1	Increased soil P availability induced by faba bean root exudation stimulates root
2	growth and P uptake in neighbouring maize
3	Deshan Zhang ¹ , Chaochun Zhang ¹ , Xiaoyan Tang ¹ , Haigang Li ¹ , Fusuo Zhang ¹ , Zed
4	Rengel ² , William R. Whalley ³ , William J. Davies ⁴ , Jianbo Shen ^{1,*}
5	¹ Department of Plant Nutrition, China Agricultural University, Key Laboratory of
6	Plant-Soil Interactions, Ministry of Education, Beijing 100193, P. R. China
7	² Soil Science & Plant Nutrition, School of Earth and Environment, The UWA Institute
8	of Agriculture, The University of Western Australia, Crawley, WA 6009, Australia
9	³ Rothamsted Research, West Common, Harpenden, Hertfordshire, AL5 2JQ, UK
10	⁴ Lancaster Environment Centre, University of Lancaster, Lancaster LA1 4YQ, UK
11	
12	
13	With 4925-4969 words, 7 figures (2 tables in the supporting material)
14	
15	*Corresponding author:
16	Prof. Jianbo Shen
17	Department of Plant Nutrition
18	China Agricultural University
19	No.2 Yuan-ming-yuan West Road,
20	Beijing 100193
21	P. R. China
22	
23	Phone: +86 10 62732406
24	Fax: +86 10 62731016
25	E-mail: <u>jbshen@cau.edu.cn</u>

26 Summary

Root growth is influenced by soil nutrients and neighbouring plants, but how
these two drivers affect root interactions and regulate plant growth dynamics is
poorly understood. Here, interactions between the roots of maize (*Zea mays*) and
faba bean (*Vicia faba*) are characterized.

Maize was grown alone (maize) or with maize (maize/maize) or faba bean
 (maize/faba bean) as competitors under <u>five levels</u> <u>variable rates</u> of phosphorus
 (P) supply, and with homogeneous or heterogeneous P distribution.

 Maize had longer root length and greater shoot biomass and P content when grown with faba bean than with maize. At each P supply rate, faba bean had a smaller root system than maize but greater exudation of citrate and acid phosphatase, suggesting greater capacity to mobilize P in the rhizosphere. Heterogeneous P availability enhanced root-length density of maize but not faba bean. Maize root proliferation in the P-rich patches was associated with increased shoot P uptake.

Increased P availability by localized P application or by the presence of faba bean
 exudation stimulated root morphological plasticity and increased shoot growth in
 maize in the maize/faba bean mixture, suggesting root interactions of
 neighbouring plants can be modified by increased P availability.

45

Key words: Phosphorus uptake, *Zea mays* (maize), *Vicia faba* (faba bean), root
interactions, heterogeneous phosphorus supply, rhizosphere processes

48 Introduction

Root interactions among plants play an important role in determining the performance 49 of individuals in natural communities as well as crop productivity in agroecological 50 51 systems. The foraging capacity of roots is a key trait in belowground competition and is dependent on morphological (root architecture) and physiological plasticity (eg. 52 exudation of organic and inorganic compounds, nutrient uptake) in response to the 53 soil environment. To compete for soil resources (water and nutrients), plants invest in 54 root growth to maximize root length density and can out-compete neighbouring plants 55 (Cahill et al., 2010; Cahill & McNickle, 2011). For example, Rumex palustris 56 increased root growth in nutrient-rich areas and as a result became the superior 57 species in competition with Agrostis stolonifera (Mommer et al., 2012). In addition to 58 root growth, root physiological plasticity (ie. activity) can be critical in obtaining a 59 long-term competitive advantage (Fransen et al., 2001). Modelling studies have 60 61 suggested that physiological responses may be more important for the capture of patchy nitrogen (N) than morphological responses (Jackson & Caldwell, 1996; 62 Robinson, 1996; Dunbabin et al., 2003). 63

Root morphological and physiological plasticity is determined by various 64 environmental factors, such as availability of soil resources and their distribution 65 66 (Jackson et al., 1990; Hutchings & de Kroon, 1994; Hodge, 2004; Cahill et al., 2010; Andersen et al., 2014). Soil nutrient availability can profoundly affect root 67 morphology and physiology. For example, cluster-root formation and citrate 68 exudation in Lupinus albus can be induced by low phosphorus (P) conditions, and 69 70 inhibited by increased P supply (Shen et al., 2005; Lambers et al., 2006; Li et al., 2008). Faba bean roots have no significant growth response to localized nutrient 71 supply (Li et al., 2014a), suggesting relatively poor morphological plasticity to 72 variable nutrient availability, and potentially a low capacity to take full advantage of 73 nutrient-rich patches. However, in many species, root growth and distribution are 74 affected by soil nutrient heterogeneity (Drew, 1975; Li et al., 2008; Jing et al., 2010). 75 In response to nutrient-rich patches, plants tend to stimulate root growth and alter root 76

distribution, with increased root proliferation in the local nutrient-rich zone (Drew, 77 78 1975; Hodge et al., 1999; Hodge, 2004). In addition, some species show root physiological responses to localized nutrient enrichment (eg. an increase in the 79 nutrient uptake capacity per unit root length) (Jackson et al., 1990). Hence, foraging 80 for nutrients is determined by root morphological and physiological dynamics in 81 response to soil environmental conditions, and may result in differences in 82 competitive ability, which, to a large extent, affects plant productivity and nutrient 83 uptake in a mixed system. 84

Growing mixtures of maize and faba bean is used widely to improve grain yield 85 and P-use efficiency in cropping ecosystems in many parts of the world (intercropping) 86 (Li et al., 2007; Zhang et al., 2010; Shen et al., 2011, 2013). Root interactions have a 87 profound impact on P uptake and yield in the maize/faba bean system (Li et al., 2003, 88 2014b), with rhizosphere acidification caused by faba bean increasing mobilization of 89 organic/inorganic P sources and facilitating P uptake by target maize (Li et al., 2007). 90 The roots of maize and faba bean intermingle and appear to grow together; hence, 91 complementarity of the spatial root distribution of intercropped species contributes to 92 interspecies facilitation in maize/faba bean intercropping compared to the 93 maize/maize monocropping system (Li et al., 2006). However, the detailed 94 mechanisms underlying root interactions in the maize/faba bean system are still 95 unclear. 96

97 Much work on root traits in response to P supply has been conducted in maize or 98 faba bean separately (Li *et al.*, 2014a), but how altered nutrient gradients and spatial 99 distribution affect root-root interactions in the mixture, and thus regulate plant growth 100 dynamics, is largely unknown. Maize and faba bean roots grow well together, but the 101 root interactions based on complementary and niche differentiation in space might be 102 enhanced with a better understanding of how spatial changes in soil nutrient 103 availability affect root growth, distribution and hence competition.

