SPECTROGRAPHIC TECHNIQUES AND ANALYSES OF PINE NEEDLES

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The spectrograph can be used to determine many elements in plants, although all elements cannot be determined equally well because of volatility, excitation, or ionization differences. Some 12 to 20 elements are rather easily determined in the open arc in plant ash in the procedure used here. They are boron, phosphorus, magnesium, lead, tin, silicon, iron, aluminum, molybdenum, calcium, vanadium, copper, silver, zinc, sodium, titanium, nickel, cobalt, wolfram, manganese, potassium, and chromium. Successful analyses can be made on very small samples on a spectrograph. Most samples are 100 milligrams in weight but smaller ones, down to 10 milligrams, can be used if necessary. This small sample size is an advantage when studying the margins and tips of leaves, especially the tips of pine needles.

In brief the spectrographic procedure consists of burning the samples in an open arc and recording the spectrum from this arc which contains the unknowns on a photographic plate. The spectrograms when properly developed and fixed constitute a permanent record of the elements in the samples. The light or radiation from the arc is dispersed through a quartz prism and each element shows up on the plate as a line of specific wave length or a group of lines characteristic of the element. The photographic emulsion also integrates the light energy from the elements and

THE OHIO JOURNAL OF SCIENCE 57(6): 345, November, 1957.

forms a dense line when a quantity of the elements is present and a very faint line when only a small amount is present. This is the basis of the quantitative nature of the procedure.

The spectra of the elements on the plates are fine lines and must be examined and measured under a magnification of about 10 to 20 diameters for best results. Hence, a microdensitometer is a necessary part of the spectrographic laboratory.

METHODS

Location of lines.—The recognition, location, and determination of the origin of spectral lines could be a long job. But it is possible to locate and recognize those lines of the elements used in quantitative analysis in a few days time. The following is a list of those lines most commonly used to determine the given elements.

Element	Angstrom	Element	Angstrom
Boron	2497.7	Copper	3274.0
Phosphorus	2535.7	Silver	3280.0
Magnesium	2779.9	Zinc	3282.3
Lead	2833.1	Sodium	3302.3
Tin	2840.0	Titanium	3349.0
Silicon	2881.6	Nickel	3414.8
Iron	3020.6	Cobalt	3453.5
Aluminum	3092.7	Wolfram	4008.8
Molybdenum	3170.3	Manganese	4030.8
Calcium	3179.3	Potassium	4044.1
Vanadium	3184.0	Chromium	4254.3

The spectral lines are located (1) by means of an angstrom scale on the spectrograph, (2) by means of a reference spectrum, usually iron on high dispersion instruments or mercury on quartz prism spectrographs, or (3) by the characteristic pattern of lines for each element. Unconsciously all three methods may be used even though the angstrom scale or a reference spectra may not be consulted, or the pattern found. For with very little practice, the cyanogen bands or other known lines or groups of lines serve as a reference point and in a short time a dozen elements can be quickly recognized and measured.

Spectrographic arc.—The theory and construction of spectrographic sources is an involved and complicated study, but the practical use of such an arc is as simple as turning on and off an electric light. Flat top purified graphite electrodes are used to support the samples, and the arc or spark is maintained between them in the proper position in front of the spectrograph.

Plate calibration.—The photographic emulsion is not an ideal integrator of light energy. It may vary with the wavelength of light and not show a true linear response with intensity. It must be calibrated to show the amount of light necessary to produce a given blackening of the emulsion. This is an involved process and requires step sector discs, constant light source, special graph paper (Seidel paper), and controlled development of the plate. The end result is a table showing the transmission of the spectral lines and the intensity of light necessary to produce them. So all one has to do is read a table.

Working curves.—Working curves are constructed by plotting intensities from known amounts of the elements being studied. They are similar to working curves of substances determined in colorimetric analysis. They are made from synthetic mixtures of salts similar in composition to plant ash, with added known amounts of the required elements to be studied. Some are very easy to construct, but others, where enhancement or suppression of lines results from the presence of certain other elements or ions in the mixture, are very difficult to obtain.

