



Yan, J., Poon, J. F., Singh, V. P., Gates, P., & Engman, L. (2015).  
Regenerable Thiophenolic Radical-Trapping Antioxidants. *Organic Letters*,  
17(24), 6162-6165. DOI: 10.1021/acs.orglett.5b03169

Peer reviewed version

Link to published version (if available):  
[10.1021/acs.orglett.5b03169](https://doi.org/10.1021/acs.orglett.5b03169)

[Link to publication record in Explore Bristol Research](#)  
PDF-document

This document is the Accepted Manuscript version of a Published Work that appeared in final form in *Organic Letters*, copyright © American Chemical Society after peer review and technical editing by the publisher. To access the final edited and published work, see DOI: 10.1021/acs.orglett.5b03169

## University of Bristol - Explore Bristol Research

### General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:  
<http://www.bristol.ac.uk/pure/about/ebr-terms.html>

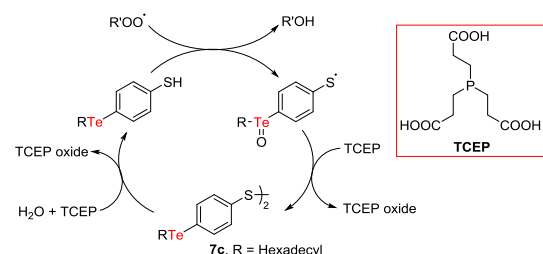
# Regenerable Thiophenolic Radical-Trapping Antioxidants

Jiajie Yan,<sup>†</sup> Jia-fei Poon,<sup>†</sup> Vijay P. Singh,<sup>†</sup> Paul Gates,<sup>‡</sup> and Lars Engman\*<sup>·†</sup>

<sup>†</sup>Uppsala University, Department of Chemistry – BMC, Box 576, SE-751 23 Uppsala, Sweden

<sup>‡</sup>University of Bristol, School of Chemistry, Bristol, BS8 1TS, United Kingdom.

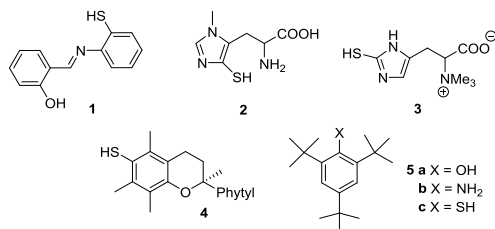
Supporting Information Placeholder



**ABSTRACT:** Diphenyl disulfides carrying alkyltelluro groups in the *o*- *m*- and *p*-positions were prepared using ortho-lithiation and lithium halogen exchange reactions. The novel antioxidants showed only minimal inhibitory effect on the azo-initiated peroxidation of linoleic acid in chlorobenzene until reduced to the corresponding thiophenols by tris-(2-carboxyethyl)phosphine (TCEP). The best in situ-generated thiophenol (from **7c**) under these conditions quenched peroxy radicals more efficiently than  $\alpha$ -tocopherol with an almost threefold increase in inhibition time.

As compared with phenols, their sulfur counterparts – the thiophenols – would seem attractive as radical trapping antioxidants. The S-H bond dissociation enthalpy (BDE) in benzenethiol (79.1 kcal/mol<sup>1a</sup>) is considerably lower than the O-H BDE in hydroxybenzene (89.9 kcal/mol<sup>1b</sup>). According to both theory<sup>2</sup> and experimentation,<sup>3</sup> thiophenol is also more reactive than phenol towards peroxy radicals.

Some arenethiol- and heteroarenethiol derivatives have been shown to function as antioxidants in biological systems. Due to its anti-inflammatory properties, salicylideneamino-2-thiophenol (**1**) has been used as a cure for pain, fever and rheumatism.<sup>4,5</sup> It is also known to stimulate bone formation.<sup>6</sup>



The heteroaromatic imidazol-4-thiol family of natural products known as the ovothiols (represented by **2**) and less polar derivatives thereof<sup>7-9</sup> have been found to act as biological antioxidants. Compounds of this sort could scavenge lipid peroxy radicals almost as efficiently as  $\alpha$ -tocopherol ( $\alpha$ -T) and vitamin C.<sup>10</sup> The natural product ergothioneine (**3**), which is a derivative of imidazol-2-thiol, has also demonstrated numerous antioxidative and cytoprotective capabilities.<sup>11,12</sup> 6-Mercapto- $\alpha$ -tocopherol (**4**) – a sulfur analog of  $\alpha$ -T – has been described in the early literature,<sup>13,14</sup> but little is known about its antioxidative properties.