To investigate the effects of heterogeneous nutrient availability on root growth, distribution and thus competition intensity in the maize/faba bean mixture system, we conducted experiments with varying P supplies and homogeneous or heterogeneous P distribution to test the hypothesis that neighbouring faba bean would not compete
strongly with target maize because faba bean roots have low morphological plasticity
that is compensated for by significant physiological plasticity in variable soil P supply.
We then demonstrate that increased maize growth in the maize/faba bean mixtures is
consistent with the hypothesis that spatial heterogeneity in P availability induced by
faba bean exudation would stimulates root morphological plasticity in maize,
resulting in improved maize shoot growth and nutrient uptake.

114 Materials and Methods

115 Experimental set-up

116 Experiment 1

To investigate how maize roots respond to faba bean in soil with variable P supply, 117 and test how soil P availability regulates root interactions between maize and faba 118 119 bean, a pot experiment was conducted in a glasshouse with five soil P supply levels and three cropping treatments. Maize was grown alone as a single species (single 120 maize treatment), mixed with other maize plants (maize/maize treatment) or 121 intercropped with faba bean (maize/faba bean treatment) in soil supplied with 0, 50, 122 100, 200 or 500 mg P kg⁻¹ soil, corresponding to soil Olsen-P values of 3.5, 36, 51, 123 123 and 270 mg kg⁻¹, respectively. We choose the soil P supply levels based on a 124 preliminary experiment with the same soil: (1) deficient: 0 and 50 mg P kg⁻¹ soil; (2) 125 moderate: 100 mg P kg⁻¹ soil; (3) adequate: 200 mg P kg⁻¹ soil, and (4) high: 500 mg P 126 kg⁻¹ soil (to mimic high-input maize cropping system in intensive agriculture of 127 China). There were 15 treatment combinations in the study, with four replicates per 128 129 treatment.

The soil was collected from Shangzhuang experimental station in Beijing, China, air-dried and passed through a 2-mm sieve. Soil properties were as follows: Olsen-P 3.5 mg kg⁻¹, organic C 11.5 g kg⁻¹, total N 0.72 g kg⁻¹, available N 8.5 mg kg⁻¹ (NO₃⁻¹ and NH₄⁺), available K 32.3 mg kg⁻¹, and pH 8.2 (the ratio of soil to CaCl₂ solution was 1:2.5). The pot was filled with 1.5 kg of air-dried soil. To ensure the nutrient supply was adequate for plant growth, soil was also fertilized with basal nutrients at the following rates (mg pot⁻¹): Ca(NO₃)₂·4H₂O, 1687; K₂SO₄, 200; MgSO₄·7H₂O, 65; Fe-EDTA, 8.78; MnSO₄·H₂O, 10; ZnSO₄·7H₂O, 15; CuSO₄·5H₂O, 3; H₃BO₃, 1; and Na₂MoO₄·5H₂O, 0.25. Phosphorus was applied as Ca(H₂PO₄)₂·H₂O.

The genotype of maize (*Zea mays* L.) was cv. ZD958, and the genotype of faba bean (*Vicia faba* L.) was cv. Lincan5. Maize and faba bean seeds were surface-sterilized in 30% v/v H_2O_2 for 20 min, washed with deionized water, soaked in CaSO₄ saturated solution for 12 hours and then germinated in Petri dishes covered with wet filter papers for 1-2 days at 22°C. All the pots were arranged in a completely randomized design, and were re-randomized weekly during the experiment. The plants were watered every day to maintain field capacity (18%, w/w).

Plants were harvested 38 days after sowing (DAS) and separated into shoots and 146 roots. Following root excavation, the soil adhering to roots was defined as rhizosphere 147 soil and was sub-sampled for carboxylate and acid phosphatase measurements. Roots 148 149 were transferred to a tube containing 50 mL of 0.2 µM CaCl₂ and gently shaken to dislodge the rhizosphere soil, followed by shaking for 5-10s to create homogeneous 150 suspension. A suspension volume of 10 mL was taken by pipette to a 10-mL 151 centrifuge tube for carboxylate analysis by HPLC (see below), and a 0.5 µL aliquot of 152 suspension was placed in a 2-mL centrifuge tube for acid phosphatase measurement 153 (see below). 154

155 *Experiment 2*

To address the question of how soil P heterogeneity affects root interactions between maize and faba bean, we set up a rhizo-box experiment comprising two P supply treatments and three cropping treatments (maize, maize/maize and maize/faba bean) with four replicates of each treatment. We chose the same soil, genotypes of maize and faba bean, and crop treatments as in experiment 1.

161 To record root growth and distribution, we constructed microcosm 162 PVC-rhizo-boxes ($20 \times 1.5 \times 35$ cm, Fig. 1), containing irrigation holes, a viscose 163 fleece for moisture distribution, transparent plastic foil for soil-covering and a Perspex 164 front lid with screws. All rhizo-boxes were filled with 1.3 kg air-dried soil.

