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Abstract

Clinical, epidemiological, and biomechanical studies suggest the involvement of the cervical facet joint in neck pain. Mechanical studies have suggested the facet capsular ligament to be at risk for subfailure tensile injury during whiplash kinematics of the neck. Ligament mechanical properties can be altered by subfailure injury and such loading can induce cellular damage. However, at present, there is no clear understanding of the physiologic context of subfailure facet capsular ligament injury and mechanical implications for whiplashrelated pain. Therefore, this study aimed to define a relationship between mechanical properties at failure and a subfailure condition associated with pain for tension in the rat cervical facet capsular ligament. Tensile failure studies of the C6/C7 rat cervical facet capsular ligament were performed using a customized vertebral distraction device. Force and displacement at failure were measured and stiffness and energy to failure were calculated. Vertebral motions and ligament deformations were tracked and maximum principal strains and their directions were calculated. Mean tensile force at failure (2.96±0.69 N) was significantly greater (p<0.005) than force at subfailure (1.17±0.48 N). Mean ligament stiffness to failure was 0.75±0.27 N/mm. Maximum principal strain at failure $(41.3\pm20.0\%)$ was significantly higher (p=0.003) than the corresponding subfailure value (23.1±9.3%). This study determined that failure and a subfailure painful condition were significantly different in ligament mechanics and findings provide preliminary insight into the relationship between mechanics and pain physiology for this ligament. Together with existing studies, these findings offer additional considerations for defining mechanical thresholds for painful injuries.

Keywords

facet joint, ligament, failure, strain, biomechanics

Comments

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Tensile Cervical Facet Capsule Ligament Mechanics:

Failure and Subfailure Responses in the Rat

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ABSTRACT

Clinical, epidemiological, and biomechanical studies suggest the involvement of the cervical facet joint in neck pain. Mechanical studies have suggested the facet capsular ligament to be at risk for subfailure tensile injury during whiplash kinematics of the neck. Ligament mechanical properties can be altered by subfailure injury and such loading can induce cellular damage. However, at present, there is no clear understanding of the physiologic context of subfailure facet capsular ligament injury and mechanical implications for whiplash-related pain. Therefore, this study aimed to define a relationship between mechanical properties at failure and a subfailure condition associated with pain for tension in the rat cervical facet capsular ligament. Tensile failure studies of the C6/C7 rat cervical facet capsular ligament were performed using a customized vertebral distraction device. Force and displacement at failure were measured and stiffness and energy to failure were calculated. Vertebral motions and ligament deformations were tracked and maximum principal strains and their directions were calculated. Mean tensile force at failure (2.96±0.69 N) was significantly greater (p<0.005) than force at subfailure $(1.17\pm0.48 \text{ N})$. Mean ligament stiffness to failure was 0.75 ± 0.27 N/mm. Maximum principal strain at failure (41.3±20.0%) was significantly higher (p=0.003) than the corresponding subfailure value (23.1±9.3%). This study determined that failure and a subfailure painful condition were significantly different in ligament mechanics and findings provide preliminary insight into the relationship between mechanics and pain physiology for this ligament. Together with existing studies, these findings offer additional considerations for defining mechanical thresholds for painful injuries.

1 INTRODUCTION

2 The cervical facet joint has been identified as a source of neck pain (Aprill and 3 Bogduk, 1992; Barnsley et al. 1994) and a likely candidate for painful whiplash injury, in 4 both clinical and biomechanical studies (Bogduk and Marsland, 1988; Kaneoka et al. 5 1999; Luan et al. 2000; Ono et al. 1997; Panjabi et al. 1998a,b; Pearson et al. 2004; 6 Siegmund et al. 2001; Winkelstein et al. 1999, 2000; Yoganandan and Pintar, 1997; 7 Yoganandan et al. 1998a, 2002). Studies of cadaveric head-neck preparations using high-8 speed imaging have demonstrated that the facet joint and its capsular ligament can 9 experience excessive motions and ligament strains during whiplash simulations (Panjabi 10 et al. 1998a,b; Pearson et al. 2004; Sundararajan et al. 2004; Yoganandan et al. 2001, 11 2002). Studies of isolated cervical spinal motion segments have also documented that 12 these cervical spine kinematics can induce facet capsule stretch and possible minor 13 ligament ruptures below the mechanical thresholds for gross failure of the ligament 14 (Siegmund et al. 2001; Winkelstein et al. 2000). However, while these mechanical 15 studies suggest that the facet capsular ligament may be at mechanical risk for painful 16 injury during some neck motions, the physiologic consequence of these injuries and their 17 relationship to the tensile mechanical response of the joint is undefined.

