

GCB Bioenergy (2016) 8, 925–940, doi: 10.1111/gcbb.12298

# Simulation of greenhouse gases following land-use change to bioenergy crops using the ECOSSE model: a comparison between site measurements and model predictions

MARTA DONDINI<sup>1</sup>, MARK I. A. RICHARDS<sup>1</sup>, MARK POGSON<sup>1,2</sup>, JON MCCALMONT<sup>3</sup>, JULIA DREWER<sup>4</sup>, RACHEL MARSHALL<sup>5</sup>, ROSS MORRISON<sup>6</sup>, SIRWAN YAMULKI<sup>7</sup>, ZOE M. HARRIS<sup>8</sup>, GIORGIO ALBERTI<sup>8,9</sup>, LUKAS SIEBICKE<sup>10</sup>, GAIL TAYLOR<sup>8</sup>, MIKE PERKS<sup>10</sup>, JON FINCH<sup>6</sup>, NIALL P. MCNAMARA<sup>5</sup>, JOANNE U. SMITH<sup>1</sup> and PETE SMITH<sup>1</sup>

<sup>1</sup>Institute of Biological and Environmental Sciences, School of Biological Sciences, University of Aberdeen, 23 St Machar Drive, Aberdeen, UK, <sup>2</sup>Academic Group of Engineering, Sports and Sciences, University of Bolton, Deane Road, Bolton, BL3 5AB, UK, <sup>3</sup>Institute of Biological, Environmental & Rural Sciences (IBERS), Aberystwyth University, Gogerddan, Aberystwyth, UK, <sup>4</sup>Centre for Ecology & Hydrology, Bush Estate, Penicuik, EH26 0QB, Midlothian, UK, <sup>5</sup>Centre for Ecology & Hydrology, Lancaster Environment Centre, Library Avenue, Bailrigg, Lancaster, UK, <sup>6</sup>Centre for Ecology & Hydrology, MacLean Building, Benson Lane, Crowmarsh Gifford, Wallingford, Oxfordshire, UK, <sup>7</sup>Forest Research, Alice Holt Lodge, Farnham, GU10 4LH, UK, <sup>8</sup>Centre for Biological Sciences, Faculty of Natural & Environmental Sciences, University of Southampton, Highfield Campus, Southampton, UK, <sup>9</sup>Department of Agricultural and Environmental Sciences, University of Udine, Udine, Italy, <sup>10</sup>Forest Research, Northern Research Station, Roslin, Midlothian, EH25 9SY, UK

## Abstract

This article evaluates the suitability of the ECOSSE model to estimate soil greenhouse gas (GHG) fluxes from short rotation coppice willow (SRC-Willow), short rotation forestry (SRF-Scots Pine) and *Miscanthus* after land-use change from conventional systems (grassland and arable). We simulate heterotrophic respiration ( $R_h$ ), nitrous oxide ( $N_2O$ ) and methane ( $CH_4$ ) fluxes at four paired sites in the UK and compare them to estimates of  $R_h$  derived from the ecosystem respiration estimated from eddy covariance (EC) and  $R_h$  estimated from chamber (IRGA) measurements, as well as direct measurements of  $N_2O$  and  $CH_4$  fluxes. Significant association between modelled and EC-derived  $R_h$  was found under *Miscanthus*, with correlation coefficient ( $r$ ) ranging between 0.54 and 0.70. Association between IRGA-derived  $R_h$  and modelled outputs was statistically significant at the Aberystwyth site ( $r = 0.64$ ), but not significant at the Lincolnshire site ( $r = 0.29$ ). At all SRC-Willow sites, significant association was found between modelled and measurement-derived  $R_h$  ( $0.44 \leq r \leq 0.77$ ); significant error was found only for the EC-derived  $R_h$  at the Lincolnshire site. Significant association and no significant error were also found for SRF-Scots Pine and perennial grass. For the arable fields, the modelled  $CO_2$  correlated well just with the IRGA-derived  $R_h$  at one site ( $r = 0.75$ ). No bias in the model was found at any site, regardless of the measurement type used for the model evaluation. Across all land uses, fluxes of  $CH_4$  and  $N_2O$  were shown to represent a small proportion of the total GHG balance; these fluxes have been modelled adequately on a monthly time-step. This study provides confidence in using ECOSSE for predicting the impacts of future land use on GHG balance, at site level as well as at national level.

**Keywords:** ECOSSE model, energy crops, greenhouse gases, land-use change, *Miscanthus*, short rotation coppice, short rotation forestry

Received 4 June 2015; revised version received 18 July 2015 and accepted 3 August 2015

## Introduction

The interest in using bioenergy crops as an alternative energy source to fossil fuels, and to reduce greenhouse

gas (GHG) emissions, has increased in recent decades (Hastings *et al.*, 2014). The commitment of the European Union is to increase the percentage of energy from renewable sources to 20% of total energy consumption by 2020 (EU, 2009). Under the Climate Change Act 2008 (Great Britain, 2008), the UK government committed to reduce GHG emissions by 80% in 2050 compared

Correspondence: M. Dondini, tel. +44 01224 273810, fax +44 01224 272703, e-mail: [marta.dondini@abdn.ac.uk](mailto:marta.dondini@abdn.ac.uk)

to 1990 levels; the use of bioenergy could contribute to this target using dedicated 'second generation' (2G) lignocellulosic crops/plantations, including short rotation coppice (SRC), *Miscanthus* and short rotation forestry (SRF) (Somerville *et al.*, 2010; McKay, 2011; DECC, 2012; Valentine *et al.*, 2012). Consequently, a substantial land-use change (LUC) may occur, and it might have considerable environmental and economic impact (Fargione *et al.*, 2008; Searchinger *et al.*, 2008; Gelfand *et al.*, 2011).

Carbon dioxide (CO<sub>2</sub>) emissions of bioenergy had previously been assumed to be zero (Gustavsson *et al.*, 1995; UK, 2008) on the assumption that emissions during combustion are balanced by the carbon (C) uptake during the growth of these bioenergy plantations, but this fails to take account of GHG emissions following LUC and subsequent crop growth. To this end, it is important to assess the GHG balance of bioenergy crops, particularly during the first years after conversion.

Two approaches have been widely used to monitor CO<sub>2</sub> fluxes: eddy covariance (EC) and the enclosure (or chamber) method. Eddy covariance (McMillen, 1988; Aubinet *et al.*, 2012) is a technique developed to estimate land-atmosphere exchange of gas and energy at ecosystem scale. The measured CO<sub>2</sub> flux, known as net ecosystem exchange (NEE), includes ecosystem respiration ( $R_{\text{eco}}$ ) which consists of heterotrophic ( $R_{\text{h}}$ ) and autotrophic ( $R_{\text{a}}$ ) respiration, and gross primary production (GPP) at ecosystem scale. As photosynthesis only occurs during daylight hours, the night time flux is typically used to partition the NEE signal between GPP and  $R_{\text{eco}}$ . A flux-partitioning algorithm that defines a short-term temperature sensitivity of  $R_{\text{eco}}$  is applied to extrapolate CO<sub>2</sub> fluxes from night to day (Reichstein *et al.*, 2005). In a plant removal experiment (Hardie *et al.*, 2009), the total  $R_{\text{h}}$  from the whole soil profile was found to be approximately between 46 and 59% of the total  $R_{\text{eco}}$ . Abdalla *et al.* (2014) used these values to simulate  $R_{\text{h}}$  from selected European peatland sites using a soil process-based model, ECOSSE.

Enclosure methods have been developed to measure CO<sub>2</sub> efflux from soil; these methods involve covering an area of soil surface with a chamber and the soil CO<sub>2</sub> efflux can be determined using two main modes: dynamic (closed or open) and closed static. In the former mode, a steady stream of air is pumped directly in to the chamber (Christensen, 1983; Skiba *et al.*, 1992). The latter mode simply involves closing the chamber for approximately 20–60 min and taking gas samples at intervals for analysis (Hutchinson & Mosier, 1981), or circulating the chamber air through a nondestructive infrared gas analyser (IRGA) for approximately 2 min (Norman *et al.*, 1992; Smith & Mullins, 2000). Several studies have used the closed chamber method com-

bined with root-exclusion methods, tree grilling or stable isotopes to understand the relative contribution of  $R_{\text{h}}$  and  $R_{\text{a}}$  to total soil respiration ( $R_{\text{tot}}$ ) under different land uses.

Byrne & Kiely (2006) demonstrated that  $R_{\text{a}}$  under grassland soil in Ireland accounted for approximately 50% of  $R_{\text{tot}}$  during the summer months and 38% during the rest of the year. Pacaldo *et al.* (2013) reported a contribution of  $R_{\text{a}}$  of about 18–33% of  $R_{\text{tot}}$  under SRC-Willow at three different development stages in the USA. In a study on commercial farms located across the UK, Koerber *et al.* (2010) reported a contribution of  $R_{\text{h}}$  on  $R_{\text{tot}}$  for wheat of approximately 32% from January to May, 79% from June to September and 67% from October to December. A meta-analysis of soil respiration partitioning studies reported values for the ratio  $R_{\text{h}}/R_{\text{tot}}$  for forest soils as ranging from 0.03 to 1.0 (Subke *et al.*, 2006). Overall, the ratio was higher for boreal coniferous forests than temperate sites. In temperate, mixed deciduous forests ranges for  $R_{\text{h}}/R_{\text{tot}}$  of 0.3–0.6 were reported (Gaudinski *et al.*, 2000; Borken *et al.*, 2006; Millard *et al.*, 2010; Heinemeyer *et al.*, 2012). Several studies have also shown that bioenergy plantations have low nitrous oxide (N<sub>2</sub>O) emissions compared to agricultural crops because of their lower nutrient requirements, thus reducing the fertilizer requirements, and more efficient nutrient uptake, thus increasing competition with microbial organisms of N<sub>2</sub>O production (Flessa *et al.*, 1998; Hellebrand *et al.*, 2010; Drewer *et al.*, 2012).

