



Slingsby, L. S., Murrell, J. C., Taylor, P. M., & Sear, J. W. (2016). Effect of intramuscular methadone on pharmacokinetic data and thermal and mechanical nociceptive thresholds in the cat. *Journal of Feline Medicine and Surgery*, 18(11), 875-881. DOI: 10.1177/1098612X15605164

Peer reviewed version

Link to published version (if available):
[10.1177/1098612X15605164](https://doi.org/10.1177/1098612X15605164)

[Link to publication record in Explore Bristol Research](#)
PDF-document

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
<http://www.bristol.ac.uk/pure/about/ebr-terms.html>

1 **Effect of intramuscular methadone on pharmacokinetic data and thermal and mechanical nociceptive thresholds in the cat.**

2 LS Slingsby. University of Bristol, School of Veterinary Science, Bristol, UK

3 JW Sear. University of Oxford, John Radcliffe Hospital, Oxford, UK

4 PM Taylor. Taylor Monroe, Ely, UK

5 JC Murrell. University of Bristol, School of Veterinary Science, Bristol, UK

6

7 Corresponding author

8 Louisa Susanne Slingsby. BVSc, PhD, MRCVS.

9 Louisa.Slingsby@bristol.ac.uk

10 University of Bristol, School of Veterinary Science, Langford House, Langford, Bristol, BS40 5DU, UK

11

12 **Abstract**

13 **Objectives**

14 The study assessed simultaneous pharmacokinetics (PK) and thermal and mechanical antinociception after intramuscular methadone (0.6 mg/kg)

15 in 10 cats.

16 **Methods**

17 Thermal (TT) and mechanical (MT) threshold testing and blood collection were conducted at baseline and up to 24 hours after administration.

18 Methadone plasma concentrations were determined by liquid chromatography - tandem mass spectrometry (LC-MS) and PK parameters were

19 estimated by a non-compartmental method. TT and MT were analysed using ANOVA ($P<0.05$). Time of maximum plasma concentration (T_{max}),
20 time of onset of antinociception and time of reaching cut out threshold (TT 55 °C; MT 30N) were determined.

21 **Results**

22 TT and MT increased above baseline from 20 to 240 minutes and 5 to 40 minutes respectively after intramuscular administration ($P<0.005$).
23 Maximum delta T (measured as TT minus baseline threshold) (mean \pm 95%CI) was 7.9 (4.3-11.6) °C at 60 minutes and maximum delta F
24 (measured as MT minus baseline threshold) was 4.2 (1.6-6.7) Newtons at 45 minutes. Intramuscular methadone concentration-time data
25 decreased curvilinearly, and gave a clearance estimate of mean 9.1 ml/kg/min (range 5.2-15.7) with median T_{max} at 20 minutes (range 5-360).

26 **Conclusions**

27 Intramuscular data followed classical disposition and elimination in all cats. Plasma concentrations after intramuscular administration were
28 associated with antinociceptive effect including negative hysteresis.

29 **Relevance**

30 These data can be used for devising dosing schedules for methadone in clinical feline practice.

31 **Key words**

32 cat, antinociception, methadone, pharmacokinetics, thermal and mechanical threshold.

33 Authors

34 LS Slingsby

35 JW Sear

36 PM Taylor

37 JC Murrell

38

39

40

41

42

43

44

45

46

47

48 **Introduction**

49 Methadone is structurally unrelated to other opium-derived analgesics and exists as a racemic mixture. Each enantiomer has a separate mode of
50 action; the d-isomer noncompetitively antagonizes the NMDA receptor and inhibits norepinephrine reuptake; the l-isomer is a μ -opioid receptor
51 agonist ¹

52

53 There is only one report of the kinetics of methadone data in the cat ² and no data enabling the relationship between methadone
54 pharmacokinetics and dynamic endpoints to be assessed. While administration of drugs IV is the most reliable route to ensure full uptake this is
55 not always practical in a clinical setting, for example, when used as anaesthetic premedication where IV access may not be possible. The present
56 study evaluated the kinetics and dynamics of a single dose of intramuscular (IM) methadone (0.6 mg/kg) in the cat. Thermal and mechanical
57 thresholds were measured at the same time as blood sampling to 24 hours post-drug administration. The study was undertaken in order to
58 provide registration data for an indication for use of IM racemic methadone in the cat.

59

60

61 **Methods**

62 All studies were conducted after approval by the local institutional review board and according to UK Home Office licence; in-life data were
63 collected in September 2009.

