Lyons, L. A., Erdman, C., Grahn, R., Hamilton, M., Carter, M., Helps, C. R., ... Gandolfi, B. (2016). Aristaless-like homeobox protein 1 (ALX1) variant associated with craniofacial structure and frontonasal dysplasia in Burmese cats. Developmental Biology, 409(2), 451-458. DOI: 10.1016/j.ydbio.2015.11.015

Lyons, L. A., Erdman, C., Grahn, R., Hamilton, M., Carter, M., Helps, C., ... Gandolfi, B. (2015). Aristaless-like homeobox protein 1 (ALX1) variant associated with craniofacial structure and frontonasal dysplasia in Burmese cats.Developmental Biology, 409(2), 451. 10.1016/j.ydbio.2015.11.015

Aristaless-like homeobox protein 1 (*ALX1)* variant associated with craniofacial structure and frontonasal dysplasia in Burmese cats.

Highlights

Cat breeds are models for mammalian frontonasal development.

A 12 bp in frame deletion in *ALX1,* c.496delCTCTCAGGACTG is 100% concordant with the craniofacial defect in cats.

The *ALX1* variant in cats has a heterozygous advantage in Burmese cat breeding.

The cat model for frontonasal dysplasia could facilitate therapeutics directed to early developmental stages.

Aristaless-like homeobox protein 1 (*ALX1)* variant associated with craniofacial structure and frontonasal dysplasia in Burmese cats.

Leslie A. Lyons,[1,6,9] Carolyn A. Erdman,[2,6] Robert A. Grahn,[3,6] Michael J. Hamilton,[4,6] Michael J. Carter,[5,6] Christopher R. Helps,[7] Hasan Alhaddad,[8] and Barbara Gandolfi,[1,6]

[1]Department of Veterinary Medicine & Surgery, College of Veterinary Medicine, University of Missouri – Columbia, Columbia, Missouri 65211, USA; [2]Department of Psychiatry, University of California - San Francisco, San Francisco, CA 94143, USA; [3]Veterinary Genetics Laboratory, School of Veterinary Medicine, University of California - Davis, Davis, California 96516, USA; [4]Department of Cell Biology and Neuroscience, Institute for Integrative Genome Biology, Center for Disease Vector Research, University of California - Riverside, Riverside, California 92521, USA; [5]MDxHealth Inc 15279 Alton Parkway, Suite # 100, Irvine, California 92618; [6]Department of Population Health and Reproduction, School of Veterinary Medicine, University of California - Davis, Davis, California 95776, USA; [7]Langford Veterinary Services, University of Bristol, Bristol, BS40 5DU, UK; [8]College of Science, Kuwait University, Safat, Kuwait

[9]Corresponding author:

Leslie A. Lyons, PhD

Department of Veterinary Medicine & Surgery, College of Veterinary Medicine
E109 Vet Med Building, 1600 E. Rollins St.
University of Missouri - Columbia
Columbia, MO 65211 USA
Lyonsla@Missouri.edu Phone: 01 573 882 9777 Fax: 01 573 884 2287

Running Title: *ALX1* variant disrupts craniofacial development

Abstract

Frontonasal dysplasia (FND) can have severe presentations that are medically and socially debilitating. Several genes are implicated in FND conditions, including *Aristaless-Like Homeobox 1 (ALX1)*, which is associated with FND3. Breeds of cats are selected and bred for extremes in craniofacial morphologies. In particular, a lineage of Burmese cats with severe brachycephyla is extremely popular and is termed Contemporary Burmese. Genetic studies demonstrated that the brachycephyla of the Contemporary Burmese is a simple co-dominant trait, however, the homozygous cats have a severe craniofacial defect that is incompatible with life. The craniofacial defect of the Burmese was genetically analyzed over a 20 year period, using various genetic analysis techniques. Family-based linkage analysis localized the trait to cat chromosome B4. Genome-wide association studies and other genetic analyses of SNP data refined a critical region. Sequence analysis identified a 12 bp in frame deletion in *ALX1,* c.496delCTCTCAGGACTG, which is 100% concordant with the craniofacial defect and not found in cats not related to the Contemporary Burmese.

[*Keywords: Cartilage homeo protein 1, CART1*, domestic cat, facial development, frontonasal dysplasia, FND, *Felis silvestris catus*]

Frontonasal dysplasia (FND) or median cleft syndrome is a heterogeneous group of disorders that describes an array of abnormalities affecting development of the maxilla-facial structures and the skull. The prevalence of FND is unknown and is considered a rare or “orphan” disease (ORPHA No.: ORPHA250), however affected children can have severe presentations that are life-long medically and socially debilitating. Three genes have been implicated in FND conditions. *Aristaless-Like Homeobox 1 (ALX1)* (OMIM:601527) is associated with FND3, which was defined in three Turkish sibs of consanguineous parents . *ALX1* is also known as *Cartilage homeoprotein-1 (CART1)* , which has been demonstrated to cause neural tube defects in mice , presenting as acrania and meroanencephaly in mice.

Domesticated animals are often selected for craniofacial variants that become breed defining traits. Conditions that would be considered abnormalities or severe craniofacial defects in humans are desired phenotypes in cats and dogs, thus companion animals are excellent models for human facial development due to their popularity. Many dog and cat breeds are bred for brachycephaly, which is assumed to be preferred due to its neotenic effect on the animal’s face. In dogs, the definition of brachycephaly has been quantified by morphological measurements and two genes have been implicated for affecting head type . The health concerns associated with canine brachycephaly have come under strong veterinary and public scrutiny , suggesting severe modifications to breeding programs to alleviate the extent of brachycephaly.

The Burmese is a cat breed with an extreme brachycephalic phenotype (**Fig. 1a**). In the late 1970’s, a male Burmese cat in the USA with a more brachycephalic head type became a highly popular sire and his lineage became known as the “Contemporary”

Burmese (**Fig. 1b**). The head type was found to be heritable, however, offspring from "Contemporary" style mating produced a craniofacial defect in 25% of offspring . The abnormality is characterized by agenesis of all derivatives of the medial nasal prominence; lateral duplication of most derivatives of the maxillary process; including the canine teeth and whiskers fields; telencephalic meningoencephalocele; and secondary ocular degeneration (**Fig. 1c - d**). The midline facial defect is autosomal recessive, however, carriers of the mutation are more brachycephalic individuals than wildtype and were positively selected in the breed, thus the trait has also been described as co-dominant. Affected kittens were generally born live and require euthanasia as the condition is incompatible with life. The heterozygous cats became the hallmark phenotype of the "Contemporary" Burmese and the predominant winners at cat shows.

The controversy of the craniofacial defect and the recognition of other health concerns in non-USA Burmese, such as hypokalemia , orofacial pain and diabetes has led to the isolation of the USA and non-USA breeds and the USA Burmese divided into "Traditional" and "Contemporary" styles; Burmese are now one of the most genetically inbred cat populations worldwide with significantly reduced popularity due to the health concerns . Genetic studies have proven to be highly efficient in populations with high linkage disequilibrium (LD) and inbreeding, particularly companion animals. The LD of the Burmese is amongst the most extended for cat breeds .

A long-term project that initiated with targeted linkage analysis, and, as domestic cat genomic resources improved, progressed to identity by descent mapping, homozygosity mapping and a genome-wide case – control association study (GWAS) suggests *ALX1*

as a major gene controlling craniofacial structure and the variant in *ALX1* is associated with the Burmese brachycephaly and the craniofacial abnormality.

Materials & Methods

Burmese cat sample collection

Cadavers of affected and normal stillborn kittens were voluntarily submitted by Burmese owners from the period of twenty years (1992 – 2012). Approximately 3 ml EDTA anti-coagulated whole blood of normal parents and siblings was also collected and submitted by the owners' veterinarians. For more recent submissions, DNA was supplied by owners on cotton swabs or cytological brushes via buccal swabbing. Pedigrees were supplied by the owners. White blood cells were isolated from the whole blood using standard techniques and DNA from white cells and tissues was isolated by phenol – chloroform extraction , salt precipitation , or using Qiagen kits (Qiagen, Valencia, CA). Genomic DNA from buccal swabs was isolated using the DNAeasy kit (Qiagen). Pedigree relationships were confirmed by parentage analyses .

Markers for Linkage Analysis

Short tandem repeat (STRs) markers for linkage analysis were selected in proximity to candidate genes, including the homeobox gene clusters (*HOXA@, HOXB@, HOXC@, HOXD@*) and *sonic hedgehog* (*SHH*). These genes are mapped by somatic cell hybrid studies to cat chromosomes that have conserved regions of synteny to human chromosomes 7, 11, 12, and 17, respectively . At that time, twenty-four STRs were publically available on cat chromosomes A2q, D1, B4q, E1 in juxtaposition to the candidate genes .

Linkage Analysis

Linkage analysis was conducted using the software package LINKAGE . The kittens with the craniofacial defect were considered congenitally affected with full penetrance of the phenotype, assuming an autosomal recessive mode of inheritance. The allele frequency of 0.5 was estimated for non-genotyped founders of the pedigree since the trait is under positive selection in the Contemporary lines of the breed.

SNP array genotyping

The initial dataset for the SNP array genome-wide analysis comprised 46 cats, including affected Burmese kittens cases that were unrelated as possible and related Burmese, and cats from the closely related breed, Bombay, for controls. Approximately 600 ng of genomic DNA from tissue, blood or buccal swab was submitted to Neogene, Inc (Lincoln, NE, USA) for genotyping on the Illumina Infinium Feline 63K iSelect DNA array (Illumina, Inc., San Diego, CA). Genotyping and analysis was performed as previously described .

Array data analyses

SNP genotyping rate and minor allele frequency was evaluated using PLINK . SNPs with a MAF < 5%, genotyping rate < 90%, and individuals genotyped for < 90% of SNPs were excluded from downstream analyses. An MDS with 2 dimensions was performed using PLINK to evaluate population substructure within cases and controls. Inflation of p-values was evaluated by calculating the genomic inflation factor (λ). The P for each individual was calculated using PLINK. To reduce λ, cats not tightly clustered and/or highly related with a p-hat > 0.3 were removed from downstream analyses. Moreover,

selection for each case to the closest control using the values from the MDS dimensions was attempted. Linkage disequilibrium from position 106,142,990 - 114,551,706 was determined and presented as a plot produced by HAPLOVIEW . To investigate the haplotype, SNPs from the haplotype block (n = 129 SNPs) were exported and visually inspected.

Identity by descent (IBD) analysis was conducted using PLINK . Segmental sharing was surveyed with the command --*segment* using a window of 25 SNPs (~1000 Kb). All the samples were included in the analysis using the function --*all-pairs*. Shared haplotypes between all sample comparisons were plotted and visually inspected.

Homozygosity analysis was conducted using PLINK . A window of 25 SNPs (~1000 Kb) was surveyed for homozygosity, allowing five missing genotypes and a single heterozygous. A homozygous block was defined by five SNPs (or ~250 Kb) and the threshold of homozygosity match was selected as 0.99. The consensus homozygosity block was defined as the overlapping homozygosity block from each individual using the command (--*homo-group*). Minor allele frequency (MAF) was calculated for each SNP using the function --*geno* in PLINK, separating cases from controls. For each SNP, the MAF was plotted along the chromosomal length.

ALX1 genomic analyses

The complete CDS of *ALX1* is publicly available and can be found on chromosome B4: 110145316 – 110165008 in *Felis catus* 6.2. *ALX1* has 4 coding exons; the full CDS and the 5 UTR and 3’ UTR was analyzed on genomic DNA. Primers were tested for efficient product amplification on a DNA Engine Gradient Cycler (MJ Research, GMI, Ramsey,

MN) and the final PCR magnesium concentrations, annealing temperatures, and amplicon sizes for each primer pair are shown in **Supplementary Table 1**. PCR and thermocycling conditions were conducted as previously described . The PCR products were purified and directly sequenced as previously described . Sequences were verified and aligned using the software sequencer version 4.10 (Gene Codes Corp., Ann Arbor, MI).

