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NMR Crystallography NMR & GIPAW (CASTEP) Signam (CASTEP



Single-crystal X-ray diffraction and NMR crystallography of a 1:1 cocrystal of dithianon and pyrimethanil

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A single-crystal X-ray diffraction structure of a 1:1 cocrystal of two fungicides, namely dithianon (DI) and pyrimethanil (PM), is reported [systematic name: 5,10-dioxo-5H,10H-naphtho[2,3-b][1,4]dithiine-2,3-dicarbonitrile-4,6-dimethyl-N-phenylpyrimidin-2-amine (1/1), $C_{14}H_4N_2O_2S_2\cdot C_{12}H_{13}N_2$]. Following an NMR crystallography approach, experimental solid-state magic angle spinning (MAS) NMR spectra are presented together with GIPAW (gauge-including projector augmented wave) calculations of NMR chemical shieldings. Specifically, experimental ¹H and ¹³C chemical shifts are determined from two-dimensional ¹H-¹³C MAS NMR correlation spectra recorded with short and longer contact times so as to probe one-bond C-H connectivities and longer-range C···H proximities, whereas H···H proximities are identified in a ¹H double-quantum (DQ) MAS NMR spectrum. The performing of separate GIPAW calculations for the full periodic crystal structure and for isolated molecules allows the determination of the change in chemical shift upon going from an isolated molecule to the full crystal structure. For the ¹H NMR chemical shifts, changes of 3.6 and 2.0 ppm correspond to intermolecular N-H···O and C-H···O hydrogen bonding, while changes of -2.7 and -1.5 ppm are due to ring current effects associated with $C-H\cdots\pi$ interactions. Even though there is a close intermolecular S···O distance of 3.10 Å, it is of note that the molecule-to-crystal chemical shifts for the involved sulfur or oxygen nuclei are small.

1. Introduction

With an increasing global population, limited availability of arable land, an increase in extreme weather events and growing pest resistance to certain existing agrochemical products, innovation in the agrochemical industry is as important as ever if we are to provide enough food for everyone. With lower usage rates, ease of use and more favourable toxicology profiles being important objectives, the search for and structure-based design of potential agrochemical products needs to become more efficient (Lamberth et al., 2013). One possibility in this regard is the usage of cocrystals formed between an active ingredient and coformers or other active ingredients via reversible noncovalent interactions. While this is an established procedure in the development of new active pharmaceutical ingredients, where it is used to increase the solubility and bioavailability (Blagden et al., 2007), there is also great potential to exploit cocrystals in the optimization and development of agrochemicals. For example, a reduced solubility could increase the agrochemical's resi-

nmr crystallography

Table 1
Experimental details.

| Crystal data | |
|--|--|
| Chemical formula | $C_{14}H_4N_2O_2S_2 \cdot C_{12}H_{13}N_3$ |
| $M_{ m r}$ | 495.59 |
| Crystal system, space group | Monoclinic, $P2_1/n$ |
| Temperature (K) | 100 |
| a, b, c (Å) | 7.1707 (2), 22.8006 (6), 13.8237 (4) |
| β (°) | 97.047 (3) |
| β (°) V (Å ³) | 2243.04 (7) |
| Z | 4 |
| Radiation type | Cu <i>Kα</i> |
| $\mu \text{ (mm}^{-1})$ | 2.45 |
| Crystal size (mm) | $0.60 \times 0.10 \times 0.02$ |
| Data collection | |
| Diffractometer | Agilent Xcalibur Onyx Ultra |
| Absorption correction | Multi-scan (<i>CrysAlis PRO</i> ; Agilent, 2014) |
| T_{\min}, T_{\max} | 0.596, 1.000 |
| No. of measured, independent and | 5143, 3160, 2667 |
| observed $[I > 2.0\sigma(I)]$ reflections | |
| $R_{ m int}$ | 0.035 |
| $\theta_{ m max}$ ($^{\circ}$) | 58.9 |
| $(\sin \theta/\lambda)_{\max} (\mathring{\mathbf{A}}^{-1})$ | 0.556 |
| Refinement | |
| $R[F^2 > 2\sigma(F^2)], wR(F^2), S$ | 0.045, 0.094, 0.98 |
| No. of reflections | 3141 |
| No. of parameters | 109 |
| No. of restraints | 3 |
| H-atom treatment | H atoms treated by a mixture of independent and constrained refinement |
| $\Delta \rho_{\rm max}$, $\Delta \rho_{\rm min}$ (e Å ⁻³) | 0.43, -0.37 |

Computer programs: CrysAlis PRO (Agilent, 2014), SUPERFLIP (Palatinus & Chapuis, 2007), CRYSTALS (Betteridge et al., 2003), CAMERON (Watkin et al., 1996) and Mercury (Macrae et al., 2006).

dence time on the respective plant and multicomponent entities could improve the release profile (and thus absolute usage), as well as allow the simultaneous delivery of two or more active components. However, the design of suitable cocrystalline materials and prediction of their properties and formed cocrystal structures is far from being trivial. Some design strategies based on the hierarchy of intermolecular interactions (Aakeroy & Salmon, 2005) or the assessment of the solubilities and saturation temperatures of the pure compounds to be included in a cocrystalline arrangement (ter Horst *et al.*, 2009) are available as a guideline. However, if multiple and different hydrogen-bonding donors and acceptors are present in the molecules, a reliable prediction of the resulting structure becomes very difficult (Bhatt *et al.*, 2009).

NMR crystallography, namely the combination of experimental solid-state magic angle spinning (MAS) NMR with calculation of NMR parameters, is finding important application to moderately sized organic molecules (Harris, 2004; Elena *et al.*, 2006; Harris *et al.*, 2009; Bonhomme *et al.*, 2012). We present here an NMR crystallography analysis of the 1:1 cocrystal of two fungicides, namely dithianon (DI) and pyrimethanil (PM). Specifically, following a preparation protocol in Sowa *et al.* (2013), a single-crystal X-ray diffraction structure determination is reported, with this structure (after DFT geometry optimization) providing the input for a calculation, using the GIPAW (gauge-including projector augmented

wave) method (Pickard & Mauri, 2001; Yates *et al.*, 2007), of the NMR chemical shieldings. The computational analysis is complemented by the recording of 1D (one-dimensional) and 2D (two-dimensional) experimental ¹H and ¹³C MAS NMR spectra. Building upon studies of pharmaceutical cocrystals by such an NMR crystallography investigation (Tatton *et al.*, 2013; Dudenko *et al.*, 2013; Stevens *et al.*, 2014; Kerr *et al.*, 2015; Sardo *et al.*, 2015; Luedeker *et al.*, 2016), we present here the application of this approach to an agrochemical cocrystal.

2. Experimental and computational details

2.1. Sample preparation

The DI-PM cocrystal was prepared according to method VII in point [0041] of Sowa *et al.* (2013), *i.e.* dry dithianon and pyrimethanil (both solids) were mixed thoroughly in a 1:1 molar ratio (0.5 g of pyrimethanil) and kept at 323 K under agitation. After a couple of hours, the powdery product had changed to a dark-olive-green colour.