Tensile failure properties of the human cervical facet capsular ligament have been previously defined (Mykelbust et al. 1988; Winkelstein et al. 1999, 2000; Yoganandan et al. 2000). Mykelbust et al. (1988) reported failure forces of 112±30 N and 72±18 N for tensile loading of isolated C3/C4 and C5/C6 ligaments, respectively. Likewise, Winkelstein et al. (2000) reported ligament failure at similar forces of 94.3±44.4 N and 82.5±33.0 N for these same joints. In that study, maximum ligament distraction at failure

24 was 5.10±0.73 mm and 6.40±0.66 mm, respectively, producing maximum principal 25 strains of 103.6±80.9% in the facet capsule (Winkelstein et al. 2000). In tensile failure 26 tests of lower cervical spine specimens (C5-T1), Yoganandan et al. (2000) reported 27 stresses of 7.4±1.3 MPa and strains of 116.0±19.6%, with corresponding stiffness and 28 energy to failure of 36.9±6.06 N/mm and 1.5±0.4 Nm, respectively. While these studies 29 present consistent data on the mechanical limits of the human cervical facet capsular 30 ligament, they are unable to provide a context for investigating the effects of these 31 mechanics on physiologic function or implications for mechanical responses at subfailure 32 conditions.

33 Capsule injury prior to gross ligamentous failure has been documented in both 34 isolated and full cervical spine specimens (Panjabi et al. 1998a; Siegmund et al. 2001; 35 Winkelstein et al. 2000; Yoganandan et al. 2001). In cervical motion segment studies, 36 subfailure minor ruptures were produced in the facet capsule, at strains ranging from 37 35.0-64.6%, for both shear and tension (Siegmund et al. 2001; Winkelstein et al. 2000). 38 Panjabi et al. (1998b) and Pearson et al. (2004) estimated C6/C7 ligament strains during 39 whiplash simulations to be $29.5\pm25.7\%$ and $39.9\pm26.3\%$, respectively. However, these 40 same specimens sustained ligament strains of only 6.2±5.6% for spinal motions within 41 normal physiologic ranges (Panjabi et al. 1998b), leading these authors to suggest that 42 whiplash injury induces ligament strains that are elevated above physiologic levels and 43 that these elevated strains may cause ligament injury, despite lack of any evidence of 44 rupture or noticeable injury in these specimens. All of these studies have hypothesized 45 subfailure injuries at the microscopic level as a potential means of nociceptor activation. 46 Nociceptive pain fibers have been identified throughout the facet joint and its capsular

ligament (Cavanaugh et al. 1989, 1996; McLain 1994; Inami et al. 2001). 47 48 Electrophysiologic studies have further shown that these fibers in the facet ligament can 49 be directly activated by tensile loading in the lumbar spine (Avramov et al. 1992; 50 Cavanaugh et al. 1989, 1996). Together, these mechanical and anatomic findings 51 suggest whiplash kinematics in the cervical spine may induce a subfailure mechanical 52 condition in the facet capsular ligament that has the potential to initiate physiologic 53 responses for pain. However, the relationship between the magnitude of relevant 54 subfailure and failure injury mechanics in the context of physiologic outcomes remains 55 unknown.

56 Our laboratory has developed an *in vivo* model of facet joint distraction in the rat 57 for investigating the physiologic sequelae of pain produced from this loading (Lee et al. 58 2004a,b). That model enables repeatable and controlled distraction across the facet joint 59 and its capsular ligament. Vertebral distractions of 0.57 ± 0.11 mm in that model are 60 adequate for producing repeatable behavioral sensitivity and pain symptoms (as measured 61 by mechanical allodynia) (Lee et al. 2004a). Such joint distractions produce strains in the 62 C6/C7 facet capsule of 27.7±11.9% (Lee et al. 2004a). While no observable rupture of 63 the ligament is produced at these magnitudes of vertebral distraction, information about 64 the relative mechanical severity of this subfailure condition compared to failure remains 65 to be determined.