64 *Pharmacokinetics of methadone*

65 Twelve adult neutered cats (3 male, 9 female; identified as M to X; mean weight 4.4, range 2.9-6.1kg; aged 34 months) received methadone (0.6
66 mg/kg) administered IM into the quadriceps femoris muscle. The day prior to their testing procedure, the cats were anaesthetised with
67 sevoflurane delivered in oxygen (induction by face mask followed by endotracheal intubation for maintenance). Over-the-needle catheters were
68 placed in both cephalic veins for ease of access in case non protocol intravenous administration of substances was required during the study and
69 as a back-up for/ as the main route for blood sampling. A modified Seldinger technique was used to place 5.5cm intravenous (IV) catheters in the
70 jugular vein in 6 cats (cats O, Q, S, U, V, X) and saphenous vein in 2 cats (cats M,W), the remaining 4 cats (N,P, R, T) were blood sampled
71 using the second cephalic catheter. Once the catheter was securely fixed the cats were returned to their holding cages for recovery from
72 anaesthesia. On the day of testing, blood samples were withdrawn from the (IV) (jugular, saphenous or cephalic) catheter before and at 5, 20,
73 40, 60 minutes, and then at 2, 3, 4, 6, 8, 12 and 24 hours after drug dosing. The volume of blood taken was adjusted for each individual cat, such
74 that over the 24 hour sampling period, the total blood volume taken was < 10% blood volume. An equal volume of 0.9% saline was injected
75 after each sample was withdrawn. Blood was taken into lithium heparin tubes, and centrifuged (2000g) for 10 minutes. Plasma was separated
76 and stored at -20°C for a maximum of 4 months.

77

78 Plasma methadone concentrations were measured at a commercial assay laboratory (Quotient Bioresearch Ltd, Cambs, UK) using liquid
79 chromatography - tandem mass spectrometry (LC-MS). In the concentration range 0.5 to 150 ng/ mL, the inter- and intra-assay coefficients of
80 variation were between 0.8 and 4.3%. Plasma methadone concentrations were linear over the range 0.5 to 150 ng/ mL. The limit of detection for
81 a 1 mL sample was 0.5 ng/ mL.

82

83 The maximum plasma concentration (C_{max}) and time of maximum concentration (T_{max}) were observed values. Pharmacokinetic parameters were
84 determined by a non-compartmental method³. Drug-concentration time profiles were subjected to nonlinear least squares regression of at least
85 three plasma concentrations to calculate the elimination half-life ($t_{1/2z}$). The area under the concentration-time curve (AUC) was calculated to the
86 final concentration time point of 24 hours (C_t) by the linear trapezoidal rule. The residual area to infinity was calculated as C_t/kz where z is the
87 terminal or elimination rate constant. From these values, other kinetic parameters were calculated: apparent clearance (Cl/F) and apparent
88 volume of distribution during the elimination phase (V_z/F) (which are given as Cl/F and V_z/F because we do not know the bioavailability (F)
89 after intramuscular dosing). Two sets of data for (Cl/F) and (V_z/F) are shown in table 1 - per kg body weight and weight independent. V_z (also
90 known as V_{area}) was used rather than V_{dss} (steady-state volume of distribution) as it is difficult to calculate the latter for the IM route.

91

92 *Pharmacodynamics of methadone*

93 Thermal and mechanical nociceptive thresholds threshold were used to determine the dynamic effects of methadone.

94 *Thermal threshold testing.* Thermal nociceptive thresholds were measured using a remote (infra-red) controlled system ⁴. The testing device
95 (WTT1, Topcat Metrology Ltd) is attached over the cat's back with a Velcro® and elastic strap. A small heating and temperature sensing probe
96 is held against the shaved thorax of the cat with a pressure bladder inside the strap. The starting skin temperature (SST) is recorded on the
97 integral display and then the heater activated so that temperature increases at 0.8°C per second. At the cat's response (skin twitch accompanied
98 by a behavioural response such as head turn, body shift) heating is stopped immediately and the temperature at the response is held on the
99 display and recorded as the thermal nociceptive threshold (TT). The probe rapidly returns to the starting skin temperature after the heating is
100 stopped. There is a safety cut out of 55°C.

101 *Mechanical threshold testing.* Mechanical nociceptive thresholds were measured using a silent, pneumatic system (MTT1, Topcat Metrology
102 Ltd) which employs a rolling diaphragm actuator ⁵ attached to a forelimb using a Velcro® and elastic bracelet. The actuator was placed on
103 shaved skin on the dorso-lateral aspect of the forelimb between carpus and elbow. Three hemispherical tipped pins (2.5 mm diameter) held in the
104 actuator were advanced against the skin at 2 N per second by increasing the pressure in the actuator manually, using an air filled syringe. A cage
105 mounted set of LED lights guides the rate of inflation (red = too fast, green = too slow, lights off = correct rate). Inflation is terminated as soon
106 as the cat's response is seen (leg shake, biting at the leg, picking up the leg) and the force (N) applied by the actuator at this point is held on the
107 display and recorded as the mechanical nociceptive threshold (MT). There is a safety cut out of 30N.

108 Four baseline thresholds were recorded at 15 minute intervals before treatment. The mean of these four readings was taken as the baseline TT or
109 MT. Thereafter TT and MT were measured at each sampling point to 24 hours post-drug administration. The increase in threshold at the various

110 time points following methadone administration was determined as delta T (calculated as the TT minus the pre-treatment TT for each cat) and
111 delta F (MT minus the pre-treatment MT).

