 1267 and chemical activity of organic contaminants in sediments and soils. *Environmental* 1268 *Toxicology and Chemistry* **2006**, *25*, (5), 1239-1245.
- 1269 154. ECHA, The use of alternatives to testing on animals for the REACH regulation: 1270 Second report under article 117(3) of the REACH regulation. ECHA-14-A-07-EN: Helsinki, 1271 Finland, **2014**.
- 1272 155. NRC, *Toxicity Testing in the 21st century: A Vision and a Strategy*. Committee on 1273 Toxicity Testing and Assessment of Environmental Agents. National Research Council, NRC 1274 (Ed.). National Academies Press: USA, **2007**, 216 p.
- 1275 156. Ehlers, G.; Loibner, A., Linking organic pollutant (bio)availability with geosorbent 1276 properties and biomimetic methodology: A review of geosorbent characterisation. and 1277 (bio)availability prediction. *Environmental Pollution* **2006**, *141*, (3), 494-512.
- 1278 157. Cui, X.; Mayer, P.; Gan, J., Methods to assess bioavailability of hydrophobic organic 1279 contaminants: Principles, operations, and limitations. *Environmental Pollution* **2013**, *172*, 1280 223-234.
- 1281 158. Hodson, M. E.; Vijver Martina G.; Peijnenburg, W. J. G. M., *Bioavalibility in soils*. In
  1282 Dealing with Contaminated Sites, Swartjes, F. A. (Ed.). Springer: Netherlands, **2011**, 7211283 746.
- 1284 159. Kelsey, J.; Kottler, B.; Alexander, M., Selective chemical extractants to predict 1285 bioavailability of soil-aged organic chemicals. *Environmental Science & Technology* **1997**, 1286 *31*, (1), 214-217.
- 1287 160. MacLeod, C.; Lee, P.; Reid, B.; Dzirasa, K.; Semple, K.; Patel, D.; Swingler, S.; 1288 Leeson, A.; Alleman, B., Bioavailability of cable insulating oil to soil biota. *Bioremediation* 1289 *Technologies For Polycyclic Aromatic Hydrocarbon Compounds* **1999**, 259-264.
- 1290 161. Hatzinger, P.; Alexander, M., Effect of ageing of chemicals in soil on their 1291 biodegradability and extractability. *Environmental Science & Technology* **1995**, *29*, (2), 537-1292 545.
- 1293 162. Hwang, S. C.; Cutright, T. J., Preliminary exploration of the relationships between soil
  1294 characteristics and PAH desorption and biodegradation. *Environment International* 2004, *29*,
  1295 (7), 887-894.
- 1296 163. Tang, J.; Liste, H.; Alexander, M., Chemical assays of availability to earthworms of 1297 polycyclic aromatic hydrocarbons in soil. *Chemosphere* **2002**, *48*, (1), 35-42.
- 1298 164. Reid, B.; Stokes, J.; Jones, K.; Semple, K., Nonexhaustive cyclodextrin-based 1299 extraction technique for the evaluation of PAH bioavailability. *Environmental Science* & 1300 *Technology* **2000**, *34*, (15), 3174-3179.
- 1301 165. Liste, H.; Alexander, M., Butanol extraction to predict bioavailability of PAHs in soil. 1302 *Chemosphere* **2002**, *46*, (7), 1011-1017.