Methane (CH<sub>4</sub>) is another important GHG that may be a substantial component of the GHG balance from several terrestrial ecosystems (van den Pol-van Dassel *et al.*, 1999). In agricultural systems, soil is typically a small net source or sink for CH<sub>4</sub> (Boeckx & Van Cleemput, 2001). Bioenergy crops usually present either a small CH<sub>4</sub> sink (Hellebrand *et al.*, 2003; Kern *et al.*, 2012) or a small CH<sub>4</sub> source (Gelfand *et al.*, 2011). The magnitude of the CH<sub>4</sub> flux is typically much smaller than CO<sub>2</sub> and N<sub>2</sub>O, in both agricultural soils (Boeckx & Van Cleemput, 2001) and bioenergy crops (Hellebrand *et al.*, 2003). However, very few studies (Hellebrand *et al.*, 2003; Gelfand *et al.*, 2011; Kern *et al.*, 2012) have reported on the contribution of CH<sub>4</sub> emission from bioenergy systems, increasing uncertainty in the direction of this small flux (Zona *et al.*, 2013).

Several factors control the GHG emissions of both bioenergy and conventional crops, such as site management, for example fertilization (Crutzen *et al.*, 2008; Hellebrand *et al.*, 2008, 2010), previous land use (Smith & Conen, 2004) and climatic conditions (Flessa *et al.*, 1998; Hellebrand *et al.*, 2003). Despite the high variability of the GHG fluxes, to our knowledge, only one study in the UK (Drewer *et al.*, 2012) has reported on all three GHG fluxes (CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub>) from soils under

bioenergy crops (*Miscanthus* and SRC-Willow) and, in particular, after transition from former conventional systems. To fill this gap, soil models are a useful tool to predict GHG fluxes when site measurements are not available, especially when studying the effects of the change in land use over time and under different climatic conditions over large areas.

However, soil models need to be extensively tested under a range of climates and soils before being applied under conditions different from those used to parameterize and calibrate the model itself. In fact, model evaluation involves running a model using input values that have not been used during the calibration process, demonstrating that it is capable of making accurate simulations under a wide range of conditions (Moriassi *et al.*, 2007). A model can only be properly evaluated against independent data and a useful model should be able to simulate those data with some degree of accuracy (Smith & Smith, 2007).

Although several soil models have been developed for conventional agricultural and forest systems, most of them have not been fully parameterized and effectively tested for application on 2G bioenergy crops, such as *Miscanthus*, SRF and SRC (Dimitriou *et al.*, 2012; Borzęcka-Walker *et al.*, 2013; Robertson *et al.*, 2015). Here, we focus on the applicability of the process-based model ECOSSE to predict soil CO<sub>2</sub> (heterotrophic respiration), N<sub>2</sub>O and CH<sub>4</sub> after transition from conventional to bioenergy crops.

The ECOSSE model was developed mainly to simulate the C and nitrogen (N) cycles using minimal input data on both mineral and organic soils (Smith *et al.*, 2010a,b). The ECOSSE model has been previously evaluated across the UK to simulate the effect on soil C of LUC to SRF (Dondini *et al.*, 2015a), *Miscanthus* and SRC-Willow (Dondini *et al.*, 2015b), to simulate soil N<sub>2</sub>O emissions in cropland sites in Europe (Smith *et al.*, 2010b; Bell *et al.*, 2012) and CO<sub>2</sub> emissions from peatlands (Abdalla *et al.*, 2014).

This article evaluates the suitability of ECOSSE for estimating soil GHG fluxes from SRC-Willow, SRF-Scots Pine and *Miscanthus* soils in the UK after LUC from conventional systems (grassland and arable). Based on previously published recommendations, a combination of graphical techniques and error statistics has been used for model evaluation (Moriassi *et al.*, 2007). Model testing is often limited by the lack of field data to which the simulations can be compared (Desjardins *et al.*, 2010). In this study, the model is evaluated against 2 years of observations at four locations in the UK, comprising one transition to SRF-Scots Pine, three transitions to SRC-Willow and two transitions to *Miscanthus*. Modelled GHG fluxes from conventional systems have also been evaluated against field measurements (three grassland and two arable fields).

## Materials and methods

### ECOSSE model

The ECOSSE model includes five pools of soil organic matter, each decomposing with a specific rate constant except for the inert organic matter (IOM) which is not affected by decomposition. Decomposition is sensitive to temperature, soil moisture and vegetation cover; soil texture (sand, silt and clay), pH and bulk density of the soil along with monthly climate and land-use data are the inputs to the model (Coleman & Jenkinson, 1996; Smith *et al.*, 1997). The ECOSSE model is able to simulate C and N cycle for six land-use categories of vegetation: arable, grassland, forestry, seminatural, *Miscanthus* and short rotation coppice willow (SRC-Willow).

The vegetation input to the soil (SI) is estimated by a subroutine in the ECOSSE model which uses a modification of the Miami model (Lieth, 1972), a simple model that links the climatic net primary production of biomass (NPP) to annual mean temperature and total precipitation (Grieser *et al.*, 2006). For a full description of the ECOSSE model and the plant input, estimates refer to Smith *et al.* (2010a) and Dondini *et al.* (2015b).

The minimum ECOSSE input requirements for site-specific simulations are as follows:

#### Climate/atmospheric data:

- 30-year average monthly rainfall, potential evapotranspiration (PET) and temperature,
- Monthly rainfall, temperature and PET.

#### Soil data:

- Initial soil C content (kg ha<sup>-1</sup>),
- Soil sand, silt and clay content (%),
- Soil bulk density (g cm<sup>-3</sup>),
- Soil pH and
- Soil depth (cm).

#### Land-use data:

- Land use for each simulation year.

The initialization of the model is based on the assumption that the soil column is at steady state under the initial land use at the start of the simulation. Previous work has used soil organic carbon (SOC) measured at steady state to determine the plant inputs that would be required to achieve an equivalent simulated value (e.g. Smith *et al.*, 2010a). This approach iteratively adjusts plant inputs until measured and simulated values of SOC converge. In the absence of additional measurements, estimated plant inputs were calculated from a feature built in the ECOSSE model which combine the NPP model Miami (Lieth, 1972, 1973), land-management practices of the initial land use and measured above-ground biomass (details are given in Dondini *et al.*, 2015b).

### Data

In 2011–2013, four sites were sampled in Britain using a paired site comparison approach (Keith *et al.*, 2015; Rowe *et al.*, 2015).

The sites and the relative measurements contribute to the ELUM (Ecosystem Land Use Modelling & Soil Carbon GHG Flux Trial) project (Harris *et al.*, 2014). Each site consisted of one reference field (arable or grassland, depending on the previous land use of the bioenergy fields) and one or more adjacent bioenergy fields (*Miscanthus*, SRC-Willow, SRF-Scots Pine), for a total of six transitions to bioenergy at four sites across UK (Table 1). A full description of the sites can be found in Drewer *et al.* (2012, 2015); J. McCalmont, N. McNamara, I. Donnison and J. Clifton-Brown (in preparation); and Z. M. Harris, G. Alberti, J. R. Jenkins, E. Clark, R. Marshall, R. Rowe, N. McNamara and G. Taylor (in preparation).

At each bioenergy and reference field, the NEE data were obtained from continuous EC measurements (McMillen, 1988; Aubinet *et al.*, 2012) using open path IRGAs (LI-7500) and sonic anemometers. All details regarding the EC data corrections, quality control, footprint and gap filling procedures can be found in Aubinet *et al.* (2003). The night time fluxes were used to partition the NEE flux measurements into GPP and  $R_{eco}$  (Reichstein *et al.*, 2005).

Soil GHG fluxes were measured on a monthly basis at eight points randomly distributed within each field. Soil CO<sub>2</sub> fluxes were measured using an IRGA connected to an SRC-1 soil respiration chamber (PP Systems, Amesbury, MA, USA). Measurements of soil CH<sub>4</sub> and N<sub>2</sub>O fluxes were made using a static chamber method (approx. 30 l) with the addition of a vent to compensate for pressure changes within the chamber during times of sampling. Gas samples were analysed by gas chromatograph. All details regarding the chamber data can be found in Drewer *et al.* (2012), Yamulki *et al.* (2013) and Case *et al.* (2014).

Measurements of soil C, soil bulk density and soil pH to 1 m soil depth, as well as information on the land-use history, were collected for each field (Keith *et al.*, 2015; Rowe *et al.*, 2015). Soil texture was measured for each site up to a depth of 30 cm; values to 1 m soil depth were extracted from the soil database (1 km resolution) described in Bradley *et al.* (2005), which is a collated soils data set for England and Wales, Scotland and Northern Ireland. Air temperature and precipitation data at

each location were extracted from the E-OBS gridded data set from the EU-FP6 project ENSEMBLES, provided by the ECA&D project (Haylock *et al.*, 2008). This data set is known as E-OBS and is publicly available (<http://eca.knmi.nl/>). For each location, monthly air temperature and precipitation for the 30 years before measurements started were used to calculate a long-term average (Table 2). At each site, air temperature and precipitation were collected during the entire study period and monthly values were used as input to the model. Monthly PET was estimated using the Thornthwaite method (Thornthwaite, 1948), which has been used in other modelling studies when direct observational data have not been available (e.g. Smith *et al.*, 2005; Dondini *et al.*, 2015a).

### Model evaluation and statistical analysis

Monthly simulations of soil CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub> fluxes were evaluated against monthly chamber measurements. In addition, the soil CO<sub>2</sub> predicted by the ECOSSE model was compared to estimates of  $R_h$  derived from the NEE measured by the EC.

At each site, the ECOSSE model has been run for the reference field (i.e. no land-use transition) and the bioenergy crop field (i.e. following transition from the reference land cover). The reference fields have been run for the conventional crop (arable, grassland) with no LUC, and the length of the simulations has been defined by the age of the plantation. At the bioenergy sites, the model has been run for the reference fields (conventional crop) with LUC to bioenergy crop; the length of the simulations was based on the time after transition to bioenergy crop. Measured soil characteristics and meteorological data have been used as inputs to drive the model (see above for input details), and the results of the simulations were compared to the GHG fluxes measured at the sites.