66 Therefore, the goal of the present study is to characterize the failure and 67 subfailure mechanical properties of the rat cervical facet capsular ligament in tension. 68 The subfailure condition selected here is defined as 0.57 mm of vertebral distraction, 69 based on previous *in vivo* studies in which this distraction predictably produced

behavioral sensitivities sustained over time as persistent pain (Lee et al. 2004a). It is hypothesized that for this ligament, the corresponding subfailure tensile condition that produces pain in the *in vivo* model is mechanically distinct from ligament failure, despite being sufficient to cause physiologic manifestation of pain. Characterization of the mechanical properties of this ligament and the relationship between its failure and subfailure loading will provide context for understanding both the mechanical and physiologic responses of injuries to this joint as well as other ligaments.

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78 METHODS

79 Specimen Preparation & Loading Procedure

80 Male Holtzman rats (n=11), weighing 325-425 g, were used in this study. All 81 experimental procedures were approved by the University of Pennsylvania Institutional 82 Animal Care and Use Committee and carried out according to the guidelines of the 83 Committee for Research and Ethical Issues of the International Association for the Study 84 of Pain (Zimmermann 1983). Rats were euthanized by CO₂ inhalation and cervical spinal 85 motion segments from C4-T2 were immediately removed en bloc. Specimens were 86 cleared of all musculature. The laminae, facet joints, and spinous processes at C6/C7 87 were exposed bilaterally under a surgical microscope (Carl Zeiss Inc., Thornwood, NY). 88 Tensile loading was performed using a customized device; distraction methods were 89 identical to previous *in vivo* investigations (Lee et al. 2004a,b). Briefly, the supraspinous 90 ligament, interspinous ligament, and ligamentum flavum were bilaterally resected at 91 C6/C7 to enable specimen attachment, fixation, and loading during testing. In addition, 92 for this study, the left capsular ligament, both longitudinal ligaments, and the

93 intervertebral disc at C6/C7 were transected and removed, to enable isolation of the right 94 facet capsular ligament only. The C6 and C7 spinous processes were rigidly attached to 95 the distraction device by microforceps (Figure 1); C7 was held fixed and C6 was 96 translated rostrally using a manual micrometer (Newport Corp., Irvine, CA). A linear variable differential transducer (LVDT) (MicroStrain Inc., Burlington, VT; 0.160 µm 97 98 resolution) and load cell (Interface Inc., Scottsdale, AZ; 0.02 N resolution) were rigidly 99 coupled to the C6 microforceps and their synchronized data were acquired at 10 Hz. 100 LVDT displacement histories were used to calculate distraction rates. Displacements 101 were applied until gross ligament failure was observed both visually and by a decrease in 102 tensile force.

103 The right facet joint and capsular ligament were imaged during distraction at 6 fps 104 using a digital video camera (QImaging, B.C. Canada), with 1280 x 1024 pixel 105 resolution. Image data were synchronized with the transducer data and acquired using 106 LabVIEW (National Instruments, Corp., Austin, TX). Polystyrene magnetic particles (diameter of 0.17±0.01 mm; Spherotech, Inc., Libertyville, IL) were affixed to the bones 107 108 and ligament to track joint motions. Particles were placed on each of the C6 and C7 109 laminae, as vertebral markers, to track bony motions across the joint (Figure 2). To 110 calculate ligament strains, nine additional particles, serving as ligament markers, were 111 placed on the posterior surface of the C6/C7 capsular ligament in a 3 x 3 grid, creating 4 112 elements (Figure 2). Ligament regions were defined by quadrants of the grid: Quadrant I 113 (QI) as caudal-medial; Quadrant II (QII) as rostral-medial; Quadrant III (QIII) as caudal-114 lateral; Quadrant IV (QIV) as rostral-lateral.

116 Data Analysis

Force at failure was measured as the maximum force at ligament failure. Based on the force-displacement response, ligament failure was defined as a drop in force with increased displacement. For each specimen, visual inspection of the ligament confirmed the existence and site of failure (Figure 3). Stiffness to failure was calculated as the slope of the force-displacement curve from 20-100% of the peak force, which represented the most linear portion of the curve for all specimens (Figure 4). Energy to failure was calculated as the area under the force-displacement curve up to the point of failure.