- 1303 166. Doick, K. J.; Dew, N. M.; Semple, K. T., Linking catabolism to cyclodextrin 1304 extractability: Determination of the microbial availability of PAHs in soil. *Environmental* 1305 *Science & Technology* **2005**, *39*, (22), 8858-8864.
- 1306 167. Stokes, J.; Wilkinson, A.; Reid, B.; Jones, K.; Semple, K., Prediction of polycyclic 1307 aromatic hydrocarbon biodegradation in contaminated soils using an aqueous 1308 hydroxypropyl-beta-cyclodextrin extraction technique. *Environmental Toxicology and* 1309 *Chemistry* **2005**, *24*, (6), 1325-1330.
- 1310 168. Jonker, M. T. O.; van der Heijden, S. A.; Kreitinger, J. P.; Hawthorne, S. B., 1311 Predicting PAH bioaccumulation and toxicity in earthworms exposed to manufactured gas 1312 plant soils with solid-phase microextraction. *Environmental Science & Technology* **2007**, *41*, 1313 (21), 7472-7478.
- 1314 169. Doick, K. J.; Clasper, P. J.; Urmann, K.; Semple, K. T., Further validation of the 1315 HPCD-technique for the evaluation of PAH microbial availability in soil. *Environmental* 1316 *Pollution* **2006**, *144*, (1), 345-354.
- 1317 170. Cuypers, C.; Pancras, T.; Grotenhuis, T.; Rulkens, W., The estimation of PAH 1318 bioavailability in contaminated sediments using hydroxypropyl-beta-cyclodextrin and Triton 1319 X-100 extraction techniques. *Chemosphere* **2002**, *46*, (8), 1235-1245.
- 1320 171. Ma, L. L.; Zhang, J.; Han, L. S.; Li, W. M.; Xu, L.; Hu, F.; Li, H. X., The effects of 1321 aging time on the fraction distribution and bioavailability of PAH. *Chemosphere* **2012**, *86*, 1322 (10), 1072-1078.
- 1323 172. White, J.; Kelsey, J.; Hatzinger, P.; Alexander, M., Factors affecting sequestration 1324 and bioavailability of phenanthrene in soils. *Environmental Toxicology and Chemistry* **1997**, 1325 16, (10), 2040-2045.
- 1326 173. Hatzinger, P.; Alexander, M., Biodegradation of organic compounds sequestered in 1327 organic solids or in nanopores within silica particles. *Environmental Toxicology and* 1328 *Chemistry* **1997**, *16*, (11), 2215-2221.
- 1329 174. ISO 18. Soil Quality—Requirements and guidance for the selection and application of
   methods for the assessment of bioavailability of contaminants in soil and soil materials. ISO
   1331 No. 17402; International Organization for Standardization: Geneva, Switzerland, 2008.
- 1332 175. Gouliarmou, V.; Mayer, P., Sorptive bioaccessibility extraction (SBE) of soils:
  1333 Combining a mobilization medium with an absorption sink. *Environmental Science* &
  1334 *Technology* 2012, 46, (19), 10682-10689.
- 1335 176. Gevao, B.; Mordaunt, C.; Semple, K. T.; Piearce, T. G.; Jones, K. C., Bioavailability 1336 of nonextractable (bound) pesticide residues to earthworms. *Environmental Science* & 1337 *Technology* **2001**, *35*, (3), 501-507.
- 1338 177. NEPC, Guideline on Site-Specific Health Risk Assessment Methodology. National 1339 Environment Protection Council (NEPC): Canberra, Australia, **2013**, (5), 92 p.
- 1340
  178. Rostami, I.; Juhasz, A. L., Assessment of persistent organic pollutant (POP)
  1341
  bioavailability and bioaccessibility for human health exposure assessment: A critical review.
  1342 *Critical Reviews in Environmental Science & Technology* 2011, *41*, (7), 623-656.
- 1343 179. Dean, J. R.; Ma, R. L., Approaches to assess the oral bioaccessibility of persistent organic pollutants: A critical review. *Chemosphere* **2007**, *68*, (8), 1399-1407.
- 1345 180. Ruby, M. V.; Lowney, Y. W.; Bunge, A. L.; Roberts, S. M.; Gomez-Eyles, J. L.; 1346 Ghosh, U.; Kissel, J. C.; Tomlinson, P.; Menzie, C., Oral bioavailability, bioaccessibility, and 1347 dermal absorption of PAHs from soil-state of the science. *Environmental Science* & 1348 *Technology* **2016**, *50*, (5), 2151-2164.
- 1349 181. Cui, X.-Y.; Xiang, P.; He, R.-W.; Juhasz, A.; Ma, L. Q., Advances in *in vitro* methods 1350 to evaluate oral bioaccessibility of PAHs and PBDEs in environmental matrices. 1351 *Chemosphere* **2016**, *150*, 378-389.
- 1352 182. Lal, V.; Peng, C.; Ng, J., A review of non-exhaustive chemical and bioavailability
  1353 methods for the assessment of polycyclic aromatic hydrocarbons in soil. *Environmental*1354 *Technology & Innovation* **2015**, *4*, 159-167.
- 1355 183. James, K.; Peters, R. E.; Cave, M. R.; Wickstrom, M.; Lamb, E. G.; Siciliano, S. D., 1356 Predicting polycyclic aromatic hydrocarbon bioavailability to mammals from incidentally

