- 1 Uranyl binding to humic acid under conditions relevant to cementitious
- 2 geological disposal of radioactive wastes
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18 Abstract

19 Few studies have sought to investigate specifically the potential for dissolved organic 20 matter (DOM) to bind (and thus potentially transport) radionuclides under the high pH 21 regimes expected from cementitious disposal. Here, we have used equilibrium 22 dialysis to investigate uranyl binding to humic acid over a pH range of ~10 to 13. The 23 experimental results provide evidence that DOM can bind uranyl ions over this pH 24 range, including in the presence of competing ions. There is a general decrease in 25 binding with increasing pH, from ~80% of total uranyl bound at pH 9.8 to ~10% at pH 26 12.9. Modelling of the system with WHAM/Model VII can yield representative results 27 up to pH ~10.5.

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29 Introduction

30 Strategies for disposal of intermediate level radioactive wastes (ILW) in several 31 countries involve geological emplacement, followed by addition of a cementitious 32 backfill. The aim is to provide a chemical barrier that retards radionuclide transport 33 from the geological disposal facility (GDF). Aqueous phase reactions, such as 34 dissolution, leaching and associated processes create a high pH environment within 35 the interstitial waters of the cement (Glasser et al., 2008). These reactions induce gradual changes to the composition of the interstitial water, from typical "young" 36 37 concrete leachate with a pH above 13 to more evolved leachate with a pH lower than 38 10 (Jacques et al., 2010).

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40 Natural dissolved organic matter (DOM) is ubiquitous in aquatic environments and is 41 present in groundwaters with concentrations dependent upon the host rock 42 environment and recharge source and pathway. It exerts a powerful control on 43 chemical conditions in waters and soils, influencing the transport and retention of 44 metals (Tipping et al., 2011). Binding of cationic metals to DOM can result in 45 increased solubility of metal salts and transport of DOM may result in simultaneous 46 transport of bound metals. The influence of these processes on radionuclide 47 migration has been recognised as a significant source of uncertainty in radioactive 48 waste management (Glaus et al., 1997).

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50 Few studies have sought to investigate specifically the potential for DOM to bind (and 51 thus potentially transport) radionuclides under the high pH regimes expected from 52 cementitious disposal. Here, we have used equilibrium dialysis to investigate uranyl 53 binding to humic acid over a pH range of ~9 to 13. We then compare the results with 54 predictions from a widely used chemical speciation model to assess the potential for 55 modelling to predict satisfactory results at high pH.

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57 Experimental

58 Reagents

59 Solutions were prepared using the following reagents; NaCl and Na₂CO₃ 60 (TraceSelect, Fluka Analytical), CuCl₂·H₂O (≥99.999%; Aldrich), CaCl₂ (99.9%; Alfa 61 Aesar). Humic acid was the reference material Suwanee River Humic acid (SRHA) 62 obtained from the International Humic Substances Society. Any pH adjustments were 63 undertaken using 1 M solutions of HCI (prepared from 34% NORMATOM grade, VWR) or NaOH (freshly prepared for each use from 99.99% purity salt; Alfa Aesar). 64 All experiments were performed in acid washed (10% v/v HNO₃) 180 ml 65 polypropylene pots. U-238 was prepared by dissolving UO₂(NO₃)₂ salt into 1 M HCI 66 and subsequent dilution to achieve a 10⁻⁷ mol L⁻¹ stock. U-232, used as a tracer 67 68 (stock concentration 400 Bq mL⁻¹), was separated from active daughter nuclides with a UTEVA column (Triskem International) using a previously reported method 69 70 (Horwitz et al., 1992).

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72 Instrumental analysis

73 Measurement of pH was performed using a Mettler Toledo Seven Multi meter and a 74 Mettler Toledo InLab Routine Pro electrode. A five point calibration was performed 75 using the following buffers; NIST traceable pH 4, 7 and 10; pH 9 (Scientific 76 Laboratory Supplies, UK), and pH 13 (Fluka Analytical). Calibration slopes below 77 98% were rejected and the calibration procedure repeated until this value was 78 achieved or exceeded. Calibration was carried out before each set of measurements. 79 Samples for liquid scintillation counting (Wallac Quantulus 1220 ultra-low level liquid 80 scintillation spectrometer) were prepared by addition of 1 mL of 1 M HCl to 1 mL of 81 sample, followed by 10 mL of ScintiSafe 3 cocktail (Fisher Scientific).

