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1           **Validation of the accuracy of needle placement as used**  
2           **in diagnostic local analgesia of the maxillary nerve for**  
3           **investigation of trigeminally mediated headshaking in horses**

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10          **Acknowledgements**

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12          Langford Veterinary Services for funding the first author's residency training.

13           **Validation of the accuracy of needle placement as used**  
14           **in diagnostic local analgesia of the maxillary nerve for**  
15           **investigation of trigeminally mediated headshaking in horses**

16           **Keywords:** trigeminal neuropathy, posterior ethmoidal nerve, infraorbital nerve, horse

17           **Summary**

18           **Objectives:** Diagnostic local anaesthesia of the maxillary nerve is a valuable aid in the diagnosis of  
19           trigeminally mediated headshaking in horses. Our objective is to validate the accuracy of needle  
20           placement in this procedure and to identify any correlation between accuracy of the technique and  
21           operator experience.

22           **Methods:** Using a small volume of contrast medium, the procedure was performed bilaterally on 30  
23           horse cadaver heads by three groups with different levels of experience with the technique. The  
24           location of deposition was then identified using computed tomography (CT).

25           **Results:** Contrast medium was deposited around the target site in 53.3% (32/60) of injections. An  
26           experienced operator succeeded in deposition around the target area significantly ( $p < 0.05$ ) more  
27           often (80%, 16/20) than did the less and non-experienced performers (40%, 16/40).

28           **Conclusions:** A negative response to diagnostic local anaesthesia of the maxillary nerve does not  
29           disprove facial dysaesthesia as the cause of headshaking in that horse as a false negative response  
30           could arise due to failure to deposit local anaesthetic around the target area. Increased experience  
31           in performing the procedure decreases the probability of false negative results.

## 32 **Introduction**

33 Headshaking in horses is commonly a clinical sign of a facial dysaesthesia (abnormal sensation),  
34 suspected to be due to a neuropathy of the maxillary branch of the trigeminal nerve (Newton and  
35 others 2000; Roberts 2011; Roberts and others 2013; Pickles and others 2014). A diagnosis of facial  
36 dysaesthesia can be made by observing a marked decrease in headshaking in response to local  
37 anaesthesia of the innervating sensory nerves (Roberts and others 2013; Pickles and others 2014).

38 Where no cause of this facial dysaesthesia can be identified on endoscopy, imaging of the head,  
39 ophthalmologic and dental examination the dysaesthesia is likely to be neuropathic (Newton and  
40 others 2000; Roberts and others 2013).

41 The maxillary nerve exits the brain via the round foramen, runs cranially and enters the maxillary  
42 foramen into the infraorbital canal to become the infraorbital nerve. The infraorbital nerve  
43 innervates the upper cheek teeth and the skin of the muzzle. The caudal nasal nerve (CNN) branches  
44 off the maxillary nerve just proximal to the maxillary foramen and enters the caudal nasal foramen  
45 before running towards the dorsal meatus of the nasal cavity to innervate the nasal mucosa (Dyce  
46 and others 2002). In previous articles the CNN has been called the 'posterior ethmoidal nerve'  
47 (Newton and others 2000; Roberts and others 2013), but due to possible confusion with the  
48 ethmoidal nerve which branches of the ophthalmic nerve rather than the maxillary nerve we will  
49 refrain from using this term.

50 Local anaesthesia of the infraorbital nerve as it leaves the infraorbital canal resulted in a decrease in  
51 headshaking in 3/19 (16%) horses only (Mair 1999). However, local anaesthesia applied around the  
52 maxillary nerve at the location of the maxillary foramen resulted in a marked decrease in  
53 headshaking in 11 of 17 (65%) and 23 of 27 (85%) of presumed idiopathic headshakers (Newton and  
54 others 2000; Roberts and others 2013). It is suspected there was a false negative result in at least

55 some of the horses, as 3/4 horses which did not respond to diagnostic local analgesia responded  
56 well to caudal compression surgery which itself carries a 49% success rate (Roberts and others  
57 2013).