2.2. Single-crystal X-ray diffraction: structure solution and refinement

Crystal data, data collection and structure refinement details are summarized in Table 1. The H atoms were all located in a difference map, but those attached to C atoms were repositioned geometrically. The H atoms were initially refined with soft restraints on the bond lengths and angles to regularize their geometry [C-H = 0.93–0.98 Å and N-H = 0.86–0.89 Å, and with $U_{\rm iso}({\rm H})$ = 1.2–1.5 $U_{\rm eq}({\rm parent})$], after which the positions were refined with riding constraints (Cooper *et al.*, 2010).

2.3. Solid-state NMR

1D ¹H MAS and 1D ¹³C cross polarization (CP) MAS experiments were performed on a Bruker Avance III spectrometer operating at ¹H and ¹³C Larmor frequencies of 600 and 150.9 MHz, respectively, using a 1.3 mm HXY (¹H MAS) or a 4 mm HX (¹³C CP MAS) Bruker probe. In all cases, a ¹H 90° pulse duration of 2.5 μs was used. 2D ¹H–¹³C HETCOR experiments were performed on a Bruker Avance III spectrometer, using a 4 mm HXY probe in double-resonance mode. In the HETCOR pulse sequence, the following phase cycling was employed: ¹H 90° pulse (90° 270°), ¹³C 180° pulse (2{0°} 2{180°}), ¹³C CP contact pulse (4{0°} 4{180°} 4{90°} 4{270°}), receiver (0° 180° 0° 180° 180° 0° 180° 0° 90° 270° 90° 270° 90° 270° 90°). For CP, a 70 to 100% ramp (Metz *et al.*, 1994) on the ¹H channel was used for the CP contact time. During acquisition of a ¹³C FID, SPINAL64 (Fung *et al.*, 2000)

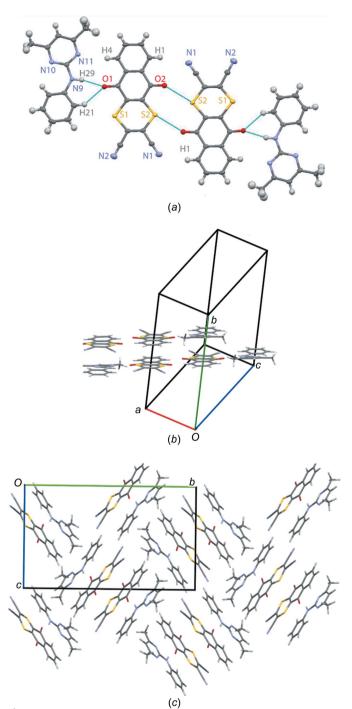


Figure 1
Representations of the crystal structure of the DI-PM cocrystal, showing (a) the intermolecular interactions within a 'chain' of molecules, with displacement ellipsoids drawn at the 50% probability level, (b) the packing of two chains of molecules as 'layers' and (c) the 'zigzag' arrangement of chains (viewed along the crystallographic a axis). In parts (b) and (c), the unit cell is shown, indicating the a, b and c unit-cell axes.

¹H heteronuclear decoupling was applied with a pulse duration of 5.9 μs at a nutation frequency of 100 kHz. A 2D ¹H DQ experiment with BABA recoupling (Sommer *et al.*, 1995; Schnell *et al.*, 1998) was performed on a Bruker Avance III spectrometer operating at a ¹H Larmor frequency of 700 MHz using a 1.3 mm HXY Bruker probe. A 16-step phase cycle was

used to select $\Delta p = \pm 2$ on the DQ excitation block and $\Delta p = -1$ on the z-filter 90° pulse, where p is the coherence order. In all 2D experiments, the States-TPPI method was used to achieve sign discrimination in F_1 . ¹³C and ¹H chemical shifts are referenced with respect to TMS using L-alanine at natural abundance as an external reference: 177.8 ppm for the ¹³C carboxylate resonance and 1.1 ppm for the ¹H methyl resonance. All experiments were performed at room temperature, though frictional effects due to MAS increase the actual sample temperature (Langer *et al.*, 1999).

2.4. DFT calculations

Calculations were performed using CASTEP (Clark et al., 2005; Academic Release Version 8.0) and employed the PBE exchange-correlational functional (Perdew et al., 1996). For both geometry optimization and NMR shielding calculations, a plane-wave basis set with ultrasoft pseudopotentials (Vanderbilt, 1990) with a maximum plane-wave cut-off energy of 700 eV was used. A Monkhorst-Pack grid of minimum sample spacing $0.05 \times 2\pi \text{ Å}^{-1}$ was used to take integrals over the Brillouin zone. Geometry optimization was performed with the unit-cell parameters fixed, starting from the singlecrystal X-ray structure. The positions of the 208 atoms in the unit cell (Z = 4, Z' = 1) were relaxed and periodic boundary conditions were applied. The space group $P2_1/n$ was preserved. All distances and angles stated in the main text of this article are for the geometry-optimized crystal structure. Note also that the geometry optimization within CASTEP causes a relabelling of the atoms - in this article, we use the CASTEP numbering; see Fig. S1 in the Supporting information for a comparison with the numbering employed in the crystallographic CIF file. The GIPAW method (Pickard & Mauri, 2001; Yates et al., 2007) was utilized for the NMR chemical-shielding calculations, which were performed on the geometry-optimized structure. For the isolated molecule calculations, a single molecule (either DI or PM) from the fully geometry optimized structure is kept in the unit cell, whose dimensions are also increased by ~5 Å in each direction - the NMR shieldings are then calculated without any further geometry optimization.

3. Results and discussion

3.1. Single-crystal X-ray diffraction structure

The single-crystal X-ray diffraction structure of the DI-PM cocrystal is schematically represented in Fig. 1. As shown in Fig. 1(a), a chain of molecules is held together by $N-H\cdots O$ and $C-H\cdots O$ hydrogen bonds (between DI and PM molecules) and by putative $S\cdots O$ interactions (Burling & Goldstein, 1992) between two DI molecules; note that the relative strengths of these interactions is investigated below (see §3.5) using GIPAW calculations of NMR chemical shieldings. The further packing of two chains of molecules as 'layers' and a 'zigzag' arrangement of chains are shown in Figs. 1(b) and 1(c), respectively. As can be seen from the representation along the crystallographic a axis in Fig. 1(c), the packing is

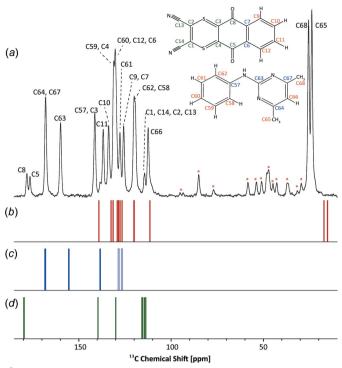


Figure 2 (a) A 1 H (600 MHz) $^{-13}$ C CP MAS (12.5 kHz) NMR spectrum of the DIPM cocrystal (* denote spinning sidebands), together with (b)–(d) stick spectra corresponding to calculated (GIPAW) 13 C chemical shifts (see Table 2). Separate stick spectra are presented according to whether correlation peaks corresponding to (b) direct C—H bonds or (c) longerrange C···H proximities are observed in the 1 H $^{-13}$ C 2D spectra presented in Fig. 4, or (d) where no experimental correlation peaks are observed. In the CP MAS experiment, a contact time of 1.4 ms was used and 1024 transients were co-added for a recycle delay of 57 s.

based on assemblies of blocks of four molecules; four molecules (PM-DI-DI-PM) are arranged in a layer (Fig. 1a), forming a block that is perpendicular to an adjacent block of four molecules, thus building up the 'zigzag' arrangement.