Whereas phenolic compounds are the most common radical trapping stabilizers for natural and man-made organic materials, aromatic thiols are rarely used. The unpleasant odour of many compounds lacking polar substituents would of course preclude most stabilizer applications, but there are also chemical problems associated with the use of arenethiols as radical trapping antioxidants. Some fifty years ago,<sup>15</sup> Ingold and co-workers compared the capacity of 2,4,6-*tert*-butylphenol (**5a**), -aniline (**5b**) and -thiophenol (**5c**) to inhibit autoxidation of cumene. All compounds **5** could quench peroxy radicals by formal H-atom transfer. However, whereas the resulting phenoxyl and phenylaminy radicals can quench a second peroxy radical, the phenylthiyl radicals disappear by rapid recombination ( $k = 5 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ <sup>16</sup>) to form a diaryl disulfide. The stoichiometric factor *n* – the number of peroxy radicals that could be destroyed per antioxidant molecule – was therefore only 0.95 for **5c**.

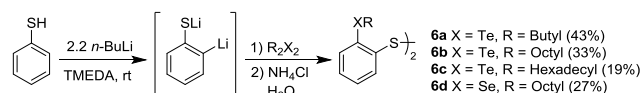
We have tried for some time to find novel antioxidants that could outperform the traditional phenols and aromatic amines when it comes to radical trapping activity and regenerability.<sup>17,18</sup> In this work, we found that alkyltelluro groups have a remarkable effect. For example, introducing an octyltelluro group next to the OH in phenol caused a ca. four orders of magnitude increase in the reactivity towards peroxy radicals. Furthermore, this modification of the structure also improved the regenerability of the antioxidant in the lipid phase by aqueous phase co-antioxidants such as *N*-acetylcysteine (NAC).

We were curious to see how alkyltelluro substitution in thiophenol would affect its radical trapping activity. For comparison, some of the corresponding alkylseleno compounds were also prepared. Obviously, we could expect that these com-

pounds would be readily transformed into inactive “dormant” diaryl disulfide antioxidants, but we were challenged by the perspective to find a suitable co-antioxidant that, “on demand”, could make them come alive again. We herein describe the synthesis of the target antioxidants as well as their radical trapping activity and regenerability in a simple two-phase peroxidation model designed to mimic a biological membrane.

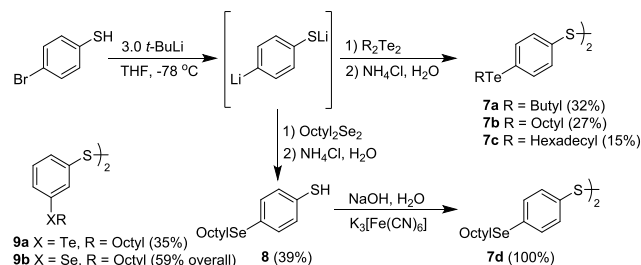
For the preparation of ortho-functionalized compounds, we decided to take advantage of the directing effect of an arenethiolate ion in lithiation.<sup>19</sup> Thiophenol was therefore treated with 2.2 equivalents of *n*-butyllithium in the presence of tetramethylethylenediamine (TMEDA). Addition of dibutyl-, dioctyl-, dihexadecyl ditelluride and dioctyl diselenide, respectively, to the *in situ*-prepared solution of 2, *S*-dilithiated thiophenol afforded diphenyl disulfides **6a-d** in modest yields (19-43%) after acidic work-up and chromatographic purification (Scheme 1). Obviously, the ortho-functionalized thiophenols initially formed are oxidized to the corresponding disulfides during work-up.

### Scheme 1. Synthesis of Compounds 6a-d



For the functionalization of thiophenol in the para- and meta-positions, we relied on lithium halogen exchange.<sup>17c</sup> 4-Bromothiophenol upon treatment with three equivalents of *tert*-butyllithium and then dibutyl-, dioctyl-, and dihexadecyl ditelluride, respectively, returned disulfides **7a-c**, again in low (15-32%) isolated yields (Scheme 2).