112 The same investigator (LS) performed all the TT and MT tests. There was no blinding as this was a single dose study.

113 *Pupil dilation, sedation scores, monitoring of adverse events* Pupil dilation and sedation were recorded at each study time-point with 4 point
114 scores:

115 Pupil dilation: 1 = constricted, 2 = normal, 3 = partially dilated, 4 = fully dilated.

116 Sedation: 0 = none, 1 = mild (cat was relaxed, but could be roused and could walk with no ataxia), 2 = moderate (cat was in sternal or lateral
117 recumbency, but could be roused and had obvious signs of ataxia), 3 = no response to stimulation.

118 Any other behaviours or adverse events were recorded as and when they occurred.

119

120 *Statistical analyses*

121 Repeated measures ANOVA was performed on the mechanical and thermal dynamic data with post hoc Dunnett's test; p values less than 0.05
122 were deemed to be statistically significant.

123 Three indices of methadone's antinociceptive effect were determined:

124 . time to achieving peak plasma drug concentration (Tmax)

125 . time to onset of antinociceptive effect of the drug and the related methadone plasma concentration (Using a conventional method to define
126 biological references ranges ⁶, antinociceptive effect in our model was defined as the baseline threshold (mechanical or thermal) plus 2 SD, all
127 calculated from the grouped data.)

128 . time to first achieving the cut-off temperature or mechanical pressure.

129 These data are presented as median and range values unless otherwise stated.

130

131 AUCs for plasma drug concentration and dynamic endpoints delta T and delta F were calculated using the linear trapezoidal method, and were
132 compared against each other using regression analysis.

133

134 **Results**

135 Prior to dosing, most cats had mildly constricted or normal diameter pupils. Administration of IM methadone resulted in profound thermal
136 antinociception with no side-effects. It caused pupillary dilation which persisted for up to 12 hours, but no detectable sedation. Due to various
137 issues full data from 2 cats (O and X) could not be collected; for this reason although 12 cats started the study, only data from 10 cats are
138 presented.

139

140 *Kinetics of methadone in the cat*

141 Unless stated otherwise all data presented are mean (SD).

142

143 Following the 0.6 mg/ kg IM dose of methadone, plasma concentration T_{max} occurred at 5 to 120 minutes post dosing in 9 of the cats (median 20
144 minutes) as presented in Table 1. The exception was cat U which is discussed below.

145

146 The C_{max} in the 10 cats was 105 (28) ng/mL (range from 54 to 139 ng/mL). The sensitivity of the assay allowed all cats to have measurable
147 plasma methadone concentrations to the last sample point of 24 hours. The AUCs to 24 hours and to infinity were 52803 (14885) and 74464
148 (25142) ng/mL/min respectively.

149

150 Examination of the derived kinetic parameters showed a normal distribution. The estimates for systemic clearance (Cl) and volume of
151 distribution during the elimination phase (V_z) are shown as a ratio of parameter divided by bioavailability (F) as an estimate for the latter was not
152 measured or cited in the literature following IM dosing. The disposition data for the 10 cats are shown in Table 1 including the location of the
153 intravenous catheter for blood sampling. There were no apparent differences seen using one way ANOVA for the main PK values between
154 animals where blood sampling had been via the cephalic, jugular or saphenous veins. However the present study was not either aimed at or
155 powered to examine this variation.

156

157 *Dynamics of methadone in the cat*

158 The delta values for MT and TT (i.e. threshold minus baseline) are shown in Figures 1a and b. There was no clinically significant change in the
159 SST over the time period 0-24 hours (variation by less than 1 °C). The group pre-treatment TT was 43.4 (2.0) °C. After intramuscular methadone
160 dosing, the TT increased significantly from 20 minutes to 4 hours. The peak TT was 54.9 (0.3) °C. The maximum threshold (safety cut out) of
161 55°C was reached in nine cats at 22 (33) minutes after treatment and remained at this value for periods between 20 minutes and 2 hours in these
162 cats.

163 The maximum delta T values in the individual cats ranged between 8.7 and 14.8 °C and occurred between 33 and 150 minutes after dosing.

164 Where cats remained at the same delta T for more than one time point, then the closest study time point to the average was taken. The plasma
165 methadone concentration at the time of maximum delta T ranged between 46.7 and 104.0 ng/mL. Thermal threshold T_{\max} values are difficult to
166 estimate in the cats that spent a period of time with thresholds that exceeded the cut out temperature. Individual T_{\max} values (where there is a
167 range, this indicates the time spent at cut out) were: M 40 minutes; N 40; P 20-120; Q 40-60; R 120; S 20-180; T 40-240; U 120; V 60; W 5-60.

168 Mechanical thresholds varied significantly over time with MT significantly raised at 5, 20 and 40 minutes compared to baseline. The pre-
169 treatment MT was 8.4 (1.6) N. The largest delta F ranged in the individual cats between 3.2 and 13.2N, and occurred between 5 and 240 minutes
170 after dosing. The plasma methadone concentration at the time of maximum delta F ranged between 23.4 and 139.0 ng/mL.

