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COMMENTARY

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The role of Rho kinase (Rock) in re-epithelialization of adult zebrafish skin wounds

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ABSTRACT

Complete re-epithelialization of full-thickness skin wounds in adult mammals takes days to complete and relies on numerous signaling cues and multiple overlapping cellular processes that take place both within the epidermis itself and in other participating tissues. We have previously shown that re-epithelialization of full-thickness skin wounds of adult zebrafish, however, is extremely rapid and largely independent of the other processes of wound healing allowing for the dissection of specific processes that occur in, or have a direct effect on, re-epithelializing keratinocytes. Recently, we have shown that, in addition to lamellipodial crawling at the leading edge, re-epithelialization of zebrafish partial- and full-thickness wounds requires long-range epithelial rearrangements including radial intercalations, flattening and directed elongation and that each of these processes involves Rho kinase (Rock) signaling. Our studies demonstrate how these coordinated signaling events allow for the rapid collective cell migration observed in adult zebrafish wound healing. Here we discuss the particular contribution of Rock to each of these processes.

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cellular rearrangements; collective cell migration; radial intercalation; Reepithelialization; Rho kinase; rock; skin; wound healing; zebrafish

The process of collective cell migration plays vital roles during several in vivo events including wound healing and embryonic morphogenetic movements during development.[1](#page-6-0) These cohesive forms of migration involve the highly coordinated movement of a sheet of epithelial cells and require the individual cells to remain closely connected to maintain epithelial barrier functions and to allow synchronised migration. $¹$ Collective cell migration</sup> during closure of adult mammalian skin wounds, for example, utilizes similar cellular processes to those required for single cell migration (such as in patrolling immune cells). These include cytoskeletal rearrangements within the leading epidermal cells that allow the formation of anteriorly positioned lamellipodia and filopodia anchoring the cell to an underlying substrate, which, when coupled with retractive cytoskeletal rearrangements at the rear of the cell, facilitate the forward movement of that cell or cells.^{[2](#page-6-1)}

The Rho family of GTPases are crucial regulators of actin cytoskeleton dynamics and control lamellipodia and filopodia formation and rear retraction of individual or groups of cells.^{[2](#page-6-1)} Rac and Cdc42 are the predominant family members controlling anterior protrusion assembly whereas RhoA and its downstream mediator Rhoassociated kinase (ROCK) are crucial for rear retrac-tion.^{[2-4](#page-6-1)} It has also been demonstrated that Rho and ROCK play a role in the planar cell polarity (PCP) pathway, which regulates epithelial polarity within a sheet of cells and coordinates collective cell migration in multiple developmental contexts.[5-8](#page-6-2) In zebrafish embryos, for example, overexpression of a dominant negative version of Rock2 results in compromised gastrulation convergence extension movements due to impaired cell elongation and orientation, phenocopying PCP component mutants.⁷ Studies involving multiple mutant analyses have also unravelled an essential role of the PCP pathway for embryonic wound closure in mammals.⁸

Recently, we have studied the roles of Rho/Rock, PCP and other pathways during the epidermal re-epithelialization of partial- and full-thickness wounds in adult zebrafish.⁹ The epidermis of late embryonic and larval zebrafish (24 hours post fertilization (hpf) - 15 d post fertilization (dpf)) consists of an inner basal layer attached to the basement membrane and an outer layer of flattened, specialized periderm cells, 10 10 10 resembling the bilayered epidermis of early embryonic mice $(E9.5-E12)^{11}$ $(E9.5-E12)^{11}$ $(E9.5-E12)^{11}$ and chicks $(E6-E14)^{12}$ $(E6-E14)^{12}$ $(E6-E14)^{12}$ in structure and function ([Fig. 1A\)](#page-3-0). As development continues, the epidermis in all

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Figure 1. (A-C) Representations of cross sections through the epidermis at different time-points and in the different models used for wound healing studies. (A) In embryonic mice and chicks the epidermis is bilayered at the stages used for embryonic wound closure experiments (E12 in mouse, E6 in chick). At these stages the epidermis consists of an inner basal layer and an outer covering of flattened periderm cells. The epidermis of late embryonic and larval zebrafish is very similar in structure and function. (B) In adult zebrafish the epidermis becomes stratified consisting of 2 to 3 layers of basal and intermediate cells and an outer superficial layer. This resembles the epidermis of mid-gestation mice (E13-E15) and chicks (E15-E17) of corresponding developmental stages. (C) The epidermis of adult mice and chicks is also stratified consisting of 4 distinct layers: the innermost basal layer, the suprabasal layer, the granulation layer where terminal differentiation commences and the outermost cornified layer consisting of terminally differentiated cornified envelopes. (D) Representation of a dorsal view of the bilayered epidermis of an embryonic chick or mouse depicting the actomyosin cable form of wound closure. For simplicity only the LE cells of the basal epidermis are shown. The first row of superficial periderm cells are depicted as semi-translucent to allow vizualization of the basal cells beneath.