58 The intended site of deposition of the local analgesic is around the maxillary and CNN, (Newton and  
59 others 2000; Roberts and others 2013) but accuracy of needle deposition at this location using the  
60 approach described by Newton et al (2000) has never been confirmed. Deposition in different  
61 locations could explain some of the false negative responses.

62 The objective of this study was to verify the exact location of injection when performing local  
63 anaesthesia of the maxillary and CNN and also to determine whether increased operator experience  
64 would result in a more accurate placement of the local analgesic.

## 65 **Materials and Methods**

66 Thirty horse cadaver heads with no known clinical abnormalities were obtained from an abattoir. No  
67 selection for breed, age or sex had been made. Decapitation had been performed at the atlanto-  
68 occipital joint. All procedures were performed within 3 hours of decapitation.

69 The heads were randomly assigned to one of three groups, with ten heads to be injected by an  
70 experienced operator (VR) who has used local anaesthesia of the maxillary and CNN routinely on  
71 clinical cases (eight years' experience with headshakers consisting 2% of caseload, total more than  
72 100 cases on which this procedure has been performed). Another 10 heads were injected by a less  
73 experienced (learning) operator (SW), who had been trained by the first operator and had  
74 performed the procedure in four clinical cases before this study. The last ten heads were injected by  
75 ten final year veterinary students with no experience in performing the procedure. Prior to  
76 performing the procedure all of the students were given access to the literature available to  
77 veterinary practitioners on the technique. This included the guidelines as described by Newton *et al.*

78 (2000), a video on placement of the injection which is included in a commercially available iPhone  
79 app (Equine Techniques)<sup>c)</sup>, anatomy books and an equine skull, but no further explanation was  
80 offered and no communication with other performers was allowed. All twelve performers were  
81 right hand dominant. The heads were positioned horizontally (ventral part of mandible resting on  
82 the table) on the CT machine <sup>a)</sup> scanning table. Injection around the maxillary and CNN was then  
83 performed bilaterally using the contrast medium amidotrizoate (Urografin 150®, Bayer)<sup>b)</sup> and all  
84 performers used the approach as first described by Newton *et al.* (2000) (technique shown in figure  
85 1); A 19 gauge 90 mm spinal needle was inserted on the ventral border of the zygomatic process of  
86 the temporal bone at the narrowest point of the zygomatic arch and directed rostromedially in the  
87 direction of the contralateral upper sixth cheek tooth. The needle was inserted to its full length or  
88 until contact with bone was made. Aspiration was attempted before contrast injection and the stylet  
89 was replaced before needle withdrawal in an attempt to keep the contrast localised. To ensure good  
90 accuracy of measurements, only a 0.1ml volume of a mixture of contrast medium was injected  
91 (Bardell and others 2010).

92 Immediately following bilateral injection, CT images were obtained using a 4<sup>th</sup> generation helical CT  
93 scanner <sup>a)</sup>. All heads were scanned from cranial to caudal using a 1.5mm diameter slice thickness  
94 with a pitch of 0.9. Settings were kV of 130, mAs 190 (effective – Siemens CareDose 4D protocol) and  
95 a rotation time of 1 second. Reconstructions were obtained at a thickness of 1.2mm to ensure that  
96 there was sufficient slice detail for multi-planar reconstructions. All images were then reviewed  
97 using commercially available DICOM viewing software <sup>d)</sup>. Images were initially reviewed in the  
98 transverse plane to locate the contrast material. Dorsal images were then reconstructed with the  
99 plane aligned with the hard palate using multiplanar reconstruction. The location of the contrast  
100 injected was then noted and the distance from the target site. In some images, due to the 3  
101 dimensional nature of the skull, thick slice reconstructions were required to bring the location of the

102 contrast injection into the same plane as the caudal nasal foramen. In order to assess placement  
103 zones correctly, the images were then reconfigured as thin slice images and rotated around the axis  
104 of the caudal nasal foramen to align the foramen and contrast injection site. All measurements were  
105 taken at the closest point of the visible contrast media to the caudal nasal foramen.