We expected a monthly underestimate of the soil CO<sub>2</sub> flux simulations because the ECOSSE model simulates  $R_h$  (from living micro-organisms + decomposition of old C sources, i.e. saprotrophic), while the CO<sub>2</sub> fluxes measured at the sites represent the total CO<sub>2</sub> efflux from the soil profile ( $R_a + R_h$ , chamber

**Table 1** Details of soil C, soil bulk density and soil pH to 1 m soil depth, as well as information on the land-use history at the study fields. Soil texture to 1 m soil depth was extracted from the soil database (1 km resolution) described in Bradley *et al.* (2005)

Site	Land use	Latitude, longitude	Establishment year	Carbon (%)	Nitrogen (%)	Bulk density (g cm <sup>-3</sup> )
West Sussex	Short rotation coppice (SRC)-Willow	50.9, -0.4	2008	0.63	0.17	1.50
	Grassland	50.9, -0.4	2000	0.53	0.17	1.55
East Grange	Short rotation forestry (SRF)-Scots Pine	56.0, -3.6	2009	0.95	0.18	1.47
	Grassland	56.0, -3.6	2009	1.30	0.17	1.49
	SRC-Willow	56.0, -3.6	2009	1.57	0.17	1.38
	Arable	56.0, -3.6	Pre-1990	1.37	0.18	1.57
Lincolnshire	SRC-Willow	53.1, -0.3	2006	1.26	0.11	1.41
	<i>Miscanthus</i>	53.1, -0.4	2006	1.30	0.13	1.53
	Arable	53.1, -0.5	Pre-1990	1.47	0.13	1.37
Aberystwyth	<i>Miscanthus</i>	52.4, -4.0	2012	0.98	0.25	1.21
	Grassland	52.4, -4.0	Pre-2007	1.16	0.26	1.45



**Table 2** Long-term (30 years) monthly rainfall, temperature, potential evapotranspiration (PET). Monthly rainfall and temperature were extracted from the E-OBS data set (Haylock *et al.*, 2008; <http://eca.knmi.nl/>). Monthly PET was estimated using the Thornthwaite method (Thornthwaite, 1948)

Month	Aberystwyth			East Grange			Lincoln			West Sussex		
	Rain (mm)	Temperature (°C)	PET (mm)	Rain (mm)	Temperature (°C)	PET (mm)	Rain (mm)	Temperature (°C)	PET (mm)	Rain (mm)	Temperature (°C)	PET (mm)
January	152	4	15	103	3	11	48	4	13	80	5	16
February	112	4	17	72	3	15	37	4	17	54	5	18
March	124	5	29	74	5	27	41	6	30	55	7	30
April	86	7	45	53	7	47	43	9	48	46	9	48
May	82	10	69	61	10	72	45	12	73	47	12	73
June	93	13	89	60	13	96	56	14	97	48	15	95
July	105	15	101	67	14	105	49	17	112	49	17	110
August	114	14	93	77	14	96	55	17	103	52	17	103
September	121	13	71	84	12	70	49	14	76	60	15	79
October	174	10	46	100	9	43	55	11	46	99	12	51
November	171	7	27	94	5	22	53	7	25	88	8	29
December	168	4	17	91	3	12	51	4	14	86	6	18

measurements) or NEE (EC measurements). To compare the modelled and measured  $R_h$ , we estimated the  $R_h$  as a proportion of the measured  $CO_2$  flux, depending on the measurement type (except EC data), vegetation type and growing season.

The EC measurements of NEE were used to derive  $R_{eco}$ ; to our knowledge, only the study by Abdalla *et al.* (2014) has reported estimates of  $R_h$  from  $R_{eco}$ . Abdalla *et al.* (2014) applied the approach proposed by Hardie *et al.* (2009) for peaty soils and reported a contribution of  $R_h$  to  $R_{eco}$  of 46–59%.

To represent the variations in  $R_h$  throughout the year, Abdalla *et al.* (2014) assumed that  $R_h$  was at the lowest value of the range (46%  $R_{eco}$ ) during the summer (June–August), the highest value (59%  $R_{eco}$ ) during the winter (December–February) and at the mean value (52.5%  $R_{eco}$ ) during the rest of the year (March–May and September–November). In this study, we used the same approach of Abdalla *et al.* (2014) to derive  $R_h$  from EC measurements from all land-use systems.

Chamber measurements represent the total  $CO_2$  flux from the soil as the sum of  $R_a$  and  $R_h$ , with the exception of grassland where exclusion of full leaves from the chamber is difficult, and therefore, above-ground plant respiration is also included in the measurements. We conducted a literature review to determine the partitioning of  $R_{tot}$  measured by the chambers under different vegetation types. Additional experiments within the ELUM project were also undertaken to directly quantify  $R_h$  and  $R_a$  at selected network sites (data not shown); where available, we used the  $R_h$  site data to estimate  $R_h$  from  $R_{tot}$  measured by the chambers (Lincolnshire – *Miscanthus*, West Sussex – SRC-Willow, Aberystwyth – *Miscanthus*). An overview of the data source and the monthly proportion of  $R_h$  for each vegetation type and at each site are shown in Table 3.

A quantitative statistical analysis was undertaken to determine the coincidence and association between measured and modelled values, following methods described in Smith *et al.* (1997) and Smith & Smith (2007). The statistical significance of the difference between model outputs and experimental

observations can be quantified if the standard error of the measured values is known (Hastings *et al.*, 2010). The standard errors (data not shown) and 95% confidence intervals around the mean measurements were calculated for all field sites.

The degree of association between modelled and measured values was determined using the correlation coefficient ( $r$ ). Values for  $r$  range from  $-1$  to  $+1$ . Values close to  $-1$  indicate a negative correlation between simulations and measurements, values of 0 indicate no correlation and values close to  $+1$  indicate a positive correlation (Smith & Smith, 2007). The significance of the association between simulations and measurements was assigned using a Student's  $t$ -test as outlined in Smith & Smith (2007).

Analysis of coincidence was undertaken to establish how different the measured and modelled values were. The degree of coincidence between the modelled and measured values was determined using the lack of fit statistic ( $LOFIT$ ), and its significance was assessed using an  $F$ -test (Whitmore, 1991) indicating whether the difference in the paired values of the two data sets is significant. The EC measurements were not replicated, so the coincidence between measured and modelled values was determined using the mean difference ( $M$ ), calculated as the sum of the differences between measured and modelled values and divided by the total number of measurements (Smith *et al.*, 1997). The variation across the different measurements was then used to calculate the value of Student's  $t$ -test and compared to the  $t$  distributions (two-tailed test) to obtain the probability that the mean difference is statistically significant. All statistical results were considered to be statistically significant at  $P < 0.05$ .

## Results

The ECOSSE model was evaluated by comparing the outputs to the EC-derived and IRGA-derived  $R_h$  fluxes from eleven fields over four sites, representing the

**Table 3** Contribution of heterotrophic respiration ( $R_h$ ) on total respiration ( $R_{tot}$ ) at the study sites

		Arable Koerber <i>et al.</i> (2010)	SRC-Willow Pacaldo <i>et al.</i> (2013)	<i>Miscanthus</i>	Grassland Byrne & Kiely (2006)	SRF-Scots Pine Millard <i>et al.</i> (2010)
Lincolnshire	January	32% $R_{tot}$	75% $R_{tot}$	41% $R_{tot}^*$		
	February	32% $R_{tot}$	75% $R_{tot}$	41% $R_{tot}^*$		
	March	32% $R_{tot}$	75% $R_{tot}$	85% $R_{tot}^*$		
	April	32% $R_{tot}$	75% $R_{tot}$	85% $R_{tot}^*$		
	May	32% $R_{tot}$	75% $R_{tot}$	85% $R_{tot}^*$		
	June	79% $R_{tot}$	75% $R_{tot}$	85% $R_{tot}^*$		
	July	79% $R_{tot}$	75% $R_{tot}$	44% $R_{tot}^*$		
	August	79% $R_{tot}$	75% $R_{tot}$	44% $R_{tot}^*$		
	September	79% $R_{tot}$	75% $R_{tot}$	44% $R_{tot}^*$		
	October	67% $R_{tot}$	75% $R_{tot}$	44% $R_{tot}^*$		
	November	67% $R_{tot}$	75% $R_{tot}$	41% $R_{tot}^*$		
	December	67% $R_{tot}$	75% $R_{tot}$	41% $R_{tot}^*$		
West Sussex	January		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	February		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	March		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	April		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	May		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	June		82% $R_{tot}^*$		40% $R_{tot}^\dagger$	
	July		82% $R_{tot}^*$		40% $R_{tot}^\dagger$	
	August		82% $R_{tot}^*$		40% $R_{tot}^\dagger$	
	September		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	October		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	November		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
	December		82% $R_{tot}^*$		60% $R_{tot}^\dagger$	
Aberystwyth	January			62% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	February			62% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	March			36% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	April			36% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	May			36% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	June			36% $R_{tot}^*$	40% $R_{tot}^\dagger$	
	July			36% $R_{tot}^*$	40% $R_{tot}^\dagger$	
	August			36% $R_{tot}^*$	40% $R_{tot}^\dagger$	
	September			36% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	October			36% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	November			62% $R_{tot}^*$	60% $R_{tot}^\dagger$	
	December			62% $R_{tot}^*$	60% $R_{tot}^\dagger$	
East Grange	January	32% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	February	32% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	March	32% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	April	32% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	May	32% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	June	79% $R_{tot}$	25% $R_{tot}$		40% $R_{tot}^\dagger$	61% $R_{tot}$
	July	79% $R_{tot}$	25% $R_{tot}$		40% $R_{tot}^\dagger$	61% $R_{tot}$
	August	79% $R_{tot}$	25% $R_{tot}$		40% $R_{tot}^\dagger$	61% $R_{tot}$
	September	79% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	October	67% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	November	67% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$
	December	67% $R_{tot}$	25% $R_{tot}$		60% $R_{tot}^\dagger$	61% $R_{tot}$

\*Values derived from direct measurements on root-exclusion plots.