165	Phosphorus was supplied as Ca(H ₂ PO ₄) ₂ ·H ₂ O in a homogeneous or heterogeneous
166	pattern. For the heterogeneous P treatment, a 3.5-cm P-rich layer (227.5 g soil)
167	containing 46.2 mg P (200 mg P kg ⁻¹ soil) was manually mixed and placed at the
168	center of the rhizo-box (Fig. 1), referred to as the P-rich patch, and the remaining soil
169	without P additions was the background soil (1072.5 g soil). For the homogeneous P
170	treatment, the same total P (46.2 mg P) was spread evenly throughout the soil (35 mg
171	P kg ⁻¹ soil).
172	Maize and faba bean seeds were handled as in experiment 1. To ensure nutrient
173	supply for plant growth, the same amounts of basal nutrients as in experiment 1 were
174	added to soil, followed by thorough mixing.
175	Before planting, all rhizo-boxes were irrigated through the bottom irrigation holes.
176	After 15 days of growth, the rhizo-boxes were irrigated every 2 days until the final
177	harvest. After 30 days of growth, the target species (maize) and neighbouring species
178	were separated, and shoots and roots of each species were collected for biomass
179	measurements. Shoot P content was analysed in maize. In the heterogeneous

Comment [z1]: How were the P additions done?

Phosphorus-rich soil (227.5 g soil) with 46.2 mg P concentrated in a single rectangular column (3.5 cm length, spanned the width and the deep of the rhizo-boxes, $3.5 \text{ cm} \times 1.5$ $cm \times 35 cm$), and the background soil (1072.5 g soil) without P added, were separately placed in the rhizo-boxes, refer to Fig. 1.

in the homogeneous treatments. 182 Both experiments were conducted in a glasshouse at China Agricultural University, 183 184 Beijing (latitude: 40° 01' N, longitude: 116°16' E). In experiment 1, temperature in the glasshouse was maintained at 21-25 °C during the day and 15-18 °C at night, with 185

treatments, roots grown in vs. out of the P-rich patch were sampled separately for

length measurement, and the corresponding roots at the same location were sampled

12-14 h daytime throughout the growth period. In experiment 2, temperature was 186

maintained at 24-28 °C during the day and 18-20 °C at night, with 14-19 h daytime. 187

Measurements 188

180

181

Plant biomass and phosphorus uptake 189

190 Shoots were oven-dried at 105°C for 30 min and then at 65°C for 3 days before weighing for dry biomass determination. Phosphorus concentration in shoots was 191 determined after digestion with a mixture of 5 mL of concentrated sulphuric acid and 192

Comment [z2]: Why were there different environmental conditions for the two experiments?

We did these two experiments separately.

193 8 mL of 30% v/v H_2O_2 . Shoot P was analyzed by the molybdovanadophosphate

method at 440 nm by spectrophotometry (Varian Vista–Pro CCD) (Johnson & Ulrich,1951959).

196 *Root parameter measurement*

Roots were washed in deionized water and then scanned with an EPSON root scanner
at 400 dots-per-inch resolution (Epson Expression 1600 pro, Model EU-35, Japan).
The total root length was analyzed with software Win-RHIZO (Regent Instruments
Inc., Quebec, QC, Canada).

201 Carboxylate and acid phosphatase exudation

Carboxylates in the rhizosphere soil were analyzed using a reversed phase high-performance liquid chromatography (HPLC) system according to a previous report (modified from Shen *et al.*, 2003 and Wang *et al.*, 2010). The chromatographic separation was conducted on a 250×4.6 mm reversed-phase column (Alltima C18, 5 Micrometers; Alltech Associates, Inc., Deerfield, IL, USA). The mobile phase was 25 mmol L⁻¹ KH₂PO₄ (pH 2.25) with a flow rate of 1 mL min⁻¹ at 31°C. Detection of carboxylates was carried out at 214 nm.

To determine the activity of acid phosphatase in the rhizosphere soil, 0.5 mL aliquots of soil suspensions were transferred into a 2-mL Eppendorf vial with 0.4 mL sodium acetate buffer and 0.1 mL *p*-nitrophenyl phosphate (NPP) substrate added. Vials were incubated at 30°C for 60 min, and the reaction was terminated by adding 0.5 mL of 0.5 M NaOH. Absorption was measured at 405 nm (Alvey *et al.*, 2000).

214 Statistical analyses

To investigate the effect of faba bean on target maize, the Relative Interaction 215 216 Intensity (RII) was calculated in experiment 1. RII can be used to compare the performance of plants growing in mixtures or alone in order to estimate the magnitude 217 of competitive responses of the target plant species (Markham & Chanway, 1996; 218 219 Howard & Goldberg, 2001; Weigelt & Jolliffe, 2003). Calculation of RII was based on the method proposed by Wilson & Keddy (1986) and modified by Markham & 220 Chanway (1996). A formula of Markham & Chanway (1996) was adopted to calculate 221 222 RII (modified from Valladares et al., 2006).

$$RII = \frac{\sum(\frac{xi-xi}{x})}{n}$$

where *xi* is shoot biomass of target maize in the presence of neighbours and *xi'* is
biomass of target maize in the absence of neighbours. X is either *xi* or *xi'*,
whichever was larger. RII is symmetrical around zero and constrained between +1
and -1 (Markham & Chanway, 1996), so that magnitudes of competition or
facilitation can be compared. If maize is unaffected by <u>its</u> neighbour, RII will be zero.
Negative RII values represent the relative competition between neighbouring plants;
positive values indicate that target maize can benefit from neighbour presence.

The *i* and *i*' are two randomly selected individuals (in the present study, i =1, ..., 4, because each treatment had 4 replicates) of the same species belonging to two different cropping treatments. The n is the number of (xi - xi') values. In the present study, n is equal to 16 because 4 replicates (4 random individuals) were set in each pair of cropping treatments (single maize and maize/maize, or single maize and maize/faba bean).

Analysis of variance (ANOVA) was conducted using the SPSS statistical software (SPSS19.0). Significant differences among means were separated by LSD at the $P \le$ 0.05 probability level. Plant growth and root length were subjected to two-way ANOVA to assess the effects of plant species, soil P supply, and their interaction in experiment 1 as well as the effects of heterogeneous/homogeneous P supply and competition in experiment 2.