species thickens to produce additional, intermediary layers between the basal and periderm layers (mouse, E13-E15; chick, E15-E17; zebrafish, 16 dpf-25 dpf).^{[10-12](#page-7-3)} In mouse and chick, terminal differentiation then commences producing a multi-layered, cornified/keratinised epidermis ([Fig. 1C](#page-3-0)).^{11,12} The epidermis of adult zebrafish, by contrast, remains composed of living cells and is similar in structure to the multi-layered, non-keratinised epidermis of late stage embryonic mice and chicks $(Fig. 1B).¹⁰⁻¹²$ $(Fig. 1B).¹⁰⁻¹²$

Contrary to what is observed in adult mammalian wound healing; re-epithelialization of small wounds in embryonic Drosophila, zebrafish, chick and mammals has been shown to occur via a purse-string mechanism rather than active lamellipodial crawling at the leading edge (LE). $^{13-19}$ $^{13-19}$ $^{13-19}$ This purse-string mechanism involves the rapid formation (within minutes of wounding) of a cable consisting of F-actin and activated myosin-II within the LE basal epidermal cells and allows prompt closure of the wound ([Fig. 1D](#page-3-0)).¹³⁻¹⁹ Rho and ROCK play vital roles during formation of this actomyosin cable in the LE cells,

with knockdown of either resulting in a lack of cable formation and consequently a failure of wound healing in vitro and in vivo.^{[20-24](#page-7-7)} Interestingly, inhibition of ROCK in vitro after actomyosin cable assembly has occurred but during cable contraction and wound closure had no affect on re-epithelialization, suggesting that ROCK activity is vital for the formation but not for the function or maintenance of the cable.^{[23](#page-7-8)}

Re-epithelialization of cutaneous wounds in adult mammals is a lengthy and step-wise process by comparison, that initiates many hours after wounding and takes 5-7 d to complete. $25-30$ Timely recovery of the wound epidermis is reliant on the proliferative capacity of the keratinocytes, the presence of a suitable substrate for lamellipodial crawling and contraction of the wound edges by the underlying granulation tissue via differentiated myofibroblasts.[25-30](#page-7-9) By contrast, the rapid re-epithelialization we observed in both full-thickness wounds $(\sim)2$ mm in diameter and down to the level of the subcutaneous adipocytes; \sim 250 μ m/h) and partial-thickness wounds (\sim 0.5 mm in diameter and retaining the basal

dermal layer; \sim 500 μ m/h) in adult zebrafish resembles what is observed in embryonic mammals in it's timing and lack of reliance on other wound healing pro-cesses.^{[9,31](#page-7-2)} Due to this rapid nature of re-epithelialization in adult zebrafish, with no apparent initial lag phase of forward movement, we sought to determine the presence of purse-string mechanisms in our injury models. Using live imaging of adult transgenic zebrafish and labeling of actin and myosin in fixed samples we could observe the formation of a purse-string in very small wounds (2-3 cell diameters) but not following our full- or partial-thickness injuries.^{[9](#page-7-2)} This suggests that in larger wounds, purse-string mechanisms are, if at all, only involved in the final steps of wound closure, when the area yet to be re-covered by keratinocytes has become very small and the number of LE keratinocytes has been reduced to a few cells.

Due to the established role of Rho and ROCK in collective cell migration we hypothesized that they may also be required during adult zebrafish reepithelialization. Indeed, chemical inhibition of Rock via Y27632 or the Rho kinase inhibitor III, Rockout, resulted in significantly slower re-epithelialization of full- and partial-thickness wounds in adult zebrafish.^{[9](#page-7-2)} As we have shown that purse-string mechanisms do not play a predominant role in the closure of larger adult zebrafish injuries, this would suggest that other Rock-dependent processes must be affected and are required for closure of these wounds. Via time-lapse imaging of live double transgenic fish following partial-thickness wounding we could reveal that the basal-most keratinocytes at the LE extended lamellipodia while superficial cells lagged behind, suggesting an active migratory mode of re-epithelialization simi-lar to adult mammals [\(Fig. 2A\)](#page-4-0).^{[9](#page-7-2)} Yet, unlike in adult mammals, this active migration occurs without any apparent lag phase (lamellipodia were observed within minutes of wounding) and at significantly higher speeds than recorded for other models.^{[9](#page-7-2)} Epithelial cells in vitro also close scratch wounds via

