

LONDON  
SCHOOL of  
HYGIENE  
& TROPICAL  
MEDICINE



Wassmer, S.C.; Grau, G.E. (2016) [Accepted Manuscript] Severe malaria: what's new on the pathogenesis front? *International journal for parasitology*. ISSN 0020-7519 DOI: <https://doi.org/10.1016/j.ijpara.2016.08.002> (In Press)

Downloaded from: <http://researchonline.lshtm.ac.uk/3331430/>

DOI: [10.1016/j.ijpara.2016.08.002](https://doi.org/10.1016/j.ijpara.2016.08.002)

#### Usage Guidelines

Please refer to usage guidelines at <http://researchonline.lshtm.ac.uk/policies.html> or alternatively contact [researchonline@lshtm.ac.uk](mailto:researchonline@lshtm.ac.uk).

Available under license: <http://creativecommons.org/licenses/by-nc-nd/2.5/>

1 **Invited Review**

2 **Severe malaria: what's new on the pathogenesis front?**

3

4

5 Samuel Crocodile Wassmer <sup>a,b,\*</sup>, Georges Emile Raymond Grau <sup>b</sup>

6

7

8 <sup>a</sup> *Department of Immunology and Infection, London School of Hygiene and Tropical Medicine, London, United*  
9 *Kingdom*

10 <sup>b</sup> *Vascular Immunology Unit, Department of Pathology, School of Medical Sciences & Marie Bashir Institute,*  
11 *The University of Sydney, Camperdown, Australia*

12

13 \*Corresponding author.

14 *E-mail address:* sam.wassmer@lshtm.ac.uk

15

16

17 **ABSTRACT**

18 *Plasmodium falciparum* causes the most severe and fatal form of malaria in humans with over half a million  
19 deaths each year. Cerebral malaria (CM), a complex neurological syndrome of severe falciparum malaria, is  
20 often fatal and represents a major public health burden. Despite vigorous efforts, the pathophysiology of CM  
21 remains to be elucidated, thereby hindering the development of adjunctive therapies. In recent years,  
22 multidisciplinary and collaborative approaches have led to groundbreaking progress both in the laboratory and  
23 in the field. Here we review the latest breakthroughs in severe malaria pathogenesis, with a specific focus on  
24 new pathogenetic mechanisms leading to CM. The most recent findings point towards specific parasite  
25 phenotypes targeting brain microvasculature, endothelial dysfunction and subsequent oedema-induced brain  
26 swelling.

27

28 *Keywords:* *Plasmodium* spp.; Pathophysiology; Cerebral malaria; Endothelial dysfunction; Sequestration;  
29 Malaria in pregnancy

30

31

32

## 33 1. Introduction

34 Malaria is still a leading cause of morbidity and mortality in the developing world. The virulence of  
35 *Plasmodium falciparum* is caused by several factors including parasite proteins on the surface of infected  
36 erythrocytes (IE). These allow the binding of these cells to the microvascular endothelium of various organs  
37 and tissues during infection. Proteins of the *P. falciparum* erythrocyte membrane protein 1 (PfEMP1) family  
38 mediate this adhesion through specific binding to multiple cell receptors. These include intercellular adhesion  
39 molecule-1 (ICAM-1), CD36, E-selectin, neural cell adhesion molecule (NCAM) and CD31 (PECAM-1) for  
40 endothelial beds, as well as chondroitin sulfate A (CSA) for placental syncytiotrophoblasts. Binding to  
41 endothelium results in widespread sequestration of IE, which can lead to endothelial activation as well  
42 as pro-inflammatory and pro-coagulant responses.

43 Severe falciparum malaria encompasses a broad range of diseases, the development of which may be  
44 influenced by age, exposure and immune status (Wassmer et al., 2015). It includes complications that affect  
45 specific organs such as the brain in cerebral malaria (CM) or the placenta in malaria in pregnancy (MiP).  
46 Histopathology and laboratory studies allowed investigators to establish a causal link between placenta-  
47 specific sequestration of *P. falciparum* and MiP. Indeed, the ability of PfEMP1 variants to target different  
48 receptors, the expression of which varies depending on the organ, could explain why some patients with  
49 malaria develop organ-specific syndromes. Researchers have speculated that a specific PfEMP1 variant could  
50 bind receptors that are preferentially expressed in cerebral microvasculature, and could account for the focal  
51 manifestations observed in CM, the most lethal complication of *P. falciparum* infection. Two recent reports  
52 simultaneously shed new light on the pathogenetic mechanisms leading to CM. First, endothelial protein C  
53 receptor (EPCR) was identified as a binding partner for PfEMP1. Second, normally low levels of EPCR in brain  
54 microvessels were shown to be further down-regulated in CM, with a loss of EPCR and thrombomodulin at  
55 sites of IE sequestration. These studies provided new clues towards parasite and host cell interactions leading  
56 to CM, and connected for the first time brain-specific sequestration of EPCR-binding parasites to the loss of the  
57 protein C anti-coagulant function and endothelial cytoprotective pathways (Aird et al., 2014).

58 While the relative frequency of severe malaria is low, its reported case fatality rate has not substantially  
59 changed over decades, especially for CM (Manning et al., 2014). Due to the lack of specific neuro- and  
60 vasculoprotective therapies, treatments for CM are currently still precariously limited to antimalarial drugs and

61 emergency supportive care. The former are quickly dwindling, as the resistance of *P. falciparum* malaria  
62 against artemisinin combination treatments, the recommended first-line therapy for infected patients, is on the  
63 rise in southeastern Asia. Multi-drug-resistant falciparum malaria is increasingly difficult to treat and new  
64 antimalarials are not expected to become available within the next few years. This underlines the necessity for  
65 molecular markers for surveillance of partner drug resistance, in conjunction with the implementation of new  
66 biomarkers for early diagnosis and outcome prediction, as well as effective adjunct therapies.

67 Here we review some recent data with a focus on newly developed research approaches aimed at a better  
68 understanding of the pathogenetic mechanisms of severe malaria in general and CM in particular.

## 69 70 **2. Parasite-brain microvasculature specificity in CM: a virulence factor?**

71 The severity of *P. falciparum* is linked to sequestration of IEs within the microvasculature of various organs  
72 including the brain. This sequestration is driven both by the expressed *var* gene in the parasite, leading to the  
73 expression of a specific variant of PfEMP-1, and the presence of its associated receptors on microvascular  
74 walls (Hviid and Jensen, 2015). Since there are considerable variations in both adhesion molecule expression  
75 and functional properties of endothelial cells depending on their position within the vascular bed of a particular  
76 tissue, it has been hypothesised that CM may result from a brain endothelial-specific adhesive type of parasite  
77 (Moxon et al., 2014). Indeed, IE expressing the domain cassettes (DC) 8 and 13 of the cytoadherent ligand  
78 PfEMP-1 adhere to EPCR (Turner et al., 2013). By interfering with EPCR anti-coagulant and pro-endothelial  
79 barrier functions, IE adhesion could promote coagulation and vascular permeability that contribute to the  
80 pathogenesis of CM (Moxon et al., 2014). To better understand parasite factors that contribute to disease  
81 severity, Gillrie et al. (2015) developed in vitro binding models for different microvascular beds to examine the  
82 adhesion of DC8- and DC13-expressing parasite lines to endothelial cells from different microvasculature, and  
83 the consequences of EPCR engagement on endothelial cell function. They reported that IE from IT4var19  
84 (DC8) and IT4var07 (DC13) parasite lines adhere to human brain, lung and dermal endothelial cells under  
85 shear stress. However, the relative contribution of EPCR to parasite cytoadherence on different types of  
86 endothelial cell varied.

87 Divergent functional outcomes for DC8 cysteine-rich interdomain region (CIDR)  $\alpha$ 1.1 and DC13 CIDR $\alpha$ 1.4  
88 domains were also observed. IT4var07 CIDR $\alpha$ 1.4 inhibited generation of activated protein C (APC) on lung

89 and dermal endothelial cells and blocked the APC-EPCR binding interaction on brain endothelial cells.  
90 IT4var19 CIDR $\alpha$ 1.1 inhibited thrombin-induced endothelial barrier dysfunction in lung endothelial cells,  
91 whereas IT4var07 CIDR $\alpha$ 1.4 inhibited the protective effect of APC on thrombin-induced permeability. Overall,  
92 these findings reveal a much greater complexity of how CIDR $\alpha$ 1-expressing parasites may  
93 modulate malaria pathogenesis through EPCR adhesion (Gillrie et al., 2015). DC8 PfEMP1 encode multiple  
94 endothelial binding domains, including binding activity for EPCR. These results show that PfEMP1 domains  
95 compete with protein C for EPCR binding but the extent of competition differs between domains.