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	Sat	mple Dat	а			Spe		Boron			
Sample No.	Year growth	Needle tips cm	Dry wt. gm	Per cent ash	B T	oron 2798 G	Lith 27 T	ium 41 G	Ratio B/Li	μg in 10 mg ash	μg in 100 mg tissue
117	1953	0-1	.8	2.23	$\begin{array}{c} 17 \\ 24 \end{array}$	$\frac{188}{129}$	$\begin{array}{c} 12.5\\ 13 \end{array}$	$\begin{array}{c} 258 \\ 248 \end{array}$.73 .52	29 18	$\begin{array}{c} 6.5 \\ 4.0 \end{array}$
118	1953	1-2	.7	1.52	$\begin{array}{c} 30 \\ 28 \end{array}$	$\begin{array}{c} 100 \\ 108 \end{array}$	$\substack{10\\9.2}$	$\begin{array}{c} 327\\ 356 \end{array}$.31 $.30$	$\begin{array}{c} 10.4 \\ 10 \end{array}$	$\begin{array}{c} 1.58 \\ 1.52 \end{array}$
119	1953	2-	10.4	1.66	$\begin{array}{c} 39 \\ 40 \end{array}$	$\begin{array}{c} 72.2 \\ 70.0 \end{array}$	$9\\9.4$	$\begin{array}{c} 365\\ 348 \end{array}$.20 .20	$\begin{array}{c} 6.5 \\ 6.5 \end{array}$	$\begin{array}{c} 1.08 \\ 1.08 \end{array}$
120	1952	0-1	.9	3.02	$\begin{array}{c} 12 \\ 16 \end{array}$	$\begin{array}{c} 269 \\ 200 \end{array}$	$\begin{array}{c} 9.8\\10\end{array}$	333 327	.81 .61	$\begin{array}{c} 34 \\ 22 \end{array}$	$\begin{array}{c} 10.3\\ 6.7\end{array}$
121	1952	1–2	1.0	2.03	$\begin{array}{c} 41 \\ 35 \end{array}$	$\begin{array}{c} 67.9\\ 83.0\end{array}$	$\begin{array}{c} 10 \\ 8.5 \end{array}$	$\begin{array}{c} 327\\ 393 \end{array}$	$\begin{array}{c} .21\\ .21\end{array}$	$\begin{array}{c} 6.7 \\ 6.7 \end{array}$	$\begin{array}{c} 1.36 \\ 1.36 \end{array}$
122	1952	2–	13.9	1.92	$\begin{array}{c} 56 \\ 53 \end{array}$	$\begin{array}{c} 42.0\\ 45.8\end{array}$	$\begin{array}{c} 10\\9 \end{array}$	$\frac{327}{365}$.13 .13	$\begin{array}{c} 3.4\\ 3.4\end{array}$	$.65 \\ .65$

Boron content of red pine needle samples, from E. L. Stone, Professor Forest Soils, Cornell University. Plot B-12, 22 year old, sampled October 22, 1953 from mid node position, height 22 feet

No. = Original sample number, from E. L. Stone.
Cm = Part of needle in each sample, from tip down.
T = Transmission in percent.
G. = Gamma value or relative intensity.
Spectrographic Plate No. 26, Book No. 11, by David Chou.