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85 Equilibrium dialysis procedure

In the dialysis experiments, the outer solution had a total volume of 150 mL, and at 86 87 the start of the experiment contained the HA and uranyl. The dialysate solution was 88 contained within pre-cleaned regenerated cellulose dialysis membrane tubing, with a 89 molecular weight cut-off of 1000 Da (SpectraPor 7) and sealed with SpectraPor clips 90 at both ends. The volume was 4 mL, and initially the ionic strength and pH were the 91 same as the outer solution, but with no HA or uranyl. The procedure differs in several ways from previously described methods (e.g., Glaus et al., 1995). The relative 92 93 volumes of dialysate and bulk solution differed significantly and the total 94 concentration of HA was lower. No additional competing ligands were added as OH⁻ 95 performs this function at high pH. This is required for many elements at lower pH 96 values to ensure that there are measurable fractions of radionuclides in both the 97 dialysate and outer fractions.

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The outer solution was prepared as two discrete solutions that were added to the 99 100 experiment simultaneously. These solutions contained HA or uranyl, with the pH and 101 ionic strength adjusted to those required for the experiment. This was done in order 102 to control the pH conditions under which the uranyl was added to the humic acid. 103 This prevents potential kinetic effects associated with dissociation of uranyl-humic 104 complexes if the pH of a binary mixture is changed. Where Cu, Ca or carbonate were added to experiments, these were included within the uranyl solution, and added so 105 that the final experimental concentrations were 10⁻⁷, 10⁻⁴ and 10⁻⁴ mol L⁻¹, 106 107 respectively. Once the solution was added to the airtight pot containing the dialysis 108 bag, the container was additionally sealed with laboratory film to further prevent 109 ingress of CO₂, and left to stand for 7 days. Solutions were prepared under ambient 110 laboratory conditions but in order to exclude carbonates as much as possible during 111 the preparation, freshly drawn MQ water was used throughout, prepared solutions 112 were stored in air evacuated containers before use and the experiments were

prepared in the shortest time possible (these effects are quantitatively evaluated inthe results and discussion section).

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116 Calculation of humic bound fraction of uranyl

The total uranyl in solution is represented by the sum of the inorganic ($[UO_2']$) and organic fractions, with the concentration of the uranyl humic complex ($[UO_2-HA]$) related to the amount bound (UO_2-HA ; mol mg⁻¹) and the humic acid concentration ([HA] mg L⁻¹). The concentration in the two compartments will be directly related to the measured tracer activity (A_{bag} and A_{bulk} ; Bq) and can be expressed as Equations 1 and 2, assuming equal binding properties of the humic acid within and outside the dialysis membrane.

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$$A_{\text{bag}} \propto [\text{UO}_2^{2^+}]_{\text{Tot-bag}} = \text{UO}_2 - \text{HA} \times [\text{HA}]_{\text{bag}} + [\text{UO}_2']$$
 (1)

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$$A_{\text{bulk}} \propto [\text{UO}_2^{2^+}]_{\text{Tot-bulk}} = \text{UO}_2 - \text{HA} \times [\text{HA}]_{\text{bulk}} + [\text{UO}_2']$$
 (2)

Where [HA]_{bag} and [HA]_{bulk} represent the humic acid concentration within and outside the dialysis membrane, respectively. Considering measured tracer activity and subtracting Equation 1 from 2, yields Equation 3.

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$$A_{UO2-HA} = (A_{bulk} - A_{bag})/([HA]_{bulk} - [HA]_{bag})$$
 (3)

130 Where A_{UO2-HA} can be considered as the tracer activity contribution of the humic 131 bound uranyl per mg L⁻¹ of HA in the experiment.

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In order to calculate the organic bound fraction of uranyl, it is first necessary to
calculate the tracer activity of the inorganic fraction (Equation 4). Using data from
either the bulk solution or the dialysate results in the same value.

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$$A_{\text{inorganic}} = A_{\text{bulk}} - A_{\text{UO2-HA}} \times [\text{HA}]_{\text{bulk}}$$
(4)

137 The fraction of humic bound metal (
$$f_{\text{organic}}$$
) can then be calculated from Equation 5.

138 $f_{\text{organic}} = A_{\text{UO2-HA}} \times [\text{HA}]_{\text{effective}} / (A_{\text{UO2-HA}} \times [\text{HA}]_{\text{effective}} + A_{\text{inorganic}})$ (5)

Where [HA]_{effective} is the effective mean concentration of HA when the total
experimental volume is considered.