3.2. Experimental and calculated ¹³C chemical shifts

Fig. 2 presents a 13 C CP MAS NMR 1D spectrum (Fig. 2a) of the DI-PM cocrystal, together with three stick spectra (Figs. 2b, 2c and 2d) that represent 13 C chemical shifts calculated using the GIPAW method for the DI-PM crystal structure. Specifically, the calculated 13 C chemical shifts are presented in three groups according to whether they correspond to direct one-bond C-H connectivities (Fig. 2b, red labels) or nonprotonated C atoms (Figs. 2c and 2d, blue and green labels, respectively). The distinction between Figs. 2(c) and 2(d) corresponds to whether cross peaks corresponding to a longer-range $C \cdots H$ proximity are observed in ^{1}H - ^{13}C 2D correlation spectra (see §3.4).

3.3. One- and two-dimensional ¹H MAS NMR spectra

Figs. 3(a) and 3(b) present ¹H NMR spectra of the DI-PM cocrystal recorded at a fast MAS frequency of 60 kHz; specifically, a one-pulse one-dimensional spectrum in Fig. 3(a), together with vertical lines corresponding to calculated

Table 2 Comparison of calculated (GIPAW)^a and experimental ¹³C and ¹H NMR chemical shifts (in ppm) in the DI-PM cocrystal^b.

| | Atom label | 1. | ³ C | ¹ H | | |
|-----|--------------------------|--------------------|--------------------|--------------------|--------------------|--|
| С | Н | $\delta_{ m calc}$ | $\delta_{ m expt}$ | $\delta_{ m calc}$ | $\delta_{ m expt}$ | |
| C65 | H22/H23/H24 ^c | 15.3 | 23.9 | 1.8 | 1.9 | |
| C68 | H26/H27/H28 ^c | 17.2 | 25.7 | 2.0 | 2.0 | |
| C66 | H25 | 111.5 | 112.6 | 3.4 | 4.0 | |
| C1 | _ | 113.8 | 114.4^{d} | _ | _ | |
| C14 | _ | 114.5 | 114.4^{d} | _ | _ | |
| C2 | _ | 115.5 | 114.4^{d} | _ | _ | |
| C13 | _ | 115.9 | 114.4^{d} | _ | _ | |
| C58 | H17 | 120.1 | 119.4 | 9.7 | 9.1 | |
| C62 | H21 | 120.2 | 120.3 | 8.4 | 8.0 | |
| C9 | H1 | 126.7 | 125.7 | 7.4 | 7.4 | |
| C7 | $H1^e$ | 126.8 | 125.7 | 7.4 | 7.4 | |
| C61 | H20 | 127.7 | 127.7 | 7.6 | 7.4 | |
| C12 | H4 | 128.5 | 129.8 | 8.5 | 8.2 | |
| C6 | $H4^e$ | 128.6 | 129.8 | 8.5 | 8.2 | |
| C60 | H19 | 129.3 | 130.2 | 7.3 | 7.8 | |
| C4 | _ | 130.1 | 131.1^{d} | - | _ | |
| C59 | H18 | 131.5 | 131.2 | 7.7 | 7.7 | |
| C10 | H2 | 132.6 | 133.9 | 5.9 | 6.2 | |
| C11 | H3 | 139.2 | 136.8 | 7.6 | 7.7 | |
| C57 | H21, H17, H29 | 138.5 | 141.5 | 8.4, 9.7, 10.5 | 8.9 | |
| C3 | _ | 139.7 | 141.4^{d} | - | _ | |
| C63 | H29 | 155.5 | 160.1 | 10.5 | 9.1 | |
| C67 | H26/H27/H28, H25 | 168.2 | 168.2 | 2.0, 3.4 | 2.8 | |
| C64 | H22/H23/H24, H25 | 168.4 | 168.2 | 1.8, 3.4 | 2.8 | |
| C5 | _ | 179.7 | 176.5^{d} | _ | - | |
| C8 | - | 179.9 | 178.2^{d} | - | - | |

Notes: (a) calculated isotropic chemical shifts are determined from calculated chemical shieldings according to $\delta_{\rm calc} = \sigma_{\rm ref} - \sigma_{\rm calc}$, where $\sigma_{\rm ref}$ equals 30.0 ppm for ¹H and 163.2 ppm for ¹³C. (b) H-atom labels and calculated and experimental ¹H chemical shifts are presented in normal font for direct one-bond C—H connectivities, while longer-range C···H proximities (corresponding to cross peaks observed in the ¹H-¹³C spectra presented in Figs. 4b and 4c) are presented in italics. (c) For CH₃ groups, the calculated ¹H chemical shifts correspond to the average over the three H atoms. (d) Experimental chemical shifts taken from the ¹³C CP MAS spectrum (Fig. 2a) since no cross peaks are observed in the ¹H-¹³C spectra presented in Figs. 4(b) and 4(c). (e) Note that the C7-H1 and C6-H4 cross peaks due to longer-range C···H proximities cannot be distinguished from the C9-H1 and C12-H4 cross peaks due to one-bond C-H connectivities – in the stick spectrum in Fig. 2(c), open bars denote the calculated (GIPAW) C7 and C6 ¹³C chemical shifts.

(GIPAW) ¹H chemical shifts, as well as a 2D DQ spectrum in Fig. 3(*b*). In addition, Fig. 3(*c*) presents a ¹H-¹³C 2D correlation spectrum of the DI-PM cocrystal; note that this spectrum has been rotated through 90° from its usual representation such that the direct (¹³C) dimension is vertical. In this way, it is possible to directly compare (see vertical dashed lines) ¹H chemical shifts of peaks in the ¹H-¹³C (Fig. 3*c*) and ¹H DQ 2D (Fig. 3*b*) and ¹H 1D (Fig. 3*a*) spectra. Two separate spectral regions are presented in Fig. 3(*c*) corresponding to (top) the methyl resonances at a ¹³C chemical shift close to 25 ppm and (bottom) the aromatic CH resonances with ¹³C chemical shifts between 110 and 140 ppm.