124 Image tracking software (Image Pro Plus; Media Cybernetics Inc, Silver Spring, MD) 125 located all vertebral and ligament marker centroids for each frame up to and including 126 failure. Vertebral distraction was defined as the linear displacement, in the rostral (x)127 direction, of the C6 vertebral marker relative to the C7 vertebral marker. This procedure 128 has previously been shown to produce distraction primarily in the rostral-caudal direction 129 (x-axis, Figure 2), with negligible motion in the medial-lateral direction (y-axis, Figure 2) 130 (Lee et al. 2004a,b). Initial positions of the ligament markers, prior to loading, were used 131 to construct a finite element mesh of 4 shell elements in LS-DYNA (LSTC, Livermore, 132 CA). The mesh divided the ligament into four quadrants (Quadrants I-IV) (Figure 2), 133 which were used to describe the location of maximum principal strain and failure. The 134 positional coordinates of ligament markers during distraction sequences were used to 135 calculate the corresponding displacement fields and Lagrangian strains within the plane 136 of the elements. For each specimen, maximum principal strain and maximum shear 137 strain in the ligament were calculated.

Matched force and strain data were also examined for each specimen at a joint distraction corresponding to the subfailure value at vertebral distraction of 0.57 mm (Figure 3). All mechanical data at this vertebral distraction were analyzed in the same manner as the failure data for comparison. One specimen (#77) did not reach this vertebral distraction prior to its ligament rupture. As such, subfailure data from this specimen were not available for analysis.

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145 Statistical Analysis

Vertebral distraction, tensile force, maximum principal strains and directions, and shear strains in the ligament were tested for normality using the Shapiro-Wilk test and compared between failure and subfailure conditions using a paired Student's t-test (Zar, 149 1999). Because Specimen #77 did not reach 0.57 mm of distraction prior to failure, its data were excluded for all statistical comparisons between failure and subfailure. All statistical analyses were performed using SYSTAT (SYSTAT Software Inc., Richmond, CA) and significance was defined as p<0.05.

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154 RESULTS

There was no significant departure from normality for any of the mechanical parameters (vertebral distraction, tensile force, maximum principal strain and direction, maximum shear strain) for either condition (failure, subfailure). For these studies, the average rate of distraction was 0.08±0.02 mm/s. The mean tensile force at failure was 2.96±0.69 N, corresponding to a vertebral distraction of 1.52±0.76 mm (Table 1). Digitization errors were 0.006±0.001 mm, only 0.4% of the imposed vertebral distractions. As such, errors did not contribute substantially to calculated distraction. Linear regression fits to force-displacement data had a mean R^2 value of 0.96±0.03. The mean C6/C7 facet capsular ligament stiffness to failure was 0.75±0.27 N/mm and mean energy to failure was 0.008±0.005 Nm (Figure 4, Table 1). For all specimens, force demonstrated a steady increase with increasing displacement until failure was reached.

166 At ligament failure, the mean maximum principal strain in the capsule was 167 41.3±20.0% (Figure 5, Table 2). The maximum principal strain was located in each of 168 the quadrants QI, QII, QIII and QIV (Table 2). The mean maximum shear strain in the 169 capsule was $23.1\pm10.9\%$, and was significantly smaller than the corresponding maximum 170 principal strain (p=0.003). Errors in determining strains were small, and had an average 171 value of $1.3\pm1.2\%$. Visual inspection of the ligaments confirmed that, for all cases, the 172 site of gross failure was located in the same quadrant (anatomic region of ligament) as the 173 site of maximum principal strain. Directions associated with the maximum principal 174 strains were oriented at a mean angle of $3.1\pm41.2^{\circ}$ off the x-axis (Figure 5B, Table 2). In 175 7 of the 11 specimens, ligament failures occurred as a small tear forming in the ligament 176 midsubstance immediately prior to failure and progressing until failure.

Subfailure vertebral distraction of 0.57 ± 0.01 mm produced 1.17 ± 0.48 N of mean tensile load. The magnitude of subfailure distraction and force were significantly lower than their corresponding values at failure (p<0.005, both cases). Mean maximum principal strain at this condition was $23.1\pm9.3\%$, which was also significantly smaller (p=0.003) than maximum strain in the ligament at failure (Figure 5). Vectors describing directions of maximum principal strain at subfailure were oriented at $28.5\pm16.9^{\circ}$ relative to the x-axis (Figure 5D, Table 2), and were not significantly different from those at failure (p=0.06). The mean maximum shear strain at subfailure was $6.9\pm7.9\%$, which was significantly smaller than the corresponding maximum principal strain at subfailure and the maximum shear strain at failure (p<0.005, both cases). For all specimens except Specimen #77, no gross ligament damage was visible at subfailure. While Specimen #77 failed at a vertebral distraction of 0.43 mm (Table 1), it ruptured in a similar manner as the majority of the other specimens, with a small tear in the midsubstance.

