

# The Manganese Transporter MntH Is a Critical Virulence Determinant for *Brucella abortus* 2308 in Experimentally Infected Mice<sup>∇</sup>

Eric S. Anderson,<sup>1</sup>† James T. Paulley,<sup>1</sup>†‡ Jennifer M. Gaines,<sup>1</sup> Michelle W. Valderas,<sup>2</sup>§  
Daniel W. Martin,<sup>1</sup> Evan Menscher,<sup>1</sup> Timothy D. Brown,<sup>1</sup>  
Colin S. Burns,<sup>3</sup> and R. Martin Roop II<sup>1\*</sup>

Department of Microbiology and Immunology, East Carolina University School of Medicine, Greenville, North Carolina 27834<sup>1</sup>;  
Department of Veterinary Pathobiology, Oklahoma State University Center for Veterinary Health Sciences, Stillwater,  
Oklahoma 74078<sup>2</sup>; and Department of Chemistry, East Carolina University, Greenville, North Carolina 27858<sup>3</sup>

Received 20 April 2009/Returned for modification 3 May 2009/Accepted 20 May 2009

The gene designated BAB1\_1460 in the *Brucella abortus* 2308 genome sequence is predicted to encode the manganese transporter MntH. Phenotypic analysis of an isogenic *mntH* mutant indicates that MntH is the sole high-affinity manganese transporter in this bacterium but that MntH does not play a detectable role in the transport of Fe<sup>2+</sup>, Zn<sup>2+</sup>, Co<sup>2+</sup>, or Ni<sup>2+</sup>. Consistent with the apparent selectivity of the corresponding gene product, the expression of the *mntH* gene in *B. abortus* 2308 is repressed by Mn<sup>2+</sup>, but not Fe<sup>2+</sup>, and this Mn-responsive expression is mediated by a Mur-like repressor. The *B. abortus mntH* mutant MWV15 exhibits increased susceptibility to oxidative killing in vitro compared to strain 2308, and a comparative analysis of the superoxide dismutase activities present in these two strains indicates that the parental strain requires MntH in order to make wild-type levels of its manganese superoxide dismutase SodA. The *B. abortus mntH* mutant also exhibits extreme attenuation in both cultured murine macrophages and experimentally infected C57BL/6 mice. These experimental findings indicate that Mn<sup>2+</sup> transport mediated by MntH plays an important role in the physiology of *B. abortus* 2308, particularly during its intracellular survival and replication in the host.

*Brucella abortus* is a gram-negative bacterium that is responsible for the zoonotic disease brucellosis. Brucellosis causes spontaneous abortion and sterility in ruminants (27) and a debilitating febrile illness in humans known as undulant fever (17). The ability of brucellae to cause disease is directly related to their capacity to establish and maintain intracellular infection in host macrophages (63). Within the phagosomal compartment in these host cells, brucellae must cope with oxidative stress, low pH, and nutrient deprivation. The availability of metal ions is restricted within this environment due in part to the activity of the host natural resistance-associated macrophage protein (NRAMP-1), which transports divalent cations out of the phagosome (40). Mn<sup>2+</sup> serves as an important cofactor for a variety of bacterial enzymes, including those involved in carbon metabolism, induction of the stringent response, and detoxification of reactive oxygen species (ROS) (55). Consequently, the inability of brucellae to acquire sufficient levels of this divalent cation may compromise their ability to successfully adapt to the environmental conditions encountered during residence in their intracellular niche.

Manganese uptake by bacteria is typically accomplished

through the activity of either ABC-type transporters such as the SitABC complex (4, 42, 59, 65) or H<sup>+</sup>-dependent manganese transporters such as MntH (37, 41, 52, 60). Many bacteria possess both types of Mn<sup>2+</sup> transporters (55), but a survey of the publicly available *Brucella* genome sequences (14, 20, 36, 57) suggests that these bacteria do not produce a SitABC-type transporter and rely solely on an MntH homolog for the high-affinity transport of Mn<sup>2+</sup>. *Escherichia coli* MntH was originally described as being able to transport both Mn<sup>2+</sup> and Fe<sup>2+</sup> (52), but subsequent studies indicated that this and other bacterial MntH proteins are highly selective Mn<sup>2+</sup> transporters that play a minor, if any, role in Fe<sup>2+</sup> transport under physiologically relevant conditions (41). To examine the role of *Brucella* MntH in Mn<sup>2+</sup> transport and virulence, the gene annotated as BAB1\_1460 in the *B. abortus* 2308 genome sequence was disrupted in this strain by gene replacement and the phenotype of the resulting mutant (MWV15) was examined. The results of these studies indicate that MntH plays a critical role in Mn<sup>2+</sup> transport in *B. abortus* 2308 and that the presence of this manganese transporter is essential for the wild-type resistance of this strain to oxidative killing in vitro and its virulence in the mouse model.

## MATERIALS AND METHODS

**Bacterial strains and culture conditions.** *Brucella abortus* 2308 and derivatives of this strain were cultivated on Schaedler agar supplemented with 5% defibrinated bovine blood (SBA) at 37°C with 5% CO<sub>2</sub> or in brucella broth at 37°C with shaking unless otherwise noted. *Escherichia coli* strain DH5α was used as the host strain for recombinant DNA procedures, and this strain was cultivated on tryptic soy agar at 37°C or in LB broth at 37°C with shaking. Growth media were supplemented with ampicillin (25 μg/ml for *B. abortus* and 100 μg/ml for *E. coli* DH5α), chloramphenicol (5 μg/ml for *B. abortus* and 30 μg/ml for *E. coli* DH5α), and/or kanamycin (45 μg/ml) as necessary. *Brucella* stock cultures were

\* Corresponding author. Mailing address: Department of Microbiology and Immunology, East Carolina University School of Medicine, 600 Moye Boulevard, Greenville, NC 27834. Phone: (252) 744-1357. Fax: (252) 744-3535. E-mail: roopr@ecu.edu.

† Present address: Department of Microbiology, Immunology, and Molecular Genetics, University of Kentucky College of Medicine, Lexington, KY 40506.

§ Present address: Lovelace Respiratory Research Institute, Albuquerque, NM 87108.

‡ Eric S. Anderson and James T. Paulley contributed equally to this work.

<sup>∇</sup> Published ahead of print on 1 June 2009.

maintained in brucella broth supplemented with 25% glycerol, and *E. coli* stock cultures were maintained in LB supplemented with 25% glycerol at  $-80^{\circ}\text{C}$ .

**Construction of an *mntH-lacZ* transcriptional fusion.** Oligonucleotide primers (forward, 5'-GAGCGGGCCATCCTTCTGAA-3'; reverse, 5'-GTTGCCGGGA TCCATATAACC-3') and PCR were used to amplify a 1,031-bp fragment of genomic DNA from *B. abortus* 2308 containing portions of the *mraW* (BAB1\_1458) and *mntH* (BAB1\_1460) genes and intervening regions. The resulting PCR fragment was digested with HindIII and KpnI, and a 239-bp HindIII/KpnI fragment containing upstream sequences and extending 8 bp into the *mntH* coding region was directionally cloned into the *lacZ* transcriptional fusion vector pMR15 (32). The authenticity of the *mntH-lacZ* fusion in the resulting plasmid, pEAM1, was verified by restriction mapping and nucleotide sequence analysis. Transcriptional activity of the  $\beta$ -galactosidase reporter fusion was determined using the methods described by Miller (53).

**Construction of the *B. abortus mur* mutant Fur2.** Plasmid pDS1 contains an 867-bp fragment of genomic DNA from *B. abortus* 2308 containing the *mur* homolog designated BAB1\_1668 cloned into pUC9 (5). An inverse PCR strategy (23) was used to generate a linear derivative of pDS1 that lacks a 338-bp region internal to the 426-bp coding region of *Brucella mur* by using the primers  $\Delta fur1$  (5'-GCCTCAGCTCTGCTCATAAT-3') and  $\Delta fur2$  (CATCACCGGCTT GAACCTTA-3'). The chloramphenicol resistance gene from pBC-SK (NEB) was ligated to the linear  $\Delta mur$  derivative of pDS1, and the resulting plasmid was used to introduce a *mur* mutation into the genome of *B. abortus* 2308 by gene replacement using the methods described by Elzer et al. (25). The genotype of the *B. abortus mur* mutant (Fur2) constructed in this fashion was confirmed by Southern blot analysis with *cat-* and *mur*-specific probes (5).