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142 Contribution of regenerated cellulose to the UV measured [HA]

143 Humic acid is measured using UV visible spectrophotometry, and the concentrations in this work are 2 mg L⁻¹ in the outer solution and typically ≤ 0.25 mg L⁻¹ in the 144 145 dialysate at the end of the experiment (measurements are made on unmodified 146 solutions at the end of the experiments). The presence of low levels of regenerated 147 cellulose degradation products yields absorbance signals in the same range as those 148 used for the determination of humic acid. Where high concentrations of HA are used, 149 this contribution will be negligible, however, this is not the case at the concentrations 150 found within the dialysate (no contribution from these degradation products is 151 detected by UV absorbance increases to the higher volume bulk solution). In order to 152 account for this contribution, experiments in the absence of HA determined the UV 153 visible absorbance from degradation of regenerated cellulose. A subsection of 27 154 experiments were selected to measure HA absorbance values within the dialysate 155 (due to the low signals in these dilute HA solutions we used the mean absorbance 156 over the wavelength range 360-410 nm). These data are then used to give a range of 157 values for the HA within the dialysate ([HA]_{bag}) which are reflected as error bars where the data are presented. 158

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160 Speciation modelling

The Windermere Humic Aqueous Model (WHAM; Tipping, 1994) is a widely used chemical speciation code used in geochemical and ecotoxicological research and in regulation. The sub-model dealing with natural organic matter binding has recently been updated (Model VII; Tipping et al, 2011) to include the most up to date information on humic and fulvic acid binding of protons and metal cations (40 cationic species). This also includes improved prediction of cation binding at higher pH than

167 the previous version, due to increased numbers of available data and stricter data 168 selection criteria than previously applied. Data are now included for proton binding 169 data up to pH ~10.5. Additional radionuclide cations have also been added to the 170 database (Stockdale et al, 2011). The uranyl binding constant is an average based 171 upon separate values derived from six experimental datasets on a range of humic 172 and fulvic acid types and a reference value based on linear free energy relationships 173 (see Tipping et al, 2011, including supporting information, for full details). It is 174 important to highlight that the Humic Ion Binding Models are first parameterised for protons. Only then are the metal datasets fitted using the generic proton conditions 175 (rather than for each specific humic or fulvic acid type). Thus, the aim of the model is 176 177 to provide a predictive capacity for a 'generic' humic or fulvic acid in a natural system. 178 Binding constants for specific humic or fulvic acid types, if available, can be substituted into the model calculations, depending upon the users requirements. 179 180 However, there are no data available for the HA used in this work and thus, we use 181 the model generic binding value. The model, subsequently referred to as 182 WHAM/Model VII, is used with default values with the exception of the uranyl 183 hydrolysis stability constants that are taken from the compilation of.Duro et al (2006). 184 Errors within the model predictions are assessed by varying the humic binding 185 constants by log ±0.3 from their default values (Lofts and Tipping, 2011).

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187 Results and discussion

188 Contribution of regenerated cellulose to the UV measured [HA]

The upper recommended pH limit for the dialysis membrane is 12. However, no increased absorbance (indicating degradation of the cellulose) was seen within the dialysate volume when exposed to pH 13 solutions for the same time interval as the HA experiments. Based upon the results of the UV-visible spectrophotometry study on the dialysate membrane only samples and the 27 HA dialysate and bulk solution samples, a mean concentration of dialysate HA was determined, as well as a 90% 195 confidence interval. The HA concentration within the bag had a mean of 4.2% of the 196 bulk solution value and a range of 0% to 13.3%. To illustrate the uncertainly in the 197 presented data, the upper and lower limits were used to obtain error ranges and these data presented as error bars. Uncertainly from the HA concentration within the 198 199 dialysate typically results in ranges of bound metal that are within <10% of the mean value. No statistically significant variations in the shape of the absorbance profiles 200 201 was observed for the different samples over the pH range of the experiment, nor 202 between the bulk solution and dialysate samples.