96 Bernabeu et al. (2016) recently investigated these parasite virulence factors in adult patients in India and  
97 demonstrated that specific EPCR-binding parasites lead to severe malaria in that population. In addition,  
98 parasite phenotype and biomass are associated with patient hospitalization and disease severity. The authors  
99 show a broad range of EPCR binding activity from severe malaria isolates and even parasite domains that  
100 partially obstructed the interaction between EPCR and APC were sufficient to interfere with the cytoprotective  
101 functions of APC (Bernabeu et al., 2016). Taken together, their findings suggest that parasites may be under  
102 selection for phenotypic variation in a key host pathway that regulates coagulation and endothelial barrier  
103 properties, and has important implications for pathogenic mechanisms in severe malaria. Full-length  
104 sequences of PfEMP1 encoding transcripts were characterized in clinical isolates from children with severe  
105 malaria admitted to hospital in Tanzania, and EPCR-binding CIDR $\alpha$ 1 domains dominated PfEMP1 transcript  
106 profiles of children suffering from CM and/or severe malarial anaemia, further strengthening the evidence for a  
107 crucial pathogenic role of the PfEMP1–EPCR interaction in severe malaria (Jespersen et al., 2016).

108 EPCR is not the only receptor suspected to play an important role in the development of severe malaria.  
109 Studies of parasite isolates have demonstrated high rates of in vitro ICAM-1 binding among wild strains but  
110 reported correlations between ICAM-1 binding and disease severity have been inconsistent. Recent results  
111 suggest that ICAM-1 is a co-receptor for a subset of EPCR-binding parasites (Avril et al., 2016). Indeed, Avril  
112 et al. (2016) showed that parasites expressing DC13 have dual binding specificity for EPCR and ICAM-1,  
113 suggesting for the first time that ICAM-1-binding variants can be grouped into CD36 and EPCR co-receptor-  
114 binding traits. This leads to different cytoadherence abilities on TNF-stimulated endothelial cells, which has  
115 important implications for understanding parasite organ-specific microvascular bed tropism in pro-inflammatory  
116 conditions.

117  
118  
119  
120  
121  
122  
123  
124  
125  
126  
127  
128  
129  
130  
131  
132  
133  
134  
135  
136  
137  
138  
139  
140  
141  
142  
143  
144

### 3. Microvascular endothelial dysfunction: new causes and repercussions

In the recent years, convincing evidence has been presented to support the role of both endothelial cell activation and platelets in modulating the pathogenesis of severe *P. falciparum* malaria. Thrombin, a common factor in both processes, is now thought to be a driver of pathology in CM. The relative contribution of EPCR-binding parasites versus loss of EPCR from the endothelial surface in mediating CM is not known, although both are associated with disease in clinical studies (Turner et al., 2013; Moxon et al., 2014). The key mediator of inflammation in both cases appears to be thrombin, which is a potent mediator of both pro-and anti-inflammatory pathways, depending on the context of endothelial signalling. Thrombin engages thrombomodulin on the plasma membrane of intact endothelium, where it promotes activation of protein C. The latter is accelerated by the presence of EPCR, which binds to protein C and presents it for optimal activation by the thrombin-thrombomodulin complex. Further to its anticoagulant activity, generated APC can also trigger numerous cell-signaling pathways initiating protective cellular responses upon exposure to pro-inflammatory, pro-apoptotic, or toxic insult. IEs expressing PfEMP1 compete with protein C and APC for EPCR, thereby down-regulating protein C activation by the thrombin-thrombomodulin complex. The resulting loss of EPCR-APC-induced cytoprotective signalling is consistent with a decrease in blood-brain barrier (BBB) properties, potentially leading to vasogenic oedema in CM.

Direct protein C pathway alteration by binding of EPCR-specific IEs is not the only cause of endothelial dysfunction in CM. Indeed, *P. falciparum* infection also initiates early endothelial and platelet activation, leading to coagulation dysregulation and microvascular lesions locally (Wassmer et al., 2011a). Furthermore, IEs elicit a myriad of signalling pathways leading to aberrant pro-coagulant effects, ultimately resulting in enhanced endothelial activation, damage and apoptosis (O'Sullivan et al., 2016). Lastly, severe falciparum malaria has been associated with a down-regulation of normal endogenous anticoagulant pathways. EC surface expression of thrombomodulin and EPCR are both reduced, likely through cytokine-enhanced shedding, leading to an increase of their soluble levels in plasma (Moxon et al., 2013). Together, these effects combine and lead to a significant reduction in generation of anti-inflammatory and cytoprotective APC on the endothelial surface. These findings suggest new avenues for acute therapeutic intervention and match well with post-mortem observations and magnetic resonance imaging (MRI) findings linked to mortality in CM.

Such an endothelial dysfunction is not only observed in severe falciparum malaria, but was also reported in severe *Plasmodium vivax* (Barber et al., 2015) and *Plasmodium knowlesi* (Yeo et al., 2007) cases, which raises the question of possible common pathogenic pathways in these various types of infection. All plasmodia species can cause severe and fatal malaria. In falciparum malaria, the most common cause of severe malaria, impaired tissue perfusion has long been recognised to arise from microvascular obstruction by IE adherent to endothelial cells. More recently recognised is the contribution of concurrent impairment of nitric oxide (NO) bioavailability, endothelial activation and microvascular dysfunction to impaired tissue perfusion and severe disease (Yeo et al., 2014). Angiopoietin-2, released from endothelial cell Weibel-Palade bodies and an NO-inhibited autocrine mediator of endothelial activation, is markedly elevated in severe falciparum malaria and consistently associated with impaired tissue perfusion and fatal outcome in both adult and paediatric severe malaria. This is independent of both total and sequestered parasite biomass, suggesting that microvascular obstruction and microvascular dysfunction make separate contributions to pathogenesis. Microvascular function, the capacity to increase flow and oxygen delivery in response to ischaemia, is decreased in severe falciparum malaria and associated with an increased risk of death. Endothelial activation, decreased endothelial NO bioavailability and microvascular dysfunction are also associated with impaired tissue perfusion in vivax and knowlesi malaria, a factor likely to contribute to organ dysfunction and severe malaria caused by infection with these species. Initial clinical trials of agents aimed at restoring NO bioavailability in severe falciparum malaria have recently been completed (Hawkes et al., 2011; Serghides et al., 2011), and microvascular function can be improved by L-arginine infusion in severe falciparum malaria (Yeo et al., 2013). These results suggest that new compounds aimed at increasing microvascular NO and microvascular functions have potential clinical benefits as adjunctive treatments in severe malaria from all species.

#### **4. Rosetting and clumping: consequences for sequestration and microvascular obstruction**

Parasite adhesion interactions in severe falciparum malaria are not restricted to the endothelium. Indeed, IEs can bind uninfected erythrocytes to form rosettes (Handunnetti et al., 1989), or platelets to form clumps (Pain et al., 2001). Both processes have been associated with severe malaria (Rowe et al., 1995; Pain et al., 2001) and CM (Carlson et al., 1990; Wassmer et al., 2008).



172 In this context, recent advances have been made in elucidating the molecular mechanisms underlying  
173 rosetting, with a view to development of rosette-reversing therapies. Rosetting generally requires the presence  
174 of soluble serum factors such as IgM, and the ability of IE to form rosettes and to bind non-immune IgM is  
175 correlated (Rowe et al., 2002). While the role of IgM binding in rosetting remains unclear, it appears to  
176 strengthen the bond of the central IE with the surrounding erythrocytes. Stevenson et al. (2015) recently  
177 reported that the serum protein  $\alpha_2$ -macroglobulin ( $\alpha_2$ M) is able to induce rosetting in vitro and ex vivo, using  
178 several parasite isolates. In contrast to IgM,  $\alpha_2$ M elicits rosetting alone, while the presence of IgM significantly  
179 lowers the concentration of  $\alpha_2$ M required. The authors of the study postulate that  $\alpha_2$ M allows the crosslinking of  
180 several individual PfEMP1 molecules, thereby increasing their combined avidity for carbohydrate receptors on  
181 surrounding erythrocytes and promoting the formation of rosettes. These results suggest that  
182 *P. falciparum* hijacks soluble host proteins for its own benefit, and avoids immune destruction by manipulating  
183 those to facilitate adhesion of IEs to low-affinity carbohydrate receptors. In addition, another report recently  
184 showed that PfEMP1 is not the only parasite ligand used in rosetting, potentially opening new rosette-  
185 disrupting approaches. Goel et al. (2015) showed in a very elegant study that the repetitive interspersed family  
186 of proteins (RIFINs) mediates binding of IEs with a preference for blood group A, leading to the formation of  
187 large rosettes of 10 or more IEs. This was not observed with IEs from group O. Indeed, blood group O is  
188 common in malaria endemic areas and IEs of that blood group were shown to form small, weak rosettes (Rowe  
189 et al., 2007). The role of RIFINs was confirmed by the disruption of rosettes in the presence of anti-RIFIN  
190 antibodies. These results suggest that RIFINs not only play a fundamental role in the development of severe  
191 malaria but also contribute to the virulence of *P. falciparum*.