**Construction and genetic complementation of the *B. abortus mntH* mutant MWV15.** A 1,566-bp region encompassing the putative *mntH* gene (BAB1\_1460) (forward primer, 5'-TTCCCTATTCCTTAACAT-3'; reverse primer, 5'-GATCGGCGTTCTATTCTTT-3') was amplified from *B. abortus* 2308 genomic DNA by PCR using *Pfx* polymerase (Invitrogen) and cloned into pGEM-T Easy (Promega). The resulting plasmid was digested with BamHI and HindIII to remove a 696-bp region internal to the predicted *mntH* coding region, treated with the Klenow fragment of DNA polymerase I, and ligated with a 987-bp SmaI/HincII fragment containing the chloramphenicol acetyltransferase (*cat*) gene from pBlueCM2 (61). This plasmid was then used to construct an *mntH* mutant from *B. abortus* 2308 via gene replacement using previously described procedures (25). The genotype of the *B. abortus mntH* mutant (designated MWV15) was confirmed by PCR analysis of genomic DNA from this strain with *mntH*-, *cat*-, and pGEM-T Easy-specific primer sets.

For genetic complementation of the *mntH* mutation in MWV15, a 1,595-bp DNA fragment containing the *mntH* coding region was amplified by PCR (forward primer, 5'-AACATACTCCCTACTCCCTTATTC-3'; reverse primer, 5'-GTATCAGATCGGCGTTCTATTCTT-3') from *B. abortus* 2308 genomic DNA by using *Pfu* polymerase (Stratagene) and cloned into pGEM-T Easy after A-tagging of the PCR fragments according to the instructions provided by the manufacturer. The cloned DNA fragment was then excised from this plasmid as an ApaI/SacI fragment and cloned into the corresponding restriction sites in pMR10 (30). This *mntH*-containing plasmid, pEA31, was introduced into *B. abortus* MWV15 by electroporation, and the resulting strain was given the designation MWV15.C.

**Capacity of  $\text{MnCl}_2$  to stimulate the growth of *B. abortus* strains in the presence of EDDA.** *B. abortus* strains were grown on SBA for 48 h at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$  and harvested into phosphate-buffered saline (PBS). The optical density of the bacterial cell suspensions at 600 nm was adjusted to 0.15 ( $10^9$  CFU/ml); 100  $\mu\text{l}$  of each cell suspension was mixed with 3 ml of tryptic soy broth containing 0.7% agar and spread evenly over the surface of tryptic soy broth solidified with 1.5% agar containing 300  $\mu\text{M}$  ethylenediamine diacetic acid (EDDA) in 100- by 15-mm petri plates. A 7-mm-diameter Whatman filter paper disk was placed in the center of each plate and impregnated with 10  $\mu\text{l}$  of a solution containing 7.95  $\mu\text{M}$ , 79.5  $\mu\text{M}$ , 795  $\mu\text{M}$ , or 7.95 mM  $\text{MnCl}_2$ . After 72 h of incubation at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$ , the diameter of the zone of bacterial growth surrounding each disk was measured to the nearest millimeter. The diameters of the zones of growth from five separate plates were measured for each bacterial strain examined.

**Growth of *B. abortus* strains in manganese- and iron-restricted media.** Low-manganese minimal medium was prepared using a modification of the protocols originally described by López-Goñi et al. (50). Briefly, a minimal medium based on Gerhardt's minimal medium (31) but supplemented with 0.5 g/liter yeast extract was treated twice with the chelator 8-hydroxyquinolone. After each treatment, the metal-chelate complexes were removed by chloroform extraction and the residual chloroform was removed from the medium by flash evaporation. Following chelator treatment, 50  $\mu\text{M}$   $\text{FeCl}_3$  and 50  $\mu\text{M}$   $\text{MgCl}_2$  (final concentrations) were added to the culture medium to ensure iron- and magnesium-replete

growth conditions. Analysis of representative samples of this medium by atomic absorption spectrophotometry indicates that it contains  $<3$   $\mu\text{M}$  manganese. Low-iron minimal medium was prepared in the same manner, with the exception that following treatment with 8-hydroxyquinolone, 50  $\mu\text{M}$   $\text{MgCl}_2$  and 50  $\mu\text{M}$   $\text{MnCl}_2$  were added to the medium. Analysis of representative samples of the low-iron minimal medium prepared in this manner indicates that it contains  $<3$   $\mu\text{M}$  iron.

*B. abortus* strains were grown on SBA at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$  for 72 h and harvested into the low-manganese minimal medium, and the optical density of the cell suspensions at 600 nm was adjusted to 0.15 ( $10^9$  CFU/ml). One hundred microliters of each suspension was used to inoculate 100 ml of low-manganese medium or low-manganese medium supplemented with 1, 5, 10, or 50  $\mu\text{M}$   $\text{MnCl}_2$ , 5  $\mu\text{M}$   $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$ , 20  $\mu\text{M}$   $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ , or 100  $\mu\text{M}$   $\text{Fe}(\text{NH}_4)_2(\text{SO}_4)_2 \cdot \text{ZnCl}_2$ , or  $\text{CuCl}_2$ . *Brucella* strains grown as described above were also harvested and inoculated into low-iron minimal medium at a cell density of  $10^6$  CFU/ml. Bacterial cultures were incubated at  $37^{\circ}\text{C}$  in 500-ml flasks with shaking at 175 rpm. Bacterial growth in these cultures was evaluated by serial dilution and plating on SBA, followed by incubation of the SBA plates at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$ .

**Resistance of *B. abortus* strains to  $\text{H}_2\text{O}_2$  and the superoxide generators paraquat and menadione in disk sensitivity assays.** *Brucella abortus* 2308, MWV15, and MWV15.C were grown to mid-log phase in brucella broth, and the cultures were adjusted to an optical density at 600 nm of 0.15 (approximately  $10^9$  CFU/ml). One hundred microliters of each cell suspension was mixed with 3 ml of tryptic soy broth containing 0.7% agar and spread evenly over the surface of either a tryptic soy agar plate (for the  $\text{H}_2\text{O}_2$  sensitivity assays) or a tryptic soy agar plate supplemented with 1,000 U/ml of bovine liver catalase (Sigma) (for the assays measuring sensitivity to the superoxide generators paraquat and menadione). A 7-mm-diameter Whatman filter paper disk was placed in the center of each plate and impregnated with 10  $\mu\text{l}$  of one of the following solutions: 30%  $\text{H}_2\text{O}_2$ , 0.5 M paraquat, or 10 mM menadione. After 72 h of incubation at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$ , the diameter of the zone of inhibition surrounding each disk on the plates was measured to the nearest millimeter. The diameters of the zones of inhibition from five separate plates were measured for each bacterial strain examined.

**Determination of superoxide dismutase (SOD) activity in *Brucella* strains.** *Brucella* strains were grown to mid-log phase in brucella broth. Twenty-five-milliliter samples from each culture were harvested by centrifugation at  $10,000 \times g$  for 10 min at  $4^{\circ}\text{C}$  and resuspended in 2 ml native gel buffer (40 mM glycine, 50 mM Tris [pH 8.9]). The cell suspensions were transferred to 2-ml screw-cap polypropylene tubes (catalog no. 72.693; Sarstedt) containing 1 g of 0.1 mM zirconia beads, and the cells were disrupted by subjecting them to 6- to 40-s cycles at 6 m/s in a Savant FastPrep 120 bead beater (Bio 101), with 1 min on ice between each cycle. Cellular debris was harvested by centrifugation at  $4^{\circ}\text{C}$  for 20 min at  $10,000 \times g$ . The cleared supernatant was removed and the protein concentration determined using the Bradford assay (9). Fifteen micrograms of each supernatant was loaded onto a 12% native acrylamide gel and subjected to electrophoresis for 1.5 h at 75 mA. Superoxide activity in the gels was determined using previously described methods (47). Briefly, the gel was soaked in 50 ml of solution 1 (50 mM phosphate buffer [pH 7.5], 28 mM *N,N,N',N'*-tetramethylethylenediamine, 0.028 mM riboflavin) for 10 min in the dark with gentle rocking. Solution 1 was poured off, 50 ml of solution 2 (50 mM phosphate buffer [pH 7.5], 2.5 mM nitroblue tetrazolium) was added, and the gel was allowed to gently rock in this solution for 10 min in the dark. Solution 2 was then poured off, and the gel was exposed to a bench lamp for 15 min. Color development in the gel was stopped by the addition of a 7% solution of acetic acid, and a flatbed scanner was used to capture images of the gels.