242 **Results**

243 Root interactions between maize and faba bean grown with variable soil P supply 244 (exp 1)

Shoot biomass and P content in target maize increased with soil P supply (Fig. 2).
Compared with the single maize treatment at five levels of P supply, shoot biomass of
target maize in the mixtures was not affected by the presence of faba bean, but was
significantly reduced when maize was grown with maize (Fig. 2a). Shoot P content of

target maize showed a similar response to shoot biomass when grown either as single 249 or with neighbours (Fig. 2b). In the maize/maize mixture, a decrease in maize shoot P 250 content was affected by soil P availability and the presence of neighbouring maize, 251 however, there was no significant interaction (Fig. 2b; Table S1). In the maize/faba 252 bean mixture, shoot P content in target maize increased with soil P supply, but not 253 with the neighbour presence; the interactive effect was not significant (Fig. 2b; Table 254 S1). Phosphorus content in shoots of target maize was higher in the maize/faba bean 255 than maize/maize mixtures at 36 and 270 mg kg⁻¹ (soil Olsen-P levels). When 256 comparing shoot P content in target maize between the maize/maize and maize/faba 257 bean treatments, we found the <u>a</u> main effects of P supply and neighbour, as well as the 258 their interaction (Fig. 2b; Table S1) had a significant influence. The interaction was 259 260 significant because the effect of faba bean on increasing maize shoot P content (compared to competing maize plants) became more pronounced as soil P supply 261 262 increased (Fig. 3).

Relative interaction intensity (RII) of maize grown with maize decreased with soil P supply, whereas in the maize/faba bean mixture, RII of target maize increased from 3.5 to 36 mg Olsen P kg⁻¹ and remained unchanged with further increases in soil P (Fig. 3). RII of maize in maize/maize was significantly lower than that of maize grown with faba bean, except in the lowest P treatment (3.5 mg Olsen P kg⁻¹) (Fig. 3). Compared with the maize/maize mixture, RII in maize/faba bean was significantly greater in treatments with Olsen P at 36, 51, 123 and 270 mg kg⁻¹.

Root length of target maize in the maize/faba bean or maize/maize mixture 270 significantly increased with P supply (Fig. 4; Table S1). In the maize/faba bean 271 mixture, root length reached the maximum at 51 mg Olsen P kg⁻¹ and remained 272 similar at higher P supplies (Fig. 4), with non-significant effects of neighbour and 273 interaction (Fig. 4; Table S1). Root length of maize when competing with maize 274 attained the highest value at 123 mg Olsen P kg⁻¹, and was significantly influenced by 275 soil P supply and neighbour, but there was no interactive effect (Fig. 4; Table S1). 276 Compared with the maize/maize mixture, root length of maize in maize/faba bean was 277 278 higher (Fig. 4). The difference in root length of target maize between maize/faba bean

Comment [z3]: Would it be more useful to talk about increased root growth in terms of relative biomass allocation? I suspect that while root growth increases with increasing P, it's a relatively smaller fraction of total biomass.

See my comments

and maize/maize mixtures depended on soil P supply, competing plant species and the
interaction (Fig. 4; Table S1), with the effect of faba bean on increasing maize root
length (relative to competing maize plants) being largest at intermediate P supply
(36-51 mg Olsen P kg⁻¹) (Fig. 4).

For the neighbouring species (faba bean or maize), root length was significantly lower in faba bean than maize, except at 3.5 mg Olsen P kg⁻¹ (Fig. 5a). There was no effect of P supply level on root length of neighbouring faba bean, but root length of neighbouring maize significantly increased with increasing P supply (Fig. 5a).

Citrate concentration and the acid phosphatase activity were significantly greater in 287 the rhizosphere of faba bean than maize (Fig. 5b, 5c). Increased P supply depressed 288 citrate exudation by faba bean. In contrast, the activity of acid phosphatase in the faba 289 bean rhizosphere was increased from the lowest P treatment (3.5 mg Olsen P kg⁻¹) to 290 123 mg Olsen P kg⁻¹, and then decreased with a further increase in P supply. There 291 were no evident effects of P treatments on citrate exudation and secretion of acid 292 293 phosphatase in maize, which were at relatively low values compared with faba bean 294 (Fig. 5b, 5c).

The effect of spatial variability in P supply on root interactions between maize and faba bean (exp. 2)

Shoot biomass of target maize in the maize/maize and maize/faba bean mixtures was significantly stimulated by heterogeneous P supply, although the total amount of P applied was the same between <u>for</u> the homogeneous and heterogeneous treatments. A similar trend was found in the single maize treatment, but the differences were not significant (Fig. 6a). When soil P was supplied heterogeneously, <u>the</u> biomass of target maize was 100% higher when grown with faba bean than with maize.

Soil P heterogeneity and neighbouring species both influenced shoot P content in target maize (Fig. 6b; Table S2). Compared to homogeneous P supply, the P content in shoots of target maize in <u>the</u> heterogeneous environment increased by 139%, 144% and 75% in single maize, maize/maize and maize/faba bean, respectively (Fig. 6b). In the maize/maize mixture, shoot P content of target maize was influenced by P heterogeneity only (Fig. 6b; Table S2). However, in mixture with faba bean, it was significantly affected by both P heterogeneity and neighbouring species (faba bean),
but there was no interactive effect (Fig. 6b; Table S2). When the two mixtures were
compared, shoot P content in target maize was higher in the maize/faba bean than
maize/maize mixtures, which was influenced by soil P supply pattern and
neighbouring species, but the interaction was not significant (Fig. 6b; Table S2). Thus,
shoot P content of maize was significantly increased by heterogeneous P supply. as
well as the presence of faba bean, compared with the maize/maize system.