Figure 2. (A) Representation of a cross section through the epidermis at the wound edge in an adult zebrafish 5 minutes post-wounding (mpw), one hour post-wounding (hpw), or 1 hpw following Rock inhibition. The epidermis initially consists of 2-3 layers of inner basal and intermediary cells and a layer of superficial cells. Within 5 mpw, the LE basal cells have begun to make lamellipodial protrusions and elongate onto the denuded area. By 1 hpw directed radial intercalations of intermediary cells into the basal layer have occurred producing a single layer of inner cells; the superficial cells have elongated to ensure coverage of the increased basal layer area; the LE cells and several cells behind are still protrusive and have undergone some cell elongation, collectively allowing the whole epidermal sheet to move forward to cover the wound. Following Rock inhibition the processes of radial intercalation and coordinated elongation are severely affected resulting in reduced forward movement even though LE protrusions are maintained. (B) Dorsal view outlines of 4 individual superficial cells taken from images from adult zebrafish at the indicated distance from a full-thickness wound (wound is to the left). In unwounded epidermis, the superficial cells are hexagonal in shape. Following wounding the superficial cells elongate in the direction of the wound to up to 10-times their original diameter. Following Rock inhibition the superficial cells change shape but in an uncoordinated manner resulting in many random shapes.

lamellipodial cell crawling and several rows of following epithelial cells are also observed to produce (cryptic) lamellipodia during collective cell migration.[32-34](#page-7-10) We observed a similar phenomenon in second and third row keratinocytes following the LE in zebrafish wounds in vivo ([Fig. 2A](#page-4-0)). 9 However, inhibition of Rock had no effect on these LE or cryptic protrusions apart from slightly randomizing their spatial orientation, most likely due to the role of Rock in the PCP pathway (see above and below) [\(Fig. 2A](#page-4-0)). 9 Thus, LE cells sometimes produced more than one major protrusion that projected more randomly and not necessarily directly toward the wound bed.

A much more striking effect on LE protrusions was obtained upon inhibition of TGF β signaling or interference with integrin-ECM binding, which compromised lamellipodial shape and stability, as well as re-epithelialization rates. TGF β has been shown to induce a more migratory phenotype in mammalian keratinocytes by inducing the expression of integrins α 5 β 1, α v β 5 and $\alpha v\beta6$ within anterior protrusions.^{[35-37](#page-8-0)} However, it has also been suggested that TGF β has a counteractive negative effect on re-epithelialization by suppressing keratinocyte proliferation, and that this effect is predominant, leading to a net increase in re-epithelialization rates upon genetically compromised TGF β signaling.^{[25,37,38](#page-7-9)} However, direct in vivo evidence for such counteractive TGF β roles is difficult to obtain due to the overlapping time-scale of keratinocyte proliferation and migration during adult mammalian wound healing.^{[25,37,38](#page-7-9)} By contrast, re-epithelialization of adult zebrafish wounds is independent of keratinocyte proliferation, indicated by its insensitivity to inhibition of DNA replication, allowing independent examination of the effect of TGF β inhibition on lamellipodial crawling of basal keratinocytes.⁹ Due to the similar results we obtained from inhibiting $TGF\beta$ signaling or integrin-ECM interactions, we hypothesized that, consistent with the studies in mammalian wound healing systems, $TGF\beta$ signaling promotes LE protrusion stability and functionality by modifying integrin populations at the basal surface of LE keratinocytes, thereby allowing a better and more dynamically regulated adhesion of protrusions with the migration substrate. $9,37$ Consistent with this notion, the effect on protrusion stability and functionality only compromised closure rates of partial-thickness wounds where a dermal substrate is still present. In turn, this suggests that other cellular mechanisms are more important for full-thickness wound re-epithelialization. Furthermore, in light of the absence of purse-string mechanisms during most phases of wound re-epithelialization, and the dispensability of Rock signaling for lamellipodial crawling at the LE (see above), we reasoned that these

other cellular mechanisms might be the main Rock targets underlying the \sim 4-fold reduction in the closure rate of full-thickness wounds upon Y27632 treatment.^{[9](#page-7-2)}

Live imaging of the entire re-epithelialization process following full-thickness wounding revealed cellular events within the intact epidermis at some distance from the wound.⁹ Indeed, close examination of the epidermis up to 2 mm away from the wound revealed that the rapid re-epithelialization observed in adult zebrafish is a result of multiple coordinated epithelial rearrangements in regions behind the LE, including radial intercalations and directed and coordinated cell flattening, polarization and elongation ([Fig. 2\)](#page-4-0). 9 Initially, basal and intermediary keratinocytes behind the LE undergo radial intercalations leading to a long-range progressive reduction of cell layers and a concomitant lateral extension of the remaining basal-most layer [\(Fig. 2A\)](#page-4-0).^{[9](#page-7-2)} This partly provides the additional keratinocytes required to cover the wound and is a process that is independent of cell proliferation and similar to events immediately adjacent to the LE described in other wound healing models. 39 Blocking Rock function with Y27632 led to severely compromised keratinocyte flattening and a failure of radial intercalations within the epidermis around the full-thickness wounds, paired with improper localization of non-mus-cle myosin within the inner keratinocytes ([Fig. 2A\)](#page-4-0). 9 This is likely to limit the capacity for forward movement of the collective cell sheet, underlying the strong reduction in wound closure rates [\(Fig. 2A\)](#page-4-0). These findings are in line with previous reports that Rock drives cell flattening during epidermal stratification and spreading in mammals and radial cell intercalations during gut morphogenesis in amphibia.[40-42](#page-8-2)