**Infection of cultured murine resident peritoneal macrophages.** Following euthanasia, macrophages were harvested from the peritoneal cavities of 9-week-old C57BL/6 mice by lavage with Dulbecco's modified Eagle's medium (DMEM)-5% fetal calf serum (FCS) by using previously described procedures (26). Pooled macrophages in 200  $\mu\text{l}$  of DMEM-5% FCS were cultivated in 96-well plates at a concentration of  $1.6 \times 10^5$  per well at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$ . Cell cultures were enriched for macrophages by washing away nonadherent cells after overnight incubation. *B. abortus* cells were opsonized for 30 min with a subglutinating dilution (1:2,000) of hyperimmune C57BL/6 mouse serum in DMEM-5% FCS. Opsonized bacteria were added to macrophages at a ratio of approximately 100 brucellae per macrophage, and the mixture was incubated at  $37^{\circ}\text{C}$  for 1.5 h to allow time for phagocytosis. The culture medium was then replaced with 200  $\mu\text{l}$  of DMEM-5% FCS containing 50  $\mu\text{g}/\text{ml}$  gentamicin and incubated for 1 h to kill extracellular brucellae. Macrophages were then washed three times with warm PBS-5% FCS and maintained in DMEM-5% FCS containing 12.5  $\mu\text{g}/\text{ml}$  gentamicin for the remainder of the experiment. At 2, 24, and 48 h after infection, the macrophages were lysed with 0.1% deoxycholate and the

number of intracellular brucellae was determined by serial dilution of the lysates in PBS and plating on SBA, followed by incubation of the SBA plates at 37°C with 5% CO<sub>2</sub>.

**Experimental infection of C57BL/6 mice.** Six-week-old female C57BL/6 mice were infected via the peritoneal route with  $5 \times 10^4$  CFU of *B. abortus* 2308, MWV15 (2308  $\Delta$ *mntH*), or MWV15.C [MWV15(pEA31)], and the spleen colonization profiles of these strains in the mice were determined using previously described methods (30).

**Statistical analysis.** All statistical analyses were performed using the two-tailed Student *t* test (64). *P* values of  $\leq 0.05$  were considered significant.

## RESULTS

**BAB1\_1460 is predicted to encode an MntH homolog, and the corresponding gene exhibits Mn-responsive repression in *B. abortus* 2308.** The gene designated BAB1\_1460 in the *B. abortus* 2308 genome sequence is predicted to encode a 456-amino-acid protein that shares 35.9% identity with the *Escherichia coli* manganese transporter MntH (52). Analysis of the amino acid sequence of the putative *Brucella* MntH homolog with the TMpred algorithm ([http://www.ch.embnet.org/software/TMPRED\\_form.html](http://www.ch.embnet.org/software/TMPRED_form.html)) predicts that this protein, like its *E. coli* counterpart, is an integral membrane protein with multiple membrane-spanning regions. All eight of the amino acid residues (Asp34, Asn37, Glu102, Asp109, Glu112, His211, Asp238, and Asn401) that have been shown by site-directed mutagenesis to be critical for Mn<sup>2+</sup> transport by *E. coli* MntH (15, 35) are conserved in *Brucella* MntH. Based on the annotation of the *B. abortus* 2308 genome sequence (14), the *Brucella mntH* homolog is located downstream of a gene (BAB1\_1459) encoding a hypothetical protein. Whether or not BAB1\_1459 is expressed, or *mntH* is cotranscribed in an operon with this gene, has not been experimentally determined.

A bioinformatics-based study described by Rodionov et al. (62) predicts the presence of a “Mur” box upstream of the *Brucella melitensis mntH* homolog BMEI0569. This conserved sequence (AATGCAAATAGTTTGCAAC) is also centered 35 nucleotides upstream of the putative *mntH* coding region (BAB1\_1460) in the *B. abortus* 2308 genome sequence. Mur is a structural homolog of the ferric uptake regulator Fur, which controls the expression of manganese transport genes in *Rhizobium leguminosarum* (21) and *Sinorhizobium meliloti* (16), two alphaproteobacteria that are close phylogenetic relatives of *Brucella* spp., in a manganese-responsive manner. As shown in Fig. 1A, the level of  $\beta$ -galactosidase production from an *mntH-lacZ* fusion in *B. abortus* 2308 during growth in a nutritionally complete growth medium (brucella broth) suggests that *mntH* exhibits a considerable degree of basal expression in this bacterium even when sufficient levels of Mn<sup>2+</sup> are present. The addition of increasing amounts of MnCl<sub>2</sub> to the culture medium ranging from 50  $\mu$ M to 1 mM represses  $\beta$ -galactosidase production in *B. abortus* 2308 (Fig. 1A) but not in the isogenic *mur* mutant Fur2 (Fig. 1B). In contrast, the addition of up to 500  $\mu$ M FeCl<sub>3</sub> fails to repress the expression of the *mntH-lacZ* fusion in either *B. abortus* 2308 or the *mur* mutant (Fig. 1B). This pattern of manganese-responsive expression is consistent with the proposed function of MntH and further indicates that Mur plays an active role in regulating the expression of the corresponding gene.

**The *B. abortus mntH* mutant MWV15 exhibits a manganese-selective defect in metal acquisition in vitro.** The *B. abortus mntH* mutant MWV15 exhibits slower growth in brucella

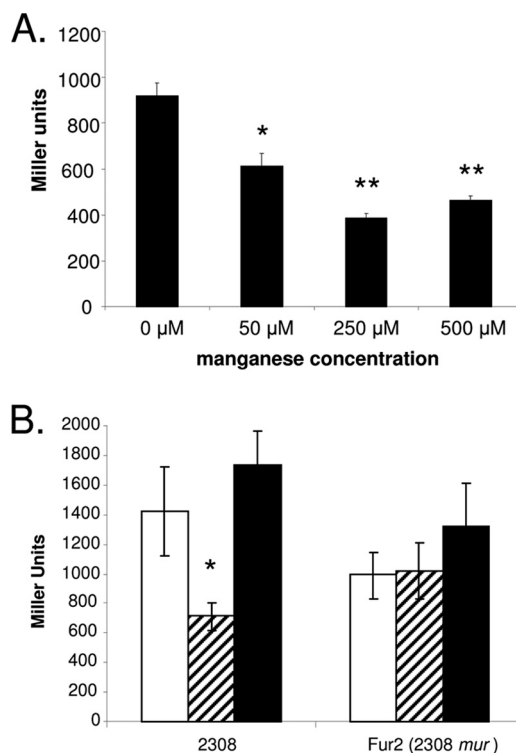


FIG. 1. Manganese-responsive repression of an *mntH-lacZ* transcriptional fusion in *B. abortus* 2308 is mediated by Mur. (A) Expression of an *mntH-lacZ* fusion in *B. abortus* 2308 following 24 h of growth in brucella broth or brucella broth supplemented with 50, 250, or 500  $\mu$ M MnCl<sub>2</sub>. (B) Activity of the *mntH-lacZ* fusion in *B. abortus* 2308 and the isogenic *mur* mutant (Fur2) following 24 h of growth in unsupplemented brucella broth (white bars), brucella broth supplemented with 500  $\mu$ M MnCl<sub>2</sub> (hatched bars), or brucella broth supplemented with 500  $\mu$ M FeCl<sub>3</sub> (black bars).  $\beta$ -Galactosidase activity is presented on the y axis in Miller units (53). The data presented are means and standard deviations for triplicate determinations from a single culture in a single experiment. The data presented are representative of multiple ( $\geq 3$ ) experiments from which equivalent results and statistical trends were obtained. \*, *P* < 0.05; \*\*, *P* < 0.005 (for comparisons of  $\beta$ -galactosidase activities in unsupplemented medium and medium supplemented with MnCl<sub>2</sub> or FeCl<sub>3</sub>).

broth, a complex growth medium, than does the parental 2308 strain or a derivative of MWV15 carrying a plasmid-borne copy of *mntH* (Fig. 2A). Consistent with the predicted function of *Brucella* MntH as a manganese transporter, supplementation of brucella broth with 50  $\mu$ M MnCl<sub>2</sub> allows MWV15 to grow with the same vigor in brucella broth as does strain 2308. Supplementation of this medium with 50  $\mu$ M FeCl<sub>2</sub>, in contrast, will not rescue the growth defect exhibited by MWV15 in brucella broth (Fig. 2A).