106 The position of the injected contrast medium was divided into 4 zones depending on its position to  
107 determine the accuracy of the injection (Figure 2). Zone 1 represented contrast around the CNN  
108 nerve within and including at the entrance of the caudal nasal foramen where the CNN nerve  
109 branches off of the maxillary nerve. Zone 2 and 3 lie between zone 1 and a line drawn between the  
110 hamulus of the pterygoid bone and the caudal lateral tip of the caudal maxillary sinus. This region  
111 was then divided into equal portions based on the distance along the maxillary tuber to give zones 2  
112 and 3 (see figure 2). Zone 4 was all regions outside this area which included the orbit and represents  
113 a failure to place the contrast near the maxillary nerve. Designation to the zones were made and  
114 recorded by a single independent observer (CWS) blinded to the identity of the performer of the  
115 injection.

116 Data are presented descriptively and a Kruskal Wallis test was performed to examine the  
117 relationship between accuracy and the three groups of performers, followed by a Mann Whitney U  
118 test to examine significance in difference between each pair of groups. When it was then found  
119 appropriate to combine groups, a Fisher's exact test was used to examine if the experienced  
120 performer did inject around the maxillary nerve (zone 1, 2 and 3) more often than the other  
121 performers did. A Wilcoxon Signed Rank test was used to determine the relationship between  
122 deposition zone and left or right side of the head. Statistical analysis was performed using SPSS  
123 Statistics for Windows version 19<sup>e</sup>). Significance was taken as  $p < 0.05$ .

## 124 **Results**

125 The position of the contrast deposition was recorded for each head and the results for each zone,  
126 side of head and group are displayed in table 1. Contrast is deposited around the maxillary nerve  
127 (zone 1, 2 and 3) in 53.3% (32/60) of injections.

128 The Kruskal Wallis test showed a significant difference in accuracy amongst the 3 groups ( $H(2) =$   
129  $12.148, P = 0.002$ ) with a mean rank of 33.50 for non-experienced, 37.60 for learning and 20.40 for  
130 the experienced performer. The Mann Whitney U test showed no significant difference in accuracy  
131 between the non-experienced and learning performers ( $P = 0.478$ ), but a significant difference  
132 between non-experienced and experienced performers ( $P = 0.018$ ) and also between the learning  
133 and experienced performers ( $P = 0.001$ ). Fig 3 shows the distribution over the zones where contrast  
134 was deposited by the experienced performer versus the other 2 groups combined.

135 When the inexperienced group was combined with the less experienced performer into one group,  
136 and zones 1, 2 and 3 were combine into a result 'hit', a Fisher's exact test could be used to compare  
137 the two groups. This confirmed that the experienced performer placed the injection around the  
138 maxillary nerve (zone 1, 2 and 3 combined) significantly ( $P = 0.006$ ) more frequently (80% (16/20  
139 injections)) than the other performers (40% (16/40 injections)).

140 There was significant difference in accuracy between injections placed on the left or right side of the  
141 head, as determined by the Wilcoxon Signed Rank test ( $P = 0.046$ ), with greater accuracy on the right  
142 as the median of the zones hit on the right hand side was lower than on the left (zone 3 and 4  
143 respectively). The injection was placed around the maxillary nerve in 43.3% of injections on the left  
144 side, and in 63.3% of injections on the right side of the head. (Table 1)



## 145 **Discussion**

146 This study shows that contrast is deposited around the maxillary nerve (zone 1, 2 and 3) in 53.3%  
147 (32/60) of injections, and we found that accuracy is significantly greater when the procedure is  
148 performed by the very experienced performer. There was no significant difference in accuracy  
149 between those with little or no experience in placing the injection in this study.

150 The use of one operator per category is not ideal and represents a limitation of the paper. However,  
151 given the limited number of experienced operators available it was not possible to include more  
152 operators in the experienced or learning group.