The following keratinocytes also undergo directed and coordinated planar polarization and elongation, predominantly in the superficial layer, which apparently does not participate in the underlying intercalation movements, but also to a lower extent in intermediary layers ([Fig. 2\)](#page-4-0).^{[9](#page-7-2)} Indeed, superficial cells elongate up to 10-times their original diameter ([Fig. 2B\)](#page-4-0).^{[9](#page-7-2)} Similar superficial cell elongations have been reported during wound closure in mouse, zebrafish and Drosophila embryos.^{[16,43-45](#page-7-11)} Following inhibition of Rock we also observed a failure in this directed superficial cell elongation, with cells managing to change their shape but in a random and uncoordi-nated manner ([Fig. 2B\)](#page-4-0).⁹ These findings would support a role for Rock in the PCP pathway to allow coordinated elongations and are similar to previously described effects of Rock perturbation during zebrafish and mouse development.^{[7,41](#page-7-0)} However, transgenic blockage of the PCP pathway did not affect wound closure to the same extent as the inhibition of Rock, although both of these treatments had comparable effects on directed

keratinocyte elongation.⁹ This suggests that the PCP pathway coordinates cell elongation but, unlike Rock, is not required for flattening and radial intercalations. In turn, this means that during the latter, and more crucial, epithelial rearrangements, Rock must be part of other pathways that have not been elucidated in this context as yet. Interestingly, a recent report suggests that ROCK functions downstream of the PCP component Celsr1 during mouse epidermal radial intercalations but that this signaling axis may be separate from other PCP driven developmental processes.^{[41](#page-8-3)} Collectively, our data indicate that the coupling of basal/intermediary radial intercalation and cell flattening and elongation, both reliant on Rock function, within the following keratinocytes is vital for, and is likely to be the driving force behind, the rapid re-epithelialization characteristic of adult zebrafish. In contrast, the PCP- and Rock-dependent directionality of cell elongations only has a minor impact on wound re-epithelialization, possibly by indirectly promoting radial cell intercalations that would be compromised if cells elongated in random directions. Of note, cell flattening and radial intercalation are also major driving forces of epiboly, one of the morphogenetic processes during gastrulation,[46](#page-8-4) which also, at least partly, depends on Rho function.²¹ Actually, the term "epiboly" was initially introduced in the context of wound clo-sure,^{[47](#page-8-5)} and was later adopted by developmental biologists in the context of gastrulation without knowing at the time of the similarities in the underlying cellular rearrangements and their genetic control.

Future studies have to address the presence and impact of such Rock-dependent long-range epithelial rearrangements during wound re-epithelialization in mammals. In addition, while our data suggest that Rock regulates these epithelial rearrangements via the actomyosin system, the regulators upstream of Rho/Rock in this context remain largely unclear. Most Rho GTPases cycle between a GTP-bound active conformation and a GDPbound inactive conformation, mediated by guanine nucleotide exchange factors (GEFs) promoting the active and GTPase-activating proteins (GAPs) promoting their inactive state.^{[48,49](#page-8-6)} In addition, Rho proteins and GEFs are regulated via post-translational modifications and their subcellular distribution, triggered by various cellsurface receptors including integrins, cadherins, cytokine receptors, receptor tyrosine kinases (RTKs) and G pro-tein-coupled receptors.^{[48,49](#page-8-6)} Our analyses have largely ruled out integrins and the RTKs of the FGFR family as such stimulators.^{[9](#page-7-2)} Once identified, the relevant cell surface receptors might serve as targets for pharmacological intervention with corresponding ligand agonists, allowing faster wound closure by accelerating the recruitment of re-epithelializing keratinocytes from skin regions

around the wound. As in adult zebrafish, such improvements would occur at the expense of the thickness of the adjacent epidermis, however, this should be a minor problem, as keratinocyte proliferation after the wound has closed could re-establish the epidermis to its normal thickness, as is the case in adult zebrafish.^{[9](#page-7-2)}

Abbreviations

Disclosure of potential conflicts of interest

No potential conflicts of interest were disclosed.