The *B. abortus mntH* mutant MWV15 also displays a significantly reduced ability to use MnCl<sub>2</sub> as a manganese source compared to the parental 2308 strain on a solid growth medium containing the chelator EDDA (Table 1). Good growth is observed for the parental 2308 strain surrounding disks containing 79.5  $\mu$ M MnCl<sub>2</sub>, but a comparable level of growth is observed for the *mntH* mutant only around disks containing 7.95 mM MnCl<sub>2</sub> in this assay. EDDA has an approximately 100-fold-greater affinity for Fe<sup>2+</sup> (equilibrium constant =  $6.45 \times 10^9$ ) than it does for Mn<sup>2+</sup> ( $5.13 \times 10^7$ ) (13). Because

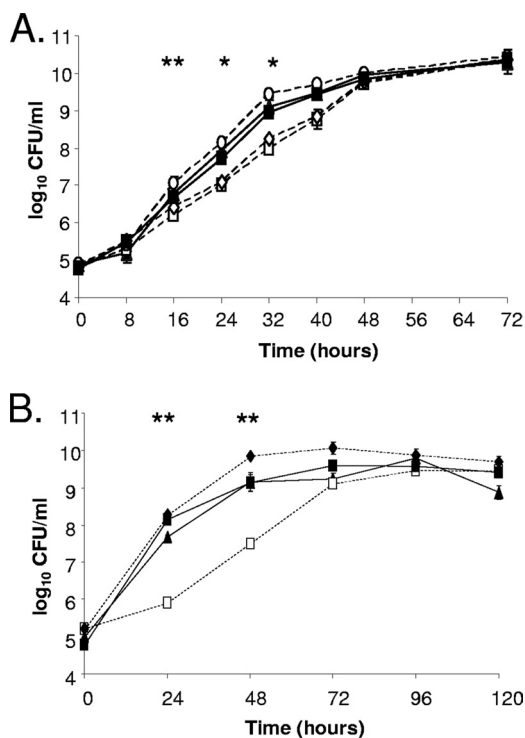


FIG. 2. Supplemental MnCl<sub>2</sub> restores wild-type growth of the *B. abortus mntH* mutant in broth culture. (A) Growth of *B. abortus* 2308 (▲), MWV15 (2308 *mntH*) (□), and MWV15.C [MWV15(pEA31)] (■) in brucella broth and MWV15 in brucella broth supplemented with 50 μM MnCl<sub>2</sub> (◆) or 50 μM FeCl<sub>2</sub> (◇). (B) Growth of *B. abortus* 2308 (▲), MWV15 (□), and MWV15.C (■) in low-manganese minimal medium and MWV15 in low-manganese medium supplemented with 50 μM MnCl<sub>2</sub> (◆). The data presented are the means and standard deviations for triplicate determinations from a single flask for each strain at each experimental time point in a single experiment. The data are representative of multiple (≥3) experiments from which equivalent results and statistical trends were obtained. \*, *P* < 0.05; \*\*, *P* < 0.005 (for comparisons of strain 2308 with MWV15 or MWV15.C).

of its high affinity for iron, EDDA is often the chelator of choice for in vitro experiments designed to detect defects in iron acquisition in bacterial mutants (18). Thus, it is notable that although the *B. abortus mntH* mutant exhibits a re-

TABLE 1. Capacity of *B. abortus* 2308, MWV15, and MWV15.C to use MnCl<sub>2</sub> as a manganese source

Strain	Growth (mm) around disk containing <sup>a</sup> :		
	79.5 μM MnCl <sub>2</sub>	795 μM MnCl <sub>2</sub>	7.95 mM MnCl <sub>2</sub>
2308	43.1 ± 0.6	52.3 ± 1	67.3 ± 0.6
MWV15	NG <sup>b</sup>	NG	42.8 ± 1.4 <sup>***</sup>
MWV15.C	43.4 ± 1.8	52.7 ± 0.3	69 ± 3.2

<sup>a</sup> The diameters of the zones of bacterial growth on a solid growth medium supplemented with 300 μM EDDA around filter disks impregnated with 10 μl of a 10-μg/ml, 100-μg/ml, or 1-mg/ml solution of MnCl<sub>2</sub> were measured after 72 h of incubation at 37°C with 5% CO<sub>2</sub>. The values presented are the means ± standard deviations of the zone sizes obtained from three separate experiments, and three separate determinations of growth were obtained for each strain in each experiment. \*\*\*, *P* ≤ 0.005 (for comparisons of MWV15 with 2308 and MWV15.C).

<sup>b</sup> NG, no growth. No zone of growth was observed surrounding the disks impregnated with 10 or 100 μg/ml MnCl<sub>2</sub> for *B. abortus* MWV15.

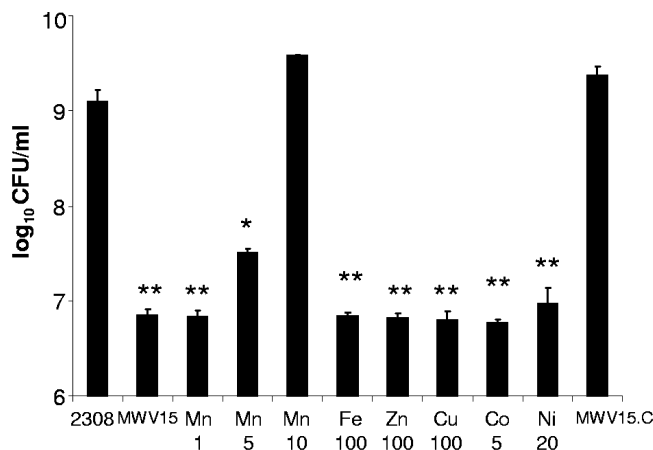


FIG. 3. Growth of the *B. abortus mntH* mutant in low-manganese medium supplemented with Mn<sup>2+</sup>, Fe<sup>2+</sup>, Zn<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, and Cu<sup>2+</sup>. Bacterial cell cultures were inoculated into low-manganese medium at cell densities of approximately 10<sup>5</sup> CFU/ml, and the number of bacteria present in these cultures following 48 h of incubation was enumerated by serial dilution and plating. Shown are the *B. abortus* parental strain (2308), the *mntH* mutant (MWV15), and the *mntH* mutant grown in low-manganese minimal medium supplemented with MnCl<sub>2</sub> at concentrations of 1, 5, and 10 μM (designated Mn1, Mn5, and Mn10, respectively). MWV15 cultures supplemented with 100 μM Fe(NH<sub>4</sub>)<sub>2</sub>(SO<sub>4</sub>)<sub>2</sub> (Fe 100), 100 μM ZnCl<sub>2</sub> (Zn 100), 5 μM CoCl<sub>2</sub> · 6H<sub>2</sub>O (Co 5), and 20 μM NiCl<sub>2</sub> · 6H<sub>2</sub>O (Ni 20) are shown as well. Growth of the *B. abortus mntH* mutant MWV15 carrying a plasmid-borne copy of *mntH* (MWV15.C) in low-manganese minimal medium is also shown. The data presented are the means and standard deviations for triplicate determinations from single flasks for each strain and experimental condition in a single experiment. The data are representative of multiple (≥3) experiments from which equivalent results and statistical trends were obtained. \*, *P* < 0.05; \*\*, *P* < 0.005 (for comparisons of strain 2308 with MWV15 or MWV15.C).

duced zone of growth around disks containing FeCl<sub>3</sub> or Fe(NH<sub>4</sub>)<sub>2</sub>(SO<sub>4</sub>)<sub>2</sub> on plates containing the same concentration of EDDA compared to strain 2308, this reduction in zone size cannot be rescued by adding increasing amounts of either iron source to the disks (data not shown).

Also consistent with its predicted function, the *B. abortus mntH* mutant exhibits delayed growth compared to 2308 when these strains are cultivated in a low-manganese minimal medium in broth culture (Fig. 2B). The addition of 50 μM MnCl<sub>2</sub> restores wild-type growth of the *mntH* mutant in this medium. In contrast, the *B. abortus mntH* mutant exhibits the same growth profile in low-iron minimal medium as the parental 2308 strain (data not shown).

To further examine the selectivity of the metal acquisition defect exhibited by *B. abortus mntH* mutant MWV15, the capacity of increasing concentrations of MnCl<sub>2</sub> or other metals that have been reported to be transported by MntH homologs (e.g., Fe<sup>2+</sup>, Zn<sup>2+</sup>, Cu<sup>2+</sup>, Co<sup>2+</sup>, and Ni<sup>2+</sup>) to alleviate the growth restriction exhibited by this strain in low-manganese medium was evaluated. As shown in Fig. 3, only MnCl<sub>2</sub> at a concentration of 10 μM was able to restore wild-type growth of the *B. abortus mntH* mutant in this medium. CoCl<sub>2</sub> and NiCl<sub>2</sub> were used at lower levels than the other divalent cation sources in these experiments because Co<sup>2+</sup> and Ni<sup>2+</sup> both inhibit the growth of *B. abortus* 2308 and MWV15 equally when used at higher concentrations.