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204 Uranyl binding to humic acid in a binary system

205 The majority of studies of natural organic matter binding to trace cations present 206 results in the form of complexation constants that relate the humic bound metal to the free ion concentration of the cation and the concentration or proton exchange 207 208 capacity of the organic matter. For uranyl in the circumneutral and slightly acidic 209 range, the organic bound fraction will dominate and reporting of experimental results 210 in terms of log complexation constants enables small variations caused by different 211 experimental conditions (e.g., ionic strength) to be assessed quantitatively. As the pH 212 increases, hydrolysis products of metal ions become more prevalent and 213 consequently decrease the free ion concentration. Changes in complexation 214 constants due to hydrolysis induced changes to the free ion concentrations may 215 obfuscate variations caused by subtle changes in organic bound metal in high pH 216 systems. For example, if the free ion decreases by orders of magnitude but the 217 organic bound changes by a factor of <10, changes to the complexation constant will 218 largely reflect the changes to the free ion concentrations. With this in mind, we report 219 results in terms of the percentage of total metal or the concentration that is bound to 220 the humic acid. Equations 1-5 reflect the fact that we are considering inorganic 221 components of the system as a whole, rather than the constituent complexes as is 222 the case in other studies (Glaus et al., 1997).

Figure 1A shows the change in relative concentrations of humic bound uranyl with pH. There is a general decrease in binding with increasing pH from ~80% of the total bound at pH 9.8 to ~10% at pH 12.9. Previous data have been obtained for uranyl binding at pH \leq 10 in the presence of competing ligands (Glaus et al, 1997). However, this is the first time that it has been practically demonstrated that binding can be significant over the pH 10 to 13 range. These are the conditions that are expected during the evolution of a cemetitious GDF.

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Previous studies have examined uranyl complexation with HA up to pH 10 (Zeh et al, 232 233 1997; Glaus et al, 1997). Zeh et al (1997) performed experiments on pH adjusted 234 natural Gorleben groundwaters to which additional uranyl was added. They observed binding in the absence of carbonate of 5.9% at pH 8.8 and 1% at pH 10.1. Glaus et 235 al (1997) reported results of 60 binding experiments at a range of ionic strengths, 236 237 organic acid types (purified Aldrich humic acid, Laurentian soil fulvic acid and 238 Suwannee River fulvic acid), and ratios of uranyl to humic/fulvic acid, performed in the presence of 0.2 - 1 \times 10⁻³ mol L⁻¹ CO₃²⁻. The fraction of uranyl bound increases 239 with the ratio of uranyl to organic acid, from 7.5 \pm 2% at 3 × 10⁻⁸ mol U mg⁻¹ HA/FA to 240 28±11% at 1.2 × 10⁻⁹ mol mg⁻¹. The results from these two studies contrast 241 242 somewhat with our experimental observations. These discrepancies may be the 243 result of the presence of a cocktail of competing metals in the case of the natural 244 Gorleben groundwater and, for the Glaus et al (1997) study, the presence of high 245 carbonate concentrations and the comparatively higher uranyl/HA ratios than in our 246 work. There also likely to be variations in binding strengths between humic acid 247 types, which could be a further reason for differences between the data sets.

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251 Uranyl binding to humic acid in the presence of Ca or Cu

252 Competitive reactions with other cationic metals may influence the concentration of 253 humic bound uranyl. In order to quantify such effects, we performed experiments where either Cu^{2+} (10⁻⁷ mol L⁻¹) or Ca^{2+} (10⁻⁴ mol L⁻¹) were added. These metal ions 254 255 were chosen due to their relative binding strengths and typical concentrations in 256 natural waters. The concentrations of these metals will be controlled by various 257 mineral solubility controls within a cement matrix. However, here we are studying only the competition effects in the simple aqueous system. Cu²⁺ typically binds very 258 259 strongly to organic matter and is present in trace concentrations in groundwater (mean European groundwater value $\sim 5 \times 10^{-8}$ mol L⁻¹; Shand and Edmunds, 2008). 260 Ca²⁺ typically has weaker binding to organic matter, but is present at greater 261 262 concentrations (mean European groundwater (~1.6 mmol L^{-1}). Competition from these metal ions in the europium-humic acid system at pH 5.5 has been assessed 263 previously (Marang et al, 2008; 2009). Panel B in Figure 1 shows that, in the 264 265 presence of either metal, uranyl is not significantly displaced from humic acid. For Cu^{2+} , only a small reduction in bound uranyl is observed at pH 12 and 10. This may 266 267 be attributed to an excess of humic binding sites available for both metals. However, this is unlikely to be the case for Ca^{2+} , where the results suggest that uranyl is able to 268 269 compete for the available binding sites. Further work is required to test the scale of 270 these effects over different relative metal concentrations and should also be 271 performed on a range of HA types.