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References

- [1] Friedl P, Gilmour D. Collective cell migration in morphogenesis, regeneration and cancer. Nat Rev Mol Cell Biol 2009; 10(7):445-57; PMID[:19546857; http://dx.doi.](http://dx.doi.org/19546857) [org/10.1038/nrm2720](http://dx.doi.org/10.1038/nrm2720)
- [2] Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, Parsons JT, Horwitz AR. Cell migration: integrating signals from front to back. Science 2003; 302 (5651):1704-9; PMID:[14657486; http://dx.doi.org/](http://dx.doi.org/14657486) [10.1126/science.1092053](http://dx.doi.org/10.1126/science.1092053)
- [3] Xu J, Wang F, Van Keymeulen A, Herzmark P, Straight A, Kelly K, Takuwa Y, Sugimoto N, Mitchison T, Bourne HR. Divergent signals and cytoskeletal assemblies regulate self-organizing polarity in neutrophils. Cell 2003; 114(2):201-14; PMID:[12887922; http://dx.doi.org/](http://dx.doi.org/12887922) [10.1016/S0092-8674\(03\)00555-5](http://dx.doi.org/10.1016/S0092-8674(03)00555-5)
- [4] Worthylake RA, Burridge K. RhoA and ROCK promote migration by limiting membrane protrusions. J Biol Chem 2003; 278(15):13578-84; PMID:[12574166; http://](http://dx.doi.org/12574166) dx.doi.org/10.1074/jbc.M211584200
- [5] Van Aelst L, Symons M. Role of Rho family GTPases in epithelial morphogenesis. Genes Dev 2002; 16:1032-54; PMID:[12000787; http://dx.doi.org/10.1101/gad.978802](http://dx.doi.org/10.1101/gad.978802)
- [6] Verdier V, Guang Chao Chen, Settleman J. Rho-kinase regulates tissue morphogenesis via non-muscle myosin and LIM-kinase during Drosophila development. BMC

Dev Biol 2006; 6:38; PMID:[16882341; http://dx.doi.org/](http://dx.doi.org/16882341) [10.1186/1471-213X-6-38](http://dx.doi.org/10.1186/1471-213X-6-38)

- [7] Marlow F, Topczewski J, Sepich D, Solnica-Krezel L. Zebrafish Rho kinase 2 acts downstream of Wnt11 to mediate cell polarity and effective convergence and extension movements. Curr Biol 2002; 12:876-84; PMID[:12062050; http://dx.doi.org/10.1016/S0960-9822](http://dx.doi.org/10.1016/S0960-9822(02)00864-3) [\(02\)00864-3](http://dx.doi.org/10.1016/S0960-9822(02)00864-3)
- [8] Caddy J, Wilanowski T, Darido C, Dworkin S, Ting SB, Zhao Q, Rank G, Auden A, Srivastava S, Papenfuss TA, et al. Epidermal wound repair is regulated by the planar cell polarity signaling pathway. Dev Cell 2011; 19:138-47; http://dx.doi.org/[10.1016/j.devcel.2010.06.008](http://dx.doi.org/10.1016/j.devcel.2010.06.008)
- [9] Richardson R, Metzger M, Knyphausen P, Ramezani T, Slanchev K, Kraus C, Schmelzer E, Hammerschmidt M. Re-epithelialization of cutaneous wounds in adult zebrafish combines mechanisms of wound closure in embryonic and adult mammals. Development 2016; 143 (12):2077-88; PMID[:27122176; http://dx.doi.org/](http://dx.doi.org/27122176) [10.1242/dev.130492](http://dx.doi.org/10.1242/dev.130492)
- [10] Le Guellec D, Morvan-Dubois G, Sire JY. Skin development in bony fish with particular emphasis on collagen deposition in the dermis of the zebrafish (Danio rerio). Int J Dev Biol 2004; 48:217-31; PMID[:15272388; http://](http://dx.doi.org/15272388) dx.doi.org/10.1387/ijdb.15272388
- [11] Richardson RJ, Hammond NL, Coulombe PA, Saloranta C, Nousiainen HO, Salonen R, Berry A, Hanley N, Headon D, Karikoski R, et al. Periderm prevents pathological epithelial adhesions during embryogenesis. J Clin Invest 2014; 124(9):3891-900; PMID:[25133425; http://dx.](http://dx.doi.org/25133425) [doi.org/10.1172/JCI71946](http://dx.doi.org/10.1172/JCI71946)
- [12] van Echten-Deckert G, Saathoff M, Kirfel G, Herzog V. Specific distribution of barrier-relevant ceramides in the emerging epidermis and the periderm/subperiderm during chicken embryogenesis. Eur J Cell Biol 2007; 86(11- 12):675-82; PMID:[17397966; http://dx.doi.org/10.1016/j.](http://dx.doi.org/10.1016/j.ejcb.2007.01.004) [ejcb.2007.01.004](http://dx.doi.org/10.1016/j.ejcb.2007.01.004)
- [13] Brock J, Midwinter K, Lewis J, Martin P. Healing of incisional wounds in the embryonic chick wing bud: characterization of the actin purse-string and demonstration of a requirement for Rho activation. J Cell Biol 1996; 135:1097-107; PMID:[8922389; http://dx.doi.org/10.1083/](http://dx.doi.org/10.1083/jcb.135.4.1097) [jcb.135.4.1097](http://dx.doi.org/10.1083/jcb.135.4.1097)
- [14] Bement WM, Forscher P, Mooseker MS. A novel cytoskeletal structure involved in purse string wound closure and cell polarity maintenance. J Cell Biol 1993; 121:565- 78; PMID:[8486737; http://dx.doi.org/10.1083/](http://dx.doi.org/10.1083/jcb.121.3.565) [jcb.121.3.565](http://dx.doi.org/10.1083/jcb.121.3.565)
- [15] Martin P, Lewis J. Actin cables and epidermal movement in embryonic wound healing. Nature 1992; 360:179-83; PMID:[1436096; http://dx.doi.org/10.1038/360179a0](http://dx.doi.org/10.1038/360179a0)
- [16] Wood W, Jacinto A, Grose R, Woolner S, Gale J, Wilson C, Martin P. Wound healing recapitulates morphogenesis in Drosophila embryos. Nat Cell Biol 2002; 4(11):907-12; PMID:[12402048; http://dx.doi.org/10.1038/ncb875](http://dx.doi.org/10.1038/ncb875)
- [17] Martin P, Parkhurst SM. Parallels between tissue repair and embryo morphogenesis. Development 2004; 131:3021-34; PMID[:15197160; http://dx.doi.org/10.1242/](http://dx.doi.org/10.1242/dev.01253) [dev.01253](http://dx.doi.org/10.1242/dev.01253)
- [18] Florian P, Schoneberg T, Schulzke JD, Fromm M, Gitter AH. Single-cell epithelial defects close rapidly by an actinomyosin purse string mechanism with functional tight