†Where  $R_{tot}$  is 60% of measured  $CO_2$  to account for plant respiration.

following land-use systems: grassland (permanent), arable (barley), *Miscanthus*, SRC-Willow and SRF-Scots Pine.

Soil CO<sub>2</sub> fluxes under *Miscanthus* were measured at two sites, Lincolnshire and Aberystwyth. At both sites, the modelled  $R_h$  followed the same seasonal pattern of measured data (Fig. 1). At the Lincolnshire site, a statistically significant association between modelled and EC-derived  $R_h$  ( $r = 0.54$ ) was found, but a small significant bias in the model simulations when tested against the EC-derived  $R_h$  was also found (Table 4). On the other hand, the IRGA-derived  $R_h$  did not correlate well with the modelled outputs ( $r = 0.29$ ), but no bias was found in the model simulations (Table 4).

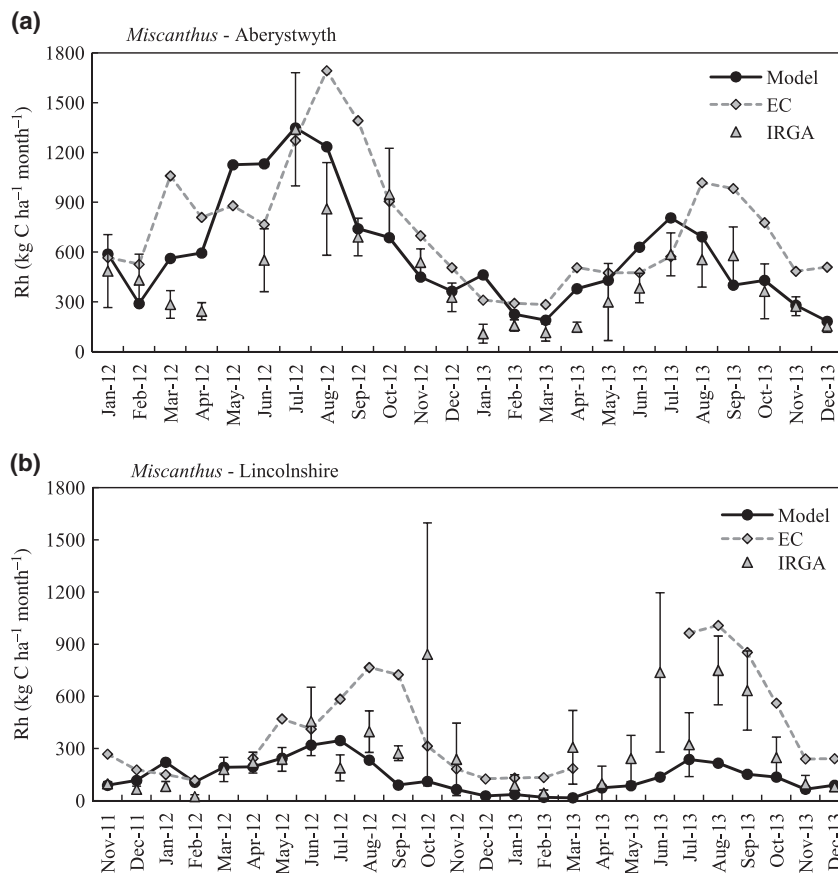
At the Aberystwyth site, significant association between modelled and measurement-derived  $R_h$  was found, regardless the type of measurement used. A slightly higher correlation coefficient was calculated correlating the modelled  $R_h$  with the EC-derived  $R_h$  ( $r = 0.70$ ) compared to the one arising from the correlation with the IRGA-derived  $R_h$  ( $r = 0.64$ ). No significant error between simulated and IRGA-derived  $R_h$

was found for this site, but a bias in the model was found when it was tested against the EC-derived  $R_h$  (Table 4).

The model performance to simulate soil CO<sub>2</sub> fluxes under SRC-Willow was tested against measurements taken at three sites: Lincolnshire, West Sussex and East Grange (Fig. 2). At all sites, a good agreement was found between simulations and measurement-derived  $R_h$  with  $r$  values ranging from 0.44 to 0.77. Also, no significant error between simulated and measurement-derived  $R_h$  was found, with the exception of the EC-derived  $R_h$  at the Lincolnshire site (Table 4).

Model performance to simulate soil CO<sub>2</sub> fluxes under SRF-Scots Pine has been evaluated against data collected at the East Grange site (Fig. 3). The modelled outputs followed the same pattern of the measured values, and the statistical analysis showed good correlation with both IRGA- and EC-derived  $R_h$ . Moreover, we found no statistically significant error between modelled and measured values as well as no bias in the model (Table 4).

Model simulations of soil  $R_h$  have also been evaluated for conventional crops (arable and grassland). Overall,



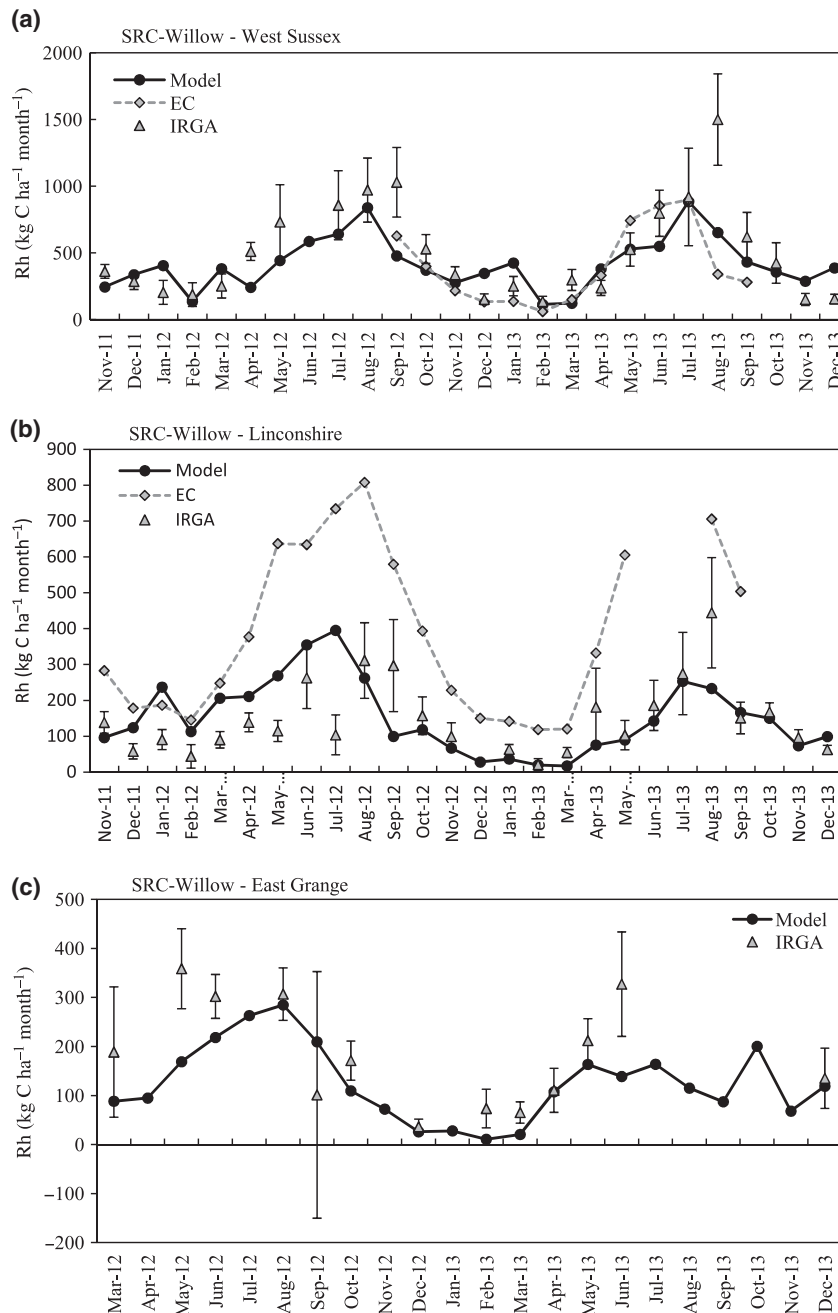
**Fig. 1** Eddy covariance derived (dotted line with diamond markers), IRGA derived (filled triangle) and modelled (solid line with circle markers) monthly heterotrophic CO<sub>2</sub> ( $R_h$ ) under *Miscanthus* plantations during the measurement period.

**Table 4** ECOSSE model performance at simulating heterotrophic respiration ( $R_h$ ) at the study sites

Land-use system	Miscanthus						SRC-Willow						SRF-Scots Pine						Grass						Arable					
	Aberystwyth		Lincolnshire		East Grange		West Sussex		East Grange		Lincolnshire		East Grange		West Sussex		Aberystwyth		East Grange		Lincolnshire		East Grange		Lincolnshire		East Grange			
Measurement type	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA	EC	IRGA				
$r$ = Correlation Coeff.	0.70	0.64	0.54	0.29	0.77	0.75	0.73	0.73	0.70	0.44	0.66	0.62	0.87	0.48	0.52	0.54	0.50	0.75	0.03											
$t$ = Student's $t$ of $r$	4.65	3.92	2.88	1.44	3.99	5.41	3.72	4.32	4.32	2.32	4.10	3.60	5.33	2.66	2.85	2.98	1.91	5.31	0.12											
$t$ -value at ( $P = 0.05$ )	2.07	2.07	2.09	2.07	2.20	2.07	2.18	2.09	2.07	2.07	2.07	2.08	2.26	2.07	2.07	2.08	2.20	2.07	2.16											
LOFIT = Lack of Fit																														
$F$	N/A	0.88	N/A	0.42	N/A	0.51	0.60	N/A	N/A	0.55	N/A	0.40	N/A	0.50	1.47	1.14	N/A	0.61	0.27											
$F$ (Critical at 5%)	N/A	1.60	N/A	1.58	N/A	1.58	1.84	N/A	N/A	1.58	N/A	1.61	N/A	1.58	1.60	1.61	N/A	1.60	1.80											
$M$ = Mean Difference (Kg C ha <sup>-1</sup> month <sup>-1</sup> )	13	-	260	-	-3	-3	-	233	-	-	-10	-	-104	-	-	-	530	-	-											
$t$ = Student's $t$ of $M$	1.89	-	4.80	-	-0.57	-0.57	-	6.14	-	3.60	-	-2.23	-	-	-	-	5.54	-	-											
$t$ -value (Critical at 2.5% - two-tailed)	2.23	-	2.09	-	2.20	2.20	-	2.09	-	2.07	-	2.26	-	-	-	-	2.20	-	-											
Number of Values	24	24	22	22	13	25	14	21	22	24	23	11	24	24	24	23	13	22	14											

Comparison of model outputs with eddy covariance (EC)-derived and IRGA-derived  $R_h$ . Association is significant for  $t > t$ -value (at  $P = 0.05$ ). Error between measured and modelled values is not significant for  $F < F$ -value (critical at 5%). Mean difference is not significant for  $t < t$ -value (Critical at 2.5% - two-tailed).