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273 Comparison of measured and modelled data

In order to test the measurements against a comprehensive speciation model we modelled the exact experimental conditions using WHAM/Model VII (Tipping et al., 2011). Figure 1 shows the model predictions in terms of the fraction of uranyl present as humic complexes vs. pH. Figure 2 shows the data expressed in terms of the log concentration of the humic bound uranyl. Error bars for the modelled values are

279 determined by variation of the humic binding constants by a fixed value of log ± 0.3 280 (Lofts and Tipping, 2011). In the uranyl only system, the model gives reasonable 281 predictions up to pH ~10.5. Greater deviation from measured values is then observed 282 as the pH increases. The data, as presented in Figure 2, shows clearly that even in 283 the case where the binding strength is varied to be higher by log 0.3 units, the model 284 still significantly underestimates binding at pH 13. Predictions of binding to organics 285 will also be affected by the accuracy of the data for the speciation of the inorganic 286 complexes.

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We stress the fact that the model is parameterised using proton binding data only up 288 to the same pH value (i.e., 10.5), and that for the majority of metal ions in the 289 290 database, binding data are calculated from experimental data at much lower pH 291 values (for example four of five uranyl humic acid binding datasets contain data at pH 292 <5, with the other dataset at pH ~5-10 and all Cu humic binding datasets fall in the 293 pH range 2.0 to 8.3). Moreover, as detailed in the theory section, we use the generic 294 Model VII value for binding of the uranyl ion. Should SRHA bind stronger than the 295 average value, this will result in experimental binding values higher than predictions. 296 Preliminary results from the authors for neptunyl binding show SRHA binds stronger 297 than purified Aldrich humic acid under the same aqueous conditions (Stockdale and 298 Bryan, unpublished results). The higher complexation capacity of the SRHA could 299 also partly explain why no competition effects are observed in the presence of Cu²⁺. 300 The high pH range is of interest in geodisposal environments, but the model has the 301 majority of its applications in natural waters where such pH regimes rarely exist. The 302 model operates on the basis that only the free ion and first hydrolysis product can 303 bind to organic matter. Increasing competition between organic binding and 304 hydrolysis will operate as the pH increases, and higher hydrolysis products that may 305 bind are not captured by the model. Figure 1C shows the inorganic speciation in the 306 modelled system of Figure 1A. This illustrates the increased hydrolysis over the

307 experimental range and the dominance of negatively charged species throughout. 308 Previous work has determined that the apparent charge of the uranyl ion can be 309 considered to be greater than 2+ (2.1 to 4; Perez-Bustamante, 1971; Rao and 310 Choppin, 1984). Thus, uranyl species with an apparent positive charge may exist at a 311 higher pH than that predicted from formation of nominally zero or negatively charged hydrolysis species and it is possible that these as well as UO_2^{2+} and UO_2OH^+ are 312 313 able to bind to humic substances. Experimental data of the type presented here will 314 enable further considerations of high pH systems when fitting and testing humic 315 binding models for high pH.

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317 Effect of carbonate on the uranyl-humic system

An important consideration in the speciation of UO_2^{2+} is the formation of carbonate 318 complexes. Our experiments were conducted in such a way as to minimise CO₂ 319 dissolution, but were not performed in CO₂ free conditions. It is therefore necessary 320 to evaluate the potential of CO_3^{2-} species to affect the humic binding results. Figure 3 321 322 shows modelling results for organically complexed uranyl in the presence of varying concentrations of CO_3^{2-} . Predicted values at 0.1 mmol L⁻¹ are within log <0.005 of the 323 values where CO_3^{2-} is absent. Increasing concentrations from 1 mmol L⁻¹ to 3 mmol 324 L⁻¹ results in a marked reduction in humic bound uranyl at all the pH values 325 326 considered. This is consistent with the experimental procedure of Glaus et al. (1997). where concentrations in this range were used in competition equilibrium dialysis 327 328 experiments for a fulvic acid uranyl system and at similar DOM : uranyl ratios. The predictions suggest that lower CO_3^{2-} concentrations (from 0.1 to 1 mmol L⁻¹) only 329 330 significantly decrease the humic bound uranyl concentration at pH 10. The fact that 331 the model underestimates uranyl binding to humic acid means that these results will 332 be conservative as it will underestimate competition between the carbonate ligand 333 and humic binding sites. Experiments performed at pH 10 with addition of 0.1 mmol L⁻¹ CO_3^{2-} (panel B of Figure 1) demonstrate that no decrease of humic bound uranyl is observed. We therefore conclude that any CO_3^{2-} present in the experiment is insufficient to suppress humic binding. We recognise that the binding to HA of mixed uranyl-carbonate complexes could also partly explain the apparent lack of competition in this system. Such mixed complexes for europium speciation have been identified by Dierckx et al (1994).