The $^{1}\text{H}-^{13}\text{C}$ correlation spectrum in Fig. 3(c) was recorded using a short CP contact time of 100 µs to transfer magnetization from ^{1}H to ^{13}C , such that cross peaks correspond to one-bond C—H connectivities. The spreading of the resonances into two dimensions in Fig. 3(c) allows the identification of two and ten resolved cross peaks for the CH₃ and aromatic CH groups, respectively. The value of such a $^{1}\text{H}-^{13}\text{C}$ correlation spectrum in resolving and assigning the experimental ^{1}H chemical shifts is thus evident. Table 2 lists the calculated (GIPAW) and experimental ^{13}C chemical shifts

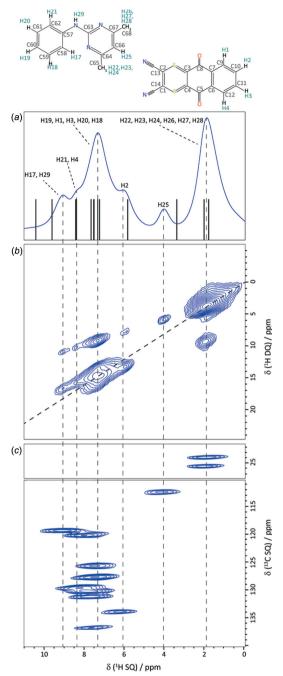


Figure 3 MAS NMR spectra of the DI-PM cocrystal, showing (a) a ¹H (600 MHz) MAS (60 kHz) one-pulse spectrum (16 transients were co-added for a recycle delay of 15 s), (b) a 2D ¹H (700 MHz) DQ MAS (60 kHz) spectrum (the dashed diagonal line indicates the $F_1 = 2F_2$ DQ-SQ diagonal) recorded using one rotor period of BABA recoupling (32 transients were co-added for each of 200 t₁ FIDs using a recycle delay of 6 s, corresponding to a total experiment time of 12 h) and (c) a ¹H (500 MHz)-13C HETCOR MAS (12.5 kHz) spectrum recorded using FSLG 1 H homonuclear decoupling in t_{1} and a short CP transfer duration of 100 µs (104 transients were co-added for each of 128 t₁ FIDs using a recycle delay of 6 s, corresponding to a total experimental time of 22 h). The vertical lines in part (a) correspond to calculated (GIPAW) 1 H chemical shifts. For the 1 H $^{-13}$ C NMR spectrum in part (c), two separate spectral regions are presented corresponding to methyl and aromatic C-H groups; note that this spectrum has been rotated through 90° from its usual representation [the 13 C dimension corresponds to direct (t_2) acquisition]. The base contour level is at (b) 7% and (c) 20% of the maximum peak height.

(sorted in order of increasing chemical shift). For directly bonded C—H connectivities, H-atom labels and calculated (GIPAW) and experimental ¹H chemical shifts are presented in normal font.

Fig. 4 compares ¹H-¹³C correlation spectra recorded with three different CP contact times of 100 µs (Fig. 4a), 500 µs (Fig. 4b) and 1 ms (Fig. 4c); Fig. 4(a) is a copy of Fig. 3(c), but presented in the normal orientation, *i.e.* with the direct (¹³C) dimension horizontal. It is evident that additional cross peaks are observed for longer CP contact times - these correspond to longer-range C···H proximities (see italics font in Table 2). Notably, cross peaks are observed at ¹³C chemical shifts of 141.5 (atom C57), 160.1 (atom C63) and 168.2 ppm (atoms C64) and C67); these all correspond to intramolecular proximities within the dithianon molecule, i.e. C57 with H17 (9.1 ppm, 2.16 Å), H21 (8.0 ppm, 2.16 Å) and H29 (9.1 ppm, 2.06 Å), C63 with H29 (9.1 ppm, 2.01 Å), C64 and C67 with H25 (4.0 ppm, 2.16 and 2.17 Å) and CH₃ protons (1.9 and 2.0 ppm, nearest distance 2.14 Å). Of most interest is the (160.1 ppm, 9.1 ppm) cross peak, which thus enables the determination of the NH ¹H chemical shift.

With all the ¹H chemical shifts assigned, let us re-examine the ¹H DQ MAS spectrum in Fig. 3(b). In such a spectrum, cross peaks are observed in the DQ dimension at the sum of the two single-quantum (SO) frequencies if there is a close proximity (typically up to 3.5 Å; Brown, 2007, 2012) between the corresponding two H atoms (a full listing of H...H proximities under 3.5 Å for the DI-PM cocrystal is given in Table S1 of the Supporting information). Consider the two lowest-ppm aromatic CH protons H25 (4.0 ppm) and H2 (6.2 ppm) for which distinct ¹H resonances are resolved in the 1 H SO dimension. For H25, the only DO peak is at 4.0 + 2.0 =6.0 ppm with the CH₃ protons, since H25 is sandwiched between two methyl-group substituents on the PM molecule. For H2, there is a DQ peak at 6.2 + 7.5 = 13.7 ppm corresponding to the intramolecular H···H proximity with the neighbouring H1 (7.4 ppm, 2.50 Å) and H3 (7.7 ppm, 2.47 Å) DI aromatic CH protons, as well as a DQ peak at 6.2 + 2.0 =8.2 ppm due to intermolecular proximities to the PM CH₃ H atoms (H23, H24, H28 and H22 at 2.90, 3.03, 3.12 and 3.12 Å, respectively). Considering the high-ppm region, DO cross peaks for the overlapping PI NH H29 (9.1 ppm) and aromatic CH H17 (9.1 ppm) resonances are observed at 9.1 + 7.7 =16.8 ppm for intramolecular H29···H21 (2.21 Å) and $H17 \cdots H18 \ (2.50 \,\text{Å})$ proximities, as well as at 9.1 + 2.0 =11.1 ppm for intermolecular proximities to PM methyl-group protons (closest distances of H17···H26 = 2.48 Å and $H29 \cdot \cdot \cdot H24 = 2.64 \text{ Å}$). For the other overlapping CH aromatic resonances, cross peaks due to intramolecular proximities with other CH aromatic resonances, as well as intermolecular proximities to the methyl protons, are also observed.

3.4. Comparison of experimental and calculated $^1\mathrm{H}$ and $^{13}\mathrm{C}$ chemical shifts

In the 1H – ^{13}C correlation spectra presented in Fig. 4, red crosses correspond to calculated (GIPAW) ^{13}C and 1H

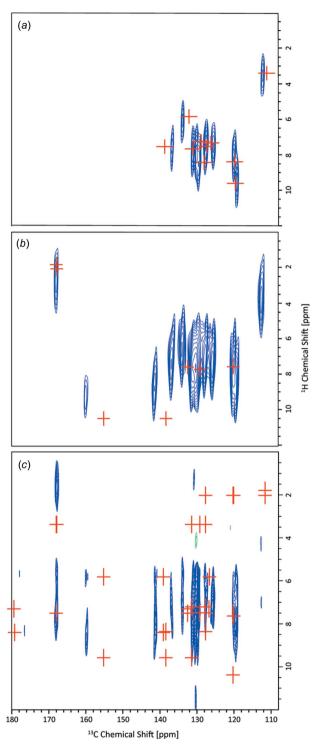


Figure 4 ¹H (500 MHz)–¹³C HETCOR MAS (12.5 kHz) spectra of the DI–PM cocrystal recorded using FSLG ¹H homonuclear decoupling (Bielecki *et al.*, 1989) in t_1 with a CP transfer duration of (a) 100 μ s, (b) 500 μ s and (c) 1 ms. The spectrum in part (a) is repeated from Fig. 3(c). 104 transients were co-added for each of (b) 128 or (c) 90 t_1 FIDs using a recycle delay of (b) 6 or (c) 5.5 s, corresponding to a total experimental time of (b) 22 or (c) 14 h. The scaling factor in F_1 was determined to be (a) and (b) 1.80 or (c) 1.73. The base contour level is at (a) 20, (b) 13 and (c) 25% of the maximum peak height. Red crosses correspond to GIPAW-calculated ¹H and ¹³C chemical shifts (see Table 2) for (a) one-bond C—H bonds and (b) and (c) C···H proximities between (b) 1.2 and 2.2 Å, and (c) 2.2 and 3.0 Å.