171 The AUC (of delta T over time) for the period 0-1440 minutes was 3772.8 (1436.9) °C.min for the intramuscular dosing; while the

172 corresponding AUC (of delta F over time) for the same period was 1769.3 (1375.6) Newtons.min. Correlation analysis between the AUC (of

173 plasma methadone concentration over time) and AUC (of antinociceptive effects over time) to 1440 minutes (24 hours) post-dosing indicated
174 significance for the delta T vs. concentration measurement ($r = 0.8573$) but not for delta F vs. concentration ($r = -0.4156$). There was no
175 significant correlation between the AUCs for the two antinociceptive measures ($r = -0.2086$).

176 *Kinetic-dynamic relationships*

177 When the measures delta T, delta F and plasma methadone concentration were plotted together against time (Figure 2), there was little difference
178 between kinetic and dynamic profiles. There did not appear to be any significant separation between drug concentration and effect - although the
179 T_{\max} (median) was 20 minutes for concentration; 40 min for MT and for TT.

180 As with most drug responses the plasma concentration-time profile and the effect-time relationship were not in phase and there was an
181 anticlockwise hysteresis loop (Figure 3).

182 Both stimulus modalities also showed a dip in response between 360 and 720 minutes (Figure 2).

183 The plasma drug concentrations associated with the onset of antinociception baseline plus 2 SD ranged between 39.2 and 124 ng/ml for
184 temperature; and 23.4 and 139 ng/ml for mechanical pressure. The comparable values for the offset of antinociception were 13.9-105 ng/ml and
185 15.1-102 ng/ml respectively.

186

187 **Discussion**

188 Methadone is a synthetic full agonist opioid, but studies have revealed that it also acts on *N*-methyl-D-aspartate receptors ⁷ and it has been used
189 as an alternative to morphine and hydromorphone in human patients with severe pain. Clinically, in cats, methadone is used for analgesia often
190 administered as part of anaesthetic premedication where it can assist in the production of sedation in combination with tranquilizers or sedatives.
191 Most comparable clinical studies have used feline ovariohysterectomy as a surgical model with methadone administered with the premedication
192 and pain/ analgesia scored by a variety of methods including wound palpation and behavioural and physiological observations. An early study
193 examined use of methadone 0.5 mg/kg IM where analgesia was reported for 1.5-6 hours from administration of methadone⁸; a later study with
194 0.6 mg/kg IM at premedication reported that 18/19 cats has adequate analgesia for the entire study period of 4 hours after surgery and only a
195 single cat required rescue analgesia after 90 minutes⁹; a third study using 0.5 mg/kg IM reported good analgesia for the study period of 6 hours
196 in 6/8 cats with 1/8 requiring rescue analgesic at 4 hours and 1/8 at 5 hours ¹⁰. Pre-anaesthetic sedation with acepromazine - methadone
197 combination was reported to be poor but similar to that seen with acepromazine - butorphanol combination ¹⁰.

198 .
199
200 The current study used threshold testing tools to assess antinociception rather than a surgical stimulus to assess pain/ analgesia. The
201 physiological processes behind a thermal stimulus applied to the thoracic skin or a mechanical stimulus applied to a forelimb are not the same as
202 the pain caused by surgery so their clinical relevance could be questioned. However nociceptive threshold testing tools are widely used and
203 accepted for pain/ analgesia research in laboratory species and in human volunteers. Our study has demonstrated clear differences between the

204 effects of methadone on the two stimulus modalities of heat and pressure; significant effects on mechanical thresholds were much shorter acting
205 than those on thermal thresholds. This is not unknown in analgesia research and may be due to a number of factors including device design and /
206 or different physiological pathways for the two stimuli. Since the thermal threshold testing tool has already demonstrated similar thermal
207 antinociceptive profiles compared to the well documented clinical analgesic profile of commonly used opioid analgesics such as buprenorphine
208 we would suggest that, for whatever reason, the thermal threshold data are more likely to be similar to the clinical profile of methadone than the
209 mechanical thresholds.

210

211 Comparison of the time-courses of kinetic and dynamic effects of the opioid show a lag time between increasing plasma concentrations and
212 increase in observable effects. This is in keeping with the site of action of methadone being at opioid receptors in tissues rather than in plasma
213 (Figure 3); with the lag being the time taken for methadone to move across the blood brain barrier to opioid receptors in the central nervous
214 system. Many other drugs demonstrate a similar 'out of phase' concentration-time and the effect-time relationship.

215

216 Much less individual variation relating to drug disposition was seen in this study compared to previous studies of other opioids (buprenorphine,
217 morphine and pethidine) in cats^{11, 12}. Maximum plasma concentration (C_{max}) and time of maximum plasma concentration (T_{max}) were very
218 consistent for all cats except for animal U. In this cat there was a late peak in concentration at 360 minutes which could indicate slow absorption
219 or perhaps injection into a poorly vascularised area of the body. The quadriceps muscle was used as the site of injection and should be well

220 vascularised but this cat was noticeably fatter than the other cats and hence injection of drug into fat tissue might be associated with a reduced
221 rate of absorption.