ALX1 mutation genotyping

The cats of the multi-generational pedigree segregating for the deformity (**Supplementary Fig. 1, 2**), as well as Burmese and other breed cats, were genotyped to confirm segregation of the variant with the craniofacial defect and to determine allele frequency. The University of California – Davis, Veterinary Genetics Laboratory and Langford Veterinary Services has offered the Burmese craniofacial mutation genetic test for approximately three years. A PCR reaction using Alx1-Fdel with a fluorescence label and Alx1-R del (**Supplementary Table 1**) was performed and electrophoretically separated on an ABI DNA analyzer (Applied Biosystems). The predicted size of the wild-type allele was 198 bp and 186 for the variant allele and verified using the software STRand . Langford Veterinary Services, primers for pyrosequencing were designed using PyroMark Assay Design Ver 2.0 (Qiagen, UK) (**Supplementary Table 1**). Pyrosequencing was undertaken after PCR amplification using GoTaq Master Mix (Promega, UK) of genomic DNA isolated from mouth swabs using the Nucleospin Blood kit (Macherey-Nagel, Germany) according to the manufacturer's instructions (PyroGold,

Qiagen) on a PyroMark Q24 (Qiagen). Pyrosequencing PCR was conducted using 95°C for 2 min, followed by 38 cycles of 95°C for 20 sec and 58°C for 40 sec.

Results

Burmese cat sample collection

The phenotype of the craniofacial defect in the affected Burmese cats is unique and distinct, with only mild variations in presentation, therefore diagnosis of affected kittens is not confounded by other congenital birth defects in cats (**Fig 1c - d.**). All affected cats used in the analyses were presented to the investigators and were phenotypically confirmed. Additional stillborn kitten littermates were often submitted and were used as normal siblings when determined phenotypically normal by gross examination. Cats were considered normal if a blood or buccal swab sample had been submitted. Over 488 samples from Burmese cats were ascertained, 83 were stillborn kittens with the craniofacial defect.

Linkage Analysis

A linkage analysis was conducted on two extended families consisting of 124 individuals, which included 47 affected and 62 normal offspring (**Supplementary Fig. 1 and 2)**. Linkage was suggested by four STRs, *FCA863*, *FCA683*, *FCA864* and *FCA866*. STR *FCA864* identified significant complete linkage ($Z = 4.63$, $\Theta = 0.00$) to the craniofacial phenotype suggesting the trait should be localized to cat chromosome B4 (**Table 1**). Fourteen additional STRs, including *FCA105, FCA124, FCA298, FCA327, FCA621, RCA656, FCA785, FCA789, FCA790, FCA791, FCA792, FCA991*, and *FCA992*, were also tested for linkage to the cranial defect data not shown). These

markers did not support linkage and generally excluded 10 – 15 cM flanking the loci. No linkage was suggested with the four *HOX@* clusters and *SHH*, although *HOXC@* is on cat chromosome B4. *FCA864* is located at chrB4: 91513852 - 91514204 in the cat reference assembly - *Felis catus* 6.2 (http://www.ncbi.nlm.nih.gov/assembly/320798).

SNP data analyses

Forty-eight cats, including 23 cases (20 Burmese, three Bombay) and 23 controls (16 Burmese and seven Bombay), were submitted for SNP genotyping. Affected Burmese, Bombay and American Shorthair cats originated from United States and healthy controls from the Burmese and Bombay breeds were selected from the USA and other countries were included in the analysis. The genotyping rate was 0.995, hence all the cats were included in the downstream analysis. After evaluating the genotype qualities of 62,897 SNPs on the array, 43,087 markers passed quality control and were included in the case-control association. Approximately 19,549 SNPs were eliminated for low MAF and 314 SNPs were eliminated for poor genotyping. For haplotype analysis, ROH and IBD, only SNPs with poor genotyping rate were removed from analyses.

Association Studies

Multi-dimensional scaling (MDS) revealed stratification of the cats used in the analysis (**Supplementary Fig. 3**). Two main clusters, one containing Burmese cases and controls and some Bombay, a second tight cluster containing only Burmese controls, and some isolated Bombay were observed. The MDS removed one case and 11 controls leaving 22 cases and 12 controls for the analysis, reducing the genomic inflation from 2.95 - 2.13. Seven cases showed a P> 0.3, and were removed, with a

decrease in inflation to 1.84. Finally, 15 cases (ten Burmese, five Bombay) and 12 controls (11 Burmese, one Bombay) were included in the association analysis and a significant association was identified with several SNPs on chromosome B4 (**Fig. 2**). After permutation testing, only three SNPs on chromosome B4 remained genome – wide significant. SNPs B4.128525117 (position 111,895,171) and B4.128576912 (position 111,938,566) had the most significant association with the trait (**Table 2**). Using the solid spine of LD analysis in Haploview , a haplotype with 92% frequency in the cases from position 106,871,872 - 111,795,395 (~ 5 Mb) was identified. In the controls, smaller blocks are detected as shown in **Fig. 2, Supplementary Fig. 4.** The haplotype was inspected from position 106,142,990 - 114,551,706 and four affected Burmese were key in refining the area containing the gene associated with the phenotype. Two Burmese cats refined the area of association from position 108,534,662 - 112,980,578, while three Bombay showed heterozygous SNPs in the haplotype and the remaining two Bombay were heterozygous for SNPs across the entire ~ 4.5 Mb region, leaving only short block for visual examination, including a 161 Kb block that contains *ALX1* (**Fig. 2, Supplementary Fig. 4**).

When comparing each case to all the other cases included in the IBD analysis, a region on chromosome B4 is shared across the majority of the cats (**Supplementary Fig. 5**). Four cats did not have the complete common ancestral allele. Other regions, such as chromosome D1 showed an extended shared allele across all the Burmese to Burmese comparisons (**Supplementary Fig. 6**).

ROH analysis was conducted on all the available cases (n = 23) and controls (n = 23) separately. Excluding the ROHs detected on the X chromosome shared in 23 cases as

well as in the controls (data not shown), a ROH was detected in 23 cases on chromosome B4 (**Supplementary Fig. 5 & 7**). The ROH spanned 162 SNPs (position 106,754,478 - 112,937,278) and covers ~ 6.2 Mb. No ROH was identified for the control group in this location (**Supplementary Fig. 5 & 7**). Other shorter ROHs were identified on several other chromosomes (**Supplementary Table 2**). Several other reductions in MAF are detected, but none exclusive to cases compared to controls (**Supplementary Fig. 7**).

ALX1 genomic analysis and variant genotyping

The entire *ALX1* CDS sequence was analyzed in ten cats, including five affected Burmese and five controls (domestic shorthair, one Persian, and three Burmese). *ALX1* has one isoform and the length of the coding region of the transcript is 981 bp in human and cats, translating into 326 amino acids. The average CDS homology between human and cat is 93.8% and the protein identity is 97.5%. A 12 bp deletion (c.496delCTCTCAGGACTG) was identified in the coding region of *ALX1* (XM_011288799.1). The variant is predicted *in silico* to be responsible for the lack of four amino acids in the homeobox domain of the protein (**Supplementary Fig. 8**) was further investigated. No other variants were identified during the sequencing effort.

All the unaffected cats in the pedigree were confirmed to be homozygous wild-type or carrier of the 12 bp deletion while all the affected cats were homozygous for the identified variant (**Supplementary Fig. 2 and 3**). Genotyping of over 3,000 Burmese type cats suggests the allele frequency is ~6% in the Burmese population. However, this estimation is biased as breeders know the at-risk cats. The variant was also

genotyped in ~2400 cats from other breeds with brachycephaly, such as Persian, Exotic Shorthair, Scottish Fold, Selkirk Rex and British shorthair, as well as random bred cats. None of the tested cats from other breeds or populations showed the deletion (**Table 3**).

Discussion

Frontonasal dysplasias are a heterogeneous group of disorders . Cases can be sporadic, however, several familial cases have been reported , with two or more of the following clinical signs: true ocular hypertelorism, broadening of the nasal root, median facial cleft affecting the nose and/or upper lip and palate, unilateral or bilateral clefting of the alae nasi, lack of formation of the nasal tip, anterior (rostral) cranium bifidum occultum and a V-shaped or widow's peak frontal hairline . The Burmese craniofacial defect has the same constellation of dysmorphologies as in humans and is a biomedical model for FND (**Fig 1 and 3**). The Burmese craniofacial abnormality was originally described as either maxillonasal hypoplasia or incomplete diprosopus and a mechanism of transformation of the medial nasal part of the frontonasal process was suggested. The dysmorphology was declared a telencephalic meningohydroencephalocele. Defects in *ALX1* are the cause of frontonasal dyspasia type 3 (FND3; OMIM: 613456) .

The genetic analyses of the craniofacial abnormality in the Burmese cats were initiated prior to the development of valuable genetic resources for the cat. Progress was incremental as the positions of the *HOX@* in the cat were identified by somatic cell hybrid analyses , STRs developed, linkage and radiation hybrid maps constructed and synteny between the cat and human genomes established . Before the understanding of the function of *ALX1*, candidate genes on cat chromosome B4, such as *SHH,* were sequenced and eliminated (data not shown). The development of the cat BAC library,

and later the first cat genome assembly also allowed the identification of regional STRs and repeated linkage studies continued to eliminate candidate genes on cat chromosome B4 (data not shown). The cat samples submitted by the Burmese breeders produced a larger, more extended pedigree containing over 300 genotyped cats that included the Burmese, Bombay and American Shorthair breeds (data not shown). Eventually, several different genetic methods identified the same chromosomal region for the craniofacial defect in the Burmese cats and the newly recognized function of *ALX1* suggested a strong candidate gene .

Initially, extended pedigrees of Burmese and Bombay cats segregating for the craniofacial defect supported linkage analyses with STRs, suggesting the craniofacial defect was linked to markers that had been mapped to cat chromosome B4. Recent, intense and rapid selection for the brachycephalic muzzle in the Burmese breed suggested that the region with the causative locus may have high linkage disequilibrium (LD). LD analyses across several cat breeds showed the Burmese had the highest LD amongst cats (~200 Kb) , implying the Burmese would be an efficient breed for analyses on the 63K cat DNA array. The first successful genome-wide association study used non-USA Burmese cats to localize the variant causing hypokalemia with 25 cases and 35 controls and a genomic inflation of 1.8 . As a disease trait, hypokalemia was not under positive selection, thus, the craniofacial defect, also autosomal recessive, would likely require fewer cats since the trait was under intense positive selection and had heterozygous advantage. After correction for sub-structure and relatedness, a GWAS with a genomic inflation of 1.84 using 15 cases and 12 controls also suggested localization of the craniofacial defect to cat chromosome B4. Both haplotype and

identity by descent analysis revealed a 5 - 10 Mb region on chromosome B4 in which four recombinant cats reduced the critical region to 161 Kb, which included the candidate gene *ALX1*. The same region on chromosome B4 was confirmed by a reduction in MAF in the cases versus the controls. Other regions also showed a remarkable reduction in MAF, but the decrease was not unique to the cases; the presence of these regions was expected, since Burmese and the closely related Bombay have other unique phenotypic features under selection. The region that contains the temperature sensitive coloration pattern in *TYR* showed a reduction spanning almost 40 Mb, indicating a historic and positive selective pressure for the trait.