junctions. J Physiol 2002; 545:485-99; PMID:[12456828;](http://dx.doi.org/12456828) <http://dx.doi.org/10.1113/jphysiol.2002.031161>

- [19] Schwayer C, Sikora M, Slováková J, Kardos R, Heisenberg CP. Actin Rings of Power. Dev Cell 2016; 37 (6):493-506; PMID:[27326928; http://dx.doi.org/](http://dx.doi.org/27326928) [10.1016/j.devcel.2016.05.024](http://dx.doi.org/10.1016/j.devcel.2016.05.024)
- [20] Yarrow JC, Totsukawa G, Charras GT, Mitchison TJ. Screening for cell migration inhibitors via automated microscopy reveals a Rho-kinase inhibitor. Chem Biol 2005; 12:385-95; PMID:[15797222; http://dx.doi.org/](http://dx.doi.org/15797222) [10.1016/j.chembiol.2005.01.015](http://dx.doi.org/10.1016/j.chembiol.2005.01.015)
- [21] Lai SL, Chang CN, Wang PJ, Lee SJ. Rho mediates cytokinesis and epiboly via ROCK in zebrafish. Mol Reprod Dev 2005; 71:186-96; PMID:[15791595; http://dx.doi.org/](http://dx.doi.org/15791595) [10.1002/mrd.20290](http://dx.doi.org/10.1002/mrd.20290)
- [22] Weiser DC, Pyati UJ, Kimelman D. Gravin regulates mesodermal cell behavior changes required for axis elongation during zebrafish gastrulation. Genes Dev 2007; 21:1559-71; PMID[:17575056; http://dx.doi.org/10.1101/](http://dx.doi.org/10.1101/gad.1535007) [gad.1535007](http://dx.doi.org/10.1101/gad.1535007)
- [23] Russo JM, Florian P, Shen L, Graham WV, Tretiakova MS, Gitter AH, Mrsny RJ, Turner JR. Distinct temporal-spatial roles for rho kinase and myosin light chain kinase in epithelial purse-string wound closure. Gastroenterology 2005; 128(4):987-1001; PMID:[15825080; http://dx.doi.org/10.1053/j.](http://dx.doi.org/10.1053/j.gastro.2005.01.004) [gastro.2005.01.004](http://dx.doi.org/10.1053/j.gastro.2005.01.004)
- [24] Abreu-Blanco MT, Verboon JM, Parkhurst SM. Coordination of Rho family GTPase activities to orchestrate cytoskeleton responses during cell wound repair. Curr Biol 2014; 24:144-55; PMID:[24388847; http://dx.doi.org/](http://dx.doi.org/24388847) [10.1016/j.cub.2013.11.048](http://dx.doi.org/10.1016/j.cub.2013.11.048)
- [25] Barrientos S, Stojadinovic O, Golinko MS, Brem H, Tomic-Canic M. Growth factors and cytokines in wound healing. Wound Repair Regen 2008; 16:585- 601; PMID:[19128254; http://dx.doi.org/10.1111/](http://dx.doi.org/10.1111/j.1524-475X.2008.00410.x) [j.1524-475X.2008.00410.x](http://dx.doi.org/10.1111/j.1524-475X.2008.00410.x)
- [26] Martin P. Wound healing–aiming for perfect skin regeneration. Science 1997; 276:75-81; PMID[:9082989; http://](http://dx.doi.org/9082989) dx.doi.org/10.1126/science.276.5309.75
- [27] Shaw TJ, Martin P. Wound repair at a glance. J Cell Sci 2009; 122:3209-13; PMID:[19726630; http://dx.doi.org/](http://dx.doi.org/19726630) [10.1242/jcs.031187](http://dx.doi.org/10.1242/jcs.031187)
- [28] Singer AJ, Clark RA. Cutaneous wound healing. N Engl J Med 1999; 341:738-46; PMID[:10471461; http://dx.doi.](http://dx.doi.org/10471461) [org/10.1056/NEJM199909023411006](http://dx.doi.org/10.1056/NEJM199909023411006)
- [29] Werner S, Grose R. Regulation of wound healing by growth factors and cytokines. Physiol Rev 2003; 83:835- 70; PMID[:12843410](http://dx.doi.org/12843410)
- [30] Werner S, Smola H, Liao X, Longaker MT, Krieg T, Hofschneider PH, Williams LT. The function of KGF in morphogenesis of epithelium and reepithelialization of wounds. Science 1994; 266:819-22; PMID[:7973639;](http://dx.doi.org/7973639) <http://dx.doi.org/10.1126/science.7973639>
- [31] Richardson R, Slanchev K, Kraus C, Knyphausen P, Eming S, Hammerschmidt M. Adult zebrafish as a model system for cutaneous wound-healing research. J Invest Dermatol 2013; 133(6):1655-65; PMID:[23325040; http://](http://dx.doi.org/23325040) dx.doi.org/10.1038/jid.2013.16
- [32] Fenteany G, Janmey PA, Stossel TP. Signaling pathways and cell mechanics involved in wound closure by epithelial cell sheets. Curr Biol 2000; 10:831-8;