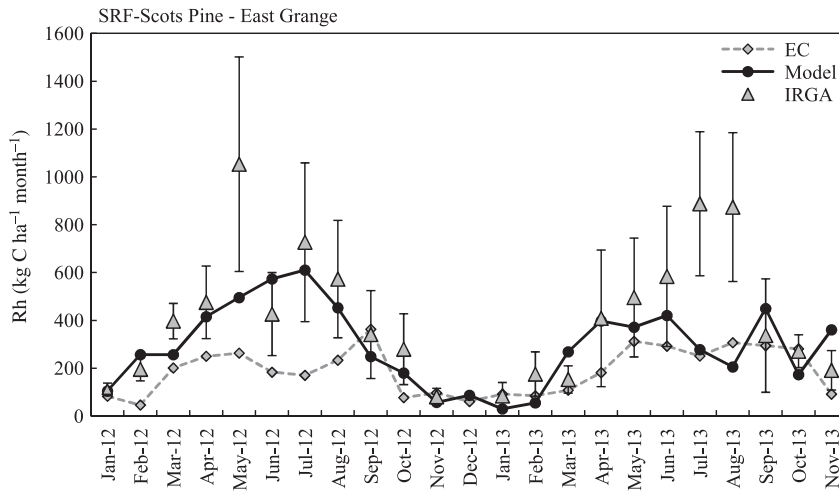


**Fig. 2** Eddy covariance derived (dotted line with diamond markers), IRGA derived (filled triangle) and modelled (solid line with circle markers) monthly heterotrophic CO<sub>2</sub> (R<sub>h</sub>) under SRC-Willow plantations during the measurement period.

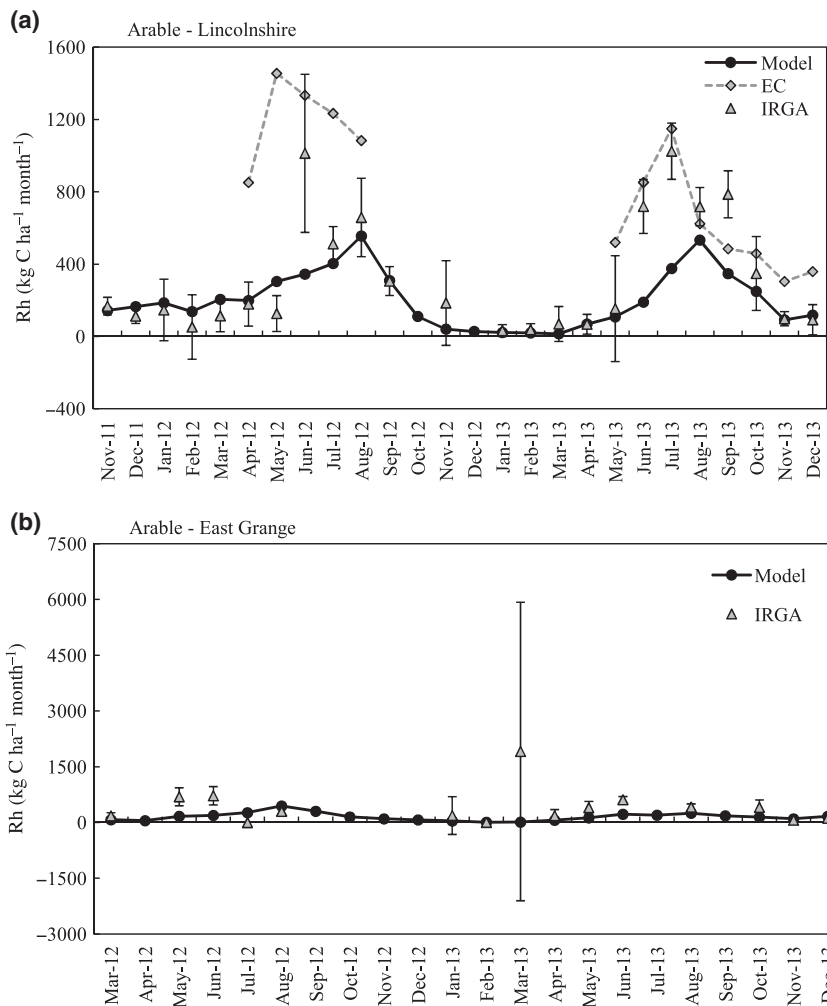
the simulated CO<sub>2</sub> follows the same pattern as the measured values at all sites (Figs 4 and 5). The statistics highlighted a significant correlation (ranging between 0.48 and 0.87 across all sites and measurements types) and no significant error between modelled and measured values as well as no model bias under perennial grass (Table 4). For the arable fields, the modelled CO<sub>2</sub> was significantly correlated to the measured value just for the IRGA-derived R<sub>h</sub> at the Lincolnshire site

( $r = 0.75$ ); however, no bias in the model was found at any site, regardless of the measurement types used for the model evaluation (Table 4).

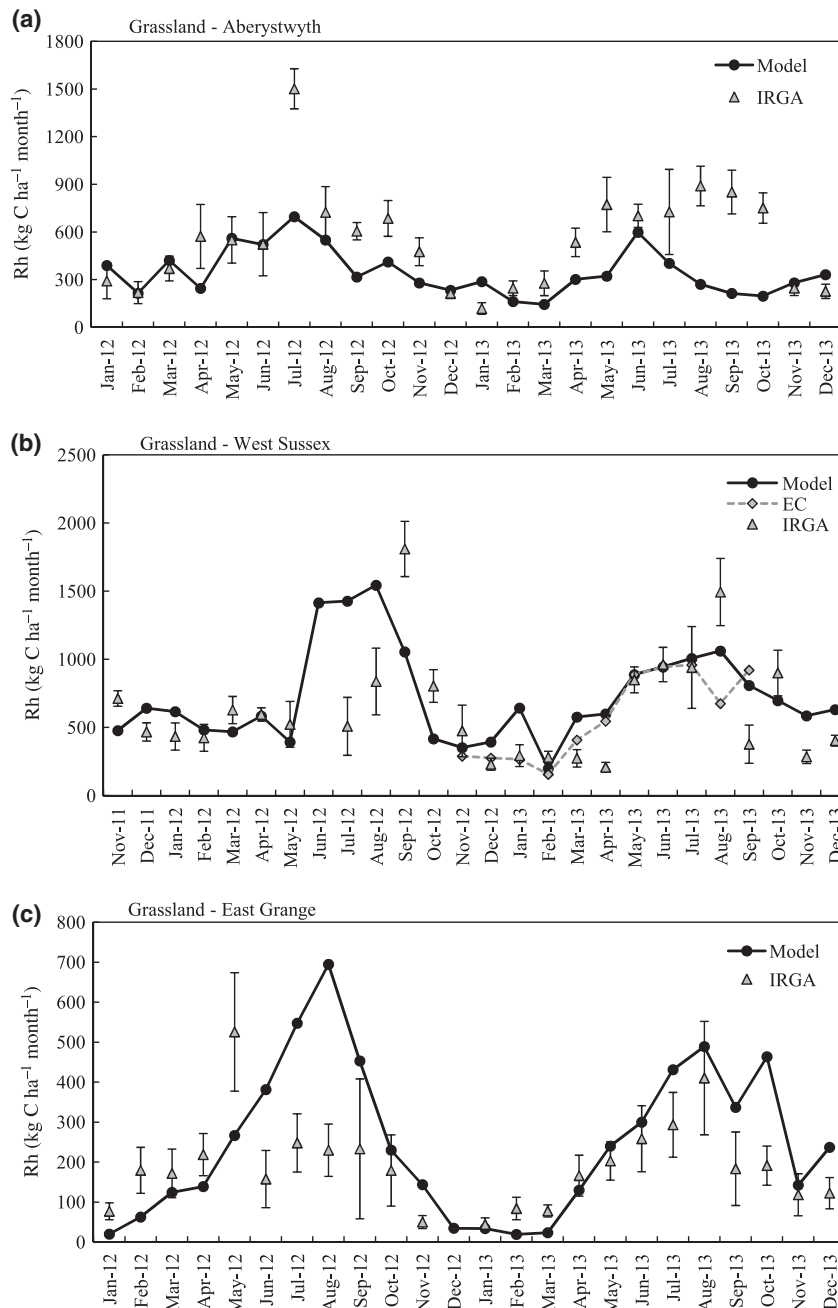
Monthly fluxes of CH<sub>4</sub> and N<sub>2</sub>O were shown to be highly variable, both spatially and temporally, across all land uses, so we present an example of the correlation between modelled and measured soil N<sub>2</sub>O and CH<sub>4</sub> fluxes for each land use. Both N<sub>2</sub>O and CH<sub>4</sub> are very small fluxes and the model outputs were within the



**Fig. 3** Eddy covariance derived (dotted line with diamond markers), IRGA derived (filled triangle) and modelled (solid line with circle markers) monthly heterotrophic CO<sub>2</sub> ( $R_h$ ) under short rotation forestry-Scots Pine plantation during the measurement period.