192 Puzzlingly, formed rosettes have never been described in the peripheral bloodstream, suggesting that they  
193 sequester in the microvasculature and aggravate microvascular obstructions during severe malaria in general,  
194 and CM in particular. Understanding the mechanisms by which rosettes sequester (i.e., by direct binding of  
195 rosetting IE to endothelial cells, to platelets on endothelial cells or to non-rosetting cytoadherent IE is  
196 fundamental, as it may open new adjunct therapies to reduce the biomass of sequestered IE, a parameter  
197 associated with disease severity (Hendriksen et al., 2012). IE of the IT/R29 strain expressing a rosette-  
198 mediating PfEMP1 variant (IT4var09) were recently shown to cytoadhere to human brain microvascular  
199 endothelial cells using heparan sulfate proteoglycans as ligands (Adams et al., 2014). This process is distinct

200 from rosetting, which is primarily mediated by interactions with between the NTS-DBL1 $\alpha$  domain of PfEMP1  
201 and complement receptor 1. This study shows for the first time that IT4var09-expressing parasites are capable  
202 of dual interactions with both endothelial cells and uninfected erythrocytes via distinct receptor-ligand  
203 interactions, and therefore could aggravate microvascular obstruction in severe malaria by facilitating the  
204 sequestration of platelet-mediated clumps.

205 One of the most recently described *P. falciparum* cytoadherence phenotypes is the ability of IEs to bind to  
206 platelets in suspension in vitro to form platelet-mediated clumps. Similar to rosetting, the formation of clumps is  
207 a common phenotype and has been reported in a variety of endemic settings (Rowe et al., 2009). This is  
208 mediated by the binding of IEs to platelet receptors which include CD36 (Pain et al., 2001), P-selectin  
209 (Wassmer et al., 2008) and globular C1q receptor (gC1qR) (Biswas et al., 2007). In all cases, the parasite  
210 ligands are unknown, although PfEMP1 is a likely candidate molecule. A recent study performed using clinical  
211 isolates from Mozambican patients to evaluate cytoadherence properties such as platelet-mediated clumping,  
212 rosetting and adhesion to purified receptors (CD36, ICAM1 and gC1qR) revealed that, compared with matched  
213 controls, prevalence of both rosetting and platelet-mediated clumping and adhesion to gC1qR was higher in  
214 severe cases (Mayor et al., 2011). Inhibition of these cytoadherence phenotypes may therefore reduce the  
215 occurrence or improve the prognosis of severe malaria outcomes. Similar to rosettes, platelet-mediated clumps  
216 have not been observed in the bloodstream and it is likely that they sequester in the microvasculature using IE  
217 or platelet receptors (or both) to cytoadhere on endothelial cells (Wassmer et al., 2011a). Since this may  
218 potentially aggravate microvascular obstruction in CM, further studies aimed at understanding the molecular  
219 process involved in their sequestration are warranted.

## 221 **5. Clinical consequences of BBB opening in CM: vasogenic oedema and brain swelling**

222 Following the report by Seydel et al. (2015), a study was initiated to investigate the different mechanisms  
223 potentially responsible for brain swelling in both pediatric and adult CM patients in India (Wassmer et al.,  
224 2015). Brain swelling was identified by MRI in over 50% of patients enrolled in the ongoing study, irrespective  
225 of their age group. The frequent occurrence of brain swelling in CM has been previously reported in separate  
226 studies on Indian adults using computed tomography (CT) (Mohanty et al., 2011), as well as in Bangladeshi  
227 adults (Maude et al., 2014) and Malawian children using MRI (Potchen et al., 2012). The cause of the swelling,

228 which can lead to brain herniation in fatal cases (Seydel et al., 2015), remains to be identified and may be due  
229 to cytotoxic or vasogenic causes. Recent results show that patients who develop CM exhibit an over-reaction  
230 of their endothelial cells to systemic inflammation, which is not observed in uncomplicated malaria (Wassmer  
231 et al., 2011b). Such vulnerability, coupled to a loss of cytoprotective EPCR in the brain, could lead to a  
232 disruption of the BBB and the leakage of fluids and proteins from the vascular system into  
233 the extracellular space. Also called vasogenic oedema, this process is consistent with the description of ring  
234 haemorrhages, a characteristic sign of BBB disruption, in pediatric CM (Taylor et al., 2004; Dorovini-Zis et al.,  
235 2011). In addition, these ring hemorrhages correlate positively between retinal and cerebral tissues, owing to  
236 their common embryological origin from the neuroectoderm (Barrera et al., 2015; Greiner et al., 2015). Malaria  
237 retinopathies seen in pediatric CM are therefore a direct reflection of the neurovascular disease process, and  
238 the high frequency of ring haemorrhages both in the brain and in the retina in fatal disease point towards the  
239 occurrence of vasogenic oedema. However, evidence for a generalised increase in BBB permeability leading  
240 to vasogenic oedema is still debated. A study performed in Thailand showed no evidence of BBB impairment,  
241 as radioactively-labelled albumin given intravenously was not found in the cerebrospinal fluid (CSF) of CM  
242 patients during coma (Warrell et al., 1986). Additionally, post-mortem analyses of adult Vietnamese patients  
243 who died of CM showed that its pathophysiology only involved subtle functional changes in BBB integrity  
244 (Brown et al., 2000), and evidence from a different cohort demonstrated that localised loss of vascular integrity  
245 did not correlate with the occurrence of pre-mortem coma (Medana et al., 2011). Brain swelling may also occur  
246 as a result of cytotoxic oedema, which can result from cells in the cerebral tissue being unable to maintain  
247 membrane potential after the failure of  $\text{Na}^+/\text{K}^+$  ATP-dependent pumps, due to hypoxia or nutrient deprivation.  
248 This may cause a redistribution of water from the extra-cellular to the intracellular compartments, ultimately  
249 leading to cell swelling, cell death and tissue damage. Ischaemic or hypoxic insults due to mechanical effects  
250 of microvascular obstruction by IEs, rosettes and platelet-mediated clumps, as well as nutrient “steal” by local  
251 metabolically active parasites could account for cytotoxic oedema in CM. This mechanism is consistent with  
252 abundant sequestration in the cerebral microvasculature during the neurologic syndrome (White et al., 2013),  
253 and the first in vivo magnetic resonance study of experimental CM revealed a preponderant role for cytotoxic  
254 oedema in fatal outcome (Penet et al., 2005).

255 An increase in intravascular fluid volume within the brain due to sludging of blood flow with sequestration of

256 IEs, rosettes and platelet-mediated clumps could also cause brain swelling and would explain the diffuse mild  
257 brain swelling reported in severe malaria patients from Bangladesh (Maude et al., 2014), without evidence of  
258 either cytotoxic or vasogenic oedema. However, cytoadherence itself may not be directly or solely responsible  
259 for this clinical syndrome (Storm and Craig, 2014).

260 Lastly, it is entirely plausible that both mechanisms are not mutually exclusive. A study in live mice using  
261 high-field MRI with whole-brain coverage showed that vasogenic oedema occurs first in infected animals, and  
262 starts in the olfactory bulb before spreading deeper into the brain along a specific path called the rostral  
263 migratory stream, eventually reaching the brain stem. Microvascular pathology and ischemic brain injury  
264 develop only secondarily, after vasogenic oedema formation (Hoffmann et al., 2016). Additional MRI studies  
265 are currently underway in CM patients from Asia and Africa, and are aiming to elucidate these pathogenetic  
266 processes (Wassmer et al., 2015).

## 268 **6. New adjunct therapies and critical care approaches in severe malaria**

269 Even under optimal conditions, the case–fatality rate in severe malaria treated with either artemisinin  
270 derivatives or quinine remains high. In addition, multi-drug-resistant falciparum malaria is increasingly difficult  
271 to treat and new effective antimalarial agents are not expected to become available within the next few years.  
272 In an effort to reduce malaria-related mortality, numerous adjunctive therapies that may alter severe malaria-  
273 induced physiological abnormalities are being evaluated, some of which have been described in other sections  
274 of this review. Such therapies are, in nature, highly specific to distinct syndromes of severe malaria, as their  
275 aim is to target precise pathophysiological processes. For example, protein C system defects inflicted by  
276 the malaria parasite protein PfEMP1 in CM can be overcome by a soluble EPCR variant (Petersen et al.,  
277 2015), and could therefore represent a revolutionary approach to dampen the pathogenetic mechanisms and  
278 decrease mortality in affected patients. Promising advances in adjunct therapies for other severe malaria  
279 syndromes such as acute respiratory distress, acute kidney injury, severe anaemia and metabolic acidosis are  
280 still lacking.