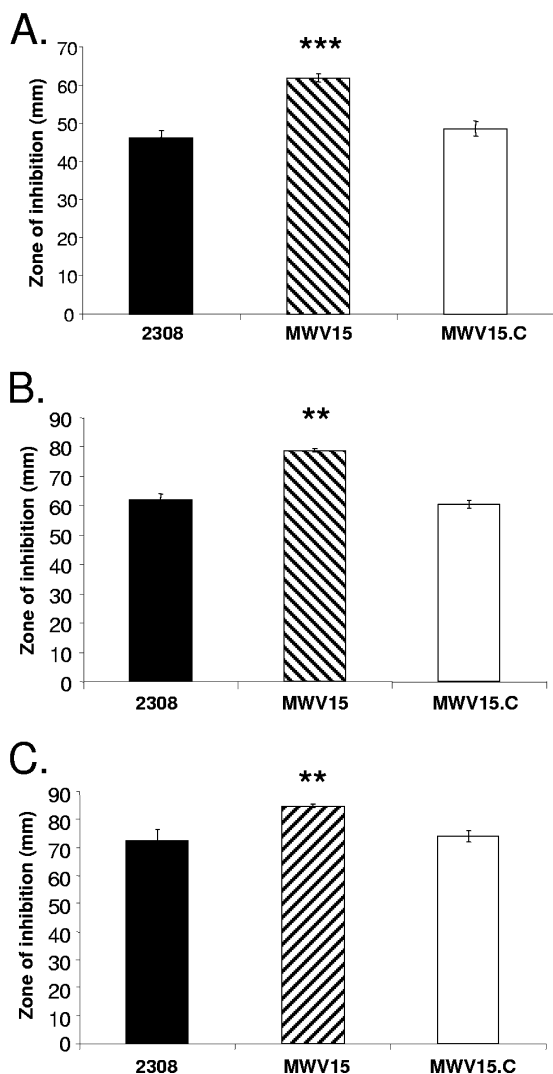


FIG. 4. Resistance of *B. abortus* 2308, MWV15 (2308 *mntH*), and MWV15.C [MWV15(pEA31)] to H<sub>2</sub>O<sub>2</sub> (A), paraquat (B), and menadione (C) in disk sensitivity assays. The data presented are zones of inhibition around disks containing H<sub>2</sub>O<sub>2</sub> (A), paraquat (B), or menadione (C). The values are the means  $\pm$  standard deviations of the zone sizes obtained from three separate experiments, and three separate determinations of growth were obtained for each strain in each experiment. \*\*,  $P \leq 0.005$ ; \*\*\*,  $P \leq 0.001$  (for comparisons of MWV15 with strain 2308 or MWV15.C).

Genetic complementation of *B. abortus* MWV15 with a plasmid-borne copy of the parental *mntH* gene restored the ability of this strain to use MnCl<sub>2</sub> as a manganese source on the EDDA-containing plates with the same efficiency as the parental 2308 strain (Table 1) and restored its ability to replicate with the same growth kinetics as 2308 in the low-manganese minimal medium (Fig. 2A).

**The *B. abortus mntH* mutant MWV15 displays reduced Mn SOD activity compared to the parental 2308 strain.** The presence of MntH is required for wild-type resistance of *B. abortus* 2308 to oxidative killing in *in vitro* assays. The parental 2308 strain is considerably more resistant to killing by both H<sub>2</sub>O<sub>2</sub> (Fig. 4A) and O<sub>2</sub><sup>-</sup> generated by the redox cyclers paraquat

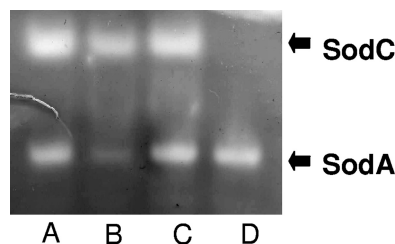


FIG. 5. SOD activity in *B. abortus* 2308, MWV15, and MWV15.C. Cu/Zn SOD (SodC) (cyanide-sensitive) and Mn SOD (SodA) (cyanide- and H<sub>2</sub>O<sub>2</sub>-resistant) activities, detected in the parental 2308 strain (lane A), the *mntH* mutant MWV15 (lane B), and a derivative of MWV15 carrying a plasmid-borne copy of *mntH* (lane C). The *B. abortus sodC* mutant MEK2 (26) was included as a control (lane D). The gel presented is representative of multiple experiments ( $\geq 3$ ) in which equivalent results were observed.

(Fig. 4B) and menadione (Fig. 4C) in disk sensitivity assays than the *mntH* mutant, and the introduction of a plasmid-borne copy of *mntH* into the mutant restores wild-type levels of resistance to H<sub>2</sub>O<sub>2</sub>, paraquat, and menadione in these assays (Fig. 4A to C). In most bacteria that have been studied, there is a strong link between the ability to acquire Mn<sup>2+</sup> and resistance to oxidative stress (39). *B. abortus* 2308 produces a manganese-containing SOD (SodA) (68), and phenotypic analysis of a *B. abortus sodA* mutant indicates that SodA is an important antioxidant in this bacterium (29). Consequently, it is possible that the *B. abortus mntH* mutant is unable to transport sufficient Mn<sup>2+</sup> to produce wild-type levels of Mn SOD activity. The results shown in Fig. 5 support this proposition. Equivalent levels of Cu/Zn SOD activity are observed in native gels for cell lysates from *B. abortus* 2308 and MWV15, but the *mntH* mutant displays a greatly reduced level of Mn SOD activity compared to the parental 2308 strain. MWV15 carrying a plasmid-borne copy of the *mntH* gene, on the other hand, displays the same level of Mn SOD activity as 2308. The basis for the increased sensitivity of the *B. abortus mntH* mutant to H<sub>2</sub>O<sub>2</sub> is unknown, but hypersensitivity to H<sub>2</sub>O<sub>2</sub> has been previously reported for other bacterial strains that are deficient in SOD activity (11).

**MntH is required for wild-type virulence of *B. abortus* 2308 in the C57BL/6 mouse model.** The *B. abortus mntH* mutant MWV15 displays extreme attenuation compared to the parent strain in both cultured murine macrophages (Fig. 6) and experimentally infected mice (Fig. 7). In both cases, the attenuation of the *B. abortus mntH* mutant is alleviated by the introduction of a plasmid-borne copy of the parental *mntH* gene into this strain.

## DISCUSSION

The experimental findings presented in this paper support the proposition that MntH serves as the sole high-affinity manganese transporter in *B. abortus* 2308. A similar role for MntH in *Bradyrhizobium japonicum*, a close phylogenetic relative of brucellae, has also recently been described (37). Many bacteria contain both proton-dependent (MntH-type) and ATP-dependent (SitABCD-type) Mn<sup>2+</sup> transporters, and mutation of both the *mntH* and *sitABCD* genes is often required before prominent defects in Mn<sup>2+</sup> utilization are observed (8, 65).

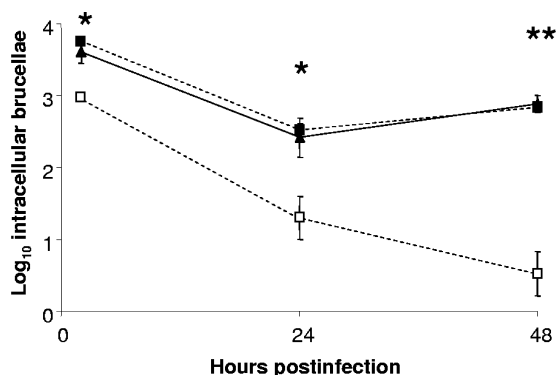


FIG. 6. Survival and replication of *B. abortus* 2308 (▲), MWV15 (2308 *mntH*) (□), and MWV15C [MWV15(pEA31)] (■) in cultured resident peritoneal macrophages from C57BL/6 mice. The data are means and standard deviations of the number of intracellular brucellae recovered for each strain from three separate wells of cultured macrophages at each experimental time point in a single experiment. The data are representative of multiple ( $\geq 3$ ) experiments from which equivalent results and statistical trends were obtained. \*,  $P < 0.05$ ; \*\*,  $P < 0.005$  (for comparisons of MWV15 with 2308 and MWV15.C).

This is clearly not the case with the *B. abortus mntH* mutant, where loss of MntH produces prominent growth defects in both rich and minimal growth media that can be relieved only by supplementation of these media with elevated levels of  $Mn^{2+}$ . It is important to note, however, that the *B. abortus mntH* mutant eventually attains the same cell density as the parental strain in a manganese-deprived minimal medium (Fig. 2), and the addition of high levels of  $MnCl_2$  (e.g., 7.95 mM) to disks will allow this strain to grow on plates containing EDDA (Table 1). These experimental findings demonstrate that *B. abortus* 2308 has an alternate means of acquiring  $Mn^{2+}$  when MntH is not present, but this alternate mechanism is apparently much less efficient at  $Mn^{2+}$  transport than MntH.

Like its counterparts in other bacteria (41), *Brucella* MntH appears to be a manganese-selective transporter. Although supplementation of the culture medium with  $Mn^{2+}$  relieves the growth defects exhibited by the *B. abortus mntH* mutant when this strain is grown in a rich medium or low-manganese minimal medium, supplementation of these media with other divalent cations reported to be transported by Nramp/MntH homologs (e.g.,  $Fe^{2+}$ ,  $Co^{2+}$ ,  $Zn^{2+}$ ,  $Ni^{2+}$ , and  $Cu^{2+}$ ) (34, 52) does not. Particular attention was paid to the possibility that MntH might also be playing a role in iron transport in *B. abortus* 2308 because other prokaryotic MntH homologs have the ability to transport  $Fe^{2+}$  at least under laboratory conditions (1, 41, 52) and efficient iron acquisition has been reported to be an important virulence determinant for this bacterium in both natural (6) and experimental (56) hosts. Although no experimental evidence linking MntH to iron transport in *B. abortus* 2308 was obtained during this study, such a role cannot be definitively ruled out without further experimental evaluation.