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191 DISCUSSION

192 While tensile failure properties of human cervical facet capsular ligaments have 193 been previously reported (Mykelbust et al. 1988; Winkelstein et al. 1999, 2000; 194 Yoganandan et al. 2000), this study is the first to report failure or subfailure properties for 195 this ligament in the rodent. Mechanical studies have shown increased joint laxity, 196 decreased ligament stiffness, and an overall change in the force-displacement response of 197 various ligaments after subfailure loading in human and rabbit tissue (Panjabi et al. 1996, 198 2001; Pollock et al. 2000). These reports suggest that subfailure loading may produce 199 microscopic ligament damage, in turn affecting the subsequent mechanical properties of 200 the ligament. Histologic study of the rat medial collateral ligament after subfailure injury 201 revealed necrosis associated with strains significantly below those necessary to induce 202 structural damage (Provenzano et al. 2002), implying that cellular damage can be induced 203 without corresponding observable ligament rupture. These mechanical and histologic 204 reports suggest that the subfailure condition applied in this study may lead to microscopic 205 damage, potentially altering the mechanical function of this ligament and may be 206 sufficient to produce pain symptoms. Tensile failure studies of cadaveric facet capsular

207 ligaments have reported a distinct drop in the force-displacement curve prior to frank 208 ligament rupture (Siegmund et al. 2001; Winkelstein et al. 2000); strains sustained at 209 these minor failures were 62% of failure values (Winkelstein et al. 2000). The current 210 study reports maximum principal strains at subfailure that are 56% of the corresponding 211 failure values for rupture of the ligament. Our study also found the subfailure tensile 212 force to be 40% of the failure force, which agrees with the corresponding force at initial 213 failure in the human cadaveric ligament which was found to be 47% of the peak force for 214 combined bending and shear (Siegmund et al. 2001). While mechanical scaling 215 relationships between the rat and the human remain undefined and may present an 216 experimental challenge, present findings demonstrate similar relative relationships exist 217 for this ligament's mechanical properties at failure and subfailure in both the rat and the 218 human. This adds further relevance to the *in vivo* findings related to the mechanical 219 injuries producing pain. Continued efforts are needed to develop an appropriate scaling 220 factor between the rat and the human for comparing mechanical data in these and other 221 species.

222 In our study, maximum principal strains in the ligament were generally directed 223 across the joint line, parallel to the direction of applied distraction, along the spine's long 224 axis. This suggests that the ligamentous fibers may be oriented across the joint line, 225 although additional mechanical and histologic studies are necessary to fully quantify the 226 orientation of ligament fibers and the precise mode of this ligament's failure. Previous 227 work has demonstrated tensile failure occurring primarily in the midsubstance of human 228 cadaveric facet capsular ligaments (Winkelstein et al. 2000). Likewise, the current study 229 reports ligament failures as small tears in the midsubstance, primarily occurring in the

medial portion of the capsule (QI and QII). While ligament deformation and strain are 230 231 not dependent on loading rate, peak load and stiffness at failure do depend on rate and are 232 higher for fast-rate loading (Winkelstein et al. 1999; Yoganandan et al. 1998b). As such, 233 the strain data reported here for quasistatic loading likely reflect those maximum 234 principal strains induced in the ligament for other loading rates. However, the location 235 of rupture may be altered for other loading rates. The distraction method used in this 236 study is purely tensile and does not fully model the coupled motions of all physiologic 237 modes of loading. Indeed, similar approaches for strain measurement have been 238 implemented recently for the human lumbar facet capsule and report both a dependence 239 on loading direction and subfailure mechanical responses for this ligament (Ianuzzi et al. 240 2004; Little and Khalsa, 2005). Accordingly, capsule strains reported in this study may 241 not be representative of those experienced during other physiologic motions.