PMID:[10899000; http://dx.doi.org/10.1016/S0960-9822](http://dx.doi.org/10.1016/S0960-9822(00)00579-0) [\(00\)00579-0](http://dx.doi.org/10.1016/S0960-9822(00)00579-0)

- [33] Farooqui R, Fenteany G. Multiple rows of cells behind an epithelial wound edge extend cryptic lamellipodia to collectively drive cell-sheet movement. J Cell Sci 2005; 118:51-63; PMID:[15585576; http://dx.doi.org/10.1242/](http://dx.doi.org/10.1242/jcs.01577) [jcs.01577](http://dx.doi.org/10.1242/jcs.01577)
- [34] Matsubayashi Y, Razzell W, Martin P. 'White wave' analysis of epithelial scratch wound healing reveals how cells mobilise back from the leading edge in a myosin-IIdependent fashion. J Cell Sci 2011; 124:1017-21; PMID:[21402875; http://dx.doi.org/10.1242/jcs.080853](http://dx.doi.org/10.1242/jcs.080853)
- [35] Gailit J, Welch MP, Clark RA. TGF- β 1 stimulates expression of keratinocyte integrins during re-epithelialization of cutaneous wounds. J Invest Dermatol 1994; 103:221-27; PMID:[8040614; http://dx.doi.org/](http://dx.doi.org/8040614) [10.1111/1523-1747.ep12393176](http://dx.doi.org/10.1111/1523-1747.ep12393176)
- [36] Zambruno G, Marchisio PC, Marconi A, Vaschieri C, Melchiori A, Giannetti A, De Luca M. Transforming growth factor- β 1 modulates β 1 and β 5 integrin receptors and induces the de novo expression of the α v β 6 heterodimer in normal human keratinocytes: implications for wound healing. J Cell Biol 1995; 129:853-65; PMID:[7537276; http://dx.doi.org/10.1083/jcb.129.3.853](http://dx.doi.org/10.1083/jcb.129.3.853)
- [37] Margadant C, Sonnenberg A. Integrin-TGF- β crosstalk in fibrosis, cancer and wound healing. EMBO Rep 2010; 11:97-105; PMID:[20075988; http://dx.doi.org/10.1038/](http://dx.doi.org/10.1038/embor.2009.276) [embor.2009.276](http://dx.doi.org/10.1038/embor.2009.276)
- [38] Ashcroft GS, Yang X, Glick AB, Weinstein M, Letterio JL, Mizel DE, Anzano M, Greenwell-Wild T, Wahl SM, Deng C, et al. Mice lacking Smad3 show accelerated wound healing and an impaired local inflammatory response. Nat Cell Biol 1999; 1:260-66; PMID:[10559937;](http://dx.doi.org/10559937) <http://dx.doi.org/10.1038/12971>
- [39] Quilhac A, Sire JY. Spreading, proliferation, and differentiation of the epidermis after wounding a cichlid fish, Hemichromis bimaculatus. Anat Rec 1999; 254:435-51; PMID:[10096676; http://dx.doi.org/10.1002/\(SICI\)1097-](http://dx.doi.org/10.1002/(SICI)1097-0185(19990301)254:3%3c435::AID-AR15%3e3.0.CO;2-D) [0185\(19990301\)254:3%3c435::AID-AR15%3e3.0.CO;2-D](http://dx.doi.org/10.1002/(SICI)1097-0185(19990301)254:3%3c435::AID-AR15%3e3.0.CO;2-D)
- [40] Vaezi A, Bauer C, Vasioukhin V, Fuchs E. Actin cable dynamics and Rho/Rock orchestrate a polarized cytoskeletal architecture in early steps of assembling a stratified