### Scheme 2. Synthesis of Compounds 7a-d and 9a-b

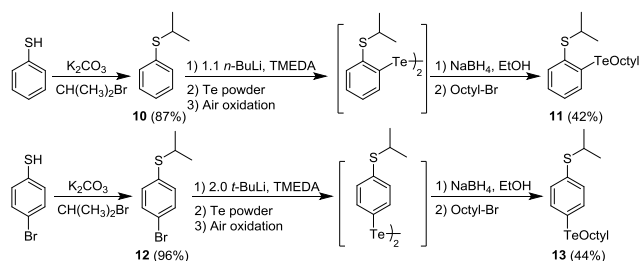


Surprisingly, when we used a similar protocol for the preparation of the corresponding octylseleno-derivative **7d**, 4-(octylseleno)thiophenol **8** was isolated in 39% yield. Oxidation with potassium ferricyanide under basic conditions afforded bis-4-(octylseleno)phenyl disulfide in quantitative yield. Compounds **9**, functionalized with octyltelluro (**9a**, 35% yield) and octylseleno groups (**9b**, 59% overall yield) in the meta-position, were also prepared by lithium halogen exchange using 3-bromothiophenol as a starting material.

For reference purposes, to see the effect of the aromatic sulfhydryl group, we also prepared two *S*-isopropylated octyltelluro-functionalized thiophenols **11** and **13**. *S*-isopropylated thiophenol (**10**) was ortho-lithiated and elemental tellurium allowed to insert into the carbon lithium bond. After air-oxidation, the resulting crude ditelluride was

reduced with sodium borohydride and the arenetellurolate formed alkylated with octyl bromide (Scheme 3).

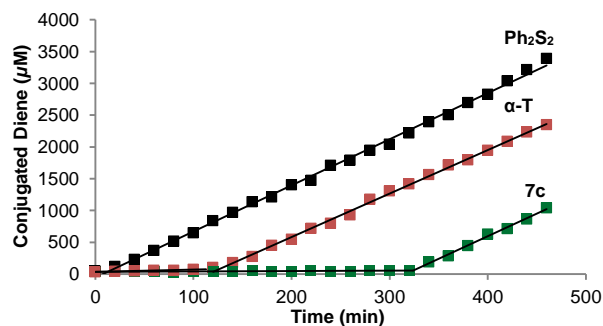
### Scheme 3. Synthesis of Compounds 11 and 13



*S*-isopropylated 4-bromothiophenol (**12**) was subjected to lithium halogen exchange and then converted to **13** using a similar protocol as described for the preparation of compound **11**.

The radical trapping capacity and regenerability of our novel thiophenol-derived antioxidants were studied in a two-phase model.<sup>20</sup> In the lower chlorobenzene layer, containing the antioxidant to be evaluated, peroxidation of linoleic acid was on-going, initiated by azo-bis-dimethylvaleronitrile. The conjugated diene formed as a result of this process was monitored by HPLC with UV detection at 234 nm. The upper aqueous layer contained a water-soluble co-antioxidant that could hopefully regenerate the active antioxidant in the organic phase. By following the increase in conjugated diene with time, information about the radical quenching activity and regenerability of the antioxidant could be obtained. Usually, two stages could be distinguished in the peroxidation traces (Figure 1). During the first inhibited phase, most of the peroxy radicals were quenched by the antioxidant and the concentration of conjugated diene increased only slowly with time ( $R_{inh}$ ). Then, in a second phase, the diene concentration with time increased suddenly to a higher constant value,  $R_{uninh}$ , which was very similar to the one recorded in the absence of any antioxidant in the chlorobenzene layer. The cross-point of the inhibited and uninhibited lines were calculated and reported as the inhibition time,  $T_{inh}$ , of the antioxidant.