The candidate gene, homeobox transcription factor *ALX1,* is within the short homozygous block spanning 161 Kb. Sequencing of the gene identified a 12 bp deletion, 496delCTCTCAGGACTG. *ALX1* contains two domains: the homeobox domain at position 133 – 191 and the OAR domain at positions 302 – 321 of the protein. The homeobox family transcription factor domain is defined by a highly conserved 60 amino acid sequence that encodes for a helix-turn-helix DNA binding domain (Gehring et al 1994). In vertebrates, *ALX1* regulates the development and survival of mesenchyme-derived elements of the face and neck and complete gene loss of *ALX1* prevents the fusion of frontonasal, nasomedial, nasolateral, and maxillary elements . Uz et al. (2010) identified a 3.7 Mb chromosomal deletion of a region containing *ALX1* that is associated with a frontonasal dysplasia. While normal development of structures originating from the frontal and nasomedial prominences are observed, the presence of bilateral cleft is suggests lack of fusion of nasomedial and nasolateral prominences. In humans, the lack of fusion of the apices of the palatal shelves suggests that embryonic

development might be disrupted before the seventh week of gestation or earlier from studies in mice due to the similar phenotype is hypothesized that *ALX1* has a similar role in early embryogenesis in the feline model. *ALX1* is tuned by several primary mesenchyme cell signals, and controls ingression genes, several skeletogenic differentiation genes and secondary mesenchyme cells specification genes . The 4 amino acid deletion in the mutated feline protein from position 68 - 71, within the homeobox binding domain alters the activity of the element, disrupting the normal development of affected Burmese craniofacial mesenchyme. This 12 bp deletion causes a desired brachcephalic presentation in the heterozygous state and the severely dysmorphic congenital abnormality when homozygous (**Fig 1 and 3)**.

ALX1 is expressed during embryogenesis in mesenchyme of craniofacial primordia (Zhao et al., 1993). *In vivo* studies of *ALX1* have demonstrated the aristaless domain of *ALX1* functions to restrain activity of this transcription factor mainly or completely through its effect on DNA binding . The aristaless domain (OAR) is essential for correct morphogenesis of the cranium and other regions of the body. The deletion of 4 amino acids of the homeobox domain in *ALX1* in the cat demonstrates disruption of the cranium morphogenesis, but only in the homozygous state. Heterozygous cats do have a brachycephalic appearance, thus all variants in *ALX1* may not be lethal.

The Burmese has its origins in cats of Thailand, historically known as the Supilak or the Copper Cat of Siam . The Burmese was accepted for stud book registration by the Cat Fancy Association (CFA) in 1936. The foundation of this breed in the United States originated the importation of a single female, "Wong Mau", from the capital of Rangoon. Wong Mau was phenotypically distinct from the Siamese cats of her homeland in that she

had a distinctively more cobby body frame with a walnut-brown coat color, exhibiting darker brown points. The Burmese has a breed defining coloration mutation at the *Color* (*C*) locus, all cats being homozygous, $c^b c^b$, for a temperature sensitive mutation in *Tyrosinase* (*TYR*) that causes the sable coloration . Genetic studies support Burmese origins from South –East Asia , as well as other closely related breeds such as Bombay, Singapura, Siamese and Korats. The Bombay, Singapura, Asian and Burmilla cat breeds are derived from the Burmese and need to be screened for the craniofacial defect, as well as hypokalemia.

Animal models offer a useful tool to understand the effects of single gene defects. Moreover, breed phenotypes under strong positive selective pressure facilitates the localization of the loci that harbor gene(s) controlling aesthetic features (Gandolfi 2013, Gandolfi 2013). The Persians are one of the oldest cat breeds, presented as the Angora in early cat shows of the late 1800's and early 1900's . Besides Burmese, the craniofacial structure of the breed was also drastically modified after World War II, replacing the moderate facial structure with the drastically brachcephalic structure of the Peked-faced Persian during the 1960's . Persians have influenced many breeds and are the major craniofacial type contributor to the Exotic Shorthair, Himalayan, Scottish Fold, Selkirk Rex and even the modern British Shorthair. Combined, these breeds represent over 60% of the registered cats in the Cat Fanciers' Association in the USA . The Burmese *ALX1* variant was not identified in these brachycephalic breeds. The drastic and rapid change in the Persian family of cat breeds suggests a second gene affecting the craniofacial structure in these breeds. Traditional Burmese have also become more brachycephalic over the past 3 decades and their phenotype cannot be

clearly distinguished from Burmese heterozygous for the *ALX1* variant. Thus, all Burmese need to be genotyped to confirm presence or absence of the variant.

Acknowledgements

This work was supported by funding by the National Center for Research Resources R24 RR016094 and is currently supported by the Office of Research Infrastructure Programs/OD R24OD010928 (LAL), NIH-NIDCR (R03-DE014965-01) and the Winn Feline Foundation (NAB – RFA 1994; 1997, 2001, W10 – 014, W11-041), Cat Health Network (D12FE-506, D12FE-508, D12FE-559**),** the Center for Companion Animal Health (2009-05-F, 2008-36-F) and the Veterinary Genetics Laboratory, School of Veterinary Medicine, University of California - Davis. We greatly appreciate the efforts of the cat breeders (nearly 80 different cat breeders and owners of affected and non-affected lines) for their supplemental funding and donation of samples over the past 20 years, especially Karen Thomas, DVM, JoAnn Arnett, Erica Graf-Webster, Art Graafmanns, Hilary Helmrich, Nancy Reeves, Roger and Rochelle Hornstein for long lasting and continued appreciation and support. We appreciate the technical assistance of Leslie Bach, Amy Young, Mark Ruhe, and Jennifer Grahn.

References

1871a. The Cat-Show. Penny Illustrated Paper, The Naturalist: 511, July 22, 22.

1871b. Crystal Palace - Summer concert today Cat Show on July 13. Penny Illustrated Paper, Amusement: 510, July 08, 11.

Alhaddad, H., Khan, R., Grahn, R.A., Gandolfi, B., Mullikin, J.C., Cole, S.A., Gruffydd-Jones, T.J., Haggstrom, J., Lohi, H., Longeri, M., Lyons, L.A., 2013. Extent of linkage disequilibrium in the domestic cat, *Felis silvestris catus*, and its breeds. PLoS One 8, e53537.

Barrett, J.C., 2009. Haploview: Visualization and analysis of SNP genotype data. Cold Spring Harb Protoc 2009, pdb ip71.

Barrett, J.C., Fry, B., Maller, J., Daly, M.J., 2005. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 21, 263-265.

Blaxter, A., Lievesley, P., Gruffydd-Jones, T., Wotton, P., 1986. Periodic muscle weakness in Burmese kittens. Vet Rec 118, 619-620.

Brouwer, A., ten Berge, D., Wiegerinck, R., Meijlink, F., 2003. The OAR/aristaless domain of the homeodomain protein Cart1 has an attenuating role *in vivo*. Mech Dev 120, 241-252.

CFA, 2004. Cat Fanciers' Association Registration Totals by Color and Breed - 2003, and 1/1/58 to 12/31/03. Cat Fanciers' Almanac 20, 72-86.

Clutterbuck, M.R., 2004. Siamese Cats: Legends and Reality. White Lotus Press, Bangkok, Thailand.

Dee, C.T., Szymoniuk, C.R., Mills, P.E., Takahashi, T., 2013. Defective neural crest migration revealed by a Zebrafish model of *Alx1*-related frontonasal dysplasia. Hum Mol Genet 22, 239-251.

Ettensohn, C.A., Illies, M.R., Oliveri, P., De Jong, D.L., 2003. Alx1, a member of the Cart1/Alx3/Alx4 subfamily of Paired-class homeodomain proteins, is an essential component of the gene network controlling skeletogenic fate specification in the sea urchin embryo. Development 130, 2917-2928.

Fryburg, J.S., Persing, J.A., Lin, K.Y., 1993. Frontonasal dysplasia in two successive generations. Am J Med Genet 46, 712-714.

Gandolfi, B., Gruffydd-Jones, T.J., Malik, R., Cortes, A., Jones, B.R., Helps, C.R., Prinzenberg, E.M., Erhardt, G., Lyons, L.A., 2012. First *WNK4*-hypokalemia animal model identified by genome-wide association in Burmese cats. PloS one 7, e53173.

Gandolfi, B., Outerbridge, C.A., Beresford, L.G., Myers, J.A., Pimentel, M., Alhaddad, H., Grahn, J.C., Grahn, R.A., Lyons, L.A., 2010. The naked truth: sphynx and Devon rex cat breed mutations in *KRT71*. Mamm Genome 21, 509-515.

Haworth, K.E., Islam, I., Breen, M., Putt, W., Makrinou, E., Binns, M., Hopkinson, D., Edwards, Y., 2001. Canine TCOF1; cloning, chromosome assignment and genetic analysis in dogs with different head types. Mamm Genome 12, 622-629.

Huber, W., Lups, P., 1968. [Biometric and mechanism of development of the symptoms of brachycephaly in the dog]. Archiv der Julius Klaus-Stiftung fur Vererbungsforschung, Sozialanthropologie und Rassenhygiene 43-44, 57-65.

Hunemeier, T., Salzano, F.M., Bortolini, M.C., 2009. TCOF1 T/Ser variant and brachycephaly in dogs. Anim Genet 40, 357-358.

Jones, B.R., Gruffydd-Jones, T.J., 1990. Hypokalemia in the cat. Cornell Vet. 80, 13-16.

Koch, D., Wiestner, T., Balli, A., Montavon, P., Michel, E., Scharf, G., Arnold, S., 2012. Proposal for a new radiological index to determine skull conformation in the dog. Schweizer Archiv fur Tierheilkunde 154, 217-220.

Kruijsen, N., Wayop, I., 2011. [Does the bulldog still have a future?]. Tijdschrift voor diergeneeskunde 136, 268-269.

Kurushima, J.D., Lipinski, M.J., Gandolfi, B., Froenicke, L., Grahn, J.C., Grahn, R.A., Lyons, L.A., 2012. Variation of cats under domestication: genetic assignment of domestic cats to breeds and worldwide random-bred populations. Anim. Genet.

Lathrop, G.M., Lalouel, J.M., 1984. Easy calculations of lod scores and genetic risks on small computers. American Journal Human Genetics 36, 460-465.

Lathrop, G.M., Lalouel, J.M., 1988. Efficient computations in multilocus linkage analysis. Am. J. Hum. Genet. 42, 498-505.

Lathrop, G.M., Lalouel, J.M., Julier, C., Ott, J., 1984. Strategies for multilocus linkage analysis in humans. Proc Nat Acad Sci USA 81, 3443-3446.

Lipinski, M., Amigues, Y., Blasi, M., Broad, T., Cherbonnel, C., Cho, G., Corley, S., Daftari, P., Delattre, D., Dileanis, S., 2007. An international parentage and identification panel for the domestic cat (Felis catus). Animal genetics 38, 371-377.

Lipinski, M.J., Froenicke, L., Baysac, K.C., Billings, N.C., Leutenegger, C.M., Levy, A.M., Longeri, M., Niini, T., Ozpinar, H., Slater, M.R., Pedersen, N.C., Lyons, L.A., 2008. The ascent of cat breeds: genetic evaluations of breeds and worldwide random-bred populations. Genomics 91, 12-21.

Lyons, L.A., Imes, D.L., Rah, H.C., Grahn, R.A., 2005. *Tyrosinase* mutations associated with Siamese and Burmese patterns in the domestic cat (*Felis catus*). Animal Genetics 36, 119-126.

Menotti-Raymond, M., David, V.A., Agarwala, R., Schaffer, A.A., Stephens, R., O'Brien, S.J., Murphy, W.J., 2003. Radiation hybrid mapping of 304 novel microsatellites in the domestic cat genome. Cytogenet Genome Res 102, 272-276.

Menotti-Raymond, M., David, V.A., Lyons, L.A., Schaffer, A.A., Tomlin, J.F., Hutton, M.K., O'Brien, S.J., 1999. A genetic linkage map of microsatellites in the domestic cat (Felis catus). Genomics 57, 9-23.

Miller, S.A., Dykes, D.D., Polesky, H.F., 1988. A simple salting out procedure for extracting DNA from human nucleated cells. Nuc. Acid Res. 16, 1215.

Moreno Fuenmayor, H., 1980. The spectrum of frontonasal dysplasia in an inbred pedigree. Clinical Genetics 17, 137-142.

Morris, D., 1999. Cat Breeds of the World. Penguin Books, New York.

Murphy, W.J., Menotti-Raymond, M., Lyons, L.A., Thompson, M.A., O'Brien, S.J., 1999. Development of a feline whole genome radiation hybrid panel and comparative mapping of human chromosome 12 and 22 loci. Genomics 57, 1-8.