222 Median plasma T_{max} was at 20 minutes. The absorption was less rapid than that seen in a presently unpublished parallel study in dogs (range 5-
223 15 minutes) although prolonged absorption time was seen in dogs when administered a higher dose of 0.5mg kg⁻¹ compared to 0.3 mg kg⁻¹.

224 Other studies with opioids in cats have demonstrated the plasma T_{max} after IM injection were 15, 3, and 10 minutes for morphine, buprenorphine
225 and pethidine respectively ¹¹ and 21 minutes for butorphanol ¹³.

226 Racemic methadone was administered at 0.6 mg/kg IM in the current study and as this was the only route of administration and the cats were not
227 also administered an IV dose it was not possible to calculate bioavailability (F) which is why our data has been corrected for F. At the time of
228 analysis we were not aware of directly measured data for the bioavailability of methadone after IM administration in the cat. Recently published
229 data ¹⁴ gives a value for F of 44.2% after buccal dosing; with the kinetics for a 0.3 mg/kg IV dose being quoted as clearance 7.2 mL/kg/min and
230 V_z 2.4/kg. Using the area under the curve in their study and applying it to our data, the estimated bioavailability in our study would be between
231 80 and 85%. From a study in cats where 0.3 mg/kg methadone was administered IV ², using their presented data we have calculated the
232 clearance as 4.3 mL/kg/min and $t_{1/2}$ as 278.4 minutes (4.6 hours). The PK values from these 2 studies are comparable with the values we have
233 reported here ($Cl/F = 9.1$ (3.3) mL/ kg/min and $V_z/F = 7.8$ (2.7) L/ kg; $t_{1/2} = 627.9$ minutes (212.2)).

234

235 Although Hedges and colleagues¹⁵ found differences in the disposition after buccal administration of buprenorphine in cats depending on the site
236 of blood sampling, we have not found any significant differences in the systemic clearance of methadone when comparing data from the three
237 sites used for blood sampling in this study. However the sample size of each group of cats was small, and to examine it further a properly
238 powered study would be needed. There are also differences in the route of drug dosing (intramuscular vs buccal) between the two studies.

239

240 The plasma methadone pharmacokinetics reported here for the cat resembles those seen in dogs. After IV administration of 0.45 mg/kg racemic
241 methadone base to greyhounds¹⁶, systemic clearance was 56.0 (9.4) mL/kg/min and V_z was 7.8 (1.9) L/ kg; after administration of 1 mg/ kg IV
242 to beagles, total body clearance was 24.1 (9.8) mL/kg/min and V_z was 3.7 (1.1) L/ kg¹⁷.

243

244 When reviewing the literature in humans it becomes clear that while there are many published studies, most publications relate to aspects of its
245 use is as a heroin replacement substance rather than a first line analgesic and therefore the number of pharmacokinetic studies of methadone in
246 healthy, non-opiate users are limited. There is also a difficulty in making direct comparisons as most animal studies report values adjusted for
247 bodyweight whereas those in humans do not. Clearance (Cl) adjusted for bioavailability (Cl/F) after oral administration in opiate-naive humans
248 was 115 mL/min (data reported as 6.9 L/ hour) and half-life estimates were 33-46 hours¹⁸. In these individuals, weight as a covariable had no
249 significant relationship to Cl/F and using the median bodyweight, the calculated Cl/F would be about 2 mL/kg/min. Another study¹⁹ reported
250 mean clearance after a 5mg IV dose as 8.3 L/ hour (138 mL/ min) and after a 10mg oral dose Cl/F = 9.8 L/ hour (163 mL/ min); adjustment for

251 mean volunteer weight (84kg) gives a Cl (IV) of 1.6 mL/kg/min and Cl/F (oral) 1.9 mL/kg/min. The estimate for oral bioavailability in this study
252 was calculated as 86% making Cl (oral) = 2.2 mL/kg/min, and the half-life was reported as 32 and 31 hours in the IV and oral groups
253 respectively. Terminal half lives in man, however, are substantially greater than in the cat, for example after effectively a 0.05 mg/ kg dose in
254 humans, the $t_{1/2}$ was 32 hours whereas after 0.6 mg/kg in the cat the $t_{1/2}$ was 10 hours, this might be due to the large V_d in humans.

255

256 A number of studies have investigated plasma concentration with respect to the antinociceptive or analgesic action of methadone in human
257 subjects; they appear to be similar for acute pain stimuli such as antinociceptive tests and surgery although much higher concentrations were
258 required for patients with chronic pain^{20,21}. Postoperative minimum analgesic concentrations in humans were 30-33 ng/ml²² whereas the EC₅₀
259 (half maximal effective concentration - the concentration of a drug which induces a response halfway between the baseline and maximum) in 8
260 chronic pain patients (5 with cancer) was 290 ng/ml²⁰. Our study findings of plasma methadone concentrations for onset (39.2 to 124 ng/ml) and
261 offset 13.9-105 ng/ml) of thermal antinociception and onset (23.4 to 139 ng/ml) and offset (15.1-102 ng/ml) of mechanical antinociception are
262 similar to those required in humans for acute pain stimuli.