Compounds **6**, **7**, **9**, **11** and **13** were first evaluated in the absence of any co-antioxidant in the aqueous phase. Diphenyl disulfide ( $\text{Ph}_2\text{S}_2$ ) and  $\alpha$ -T were also included for reference



**Figure 1.** Peroxidation traces (conjugated diene concentration vs time) recorded with compounds **7c**,  $\text{Ph}_2\text{S}_2$ , and  $\alpha$ -T (40  $\mu\text{M}$ ) as antioxidants in the chlorobenzene layer in the presence of TCEP (0.5 mM) in the aqueous phase.

purposes (Table 1). Except for  $\alpha$ -T ( $R_{inh} = 28 \mu\text{M/h}$  and  $T_{inh} = 109 \text{ min}$ ), none of the compounds tested could inhibit peroxidation. Rather, they acted as poor retarders of peroxidation and no inhibition time could be recorded.

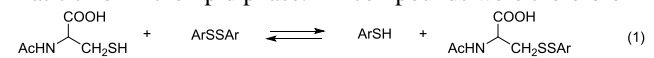
**Table 1. Inhibited Rates of Conjugated Diene Formation ( $R_{inh}$ ) and Inhibition Times ( $T_{inh}$ ) with TCEP (0.5 mM), NAC (1.0 mM), and without Co-antioxidants**

AO <sup>a</sup>	with TCEP		with NAC		without co-antioxidant	
	$R_{inh}^b$ ( $\mu\text{M/h}$ )	$T_{inh}^c$ (min)	$R_{inh}^b$ ( $\mu\text{M/h}$ )	$T_{inh}^c$ (min)	$R_{inh}^b$ ( $\mu\text{M/h}$ )	$T_{inh}^c$ (min)
<b>6a</b>	9.7 ± 1	150 ± 5	43 ± 3	66 ± 3	437	0
<b>6b</b>	8.2 ± 2	148 ± 0	32 ± 3	73 ± 6	333	0
<b>6c</b>	7.3 ± 1	156 ± 5	19 ± 2	84 ± 1	318	0
<b>6d</b>	307	0	300	0	371	0
<b>7a</b>	3.9 ± 0	284 ± 7	19 ± 3	132 ± 3	358	0
<b>7b</b>	3.7 ± 1	304 ± 4	20 ± 1	146 ± 3	321	0
<b>7c</b>	2.3 ± 1	324 ± 5	15 ± 1	170 ± 4	342	0
<b>7d</b>	304	0	343	0	385	0
<b>9a</b>	6.9 ± 1	164 ± 2	20 ± 1	88 ± 7	367	0
<b>9b</b>	338	0	371	0	436	0
<b>11<sup>d</sup></b>	81 ± 2	37 ± 2	72 ± 2	78 ± 4	390	0
<b>13<sup>d</sup></b>	118 ± 5	32 ± 3	23 ± 1	101 ± 3	388	0
Ph <sub>2</sub> S <sub>2</sub>	431	0	372	0	421	0
$\alpha$ -T	21 ± 2	123 ± 7	25 ± 1	97 ± 5	28 ± 2	109 ± 2

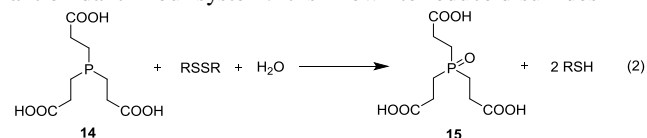
<sup>a</sup>AO = Antioxidant. 40  $\mu\text{M}$  was used in the assay. <sup>b</sup>Rate of peroxidation during the inhibited phase. Uninhibited rate is 453  $\mu\text{M/h}$  with TCEP, 425  $\mu\text{M/h}$  with NAC, and 479  $\mu\text{M/h}$  without antioxidant and co-antioxidant. Errors correspond to  $\pm$  SD for triplicates. <sup>c</sup>Inhibited phase of peroxidation. Reactions were monitored for 460 min. Errors correspond to  $\pm$  SD for triplicates. <sup>d</sup>80  $\mu\text{M}$  of antioxidant was used in the assay.

Previously, we have found that the water-soluble thiol NAC could significantly prolong the  $T_{inh}$  for many chalcogen-containing antioxidants.<sup>17</sup> The thiol probably serves to reduce the chalcogen from the tetravalent to the divalent state. It was our hope that NAC, via thiol exchange with diaryl disulfides (eq 1), would also serve to increase the concentration of aromatic thiol in the lipid phase. All compounds were therefore

tested again in the presence of NAC (1 mM). Considerable improvement was seen. Some of the catalysts (**6c**, **7a-c**, **9a**) could now match  $\alpha$ -T when it comes to inhibited rate of peroxidation and inhibition time. As noted before, NAC did not show any inhibiting effect on peroxidation when tested alone in a control experiment and it did not serve to extend the  $T_{inh}$  for  $\alpha$ -T. In a similar vein, all catalysts were also tested in the presence of 0.5 mM of the water-soluble dithiol dithiothreitol (DTT), acidified with acetic acid to the same pH (3.3;  $R_{inh}$  and  $T_{inh}$  showed some pH-dependence).