Root length density (RLD) of target maize was greater in the P-rich soil volume in 316 the heterogeneous than that in the same place in homogeneous treatments $(\frac{3.9, 2.9}{3.9, 2.9})$ 317 and 2.4 times greater in the single maize, maize/maize and maize/faba bean systems, 318 respectively, Fig. 7a). In heterogeneous environment, RLD of target maize did not 319 320 differ among the single maize, maize/maize and maize/faba bean treatments, whereas RLD of maize grown in homogeneous environment was higher in the maize/faba bean 321 than single maize treatment. Root length density (RLD) of target maize in the P-rich 322 323 soil volume was significantly influenced by heterogeneous P supply and not by neighbour competition in either maize/maize or maize/faba bean systems (Fig. 7a; 324 325 Table S2).

Outside the P-rich soil volume, RLD of target maize was increased significantly 326 with heterogeneous compared with homogeneous P supply in the maize/maize 327 treatment, but the effect was not significant in the single maize and maize/faba bean 328 treatments (Fig. 7b). The increased RLD outside the P-rich soil volume of maize in 329 maize/maize was caused by heterogeneity, but there was no effect of neighbour 330 presence (Fig. 7b; Table S2). The heterogeneous P supply increased maize RLD 331 332 outside the P-rich soil volume in maize/faba bean in comparison with single maize, but there was no significant difference between the maize/faba bean and maize/maize 333 mixtures (Fig. 7b). In the maize/faba bean system, maize RLD outside the P-rich soil 334 volume was enhanced by neighbour presence, but not by heterogeneity (Fig. 7b; Table 335 S2). Compared with the maize/maize and maize/faba bean treatments, RLD of target 336 maize outside the P-rich soil volume was influenced by soil P heterogeneity and 337 338 neighbouring species (Fig. 7b; Table S2).

Comment [z4]: Does not make sense-please revise

339 Discussion

In the present study, the maize/faba bean treatment was compared to maize alone or 340 maize/maize treatment. Any neighbours with maize would potentially represent 341 342 competition, but our findings indicated neighbouring faba bean was less competitive 343 than neighbouring maize (Fig. 3). With increasing P supply level, relative interaction 344 intensity (RII) of target maize in maize/maize mixtures significantly decreased (Fig. 3), suggesting an increase in competition intensity with soil P supply from 3.5 to 270 345 mg Olsen-P kg⁻¹. According to Hess & de Kroon (2007), plants can sense and 346 347 coordinate root growth based on available soil volume and a nutrient supply pattern. In the present study, in the maize/maize mixtures at low P supply, maize shoot growth 348 was P-limited at low P supply (Fig. 2a), which would have lowered the translocation 349 of photosynthetic products to belowground parts for production of new rootswas also 350 relatively consistent with low root length (Fig. 4), resulting in low competition for the 351 352 same soil P in the maize/maize mixtures. However, as root growth significantly 353 increased with an increase in soil P supply level, the intensity of maize competition in the maize/maize mixtures for the same available soil volume was strengthened, 354 355 resulting in a lower more negative RII and thus high competition at higher soil P 356 supply.

357 In the maize/faba bean mixture, the competition intensity remained unimodal and was lower in the maize/faba bean than maize/maize system at the same soil P supply 358 (Fig. 3). The effect of soil P availability on shoot P content and root length of target 359 maize was significantly dependent on neighbouring species, and was greater in the 360 361 maize/faba bean than maize/maize mixtures (Table S1). Target maize shoot P content was significantly influenced decreased by neighbouring maize (P<0.001) but not faba 362 363 bean (P=0.071) (Table S1), suggesting that the presence of faba bean resulted in lower competition intensity than the presence of maize. Previous studies indicated that plant 364 species win competition for a shared resource by using of the resources available in 365 hotspots more rapidly as a result of greater root growth plasticity (Grime, 1994; 366 Craine et al., 2005), or by depleting the resource to lower concentrations than their 367

Comment [z5]: Specify increased soil P supply.

Comment [z6]: This is speculative—you have actual root length numbers, why not refer to them?

Revised here.

competitors (Tilman, 1982). In the present study, the difference between root 368 369 competition in the maize/faba bean and maize/maize mixtures could be explained in two ways. Firstly, lower root length of faba bean compared to neighbouring maize 370 could provide a greater available soil volume for target maize roots to exploit, 371 indicating the competitive importance of morphological root responses to the abiotic 372 environment (similarly, plasticity in root morphology of Pseudoroegneria spicata 373 improved its capacity to withstand competition from the more vigorous, but less 374 plastic, Agropyron desertorum; Huber-Sannwald et al., 1996). Secondly, a larger 375 amount of citrate and acid phosphatase in the faba bean rhizosphere soil would have 376 improved mobilization of sparingly-soluble soil P, thus effectively increasing the 377 amount of accessible P for target maize (see: Hinsinger, 2001; Jones et al., 2003). 378 379 Hence, nutrient mobilization by root exudation could lead to increased acquisition of 380 limiting resources by decreasing intensity of interspecies competition through niche complementarity (i.e. maize and faba bean accessing different P fractions: it was 381 382 found that faba bean can access to sparingly soluble P (or unavailable for maize) through root exudation, but maize can mainly use soluble or available soil P, 383 indicating a niche complementarity) (Hinsinger et al., 2011; Shen et al., 2011, 2013; 384 Brooker et al., 2014; Li et al., 2014b). This result supports hypothesis 1 that 385 neighbouring faba bean provides low competition intensity to target maize by its 386 relatively low root morphological plasticity, which was compensated for by high 387 physiological plasticity (strong exudation) to alter soil P availability in the maize/faba 388 389 bean system.

In response to heterogeneous P supply in the present study, maize showed 390 391 significant root proliferation in the nutrient-rich soil volume (Fig. 7a), which conferred competitive advantage to maize in heterogeneous compared to 392 homogeneous nutrient supply (see also Robinson et al., 1999); hence, shoot P content 393 394 in maize was greater in the heterogeneous than homogeneous P treatments in the maize/faba bean mixture (Fig. 6b). The reasons why faba bean contributed to 395 increased shoot P content in maize in heterogeneous P supply could be explained as 396 397 follows. Firstly, heterogeneous P supply increased maize root proliferation (but not **Comment [z7]:** Aren't the roots accessing the same P fractions, once the faba exudates have made the P available?