281 The potential clinical benefit of fluid resuscitation was recently investigated and liberal fluid loading in adults  
282 with severe malaria showed no improvement in the acidosis and acute kidney injury, but increased the  
283 incidence of acute pulmonary oedema (Hanson et al., 2014). However, a more conservative fluid strategy,

284 using a simple weight-based algorithm, led to a low incidence of acute respiratory distress without significant  
285 deterioration in acid-base status, renal function, electrolyte profile or systemic haemodynamics, and was  
286 associated with increased survival (Aung et al., 2015).

287 In addition to *P. falciparum*, severe cases of *P. vivax* and *P. knowlesi* have also been reported, although  
288 sequestration is not a specific feature of either infection and the relative contribution of co-morbidities to clinical  
289 manifestations, particularly in vivax malaria, remains to be investigated (Wassmer et al., 2015). Both parasitic  
290 infections can cause acute pulmonary oedema with a clinical phenotype similar to that seen in severe  
291 falciparum malaria. No studies of fluid resuscitation have been performed in these patients, but those are  
292 warranted.

## 293

### 294 **7. Novel parasite factors involved in malarial pathogenesis and possible therapeutic targets**

295 In parallel to the development of adjunct therapies, a growing effort in the search for new and effective  
296 pharmacotherapies has been triggered by the emergence of multi-drug resistance in *P. falciparum*. While still  
297 in their infancy, these approaches are promising and provide a wide range of new therapeutic targets. Several  
298 new parasite factors were recently identified and have emerged as potential drug target candidates. Among  
299 those, G-quadruplex (G4) DNA motifs and RecQ helicases are newly described players in virulence gene  
300 control in *P. falciparum*. G4s are four-stranded structures formed by the stacking of quartets of guanines, and  
301 recent work has shown that they can form in vivo as well as in vitro, affecting DNA replication, transcription,  
302 translation and telomere maintenance. Harris and Merrick (2015) first demonstrated that DNA helicases, as  
303 well as histone-modifying proteins, can influence *var* gene dynamics in *P. falciparum*. Understanding the G4-  
304 mediated regulation of the parasite virulence may open the door to novel therapeutic interventions.

305 *Plasmodium falciparum* exports parasite-encoded proteins involved in structural and functional remodelling  
306 of the host cell. This process is essential for the development of the parasite and is also associated with its  
307 virulence (Maier et al., 2009). Molecular chaperones of the heat shock protein (Hsp) family are prominent  
308 members of the exportome, including various Hsp40s and one Hsp70. The first biochemical evidence for a  
309 specific functional co-chaperone interaction between the exported malarial PFA0660w and PfHsp70-x was  
310 recently reported (Daniyan et al., 2016). PFA0660w can stimulate the ATPase activity of PfHsp70-x and work  
311 additively with it in a co-chaperone/chaperone interaction, resulting in protein aggregation suppression. The

312 authors also showed that PFA0660w could potentially act independently as a chaperone. These findings  
313 support the proposed role of PfHsp70-x and PFA0660w in parasite protein trafficking and folding in IEs. Further  
314 studies are underway to determine the molecular basis for the specificity of this interaction, and to identify  
315 inhibitors capable of disrupting it. All of the modifications to the IEs are induced by parasite proteins, most of  
316 which initially traffic from the parasite via the secretory pathway to the parasitophorous vacuole. There,  
317 proteins interact with a translocon complex called PTEX (*Plasmodium* translocon of exported proteins) that  
318 transports them across the vacuole membrane into the IE. Blocking protein export through blocking PTEX  
319 function leads to the arrest of parasite growth and loss of virulence (Kalanon et al., 2016), making PTEX an  
320 excellent drug target (Gilson et al., 2016).

321 Post-translational modification of histones is one of the key gene regulation mechanisms during the intra-  
322 erythrocyte development cycle of *P. falciparum*. Studies of proteins, which recognise and interact with histone  
323 post-translational modifications, are pivotal for understanding *P. falciparum* pathogenesis. Bromodomain  
324 proteins bind to acetylated lysines, often on histones, and frequently play a role in regulation of gene  
325 expression. *Plasmodium falciparum*-specific bromodomain protein 1 (PfBDP1) binds to chromatin at  
326 transcriptional start sites of invasion-related genes and directly controls their expression. Conditional PfBDP1  
327 knockdown causes a significant defect in parasite invasion and growth (Josling et al., 2015). In parallel to these  
328 studies, several small molecule inhibitors have recently been reported to have a high affinity and specificity to  
329 bromodomains, and could represent a new therapeutic avenue in *P. falciparum* infection (Padmanabhan et al.,  
330 2016).

331 The unique plasticity of the epigenetic regulation in *P. falciparum* has also emerged as a pivotal virulence  
332 and pathogenicity factor in recent years. Karmodyia et al. (2015) performed genome-wide mapping of multiple  
333 histone modifications of *P. falciparum* and reported H3K36me2 as a global repressive mark, with gene  
334 regulation being fine-tuned by the ratio of activation marks to H3K36me2 (Karmodyia et al., 2015).  
335 Moreover, *var* genes are mostly poised and marked by a unique set of activation (H4ac) and repression  
336 (H3K9me3) marks, which are mutually exclusive to other *P. falciparum* housekeeping genes. A better  
337 characterization of epigenetic regulation in *P. falciparum* will lead to the identification of potential therapeutic  
338 targets (Ay et al., 2015). Indeed, disrupting the function of proteins responsible for maintaining  
339 heterochromatin, such as HP1 (Brancucci et al., 2014), could be an effective strategy to block parasite

340 replication during the asexual cycle. PfAP2-G, a transcription factor shown to drive gametocytogenesis, also  
341 represents a promising target to disrupt malaria transmission (Coleman et al., 2014). Extensive  
342 characterisations of the parasite epigenetic factors, as well as its post-transcriptional and translational control  
343 processes, are likely to open new avenues for drug development against *P. falciparum*.

## 345 **8. Pathologies other than CM: MiP**

346 *Plasmodium falciparum* infection during pregnancy can result in ~~MiP~~, a pathology ~~resulting from~~caused by  
347 the accumulation of IEs in the placental intervillous space and the infiltration of maternal  
348 monocytes/macrophages (Rogerson et al., 2003), with detrimental outcomes for both the mother and the  
349 foetus. Expression of PfEMP1-var2csa at the surface of IEs mediates their adhesion to the placenta. Adaptive  
350 immunity is progressively acquired during sequential malaria infections in pregnancy and is mediated by the  
351 production of anti-VAR2CSA antibodies, which promote IE adhesion blocking and opsonisation (Desai et al.,  
352 2007). This naturally acquired immunity is the key basis for development of a vaccine to protect women during  
353 pregnancy, and VAR2CSA is currently the leading candidate (Fried and Duffy, 2015). However, recent findings  
354 indicate that broadly neutralising antibodies of multigravidae are not depleted on VAR2CSA recombinant  
355 antigens. Using a new approach to assess VAR2CSA domains for functional epitopes recognized by naturally  
356 acquired antibodies, Doritchamou et al. (2016) recently demonstrated that different Duffy binding-like (DBL)  
357 domain-specific IgG could react to both homologous as well as heterologous antigens and parasites,  
358 suggesting that conserved epitopes are shared between allelic variants. In addition, IE binding was blocked by  
359 ID1-DBL2-ID2a, DBL4 and DBL5-specific IgG, while partial cross-inhibition activity was observed with purified  
360 IgG specific to ID1-DBL2-ID2a and DBL4 antigens. Interestingly, plasma from patients still showed broadly  
361 adherence-blocking activity after complete depletion of these VAR2CSA specificities. These results suggest  
362 that VAR2CSA vaccines based on a single construct and variant might induce antibodies with limited broad  
363 blocking activity, and confirm that a multivalent vaccine comprised of up to five different alleles or with the  
364 addition of multiple placental malaria vaccine candidates may be needed to elicit the broad blocking activity  
365 observed in African multigravidae (Avril et al., 2010; Hommel et al., 2010).

366 The generation of protective vaccines is becoming a priority, especially in areas where the prevalence of  
367 malaria has decreased due to control and elimination campaigns. Indeed, a recent study conducted in

368 Mozambique showed a close relationship between antibody levels and the intensity of malaria transmission.  
369 Mayor et al. (2015) showed convincing evidence that a decline in the prevalence of malaria documented in the  
370 study area was accompanied by reductions in levels of IgG antibodies not only against VAR2CSA, but also  
371 against non-pregnancy-specific malaria antigens. In pregnant women with MiP, this was associated with an  
372 increase in parasite densities and a higher adverse effect of *P. falciparum* infection on maternal haemoglobin  
373 levels and newborn weights (Mayor et al., 2015). Although they also suggest that immunity may be regained as  
374 exposure increases, the findings of this study indicate that malaria control and elimination programmes could  
375 precede a resurgence of pregnancy-associated malaria pathologies.