- 1357 ingested soils using partitioning and fugacity. *Environmental Science & Technology* 2016,1358 50, (3), 1338-1346.
- 1359 184. James, K.; Peters, R. E.; Laird, B. D.; Ma, W. K.; Wickstrom, M.; Stephenson, G. L.;

Siciliano, S. D., Human Exposure Assessment: A case study of 8 PAH contaminated soils
using in vitro digestors and the juvenile swine model. *Environmental Science & Technology* **2011**, *45*, (10), 4586-4593.

1363 185. Juhasz, A. L.; Aleer, S.; Adetutu, E. M., Predicting PAH bioremediation efficacy using
1364 bioaccessibility assessment tools: Validation of PAH biodegradation-bioaccessibility
1365 correlations. *International Biodeterioration & Biodegradation* 2014, *95*, 320-329.

1366 186. Juhasz, A. L.; Tang, W.; Smith, E., Using *in vitro* bioaccessibility to refine estimates
of human exposure to PAHs via incidental soil ingestion. *Environmental research* 2016, 145,
1368 145-153.

- 1369 187. Brand, E.; Lijzen, J.; Peijnenburg, W.; Swartjes, F., Possibilities of implementation of
  bioavailability methods for organic contaminants in the Dutch soil quality assessment
  1371 framework. *Journal of Hazardous Materials* **2013**, *261*, 833-839.
- 1372 188. Ortega-Calvo, J. J.; Tejeda-Agredano, M. C.; Jimenez-Sanchez, C.; Congiu, E.; 1373 Sungthong, R.; Niqui-Arroyo, J. L.; Cantos, M., Is it possible to increase bioavailability but 1374 not environmental risk of PAHs in bioremediation? *Journal of Hazardous Materials* **2013**, 1375 261, 733-745.
- 1376 189. Cachada, A.; Ferreira da Silva, E.; Duarte, A. C.; Pereira, R., Risk assessment of 1377 urban soils contamination: The particular case of polycyclic aromatic hydrocarbons. *Science* 1378 of *The Total Environment* **2016**, *551–552*, 271-284.
- 1379 190. Swartjes, F. A.; Rutgers, M.; Lijzen, J. P. A.; Janssen, P. J. C. M.; Otte, P. F.; 1380 Wintersen, A.; Brand, E.; Posthuma, L., State of the art of contaminated site management in 1381 The Netherlands: Policy framework and risk assessment tools. *Science of the Total* 1382 *Environment* **2012**, *427*, 1-10.
- 1383 191. NEPC, National Environmental Protection Measure (NEPM): Guideline on 1384 investigation levels for soil and groundwater. Schedule B1, F2013C00288. National 1385 Environment Protection Council (NEPC): Australia, **1999**, 87 p.
- 1386 192. Latawiec, A. E.; Simmons, P.; Reid, B. J., Decision-makers' perspectives on the use
  of bioaccessibility for risk-based regulation of contaminated land. *Environment International*2010, *36*, (4), 383-389.
- 1389 193. ATSDR, *Toxicological profile for polycyclic aromatic hydrocarbons (PAHs)*. Agency 1390 for Toxic Substances and Disease Registry (ATSDR), **1995**.
- 1391 194. Chung, N.; Alexander, M., Effect of concentration on sequestration and bioavailability
  1392 of two polycyclic aromatic hydrocarbons. *Environmental Science & Technology* **1999**, *33*,
  1393 (20), 3605-3608.
- 1394
  195. Johnson, D.; Jones, K.; Langdon, C.; Piearce, T.; Semple, K., Temporal changes in
  1395
  earthworm availability and extractability of polycyclic aromatic hydrocarbons in soil. Soil
  1396
  Biology & Biochemistry 2002, 34, (9), 1363-1370.
- 1397 196. White, J.; Hunter, M.; Pignatello, J.; Alexander, M., Increase in bioavailability of aged 1398 phenanthrene in soils by competitive displacement with pyrene. *Environmental Toxicology* 1399 *and Chemistry* **1999**, *18*, (8), 1728-1732.
- 1400 197. Hawthorne, S. B.; Grabanski, C. B.; Martin, E.; Miller, D. J., Comparisons of Soxhlet
  1401 extraction, pressurized liquid extraction, supercritical fluid extraction and subcritical water
  1402 extraction for environmental solids: recovery, selectivity and effects on sample matrix.
  1403 *Journal of Chromatography A* 2000, *892*, (1-2), 421-433.
- 1404
  198. Weber, W. J.; Young, T. M., A distributed reactivity model for sorption by soils and
  1405
  sediments. 6. Mechanistic implications of desorption under supercritical fluid conditions.
  1406 *Environmental Science & Technology* **1997**, *31*, (6), 1686-1691.
- 1407 199. Latawiec, A. E.; Reid, B. J., Sequential extraction of polycyclic aromatic 1408 hydrocarbons using subcritical water. *Chemosphere* **2010**, *78*, (8), 1042-1048.
- 1409 200. Latawiec, A. E.; Swindell, A. L.; Reid, B. J., Environmentally friendly assessment of
  organic compound bioaccessibility using sub-critical water. *Environmental Pollution* 2008,
  1411 156, (2), 467-473.