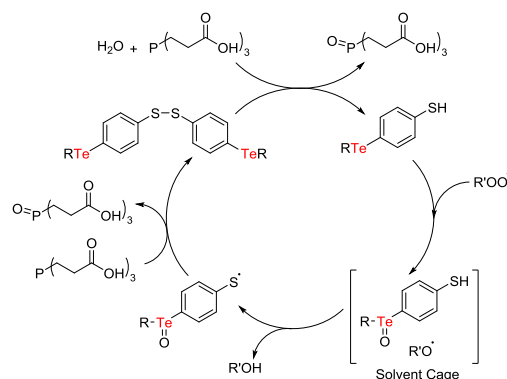
as in 1 mM NAC. However, the  $R_{inh}$ - and  $T_{inh}$  data recorded were almost a blueprint of those obtained with NAC (Table S1 in the Supporting Information). It may be that the thiol exchange of NAC and DTT with the diaryl disulfides is not favourable enough that the compounds could exert their full antioxidant capacity. Therefore, a better water-soluble disulfide reducing agent was sought for. It occurred to us that tris-(2-carboxyethyl)phosphine (TCEP, **14**) could be a useful co-antioxidant in our system. It is known to reduce disulfides



as shown in eq 2.<sup>21</sup> Interestingly, when TCEP hydrochloride (0.5 mM; pH = 3.27) was added to the aqueous phase,  $R_{inh}$  was notably reduced and  $T_{inh}$  prolonged for most of the antioxidants tested. This would indicate efficient reduction of diaryl disulfide and facile regeneration of the antioxidant under the conditions of the assay. Whenever the alkyl part of the alkyltelluro moiety was varied in a systematic way (disulfides **6** and **7**) the performance increased slightly with increasing chain-length/lipophilicity (butyl < octyl < hexadecyl). Overall, the *p*-alkyltelluro functionalized disulfides **7a-c** showed lower  $R_{inh}$  and longer  $T_{inh}$  than their *o*-substituted counterparts **6**. The antioxidant activity of the only *m*-alkyltelluro functionalized derivative **9a** prepared was in between those of the corresponding *o*- and *p*-derivatives. The best antioxidant, *p*-disubstituted diphenyl disulfide **7c**, inhibited peroxidation for 324 min with  $R_{inh}$  as low as 2.3  $\mu\text{M/h}$  (see Figure 1). In contrast, none of the analogous alkylseleno substituted diphenyl disulfides **6d**, **7d**, and **9b** or diphenyl disulfide could act as radical trapping agents in the two-phase model. Apparently, tellurium is a vital constituent in these antioxidants.

We postulate below that alkyltelluro thiophenols are the active antioxidants generated in situ under the conditions of the two-phase model. It was therefore of interest to study the performance of some *S*-alkylated thiophenols. In the presence

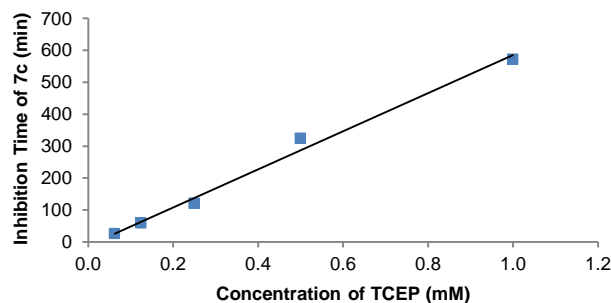
#### Scheme 4. Proposed Catalytic Mechanism for Peroxyl Radical Trapping by Bis-4-(alkyltelluro)phenyl disulfides in the Presence of TCEP.