376 Additionally, MiP has also recently been linked to placental pathology in a low malaria transmission area in  
377 Brazil, where *P. vivax* is predominant (Souza et al., 2013). While MiP was not associated with severe outcome  
378 in a second study performed by the same team in this region, an increased ratio of peripheral receptor tyrosine  
379 kinase Tie-2 to angiopoietin (Ang-1) was associated with the occurrence of MiP. Both Ang-1 and Ang-2 had  
380 similar magnitudes but inverse associations with placental barrier thickness. MiP is an effect modifier of the  
381 association between Ang-1 and placental barrier thickness (Ataide et al., 2015). These findings provide a  
382 possible pathway through which placental pathological changes occur during MiP.

383 Lastly, further to imbalances in cytokine cascades, IE cytoadhesion and angiogenic dysregulation,  
384 excessive or dysregulated complement activation as part of the host innate immune response to malaria  
385 infection can also exacerbate the severity of MiP, leading to poor pregnancy outcomes (McDonald et al.,  
386 2015b). Using an experimental model of MiP in conjunction with micro-CT and HPLC analysis of  
387 neurotransmitter levels, McDonald et al. (2015a) showed that complement activation, in particular C5a,  
388 contributes to foetal neuropathologic outcomes during MiP. The offspring of infected animals showed  
389 persistent neurocognitive deficits in memory and affective-like behaviour compared with unexposed controls  
390 (McDonald et al., 2015a). These impairments were linked with decreased tissue levels of neurotransmitters in  
391 regions of the brain associated with the deficits. The inhibition of maternal C5a complement receptor signaling  
392 restored the levels of neurotransmitters and reversed the associated phenotype, suggesting new targets for  
393 intervention in MiP aimed at decreasing foetal neuropathologic outcomes.

## 394 395 **9. New investigative tools and experimental models**



396 A vast array of new tools and models has recently become available to facilitate the investigation of severe  
397 malaria pathogenesis, with a particular focus on CM. These are detailed elsewhere (Sahu et al., 2015). Further  
398 to this, the rise of the 'omic' era during the past decade has provided the malaria research community with  
399 unprecedented approaches and technologies to better understand the biology, evolution and pathogenesis of  
400 different *Plasmodium* spp. The Malaria Host-Pathogen Interaction Centre (MaHPIC) is a large international  
401 systems biology consortium developed in 2012 and based in the USA  
402 (<http://www.systemsbiology.emory.edu/index.html>), which allowed the development of a variety of distinctive  
403 hypothesis-generating and hypothesis-driven collaborations with scientific teams in malaria-endemic regions.  
404 These collaborations involve investigators from several countries in South America, southeastern Asia and  
405 sub-Saharan Africa, with a focus on infections caused by the predominant, less predominant, or mixed species  
406 of *Plasmodium* (Anderson et al., 2015; Lapp et al., 2015), to study disease states, pathogenesis and  
407 physiological or immunobiological questions through the use of untargeted high-resolution metabolomics, as  
408 well as clinical and demographic metadata (Salinas et al., 2014). In addition, the MaHPIC team is intensively  
409 studying malaria using non-human primate model systems (macaque and New World monkey species, (Joyner  
410 et al., 2015)). An overarching goal, beyond the team's specific research quests, is to develop and make  
411 available unique large 'omic' datasets (e.g., transcripts, proteins, lipids, metabolites and immune responses),  
412 integrated models and tools for the use and benefit of the research community at large. This unique and  
413 integrated combination of 'omic' approaches represents a revolutionary platform to identify, develop and  
414 evaluate new diagnostic tools, antimalarial drugs and vaccines for different types of malaria parasites.

415 In parallel, live imaging, including multi-photon approaches, has contributed to the understanding of CM  
416 pathogenesis, notably its haemodynamics (Nacer et al., 2014) and its immune cell mobilisation (Pai et al.,  
417 2014) components. Recently, in experimental CM, vibrational spectroscopies provided evidence of  
418 peroxidative stress and protein oxidation within cerebellar gray matter, which were co-localised with elevated  
419 non-haeme iron at the site of microhaemorrhages. A novel combination of chemical probe-free, multimodal  
420 imaging to quantify molecular markers of disturbed energy metabolism and peroxidative stress thus provides  
421 new insights into understanding CM pathogenesis (Hackett et al., 2015).

## 423 **10. Conclusions and future perspectives**

424 Despite the recent leap in our understanding of pathogenetic mechanisms leading to severe malaria, the  
425 translational outputs to improve the clinical outcome of patients remain meager. Collaborative and  
426 multidisciplinary approaches using clinical samples from field sites in endemic areas, in vitro and ex vivo  
427 models, as well as animal models of the disease, are crucial to allow global advances in the fight not only  
428 against severe falciparum malaria, but also emerging public health issues such as severe malaria caused by *P.*  
429 *vivax* and *P. knowlesi*.

### 431 **Acknowledgements**

432 Research reported in this publication was supported by the National Institute of Allergy and Infectious  
433 Diseases of the National Institutes of Health (USA) under Award Number U19AI089676, as well as by the  
434 National Health and Medical Research Council (Australia), the Rebecca L. Cooper Foundation (Australia) and  
435 the Australian Research Council (Australia).

439 **References**

- 440 Adams, Y., Kuhnrae, P., Higgins, M.K., Ghumra, A., Rowe, J.A., 2014. Rosetting *Plasmodium falciparum*-  
441 infected erythrocytes bind to human brain microvascular endothelial cells in vitro, demonstrating a dual  
442 adhesion phenotype mediated by distinct *P. falciparum* erythrocyte membrane protein 1 domains. Infect  
443 Immun 82, 949-959.
- 444 Aird, W.C., Mosnier, L.O., Fairhurst, R.M., 2014. *Plasmodium falciparum* picks (on) EPCR. Blood 123, 163-  
445 167.
- 446 Anderson, D.C., Lapp, S.A., Akinyi, S., Meyer, E.V., Barnwell, J.W., Korir-Morrison, C., Galinski, M.R., 2015.  
447 *Plasmodium vivax* trophozoite-stage proteomes. J Proteomics 115, 157-176.
- 448 Ataide, R., Murillo, O., Dombrowski, J.G., Souza, R.M., Lima, F.A., Lima, G.F., Hristov, A.D., Valle, S.C., Di  
449 Santi, S.M., Epiphonio, S., Marinho, C.R., 2015. Malaria in Pregnancy Interacts with and Alters the  
450 Angiogenic Profiles of the Placenta. PLoS Negl Trop Dis 9, e0003824.
- 451 Aung, N.M., Kaung, M., Kyi, T.T., Kyaw, M.P., Min, M., Htet, Z.W., Anstey, N.M., Kyi, M.M., Hanson, J., 2015.  
452 The Safety of a Conservative Fluid Replacement Strategy in Adults Hospitalised with Malaria. PloS One  
453 10, e0143062.
- 454 Avril, M., Bernabeu, M., Benjamin, M., Brazier, A.J., Smith, J.D., 2016. Interaction between Endothelial Protein  
455 C Receptor and Intercellular Adhesion Molecule 1 to Mediate Binding of *Plasmodium falciparum*-Infected  
456 Erythrocytes to Endothelial Cells. mBio 7, e00615.
- 457 Avril, M., Cartwright, M.M., Hathaway, M.J., Hommel, M., Elliott, S.R., Williamson, K., Narum, D.L., Duffy, P.E.,  
458 Fried, M., Beeson, J.G., Smith, J.D., 2010. Immunization with VAR2CSA-DBL5 recombinant protein elicits  
459 broadly cross-reactive antibodies to placental *Plasmodium falciparum*-infected erythrocytes. Infect Immun  
460 78, 2248-2256.
- 461 Ay, F., Bunnik, E.M., Varoquaux, N., Vert, J.P., Noble, W.S., Le Roch, K.G., 2015. Multiple dimensions of  
462 epigenetic gene regulation in the malaria parasite *Plasmodium falciparum*: gene regulation via histone  
463 modifications, nucleosome positioning and nuclear architecture in *P. falciparum*. BioEssays : News Rev  
464 Mol Cell Dev Biol 37, 182-194.

465 Barber, B.E., William, T., Grigg, M.J., Parameswaran, U., Piera, K.A., Price, R.N., Yeo, T.W., Anstey, N.M.,  
466 2015. Parasite biomass-related inflammation, endothelial activation, microvascular dysfunction and  
467 disease severity in vivax malaria. PLoS Pathog 11, e1004558.