chemical shifts. Specifically, in Fig. 4(a), red crosses correspond to direct C-H one-bond connectivities (C-H distances under 1.2 Å), while in Figs. 4(b) and 4(c), red crosses are presented for C-H proximities between 1.2 and 2.2 Å (Fig. 4b), and between 2.2 and 3.0 Å (Fig. 4c). We comment here on the level of agreement between experimental and calculated (GIPAW) chemical shifts. Starting with a consideration of the aromatic CH moieties (see Fig. 4a and Table 2), the discrepancy between experiment and calculation is within 2 ppm for the ¹³C chemical shifts (except for C11, where the difference is 2.4 ppm); this corresponds to the established observation that the discrepancy is within 1% of the chemical shift range (~200 ppm for ¹³C chemical shifts of diamagnetic molecules). For the ¹H chemical shifts, while most are within the usual 0.3 ppm, some exhibit slightly larger discrepancies, notably 0.6 ppm for atoms H17 and H25.

For the two CH₃ groups (see Figs. 2 and 3a, and Table 2), there is excellent agreement for the ¹H chemical shifts (within 0.1 ppm), whereas the calculated ¹³C chemical shifts are both 8.5 ppm lower than the experimental values, although the experimental difference in ¹³C chemical shifts between atoms C65 and C68 of 1.8 ppm is reproduced by the calculation (difference of 1.9 ppm). The explanation for this is well known, namely, the gradient of a plot of experimental ¹³C chemical shifts against calculated shielding deviates slightly from -1 (Harris et al., 2007; Ashbrook & McKay, 2016), such that calculated ¹³C chemical shifts are too low and too high compared to experiment for low-ppm and high-ppm resonances if, as here (see Fig. 2), the gradient is constrained to -1and a single reference shielding is used. An alternative approach would be to use different reference shieldings for different regions of the spectrum (Webber, Emsley et al.,

Returning to the ¹H chemical shifts, the biggest discrepancy is for the NH proton (H29), where the calculated ¹H chemical shift of 10.5 ppm is 1.4 ppm higher than the experimental value of 9.1 ppm. Such a large difference is explained by a known temperature dependence (the experimental ¹H chemical shift increases upon reducing the temperature) for hydrogen-bonded protons (Brown *et al.*, 2001; Pickard *et al.*, 2007; Webber, Elena *et al.*, 2010), considering that the calculation corresponds to 0 K.

3.5. Calculated molecule-to-crystal changes in chemical shifts

For cases such as the DI–PM cocrystal in this article, an NMR crystallography study is able to provide new insight by means of a comparison of chemical shifts calculated for the full crystal structure with those calculated for an isolated molecule (as extracted from the geometry-optimized crystal structure) (Yates *et al.*, 2005; Schmidt *et al.*, 2006; Mafra *et al.*, 2012). Specifically, a molecule-to-crystal difference in chemical shift is indicative of a combination of intermolecular interactions, notably hydrogen bonding and ring currents due to $C-H\cdots\pi$ interactions, whereby the latter can be separately quantified by means of the nucleus independent chemical shift (NICS)

Table 3
Comparison of experimental ¹H chemical shifts with calculated^a (GIPAW) values (all in ppm) for the DI-PM cocrystal for the full crystal structure and an isolated dithianon or pyrimethanil molecule.

| Atom | $\delta_{ m exp}$ | $\delta_{ m crystal}$ | $\delta_{ m molecule}$ | $\Delta \delta_{ m crystal-molecule}$ |
|------------------------|-------------------|-----------------------|------------------------|---------------------------------------|
| H1 | 7.4 | 7.4 | 7.8 | -0.4 |
| H2 | 6.2 | 5.9 | 7.4 | -1.5 |
| H3 | 7.7 | 7.6 | 7.4 | 0.2 |
| H4 | 8.2 | 8.5 | 7.8 | 0.7 |
| H17 | 9.1 | 9.7 | 9.2 | 0.5 |
| H18 | 7.7 | 7.7 | 7.0 | 0.7 |
| H19 | 7.8 | 7.3 | 6.6 | 0.7 |
| H20 | 7.4 | 7.6 | 7.0 | 0.6 |
| H21 | 8.0 | 8.4 | 6.4 | 2.0 |
| H22/23/24 ^b | 1.9 | 1.8 | 1.9 | -0.1 |
| H25 | 4.0 | 3.4 | 6.1 | -2.7 |
| H26/27/28 ^b | 2.0 | 2.0 | 1.8 | 0.2 |
| H29 | 9.1 | 10.5 | 6.9 | 3.6 |

Notes: (a) calculated isotropic chemical shieldings are determined from calculated chemical shieldings according to $\delta_{\rm calc} = \sigma_{\rm ref} - \sigma_{\rm calc}$, where $\sigma_{\rm ref}$ equals 30.0 ppm; (b) for CH₃ groups, the calculated $^1{\rm H}$ chemical shifts correspond to the average over the three H atoms.

(von Ragué Schleyer et al., 1996; Sebastiani, 2006; Uldry et al., 2008; Mafra et al., 2012). Consider Table 3, which presents the change in ¹H chemical shift upon going from an isolated molecule to the full crystal, $\Delta \delta_{\text{crystal-molecule}}$, for the different H atoms in the DI-PM cocrystal. The largest positive change of 3.6 ppm is observed for the NH (H29) atom that is involved in an intermolecular N-H···O hydrogen bond to atom O1 (see Fig. 1a; the N···O and H···O distances are 2.95 and 1.96 Å, respectively, with a 162° N-H···O angle). Interestingly, $\Delta \delta_{crystal-molecule} = 2.0$ ppm for the aromatic CH H21 atom, for which Fig. 1(a) identifies an intermolecular $C-H \cdots O$ socalled weak hydrogen-bonding (Desiraju & Steiner, 1999; Yates et al., 2005; Uldry et al., 2008) interaction (the $C \cdot \cdot \cdot O$ and H···O distances are 3.24 and 2.35 Å, respectively, with a 138° C-H···O angle). The other H atoms, for which the magnitude of $\Delta \delta_{\text{crystal-molecule}}$ exceeds 1 ppm, are H25 (-2.7 ppm) and H2 (-1.6 ppm); as shown in Fig. 5, these marked changes

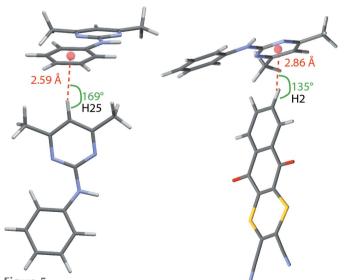


Figure 5 Schematic representations showing $C-H\cdots\pi$ interactions for aromatic atoms (a) H25 and (b) H2.