See my comments

that of faba bean, data not shown) in P-enriched soil (Fig. 7), as in the earlier study 398 399 (Li et al., 2014a) in which heterogeneous nutrient supply did not influence faba bean root growth in the single faba bean treatments. Increased maize root proliferation in 400 the localized fertilizer zone could cause higher competition intensity in maize/maize 401 compared with maize/faba bean. Secondly, faba bean roots were located mostly 402 outside P-enriched zones (i.e. in soil not fertilized with P), and shoot biomass and P 403 content in faba bean were similar in heterogeneous and homogeneous P supply 404 treatments (data not shown), which is consistent with the previous findings that shoot 405 growth and nutrient uptake of single faba bean was not influenced by heterogeneous P 406 availability (Li et al., 2014a). 407

The high physiological plasticity of faba bean in terms of exudation of carboxylates 408 409 and acid phosphatase could mobilize soil P and increase P availability to provide adequate P supply for its own growth and could even contribute to satisfying demand 410 of neighbouring plants (see experiment 1). A shallow root system of faba bean (Li et 411 412 al., 2006) provided a niche for maize roots to increase root length density (Fig. 7b) and P uptake (Fig. 6b) compared with maize in the maize/maize treatment. Hence, 413 414 increased P uptake by maize may be attributed to root interactions between maize and faba bean being facilitated in the heterogeneous P supply treatment. The results 415 indicated that spatial heterogeneity in P availability the increased soil P availability 416 417 induced by physiological root plasticity in faba bean stimulated morphological root plasticity in maize, resulting in improved maize growth and P uptake. In contrast, in 418 the maize/maize mixture, high morphological root plasticity caused strong 419 competition, further intensified in the heterogeneous P treatment by greater root 420 421 proliferation in the P-rich patches, and resulting in lower shoot growth and nutrient uptake compared with maize grown with faba bean. The results provide support for 422 hypothesis 2 that maize would benefit from the root physiological traits of 423 424 neighbouring faba bean as well as from heterogeneous P supply.

Even though the present study showed that root interactions could be significantly influenced by heterogeneous P supply and root traits of neighbouring species in the maize/faba bean and maize/maize mixtures, we do not know yet how common such

interactions might be in other agroecological systems. In maize/faba bean 428 429 intercropping, a more asymmetric root distribution between maize and faba bean would occur in heterogeneous than homogeneous nutrient supply (cf. Schenk, 2006), 430 probably resulting in an increasing potential for spatial niche complementarity as 431 mentioned above. Interspecies facilitation based on nutrient-mobilizing mechanisms 432 (Shen et al., 2011, 2013; Li et al., 2014b) may be used to help explore potential P 433 resources in soils through selecting neighbouring species or optimizing plant 434 interactions based on root proliferation and the rhizosphere processes to alter 435 competition. Alternatively, a plant species can achieve competitive superiority by 436 depleting nutrient resources to lower concentrations than its competitors (Tilman, 437 1982). In the present study, maize had competitive superiority for nutrient acquisition 438 439 not only by stimulating root growth in the presence of faba bean, but also by 440 exploiting additional nutrient resources made available in the rhizosphere soil of faba bean. Nutrient spatial heterogeneity availability induced increased by neighbouring 441 442 faba bean plants could modify maize root growth and thus change root interactions in the maize/faba bean mixture. This study provided novel support for the 443 444 nutrient-driven root interactions regulating growth dynamics of plant species in the mixed stand. Furthermore, heterogeneous nutrient supply could be considered a useful 445 strategy for modifying root/rhizosphere interactions to optimize plant combinations 446 and underpin improved crop productivity and nutrient uptake in the agroecological 447 448 systems.

449 Conclusions

Both nutrient supply and neighbouring species significantly modified root interactions between maize and faba bean, influencing P uptake and biomass production. At different soil P <u>suppliessupply</u>, faba bean exhibited relatively poor morphological, but strong physiological root plasticity. This contributed to relatively low competition intensity in the maize/faba bean compared with maize/maize mixture. Localized P supply enhanced root proliferation of target maize (but not faba bean) in the

maize/faba bean mixture. Enhanced root exudation of citrate and acid phosphatase by 456 faba bean could facilitate soil P availability to benefit maize growth in the maize/faba 457 bean mixture. The study provided the new insights into root/rhizosphere interactions 458 in the maize/faba bean intercropping in the systems with variable P supply, which is 459 important in developing strategies for rhizosphere management through optimizing 460 plant combinations and soil nutrient supply to increase crop productivity and 461 nutrient-use efficiency. The challenge of enhancing food supply without using extra 462 463 nutrients could be appropriate accomplished by maximizing the efficacy of intercropping as a means of delivering more crop production per drop-unit of fertilizer 464 465 rate.

466 Acknowledgements

This study was supported by the National Natural Science Foundation of China (31330070, 30925024, 31210103906), the National Basic Research Program (973-2015CB150405) and the Innovative Group Grant of the National Science Foundation of China (31421092). ZR is supported by Australian Research Council (DP130104825). WRW is supported at Rothamsted Research by the 20:20 Wheat® project.

473 **References**

Alvey S, Bagayoko M, Neumann G, Buerkert A. 2000. Cereal/legume rotation
effects in two West African soils under controlled conditions. *Plant and Soil* 231:
45-54.

- Andersen SN, Dresbøll DB, Thorup-Kristensen K. 2014. Root interactions between
 intercropped legumes and non-legumes-a competition study of red clover and red
 beet at different nitrogen levels. *Plant and Soil* 378: 59-72.
- 480 Brook RW, Bennett AE, Cong WF, Daniell TJ, George TS, Hallett PD, Hawes C,
- 481 Iannetta PPM, Jones HG, Karley AJ, Li L, McKenzie BM, Pakeman RJ,
- 482 Paterson E, Schöb C, Shen J, Squire G, Watson CA, Zhang CC, Zhang FS,

Comment [z8]: too conversational.