epithelium. Dev Cell 2002; 3:367-81; PMID:[12361600;](http://dx.doi.org/12361600) [http://dx.doi.org/10.1016/S1534-5807\(02\)00259-9](http://dx.doi.org/10.1016/S1534-5807(02)00259-9)

- [41] Panousopoulou E, Hobbs C, Mason I, Green JB, Formstone CJ. Epiboly generates the epidermal basal monolayer and spreads the nascent mammalian skin to enclose the embryonic body. J Cell Sci 2016; 129 (9):1915-27; PMID:[26989131; http://dx.doi.org/](http://dx.doi.org/26989131) [10.1242/jcs.180703](http://dx.doi.org/10.1242/jcs.180703)
- [42] Reed RA, Womble MA, Dush MK, Tull RR, Bloom SK, Morckel AR, Devlin EW, Nascone-Yoder NM. Morphogenesis of the primitive gut tube is generated by Rho/ Rock/Myosin II-mediated endoderm rearrangements. Dev Dyn 2009; 238:3111-25; PMID[:19924810; http://dx.](http://dx.doi.org/19924810) [doi.org/10.1002/dvdy.22157](http://dx.doi.org/10.1002/dvdy.22157)
- [43] Gault WJ, Enyedi B, Niethammer P. Osmotic surveillance mediates rapid wound closure through nucleotide release. J Cell Biol 2014; 207:767-82; PMID[:25533845; http://dx.](http://dx.doi.org/25533845) [doi.org/10.1083/jcb.201408049](http://dx.doi.org/10.1083/jcb.201408049)
- [44] McCluskey J, Martin P. Analysis of the tissue movements of embryonic wound healing-DiI studies in the limb bud stage mouse embryo. Dev Biol 1995; 170:102-14; PMID[:7601301; http://dx.doi.org/10.1006/](http://dx.doi.org/10.1006/dbio.1995.1199) [dbio.1995.1199](http://dx.doi.org/10.1006/dbio.1995.1199)
- [45] Razzell W, Wood W, Martin P. Recapitulation of morphogenetic cell shape changes enables wound re-epithelialisation. Development 2014; 141:1814-20; PMID:[24718989; http://dx.doi.org/10.1242/dev.107045](http://dx.doi.org/10.1242/dev.107045)
- [46] Warga RM, Kimmel CB. Cell movements during epiboly and gastrulation in zebrafish. Development 1990; 108:569-80; PMID:[2387236](http://dx.doi.org/2387236)
- [47] Stenn KS. Epibolin: a protein of human plasma that supports epithelial cell movement. Proc Natl Acad Sci U S A 1981; 78:6907; PMID[:6171824; http://dx.doi.org/10.1073/](http://dx.doi.org/10.1073/pnas.78.11.6907) [pnas.78.11.6907](http://dx.doi.org/10.1073/pnas.78.11.6907)
- [48] Rossman KL, Der CJ, Sondek J. GEF means go: turning on RhoGTPases with guanine nucleotide-exchange factors. J Nat Rev Mol Cell Biol 2005; 6:167-80; http://dx. doi.org/[10.1038/nrm1587](http://dx.doi.org/10.1038/nrm1587)
- [49] Hodge RG, Ridley AJ. Regulating Rho GTPases and their regulators. Nat Rev Mol Cell Biol 2016; 17(8):496-510: advance online publication; PMID[:27301673; http://dx.](http://dx.doi.org/27301673) [doi.org/10.1038/nrm.2016.67](http://dx.doi.org/10.1038/nrm.2016.67)