468 Barrera, V., Hiscott, P.S., Craig, A.G., White, V.A., Milner, D.A., Beare, N.A., MacCormick, I.J., Kamiza, S.,  
469 Taylor, T.E., Molyneux, M.E., Harding, S.P., 2015. Severity of retinopathy parallels the degree of parasite  
470 sequestration in the eyes and brains of malawian children with fatal cerebral malaria. J Infect Dis 211,  
471 1977-1986.

472 Bernabeu, M., Danziger, S.A., Avril, M., Vaz, M., Babar, P.H., Brazier, A.J., Herricks, T., Maki, J.N., Pereira, L.,  
473 Mascarenhas, A., Gomes, E., Chery, L., Aitchison, J.D., Rathod, P.K., Smith, J.D., 2016. Severe adult  
474 malaria is associated with specific PfEMP1 adhesion types and high parasite biomass. Proc Natl Acad Sci  
475 (USA), 113, E3270-E3279.

476 Biswas, A.K., Hafiz, A., Banerjee, B., Kim, K.S., Datta, K., Chitnis, C.E., 2007. *Plasmodium falciparum* uses  
477 gC1qR/HABP1/p32 as a receptor to bind to vascular endothelium and for platelet-mediated clumping.  
478 PLoS Pathog 3, 1271-1280.

479 Brancucci, N.M., Bertschi, N.L., Zhu, L., Niederwieser, I., Chin, W.H., Wampfler, R., Freymond, C., Rottmann,  
480 M., Felger, I., Bozdech, Z., Voss, T.S., 2014. Heterochromatin protein 1 secures survival and transmission  
481 of malaria parasites. Cell Host Microbe 16, 165-176.

482 Brown, H.C., Chau, T.T., Mai, N.T., Day, N.P., Sinh, D.X., White, N.J., Hien, T.T., Farrar, J., Turner, G.D.,  
483 2000. Blood-brain barrier function in cerebral malaria and CNS infections in Vietnam. Neurology 55, 104-  
484 111.

485 Carlson, J., Helmsby, H., Hill, A.V., Brewster, D., Greenwood, B.M., Wahlgren, M., 1990. Human cerebral  
486 malaria: association with erythrocyte rosetting and lack of anti-rosetting antibodies. Lancet 336, 1457-  
487 1460.

488 Coleman, B.I., Skillman, K.M., Jiang, R.H., Childs, L.M., Altenhofen, L.M., Ganter, M., Leung, Y., Goldowitz, I.,  
489 Kafsack, B.F., Marti, M., Llinas, M., Buckee, C.O., Duraisingh, M.T., 2014. A *Plasmodium falciparum*  
490 histone deacetylase regulates antigenic variation and gametocyte conversion. Cell Host Microbe 16, 177-  
491 186.

492 Daniyan, M.O., Boshoff, A., Prinsloo, E., Pesce, E.R., Blatch, G.L., 2016. The Malarial Exported PFA0660w Is  
493 an Hsp40 Co-Chaperone of PfHsp70-x. PloS One 11, e0148517.

494 Desai, M., ter Kuile, F.O., Nosten, F., McGready, R., Asamo, K., Brabin, B., Newman, R.D., 2007.  
495 Epidemiology and burden of malaria in pregnancy. Lancet. Infect Dis 7, 93-104.

496 Doritchamou, J.Y., Herrera, R., Aebig, J.A., Morrison, R., Nguyen, V., Reiter, K., Shimp, R., MacDonald, N.J.,  
497 Narum, D.L., Fried, M., Duffy, P.E., 2016. VAR2CSA domain-specific analysis of naturally acquired  
498 functional antibodies to *P. falciparum* placental malaria. J Infect Dis, 214. 577-586.

499 Dorovini-Zis, K., Schmidt, K., Huynh, H., Fu, W., Whitten, R.O., Milner, D., Kamiza, S., Molyneux, M., Taylor,  
500 T.E., 2011. The neuropathology of fatal cerebral malaria in malawian children. Am J Pathol 178, 2146-  
501 2158.

502 Fried, M., Duffy, P.E., 2015. Designing a VAR2CSA-based vaccine to prevent placental malaria. Vaccine 33,  
503 7483-7488.

504 Gillrie, M.R., Avril, M., Brazier, A.J., Davis, S.P., Stins, M.F., Smith, J.D., Ho, M., 2015. Diverse functional  
505 outcomes of *Plasmodium falciparum* ligation of EPCR: potential implications for malarial pathogenesis.  
506 Cell Micro 17, 1883-1899.

507 Gilson, P.R., Chisholm, S.A., Crabb, B.S., de Koning-Ward, T.F., 2016. Host cell remodelling in malaria  
508 parasites: a new pool of potential drug targets. Int J Parasitol. [This issue].

509 Goel, S., Palmkvist, M., Moll, K., Joannin, N., Lara, P., Akhouri, R.R., Moradi, N., Ojemalm, K., Westman, M.,  
510 Angeletti, D., Kjellin, H., Lehtio, J., Blixt, O., Idestrom, L., Gahmberg, C.G., Storry, J.R., Hult, A.K., Olsson,  
511 M.L., von Heijne, G., Nilsson, I., Wahlgren, M., 2015. RIFINs are adhesins implicated in severe  
512 *Plasmodium falciparum* malaria. Nat Med 21, 314-317.

513 Greiner, J., Dorovini-Zis, K., Taylor, T.E., Molyneux, M.E., Beare, N.A., Kamiza, S., White, V.A., 2015.  
514 Correlation of hemorrhage, axonal damage, and blood-tissue barrier disruption in brain and retina of  
515 Malawian children with fatal cerebral malaria. Front Cell Infect Microbiol 5, 18.

516 Hackett, M.J., Aitken, J.B., El-Assaad, F., McQuillan, J.A., Carter, E.A., Ball, H.J., Tobin, M.J., Paterson, D., de  
517 Jonge, M.D., Siegele, R., Cohen, D.D., Vogt, S., Grau, G.E., Hunt, N.H., Lay, P.A., 2015. Mechanisms of  
518 murine cerebral malaria: Multimodal imaging of altered cerebral metabolism and protein oxidation at  
519 hemorrhage sites. Sci Adv 1, e1500911.

520 Handunnetti, S.M., David, P.H., Perera, K.L., Mendis, K.N., 1989. Uninfected erythrocytes form "rosettes"  
521 around *Plasmodium falciparum* infected erythrocytes. Am J Trop Med Hyg 40, 115-118.

522 Hanson, J., Anstey, N.M., Bihari, D., White, N.J., Day, N.P., Dondorp, A.M., 2014. The fluid management of  
523 adults with severe malaria. Crit Care 18, 642.

524 Harris, L.M., Merrick, C.J., 2015. G-quadruplexes in pathogens: a common route to virulence control? PLoS  
525 Pathog 11, e1004562.

526 Hawkes, M., Opoka, R.O., Namasopo, S., Miller, C., Conroy, A.L., Serghides, L., Kim, H., Thampi, N., Liles,  
527 W.C., John, C.C., Kain, K.C., 2011. Nitric oxide for the adjunctive treatment of severe malaria: hypothesis  
528 and rationale. Med Hypotheses 77, 437-444.

529 Hendriksen, I.C., Mwanga-Amumpaire, J., von Seidlein, L., Mtove, G., White, L.J., Olaosebikan, R., Lee, S.J.,  
530 Tshetu, A.K., Woodrow, C., Amos, B., Karema, C., Saiwaew, S., Maitland, K., Gomes, E., Pan-Ngum, W.,  
531 Gesase, S., Silamut, K., Reyburn, H., Joseph, S., Chotivanich, K., Fanello, C.I., Day, N.P., White, N.J.,  
532 Dondorp, A.M., 2012. Diagnosing severe falciparum malaria in parasitaemic African children: a  
533 prospective evaluation of plasma PfHRP2 measurement. PLoS Med 9, e1001297.

534 Hoffmann, A., Pfeil, J., Alfonso, J., Kurz, F.T., Sahm, F., Heiland, S., Monyer, H., Bendszus, M., Mueller, A.K.,  
535 Helluy, X., Pham, M., 2016. Experimental Cerebral Malaria Spreads along the Rostral Migratory Stream.  
536 PLoS Pathog 12, e1005470.

537 Hommel, M., Elliott, S.R., Soma, V., Kelly, G., Fowkes, F.J., Chesson, J.M., Duffy, M.F., Bockhorst, J., Avril,  
538 M., Mueller, I., Raiko, A., Stanistic, D.I., Rogerson, S.J., Smith, J.D., Beeson, J.G., 2010. Evaluation of the  
539 antigenic diversity of placenta-binding *Plasmodium falciparum* variants and the antibody repertoire among  
540 pregnant women. Infect Immun 78, 1963-1978.