- Zhang JL, White PJ. 2014. Improving intercropping: a synthesis of research in
 agronomy, plant physiology and ecology. *New Phytologist* 3: 1-11.
- 485 Cahill JF, McNickle GG, Haag JJ, Lamb EG, Nyanumba SM, Clair CCS. 2010.
- 486 Plants integrate information about nutrients and neighbors. *Science* **328**: 1657.
- Cahill JF, McNickle GG. 2011. The behavioral ecology of nutrient foraging by
 plants. *Annual Review of Ecology, Evolution, and Systematics* 42: 289-311.
- 489 Craine JM, Fargione J, Sugita S. 2005. Supply preemption, not concentration
 490 reduction, is the mechanism of competition for nutrients. *New Phytologist* 166:
 491 933-940.
- 492 Drew MC. 1975. Comparison of the effects of localized supply of phosphorus, nitrate,
 493 ammonium and potassium on the growth of the seminal root system, and the
 494 shoot, in barley. *New Phytologist* 75: 479-490.
- Dunbabin V, Diggle A, Rengel Z. 2003. Is there an optimal root architecture for
 nitrate capture in leaching environments? *Plant, Cell and Environment* 26:
 835-844.
- Fransen B, de Kroon H, Berendse F. 2001. Soil nutrient heterogeneity alters
 competition between two perennial grass species. *Ecology* 82: 2534-2546.
- 500 Grime JP. 1994. The role of plasticity in exploiting environmental heterogeneity. In:
- 501 Caldwell MM, Pearcy RW, editors. *Exploitation of environmental heterogeneity*
- by plants: ecophysiological processes above- and below-ground. San Diego, CA,
 USA: Academic Press, 1-19.
- Hess L, de Kroon H. 2007. Effects of rooting volume and nutrient availability as an
 alternative explanation for root self/non-self discrimination. *Journal of Ecology*95: 241-251.
- Hinsinger P. 2001. Bioavailability of soil inorganic P in the rhizosphere as affected
 by root-induced chemical changes: a review. *Plant and Soil* 237: 173-195.
- Hinsinger P, Betencourt E, Bernard L, Brauman A, Plassard C, Shen JB, Tang
 XY, Zhang FS. 2011. P for two, sharing a scarce resource: soil phosphorus
 acquisition in the rhizosphere of intercropped species. *Plant Physiology* 156:
 1078-1086.

Hodge A. 2004. The plastic plant: root responses to heterogeneous supplies of
nutrients. *New Phytologist* 162: 9-24.

- 515 Hodge A, Robinson D, Griffiths BS, Fitter AH. 1999. Why plants bother: root
- proliferation results in increased nitrogen capture from an organic patch when

517 two grasses compete. *Plant, Cell & Environment* **22**: 811-820.

- 518 Howard TG, Goldberg DE. 2001. Competitive response hierarchies for germination,
- growth, and survival and their influence on abundance. *Ecology* **82**: 979-990.
- Huber-Sannwald E, Pyke DA, Caldwell MM. 1996. Morphological plasticity
 following species-specific recognition and competition in two perennial grasses. *Annals of Botany* 83: 919-931.
- 523 Hutchings MJ, de Kroon H. 1994. Foraging in plants: the role of morphological
- 524 plasticity in resource acquisition. *Advances in Ecological Research* **25**: 159-238.
- Jackson RB, Manwaring JH, Caldwell MM. 1990. Rapid physiological adjustment
 of roots to localized soil enrichment. *Nature* 344:58-60.
- Jackson RB, Caldwell MM. 1996. Integrating resource heterogeneity and plant
 plasticity: modelling nitrate and phosphate uptake in a patchy soil environment.
 Journal of Ecology 84: 891-903.
- Jing JY, Rui Y, Zhang FS, Rengel Z, Shen JB. 2010. Localized application of
 phosphorus and ammonium improves growth of maize seedlings by stimulating
 root proliferation and rhizosphere acidification. *Field Crops Research* 119:
 335-364.
- Jones DL, Dennis PG, Owen AG, van Hees PAW. 2003. Organic acid behaviour in
 soils: misconceptions and knowledge gaps. *Plant and Soil* 248: 31-41.
- **Johnson CM, Ulrich A. 1959.** Analytical methods for use in plant analysis.
- 537 University of California, Agricultural Experiment Station, Berkeley, CA.
- Lambers H, Shane MW, Cramer MD, Pearse SJ, Veneklaas EJ. 2006. Root
 structure and functioning for efficient acquisition of phosphorus: Matching
- 540 morphological and physiological traits. *Annals of Botany* **98**: 693-713.
- Li HB, Ma QH, Li HG, Zhang FS, Rengel Z, Shen JB. 2014a. Root morphological
 responses to localized nutrient supply differ among crop species with contrasting

543 root traits. *Plant and Soil* **376**: 151-163.