TABLE	2
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* No.	Р 2536 G	Mg 2802 G	Pb 2833 G	Si 2882 G	Fe 3020 G	A1 3092 G	Ca 3179 G	Cu 3274 G	Ag 3280 G	Mn 4035 G	K 4044 G	Zn 4810 G	Li 2741 G
117	$53 \\ 47$	$\begin{array}{c} 158\\ 136 \end{array}$	· 62 66	$\begin{array}{c} 269 \\ 248 \end{array}$	$\begin{array}{c} 142 \\ 136 \end{array}$	129 123	77 66	177 167	$\begin{array}{c} 41 \\ 43 \end{array}$	$\begin{array}{c} 100\\ 86 \end{array}$	$\begin{array}{c} 15\\14 \end{array}$	$\begin{array}{c} 25 \\ 26 \end{array}$	$\begin{array}{c} 258\\ 248 \end{array}$
118	$\begin{array}{c} 46 \\ 50 \end{array}$	$\begin{array}{c} 149 \\ 149 \end{array}$	$\begin{array}{c} 62 \\ 68 \end{array}$	80 83	$\begin{array}{c} 50 \\ 50 \end{array}$	77 80	86 93	$\begin{array}{c} 230\\ 230\end{array}$	$\begin{array}{c} 49 \\ 52 \end{array}$	72 68	$\frac{24}{24}$	$\begin{array}{c} 19 \\ 18 \end{array}$	$\begin{array}{c} 327\\ 356 \end{array}$
119	$\begin{array}{c} 48 \\ 42 \end{array}$	$\begin{array}{c} 142 \\ 136 \end{array}$	$\begin{array}{c} 55\\62\end{array}$	70 66	$\begin{array}{c} 230\\113 \end{array}$	$\begin{array}{c} 60 \\ 53 \end{array}$	66 66	89 96	$\begin{array}{c} 49 \\ 53 \end{array}$	$\begin{array}{c} 64 \\ 53 \end{array}$	$29 \\ 25$	15 _14	$\begin{array}{c} 365\\ 348 \end{array}$
120	53 37	89 83	$\begin{array}{c} 43 \\ 68 \end{array}$	$\begin{array}{c} 258 \\ 230 \end{array}$	$\begin{array}{c} 129 \\ 129 \end{array}$	$\begin{array}{c} 136 \\ 129 \end{array}$	$\begin{array}{c} 108\\93 \end{array}$	$\begin{array}{c} 136\\ 136\end{array}$	33 37	177 177	$9 \\ 8$	$\begin{array}{c} 14 \\ 12 \end{array}$	333 327
121	33 37	72 75	$\begin{array}{c} 46 \\ 50 \end{array}$	$\begin{array}{c} 68 \\ 68 \end{array}$	$\begin{array}{c} 45\\ 49\end{array}$	$\begin{array}{c} 75 \\ 80 \end{array}$	$\begin{array}{c} 123 \\ 123 \end{array}$	$\begin{array}{c} 113\\ 123 \end{array}$	36 37	. 89 89	$\begin{array}{c} 15\\ 20 \end{array}$	$\frac{8}{6}$	$\begin{array}{c} 327\\ 393 \end{array}$
122	$\begin{array}{c} 37 \\ 42 \end{array}$	$\begin{array}{c} 113\\118\end{array}$	83 108	93 108	$\begin{array}{c} 50 \\ 60 \end{array}$	70 80	89 104	80 96	36 45	96 104	$\begin{array}{c} 18 \\ 24 \end{array}$	9 10	$\begin{array}{c} 327\\ 365 \end{array}$

Elements in red pine needle samples

G = Gamma values or relative intensity.

= See table 1 for other data on same samples.

Sample No.	1953 growth	Needle tips cm	В 2498 G	Si 2507 G	Р 2536 G	Mg 2779 G	Mn 2801 G	РЪ 2833 G	Fe 3020 G	A1 3093 G	Ca 31 5 9 G	Cu 3274 G	Ag 3280 G	Zn 3282 G	К 4044 G	Li 2741 G
236	Apical 2 cm	05	117 142	167 158	28 24	32 31	108 104	22 20	83 80	356 333	20 18	43 43	19 18	13 12	8 9	53 52
237	Apical 2 cm	.5-1.0	$\begin{array}{c} 75 \\ 62 \end{array}$	$\begin{array}{c} 72 \\ 62 \end{array}$	$\frac{24}{22}$	37 37	100 93	$\frac{21}{22}$	100 89	200 182	24 22	47 46	16 17	14 12	10 10	53 55
238	Apical 2 cm	1.0-	28 22	20 17	30 24	59 52	72 64	22 29	38 34	142 129	22 19	22 25	13 14	12 12	12 12	57 62
239	Basal 5 cm	0~.5	50 39	149 136	26 24	31 31	93 96	20 21	89 104	356 327	18 17	47 52	14 14	10 10	7 7	53 57
240	Basal 5 cm	.5-1.0	31 24	41 35	24 18	32 29	93 83	18 17	46 42	158 158	22 20	43 39	14 14	11 10	11 12	64 64
241	Basal 5 cm	1.0	22 15	28 22	30 22	44 39	66 60	17 20	43 36	123 118	20 18	22 27	12 11	11 12	12 12	57 62

TABLE 3 Elements in white pine needle samples from E. L. Stone, Professor Forest Soils, Cornell University. Plot B-22, 19 year old, sampled October 31, 1953

G = Gamma value or relative intensity.

Spectrographic Plate No. 27, Book No. 11, David Chou.