541 Hviid, L., Jensen, A.T., 2015. PfEMP1 - A Parasite Protein Family of Key Importance in *Plasmodium falciparum*  
542 Malaria Immunity and Pathogenesis. Adv Parasitol 88, 51-84.

543 Jespersen, J.S., Wang, C.W., Mkumbaye, S.I., Minja, D.T., Petersen, B., Turner, L., Petersen, J.E., Lusingu,  
544 J.P., Theander, T.G., Lavstsen, T., 2016. *Plasmodium falciparum* var genes expressed in children with  
545 severe malaria encode CIDRalpha1 domains. EMBO Mol Med 8, 839-850.

546 Josling, G.A., Petter, M., Oehring, S.C., Gupta, A.P., Dietz, O., Wilson, D.W., Schubert, T., Langst, G., Gilson,  
547 P.R., Crabb, B.S., Moes, S., Jenoe, P., Lim, S.W., Brown, G.V., Bozdech, Z., Voss, T.S., Duffy, M.F.,

548 2015. A *Plasmodium falciparum* Bromodomain Protein Regulates Invasion Gene Expression. Cell Host  
549 Microbe 17, 741-751.

550 Joyner, C., Barnwell, J.W., Galinski, M.R., 2015. No more monkeying around: primate malaria model systems  
551 are key to understanding *Plasmodium vivax* liver-stage biology, hypnozoites, and relapses. Front Microbiol  
552 6, 145.

553 Kalanon, M., Bargieri, D., Sturm, A., Matthews, K., Ghosh, S., Goodman, C.D., Thiberge, S., Mollard, V.,  
554 McFadden, G.I., Menard, R., de Koning-Ward, T.F., 2016. The *Plasmodium* translocon of exported  
555 proteins component EXP2 is critical for establishing a patent malaria infection in mice. Cell Microbiol 18,  
556 399-412.

557 Karmodiya, K., Pradhan, S.J., Joshi, B., Jangid, R., Reddy, P.C., Galande, S., 2015. A comprehensive  
558 epigenome map of *Plasmodium falciparum* reveals unique mechanisms of transcriptional regulation and  
559 identifies H3K36me2 as a global mark of gene suppression. Epigenetics Chromatin 8, 32.

560 Lapp, S.A., Mok, S., Zhu, L., Wu, H., Preiser, P.R., Bozdech, Z., Galinski, M.R., 2015. *Plasmodium knowlesi*  
561 gene expression differs in ex vivo compared to in vitro blood-stage cultures. Malaria J 14, 110.

562 Maier, A.G., Cooke, B.M., Cowman, A.F., Tilley, L., 2009. Malaria parasite proteins that remodel the host  
563 erythrocyte. Nat Rev Microbiol 7, 341-354.

564 Manning, L., Laman, M., Davis, W.A., Davis, T.M., 2014. Clinical features and outcome in children with severe  
565 *Plasmodium falciparum* malaria: a meta-analysis. PloS One 9, e86737.

566 Maude, R.J., Barkhof, F., Hassan, M.U., Ghose, A., Hossain, A., Abul Faiz, M., Choudhury, E., Rashid, R., Abu  
567 Sayeed, A., Charunwatthana, P., Plewes, K., Kingston, H., Maude, R.R., Silamut, K., Day, N.P., White,  
568 N.J., Dondorp, A.M., 2014. Magnetic resonance imaging of the brain in adults with severe falciparum  
569 malaria. Malaria J 13, 177.

570 Mayor, A., Bardaji, A., Macete, E., Nhampossa, T., Fonseca, A.M., Gonzalez, R., Maculuve, S., Cistero, P.,  
571 Ruperez, M., Campo, J., Vala, A., Sigauque, B., Jimenez, A., Machevo, S., de la Fuente, L., Nhama, A.,  
572 Luis, L., Aponte, J.J., Acacio, S., Nhacolo, A., Chitnis, C., Dobano, C., Sevene, E., Alonso, P.L.,  
573 Menendez, C., 2015. Changing Trends in *P. falciparum* Burden, Immunity, and Disease in Pregnancy.  
574 New Engl J Med 373, 1607-1617.

575 Mayor, A., Hafiz, A., Bassat, Q., Rovira-Vallbona, E., Sanz, S., Machevo, S., Aguilar, R., Cistero, P., Sigauque,  
576 B., Menendez, C., Alonso, P.L., Chitnis, C.E., 2011. Association of severe malaria outcomes with platelet-  
577 mediated clumping and adhesion to a novel host receptor. PLoS One 6, e19422.

578 McDonald, C.R., Darling, A.M., Conroy, A.L., Tran, V., Cabrera, A., Liles, W.C., Wang, M., Aboud, S., Urassa,  
579 W., Fawzi, W.W., Kain, K.C., 2015a. Inflammatory and Angiogenic Factors at Mid-Pregnancy Are  
580 Associated with Spontaneous Preterm Birth in a Cohort of Tanzanian Women. PLoS One 10, e0134619.

581 McDonald, C.R., Tran, V., Kain, K.C., 2015b. Complement Activation in Placental Malaria. Front Microbiol 6,  
582 1460.

583 Medana, I.M., Day, N.P., Sachanonta, N., Mai, N.T., Dondorp, A.M., Pongponratn, E., Hien, T.T., White, N.J.,  
584 Turner, G.D., 2011. Coma in fatal adult human malaria is not caused by cerebral oedema. Malaria J, 10,  
585 267.

586 Mohanty, S., Mishra, S.K., Patnaik, R., Dutt, A.K., Pradhan, S., Das, B., Patnaik, J., Mohanty, A.K., Lee, S.J.,  
587 Dondorp, A.M., 2011. Brain swelling and mannitol therapy in adult cerebral malaria: a randomized trial.  
588 Clin Infect Dis 53, 349-355.

589 Moxon, C.A., Chisala, N.V., Wassmer, S.C., Taylor, T.E., Seydel, K.B., Molyneux, M.E., Faragher, B.,  
590 Kennedy, N., Toh, C.H., Craig, A.G., Heyderman, R.S., 2014. Persistent endothelial activation and  
591 inflammation after *Plasmodium falciparum* Infection in Malawian children. J Infect Dis 209, 610-615.

592 Moxon, C.A., Wassmer, S.C., Milner, D.A., Jr., Chisala, N.V., Taylor, T.E., Seydel, K.B., Molyneux, M.E.,  
593 Faragher, B., Esmon, C.T., Downey, C., Toh, C.H., Craig, A.G., Heyderman, R.S., 2013. Loss of  
594 endothelial protein C receptors links coagulation and inflammation to parasite sequestration in cerebral  
595 malaria in African children. Blood 122, 842-851.

596 Nacer, A., Movila, A., Sohet, F., Girgis, N.M., Gundra, U.M., Loke, P., Daneman, R., Frevert, U., 2014.  
597 Experimental cerebral malaria pathogenesis--hemodynamics at the blood brain barrier. PLoS Pathog 10,  
598 e1004528.

599 O'Sullivan, J.M., Preston, R.J., O'Regan, N., O'Donnell, J.S., 2016. Emerging roles for hemostatic dysfunction  
600 in malaria pathogenesis. Blood 127, 2281-2288.

601 Padmanabhan, B., Mathur, S., Manjula, R., Tripathi, S., 2016. Bromodomain and extra-terminal (BET) family  
602 proteins: New therapeutic targets in major diseases. J Biosci 41, 295-311.



603 Pai, S., Qin, J., Cavanagh, L., Mitchell, A., El-Assaad, F., Jain, R., Combes, V., Hunt, N.H., Grau, G.E.,  
604 Weninger, W., 2014. Real-time imaging reveals the dynamics of leukocyte behaviour during experimental  
605 cerebral malaria pathogenesis. PLoS Pathog 10, e1004236.

606 Pain, A., Ferguson, D.J., Kai, O., Urban, B.C., Lowe, B., Marsh, K., Roberts, D.J., 2001. Platelet-mediated  
607 clumping of *Plasmodium falciparum*-infected erythrocytes is a common adhesive phenotype and is  
608 associated with severe malaria. Proc Natl Acad Sci (USA) 98, 1805-1810.

609 Penet, M.F., Viola, A., Confort-Gouny, S., Le Fur, Y., Duhamel, G., Kober, F., Ibarrola, D., Izquierdo, M., Coltel,  
610 N., Gharib, B., Grau, G.E., Cozzone, P.J., 2005. Imaging experimental cerebral malaria in vivo: significant  
611 role of ischemic brain edema. J Neurosci 25, 7352-7358.