- Li HG, Shen JB, Zhang FS, Tang CX, Lambers H. 2008. Is there a critical level of
 shoot phosphorus concentration for cluster-root formation in *Lupinus albus*? *Functional Plant Biology* 35: 328-336.
- Li L, Zhang FS, Li XL, Christie P, Yang SC, Tang CX. 2003. Interspecific
 facilitation of nutrient uptakes by intercropped maize and faba bean. *Nutrient Cycling in Agroecosystems* 65: 61-71.
- Li L, Sun JH, Zhang FS, Guo TW, Bao XG, Smith FA, Smith SE. 2006. Root
 distribution and interactions between intercropped species. *Oecologia* 147:
 280-290.
- Li L, Li SM, Sun JH, Zhou LL, Bao XG, Zhang HG, Zhang FS. 2007. Diversity
 enhances agricultural productivity via rhizosphere phosphorus facilitation on
 phosphorus-deficient soils. *Proceedings of the National Academy of Sciences*,
 USA 104: 11192-11196.
- Li L, Tilman D, Lambers H, Zhang FS. 2014b. Biodiversity and overyielding:
 Insights from below-ground facilitation of intercropping in agriculture. *New Phytologist* 203: 63-69.
- Markham JH, Chanway CP. 1996. Measuring plant neighbour effects. *Functional Ecology* 10: 548-549.
- Mommer L, Van Ruijven J, Jansen C, Van de steeg HM, de Kroon H. 2012.
 Interactive effects of nutrient heterogeneity and competition: implications for
- root foraging theory? *Functional Ecology* **26**: 66-73.
- **Robinson D. 1996.** Resource capture by localized root proliferation: Why do plants
 bother? *Annals of Botany* **77:** 179-185.
- **Robinson D, Hodge A, Griffiths BS, Fitter AH. 1999.** Plant root proliferation in
 nitrogen-rich patches confers competitive advantage. *Proceedings of the Royal Society of London* 266: 431-435.
- 570 Schenk HJ. 2006. Root competition: beyond resource depletion. *Journal of Ecology*571 94: 725-739.
- 572 Shen JB, Rengel Z, Tang CX, Zhang FS. 2003. Role of phosphorus nutrition in 20

development of cluster roots and release of carboxylates in soil-grown *Lupinus albus*. *Plant and Soil* 248: 199-206.

- Shen JB, Li HG, Neumann G, Zhang FS. 2005. Nutrient uptake, cluster root
 formation and exudation of protons and citrate in *Lupinus albus* as affected by
 localized supply of phosphorus in a split-root system. *Plant Science* 168:
 837-845.
- Shen JB, Yuan LX, Zhang JL, Li HG, Bai ZH, Chen XP, Zhang WF, Zhang FS.
 2011. Phosphorus dynamics: from soil to plant. *Plant Physiology* 156: 997-1005.
- Shen JB, Li CJ, Mi GH, Li L, Yuan LX, Jiang RF, Zhang FS. 2013. Maximizing
 root/rhizosphere efficiency to improve crop productivity and nutrient use
 efficiency in intensive agriculture of China. *Journal of Experimental Botany*64:1181-1192.
- Tilman D. 1982. Resource competition and community structure. Monographs in
 population biology. Princeton, New Jersey, USA: Princeton University Press.
- Valladares F, Sanchez D, Zavala MA. 2006. Quantitative estimation of phenotypic
 plasticity: bridging the gap between the evolutionary concept and its ecological
 applications. *Journal of Ecology* 94: 1103-1116.
- 590 Wang BL, Tang XY, Cheng LY, Zhang AZ, Zhang WH, Zhang FS, Liu JQ, Cao Y,
- Allan DL, Vance CP, Shen JB. 2010. Nitric oxide is involved in phosphorus
 deficiency-induced cluster-root development and citrate exudation in white lupin. *New Phytologist* 187: 1112-1123.
- Weigelt A, Jolliffe P. 2003. Indices of plant competition. Journal of Ecology 91:
 707-720.
- Wilson SD, Keddy PA. 1986. Measuring diffuse competition along an environmental
 gradient: results from a shoreline plant community. *The American Naturalist* 127:
 862-869.
- Zhang FS, Shen JB, Zhang JL, Zuo YM, Li L, Chen XP. 2010. Rhizosphere
 processes and management for improving nutrient use efficiency and crop
 productivity: Implications for China. In Donald L. Sparks, editor. *Advances in Agronomy*. Burlington: Academic Press, 107: 1-32.

603 Figure legends

604 **Fig. 1** A schematic diagram of rhizo-box.

Fig. 2 Experiment 1: effects of neighbour on shoot biomass (a) and shoot P content (b) of target maize grown at five levels of P supply. The error bars indicate \pm standard errors. Within a P supply, different letters denote significant differences among target maize data in the single maize, maize/maize and maize/faba bean treatments ($P \le 0.05$).

Fig. 3 Experiment 1: relative interaction intensity (RII) of target maize with its neighbours in the maize/maize and maize/faba bean mixtures. The error bars indicate \pm standard errors. Within a P supply, stars denote significant differences between the maize/maize and maize/faba bean mixtures, ***P*<0.01, ns = not significant.

Fig. 4 Experiment 1: effects of neighbour on root length of target maize grown at five levels of P supply. The error bars indicate \pm standard errors. Within a P supply, stars denote significant differences between the maize/maize and maize/faba bean mixtures, * $P \le 0.05$, **P < 0.01, ns = not significant.

Fig. 5 Experiment 1: root length (a), and citrate concentration (b) and acid phosphatase activity (c) in the rhizosphere of neighbours (maize and faba bean). The error bars indicate \pm standard errors. Within a P supply, stars denote significant differences between neighbouring species (maize *vs* faba bean), **P*≤0.05, ***P*<0.01, ns = not significant.

Fig. 6 Experiment 2: effects of homogeneous vs heterogeneous P supply on shoot biomass (a) and shoot P content (b) of target maize. Hom: homogeneous P supply; Het: heterogeneous P supply. The error bars indicate standard errors. Different lower case letters denote significant differences among target maize ($P \le 0.05$) data in the single maize, maize/maize and maize/faba bean treatments in the homogeneous treatments, and capitals denote significant differences in the heterogeneous treatments. For each parameter and species treatment, t-tests were run separately to detect difference between homogeneous and heterogeneous treatments; stars indicate significant differences, * $P \le 0.05$.

631 Fig. 7 Experiment 2: effects of homogeneous/heterogeneous P supply on target maize root length density within (a) and outside P-enriched soil volume (b). Hom: 632 homogeneous P supply; Het: heterogeneous P supply. The error bars indicate standard 633 errors. Different lower case letters denote significant differences among target maize 634 (P≤0.05) data in single maize, maize/maize and maize/faba bean mixtures in the 635 636 homogeneous treatments, and capitals denote significant differences in the heterogeneous treatments. For each soil volume and each species treatment, t-tests 637 638 were run separately to detect significant difference between homogeneous and 639 heterogeneous treatments; stars indicate significant differences, * $P \leq 0.05$.