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## Metagenomics Reveals Planktonic Bacterial Community Shifts across a Natural CO<sub>2</sub> Gradient in the Mediterranean Sea

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Bacterial communities at a  $CO_2$  vent (pH 6.7) were compared with those at control (pH 8.0) and transition sites (pH 7.6) using 16S rRNA metagenomics. *Firmicutes* and unclassified bacteria dominated across all sites, *Proteobacteria*, especially *Gammaproteobacteria*, declined, and *Epsilonproteobacteria* increased in the vent with an increase in *Bacteroidetes* at both the vent and transition sites.

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The current average oceanic surface pH of 8.1 is falling rapidly, at an unprecedented rate, due to increasing anthropogenic  $CO_2$  emissions (1), impacting biodiversity and fundamentally altering food webs (2). It is difficult to simulate ocean acidification for sufficient periods to monitor ecosystem level effects, so submarine  $CO_2$  vent ecosystems are being used as "natural laboratories" to gauge acidification impacts (3–7). Because marine microorganisms recycle organic matter, making it available to higher life (8), pH shifts can have dramatic and long-lasting impacts on microbially mediated processes. We used amplicon-based metagenomic sequencing to assess planktonic microbial communities at a vent off Vulcano Island (38°25′08.52″N, 14°57′39.13″E), Italy, where gaseous emissions comprise >98%  $CO_2$ , causing a pH gradient reaching ambient conditions (pH ~8.1), ca. 350 m from the intense  $CO_2$  leakage site (4).

Replicate surface water samples (n = 3 to 5) from the Vulcano vent ecosystem were collected from 300-m transects using a peristaltic pump into sterilized and dissolved organic carbon (DOC) clean containers. CO<sub>2</sub> concentration gradients from where the samples were collected ranged from modern levels to projected late-21st-century conditions (pH ~7.6); thus, reference samples had mean pH 8.0, transition pH 7.6, and vent pH 6.7, respectively. Samples were filtered through 0.22-µm pore-size filters and shipped overnight to the Chauhan laboratory, where metagenomic DNA was extracted from the filters with a PowerWater DNA isolation kit (Mo Bio Laboratories, Inc., Carlsbad, CA, USA), and pyrotag sequencing of the 16S rRNA gene V1-V3 region was performed using standard protocols (9). Sequences obtained using a Roche 454 FLX instrument (Roche, Indianapolis, IN, USA) were processed using MOTHUR (10) as demonstrated (11), and a total of 95.1 Mb data containing 79,078 16S rRNA gene sequences were obtained and identified using RDP (12) and MG-RAST (13).

Heatmap and UniFrac analysis revealed that the vent microbiota were taxonomically distinct from those in the transition and reference samples. Interestingly, 8% to 37% of the retrieved sequences remained taxonomically unresolved, especially from the vent, indicating the presence of potentially novel bacteria within this Mediterranean ecosystem. Relative abundances of *Firmicutes* were highest across all sites (37% to 70%), which is rather unusual for marine waters. However, previous studies have, in fact, identified bacteria, such as novel *Bacillus vulcani* and *Bacillus aeolius* (14, 15), from the Gram-positive *Firmicutes* phylum from this ecosystem.

Of particular interest was the decline of Proteobacteria, especially Gammaproteobacteria, and the increase in Epsilonproteobacteria at the vent relative to the transition and reference sites. It is likely that the high CO<sub>2</sub>-low pH environment favors chemosynthetic Epsilonproteobacteria, many of which are human and animal pathogens (16, 17). The abundances of Bacteroidetes in both the vent and transition sites also increased compared with the reference site, suggesting enhanced availability of high-molecularweight dissolved organic matter (DOM) on which Bacteroidetes are known to thrive (18, 19). Because the gas composition at the Vulcano seep consists of >98% carbon dioxide (4), elevated CO<sub>2</sub> potentially stimulates DOM production and consumption, as demonstrated previously in mesocosms held at different CO<sub>2</sub> levels (20). Further metagenomic analysis of this high CO<sub>2</sub>-low pH "natural laboratory" may strengthen our ability to predict the impacts of ocean acidification on marine microorganisms.

**Nucleotide sequence accession number.** The DNA sequences from this metagenomic project were deposited in the Sequence Read Archive under the accession no. SRP050984.