Nevin, N.C., Leonard, A.G., Jones, B., 1999. Frontonasal dysostosis in two successive generations. Am J Med Genet 87, 251-253.

Noden, D.M., Evans, H.E., 1986. Inherited homeotic midfacial malformations in Burmese cats. J Craniofac Genet Dev Biol Suppl 2, 249-266.

O'Brien, S.J., Cevario, S.J., Martenson, J.S., Thompson, M.A., Nash, W.G., Chang, E., Graves, J.A., Spencer, J.A., Cho, K.W., Tsujimoto, H., Lyons, L.A., 1997a. Comparative gene mapping in the domestic cat *(Felis catus*). J Hered 88, 408-414.

O'Brien, S.J., Wienberg, J., Lyons, L.A., 1997b. Comparative genomics: lessons from cats. Trends Genet 13, 393-399.

Oechtering, G.U., Schluter, C., Lippert, J.P., 2010. [Brachycephaly in dog and cat: a "human induced" obstruction of the upper airways]. Pneumologie 64, 450-452.

Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M.A., Bender, D., Maller, J., Sklar, P., de Bakker, P.I., Daly, M.J., Sham, P.C., 2007. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am. J. Hum. Genet. 81, 559-575.

Rand, J.S., Bobbermien, L.M., Hendrikz, J.K., Copland, M., 1997. Over representation of Burmese cats with diabetes mellitus. Australian veterinary journal 75, 402-405.

Regodon, S., Robina, A., Franco, A., Vivo, J.M., Lignereux, Y., 1991. [Radiological and statistical determination of the morphological type of the canine skull: dolichocephaly, mesocephaly and brachycephaly]. Anatomia, histologia, embryologia 20, 129-138.

Roberts, T., McGreevy, P., Valenzuela, M., 2010. Human induced rotation and reorganization of the brain of domestic dogs. PloS one 5, e11946.

Rusbridge, C., Heath, S., Gunn-Moore, D.A., Knowler, S.P., Johnston, N., McFadyen, A.K., 2010. Feline orofacial pain syndrome (FOPS): a retrospective study of 113 cases. Journal of feline medicine and surgery 12, 498-508.

Sambrook, J., Russel, D.W., 2001. Preparation and analysis of eukaryotic genomic DNA., Molecular Cloning, 3rd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, pp. 6.1-6.30.

Schmidt, M.J., Neumann, A.C., Amort, K.H., Failing, K., Kramer, M., 2011. Cephalometric measurements and determination of general skull type of Cavalier King Charles Spaniels. Veterinary radiology & ultrasound : the official journal of the American College of Veterinary Radiology and the International Veterinary Radiology Association 52, 436-440.

Schoenebeck, J.J., Hutchinson, S.A., Byers, A., Beale, H.C., Carrington, B., Faden, D.L., Rimbault, M., Decker, B., Kidd, J.M., Sood, R., Boyko, A.R., Fondon, J.W., 3rd, Wayne, R.K., Bustamante, C.D., Ciruna, B., Ostrander, E.A., 2012. Variation of BMP3 contributes to dog breed skull diversity. PLoS genetics 8, e1002849.

Sedano, H.O., Cohen, M.M., Jr., Jirasek, J., Gorlin, R.J., 1970. Frontonasal dysplasia. J Pediatr 76, 906-913.

Sedano, H.O., Gorlin, R.J., 1988. Frontonasal malformation as a field defect and in syndromic associations. Oral Surg Oral Med Oral Pathol 65, 704-710.

Sekeles, E., 1981. Craniofacial and skeletal malformaitons in a cat. Feline Prac 11.

Sponenberg, D.P., Graf-Webster, E., 1986. Hereditary meningoencephalocele in Burmese cats. J Hered 77, 60.

Toonen, R.J., Hughes, S., 2001. Increased throughput for fragment analysis on an ABI PRISM 377 automated sequencer using a membrane comb and STRand software. Biotechniques 31, 1320-1324.

Toriello, H.V., Higgins, J.V., Walen, A., Waterman, D.F., 1985. Familial occurrence of a developmental defect of the medial nasal processes. American Journal Medical Genetics 21, 131-135.

Twigg, S.R., Versnel, S.L., Nurnberg, G., Lees, M.M., Bhat, M., Hammond, P., Hennekam, R.C., Hoogeboom, A.J., Hurst, J.A., Johnson, D., Robinson, A.A., Scambler, P.J., Gerrelli, D., Nurnberg, P., Mathijssen, I.M., Wilkie, A.O., 2009. Frontorhiny, a distinctive presentation of frontonasal dysplasia caused by recessive mutations in the ALX3 homeobox gene. Am J Hum Genet 84, 698-705.

Uz, E., Alanay, Y., Aktas, D., Vargel, I., Gucer, S., Tuncbilek, G., von Eggeling, F., Yilmaz, E., Deren, O., Posorski, N., Ozdag, H., Liehr, T., Balci, S., Alikasifoglu, M., Wollnik, B., Akarsu, N.A., 2010. Disruption of ALX1 causes extreme microphthalmia and severe facial clefting: expanding the spectrum of autosomal-recessive ALX-related frontonasal dysplasia. Am J Hum Genet 86, 789-796.

Wu, E., Vargevik, K., Slavotinek, A.M., 2007. Subtypes of frontonasal dysplasia are useful in determining clinical prognosis. Am J Med Genet A 143A, 3069-3078.

Zhao, G.Q., Eberspaecher, H., Seldin, M.F., de Crombrugghe, B., 1994. The gene for the homeodomain-containing protein Cart-1 is expressed in cells that have a chondrogenic potential during embryonic development. Mech Dev 48, 245-254.

Zhao, G.Q., Zhou, X., Eberspaecher, H., Solursh, M., de Crombrugghe, B., 1993. Cartilage homeoprotein 1, a homeoprotein selectively expressed in chondrocytes. Proceedings of the National Academy of Sciences of the United States of America 90, 8633-8637.

Zhao, Q., Behringer, R.R., de Crombrugghe, B., 1996. Prenatal folic acid treatment suppresses acrania and meroanencephaly in mice mutant for the Cart1 homeobox gene. Nat Genet 13, 275-283.

Zook, B.C., 1983. Encephalocele and other congenital craniofacial anomalies in Burmese cats. Vet Med/Small Anim Clin 78, 695-701.

Tables

Table 1. Linkage of feline STRs on chromosome B4 to the Burmese craniofacial defect.

		θ					Max	Max
Marker 1	**Marker 2**	**0.00**	**0.01**	**0.05**	**0.10**	**0.20**	**theta, θ**	**LOD Z**
FCA683	Defect	- ∞	-0.82	2.10	2.87	2.83	0.140	3.00
FCA863	Defect	- ∞	-10.57	-4.66	-2.44	-0.75	---------	--------
FCA864	Defect	4.63	4.56	4.23	3.81	2.87	0.000	4.63
FCA866	Defect	- ∞	0.49	1.56	1.73	1.41	0.097	1.73
FCA683	FCA863	- ∞	1.62	3.22	3.49	3.02	0.099	3.49
FCA683	FCA864	- ∞	1.00	1.51	1.59	1.41	0.096	1.59
FCA683	FCA866	- ∞	-0.57	0.55	0.81	0.75	0.127	0.84
FCA863	FCA864	- ∞	--6.70	-3.33	-1.96	-0.74	---------	--------
FCA863	FCA866	- ∞	-8.63	-3.96	-2.15	-0.71	---------	--------
FCA864	FCA866	- ∞	-4.99	-2.83	-1.88	-0.93	---------	--------

Table 2. Genome-wide significantly associated SNPS after 100,000 permutation testing.

SNP ID	Chromosome	Position	*P* value	P_{genome} value
B4.128525117	B4	111895171	$6.38e^{-7}$	0.014
B4.128576912	B4	111938566	$1.33e^{-6}$	0.031
B4.128654054	B4	112016581	$2.23e^{-6}$	0.045

Table 3. Frequency of the *ALX1* variant in different cat breeds*.

Group	Tested	Carrier	Wildtype
Pedigree	230	119	28
Asian	2	0	2
Australian Mist	1	0	1
Bombay	133	43	90
Burmese	3250	151	3099
Burmilla	5	0	5
Tonkinese	6	0	6
Burmese Breeds	3264	194	3070
Other Breeds	2456	0	2456

*Cats homozygous for the variant are affected and stillborn. From the pedigree, 83 affected kittens were all homozygous for the *ALX1* variant. The total number of pedigree cats tested was increased after the linkage analyses. Testing of other cat breeds was performed at the University of California, Veterinary Genetics Laboratory, California, USA and the Langford Veterinary Services, Bristol, UK.

Figure Legends

Figure 1. Variation of the Burmese cat breed's craniofacial structure. A) Traditional lines are not as extreme, but selection has continued for the past 30 years for a more extreme type that is not associated with congenital abnormalities. Some Traditional lines and contemporary lines are now difficult to distinguish phenotypically. Thus, all Burmese need to be genotyped to confirm presence or absence of the variant.

B) The Contemporary style Burmese has extreme brachycephyla and the phenotype is association with the craniofacial defect. C) Frontal view displays duplication of the maxillary processes and agenesis of the medial nasal prominence. D) Lateral view displays abnormal development of the maxillary processes and ocular degeneration. Photographs courtesy of Nancy Reeves, Isabelle Marchand and Richard Katris – Chanan Photography.

Figure 2. Manhattan plot of the Burmese head deformity GWAS and SNPs genotypes within chromosome B4 haplotype. a. The plot represents the P_{raw} (top) and P_{genome} (bottom) values of each SNP included in the case-control association study. The association study compared the affected Burmese and Bombay cats. A significant association with chromosome B4 was detected. b. The area from SNP B4.121572441 (position 106,142,990) to SNP B4.114551707 (position 114,551,707) spans ~ 8.4 Mb. The two red vertical dashed lines represent the region of the single haplotype containing *ALX1*, from SNP B4.126353636 (position 110,094,604) to SNP B4.126530474 (position 110,255,914) spanning 161 Kb. Each SNP is represented by two squares where markers are on the x-axis and individuals on the y-axis. Gray boxes represent the major allele in the cases and black squares represent the minor.

Figure 3. Three dimensional CT reconstructions of Burmese cat crania. Top) Frontal views of normal stillborn littermate of a Burmese with the craniofacial defect. Bottom) Lateral and dorsal-ventral view of Burmese kitten with a craniofacial defect. Normal kittens may carry the *ALX1* variant. Affected kittens are homozygous for the variant and have the hallmark features of FND.

Supplementary Figure Legends

Supplementary Figure 1. Pedigree segregating for the Burmese Craniofacial Defect. Circles represent females, squares represent males, diamonds are unknown gender. Open symbols indicate phenotypically normal animals, filled symbols indicate affected cats, half-filled are obligate carriers. A small filled circle represents a "breeding node" for parental cats. Numbers under the symbols represent the laboratory sample number. Genotypes for the linked marker *FCA864* are represented below the identification numbers or names. The base pair size of the microsatellite marker was converted to a single number to distinguish the allele. No data is represented by dashes, "--".

Supplementary Figure 2. Pedigree segregating for the Burmese Craniofacial Defect. Circles represent females, squares represent males, diamonds are unknown gender. Open symbols indicate phenotypically normal animals, filled symbols indicate affected cats, half-filled are obligate carriers. A small filled circle represents a "breeding node" for parental cats. Numbers under the symbols represent the laboratory sample number. Genotypes for the linked marker *FCA864* are represented below the identification numbers or names. The base pair size of the microsatellite marker was converted to a single number to distinguish the allele. No data is represented by dashes, "--".

Supplementary Figure 3. Multidimensional scaling (MDS) analysis for population stratification of Burmese. Forty-six samples were plotted for principle components 1 and 2, a. represents the distribution of cases and controls, b. shows the distribution of the samples based on breed, and c. The Burmese controls on the lower left of each plot were eliminated from the case-control analysis.