Mur, rather than MntR, appears to be the major regulator of  $Mn^{2+}$  acquisition genes in most of the alphaproteobacteria that have been examined (16, 21, 37, 45, 58, 62). Thus, the fact that *mntH* expression in *B. abortus* 2308 is regulated by manganese in a Mur-dependent manner, but is not responsive to iron, is also consistent with the role of MntH as a manganese-

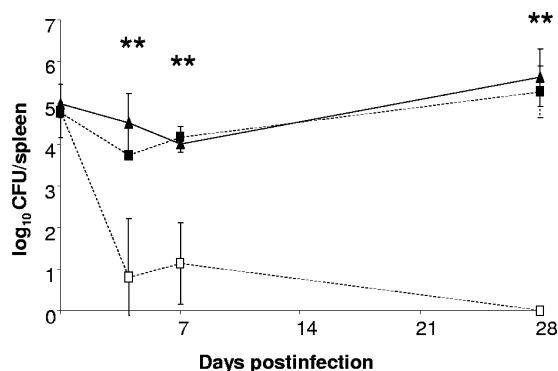


FIG. 7. Spleen colonization profiles of *B. abortus* 2308 (▲), MWV15 (2308 *mntH*) (□), and MWV15.C [MWV15(pEA31)] (■) in C57BL/6 mice. The data presented are means and standard deviations of the number of brucellae detected in the spleens of five mice infected with each strain at each experimental time point in a single experiment. \*\*,  $P < 0.005$  (for comparisons of MWV15 with 2308 and MWV15.C).

selective transporter. The observation that *mntH* apparently exhibits a relatively high level of basal expression during growth in a nutritionally replete medium may be a reflection of the critical role that MntH plays in providing this bacterium with sufficient levels of  $Mn^{2+}$  to meet its physiologic needs and the low toxicity exhibited by  $Mn^{2+}$  compared to other divalent cations (43). The precise nature of the regulatory link between *mntH*, Mur, and cellular Mn levels in *B. abortus* 2308 is presently being examined. A computational analysis of the *B. melitensis* 16M genome sequence suggests that the *Brucella* Mur regulon may be limited to only a few genes other than *mntH* (62).

In most bacteria that have been studied, there is a strong link between the ability to acquire  $Mn^{2+}$  and resistance to oxidative stress (39), and this same relationship is observed for *B. abortus* 2308 and the isogenic *mntH* mutant MWV15. Activity gels indicate that MWV15 produces considerably lower levels of Mn SOD activity than the parental strain, and this deficit in SodA activity is consistent with the increased sensitivity of the *B. abortus mntH* mutant to the redox cyclers menadione and paraquat, which generate  $O_2^-$  in the cytoplasmic compartment of bacterial cells. This link between inefficient  $Mn^{2+}$  transport and suboptimal levels of SOD activity is also similar to the one described for *Sinorhizobium meliloti sitA* mutants (19). It is quite possible, however, that the increased sensitivity of the mutant *B. abortus mntH* to oxidative stress results from defects in addition to its inability to produce wild-type levels of SodA. Anjem et al. (2), for example, recently presented evidence suggesting that MntH-mediated  $Mn^{2+}$  transport allows *E. coli* to metalate key metabolic enzymes with this divalent cation instead of  $Fe^{2+}$ , possibly protecting these enzymes from oxidative damage via Fenton chemistry. Those authors also showed that MntH-dependent  $Mn^{2+}$  transport is an important component of this bacterium's OxyR-mediated response to  $H_2O_2$  exposure. Intracellular levels of  $Mn^{2+}$  can also influence the activity of transcriptional regulators such as PerR that control the expression of genes important for resistance to oxidative stress (38, 49, 70). Preliminary studies suggest that the gene annotated as BAB1\_0393 in the *B. abortus* 2308 genome sequence may be a PerR homolog (E. S. Anderson,

unpublished data), but the nature of the genes that are controlled by this putative regulator and whether or not the activity of the regulator is influenced by cellular  $Mn^{2+}$  levels remain to be determined experimentally.  $Mn^{2+}$  has also been shown to be able to directly detoxify  $H_2O_2$  and  $O_2^-$  in vitro assays (3, 69). It has been postulated, however, that only a few bacteria such as lactobacilli have the capacity to accumulate sufficient levels of  $Mn^{2+}$  (e.g., mM) to make direct intracellular detoxification of ROS by this metal biologically significant (39). Previous studies have shown that wild-type *Brucella* strains require low levels of manganese for growth (28, 66), and those findings are supported by the growth properties exhibited by *B. abortus* 2308 in the study reported here (Fig. 1). Thus, it seems unlikely that direct detoxification of ROS by intracellular  $Mn^{2+}$  plays a major role in oxidative defense in *Brucella* strains.

Nramp1-mediated efflux of  $Mn^{2+}$  and other divalent cations from the phagosomal compartments of macrophages has been proposed to be an important component of host defense against intracellular pathogens such as *Mycobacterium tuberculosis* and *Salmonella enterica* serovar Typhimurium (12, 40). Studies have shown, however, that Nramp1 does not play a prominent role in protecting experimentally infected mice from *Brucella* infections (33). Studies of ruminants have also shown that the contributions of this divalent cation transporter to host defense against *Brucella* infections vary depending on the species of ruminant being examined (10, 54). Thus, the severe attenuation exhibited by the *B. abortus mntH* mutant in C57BL/6 mice (which lack a functional Nramp1) (67) and cultivated macrophages obtained from these mice is particularly striking. This is especially true considering the fact that bacterial *mntH* mutants often exhibit subtle, if any, attenuation in experimental hosts (8, 22), and significant attenuation of *S. Typhimurium mntH sitABCD* double mutants (which lack both of their high-affinity manganese transporters) is seen only in mice that have a functional Nramp1 locus (72). These findings suggest that  $Mn^{2+}$ , like  $Mg^{2+}$  and  $Zn^{2+}$  (44, 48, 71), represents an important micronutrient for brucellae during residence in the mammalian host. They also suggest that the levels of  $Mn^{2+}$  present in the tissues of experimentally infected C57BL/6 mice are insufficient to meet the physiologic requirements of the *B. abortus mntH* mutant for this divalent cation even in the absence of a functional Nramp1 in this mammalian host. Considering the dramatic effect that the loss of MntH has on virulence in *B. abortus* 2308, it is intriguing that brucellae rely on a single high-affinity divalent cation transporter to meet their physiologic needs for  $Mn^{2+}$  during intracellular replication. Indeed, having only one high-affinity  $Mn^{2+}$  transporter appears to be an "Achilles' heel" for these bacteria during residence in the mammalian host. Most other bacterial pathogens that have been studied possess both MntH- and SitABCD-type  $Mn^{2+}$  transporters (55). The bacterial SitABCD-type transporters that have been studied in detail, however, have been reported to be unable to transport  $Mn^{2+}$  at an acidic pH (42, 55), and thus, this type of transporter might be of limited utility to brucellae during the early stages of their intracellular residence in host macrophages when these bacteria occupy acidified compartments (7, 46). In contrast, if *Brucella* MntH is similar to its *Salmonella* counterpart (41) and exhibits optimum  $Mn^{2+}$  transport at an acidic pH, it would

appear to be well suited to the intracellular lifestyle of brucellae in their mammalian hosts.

There are multiple reasons that an insufficient level of intracellular  $Mn^{2+}$  could lead to attenuation in the *B. abortus mntH* mutant. Based on its sensitivity to oxidative killing, the *B. abortus mntH* mutant may be compromised in its ability to withstand the oxidative stresses it encounters during its interactions with host phagocytes. *Brucella* spp. also produce a single bifunctional (p)ppGpp synthetase/hydrolase that has been given the designation Rsh (*RelA/SpoT* hybrid) (24), and the enzymatic activity of this class of bacterial proteins is  $Mn^{2+}$  dependent (55). Because production of (p)ppGpp is required for a stringent response in bacteria (51), reduced Rsh activity due to inadequate  $Mn^{2+}$  levels in the *B. abortus mntH* mutant may interfere with this strain's ability to cope with the nutrient deprivation encountered during long-term residence in the phagosome of host macrophages (46, 63). The presence of a functional Rsh protein has also been shown to be required for the proper expression of the *virB* genes, which encode the type IV secretion machinery in *B. melitensis* 16M and *Brucella suis* 1330 (24). Consequently, inefficient expression of the *virB* genes due to reduced Rsh function could also be contributing to the attenuation exhibited by the *B. abortus mntH* mutant in macrophages and mice. Preliminary studies suggest that *virB4* and *virB5* expression levels are indeed reduced in the *B. abortus mntH* mutant MWV15 compared to those in the parental 2308 strain (J. Gaines and E. Anderson, unpublished data), but the nature of the link between MntH and *virB* expression remains to be experimentally determined.