263

264 Pharmacokinetics can be defined as what the body does with an administered drug and pharmacodynamics as what the drug does to the body.

265 The aim when investigating potentially useful clinical analgesics is to determine the optimum plasma concentration associated with analgesia,

266 PK and PD data may then be used to determine optimum route of administration, dose and dosing interval in order to maintain this analgesic

267 plasma concentration. In this study, we have shown that the kinetics of a single dose of IM methadone in the cat are similar to that reported in
268 the dog. Drug uptake is rapid in the cat, but systemic clearance for both species is greater than that reported for man. We have also determined
269 analgesic plasma concentrations of methadone. These data may be used in combination to suggest that analgesia from an intramuscular dose of
270 methadone at 0.6 mg/kg would be expected to provide four hours of analgesia and that the target plasma concentration for onset of analgesia lies
271 between 40 and 124 ng/ml.

272

273 **Acknowledgements**

274 This study was funded by Eurovet Animal Health B.V

- 275 1. Davis AM and Inturrisi CE. d-Methadone Blocks Morphine Tolerance and N-Methyl-d-Aspartate-Induced Hyperalgesia. *J Pharmacol Exp Ther.* 1999;
276 289: 1048-53.
- 277 2. Ferreira TH, Rezende ML, Mama KR, Hudachek SF and Aguiar AJA. Plasma concentrations and behavioral, antinociceptive, and physiologic effects of
278 methadone after intravenous and oral transmucosal administration in cats. *Am J Vet Res.* 2011; 72: 764-71.
- 279 3. Benet LZ and Galeazzi RL. Noncompartmental determination of the steady-state volume of distribution. *J Pharm Sci.* 1979; 68: 1071-4.
- 280 4. Dixon M, Taylor P, Slingsby L and Waterman-Pearson A. Development of a remote controlled thermal threshold testing system for the cat. *Vet*
281 *Anaesth Analg.* 2007; 34: 70.
- 282 5. Dixon MJ, Taylor PM, Slingsby L, Hoffmann MV, Kastner SBR and Murrell J. A small, silent, low friction, linear actuator for mechanical nociceptive
283 testing in veterinary research. *Lab Anim.* 2010; 44: 247-53.
- 284 6. Kaneko JJ, Harvey JW and Bruss M. Clinical Biochemistry of Domestic Animals. Academic Press, 2008.
- 285 7. Inturrisi CE. Pharmacology of methadone and its isomers. *Minerva Anesthesiol.* 2005; 71: 435-7.
- 286 8. Dobromylskyj P. Assessment of methadone as an anaesthetic premedicant in cats. *J Small Anim Pract.* 1993; 34: 604-8.
- 287 9. Bley CR, Neiger-Aeschbacher G, Busato A and Schatzmann U. Comparison of perioperative racemic methadone, levo-methadone and
288 dextromoramide in cats using indicators of post-operative pain. *Vet Anaesth Analg.* 2004; 31: 175-82.

- 289 10. Bortolami E, Murrell JC and Slingsby LS. Methadone in combination with acepromazine as premedication prior to neutering in the cat. *Vet Anaesth*
290 *Analg.* 2013; 40: 181-93.
- 291 11. Taylor P, Robertson S, Dixon M, et al. Morphine, pethidine and buprenorphine disposition in the cat. *J Vet Pharmacol Ther.* 2001; 24: 391-8.
- 292 12. Robertson SA, Lascelles BDX, Taylor PM and Sear JW. PK-PD modeling of buprenorphine in cats: intravenous and oral transmucosal administration. *J*
293 *Vet Pharmacol Ther.* 2005; 28: 453-60.
- 294 13. Wells SM, Glerum LE and Papich MG. Pharmacokinetics of butorphanol in cats after intramuscular and buccal transmucosal administration. *Am J Vet*
295 *Res.* 2008; 69: 1548-54.
- 296 14. Pypendop BH, Ilkiw JE and Shilo-Benjamini Y. Bioavailability of morphine, methadone, hydromorphone, and oxymorphone following buccal
297 administration in cats. *J Vet Pharmacol Ther.* 2014; 37: 295-300.
- 298 15. Hedges AR, Pypendop BH, Shilo-Benjamini Y, Stanley SD and Ilkiw JE. Pharmacokinetics of buprenorphine following intravenous and buccal
299 administration in cats, and effects on thermal threshold. *J Vet Pharmacol Ther.* 2014; 37: 252-9.
- 300 16. Kukanich B and Borum SL. The disposition and behavioral effects of methadone in Greyhounds. *Vet Anaesth Analg.* 2008; 35: 242-8.
- 301 17. Kukanich B, Lascelles BDX, Aman AM, Mealey KL and Papich MG. The effects of inhibiting cytochrome P450 3A, p-glycoprotein, and gastric acid
302 secretion on the oral bioavailability of methadone in dogs. *J Vet Pharmacol Ther.* 2005; 28: 461-6.
- 303 18. Wolff K, Rostami-Hodjegan A, Shires S, et al. The pharmacokinetics of methadone in healthy subjects and opiate users. *Br J Clin Pharmacol.* 1997;
304 44: 325-34.
- 305 19. Dale O, Sheffels P and Kharasch ED. Bioavailabilities of rectal and oral methadone in healthy subjects. *Br J Clin Pharmacol.* 2004; 58: 156-62.
- 306 20. Inturrisi CE, Colburn WA, Kaiko RF, Houde RW and Foley KM. Pharmacokinetics and pharmacodynamics of methadone in patients with chronic pain.
307 *Clinical Pharmacology and Therapeutics.* 1987; 41: 392-401.
- 308 21. Inturrisi CE, Portenoy RK, Max MB, Colburn WA and Foley KM. Pharmacokinetic-pharmacodynamic relationships of methadone infusions in patients
309 with cancer pain. *Clin Pharmacol Ther.* 1990; 47: 565-77.
- 310 22. Gourlay GK, Wilson PR and Glynn CJ. Pharmacodynamics and Pharmacokinetics of Methadone during the Perioperative Period. *Anesthesiology.*
311 1982; 57: 458-67.