Supplementary Figure 4. Haplotype analysis of Burmese cases and controls for the craniofacial defect. Position 106,871,872 - 111,795,156 of chromosome B4 in cases and controls. a. LD block identified by HAPLOVIEW across all the cases, spanning 4,923 Kb. b. Haplotypes sequence and frequencies across the 4,923 Kb regions. The main haplotype is squared in red and shows a frequency of 92% across cases. c. LD blocks identified by HAPLOVIEW in the correspondent region across all controls included in the study. d. Haplotypes sequence and frequencies for each identified LD block within the 4,923 Kb region in the control cats.

Supplementary Figure 5. **Identity by descent (IBD) and minor allele frequency (MAF) analyses for chromosome B4**. The horizontal lines in the graph represent all the IBD regions (shared alleles) on chromosome B4 between all the cats included in the analysis. a. Each case is compared to all the other cases included in the analysis. Each group of comparison (breed to breed) is color-coded. Vertical black dashed lines represent a shared IDB region in common between almost all cases. b. Each case is compared to all the controls included in the study. c. Controls versus controls comparison of shared IBD. Cases versus controls and controls versus control comparisons do not show any shared IBD across all the specimens. (Bottom) Graphical representation of the MAF differences within the affected samples (black line) and the control

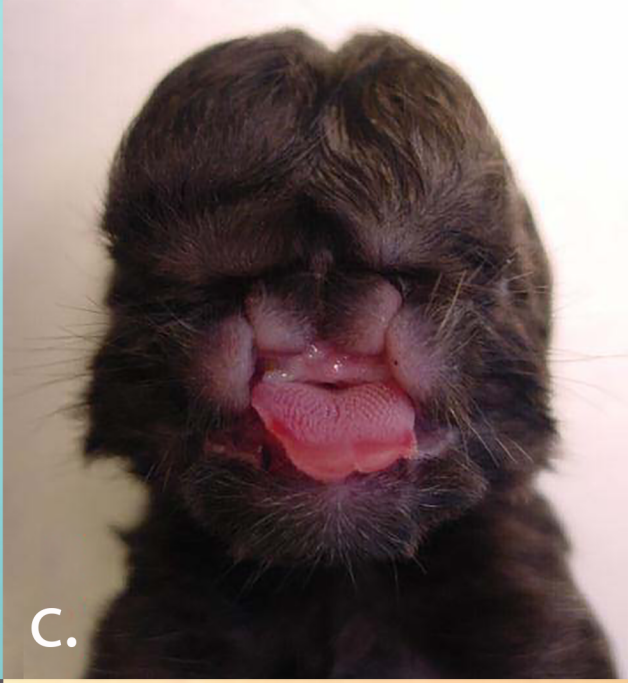
samples (red line) across all the *Felis catus* chromosomes. The black line represents the MAF within the cases and is compared with the MAF within the controls for each SNP. The red dashed line represent the MAF mean for the chromosome within the cases and the black dashed line the MAF mean within the controls. Several gaps are present in the current genome assembly, thus SNPs surrounding the gaps are connected with straight lines.

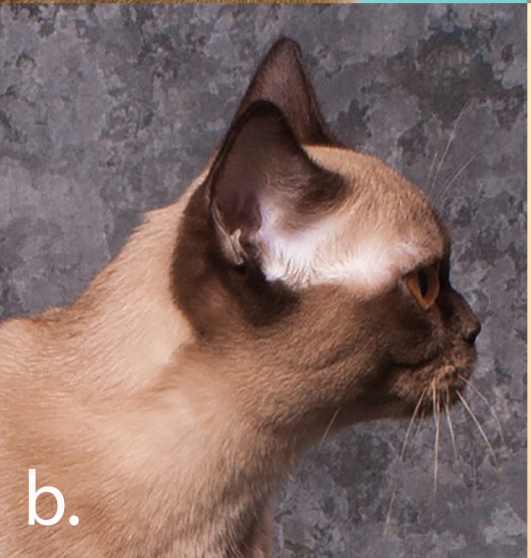
Supplementary Figure 6. Identity by descent (IBD) analysis for chromosome D1. The horizontal lines in the graph represent all the IBD regions (shared alleles) on chromosome D1 between all the cats included in the analysis. a. Each case is compared to all the other cases included in the analysis. Each group of comparison (breed to breed) is color-coded. b. Each case is compared to all the controls included in the study. c. Controls versus controls comparison of shared IBD. A common IBD is shared across the majority of East Asian breeds. The trait contained in the IBD region is a phenotypic trait responsible for the Burmese point coloration, fixed within the breed and confirmed by other analysis included in this study.

Supplementary Figure 7. Full chromosomal minor allele frequency (MAF) comparison within cases and controls. Graphical representation of the MAF differences within the affected samples (black line) and the control samples (red line) across all the *Felis catus* chromosomes. The black line represents the MAF within the cases and is compared with the MAF within the controls for each SNP. The red dashed line represent the MAF mean for the chromosome within the cases and the black dashed line the MAF mean within the controls. Several gaps are present in the current genome assembly, thus SNPs surrounding the gaps are connected with straight lines.

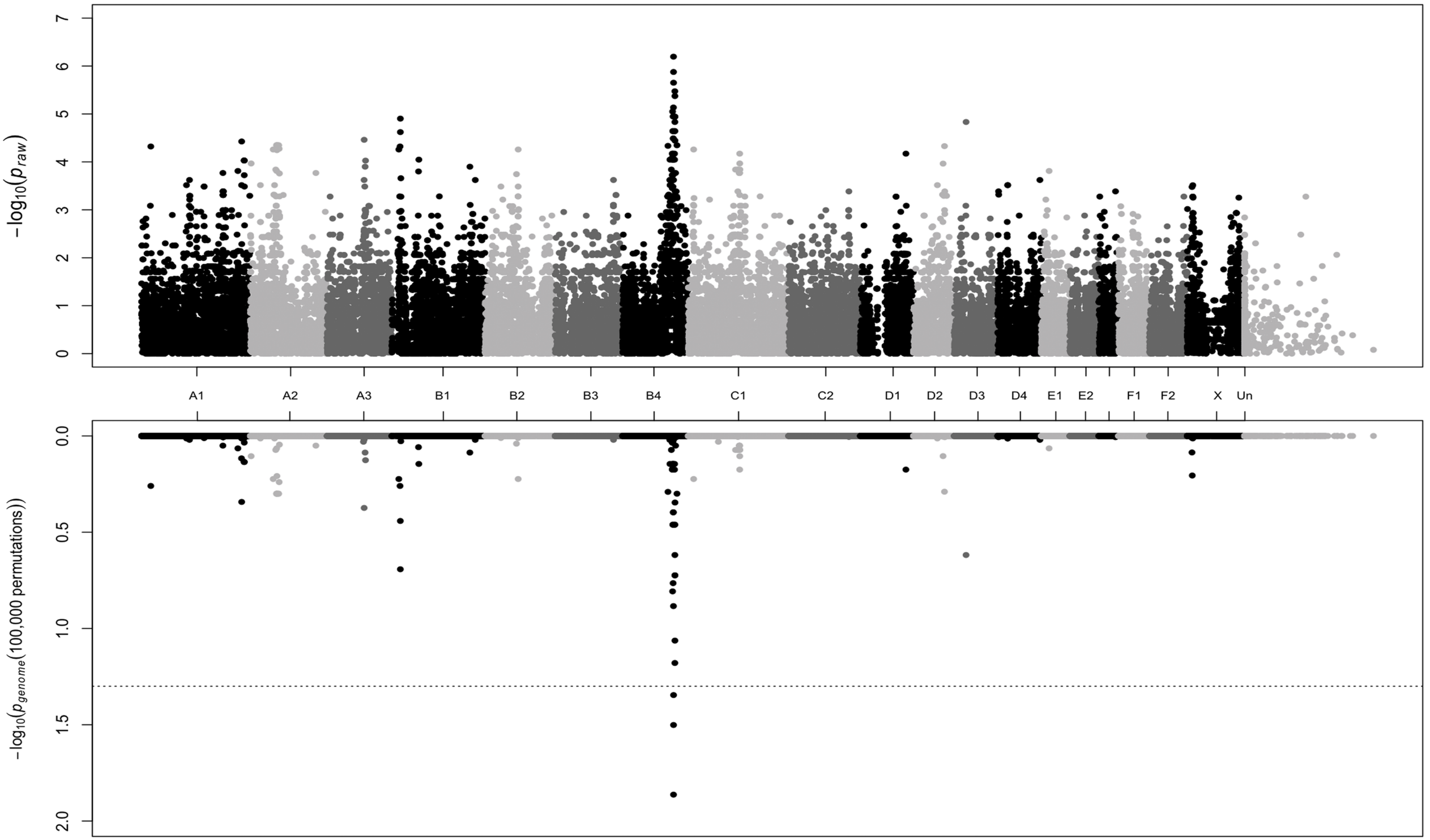
Supplementary Figure 8: **Protein alignment of the *Cart1* wildtype and mutated alleles**. The mutation, underlined in red, is responsible for the lack of 4 amino acids (*in silico* prediction) in the homeobox domain. In the human protein, the homeobox domain starts at position 132 and ends at position 191 of the amino acid chain. Underlined in blue is the *Cart1* OAR domain, which starts at position 306 and ends at position 319 of the Cart1 protein.


a.
© Chanan

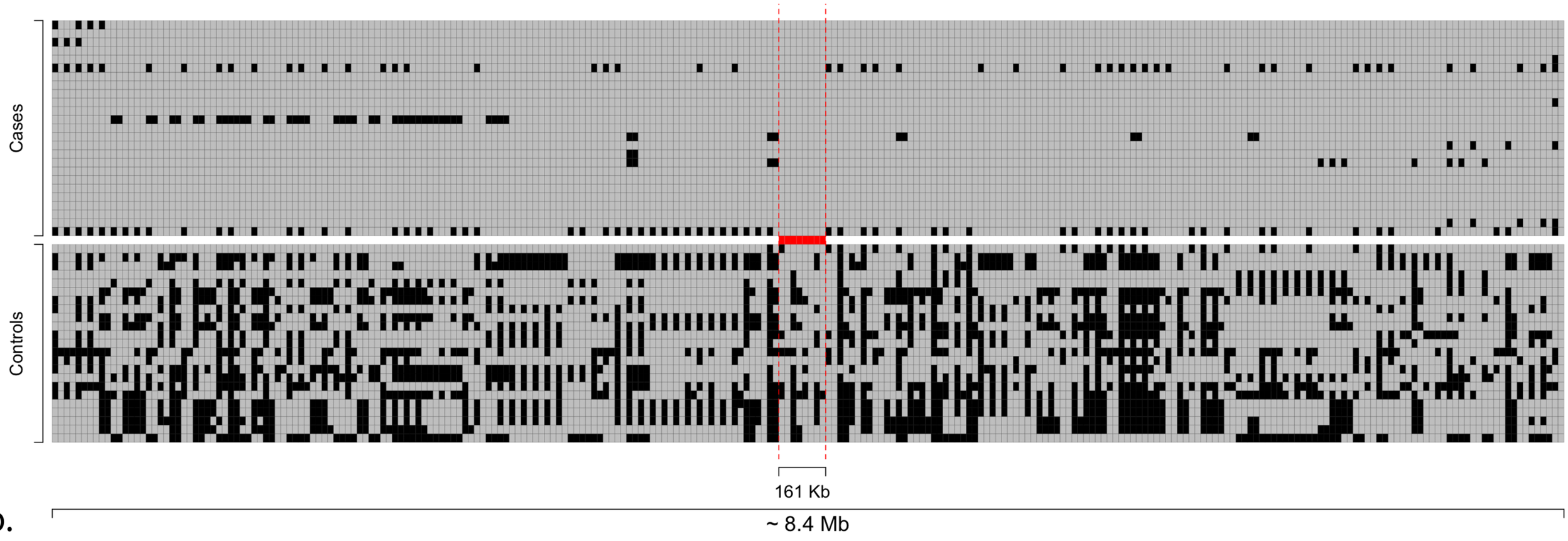

c.


b.