of NAC, *S*-isopropylated compound **13** (at 80  $\mu\text{M}$ ) showed the same antioxidant characteristics as  $\alpha$ -T (at 40  $\mu\text{M}$ ). The performance of the corresponding *o*-disubstituted compound **11** was not so impressive. Although an inhibited phase of peroxi-

ation could be distinguished for both **11** and **13**, the  $R_{inh}$ - and  $T_{inh}$  values were not near those recorded for the corresponding disulfides **6b** and **7b** (at 40  $\mu$ M). It may be that these compounds quench peroxy radicals by donating electrons from tellurium. In the absence of an aqueous-phase reducing agent they are likely to be oxidized to the corresponding inactive telluroxides by residual amounts of linoleic acid hydroperoxide always present in linoleic acid. Our proposed catalytic antioxidant mechanism is shown in Scheme 4. TCEP-induced reduction of disulfide will produce alkyltelluro thiophenol which quenches peroxy radicals by transfer of oxygen to tellurium.<sup>18</sup> Then, in a solvent cage, the resulting alkoxy radical will abstract a hydrogen atom from the thiophenol. Recombination of thiyl radicals will follow, accompanied by reduction of tetravalent tellurium (shown as a telluroxide) to reform the disulfide antioxidant. In fact, it is known that trivalent phosphorous compounds could act as reducing agents towards telluroxides.<sup>22</sup> No attempt was made to isolate intermediates proposed in Scheme 4.

We were also curious to see the performance of our catalysts in the presence of varying (0.0625 mM, 0.125 mM, 0.25 mM, 0.5 mM, 1.0 mM) amounts of TCEP (Table S2 in the Supporting Information). As shown in Figure 2,  $T_{inh}$  increased linearly with the concentration of the co-antioxidant. On the other hand, the corresponding  $R_{inh}$  values decreased to reach a constant value above 0.5 mM TCEP. Obviously, at the lower concentrations of TCEP, the thiol concentration in the chlorobenzene phase is so low that the peroxy radicals cannot be efficiently quenched.



**Figure 2.** Inhibition Time of Compound **7c** versus Concentration of TCEP

In conclusion, we have shown that (alkyltelluro)thiophenols generated in situ in a lipid phase can quench peroxy radicals more efficiently than  $\alpha$ -T. The tris-(2-carboxyethyl)phosphine used to bring about disulfide to thiol reduction has an additional role: it reduces the telluroxide form of the catalyst to the corresponding telluride and thus allows for antioxidant regeneration and a catalytic mode of action. What makes these antioxidants unique is the possibility to keep them turned off under non-reducing conditions and, whenever needed, make them come alive. It remains to be seen if this capability can be exploited in biological systems.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at.

Experimental section including spectral data for all compounds prepared (PDF).

## AUTHOR INFORMATION

### Corresponding Author

\* lars.engman@kemi.uu.se

### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENT

The Swedish Research Council (621-2011-4006) and Carl Tryggers Stiftelse för Vetenskaplig Forskning (CTS:120) are gratefully acknowledged for financial support.