**Fig. 4** Eddy covariance derived (dotted line with diamond markers), IRGA derived (filled triangle) and modelled (solid line with circle markers) monthly heterotrophic CO<sub>2</sub> ( $R_h$ ) under arable plantations during the measurement period.



**Fig. 5** Eddy covariance derived (dotted line with diamond markers), IRGA derived (filled triangle) and modelled (solid line with circle markers) monthly heterotrophic  $\text{CO}_2$  ( $R_h$ ) under grassland plantation during the measurement period.

errors of the measurements, for both GHGs and at all sites (data not shown). However, low correlation between measured and modelled values has been found for the majority of the sites, ranging from  $-0.02$  to  $0.61$  for  $\text{N}_2\text{O}$  and from  $-0.29$  to  $0.53$  for  $\text{CH}_4$ . The high variability of the measured  $\text{N}_2\text{O}$  and  $\text{CH}_4$  fluxes led to a statistically significant error between simulated and measured values at most of the study sites (Tables 5 and 6).

## Discussion

Soil  $\text{CO}_2$  emissions under *Miscanthus* have been quantified at two sites (Lincolnshire and Aberystwyth) using two different sampling methods (EC and IRGA methods). At both sites, we found a high correlation between measured and modelled  $R_{h,v}$ , ranging from  $0.54$  to  $0.60$ , except for the IRGA values at Lincolnshire site ( $r = 0.29$ , Table 4). The lack of association at this site

**Table 5** ECOSSE model performance at simulating N<sub>2</sub>O fluxes at the study sites

Land-use system	Miscanthus		SRC-Willow		SRF-Scots Pine				Arable		
	Aberystwyth	Lincolnshire	Lincolnshire	East Grange	West Sussex	East Grange	West Sussex	Aberystwyth	East Grange	Lincolnshire	East Grange
$r =$ Correlation Coeff.	0.34	-0.15	-0.13	0.12	-0.02	0.19	0.25	0.06	-0.12	-0.20	0.61
$t =$ Student's $t$ of $r$	1.72	0.64	0.66	0.48	0.08	0.86	1.24	0.30	0.56	0.97	3.25
$t$ -value at ( $P = 0.05$ )	2.07	2.10	2.06	2.12	2.06	2.08	2.06	2.07	2.08	2.07	2.10
LOFIT = Lack of Fit											
$F$	0.37	3.34	54.66	22.62	0.37	40.75	0.62	0.68	312.92	0.43	0.25
$F$ (Critical at 5%)	1.63	1.69	1.59	1.74	1.59	1.63	1.59	1.62	1.63	1.60	1.69
Number of values	24	20	26	18	26	23	26	24	23	25	20

Association is significant for  $t > t$ -value (at  $P = 0.05$ ). Error between measured and modelled values is not significant for  $F < F$ -value (critical at 5%).

was mainly due to differences between modelled and IRGA-derived  $R_h$  in the year 2013 (Fig. 1b). In April 2013, the soil was harrowed and disked to break up the rhizomes for improved yield, so the system was out of balance; the farmer also applied waste wood products, which led to high CO<sub>2</sub> emissions, undetected by the model (May–August 2013 in Fig. 1b) as this was not included in the management file. In the ECOSSE model, the patterns of C and N debris return during the growing season follow a standard exponential relationship, as originally derived by Bradbury *et al.* (1993). Any alteration, such as harrowing or waste application, cannot be easily entered by the user. The scope of the present study is to evaluate the model using independent data which has not been used to develop the model. Therefore, we deliberately chose not to apply any modifications to the model to fit the measured data. However, the model was able to simulate independent data derived from two different sources with a good degree of accuracy.

Soil CO<sub>2</sub> emissions under SRC-Willow and SRF-Scots Pine plantations have been quantified using the same sampling methods. At all sites, the modelled  $R_h$  significantly correlated with all types of measurements, showing no significant error between measured and modelled values (Fig. 2).

The model has also been tested against CO<sub>2</sub> fluxes measured under conventional crops. At all three grassland sites (West Sussex, Aberystwyth and East Grange), the measured CO<sub>2</sub> fluxes correlate significantly with the modelled values and the statistical analysis showed no error between measured and modelled values, and no bias in the model (Fig. 5). This is a striking result which underlines the good quality of the data provided for the model evaluation, as well as the good model performance to simulate soil CO<sub>2</sub> fluxes.

Under grassland,  $R_h$  derived from the IRGA measurements does not always show a high correlation with the modelled values, particularly during the summer months (Fig. 5). This lack of correlation is mainly due to the difficulties in the separation of soil respiration from grassland, due to the possible inclusion of vegetation within the chamber. When deriving  $R_h$  from grassland, we estimated that 60% of the measured CO<sub>2</sub> can be attributed to plant (leaf) respiration, as reported by Byrne & Kiely (2006), but this crude estimate does not always reflect the field conditions. For an accurate quantification of the proportion of the CO<sub>2</sub> derived from the plant occluded in the chambers, field experiments would be needed to explicitly quantify plant respiration and biomass.

The analysis of the soil  $R_h$  fluxes from the arable fields reveals reasonable model performance at the Lincolnshire site, while at the East Grange site, correlation

Table 6 ECOSSE model performance at simulating CH<sub>4</sub> fluxes at the study sites

Land-use system	SRC-Willow		SRC-Scots Pine		Grass West Sussex		Arable Lincolnshire		East Grange	
	Miscanthus Aberystwyth	Lincolnshire	Lincolnshire	East Grange	West Sussex	West Sussex	Lincolnshire	Lincolnshire	East Grange	East Grange
<i>r</i> = Correlation Coeff.	0.31	0.28	0.18	0.53	0.18	0.27	0.51	-0.29	0.41	0.05
<i>t</i> = Student's <i>t</i> of <i>r</i>	1.52	1.28	0.88	2.51	0.91	1.40	2.81	1.44	1.91	0.20
<i>t</i> -value at ( <i>P</i> = 0.05)	2.07	2.09	2.07	2.12	2.06	2.06	2.07	2.07	2.10	2.10
LOFIT = Lack of Fit	0.33	3.61	6.50	0.53	0.61	0.30	0.34	0.66	4.09	0.76
<i>F</i> ( <i>F</i> Critical at 5%)	1.62	1.65	1.60	1.74	1.59	1.59	1.62	1.62	1.63	1.69
Number of values	24	22	25	18	26	26	24	24	23	20

Association is significant for *t* > *t*-value (at *P* = 0.05). Error between measured and modelled values is not significant for *F* < *F*-value (critical at 5%).

between modelled and measured IRGA values was poor (Table 4). This discrepancy between modelled and measurement-derived *R<sub>h</sub>* appears to be due to the nature of the source data; in fact, the IRGA-derived *R<sub>h</sub>* is estimated from a single data point which is taken to represent monthly CO<sub>2</sub> fluxes. Therefore, the monthly CO<sub>2</sub> flux might not be properly represented if high flux variation occurred within the month. Another explanation could also be the discontinuity of the IRGA measurements taken at the East Grange site (Fig. 4b). The latter hypothesis is supported by the *R<sub>h</sub>* results of the arable field at the Lincolnshire site. In fact, the IRGA measurements at the Lincolnshire site have been taken over a 2-year period, and the statistical analysis shows a good correlation against the model output (*r* = 0.75; Table 4). Therefore, we conclude that the low correlation at the East Grange arable field is mainly due to the variability and quantity of the measurements, and that the model accurately describes the CO<sub>2</sub> emissions from arable crop.

Generally, the model was able to predict seasonal trends in *R<sub>h</sub>* at most of the sites; however, the model occasionally over/underestimated the flux values during the warm weather in spring and summer. This is particularly evident at the Lincolnshire site, resulting in a high mean difference between modelled and EC-derived *R<sub>h</sub>* (Table 4). Despite using a generic method to estimate *R<sub>h</sub>* from *R<sub>eco</sub>*, therefore providing a challenging test for the model, we found no significant mean difference between modelled and EC-derived *R<sub>h</sub>* at three sites (for a total of four land uses), proving that the model adequately simulates soil processes under different land-use systems and climate/soil conditions.

Low correlation between measurements and model simulations arose predominantly when comparing model outputs against the IRGA-derived data set; this is mainly due to the nature of the measurements (single data point representing total monthly CO<sub>2</sub> flux), an aspect not related to the soil processes described in the model. However, it is to notice that the IRGA-derived *R<sub>h</sub>* has been estimated from direct measurements of total soil respiration and the degree of correlation between measured and modelled *R<sub>h</sub>* is also related to the *R<sub>h</sub>* : *R<sub>tot</sub>* ratio adopted. On the other hand, the EC-derived *R<sub>h</sub>* was estimated from the *R<sub>eco</sub>* during daytime, which is a modelled flux driven by air temperature and other environmental factors. Further model evaluation should be based on comparison of the model output with direct measurements of soil *R<sub>h</sub>* fluxes, possibly using automatic chambers on soil plots where roots have been excluded. This measurement technique would provide continuous *R<sub>h</sub>* measurements which would be directly comparable to the model outputs and therefore would provide a more accurate evaluation of the performance of the



model. However, given the very limited input data used to run the model and the number of sites/locations used for the model evaluation, we conclude that the simulations are robust and the model adequately simulate soil CO<sub>2</sub> fluxes under five land-use systems.