242 The subfailure vertebral distraction magnitude selected in this study was based on 243 existing in vivo facet distraction-mediated pain models and cadaveric data obtained 244 during whiplash simulations (Lee et al. 2004a, Panjabi et al. 1998b, Pearson et al. 2004). 245 In vivo tensile vertebral distractions matching those applied in the current study have 246 been previously demonstrated to produce ligament strains (27.7±11.9%) (Lee et al. 247 2004b) that are similar to those produced in the C6/C7 ligament (29.5-39.9%) during 248 whiplash simulations (Panjabi et al. 1998b, Pearson et al. 2004). These in vivo vertebral 249 distractions elicited behavioral sensitivity, which remained elevated above physiologic 250 levels for 14 days (Lee et al. 2004a). Moreover, significantly increased astrocytic 251 activation was also observed in the spinal cord of these rats after these subfailure 252 vertebral distractions (Lee et al. 2004a), suggesting sustained cellular reactivity for this

253 loading. The vertebral distractions in the present subfailure study produced ligament 254 strains of 23.1±9.3% (Table 2) that are similar to those produced *in vivo* causing pain. In 255 fact, in directly comparing the strain data from the current study with the previously 256 reported *in vivo* work, a Student's t-test reveals no statistical difference (p=0.42). This 257 suggests that the subfailure ligament distractions used here are sufficient to produce pain 258 symptoms in an *in vivo* condition. It should be noted, however, that while the current 259 study involved unilateral facet capsule distraction, the distraction applied in the *in vivo* 260 study was applied across both the right and left capsules. Demonstration of the presence 261 of nociceptors (Cavanaugh et al. 1989, 1996; Inami et al. 2001; McLain 1994) and their 262 activation (Avramov et al. 1992; Cavanaugh et al. 1996) in the ligament further suggests 263 a role for this joint in pain signaling. Given the histologic and electrophysiologic 264 evidence of this ligament's involvement in nociception, the current study suggests that 265 subfailure tensile loading of the ligament may lead to nociceptive physiologic changes in 266 the spine, despite lack of its mechanical injury.

267 Visual inspection of image data at subfailure revealed no evidence of ligamentous 268 damage. However, it remains unclear whether subfailure loading at this magnitude 269 produces small, or even microscopic tears, which would not be visible to the naked eye. 270 As such, while this study did not histologically examine the ligament for evidence of 271 damage at subfailure, we have previously demonstrated that stiffness is not altered for 272 repeated distraction at these levels (Franklin et al. 2004), suggesting no gross structural 273 damage occurs at these distraction levels. Further mechanical or histologic investigations 274 of the ligament would provide further characterization and interpretation of the 275 mechanical and physiologic meaning of these loading conditions. Of note, while tensile

276 failure of cadaveric ligaments has been reported to produce a noticeable subfailure event 277 in the force-displacement curve during loading (Siegmund et al. 2001; Winkelstein et al. 278 2000), such a distinct event was not detected at any point in the loading responses for the 279 ligaments in this study (Figure 4). This suggests that the mechanical response of the 280 fibers within the rat ligament may be different than that of the matched human ligament. 281 Also, while Specimen #77 failed prior to its reaching the subfailure value, its structural 282 properties at failure were within the range of the values determined for the other 283 specimens (Table 2).

284 The findings presented here offer a foundation for interpreting existing 285 physiologic data obtained during subfailure ligament distraction (Lee et al. 2004a). Data 286 show that for this subfailure distraction condition, ligament forces and strains are 287 significantly smaller than those produced at failure, suggesting no structural damage at 288 these levels and that this subfailure distraction in an *in vivo* setting is not sufficient to 289 produce gross injury. However, in the context of *in vivo* work (Lee et al. 2004a), this 290 subfailure condition *does* produce sustained cellular responses in the spinal cord and pain 291 symptoms, suggesting a threshold for physiologic damage or nociceptive modulation at 292 this level of mechanical loading for this joint. While distractions may not produce 293 detectable alteration in mechanical responses for the rat facet capsular ligament, they may 294 indeed be sufficient to trigger the physiologic sequelae of pain and its symptoms. This 295 study provides context for existing physiologic data obtained at subfailure levels, offers 296 new insight into mechanical thresholds of facet joint injury, and lays the groundwork for 297 further investigation into the physiologic implications of this and other subfailure 298 conditions in ligamentous loading.