612 Petersen, J.E., Bouwens, E.A., Tamayo, I., Turner, L., Wang, C.W., Stins, M., Theander, T.G., Hermida, J.,  
613 Mosnier, L.O., Lavstsen, T., 2015. Protein C system defects inflicted by the malaria parasite protein  
614 PfEMP1 can be overcome by a soluble EPCR variant. Thromb Haemost 114, 1038-1048.

615 Potchen, M.J., Kampondeni, S.D., Seydel, K.B., Birbeck, G.L., Hammond, C.A., Bradley, W.G., DeMarco, J.K.,  
616 Glover, S.J., Ugorji, J.O., Latourette, M.T., Siebert, J.E., Molyneux, M.E., Taylor, T.E., 2012. Acute brain  
617 MRI findings in 120 Malawian children with cerebral malaria: new insights into an ancient disease. AJNR  
618 Am J Neuroradiol 33, 1740-1746.

619 Rogerson, S.J., Pollina, E., Getachew, A., Tadesse, E., Lema, V.M., Molyneux, M.E., 2003. Placental  
620 monocyte infiltrates in response to *Plasmodium falciparum* malaria infection and their association with  
621 adverse pregnancy outcomes. American J Trop Med Hyg 68, 115-119.

622 Rowe, A., Obeiro, J., Newbold, C.I., Marsh, K., 1995. *Plasmodium falciparum* rosetting is associated with  
623 malaria severity in Kenya. Infect Immun 63, 2323-2326.

624 Rowe, J.A., Claessens, A., Corrigan, R.A., Arman, M., 2009. Adhesion of *Plasmodium falciparum*-infected  
625 erythrocytes to human cells: molecular mechanisms and therapeutic implications. Expert Rev Mol Med 11,  
626 e16.

627 Rowe, J.A., Handel, I.G., Thera, M.A., Deans, A.M., Lyke, K.E., Kone, A., Diallo, D.A., Raza, A., Kai, O.,  
628 Marsh, K., Plowe, C.V., Doumbo, O.K., Moulds, J.M., 2007. Blood group O protects against severe  
629 *Plasmodium falciparum* malaria through the mechanism of reduced rosetting. Proc Natl Acad Sci (USA)  
630 104, 17471-17476.

631 Rowe, J.A., Shafi, J., Kai, O.K., Marsh, K., Raza, A., 2002. Nonimmune IgM, but not IgG binds to the surface of  
632 *Plasmodium falciparum*-infected erythrocytes and correlates with rosetting and severe malaria. Am J Trop  
633 Med Hyg 66, 692-699.

634 Sahu, P.K., Satpathi, S., Behera, P.K., Mishra, S.K., Mohanty, S., Wassmer, S.C., 2015. Pathogenesis of  
635 cerebral malaria: new diagnostic tools, biomarkers, and therapeutic approaches. Front Cell Infect Microbiol  
636 5, 75.

637 Salinas, J.L., Kissinger, J.C., Jones, D.P., Galinski, M.R., 2014. Metabolomics in the fight against malaria.  
638 Memorias do Instituto Oswaldo Cruz 109, 589-597.

639 Serghides, L., Kim, H., Lu, Z., Kain, D.C., Miller, C., Francis, R.C., Liles, W.C., Zapol, W.M., Kain, K.C., 2011.  
640 Inhaled nitric oxide reduces endothelial activation and parasite accumulation in the brain, and enhances  
641 survival in experimental cerebral malaria. PloS One 6, e27714.

642 Seydel, K.B., Kampondeni, S.D., Valim, C., Potchen, M.J., Milner, D.A., Muwalo, F.W., Birbeck, G.L., Bradley,  
643 W.G., Fox, L.L., Glover, S.J., Hammond, C.A., Heyderman, R.S., Chilingulo, C.A., Molyneux, M.E., Taylor,  
644 T.E., 2015. Brain swelling and death in children with cerebral malaria. New Engl J Med 372, 1126-1137.

645 Souza, R.M., Ataide, R., Dombrowski, J.G., Ippolito, V., Aitken, E.H., Valle, S.N., Alvarez, J.M., Epiphonio, S.,  
646 Marinho, C.R., 2013. Placental histopathological changes associated with *Plasmodium vivax* infection  
647 during pregnancy. PLoS Negl Trop Dis 7, e2071.

648 Stevenson, L., Laursen, E., Cowan, G.J., Bandoh, B., Barfod, L., Cavanagh, D.R., Andersen, G.R., Hviid, L.,  
649 2015. alpha2-Macroglobulin Can Crosslink Multiple *Plasmodium falciparum* Erythrocyte Membrane Protein  
650 1 (PfEMP1) Molecules and May Facilitate Adhesion of Parasitized Erythrocytes. PLoS Pathog 11,  
651 e1005022.

652 Storm, J., Craig, A.G., 2014. Pathogenesis of cerebral malaria-inflammation and cytoadherence. Front Cell  
653 Infect Microbiol 4, 100.

654 Taylor, T.E., Fu, W.J., Carr, R.A., Whitten, R.O., Mueller, J.S., Fosiko, N.G., Lewallen, S., Liomba, N.G.,  
655 Molyneux, M.E., 2004. Differentiating the pathologies of cerebral malaria by postmortem parasite counts.  
656 Nat Med, 10, 143-145.

657 Turner, L., Lavstsen, T., Berger, S.S., Wang, C.W., Petersen, J.E., Avril, M., Brazier, A.J., Freeth, J.,  
658 Jespersen, J.S., Nielsen, M.A., Magistrado, P., Lusingu, J., Smith, J.D., Higgins, M.K., Theander, T.G.,

659 2013. Severe malaria is associated with parasite binding to endothelial protein C receptor. *Nature* 498,  
660 502-505.

661 Warrell, D.A., Looareesuwan, S., Phillips, R.E., White, N.J., Warrell, M.J., Chapel, H.M., Areekul, S.,  
662 Tharavanij, S., 1986. Function of the blood-cerebrospinal fluid barrier in human cerebral malaria: rejection  
663 of the permeability hypothesis. *Am J Trop Med Hyg* 35, 882-889.

664 Wassmer, S.C., Combes, V., Grau, G.E., 2011a. Platelets and microparticles in cerebral malaria: the unusual  
665 suspects. *Drug Discov Today Dis Mech* 8, e15-e23.

666 Wassmer, S.C., Moxon, C.A., Taylor, T., Grau, G.E., Molyneux, M.E., Craig, A.G., 2011b. Vascular endothelial  
667 cells cultured from patients with cerebral or uncomplicated malaria exhibit differential reactivity to TNF.  
668 *Cell Microbiol* 13, 198-209.

669 Wassmer, S.C., Taylor, T., Maclennan, C.A., Kanjala, M., Mukaka, M., Molyneux, M.E., Grau, G.E., 2008.  
670 Platelet-induced clumping of *Plasmodium falciparum*-infected erythrocytes from Malawian patients with  
671 cerebral malaria-possible modulation in vivo by thrombocytopenia. *J Infect Dis* 197, 72-78.

672 Wassmer, S.C., Taylor, T.E., Rathod, P.K., Mishra, S.K., Mohanty, S., Arevalo-Herrera, M., Duraisingh, M.T.,  
673 Smith, J.D., 2015. Investigating the Pathogenesis of Severe Malaria: A Multidisciplinary and Cross-  
674 Geographical Approach. *Am J Trop Med Hyg* 93, 42-56.

675 White, N.J., Turner, G.D., Day, N.P., Dondorp, A.M., 2013. Lethal malaria: Marchiafava and Bignami were  
676 right. *J Infect Dis* 208, 192-198.

677 Yeo, T.W., Lampah, D.A., Gitawati, R., Tjitra, E., Kenangalem, E., McNeil, Y.R., Darcy, C.J., Granger, D.L.,  
678 Weinberg, J.B., Lopansri, B.K., Price, R.N., Duffull, S.B., Celermajer, D.S., Anstey, N.M., 2007. Impaired  
679 nitric oxide bioavailability and L-arginine reversible endothelial dysfunction in adults with falciparum  
680 malaria. *J Exp Med* 204, 2693-2704.

681 Yeo, T.W., Lampah, D.A., Kenangalem, E., Tjitra, E., Weinberg, J.B., Granger, D.L., Price, R.N., Anstey, N.M.,  
682 2014. Decreased endothelial nitric oxide bioavailability, impaired microvascular function, and increased  
683 tissue oxygen consumption in children with falciparum malaria. *J Infect Dis* 210, 1627-1632.

684 Yeo, T.W., Lampah, D.A., Rooslamati, I., Gitawati, R., Tjitra, E., Kenangalem, E., Price, R.N., Duffull, S.B.,  
685 Anstey, N.M., 2013. A randomized pilot study of L-arginine infusion in severe falciparum malaria:  
686 preliminary safety, efficacy and pharmacokinetics. *PloS One* 8, e69587.