153 Injection did not always occur around the maxillary nerve as intended. Thus, a negative response to  
154 the local anaesthesia of the maxillary nerve in horses could result from failure to deposit local  
155 anaesthetic around the nerve. In clinical cases, accurate placement of the local anaesthetic can be  
156 confirmed to some extent by assessing sensitivity of the facial skin and nasal passage, which may  
157 help to distinguish a true negative from a false negative response.

158 Injections placed on the right side of the head were significantly more accurate than injections  
159 placed on the left side of the head. All performers were right hand dominant. In clinical cases a false  
160 negative response to bilateral local anaesthesia of the maxillary nerve and CNN would be more likely  
161 for remaining left-sided facial dysaesthesia. Signs of facial dysaesthesia in headshaking horses are  
162 usually bilateral (Aleman and others 2013). Therefore unilateral application of the local anaesthesia  
163 has not been attempted by the authors in clinical cases.

164 When performing local anaesthesia of the maxillary nerve and CNN in horses, a greater volume of  
165 local anaesthetic (5ml) is used than the volume of contrast medium in this study (Newton and others  
166 2000). Previous to this study, an attempt was made by the authors to analyse the location of  
167 deposition of 5ml contrast medium, but description of anatomical structures involved proved

168 difficult, complicating ensuing statistical analysis had this larger volume been used in the study. The  
169 use of a small volume of contrast medium and imaging modalities rather than dye and dissection  
170 techniques enabled us to accurately locate into which zone the injection had been placed. The  
171 larger volume used in clinical cases will result in analgesia of a much larger area as the anaesthetic  
172 will diffuse along tissues, including more branches of the maxillary nerve than could be visualised in  
173 this study. Therefore in clinical cases success rate of anaesthetising the maxillary and CNN will be  
174 greater than reported in this study. The use of zones rather than absolute distances enabled us to  
175 overcome the variation in head size by standardising each head to itself and helped verify the  
176 accuracy of placement, although the distance of diffusion could not be accounted for.

177 However even when using a larger volume, deposition of local anaesthetic in zone 4 is unlikely to  
178 result in desensitization of the maxillary nerve and will not lead to a decrease in headshaking even if  
179 this is due to facial dysaesthesia. This occurred in 24/40 (60%) of injections by the non-experienced  
180 or less experienced performers, and in 4/20 (20%) of the injections performed by the experienced  
181 performer, suggesting experience is essential when performing local anaesthesia of the maxillary  
182 nerve and CNN.

183 Possible complications of inaccurate injection of local anaesthesia around the maxillary nerve that  
184 have been observed by the authors in clinical cases include temporary loss of sight when the  
185 anaesthetic diffuses to or is deposited around the optic nerve, swelling of the retrobulbar fossa  
186 when disruption of an artery or vein results in a haematoma, or sudden distress when the needle  
187 touches the nerve. These side effects do resolve with time (Tremaine 2007).

188

189 We consider it to be good practice to perform diagnostic local anaesthesia around the maxillary  
190 nerve and CNN on presumed idiopathic headshakers (Roberts and others 2013). A marked decrease

191 in headshaking in response to application of diagnostic local anaesthesia around the maxillary nerve  
192 and CNN without other pathology in the innervated area will support the diagnosis of trigeminal  
193 (maxillary) neuropathy to the veterinarian, the owner of the horse, and may also be used as proof  
194 for insurers. However, the possibility of a false negative outcome must be considered and preferably  
195 explained to the client before performing the diagnostic local anaesthesia around the maxillary  
196 nerve and CNN.

## 197 **Manufacturers' details**

- 198 a) Siemens Somatom Emotion, Erlanger, Germany.
- 199 b) Bayer plc, Newbury, Berkshire, UK.
- 200 c) Veterinary advances ltd, Ireland.
- 201 d) Osirix, Pixmeo, Switzerland
- 202 e) IBM Corporation, New York, United States.

## 203 **Acknowledgements**

204 The Langford Trust for Animal Health and Welfare for funding this study.

205 Langford Veterinary Services for funding the first author's residency training.

## 206 **List of figure legends**

207 Fig 1: Location of intended placement location of 19 Gauge 19mm spinal needle demonstrated in a  
208 horse skull and a clinical case.