Table 4
Comparison of calculated (GIPAW) NMR chemical shieldings (in ppm) for the DI-PM cocrystal for the full crystal structure and an isolated dithianon or pyrimethanil molecule.

| Atom | $\sigma_{ m molecule}$ | $\sigma_{ m crystal}$ | $\sigma_{ m crystal-moleculo}$ |
|------|------------------------|-----------------------|--------------------------------|
| N1 | -106.4 | -88.9 | 17.5 |
| N2 | -107.2 | -88.9 | 18.3 |
| N9 | 98.9 | 91.4 | -7.4 |
| N10 | -30.1 | -33.0 | -2.9 |
| N11 | -44.5 | -42.4 | 2.1 |
| O1 | -363.4 | -265.9 | 97.5 |
| O2 | -345.3 | -322.2 | 23.1 |
| S1 | 330.7 | 305.6 | -25.1 |
| S2 | 333.8 | 320.6 | -13.2 |

in the 1 H chemical shift are a consequence of ring current effects associated with the proton pointing towards the centre of a six-membered aromatic ring of a nearby PM molecule in a $C-H\cdots\pi$ interaction, as has been noted previously in a number of other cases (Brouwer *et al.*, 2008; Mafra *et al.*, 2012; Brown, 2012).

In the above discussion in §3.1, a close $S \cdots O$ distance, equal to 3.10 Å, between the O2 and S2 atoms of neighbouring DI molecules was noted; this is less than the sum of the van der Waals radii (3.32 Å) (Beno et al., 2015; Zhang et al., 2015). Indeed, there is a growing literature discussing S···O interactions (Burling & Goldstein, 1992; Iwaoka et al., 2002; Beno et al., 2015). While we have not carried out ¹⁷O or ³³S solidstate NMR experiments as part of this study, an NMR crystallography approach enables the effect of such a putative S...O interaction on the oxygen and sulfur NMR chemical shieldings to be investigated by means of the GIPAW calculation that reports on all nuclei in the solid-state structure. An inspection of Table 4 shows that it is interesting that $\Delta \delta_{ ext{crystal-molecule}}$ (note that this is the negative of the difference in calculated absolute shielding, with the latter being stated in Table 4) is much larger for O1 (-98 ppm), which is involved in a N-H···O intermolecular hydrogen bonding, as compared to that for O2 (-23 ppm). Moreover, the change for S2 (13 ppm) is less than that for S1 (25 ppm), with both changes being small, though there is limited information on the range of experimentally observed solid-state NMR 33S chemical shifts (Hansen et al., 2008). We conclude that even though there is a close intermolecular $S \cdots O$ distance of 3.10 Å in the DI-PM cocrystal, there is not a marked effect on the calculated NMR chemical shieldings for the O2 and S2 nuclei.

4. Summary

In summary, we have presented here an NMR crystallography study of an agrochemical cocrystal. Specifically in combination with a GIPAW calculation of the NMR shieldings, $^1H^{-13}C$ 2D correlation spectra enable the resolution and assignment of the NH, aromatic CH and methyl resonances for the DI-PM cocrystal, while specific intra- and intermolecular $H\cdots H$ proximities are identified in a 1H DQ MAS spectrum. The performing of separate GIPAW calculations for the full crystal

structure and isolated DI and PM molecules yields the change in the NMR chemical shift upon going from the molecule to the crystal structure, thus allowing the quantitation of specific $N-H\cdots O$, $C-H\cdots O$ and $C-H\cdots \pi$ interactions.

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Single-crystal X-ray diffraction and NMR crystallography of a 1:1 cocrystal of dithianon and pyrimethanil

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Computing details

Data collection: *CrysAlis PRO* (Agilent, 2014); cell refinement: *CrysAlis PRO* (Agilent, 2014); data reduction: *CrysAlis PRO* (Agilent, 2014); program(s) used to solve structure: SUPERFLIP (Palatinus & Chapuis, 2007); program(s) used to refine structure: *CRYSTALS* (Betteridge *et al.*, 2003); molecular graphics: *CAMERON* (Watkin *et al.*, 1996) and *Mercury* (Macrae *et al.*, 2006); software used to prepare material for publication: *CRYSTALS* (Betteridge *et al.*, 2003).

5,10-Dioxo-5*H*,10*H*-naphtho[2,3-*b*][1,4]dithiine-2,3-dicarbonitrile–4,6-dimethyl-*N*-phenylpyrimidin-2-amine (1/1)

Crystal data

 $C_{14}H_4N_2O_2S_2 \cdot C_{12}H_{13}N_3$ $M_r = 495.59$ Monoclinic, $P2_1/n$ Hall symbol: -P 2yn a = 7.1707 (2) Å b = 22.8006 (6) Å c = 13.8237 (4) Å $\beta = 97.047$ (3)° V = 2243.04 (7) Å³ Z = 4

Data collection

Agilent Xcalibur Onyx Ultra diffractometer Mirror monochromator $\omega/2\theta$ scans Absorption correction: multi-scan (CrysAlis PRO; Agilent, 2014) $T_{\min} = 0.596$, $T_{\max} = 1.000$ 5143 measured reflections

Refinement

Refinement on F^2 Least-squares matrix: full $R[F^2 > 2\sigma(F^2)] = 0.045$ $wR(F^2) = 0.094$ S = 0.983141 reflections F(000) = 1024 $D_x = 1.467 \text{ Mg m}^{-3}$ $Cu \ K\alpha \text{ radiation, } \lambda = 1.54184 \text{ Å}$ Cell parameters from 2711 reflections $\theta = 5.0-62.6^{\circ}$ $\mu = 2.45 \text{ mm}^{-1}$ T = 100 KPlate, purple $0.60 \times 0.10 \times 0.02 \text{ mm}$

3160 independent reflections 2667 reflections with $I > 2.0\sigma(I)$ $R_{\text{int}} = 0.035$ $\theta_{\text{max}} = 58.9^{\circ}, \ \theta_{\text{min}} = 3.2^{\circ}$ $h = -5 \rightarrow 7$ $k = -25 \rightarrow 25$ $l = -14 \rightarrow 15$

109 parameters
3 restraints
Primary atom site location: other
Hydrogen site location: difference Fourier map
H atoms treated by a mixture of independent
and constrained refinement

Method, part 1, Chebychev polynomial [weight] $= 1.0/[A_0*T_0(x) + A_1*T_1(x) \cdots + A_{n-1}]*T_{n-1}(x)]$ where A_i are the Chebychev coefficients listed below and x = F/Fmax Method = Robust Weighting $W = [weight] * [1-(deltaF/6*sigmaF)^2]^2 A_i$ are: 0.138E+04 0.207E+04 0.111E+04 326. $(\Delta/\sigma)_{max} = 0.001$ $\Delta\rho_{max} = 0.43$ e Å $^{-3}$ $\Delta\rho_{min} = -0.37$ e Å $^{-3}$

Special details

Experimental. The crystal was placed in the cold stream of an Oxford Cryosystems open-flow nitrogen cryostat (Cosier & Glazer, 1986) with a nominal stability of 0.1K. Cosier, J. & Glazer, A.M., 1986. J. Appl. Cryst. 105-107.