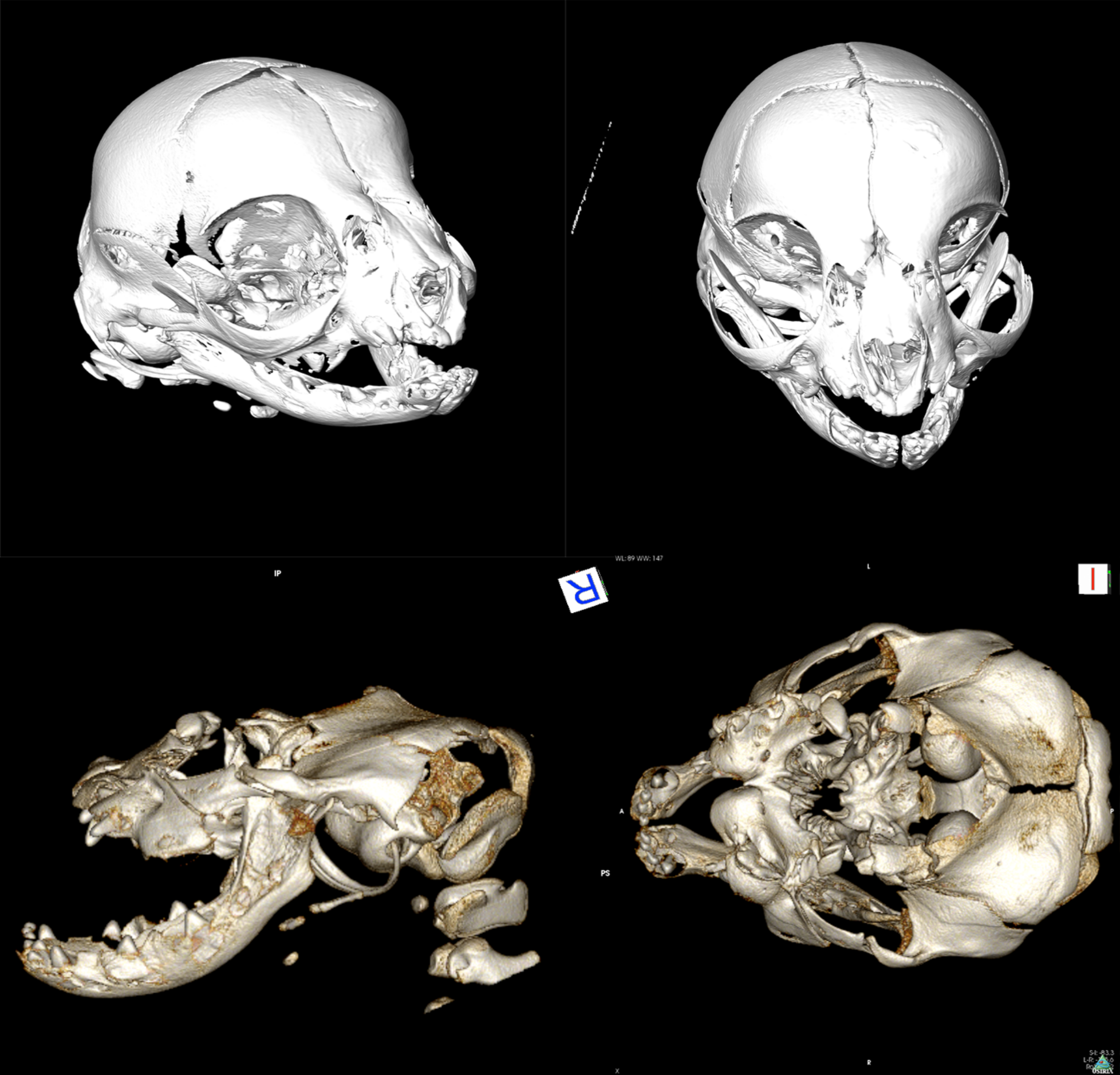

d.
27 2:49


$-\log_{10}(p_{raw})$
0
1
2
3
4
5
6
7
A1
A2
A3
B1
B2
B3
B4
C1
C2
D1
D2
D3
D4
E1
E2
F1
F2
X
Un
$-\log_{10}(p_{genome}(100{,}000\ permutations))$
0.0
0.5
1.0
1.5
2.0

a.


Cases
Controls
161 Kb
~ 8.4 Mb

b.


WL: 89 WW: 147
IP
L
R
A
P
PS
R
X

Supplementary Table 1. PCR and primers for analysis of cat *ALX1*.

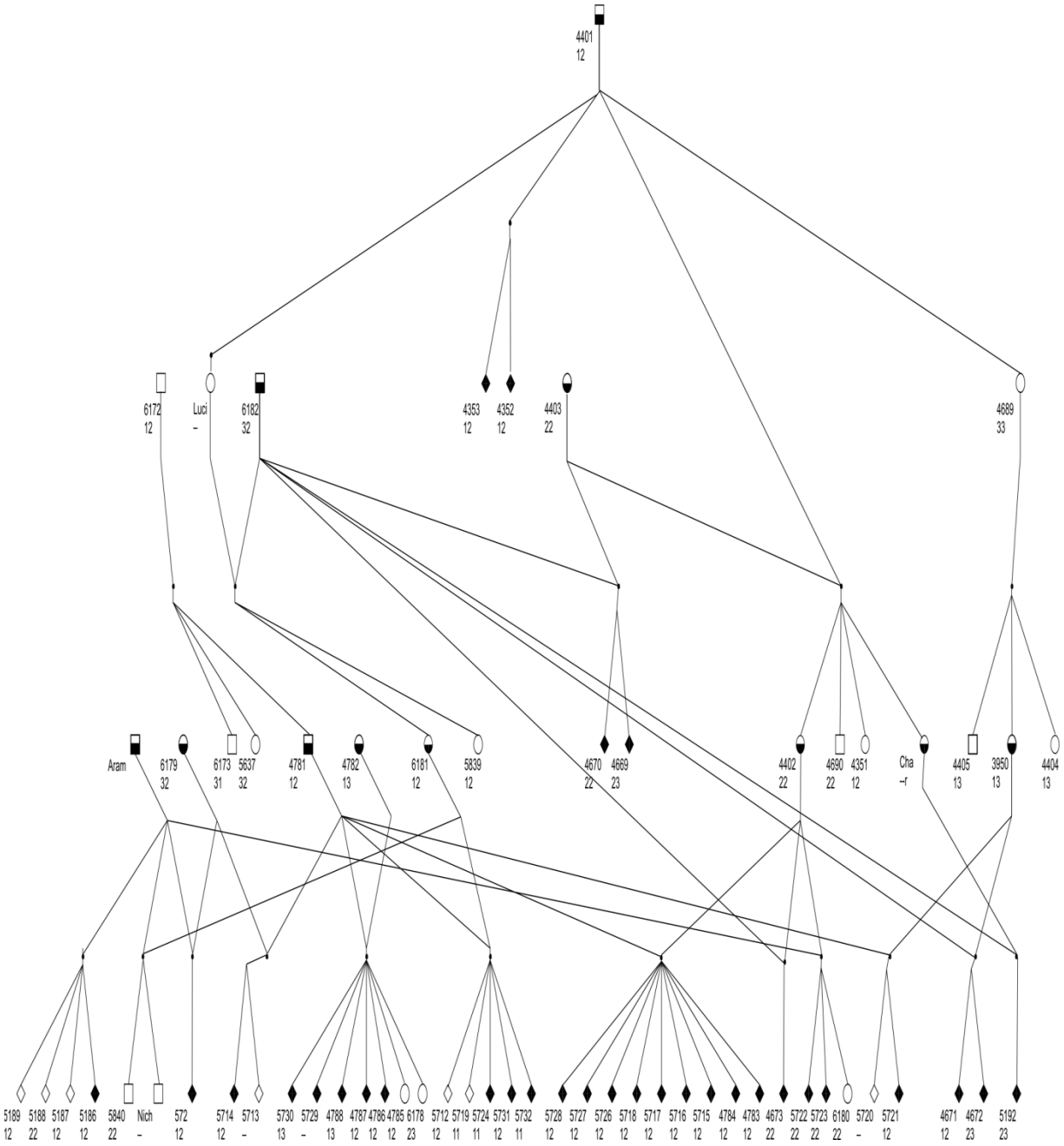
Forward	Reverse	Mg	Tm
GGACGTATTAAGGGCTCGGAGC	TAAAACGCTCGCAGTTCCACCG	1.5 mM	58 °C
AAATCATTAACAGACTGCTTTCCTGA	ATGGTTCTAGTCTTTAGTGAGAGGATCA*	2 mM	58 °C
TTAGTGATTTTGTTGACCTGGTTTGTGT	TAAAATGCTCTCCTGGCACCTGG	1.5 mM	60 °C
TAAGGGGACAAAAGTGAGAATGCG	CGTTTGTGGAGACTGATGGATGGT	1.5 mM	60 °C
Pyrosequencing			
Biotin-GAAAACCCATTACCCGGATGTAT	CTTCATTTGGCTCCTACCTGGA		
CCTGGACTCTGGCCTCCGTGA[$]			

*for genotyping, this primer was labelled with FAM dye.
[$]This primer is for sequencing in the pyrosequencing reaction.

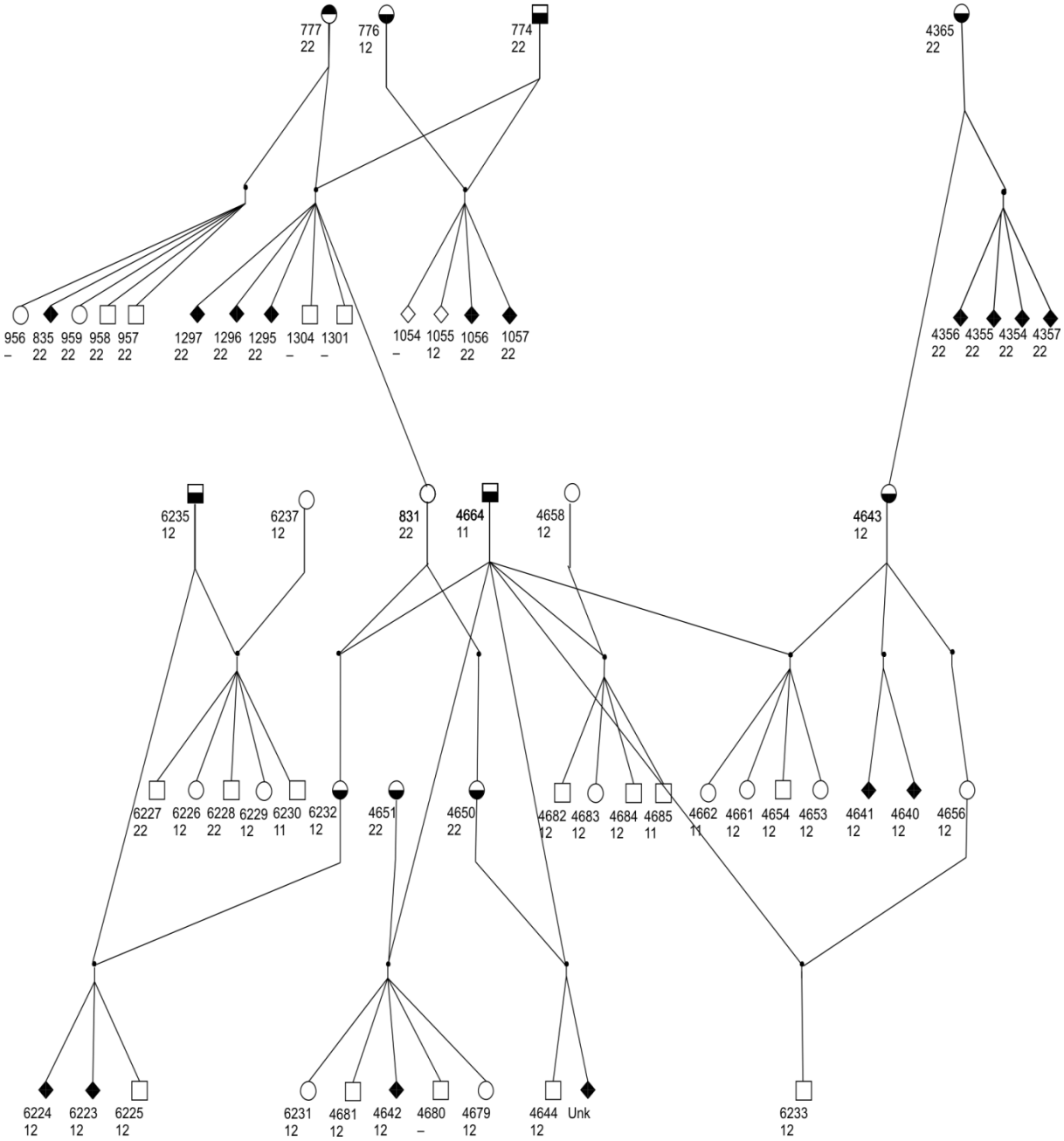
Supplementary Table 2. Consensus details homozygous regions across the affected cats.