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## REFERENCES

- Caldeira K, Wickett ME. 2003. Anthropogenic carbon and ocean pH. Nature 425:365. http://dx.doi.org/10.1038/425365a.
- Wittmann AC, Pörtner HO. 2013. Sensitivities of extant animal taxa to ocean acidification. Nat Clim Change 3:995–1001. http://dx.doi.org/ 10.1038/nclimate1982.
- Hall-Spencer JM, Rodolfo-Metalpa R, Martin S, Ransome E, Fine M, Turner SM, Rowley SJ, Tedesco D, Buia MC. 2008. Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. Nature 454: 96–99. http://dx.doi.org/10.1038/nature07051.
- Boatta F, D'Alessandro W, Gagliano AL, Liotta M, Milazzo M, Rodolfo-Metalpa R, Hall-Spencer JM, Parello F. 2013. Geochemical survey of Levante Bay, Vulcano Island (Italy), and its suitability as a natural laboratory for the study of ocean acidification. Mar Pollut Bull 73:485–494. http://dx.doi.org/10.1016/j.marpolbul.2013.01.029.
- Taylor JD, Ellis R, Milazzo M, Hall-Spencer JM, Cunliffe M. 2014. Intertidal epilithic bacteria diversity changes along a naturally occurring carbon dioxide and pH gradient. FEMS Microbiol Ecol 89:670–678. http://dx.doi.org/10.1111/1574-6941.12368.
- Ziveri P, Passaro M, Incarbona A, Milazzo M, Rodolfo-Metalpa R, Hall-Spencer JM. 2014. Decline in coccolithophore diversity and impact on coccolith morphogenesis along a natural CO<sub>2</sub> gradient. Biol Bull 226: 282–290.
- Kerfahi D, Hall-Spencer JM, Tripathi BM, Milazzo M, Lee J, Adams JM. 2014. Shallow water marine sediment bacterial community shifts along a natural CO<sub>2</sub> gradient in the Mediterranean Sea off Vulcano, Italy. Microb Ecol 67:819–828. http://dx.doi.org/10.1007/s00248-014-0368-7.
- Azam F. 1998. Microbial control of oceanic carbon flux: the plot thickens. Science 280:694–696. http://dx.doi.org/10.1126/science.280.5364.694.
- Handl S, Dowd SE, Garcia-Mazcorro JF, Steiner JM, Suchodolski JS. 2011. Massive parallel 16S rRNA gene pyrosequencing reveals highly diverse fecal bacterial and fungal communities in healthy dogs and cats. FEMS Microbiol Ecol 76:301–310. http://dx.doi.org/10.1111/j.1574 -6941.2011.01058.x.
- Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson CJ, Sahl JW, Stres B, Thallinger GG, Van Horn DJ, Weber CF. 2009. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. Appl Environ Microbiol 75:7537–7541. http://dx.doi.org/10.1128/AEM.01541-09.
- 11. Chauhan A, Wafula D, Lewis DE, Pathak A. 2014. Metagenomic assess-

ment of the eastern oyster-associated microbiota. Genome Announc 2(5): e01083-14. http://dx.doi.org/10.1128/genomeA.01083-14.

- Cole JR, Chai B, Farris RJ, Wang Q, Kulam SA, McGarrell DM, Garrity GM, Tiedje JM. 2005. The Ribosomal Database project (RDP-II): sequences and tools for high-throughput rRNA analysis. Nucleic Acids Res 33:D294–D296. http://dx.doi.org/10.1093/nar/gki038.
- 13. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.
- Caccamo D, Gugliandolo C, Stackebrandt E, Maugeri TL. 2000. Bacillus vulcani sp. nov., a novel thermophilic species isolated from a shallow marine hydrothermal vent. Int J Syst Evol Microbiol 50:2009–2012. http:// dx.doi.org/10.1099/00207713-50-6-2009.
- Gugliandolo C, Maugeri TL, Caccamo D, Stackebrandt E. 2003. Bacillus aeolius sp. nov. a novel thermophilic, halophilic marine Bacillus species from Eolian Islands (Italy). Syst Appl Microbiol 26:172–176. http:// dx.doi.org/10.1078/072320203322346001.
- Nakagawa S, Takaki Y, Shimamura S, Reysenbach AL, Takai K, Horikoshi K. 2007. Deep-sea vent epsilon-proteobacterial genomes provide insights into emergence of pathogens. Proc Natl Acad Sci U S A. 104: 12146–12150. http://dx.doi.org/10.1073/pnas.0700687104.
- Pérez-Rodríguez I, Bolognini M, Ricci J, Bini E, Vetriani C. 2014 Nov 14. From deep-sea volcanoes to human pathogens: a conserved quorumsensing signal in Epsilonproteobacteria. ISME J. http://dx.doi.org/ 10.1038/ismej.2014.214.
- Teeling H, Fuchs BM, Becher D, Klockow C, Gardebrecht A, Bennke CM, Kassabgy M, Huang S, Mann AJ, Waldmann J, Weber M, Klindworth A, Otto A, Lange J, Bernhardt J, Reinsch C, Hecker M, Peplies J, Bockelmann FD, Callies U. 2012. Substrate-controlled succession of marine bacterioplankton populations induced by a phytoplankton bloom. Science 336:608–611. http://dx.doi.org/10.1126/science.1218344.
- Fernández-Gómez B, Richter M, Schüler M, Pinhassi J, Acinas SG, González JM, Pedrós-Alió C. 2013. Ecology of marine *Bacteroidetes*: a comparative genomics approach. ISME J 7:1026–1037. http://dx.doi.org/ 10.1038/ismej.2012.169.
- Riebesell U, Schulz KG, Bellerby RG, Botros M, Fritsche P, Meyerhöfer M, Neill C, Nondal G, Oschlies A, Wohlers J, Zöllner E. 2007. Enhanced biological carbon consumption in a high CO<sub>2</sub> ocean. Nature 450:545–548. http://dx.doi.org/10.1038/nature06267.