- 1412 201. Hickman, Z.; Reid, B., Towards a more appropriate water based extraction for the 1413 assessment of organic contaminant availability. *Environmental Pollution* **2005**, *138*, (2), 299-1414 306.
- 1415 202. Papadopoulos, A.; Paton, G. I.; Reid, B. J.; Semple, K. T., Prediction of PAH 1416 biodegradation in field contaminated soils using a cyclodextrin extraction technique. *Journal* 1417 of *Environmental Monitoring* **2007**, *9*, (6), 516-522.
- 1418 203. Cuypers, C.; Clemens, R.; Grotenhuis, T.; Rulkens, W., Prediction of petroleum 1419 hydrocarbon bioavailability in contaminated soils and sediments. *Soil & Sediment* 1420 *Contamination* **2001**, *10*, (5), 459-482.
- 1421 204. Dandie, C.; Weber, J.; Aleer, S.; Adetutu, E.; Ball, A.; Juhasz, A., Assessment of five
  1422 bioaccessibility assays for predicting the efficacy of petroleum hydrocarbon biodegradation
  1423 in aged contaminated soils. *Chemosphere* **2010**, *81*, (9), 1061-1068.
- 1424 205. Juhasz, A. L.; Naidu, R., Bioremediation of high molecular weight polycyclic aromatic 1425 hydrocarbons: a review of the microbial degradation of benzo[a]pyrene. *International* 1426 *Biodeterioration & Biodegradation* **2000**, *45*, (1-2), 57-88.
- 1427 206. Couling, N.; Towell, M.; Semple, K., Biodegradation of PAHs in soil: Influence of 1428 chemical structure, concentration and multiple amendment. *Environmental Pollution* **2010**, 1429 158, (11), 3411-3420.
- 1430 207. Wilson, S. C.; Jones, K. C., Bioremediation of soil contaminated with polynuclear 1431 aromatic hydrocarbons (PAHs): A review. *Environmental Pollution* **1993**, *81*, (3), 229-249.
- 1432 208. Dew, N.; Paton, G.; Semple, K., Prediction of [3-C-14]phenyldodecane
  1433 biodegradation in cable insulating oil-spiked soil using selected extraction techniques.
  1434 *Environmental Pollution* 2005, 138, (2), 316-323.
- 1435 209. Obuekwe, I.; Semple, K., Impact of Zn, Cu, Al and Fe on the partitioning and 1436 bioaccessibility of C-14-phenanthrene in soil. *Environmental Pollution* **2013**, *180*, 180-189.
- 1437 210. Obuekwe, I.; Semple, K., Impact of Al and Fe on the development of phenanthrene
- 1438 catabolism in soil. *Journal of Soils and Sediments* **2013**, *13*, (9), 1589-1599.