Chr.	SNP1	SNP2	bp start	bp end	Kb	# SNPs	# cats
X	chrX.38990530	chrX.40531503	31154766	32362304	1207.54	26	23
X	chrX.36970662	chrX.38733350	29539864	30953514	1413.65	38	23
B4	chrB4.122309957	chrB4.129821393	106754478	112937278	6182.8	162	23
X	chrX.17358465	chrX.26762022	13769102	21616256	7847.15	201	21
D1	chrD1.29042292	chrD1.31719770	23604822	25470200	1865.38	61	21
B3	chrUn.36239579	chrB3.37818450	30716536	32150500	1433.96	38	21
D3	chrD3.10607895	chrA1.170901230	8276516	10296770	2020.25	55	20
D1	chrD1.74373646	chrD1.77044135	47727880	49998734	2270.85	69	20
B4	chrB4.133640812	chrB4.133943775	116050184	116294718	244.534	5	20
B1	chrB1.58452923	chrB1.58795359	45352082	45619806	267.724	9	20
B1	chrB1.55781749	chrB1.56493341	43293324	43849984	556.66	18	20

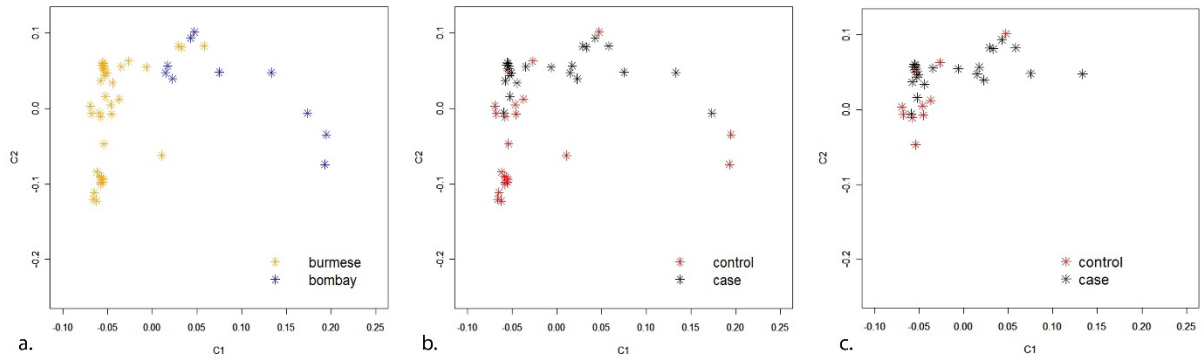
Supplementary Figures:



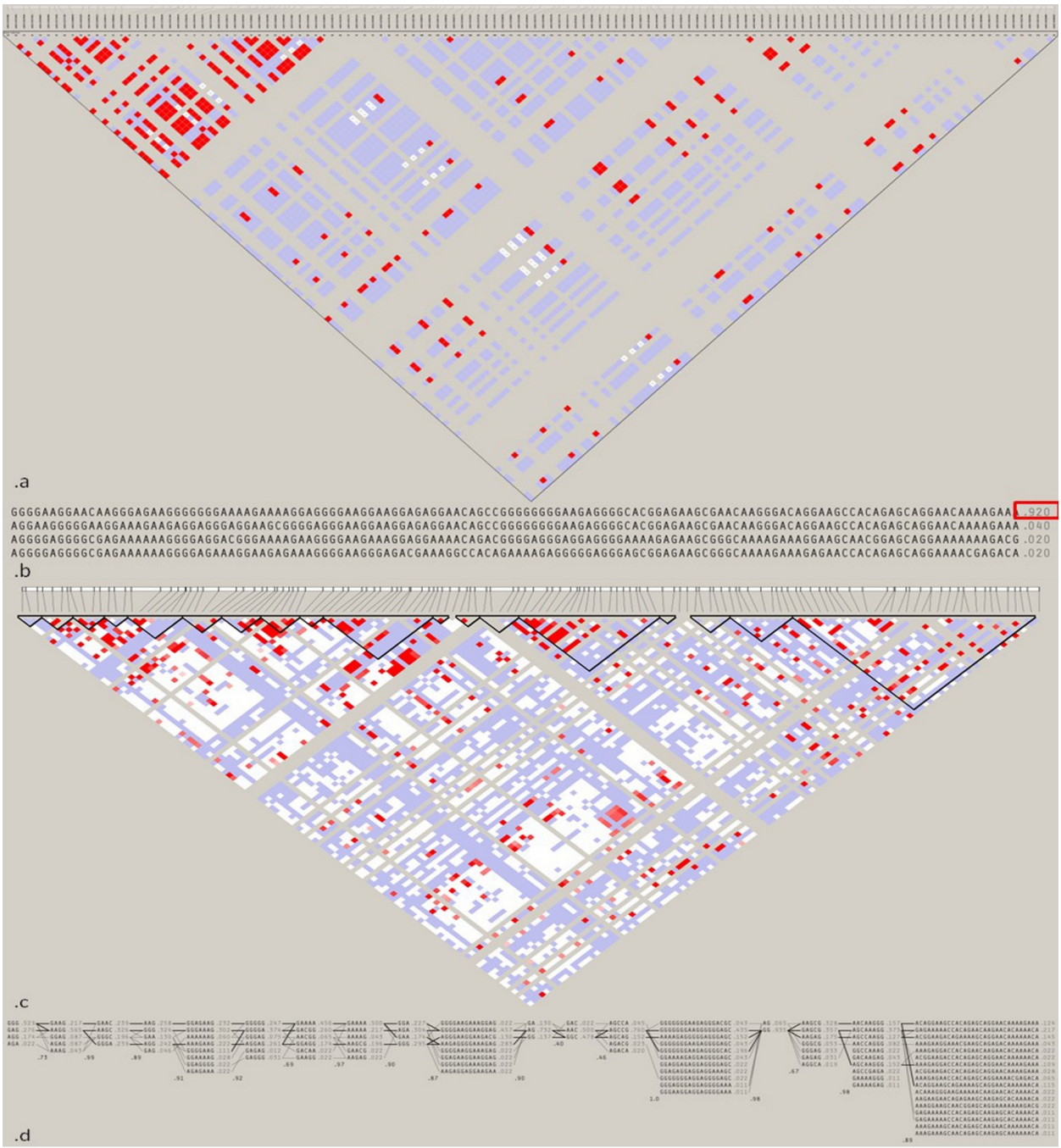
Supplementary Figure 1. Pedigree segregating for the Burmese Craniofacial Defect. Circles represent females, squares represent males, diamonds are unknown gender. Open symbols indicate phenotypically normal animals, filled symbols indicate affected cats, half-filled are obligate carriers. A small filled circle represents a "breeding node" for parental cats. Numbers under the symbols represent the laboratory sample number. Genotypes for the linked marker *FCA864* are represented below the identification numbers or names. The base pair size of the microsatellite marker was converted to a single number to distinguish the allele. No data is represented by dashes, "--".



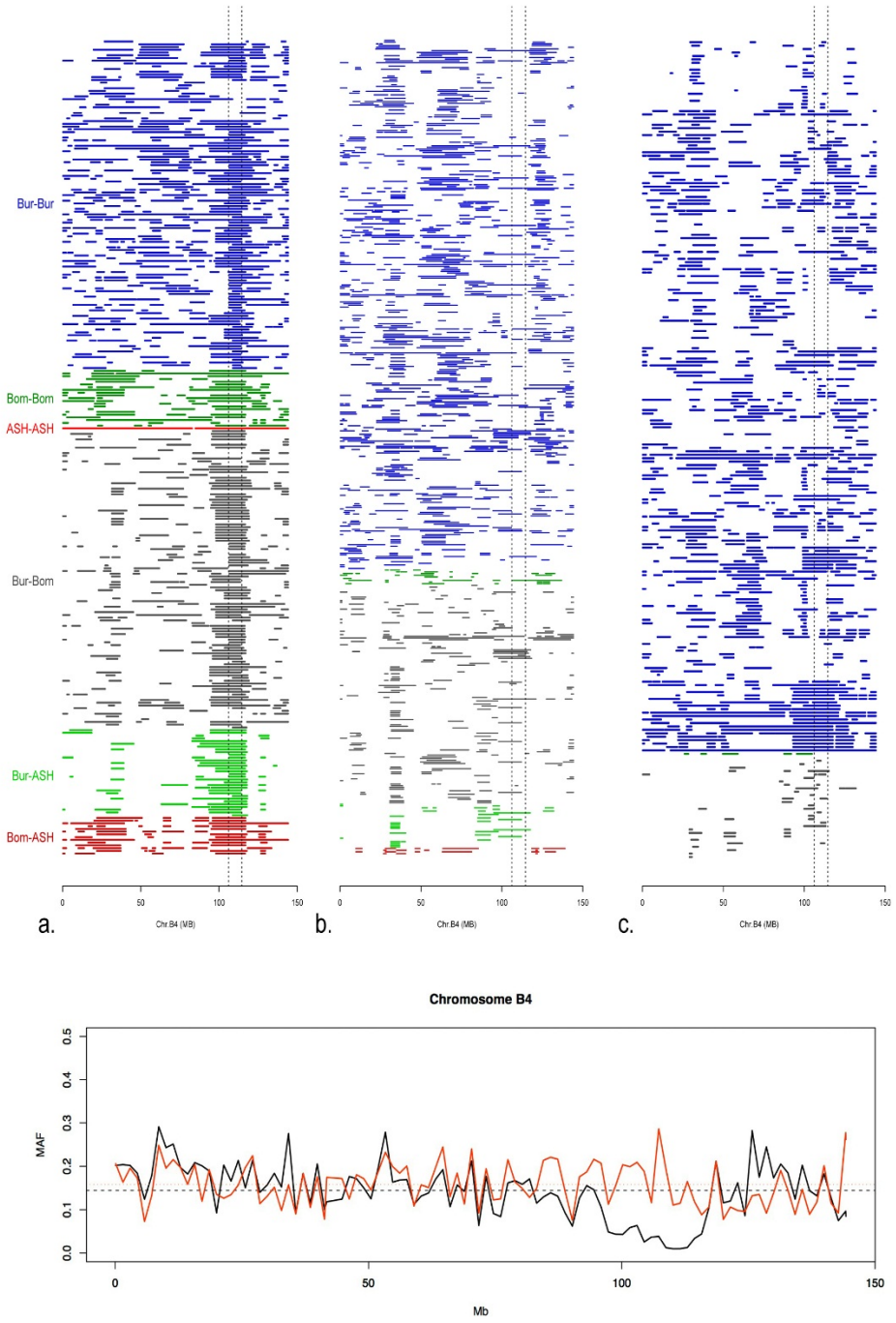
Supplementary Figure 2. Pedigree segregating for the Burmese Craniofacial Defect. Circles represent females, squares represent males, diamonds are unknown gender. Open symbols indicate phenotypically normal animals, filled symbols indicate affected cats, half-filled are obligate carriers. A small filled circle represents a “breeding node” for parental cats. Numbers under the symbols represent the laboratory sample number. Genotypes for the linked marker *FCA864* are represented below the identification numbers or names. The base pair size of the microsatellite marker was converted to a single number to distinguish the allele. No data is represented by dashes, “--”.



