

1	Tuft Cells: a new flavor in innate epithelial immunity
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3	Richard K. Grencis ¹ & John J. Worthington ²
4	¹ Faculty of Life Sciences, University of Manchester, UK.
5	² Faculty of Health and Medicine, University of Lancaster, UK.
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7	Correspondence to: <u>richard.grencis@manchester.ac.uk</u> (R. K. Grencis) and
8	j.j.worthington@lancaster.ac.uk (J. J. Worthington)
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10	Abstract
11	How host cells sense intestinal parasitic infection and initiate the appropriate
12	immune response has long been a focus of many immunologists. Three new
13	papers now identify a critical role for tuft cells, an epithelial cell involved in
14	perception of taste, as key players that kick start Type 2 immunity.
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16	Main text
17	Infection with intestinal dwelling helminths is commonly associated with the
18	generation of type 2 immunity. The cytokine interleukin (IL-) 13 is critical in
19	driving this characteristic 'allergic' immune response[1] and is secreted by type
20	two innate lymphoid cells (ILC2) and CD4+T cells. ILC2 are believed to be major
21	initiators of type 2 immunity following parasitic infection[2], although a long-
22	standing question that still remains to be answered is how the expansion and
23	proliferation of ILC2, via production of the key epithelial cytokines IL-33 and IL-
24	25, is orchestrated?
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26	Three recent papers now provide a significant advance in our understanding of
27	this process. All have used intestinal dwelling nematodes (Nippostrongylus
28	brasiliensis and/or Heligmosomoides polygyrus), while one study used the enteric
29	protozoan (Tritrichomonas muris) as drivers of type 2 immunity. Coupled with
30	the use of a variety of transgenic mouse strains this powerful combination has
31	allowed a series of elegant and definitive experiments to collectively show that

- 32 the expansion of ILC2s is dependent on a much-neglected epithelial cell type –
- the tuft cell. Although discovered 60 years ago, relatively little is known about

tuft cell (also known as brush cell) function. It has been postulated that they play
a chemosensory role and indeed tuft cells encode genes involved in the
transduction of bitter and umami tastes [3].

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38 Using a "knock-in" mouse (Flare 25 – flox and reporter of *Il25*) von Moltke et al. 39 [4] were able to identify that the cells in the epithelium of the digestive tract (as 40 well as lung and gall bladder) expressing IL-25, under normal homeostatic 41 conditions, were indeed tuft cells. Importantly, N. brasiliensis infected Flare 25 42 mice exhibited a dramatic hyperplasia of tuft cells in the small intestine, 43 returning to homeostatic levels after worm expulsion. Infection with *H. polyayrus* 44 was also associated with a small intestinal tuft cell hyperplasia. Further studies 45 of *N. brasiliensis* infection in a panel of immunologically compromised mutant and transgenic mice confirmed that IL-13 was a key cytokine in tuft cell 46 47 hyperplasia operating through IL-4Rα. Moreover, studies using cytokine 48 administration in vivo and ex vivo intestinal organoids showed that IL-25 from 49 tuft cells acted upon ILC2 to induce IL-13 production which in turn promoted 50 further tuft cell hyperplasia in a feed-forward loop. These data suggest that 51 waves of tuft cell hyperplasia emanate from stem cell differentiation decisions in 52 the intestinal crypts, evident from the observed organoid tuft cell hyperplasia 53 upon Notch signalling inhibition.