312

313

314 Table 1: Pharmacokinetic parameters for 10 cats after IM administration of 0.6mg/kg methadone. Clearance and volume of distribution are
 315 shown as both body weight dependent and independent. Sampling route, Cep = cephalic; Jug = jugular; Sap = saphenous

316

cat	sampling route	Cmax ng/ml	Tmax min	Cl/F ml/min	Cl/F ml/kg/min	t _{1/2 z} min	Vz/F litres	Vz/F l/kg	AUC24 ng/mL/min
N	Cep	132	20	34	7.9	350	17	4	62446
P	Cep	92	20	54.2	10.1	412	32	6	53642
R	Cep	122	5	26.4	6.3	824	31	7	68647
T	Cep	125	5	30.2	7	416	18	4	41746
Q	Jug	100	120	27.9	5.2	906	36	7	74022
S	Jug	71	40	50.2	12.5	549	40	10	38990
U	Jug	54	360	40.3	8.6	894	52	11	44206
V	Jug	139	40	25.8	6.1	801	30	7	69644
M	Sap	92	20	45.4	15.7	513	34	12	32319
W	Sap	121	20	43.4	11.1	613	38	10	42370
	mean	105	65	37.8	9.1	628	33	8	52803
	SD	28	109	10.3	3.3	212	10	3	14885
	median	111	20	37.1	8.2	581	33	7	48924

317

318

319

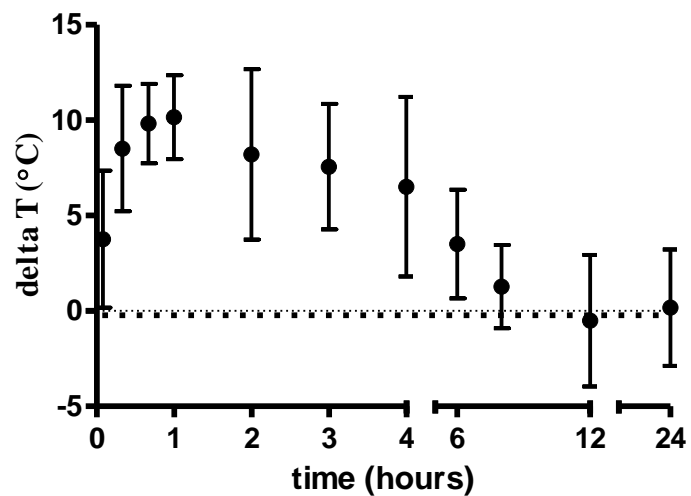
320

321 Figure 1

322 Delta T (Fig 1a) and delta F (Fig 1b) threshold values for 10 cats after administration of methadone 0.6mg/kg (mean and standard deviation).
323 Significant differences from baseline thermal threshold from 20 minutes up to and including 4 hours (1a) and from baseline mechanical
324 threshold at 5, 20 and 40 minutes (1b)