Fractional atomic coordinates and isotropic or equivalent isotropic displacement parameters (\mathring{A}^2)

| | x | у | Z | $U_{ m iso}$ */ $U_{ m eq}$ | |
|-----|--------------|--------------|--------------|-----------------------------|--|
| S1 | 0.57630 (11) | 0.51819 (3) | 0.79844 (6) | 0.0223 | |
| C2 | 0.3685 (4) | 0.48079 (13) | 0.7529 (2) | 0.0212 | |
| C3 | 0.2072 (4) | 0.47646 (13) | 0.7926(2) | 0.0202 | |
| S4 | 0.14929 (11) | 0.50645 (3) | 0.90297 (5) | 0.0206 | |
| C5 | 0.3411 (4) | 0.55384 (12) | 0.9352(2) | 0.0177 | |
| C6 | 0.5006 (4) | 0.55878 (12) | 0.8951 (2) | 0.0175 | |
| C7 | 0.6476 (4) | 0.60247 (12) | 0.9342(2) | 0.0179 | |
| O8 | 0.7809(3) | 0.61031 (9) | 0.88850 (15) | 0.0221 | |
| C9 | 0.6255 (4) | 0.63243 (12) | 1.0265 (2) | 0.0167 | |
| C10 | 0.4604 (4) | 0.62563 (12) | 1.0704(2) | 0.0176 | |
| C11 | 0.3051 (4) | 0.58862 (13) | 1.0216(2) | 0.0176 | |
| O12 | 0.1533 (3) | 0.58453 (9) | 1.05284 (15) | 0.0237 | |
| C13 | 0.4420 (4) | 0.65186 (13) | 1.1586 (2) | 0.0214 | |
| C14 | 0.5889 (5) | 0.68550 (14) | 1.2043 (2) | 0.0247 | |
| C15 | 0.7515 (5) | 0.69313 (13) | 1.1606 (2) | 0.0235 | |
| C16 | 0.7717 (4) | 0.66662 (13) | 1.0719 (2) | 0.0211 | |
| C17 | 0.0513 (5) | 0.44419 (13) | 0.7438 (2) | 0.0227 | |
| N18 | -0.0763(4) | 0.41868 (13) | 0.7075 (2) | 0.0334 | |
| C19 | 0.3871 (5) | 0.45136 (14) | 0.6633 (2) | 0.0244 | |
| N20 | 0.4065 (4) | 0.42677 (13) | 0.5922(2) | 0.0372 | |
| N21 | 0.1269 (3) | 0.65896 (11) | 0.81861 (18) | 0.0185 | |
| C22 | 0.1456 (4) | 0.63048 (12) | 0.7301 (2) | 0.0193 | |
| C23 | 0.3046 (5) | 0.63170 (13) | 0.6818 (2) | 0.0227 | |
| C24 | 0.3041 (5) | 0.60183 (14) | 0.5940(2) | 0.0281 | |
| C25 | 0.1478 (5) | 0.57112 (14) | 0.5532(2) | 0.0305 | |
| C26 | -0.0103(5) | 0.56959 (14) | 0.6017 (2) | 0.0289 | |
| C27 | -0.0132(5) | 0.59861 (13) | 0.6889 (2) | 0.0227 | |
| C28 | 0.2398 (4) | 0.69992 (12) | 0.8710(2) | 0.0172 | |
| N29 | 0.4071 (3) | 0.71395 (10) | 0.84429 (17) | 0.0189 | |
| C30 | 0.5047 (4) | 0.75525 (13) | 0.8996 (2) | 0.0204 | |
| C31 | 0.4355 (4) | 0.78043 (13) | 0.9782(2) | 0.0240 | |

| C32 | 0.2624 (5) | 0.76203 (13) | 1.0009 (2) | 0.0233 |
|------|------------|--------------|--------------|---------|
| N33 | 0.1612 (3) | 0.72147 (11) | 0.94755 (18) | 0.0203 |
| C34 | 0.1768 (5) | 0.78633 (16) | 1.0859 (3) | 0.0350 |
| C35 | 0.6919 (4) | 0.77160 (14) | 0.8698 (2) | 0.0256 |
| H131 | 0.3325 | 0.6473 | 1.1887 | 0.0272* |
| H141 | 0.5773 | 0.7037 | 1.2645 | 0.0304* |
| H151 | 0.8471 | 0.7168 | 1.1907 | 0.0275* |
| H161 | 0.8807 | 0.6722 | 1.0421 | 0.0261* |
| H231 | 0.4128 | 0.6519 | 0.7083 | 0.0274* |
| H241 | 0.4121 | 0.6020 | 0.5619 | 0.0339* |
| H251 | 0.1519 | 0.5519 | 0.4941 | 0.0370* |
| H261 | -0.1170 | 0.5500 | 0.5748 | 0.0342* |
| H271 | -0.1197 | 0.5972 | 0.7217 | 0.0260* |
| H311 | 0.5042 | 0.8085 | 1.0170 | 0.0294* |
| H342 | 0.0443 | 0.7782 | 1.0832 | 0.0544* |
| H341 | 0.1920 | 0.8277 | 1.0869 | 0.0549* |
| H343 | 0.2423 | 0.7710 | 1.1438 | 0.0548* |
| H352 | 0.7389 | 0.8070 | 0.9003 | 0.0411* |
| H353 | 0.6847 | 0.7771 | 0.8015 | 0.0418* |
| H351 | 0.7800 | 0.7420 | 0.8872 | 0.0415* |
| H211 | 0.022(3) | 0.6531 (11) | 0.8421 (16) | 0.0231* |
| | | | | |

Atomic displacement parameters (\mathring{A}^2)

| | U^{11} | U^{22} | U^{33} | U^{12} | U^{13} | U^{23} |
|-----|----------|----------|----------|----------|----------|----------|
| S1 | 0.0223 | 0.0220 | 0.0224 | -0.0001 | 0.0015 | -0.0055 |
| C2 | 0.0274 | 0.0150 | 0.0197 | 0.0019 | -0.0035 | 0.0014 |
| C3 | 0.0236 | 0.0150 | 0.0203 | -0.0003 | -0.0043 | 0.0020 |
| S4 | 0.0215 | 0.0185 | 0.0211 | -0.0044 | 0.0004 | -0.0018 |
| C5 | 0.0202 | 0.0125 | 0.0189 | 0.0022 | -0.0038 | 0.0041 |
| C6 | 0.0214 | 0.0132 | 0.0169 | 0.0009 | -0.0021 | 0.0022 |
| C7 | 0.0197 | 0.0133 | 0.0204 | 0.0045 | 0.0009 | 0.0041 |
| O8 | 0.0219 | 0.0229 | 0.0214 | -0.0035 | 0.0027 | -0.0011 |
| C9 | 0.0188 | 0.0127 | 0.0171 | 0.0016 | -0.0041 | 0.0010 |
| C10 | 0.0199 | 0.0135 | 0.0184 | 0.0016 | -0.0015 | 0.0037 |
| C11 | 0.0187 | 0.0143 | 0.0198 | 0.0015 | 0.0023 | 0.0054 |
| O12 | 0.0250 | 0.0224 | 0.0244 | -0.0031 | 0.0062 | 0.0008 |
| C13 | 0.0242 | 0.0199 | 0.0206 | 0.0015 | 0.0041 | 0.0008 |
| C14 | 0.0338 | 0.0223 | 0.0175 | -0.0007 | 0.0016 | -0.0052 |
| C15 | 0.0277 | 0.0184 | 0.0225 | -0.0036 | -0.0042 | -0.0039 |
| C16 | 0.0206 | 0.0174 | 0.0244 | 0.0004 | -0.0003 | 0.0021 |
| C17 | 0.0271 | 0.0205 | 0.0195 | -0.0001 | -0.0013 | 0.0002 |
| N18 | 0.0413 | 0.0324 | 0.0255 | -0.0073 | 0.0002 | -0.0019 |
| C19 | 0.0281 | 0.0203 | 0.0244 | 0.0027 | 0.0018 | 0.0001 |
| N20 | 0.0423 | 0.0377 | 0.0314 | 0.0023 | 0.0037 | -0.0076 |
| N21 | 0.0157 | 0.0194 | 0.0204 | -0.0024 | 0.0017 | -0.0019 |
| C22 | 0.0294 | 0.0114 | 0.0161 | 0.0038 | -0.0013 | 0.0021 |
| C23 | 0.0284 | 0.0181 | 0.0215 | 0.0000 | 0.0027 | 0.0018 |