#### ACKNOWLEDGMENTS

This work was supported by grants (AI48499 and AI63516) from the National Institutes of Health to R.M.R.

#### REFERENCES

- Agranoff, D., I. M. Monahan, J. A. Mangan, P. D. Butcher, and S. Krishna. 1999. *Mycobacterium tuberculosis* expresses a novel pH-dependent divalent cation transporter belonging to the Nramp family. *J. Exp. Med.* **190**:717–724.
- Anjem, A., S. Varghese, and J. A. Imlay. 2009. Manganese import is a key element of the OxyR response to hydrogen peroxide in *Escherichia coli*. *Mol. Microbiol.* **72**:844–858.
- Archibald, F. S., and I. Fridovich. 1982. The scavenging of superoxide radical by manganese complexes: in vitro. *Arch. Biochem. Biophys.* **214**:452–463.
- Bearden, S. W., and R. D. Perry. 1999. The Yfe system of *Yersinia pestis* transports iron and manganese and is required for full virulence of plague. *Mol. Microbiol.* **32**:403–414.
- Bellaire, B. H. 2001. Ph.D. thesis. Louisiana State University Health Sciences Center, Shreveport.
- Bellaire, B. H., P. H. Elzer, S. Hagius, J. Walker, C. L. Baldwin, and R. M. Roop II. 2003. Genetic organization and iron-responsive regulation of the *Brucella abortus* 2,3-dihydroxybenzoic acid biosynthesis operon, a cluster of genes required for wild-type virulence in pregnant cattle. *Infect. Immun.* **71**:1794–1803.
- Bellaire, B. H., R. M. Roop II, and J. A. Cardelli. 2005. Oposonized virulent *Brucella abortus* replicates within nonacidic, endoplasmic reticulum-negative, LAMP-1-positive phagosomes in human monocytes. *Infect. Immun.* **73**:3702–3713.
- Boyer, E., I. Bergevin, D. Malo, P. Gros, and M. F. M. Cellier. 2002. Acquisition of Mn(II) in addition to Fe(II) is required for full virulence of *Salmonella enterica* serovar Typhimurium. *Infect. Immun.* **70**:6032–6042.
- Bradford, M. M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* **72**:248–254.
- Capparelli, R., F. Alfano, M. Grazia Amoroso, G. Borriello, D. Fenizia, A. Bianco, S. Roperto, F. Roperto, and D. Iannelli. 2007. Protective effect of the Nramp1 BB genotype against *Brucella abortus* in the water buffalo (*Bubalus bubalis*). *Infect. Immun.* **75**:988–996.
- Carlizo, A., and D. Touati. 1986. Isolation of superoxide dismutase mutants in *Escherichia coli*: is superoxide dismutase necessary for aerobic life? *EMBO J.* **5**:623–630.