	White pine	e needles, n	ear greenho	use, Ohio A	1gr. Expt.	Station Ma	y, 1955	<u> </u>
	В 2798 G	Si 2881 G	Fe 3020 G	A1 3093 G	Mn 4035 G	Mg 2802 G	 1	Li 2741 G
$\begin{array}{c c} Tip & 1 \\ & 2 \\ & 3 \\ Base & 4 \end{array}$	$ \begin{array}{r} 108 \\ 24 \\ 10 \\ 5 \end{array} $	$136 \\ 41 \\ 50 \\ 35$	$ \begin{array}{r} 104 \\ 64 \\ 68 \\ 46 \end{array} $	248 86 104 70	96 83 68 35	80 86 96 64		72 75 104 104
	Р 2536 G	Pb 2833 G	Ca 3178 G	Cu 3274 G	Na 3302 G	Ag 3280 G	Zn 3282 G	К 4044 G
$\begin{array}{c c} Tip & 1 \\ & 2 \\ & 3 \\ Base & 4 \end{array}$	الم 8 11 8	26 30 41 33	12 18 32 35	37 39 53 53	$\begin{array}{c}12\\16\\20\\22\end{array}$	7 7 12 12	6 5 8 8	$\begin{array}{r} 32\\41\\64\\66\end{array}$

TABLE 4

Spectrographic Plate No. 52, Book No. 11, A. R. Correll.

Accuracy and expression of results.-Spectrographic analyses can be very accurate for certain elements, especially when these are present in very small amounts. The National Bureau of Standards accepts the spectrographic results over chemical results in many instances where only small amounts are involved. In most of the work reported in this paper only large differences in concentration are discussed.

The results of spectrographic analysis can be expressed just as any other analysis, in percent or parts per million, but sometimes this is very difficult because of the very small amounts involved. It is often better to express the results in comparative amounts rather than in absolute values. One should know the copper constant of an ore very accurately; but since all biological material contains a little copper, it is often more important to know the relative amount of copper in one tissue as compared to another than to have the absolute amount in each. And it is much easier to do this.

Starting with the spectra of a series of plant samples on a spectrographic plate, one can first of all visually examine the density of the lines in the different spectra and see whether there is more in one than in another. Next the lines can be measured with the densitometer and compared. The values of the density measurements can be corrected for emulsion response, and for spectrogram variations (internal standard), and further compared. The values can be expressed in absolute amounts in the sample mixture from prepared working curves. Finally, if the ash content of the dried tissue is known, they can be expressed in amount per unit of dry tissue and if moisture were determined on the wet plant tissue, then in amounts in the wet tissue.

RESULTS

This work, resulting from a rather extensive program on the study of mineral elements in the margins and tips of corn leaves, has been extended to many other plants such as sorghum, sugarcane, wheat, oats, grass, iris, lily, wheat awns, and pine needles.

Only the results on pine needles are discussed here. The samples were obtained from New York State through the kindness of E. L. Stone at Cornell and from the campus and arboretum here. Several graduate students and technicians besides myself have worked on the samples.

The results of the boron analyses in the first set of samples from New York State are tabulated in table 1 in some detail to show how is it done. These data show the high accumulation of boron in the tips of needles.

Table 2 shows the results of spectrographic examination of the same set of samples for eleven other elements besides boron. These data are summarized as gamma values or relative intensities only to save space. They show that, in addition to boron, silicon, manganese, iron, copper, and aluminum are higher in the tips than in the rest of the needle. Phosphorus, magnesium, lead, calcium, silver, or zinc, show no difference; potassium is slightly lower in the tips than in the other parts of the tissue.

The data in table 3 show the elements examined in white pine needles. The sample size was too small for a 90–10 buffer mixture, so a mixture of 100 milligrams of LiF and 100 milligrams of tissue were used. The results are recorded in relative intensities and they show a high accumulation of boron, silicon, aluminum, manganeses, iron, and copper in the tips and about equal amounts of the other elements throughout the needle, with the exception of potassium which is lower in the tips.

A set of white pine needle samples was obtained by Tzu-liang Yuan at Wooster and separation or dissection made into 4 parts. These samples were put on Plate No. 52 by A. R. Correll and the results are shown in table 4. Here again there was an accumulation of boron, silicon, aluminum, iron, and manganese in the tips of the needles, with six of the elements about equally distributed in the needle, and potassium again lower in the tips then elswhere.

SUMMARY

It appears from these studies on pine needles that boron, silicon, manganese, aluminum, iron, and copper accumulate in the tips; that phosphorus, magnesium, lead, calcium, silver, and zinc are uniformly distributed and that potassium is lower in the tips and high in the base of the needle.