299

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Specimen	Weight (g)	Vertebral Distraction (mm)	Tensile Load (N)	Tensile Stiffness (N/mm)	Energy at Failure (Nm)	
52	326	1.88	3.56	0.99	0.006	
75	404	1.03	2.51	0.68	0.005	
76	404	3.40	3.92	0.51	0.016	
77	346	0.43	2.47	1.05	0.003	
78	392	0.92	3.75	1.21	0.006	
79	418	1.09	2.52	0.77	0.005	
112	398	1.56	2.86	0.42	0.008	
114	392	1.37	3.81	0.63	0.019	
115	410	1.72	2.98	1.00	0.006	
116	396	1.47	2.17	0.55	0.007	
117	368	1.89	2.04	0.43	0.010	
Average (SD)	391 (26)	1.52 (0.76)	2.96 (0.69)	0.75 (0.27)	0.008 (0.005)	

Table 1. Summary of failure properties of isolated rodent facet capsules.

		FAILURE	SUBFAILURE				
Specimen	Maximum Principal Strain (%)	Maximum Principal Strain Direction (relative to x-axis) (°)	Maximum Shear Strain (%)	Location of Maximum Principal Strain (Quadrant)	Maximum Principal Strain (%)	Maximum Principal Strain Direction (relative to x-axis) (°)	Maximum Shear Strain (%)
52	47.8	45.0	33.2	Ι	21.9	25.7	11.0
75	38.1	9.8	15.4	Ι	37.2	57.9	15.5
76	64.4	-36.2	44.6	IV	15.8	10.1	3.5
77	19.9	23.5	11.2	II	*	*	*
78	27.2	26.6	20.9	IV	17.3	25.0	14.4
79	55.1	30.3	26.2	II	34.9	20.1	15.0
112	32.6	-89.4	5.1	IV	15.3	54.3	-9.8
114	36.0	43.4	25.9	III	30.2	29.1	6.2
115	83.9	12.7	22.2	Ι	30.8	3.8	5.3
116	18.4	6.5	18.7	II	11.2	30.4	-0.6
117	31.0	-37.5	31.1	II	16.7	28.8	8.5
Average (SD)	41.3 (20.0)	3.14 (41.2)	23.1 (10.9)	-	23.1 (9.3)	28.5 (16.9)	6.9 (7.9)

Table 2. Summary of capsule strain data.

* Subfailure data not available for Specimen #77, as it failed prior to reaching subfailure distraction.

Figure Legends

Figure 1. Facet distraction device, with microforceps, micrometer, LVDT, and load cell. A surgical microscope is mounted above the setup to acquire image data. For distraction, the C7 microforceps are held rigidly in place while the C6 microforceps are translated rostrally, using the micrometer.

Figure 2. Schematic illustrating the posterior view of the C6/C7 facet joint and its capsule (**A**). Two sets of markers are used: vertebral markers (large circles) are placed on the C6 and C7 laminae to track bony motions and nine ligament markers (small circles) are placed on the facet capsular ligament and define a grid for finite element analysis. The grid contains four quadrants, labeled I-IV. A representative image is also shown for a typical ligament exposure (Specimen #52), demonstrating the relevant markers and anatomy (**B**). X- and y- directions are shown in (**A**) for reference.

Figure 3. A series of *ex vivo* images obtained during tensile failure. In the reference condition (**A**), the vertebral separation is represented by *x*. Vertebral distraction is calculated using the x-coordinate of each vertebral marker. Subfailure vertebral distraction (**B**) is calculated as $(x_{sf} - x)$ and vertebral distraction at failure (**C**) was calculated as $(x_{f} - x)$. This test specimen (#52) sustained vertebral distractions of 0.57 mm and 1.88 mm for subfailure and failure, respectively; maximum principal strains in the capsule were 17.3% (subfailure) and 27.2% (failure). The location of failure occurred in Quadrant I (**C**) and is indicated by an arrow.

Figure 4. A representative force-displacement curve (Specimen #78) indicating ligament failure. The linear regression fit (R^2 =0.98) to the data between 20-100% of failure load estimates the capsule stiffness to failure. Failure occurred at 3.75 N, with a stiffness of 1.21 N/mm.

Figure 5. Representative maximum principal strains in the capsular ligament (Specimen #78) at failure (maximum 27.2%) (**A**) and subfailure (maximum 17.3%) (**C**). Also shown are the corresponding direction vectors of these strains for failure (**B**) and subfailure (**D**), indicating the primary direction across the joint. Quadrants I-IV are shown in (**A**) for reference. For this specimen, failure occurred as a midsubstance tear in QIV, corresponding to the location of maximum strain in (**A**).















Figure 5



(B)



(D)