## REFERENCES

- (1) S-H and O-H BDEs given were estimated as described by Bordwell and are based on a thermodynamic cycle which rely on pK<sub>a</sub>-values for phenol/thiophenol and oxidation potentials of the corresponding conjugate bases. These BDE-values are in excellent agreement with experiment. (a) Bordwell, F. G.; Zhang, X.; Satish, A. V.; Cheng, J.-P. *J. Am. Chem. Soc.* **1994**, *116*, 6605-6610. (b) Bordwell, F. G.; Cheng, J. *J. Am. Chem. Soc.* **1991**, *113*, 1736-1743.
- (2) Denisov, E.; Chatgililoglu, C.; Shestakov, A.; Denisova, T. *Int. J. Chem. Kin.* **2009**, *41*, 284-293.
- (3) Chenier, J. H. B.; Furimsky, E.; Howard, J. A. *Can. J. Chem.* **1974**, *52*, 3682-3688.
- (4) Chung, J.; Lee H. S.; Chung, H. Y.; Yoon, T. R.; Kim H. K. *Biotechnol. Lett.* **2008**, *30*, 1553-1558.
- (5) Park, S.; Sung, B.; Jang, E. J.; Kim, D. H.; Park, C. H.; Choi, Y. J.; Ha, Y. M.; Kim, M. K.; Kim, N. D.; Yu, B. P.; Chung, H. Y. *Arch. Pharm. Res.* **2013**, *36*, 880-889.
- (6) Kim, H. K.; Park, K. S.; Lee, J. S.; Kim, J. H.; Park, D. S.; Shin, J.; Yoon, T. R. *J. Cell. Biochem.* **2012**, *113*, 1833-1841.
- (7) Holler, T. P.; Hopkins, P. B. *J. Am. Chem. Soc.* **1988**, *110*, 4837-4838.
- (8) Zoete, V.; Bailly, F.; Catteau, J.; Bernier, J. *J. Chem. Soc. Perkin Trans. I* **1997**, 2983-2988.
- (9) Zoete, V.; Vezin, H.; Bailly, F.; Vergoten, G.; Catteau, J.; Bernier, J. *Free Rad. Res.* **2000**, *32*, 525-533.
- (10) Marjanovic, B.; Simic, M. G.; Jovanovic, S. V. *Free Rad. Biol. Med.* **1995**, *18*, 679-685.
- (11) Cheah, I. K.; Halliwell, B. *Biochim. Biophys. Acta* **2012**, *1822*, 784-793.
- (12) Servillo, L.; Castaldo, D.; Casale, R.; D'Onofrio, N.; Giovane, A.; Cautela, D.; Balestrieri, M. L.; *Free Rad. Biol. Med.* **2015**, *79*, 228-236.
- (13) Hromata, O.; Kirmig, I. *Monatsh. Chem.* **1954**, *85*, 235-40.
- (14) Csallany, A.; Saari, A.; Draper, H. H. *Int. J. Vitam. Nutr. Res.* **1971**, *41*, 368-375.
- (15) Gardner, D. V.; Howard, J. A.; Ingold, K. U. *Can. J. Chem.* **1964**, *42*, 2847-2851.
- (16) Scott, T. W.; Liu, S. N. *J. Phys. Chem.* **1989**, *93*, 1393-1396.
- (17) (a) Kumar, S.; Johansson, H.; Kanda, T.; Engman, L.; Müller, T.; Jonsson, M.; Pedulli, G. F.; Petrucci, S.; Valgimigli, L. *Org. Lett.* **2008**, *10*, 4895-4898. (b) Kumar, S.; Johansson, H.; Kanda, T.; Engman, L.; Müller, T.; Bergenudd, H.; Jonsson, M.; Pedulli, G. F.; Amorati, R.; Valgimigli, L. *J. Org. Chem.* **2010**, *75*, 716-725. (c) Johansson, H.; Shanks, D.; Engman, L.; Amorati, R.; Pedulli, G. F.; Valgimigli, L. *J. Org. Chem.* **2010**, *75*, 7535-7541. (d) Singh, V. P.; Poon, J.; Engman, L. *J. Org. Chem.* **2013**, *78*, 1478-1487. (e) Poon, J.; Singh, V. P.; Engman, L. *J. Org. Chem.* **2013**, *78*, 6008-6015. (f) Singh, V. P.; Poon, J.; Engman, L. *Org. Lett.* **2013**, *15*, 6274-6277. (g) Poon, J.; Singh, V. P.; Yan, J.; Engman, L. *Chem. Eur. J.* **2015**, *21*, 2447-2457.
- (18) Amorati, R.; Valgimigli, L.; Dinér, P.; Bakhtiari, K.; Saeedi, M.; Engman, L. *Chem. Eur. J.* **2013**, *19*, 7510-7522.

- (19) Smith, K.; Lindsay, C. M.; Pritchard, G. J. *J. Am. Chem. Soc.* **1989**, *111*, 665-669.
- (20) Shanks, D.; Amorati, R.; Fumo, M. G.; Pedulli, G. F.; Valgimigli, L.; Engman, L. *J. Org. Chem.* **2006**, *71*, 1033-1038.
- (21) (a) Levison, M. E.; Josephson, A. S.; Kirschenbaum, D. M.; *Experientia*, **1969**, *25*, 126-127. (b) Burns, J. A.; Butler, J. C.; Moran, J.; Whitesides, G. M. *J. Org. Chem.* **1991**, *56*, 2648-2650.
- (22) Oba, M.; Okada, Y.; Nishiyama, K.; Ando, W. *Org. Lett.* **2009**, *11*, 1879-1881.