Supplementary Figure 3. Multidimensional scaling (MDS) analysis for population stratification of Burmese. Forty-six samples were plotted for principle components 1 and 2,a. represents the distribution of cases and controls, b. shows the distribution of the samples based on breed. c. The Burmese controls on the lower left of each plot were eliminated from the case-control analysis.

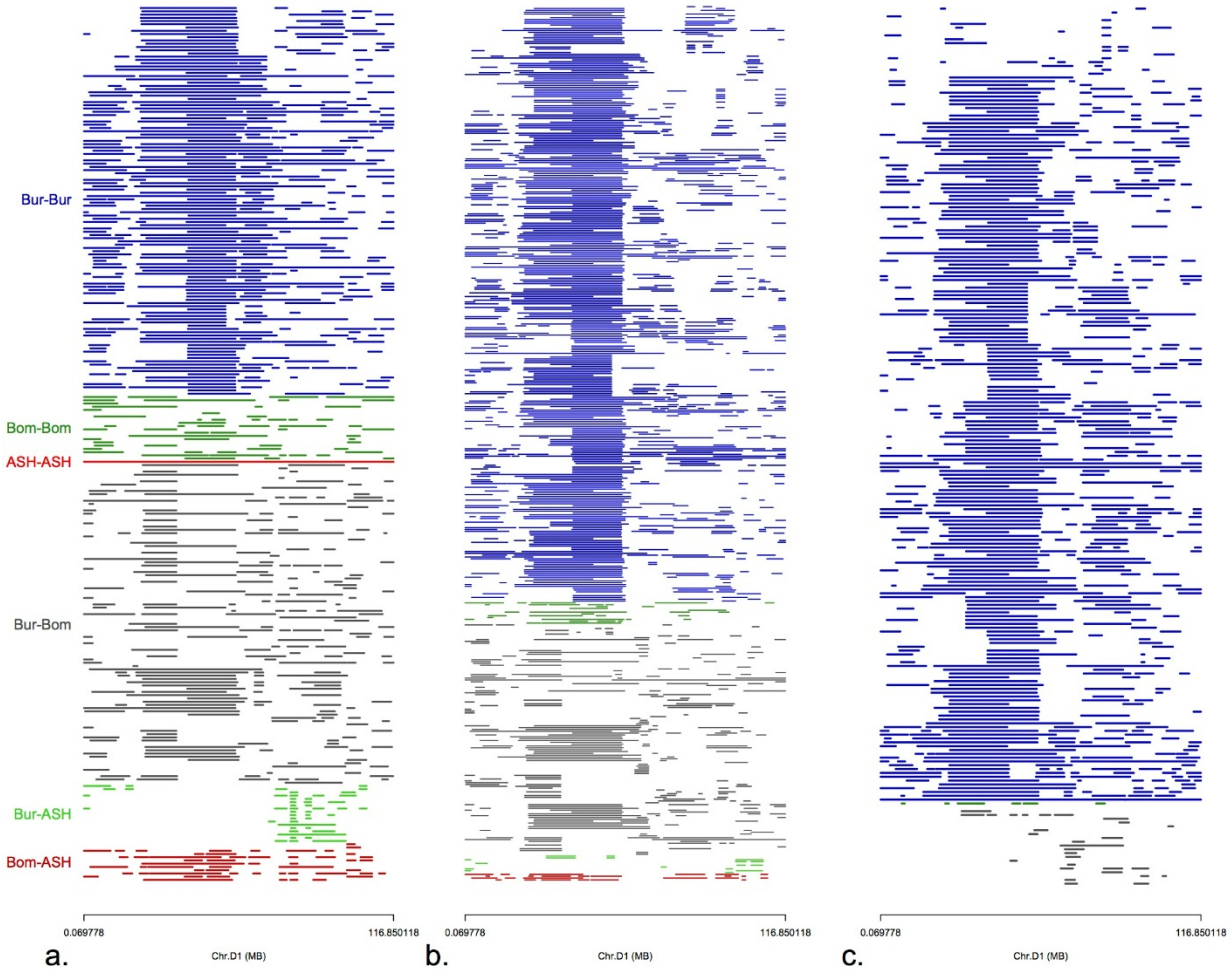


Supplementary Figure 4. Haplotype analysis of Burmese cases and controls for the craniofacial defect. Position 106,871,872 to position 111,795,156 of chromosome B4 in cases and controls. a. LD block identified by HAPLOVIEW across all the cases, spanning 4,923 Kb. b. Haplotypes sequence and frequencies across the 4,923 Kb regions. The main haplotype is squared in red and shows a frequency of 92% across cases. c. LD blocks identified by HAPLOVIEW in the correspondent region across all controls included in the study. d. Haplotypes sequence and frequencies for each identified LD block within the 4,923 Kb region in the control cats.

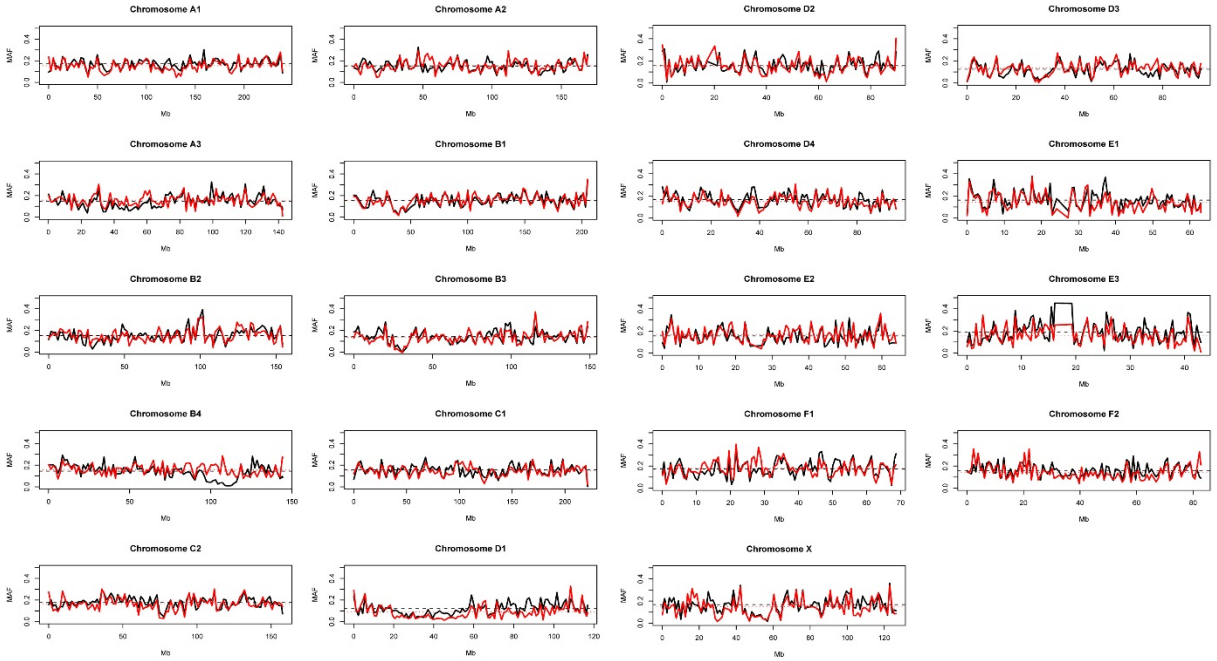


Supplementary Figure 5. **Identity by descent (IBD) and MAF analyses for chromosome B4**. The horizontal lines in the graph represent all the IBD regions (shared alleles) on chromosome B4 between all the cats included in the analysis. a. Each case is compared to all the other cases included in the analysis. Each group of comparison (breed to breed) is color-coded. Vertical black dashed lines represent a shared IDB region in common between almost all cases. b. Each case is compared to all the controls included in the study. c. Controls versus controls comparison of shared IBD. Cases versus controls and controls versus control comparisons do not show any shared IBD across all the specimens. (Bottom) Graphical representation of the MAF differences within the affected samples (black line) and the control

samples (red line) across all the *Felis catus* chromosomes. The black line represents the MAF within the cases and is compared with the MAF within the controls for each SNP. The red dashed line represent the MAF mean for the chromosome within the cases and the black dashed line the MAF mean within the controls. Several gaps are present in the current genome assembly, thus SNPs surrounding the gaps are connected with straight lines.



Supplementary Figure 6. Identity by descent (IBD) analysis for chromosome D1. The horizontal lines in the graph represents all the IBD regions (shared alleles) on chromosome D1 between all the cats included in the analysis. a. Each case is compared to all the other cases included in the analysis. Each group of comparison (breed to breed) is color-coded. b. Each case is compared to all the controls included in the study. c. Controls versus controls comparison of shared IBD. A common IBD is shared across the majority of East Asian breeds. The trait contained in the IBD region is a phenotypic trait responsible for the Burmese point coloration, fixed within the breed and confirmed by other analysis included in this study.



Supplementary Figure 7. Full chromosomal MAF comparison within cases and controls. Graphical representation of the MAF differences within the affected samples (black line) and the control samples (red line) across all the *Felis catus* chromosomes. The black line represents the MAF within the cases and is compared with the MAF within the controls for each SNP. The red dashed line represent the MAF mean for the chromosome within the cases and the black dashed line the MAF mean within the controls. Several gaps are present in the current genome assembly, thus SNPs surrounding the gaps are connected with straight lines.

```
                1                                                          60
Wild-type       MEFLSEKFALKSPPSKNSDFYMGAGGALEHVMETLDNESFYSKASAGKCVQAFGPLPRAE
Mutant          MEFLSEKFALKSPPSKNSDFYMGAGGALEHVMETLDNESFYSKASAGKCVQAFGPLPRAE

               61                                                         120
Wild-type       HHVRLERASPCQDSGVNYGITKGEGQPLHPELNRAMDNCNSLRMSPVKGMPEKGELDELG
Mutant          HHVRLERASPCQDSGVNYGITKGEGQPLHPELNRAMDNCNSLRMSPVKGMPEKGELDELG

               121                                                        180
Wild-type       DKCDSNVSSSKKRRHRTTFTSLQLEELEKVFQKTHYPDVYVREQLALRTELTEARVQVWF
Mutant          DKCDSNVSSSKKRRHRTTFTSLQLEELEKVFQKTHYPDVYVREQLEL----TEARVQVWF

               181                                                        240
Wild-type       QNRRAKWRKRERYGQIQQAKSHFAATYDISVLPRTDSYPQIQNNLWAGNASGGSVVTSGM
Mutant          QNRRAKWRKRERYGQIQQAKSHFAATYDISVLPRTDSYPQIQNNLWAGNASGGSVVTSGM

               241                                                        300
Wild-type       LPRDTSSCMTPYSHSPRTDSSYTGFSHHQNQFSHVPLNNFFTDSLLTGATNGHAFETKPE
Mutant          LPRDTSSCMTPYSHSPRTDSSYTGFSHHQNQFSHVPLNNFFTDSLLTGATNGHAFETKPE

               301                     327
Wild-type       FERRSSSIAVLRMKAKEHTANISWAM
Mutant          FERRSSSIAVLRMKAKEHTANISWAM

Homeobox domain
OAR domain
```

Supplementary Figure 8: **Protein alignment of the *Cart1* wildtype and mutated alleles**. The mutation, underlined in red, is responsible for the lack of 4 amino acids (*in silico* prediction) in the homeobox domain. In the human protein, the homeobox domain starts at position 132 and ends at position 191 of the amino acid chain. Underlined in blue is the *Cart1* OAR domain, which starts at position 306 and ends at position 319 of the Cart1 protein.