209 Fig 2: Dorsal view of an equine head at the level of the maxillary foramen, with the location of zones  
210 used in this study. The caudal part of the infraorbital nerve and the caudal nasal nerve are  
211 highlighted in red. Zone 1 represented contrast around the CNN nerve within and including at the  
212 entrance of the caudal nasal foramen where the CNN nerve branches off of the maxillary nerve.  
213 Zone 2 and 3 lie between zone 1 and a line drawn between the hamulus of the pterygoid bone and  
214 the caudal lateral tip of the caudal maxillary sinus. This region was then divided into equal portions  
215 based on the distance along the maxillary tuber to give zones 2 and 3 (see figure 2). Zone 4 was all  
216 regions outside this area which included the orbit and represents a failure to place the contrast near  
217 the maxillary nerve.

218 Fig 2b: Dorsal view of an equine head at the level of the maxillary foramen, with yellow circles  
219 highlighting the contrast medium in zone 1 (left) and zone 2 (right).

220 Fig 3: The distribution over the zones where contrast was deposited by the experienced performer  
221 versus the other 2 groups combined.

222 **Tables**

		Zone 1	Zone 2	Zone 3	Zone 4	Zone 1+2+3	Total
Left	Non-experienced	0 (0%)	2 (20%)	1 (10%)	7 (70%)	3 (30%)	10
	Learning	0 (0%)	1 (10%)	2 (20%)	7 (70%)	3 (30%)	10
	Experienced	0 (0%)	6 (60%)	1 (10%)	3 (30%)	7 (70%)	10
	Non-exp + learning	0 (0%)	3 (15%)	3 (15%)	14 (70%)	6 (30%)	20
	All performers <sup>§</sup>	0 (0%)	9 (30%)	4 (13.3%)	17 (56.7%)	13 (43.3%)	30
Right	Non-experienced	1 (10%)	2 (20%)	3 (30%)	4 (40%)	6 (60%)	10
	Learning	1 (10%)	0 (0%)	3 (30%)	6 (60%)	4 (40%)	10
	Experienced	2 (20%)	5 (50%)	2 (20%)	1 (10%)	9 (90%)	10
	Non-exp + learning	2 (10%)	2 (10%)	6 (30%)	10 (50%)	10 (50%)	20
	All performers <sup>§§</sup>	4 (13.3%)	7 (23.3%)	8 (26.7%)	11 (36.7%)	19 (63.3%)	30
Both sides	Non-experienced <sup>*</sup>	1 (5%)	4 (20%)	4 (20%)	11 (55%)	9 (45%)	20
	Learning <sup>*</sup>	1 (5%)	1 (5%)	5 (25%)	13 (65%)	7 (35%)	20
	Experienced <sup>**</sup>	2 (10%)	11 (55%)	3 (15%)	4 (20%)	16 (80%) <sup>†</sup>	20
	Non-exp + learning	2 (5%)	5 (12.5%)	9 (22.5%)	24 (60%)	16 (40%) <sup>††</sup>	40
	All performers	4 (6.7%)	16 (26.7%)	12 (20%)	28 (46.7%)	32 (53.3%)	60

223 TABLE 1: Number of times contrast was located in each zone at each side of the head for each group  
 224 of performers.

225 Zone 1 represented contrast around the CNN nerve within and including at the entrance of the  
 226 caudal nasal foramen where the CNN nerve branches of from the maxillary nerve. Zone 2 and 3 lay  
 227 between zone 1 and a line drawn between the hamulus of the pterygoid bone and the caudal lateral  
 228 tip of the caudal maxillary sinus. This region was then divided into equal portions based on the  
 229 distance along the maxillary tuber to give zones 2 and 3 (see figure 2). Zone 4 was all regions outside  
 230 this area which included the orbit and represents a failure to place the contrast near the maxillary  
 231 nerve.

232 The \*, § and † symbols indicate which groups were compared with each other, where an equal  
 233 number of symbols represents no significant difference and an different number of the same  
 234 symbols represent a significant difference.

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