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341 Conclusions

The experimental results presented here provide evidence that within simple high pH environments dissolved organic matter can bind uranyl ions, including in the presence of competing ions. The results therefore, suggest that DOM has the potential to be important in the transport of uranium from a cementitious (alkali) geological disposal facility, despite formation of highly hydrolysed species.

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Using the default Model VII uranyl binding values we can not accurately predict uranyl humic interactions for the HA used in this work at pH values above ~10.5.. The reason for the discrepancies may be due to a possible underestimation of the binding of more hydrolysed species, or be related to higher binding strengths for the specific HA used in this work. Further experimental work of the type presented here, and on a range of HA types, will enable further considerations of high pH environments when fitting and testing humic binding models for suchpH systems.

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410 Figures

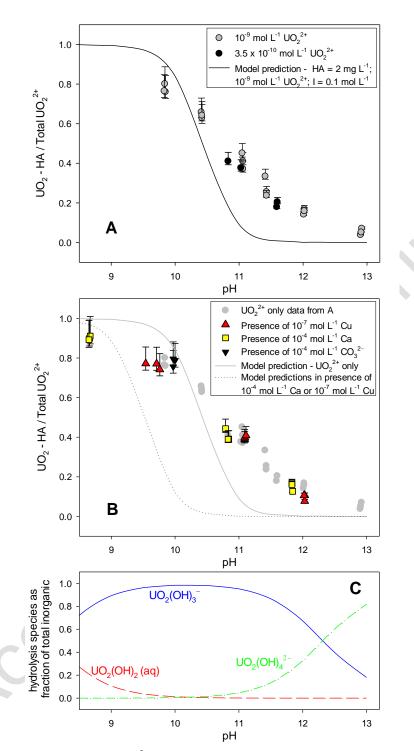




Figure 1. Complexation of UO_2^{2+} with Suwannee River humic acid (SRHA). Panel A 412 shows the fraction of total uranyl bound to SRHA in a solution containing 2 mg L⁻¹ 413 SRHA (added electrolyte, 0.1 mol L⁻¹ NaCl), the solid line represents the predicted 414 values using WHAM/Model VII. Panel B show the fraction of uranyl bound to SRHA 415 in the presence of competing metal ions or carbonate. Only one model prediction line 416 is shown for both the Cu²⁺ and Ca²⁺ systems due to very similar effects at the metal 417 418 ion concentrations considered. Panel C shows the modelled changes to the 419 hydrolysis in the inorganic system over the pH range considered in the experiments 420 and modelling.

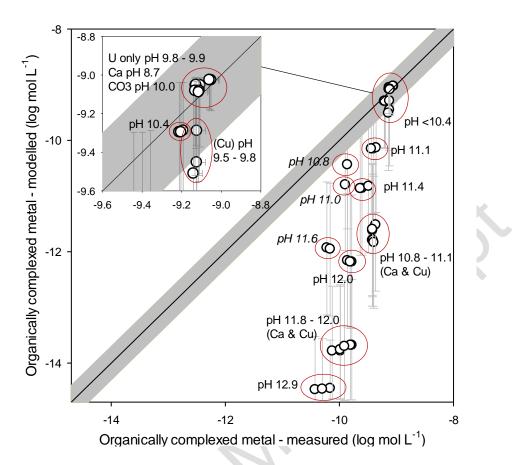
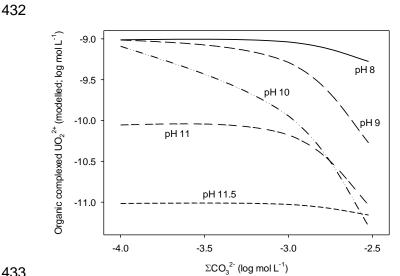




Figure 2. Comparison of WHAM/Model VII calculated concentrations of organically complexed metal and those derived from experimental data. The solid line represents the 1:1 line and the grey area shows those points that fall within 0.3 of a log unit from this line. Experimental conditions are defined in Figure 1. Italicised data indicate those points in Figure 1A where a UO_2^{2+} concentration of 3.5 × 10^{-10} mol L⁻¹ was used.

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435 Figure 3. Modelled humic complexation of uranyl in the presence of varying 436 concentrations of carbonate. Concentrations are the same as the grey circles in Figure 1A. See text for discussion. 437