325

326 Fig 1a



328

329

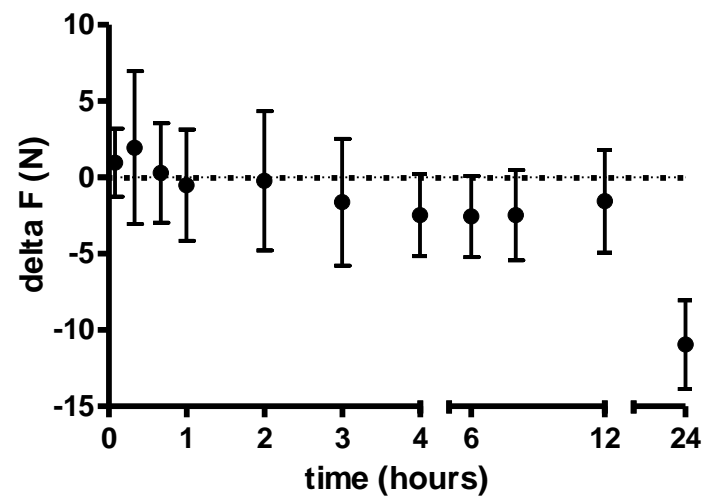
330

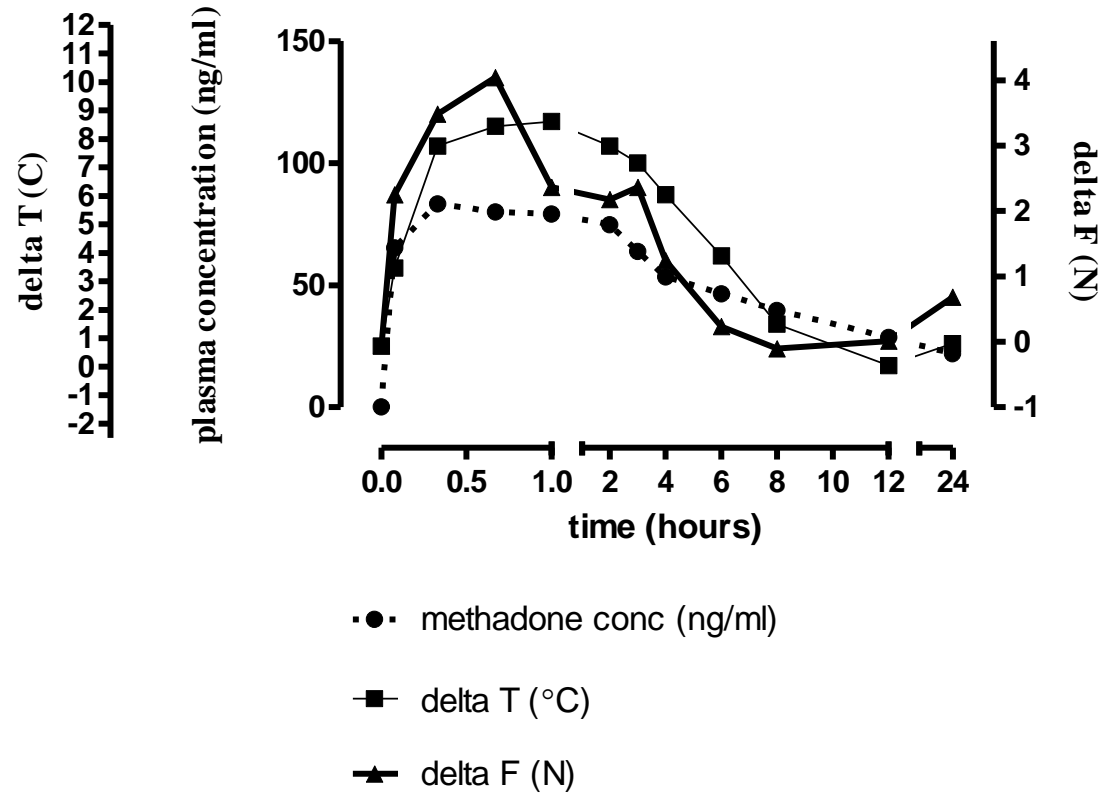
331

332

333

Fig 1b





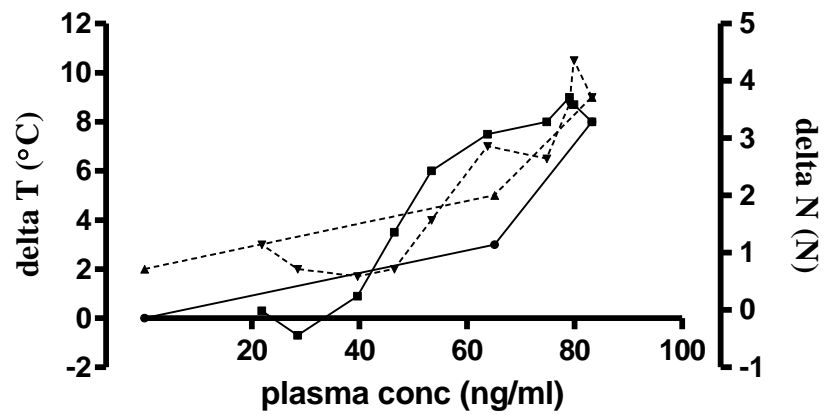
335

336

337 Figure 2

338 Methadone plasma concentration and changes in mechanical and thermal threshold (delta T and F)

339
340
341
342
343
344



- delta T (°C) up
- delta T (°C) down
- - -▲- - - delta F (N) up
- - -▼- - - delta F (N) down

345
346

347 Figure 3 plasma concentration-time profile and the effect-time relationship showing anticlockwise hysteresis loop (effect data derived when
348 concentrations were increasing are called 'up' and those when concentrations were decreasing were called 'down')

349