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55 Data from Gerbe and colleagues [5] was very much in accordance with that of von 56 Moltke et al. [4] published in the same issue of Nature and demonstrating a 57 significant intestinal hyperplasia in tuft cells following helminth infection in 58 mice. They demonstrated that tuft cells expressed the Pou domain class 2, 59 transcription factor 3 (Pou2f3) and that Pou2f3 null mice contained no tuft cells 60 and depressed IL-25 expression even after infection with intestinal helminths. 61 This was also associated with a marked depression in a number of type 2 62 immune mediated changes, including goblet cell hyperplasia, Retnlß expression, 63 ILC2 expansion and IL-13 expression in intestinal tissue. Taken together, this 64 data supports a strong link between tuft cell hyperplasia and goblet cell 65 hyperplasia, the latter key to worm expulsion[6]. Accordingly, Pou2f3 null mice 66 showed a highly significant delay in *N. brasiliensis* expulsion with some worms

- 67 remaining in the intestine until day 42 post infection. Organoid cultures and
- 68 cytokine add back experiments using the Pou2f3 null mice confirmed that IL-13
- 69 acted downstream of the tuft cell lineage. This supported the concept of a
- 70 positive feedback loop in which ILC2 expansion, driven by tuft cell derived IL-25,
- 71 promoted IL-13 driven tuft cell hyperplasia with the resultant goblet cell
- 72 hyperplasia driving parasite expulsion from the intestinal tract.
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74 The paper from Howitt *et al.* [7]stemmed from observations of the number of 75 tuft cells (Dclk1 positive cells) in the intestinal tract, as part of a study into the 76 role of taste-chemosensory cells in recognition of intestinal microbes via G 77 protein couple receptors. They observed that mice bred in their facility exhibited 78 elevated numbers of tuft cells in comparison to previously published work. A 79 series of experiments identified that this was due to the presence of the common 80 protozoan T. muris. As intestinal helminths are also frequent infections of the 81 mammalian intestine, infection by N. brasiliensis, H. polygyrus or Trichinella 82 spiralis confirmed and extended the parasite driven tuft cell hyperplasia data 83 from the other two groups. The role of tuft cells as chemosensory cells was 84 explored using mice which lacked either the taste-specific G protein subunit 85 gustducin, or the transient receptor potential cation channel, subfamily M, 86 member 5 (TRMP5), a cation channel known to be important in the signaling 87 cascade of chemosensory cells in the gut. Gustducin null mice had significantly 88 fewer tuft cells after *T. muris* infection, while TRMP5 null mice also showed 89 blunted tuft cell and goblet cell hyperplasia. In vivo cytokine and ex vivo organoid 90 cultures in TRMP5 null mice showed IL-25 was produced by tuft cells and that 91 ILC2 were a critical part of tuft cell hyperplasia through secreting IL-13. The 92 authors concluded that the tuft cells may detect protozoans (and presumably 93 metazoan parasites) through TRMP5 taste chemoreception to initiate the tuft 94 cell/ILC2 feed-forward loop.

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96 These three papers make a major contribution to our understanding of the
97 initiation of type 2 immune responses to metazoan and protozoan enteric
98 parasites and provide a clear link between the epithelial barrier and the innate
99 immune response (Fig. 1). Whether this is the only function of tuft cells during

- 100 infection is unknown; tuft cells also possess the cellular machinery to influence
- 101 intestinal smooth muscle contraction, blood pressure and water balance, being
- 102 closely associated with nerve fibers and secreting leukotriene C4 and certain
- 103 opioids [8, 9]. Also, we are still to define the precise nature of the parasite
- 104 derived or infection induced molecules that are 'tasted' by the tuft cell.
- 105 Regardless, these studies highlight the rich biology of 'rare' cell populations in
- 106 the generation of immune responses to infection.
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- **Figure 1. Taste your parasites.** Helminth or infection induced molecules are
- 110 sensed via tuft cells expressing the transient receptor potential cation channel,
- subfamily M, member 5 (TRMP5) and gustducin-α. In response, tuft cells release
- the alarmin IL-25 which increases type two innate lymphoid cells (ILC2)
- 113 numbers and their secretion of the cytokine IL-13. In turn IL-13 signals to the
- 114 pluripotent stem cell niche within the intestinal crypt and promotes the
- 115 differentiation of tuft cells, possibly by altering Notch signaling. This tuft cell
- 116 hyperplasia causes a feed-forward loop and concurrent goblet cell hyperplasia,
- 117 leading to worm expulsion.
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