Model simulations of N<sub>2</sub>O and CH<sub>4</sub> fluxes resulted in low correlation and association at most of the study sites (Tables 5 and 6), which is expected with such low fluxes, and does not represent a failure of the model. In fact, the measured N<sub>2</sub>O and CH<sub>4</sub> fluxes are pooled from sample data points containing outliers and extreme variation between sample points in each site, which results in a high standard error of the measured values. But the N<sub>2</sub>O and CH<sub>4</sub> flux simulations are within the 95% confidence interval of the measured values, showing that the model cannot be improved to better fit these data and suggesting that the lack of correlation between modelled and measured values is due to the high variation in the measured fluxes, which is a common phenomenon verified in many N<sub>2</sub>O (e.g. Oenema *et al.*, 1997; Skiba *et al.*, 2013; Cowan *et al.*, 2015) and CH<sub>4</sub> flux measurement experiments (Parkin *et al.*, 2012; Savage *et al.*, 2014). Moreover, if the measured values do not show any seasonal trend, a significant correlation with the model outputs cannot be obtained (Smith & Smith, 2007) and low correlation is expected.

Measured fluxes of CH<sub>4</sub> were shown to be negligible across all land uses and their contribution to the total GHG balance, when converted to CO<sub>2</sub> equivalent, was on average <0.2%, except for the *Miscanthus* field at the Aberystwyth site (3% of the total GHG balance). The high mean value recorded for *Miscanthus* in 2012 is driven by one replicate with very high CH<sub>4</sub> production and there was large standard error associated with the measurements. In general, CH<sub>4</sub> production or consumption was negligible also for this field.

Across all land uses, measured fluxes of N<sub>2</sub>O represent a small proportion (<1.5%) of the total GHG balance, with the exception of the arable field at the Lincolnshire site and the *Miscanthus* field at the Aberystwyth site (6% of the total GHG balance over the 2 years measurement period at both fields). Due to technical issues and issues regarding access to sites for sampling, the data set for the arable and SRC-Willow fields at East Grange is missing a substantial number of months, and therefore, it was not possible to determine the annual GHG balance.

Despite the very low values of the CH<sub>4</sub> and N<sub>2</sub>O fluxes, and their small contribution to the total GHG balance at all experimental sites, both fluxes have been modelled adequately on a monthly time-step and no improvements can be made to the model with the available flux data.

In this study, all major GHG fluxes from five land-use systems were reasonably well estimated using the ECOSSE model. The results from this evaluation exercise show that ECOSSE is robust for simulating GHG fluxes from cropland, grassland, SRC-Willow, SRF-Scots Pine and *Miscanthus* (and transitions from the former two land uses to the latter three energy crops). This validation builds confidence that the model can be used to investigate the impacts of land-use transitions spatially in the UK and to investigate the effects of converting large areas to grow bioenergy crops.

## Acknowledgements

This work contributes to the ELUM (Ecosystem Land Use Modelling & Soil Carbon GHG Flux Trial) project, which was commissioned and funded by the Energy Technologies Institute (ETI). We acknowledge the E-OBS data set from the EU-FP6 project ENSEMBLES (<http://ensembles-eu.metoffice.com>) and the data providers in the ECA&D project (<http://www.ecad.eu>).

## References

- Abdalla M, Hastings A, Bell MJ *et al.* (2014) Simulation of CO<sub>2</sub> and attribution analysis at six European Peatland sites using the ECOSSE model. *Water, Air and Soil Pollution*, **225**, 2182.
- Aubinet M, Clement R, Elbers JE *et al.* (2003) Methodology for data acquisition, storage and treatment. In: *Fluxes of Carbon, Water and Energy of European Forests* (ed. Valentini R), pp. 9–35. Ecological Studies, Springer, Berlin, Heidelberg, Germany.
- Aubinet M, Vesala T, Papale DE (2012) *Eddy Covariance: A Practical Guide to Measurement and Data Analysis*. *Advances in Atmospheric Sciences*, pp. 2194–5217. Springer, Dordrecht.
- Bell MJ, Jones E, Smith J *et al.* (2012) Simulation of soil nitrogen, nitrous oxide emissions and mitigation scenarios at 3 European cropland sites using the ECOSSE model. *Nutrient Cycling in Agroecosystems*, **92**, 161–181.
- Boeckx P, Van Cleemput O (2001) Estimates of N<sub>2</sub>O and CH<sub>4</sub> fluxes from agricultural land in various regions of Europe. *Nutrient Cycling in Agroecosystems*, **60**, 35–47.
- Borken W, Savage K, Davidson EA, Trumbore SE (2006) Effects of experimental drought on soil respiration and radiocarbon efflux from a temperate forest soil. *Global Change Biology*, **12**, 177–193.
- Borzęcka-Walker M, Borek R, Faber A, Pudelko R, Kozyra J, Syp A, Matyka M (2013) Carbon and nitrogen balances in soil under SRC willow using DNDC model. *Journal of Food, Agriculture & Environment*, **11**, 1920–1925.
- Bradbury NJ, Whitmore AP, Hart PBS, Jenkinson DS (1993) Modelling the fate of nitrogen in crop and soil in the years following application of 15N-labelled fertilizer to winter wheat. *Journal of Agricultural Science*, **121**, 363–379.
- Bradley RI, Milne R, Bell J, Lilly A, Jordan C, Higgins A (2005) A soil carbon and land use database for the United Kingdom. *Soil Use and Management*, **21**, 363–369.
- Byrne KA, Kiely G (2006) Partitioning of respiration in an intensively managed grassland. *Plant and Soil*, **282**, 281–289.
- Case SDC, McNamara NP, Reay DS, Whitaker J (2014) Can biochar reduce soil greenhouse gas emissions from a *Miscanthus* bioenergy crop? *GCB Bioenergy*, **6**, 76–89.
- Christensen S (1983) Nitrous oxide emission from a soil under permanent grass: seasonal and diurnal fluctuations as influenced by manuring and fertilisation. *Soil Biology Biochemistry*, **15**, 531–533.
- Coleman KW, Jenkinson DS (1996) RothC-26.3 – a model for the turnover of carbon in soil. In: *Evaluation of Soil Organic Matter Models Using Existing Long-Term Datasets* (eds Powlson DS, Smith P, Smith J), pp. 237–246. Springer-Verlag, Heidelberg.
- Cowan NJ, Norman P, Farmulari D, Levy PE, Rey DS, Skiba UM (2015) Spatial variability and hotspots of soil N<sub>2</sub>O fluxes from intensively grazed grassland. *Biogeosciences*, **12**, 1585–1596.
- Crutzen PJ, Mosier AR, Smith KA, Winiwarter W (2008) N<sub>2</sub>O release from agrobiological production negates global warming reduction by replacing fossil fuels. *Atmospheric Chemistry and Physics*, **8**, 389–395.