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| C24 | 0.0429 | 0.0202 | 0.0223 | 0.0053 | 0.0090 | 0.0021 |
|-----|--------|--------|--------|---------|---------|---------|
| C25 | 0.0517 | 0.0213 | 0.0177 | 0.0021 | 0.0016 | -0.0017 |
| C26 | 0.0429 | 0.0167 | 0.0242 | -0.0026 | -0.0075 | -0.0005 |
| C27 | 0.0289 | 0.0183 | 0.0198 | 0.0015 | -0.0020 | 0.0034 |
| C28 | 0.0207 | 0.0113 | 0.0185 | 0.0042 | -0.0016 | 0.0037 |
| N29 | 0.0213 | 0.0152 | 0.0199 | 0.0012 | 0.0007 | 0.0033 |
| C30 | 0.0215 | 0.0145 | 0.0238 | 0.0007 | -0.0033 | 0.0047 |
| C31 | 0.0283 | 0.0158 | 0.0267 | -0.0028 | -0.0013 | -0.0036 |
| C32 | 0.0270 | 0.0190 | 0.0235 | 0.0016 | 0.0019 | -0.0027 |
| N33 | 0.0224 | 0.0175 | 0.0210 | 0.0013 | 0.0029 | -0.0031 |
| C34 | 0.0366 | 0.0326 | 0.0373 | -0.0031 | 0.0103 | -0.0140 |
| C35 | 0.0244 | 0.0239 | 0.0278 | -0.0050 | 0.0004 | 0.0034 |
| | | | | | | |

Geometric parameters (Å, °)

| Geometrie parameters (11, | | | |
|---------------------------|-------------|--------------|------------|
| S1—C2 | 1.764 (3) | N21—H211 | 0.864 (17) |
| S1—C6 | 1.764 (3) | C22—C23 | 1.391 (4) |
| C2—C3 | 1.343 (4) | C22—C27 | 1.410 (4) |
| C2—C19 | 1.429 (4) | C23—C24 | 1.390 (4) |
| C3—S4 | 1.767 (3) | C23—H231 | 0.938 |
| C3—C17 | 1.436 (4) | C24—C25 | 1.382 (5) |
| S4—C5 | 1.763 (3) | C24—H241 | 0.939 |
| C5—C6 | 1.336 (4) | C25—C26 | 1.387 (5) |
| C5—C11 | 1.483 (4) | C25—H251 | 0.930 |
| C6—C7 | 1.502 (4) | C26—C27 | 1.378 (4) |
| C7—O8 | 1.223 (4) | C26—H261 | 0.923 |
| C7—C9 | 1.473 (4) | C27—H271 | 0.936 |
| C9—C10 | 1.404 (4) | C28—N29 | 1.336 (4) |
| C9—C16 | 1.393 (4) | C28—N33 | 1.350 (4) |
| C10—C11 | 1.491 (4) | N29—C30 | 1.353 (4) |
| C10—C13 | 1.379 (4) | C30—C31 | 1.375 (4) |
| C11—O12 | 1.222 (4) | C30—C35 | 1.499 (4) |
| C13—C14 | 1.390 (4) | C31—C32 | 1.382 (5) |
| C13—H131 | 0.938 | C31—H311 | 0.935 |
| C14—C15 | 1.388 (5) | C32—N33 | 1.339 (4) |
| C14—H141 | 0.944 | C32—C34 | 1.498 (4) |
| C15—C16 | 1.390 (4) | C34—H342 | 0.964 |
| C15—H151 | 0.931 | C34—H341 | 0.950 |
| C16—H161 | 0.936 | C34—H343 | 0.944 |
| C17—N18 | 1.146 (4) | C35—H352 | 0.952 |
| C19—N20 | 1.155 (4) | C35—H353 | 0.949 |
| N21—C22 | 1.406 (4) | C35—H351 | 0.935 |
| N21—C28 | 1.382 (4) | | |
| | | | |
| C2—S1—C6 | 101.45 (14) | N21—C22—C27 | 115.5 (3) |
| S1—C2—C3 | 128.4 (2) | C23—C22—C27 | 119.0(3) |
| S1—C2—C19 | 111.8 (2) | C22—C23—C24 | 119.6 (3) |
| C3—C2—C19 | 119.7 (3) | C22—C23—H231 | 120.6 |
| | | | |

| 119.7 121.3 (3) 119.7 118.9 119.0 (3) 119.1 |
|--|
| 119.7 118.9 119.0 (3) |
| 118.9 119.0 (3) |
| 119.0 (3) |
| |
| 119.1 |
| |
| 121.9 |
| 120.8 (3) |
| 120.2 |
| 118.9 |
| 120.2 (3) |
| 119.3 |
| 120.5 |
| 120.3 (3) |
| 112.5 (3) |
| 127.2 (3) |
| 115.5 (3) |
| 121.7 (3) |
| 115.9 (3) |
| 122.4 (3) |
| 118.3 (3) |
| 121.5 |
| 120.2 |
| 121.7 (3) |
| 122.1 (3) |
| 116.2 (3) |
| 115.6 (3) |
| 113.2 |
| 108.8 |
| 107.6 |
| 108.7 |
| 110.3 |
| 108.1 |
| 111.7 |
| 111.4 |
| 107.6 |
| 110.5 |
| |
| 107.8 |
| |
| |

Hydrogen-bond geometry (Å, °)

| <i>D</i> —H··· <i>A</i> | <i>D</i> —Н | HA | D··· A | <i>D</i> —H··· <i>A</i> |
|-------------------------|-------------|------|-----------|-------------------------|
| C23—H231···N29 | 0.94 | 2.36 | 2.950 (4) | 121 (1) |

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| C27—H271···O8 ⁱ | 0.94 | 2.51 | 3.296 (4) | 141 (1) |
|----------------------------|------|------|-----------|---------|
| N21—H211···O8i | 0.86 | 2.15 | 2.985 (4) | 162 (2) |

Symmetry code: (i) x-1, y, z.