- DECC (2012) UK Bioenergy Strategy. Department for Transport, DECC, DEFRA. Crown copyright. Available at: <https://www.gov.uk/government/publications/uk-bioenergy-strategy> (accessed 20 March 2015).
- Desjardins RL, Pattey E, Smith WN *et al.* (2010) Multiscale estimates of N<sub>2</sub>O emissions from agricultural lands. *Agricultural and Forest Meteorology*, **150**, 817–824.
- Dimitriou I, Mola-Yudego B, Aronsson P, Eriksson J (2012) Changes in organic carbon and trace elements in the soil of willow short-rotation coppice plantations. *Bioenergy Research*, **5**, 563–572.
- Dondini M, Jones EO, Richards M *et al.* (2015a) Evaluation of the ECOSSE model for simulating soil carbon under short rotation forestry energy crops in Britain. *GCB Bioenergy*, **7**, 527–540.
- Dondini M, Richards M, Pogson M *et al.* (2015b) Evaluation of the ECOSSE model for simulating soil carbon under Miscanthus and short rotation coppice – willow crops in Britain. *GCB Bioenergy*, doi:10.1111/gcbb.12286.
- Drewer J, Finch JW, Lloyd CR, Baggs EM, Skiba U (2012) How do soil emissions of N<sub>2</sub>O, CH<sub>4</sub> and CO<sub>2</sub> from perennial bioenergy crops differ from arable annual crops? *GCB Bioenergy*, **4**, 408–419.
- Drewer J, Yamulki S, Leeson SR, Convery C, Anderson M, Skiba UM, Perks MP (2015) Influence of transition from arable agricultural land to two perennial bioenergy crops short rotation coppice (SRC) willow and short rotation forest (SRF) as well as fallow on greenhouse gas fluxes in central Scotland. *GCB Bioenergy*, under review.
- EU (2009) Directive 2009/28/EC of the European Parliament and of the Council of 23 April 2009 on the promotion of the use of energy from renewable sources and amending and subsequently repealing Directives 2001/77/EC and 2003/30/EC. O. J. o. t. E. Union (ed. Union OjotE). EU, Brussels.
- Fargione J, Hill J, Tilman D, Polasky S, Hawthorne P (2008) Land clearing and the biofuel carbon debt. *Science*, **319**, 1235–1238.
- Flessa H, Beese F, Brumme R, Priesack E, Przemek E, Lay JP (1998) *Freisetzung und Verbrauch der Klimarelevanten Spurengase N<sub>2</sub>O und CH<sub>4</sub> Beim Anbau Nachwachsender Rohstoffe*. Zeller Verlag, Osnabrück, Germany.
- Gaudinski JB, Trumbore SE, Davidson EA, Zheng S (2000) Soil carbon cycling in a temperate forest: radiocarbon-based estimates of residence times, sequestration rates and partitioning of fluxes. *Biogeochemistry*, **51**, 33–69.
- Gelfand I, Zenone T, Jasrotia P, Chen J, Hamilton SK, Robertson GP (2011) Carbon debt of Conservation Reserve Program CRP grasslands converted to bioenergy production. *Proceedings of the National Academy of Sciences*, **108**, 13864–13869.
- Great Britain (2008) *Climate Change Act 2008: Elizabeth II, Chapter 27*. The Stationery Office, London.
- Grieser J, Gomme R, Bernardi M (2006) *The Miami Model of Climatic Net Primary Production of Biomass*. FAO of the UN, Italy.
- Gustavsson L, Börjesson P, Johansson B, Sverningsson P (1995) Reducing CO<sub>2</sub> emissions by substituting biomass for fossil fuels. *Energy*, **20**, 1097–1113.
- Hardie SML, Garnett MH, Fallick AE, Ostle NJ, Rowland AP (2009) Bomb <sup>14</sup>C analysis of ecosystem respiration reveals that peatland vegetation facilitates release of old carbon. *Geoderma*, **153**, 393–401.
- Harris ZM, McNamara NP, Rowe R *et al.* (2014) Research spotlight: The ELUM project: ecosystem land-use modeling and soil carbon GHG flux trial. *Biofuels*, **5**, 111–116.
- Hastings A, Wattenbach M, Eugster W, Li C, Buchmann N, Smith P (2010) Uncertainty propagation in soil greenhouse gas emission models: an experiment using the DNDC model and at the Oensingen Cropland site. *Agriculture Ecosystem & Environment*, **136**, 97–110.
- Hastings A, Tallis MJ, Casella E *et al.* (2014) The technical potential of Great Britain to produce ligno-cellulosic biomass for bioenergy in current and future climates. *GCB Bioenergy*, **6**, 108–122.
- Haylock MR, Hofstra N, Klein Tank AMG, Klok EJ, Jones PD, New M (2008) A European daily high-resolution gridded dataset of surface temperature and precipitation for 1950–2006. *Journal of Geophysical Research (Atmospheres)*, **113**, D20119.
- Heinemeyer A, Wilkinson M, Vargas R, Subke J-A, Casella E, Morison JIL, Ineson P (2012) Exploring the “overflow tap” theory: linking forest soil CO<sub>2</sub> fluxes and individual mycorrhizosphere components to photosynthesis. *Biogeosciences*, **9**, 79–95.
- Hellebrand HJ, Kern J, Scholz V (2003) Long-term studies on greenhouse gas fluxes during cultivation of energy crops on sandy soils. *Atmospheric Environment*, **37**, 1635–1644.
- Hellebrand HJ, Scholz V, Kern J (2008) Fertiliser induced nitrous oxide emissions during energy crop cultivation on loamy sand soils. *Atmospheric Environment*, **42**, 8403–8411.
- Hellebrand HJ, Strähle M, Scholz V, Kern J (2010) Soil carbon, soil nitrate, and soil emissions of nitrous oxide during cultivation of energy crops. *Nutrient Cycling in Agroecosystems*, **87**, 175–186.
- Hutchinson GL, Mosier AR (1981) Improved soil cover method for field measurement of nitrous oxide fluxes. *Soil Science Society of America Journal*, **45**, 311–316.
- Keith AM, Rowe RL, Parmar K, Perks MP, Mackie E, Dondini M, McNamara NP (2015) Implications of land-use change to Short Rotation Forestry in Great Britain for soil and biomass carbon. *GCB Bioenergy*, **7**, 541–552.
- Kern J, Hellebrand HJ, Gommel M, Ammon C, Berg W (2012) Effects of climatic factors and soil management on the methane flux in soils from annual and perennial energy crops. *Biology and Fertility of Soils*, **48**, 1–8.
- Koerber GR, Hill PW, Edwards-Jones G, Jones DL (2010) Estimating the component of soil respiration not depending on living plant roots: comparison of the indirect y-intercept regression approach and direct bare plot approach. *Soil Biology & Biochemistry*, **42**, 1835–1841.
- Lieth H (1972) Modelling the primary productivity of the world. *Nature and Resources*, UNESCO, VIII, 2, 5–10.
- Lieth H (1973) Primary production: terrestrial ecosystems. *Human Ecology*, **1**, 303–332.
- McKay H (2011) Short rotation forestry: review of growth and environmental impacts. *Forest Research Monograph*, **2**, Forest Research, Surrey. Available at: [http://www.forestry.gov.uk/pdf/frmg002\\_short\\_rotation\\_forestry.pdf](http://www.forestry.gov.uk/pdf/frmg002_short_rotation_forestry.pdf) (accessed 17 July 2015).
- McMillen RT (1988) An eddy correlation technique with extended applicability to non-simple terrain. *Boundary-Layer Meteorology*, **43**, 231–245.
- Millard P, Midwood AJ, Hunt JE, Barbour MM, Whitehead D (2010) Quantifying the contribution of soil organic matter turnover to forest soil respiration, using natural abundance  $\delta^{13}\text{C}$ . *Soil Biology & Biochemistry*, **42**, 935–943.
- Moriasi DN, Arnold JG, Van Liew MW, Bingner RL, Harmel RD, Veith TL (2007) Model evaluation guidelines for systematic quantification of accuracy in watershed simulations. *American Society of Agricultural and Biological Engineers*, **50**, 885–900.
- Norman JM, Garcia R, Verma SB (1992) Soil surface CO<sub>2</sub> fluxes and the carbon budget of a grassland. *Journal of Geophysical Research*, **97**, 18845–18853.
- Oenema O, Velthof GL, Yamulki S, Jarvis SC (1997) Nitrous oxide emissions from grazed grassland. *Soil Use Management*, **13**, 288–295.
- Pacaldo RS, Volk TA, Briggs RD, Abrahamson LP, Bevilacqua E, Fabio ES (2013) Soil CO<sub>2</sub> fluxes, temporal and spatial variations, and root respiration in shrub willow biomass crops fields along a 19-years chronosequence as affected by regrowth and removal treatments. *GCB Bioenergy*, **6**, 488–498.
- Parkin TB, Venterea RT, Hargreaves SK (2012) Calculating the detection limits of chamber-based soil greenhouse gas flux measurements. *Journal of Environmental Quality*, **41**, 705–715.
- van den Pol-van Dasselaar A, van Beusichem ML, Oenema O (1999) Effects of nitrogen input and grazing on methane fluxes of extensively and intensively managed grasslands in the Netherlands. *Biology and Fertility of Soils*, **29**, 24–30.
- Reichstein M, Falge E, Baldocchi D *et al.* (2005) On the separation of net ecosystem exchange into assimilation and ecosystem respiration: review and improved algorithm. *Global Change Biology*, **11**, 1424–1439.
- Robertson AD, Davies CA, Smith P, Dondini M, McNamara NP (2015) Modelling the carbon cycle of Miscanthus plantations: existing models and potential for their improvement. *GCB Bioenergy*, **7**, 405–421.
- Rowe R, Keith AM, Elias D, Dondini M, Smith P, Oxley J, McNamara NP (2015) Initial soil C and land use history determine soil C sequestration under bioenergy crops. *GCB Bioenergy*, (under review).
- Savage K, Phillips R, Davidson E (2014) High temporal frequency measurements of greenhouse gas emissions from soils. *Biogeosciences*, **11**, 2709–2720.
- Searchinger T, Heimlich R, Houghton RA *et al.* (2008) Use of U.S. croplands for bio-fuels increases greenhouse gases through emissions from land-use change. *Science*, **319**, 1238–1240.
- Skiba U, Hargreaves KJ, Fowler D, Smith KA (1992) Fluxes of nitric and nitrous oxides from agricultural soils in a cool temperate climate. *Atmospheric Environment*, **26**, 2477–2488.
- Skiba U, Jones SK, Drewer J *et al.* (2013) Comparison of soil greenhouse gas fluxes from extensive and intensive grazing in a temperate maritime climate. *Biogeosciences*, **10**, 1231–1241.
- Smith KA, Conen F (2004) Impacts of land management on fluxes of trace greenhouse gases. *Soil Use and Management*, **20**, 255–263.
- Smith KA, Mullins CE (eds) (2000) *Soil and Environmental Analysis: Physical Methods* (2nd edn). Marcel Dekker, Inc., New York.

- Smith JU, Smith P (2007) *Environmental Modelling. An Introduction*. Oxford University Press, Oxford.
- Smith P, Smith JU, Powlson DS *et al.* (1997) A comparison of the performance of nine soil organic matter models using datasets from seven long-term experiments. *Geoderma*, **81**, 153–225.
- Smith P, Smith JU, Wattenbach M *et al.* (2005) Projected changes in mineral soil carbon of European forests, 1990–2100. *Canadian Journal of Soil Science*, **86**, 159–169.
- Smith JU, Gottschalk P, Bellarby J *et al.* (2010a) Estimating changes in national soil carbon stocks using ECOSSE-a new model that includes upland organic soils. Part I. Model description and uncertainty in national scale simulations of Scotland. *Climate Research*, **45**, 179–192.
- Smith JU, Gottschalk P, Bellarby J *et al.* (2010b) Estimating changes in national soil carbon stocks using ECOSSE-a new model that includes upland organic soils. Part II. Application in Scotland. *Climate Research*, **45**, 193–205.
- Somerville C, Youngs H, Taylor C, Davis SC, Long SP (2010) Feedstocks for lignocellulosic biofuels. *Science*, **329**, 790–792.
- Subke J-A, Inghima I, Cotrufo F (2006) Trends and methodological impacts in soil CO<sub>2</sub> efflux partitioning: a metaanalytical review. *Global Change Biology*, **12**, 1–23.
- Thornthwaite CW (1948) An approach toward a rational classification of climate. *Geographical Review*, **38**, 55–94.
- UK (2008) *Renewable Fuels Agency, Carbon and Sustainability Reporting Within the Renewable Transport Fuel Obligation Technical Guidance Part Two Carbon Reporting, Default Values and Fuel Chains* Publisher, London, UK.
- Valentine J, Clifton-Brown J, Hastings A, Robson P, Allison G, Smith P (2012) Food vs. fuel: the use of land for lignocellulosic 'next generation' energy crops that minimize competition with primary food production. *GCB Bioenergy*, **4**, 1–19.
- Whitmore AP (1991) A method for assessing the goodness of computer simulations of soil processes. *Journal of Soil Science*, **42**, 289–299.
- Yamulki S, Anderson R, Peace A, Morison JIL (2013) Soil CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes from an afforested lowland raised peatbog in Scotland: implications for drainage and restoration. *Biogeosciences*, **10**, 1051–1065.
- Zona D, Janssens IA, Aubinet M, Gioli B, Vicca S, Fichot R, Ceulemans R (2013) Fluxes of the greenhouse gases (CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O) above a short-rotation poplar plantation after conversion from agricultural land. *Agricultural and Forest Meteorology*, **169**, 100–110.