12. Cellier, M. F., P. Courville, and C. Campion. 2007. Nramp1 phagocyte intracellular metal withdrawal defense. *Microbes Infect.* **9**:1662–1670.
13. Chaberek, S., and A. E. Martell. 1959. Organic sequestering agents. John Wiley & Sons, New York, NY.
14. Chain, P. S. G., D. J. Comerci, M. E. Tomalsky, F. W. Larimer, S. A. Malfatti, L. M. Vergez, F. Aguerro, M. L. Land, R. A. Ugalde, and E. Garcia. 2005. Whole-genome analyses of speciation events in pathogenic brucellae. *Infect. Immun.* **73**:8353–8361.
15. Chaloupka, R., P. Courville, F. Veyrier, B. Knudsen, T. A. Tompkins, and M. F. M. Cellier. 2005. Identification of functional amino acids in the Nramp family by a combination of evolutionary analysis and biophysical studies of metal and proton cotransport *in vivo*. *Biochemistry* **44**:726–733.
16. Chao, T.-C., A. Becker, J. Buhrmester, A. Pühler, and S. Weidner. 2004. The *Sinorhizobium meliloti* fur gene regulates, with dependence on Mn(II), transcription of the *sitABCD* operon, encoding a metal-type transporter. *J. Bacteriol.* **186**:3609–3620.
17. Corbel, M. J. 1997. Brucellosis: an overview. *Emerg. Infect. Dis.* **3**:213–221.
18. Cox, C. D. 1994. Deferration of laboratory media and assays for ferric and ferrous ions. *Methods Enzymol.* **235**:315–329.
19. Davies, B. W., and G. C. Walker. 2007. Disruption of *sitA* compromises *Sinorhizobium meliloti* for manganese uptake required for protection against oxidative stress. *J. Bacteriol.* **186**:2101–2109.
20. DelVecchio, V. G., V. Kapatral, R. J. Redkar, G. Patra, C. Mujer, T. Los, N. Ivanova, I. Anderson, A. Bhattacharyya, A. Lykidis, G. Reznik, L. Jablonski, N. Larsen, M. D'Souza, A. Bernal, M. Mazur, E. Goltsman, E. Selkov, P. H. Elzer, S. Hagius, D. O'Callaghan, J. J. Letesson, R. Haselkorn, N. Kyrpides, and R. Overbeek. 2002. The genome sequence of the facultative intracellular pathogen *Brucella melitensis*. *Proc. Natl. Acad. Sci. USA* **99**:443–448.
21. Diaz-Mireles, E., M. Wexler, G. Sawers, D. Bellini, J. D. Todd, and A. W. B. Johnston. 2004. The Fur-like protein Mur of *Rhizobium leguminosarum* is a Mn(2+)-responsive transcriptional regulator. *Microbiology* **150**:1447–1456.
22. Domenech, P., A. S. Pym, M. Cellier, C. E. Barry III, and S. T. Cole. 2002. Inactivation of the *Mycobacterium tuberculosis* Nramp orthologue (*mntH*) does not affect virulence in the mouse model of tuberculosis. *FEMS Microbiol. Lett.* **207**:81–86.
23. Dorrell, N., V. G. Gyselman, S. Foyne, S. R. Li, and B. W. Wren. 1996. Improved efficiency of inverse PCR mutagenesis. *BioTechniques* **21**:604–608.
24. Dozot, M., R.-A. Boige grain, R.-M. Delrue, R. Hallez, S. Ouahrani-Bettache, I. Danese, J.-J. Letesson, X. de Bolle, and S. Köhler. 2006. The stringent response mediator Rsh is required for *Brucella melitensis* and *Brucella suis* virulence, and for expression of the type IV secretion system *virB*. *Cell. Microbiol.* **8**:1791–1802.
25. Elzer, P. H., R. W. Phillips, M. E. Kovach, K. M. Peterson, and R. M. Roop II. 1994. Characterization and genetic complementation of a *Brucella abortus* high-temperature-requirement A (*htrA*) deletion mutant. *Infect. Immun.* **62**:4135–4139.
26. Elzer, P. H., R. W. Phillips, G. T. Robertson, and R. M. Roop II. 1996. The HtrA stress response protease contributes to the resistance of *Brucella abortus* to killing by murine phagocytes. *Infect. Immun.* **64**:4838–4841.
27. Enright, F. M. 1990. The pathogenesis and pathobiology of *Brucella* infections in domestic animals, p. 301–320. *In* K. H. Nielsen and J. R. Duncan (ed.), *Animal brucellosis*. CRC Press, Boca Raton, FL.
28. Evenson, M. A., and P. Gerhardt. 1955. Nutrition of brucellae: utilization of iron, magnesium and manganese for growth. *Proc. Soc. Exp. Biol. Med.* **89**:678–680.
29. Gee, J. M. 2004. Ph.D. thesis. Louisiana State University Health Sciences Center, Shreveport.
30. Gee, J. M., M. W. Valderas, M. E. Kovach, V. K. Grippe, G. T. Robertson, W.-L. Ng, J. M. Richardson, M. E. Winkler, and R. M. Roop II. 2005. The *Brucella abortus* Cu/Zn superoxide dismutase is required for optimal resistance to oxidative killing by murine macrophages and wild-type virulence in experimentally infected mice. *Infect. Immun.* **73**:2873–2880.
31. Gerhardt, P. 1958. The nutrition of brucellae. *Microbiol. Rev.* **22**:81–98.
32. Gober, J. W., and L. Shapiro. 1992. A developmentally regulated *Caulobacter* flagellar promoter is activated by 3' enhancer and IHF binding elements. *Mol. Biol. Cell* **3**:913–926.
33. Guilloteau, L. A., J. Dornand, A. Gross, M. Olivier, F. Cortade, Y. L. Vern, and D. Kerboeuf. 2003. Nramp1 is not a major determinant in the control of *Brucella melitensis* infection in mice. *Infect. Immun.* **71**:621–628.
34. Gunshin, H., B. Mackenzie, U. V. Berger, Y. Gunshin, M. F. Romero, W. F. Boron, S. Nussberger, J. L. Gollan, and M. A. Hediger. 1997. Cloning and characterization of a mammalian proton-coupled metal-ion transporter. *Nature* **388**:482–488.
35. Haemig, H. A. H., and R. J. Brooker. 2004. Importance of conserved acidic residues in MntH, the Nramp homolog of *Escherichia coli*. *J. Membr. Biol.* **201**:97–107.
36. Halling, S. M., B. D. Peterson-Burch, B. J. Bricker, R. L. Zuerner, Z. Qing, L. L. Li, V. Kapur, D. P. Alt, and S. C. Olsen. 2005. Completion of the genome sequence of *Brucella abortus* and comparison to the highly similar genomes of *Brucella melitensis* and *Brucella suis*. *J. Bacteriol.* **187**:2715–2726.
37. Hohle, T. M., and M. R. O'Brian. 2009. The *mntH* gene encodes the major Mn<sup>2+</sup> transporter in *Bradyrhizobium japonicum* and is regulated by manganese via the Fur protein. *Mol. Microbiol.* **72**:399–409.
38. Horsburgh, M. J., S. J. Wharton, A. G. Cox, E. Ingham, S. Peacock, and S. J. Foster. 2002. MntR modulates expression of the PerR regulon and superoxide resistance in *Staphylococcus aureus* through control of manganese uptake. *Mol. Microbiol.* **44**:1269–1286.
39. Horsburgh, M. J., S. J. Wharton, M. Karavolos, and S. J. Foster. 2002. Manganese: elemental defence for a life with oxygen. *Trends Microbiol.* **10**:496–501.
40. Jabado, N., A. Jankowski, S. Dougaparsad, V. Picard, S. Grinstein, and P. Gros. 2000. Natural resistance to intracellular infections: natural resistance-associated macrophage protein 1 (NRAMP1) functions as a pH-dependent manganese transporter at the phagosomal membrane. *J. Exp. Med.* **192**:1237–1247.
41. Kehres, D. G., M. L. Zaharik, B. B. Finlay, and M. E. Maguire. 2000. The NRAMP proteins of *Salmonella typhimurium* and *Escherichia coli* are selective manganese transporters involved in the response to reactive oxygen. *Mol. Microbiol.* **36**:1085–1100.
42. Kehres, D. G., A. Janakiraman, J. M. Schlauch, and M. E. Maguire. 2002. SitABCD is the alkaline Mn<sup>2+</sup> transporter of *Salmonella enterica* serovar Typhimurium. *J. Bacteriol.* **184**:3159–3166.
43. Kehres, D. G., and M. E. Maguire. 2003. Emerging themes in manganese transport, biochemistry and pathogenesis in bacteria. *FEMS Microbiol. Rev.* **27**:263–290.
44. Kim, S., K. Watanabe, T. Shirahata, and M. Watarai. 2004. Zinc uptake system (*znuA* locus) of *Brucella abortus* is essential for intracellular survival and virulence in mice. *J. Vet. Med. Sci.* **66**:1059–1063.
45. Kitphati, W., P. Ngok-Ngam, S. Suwanmaneerat, R. Sukchawalit, and S. Mongolsuk. 2007. *Agrobacterium tumefaciens fur* has important physiological roles in iron and manganese homeostasis, the oxidative stress response, and full virulence. *Appl. Environ. Microbiol.* **73**:4760–4768.
46. Köhler, S., S. Michaux-Charachon, F. Porte, M. Ramuz, and J.-P. Liautard. 2003. What is the nature of the replicative niche of a stealthy bug named *Brucella*? *Trends Microbiol.* **11**:215–219.
47. Langford, P. R., A. Sansone, P. Valenti, A. Battistoni, and J. S. Kroll. 2002. Bacterial superoxide dismutases and virulence. *Methods Enzymol.* **349**:155–166.
48. Lavigne, J.-P., D. O'Callaghan, and A.-B. Blanc-Potard. 2005. Requirement of MgTC for *Brucella suis* intramacrophage growth: a potential mechanism shared by *Salmonella enterica* and *Mycobacterium tuberculosis* for adaptation to a low-Mg<sup>2+</sup> environment. *Infect. Immun.* **73**:3160–3163.
49. Lee, J.-W., and J. D. Helmann. 2006. Biochemical characterization of the structural Zn<sup>2+</sup> site in the *Bacillus subtilis* peroxide sensor PerR. *J. Biol. Chem.* **281**:23567–23578.
50. López-Goñi, I., I. Moriyón, and J. B. Neilands. 1992. Identification of 2,3-dihydroxybenzoic acid as a *Brucella abortus* siderophore. *Infect. Immun.* **60**:4496–4503.
51. Magnusson, L. U., A. Farewell, and T. Nystrom. 2005. ppGpp: a global regulator in *Escherichia coli*. *Trends Microbiol.* **13**:236–242.
52. Makui, H., E. Roig, S. T. Cole, J. D. Helmann, P. Gros, and M. F. M. Cellier. 2000. Identification of the *Escherichia coli* K-12 Nramp orthologue (MntH) as a selective divalent metal ion transporter. *Mol. Microbiol.* **35**:1065–1078.
53. Miller, J. H. 1972. Experiments in molecular genetics, p. 352–355. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
54. Paixão, T. A., F. P. Poester, A. V. Carvalho Neta, A. M. Borges, A. P. Lage, and R. L. Santos. 2007. NRAMP1 3' untranslated region polymorphisms are not associated with natural resistance to *Brucella abortus* in cattle. *Infect. Immun.* **75**:2493–2499.
55. Papp-Wallace, K. M., and M. E. Maguire. 2006. Manganese transport and the role of manganese in virulence. *Annu. Rev. Microbiol.* **60**:187–209.
56. Paulley, J. T., E. S. Anderson, and R. M. Roop II. 2007. *Brucella abortus* requires the heme transporter BhuA for maintenance of chronic infection in BALB/c mice. *Infect. Immun.* **75**:5248–5254.
57. Paulsen, I. T., R. Seshadri, K. E. Nelson, J. A. Eisen, J. F. Heidelberg, T. D. Read, R. J. Dodson, L. Umayam, L. M. Brinkay, M. J. Beanan, S. C. Daugherty, R. T. Deboy, A. S. Durkin, J. F. Dolonay, R. Madupu, W. C. Nelson, B. Ayodeji, M. Kraul, J. Shetty, J. Malek, S. E. Van Aken, S. Riedmuller, H. Tettelin, S. R. Gill, O. White, S. L. Salzberg, D. L. Hoover, L. E. Lindler, S. M. Halling, S. M. Boyle, and C. M. Fraser. 2002. The *Brucella suis* genome reveals fundamental similarities between animal and plant pathogens and symbionts. *Proc. Natl. Acad. Sci. USA* **99**:231–235.
58. Platero, R., L. Peixoto, M. R. O'Brian, and E. Fabiano. 2004. Fur is involved in manganese-dependent regulation of *mntA* (*sitA*) expression in *Sinorhizobium meliloti*. *Appl. Environ. Microbiol.* **70**:4349–4355.
59. Platero, R. A., M. Jauregui, F. J. Battistoni, and E. R. Fabiano. 2003. Mutations in *sitB* and *sitD* genes affect manganese-growth requirements in *Sinorhizobium meliloti*. *FEMS Microbiol. Lett.* **218**:65–70.
60. Que, Q., and J. D. Helmann. 2000. Manganese homeostasis in *Bacillus subtilis* is regulated by MntR, a bifunctional regulator related to the diphtheria toxin repressor family of proteins. *Mol. Microbiol.* **35**:1454–1468.
61. Robertson, G. T., M. E. Kovach, C. A. Allen, T. A. Ficht, and R. M. Roop II. 2000. The *Brucella abortus* Lon functions as a generalized stress response



- protease and is required for wild-type virulence in BALB/c mice. *Mol. Microbiol.* **35**:577–588.
62. **Rodionov, D. A., M. S. Gelfand, J. D. Todd, A. R. J. Curson, and A. W. B. Johnston.** 2006. Computational reconstruction of iron- and manganese-responsive transcriptional networks in  $\alpha$ -proteobacteria. *PLoS Comput. Biol.* **2**:1568–1585.
63. **Roop, R. M., II, B. H. Bellaire, M. W. Valderas, and J. M. Cardelli.** 2004. Adaptation of the brucellae to their intracellular niche. *Mol. Microbiol.* **52**:621–630.
64. **Rosner, B.** 2000. *Fundamentals of biostatistics*, 5th ed. Duxbury, Pacific Grove, CA.
65. **Runyen-Janecky, L., E. Dazenski, S. Hawkins, and L. Warner.** 2006. Role and regulation of the *Shigella flexneri* Sit and MntH systems. *Infect. Immun.* **74**:4666–4672.
66. **Sanders, T. H., K. Higuchi, and C. R. Brewer.** 1953. Studies on the nutrition of *Brucella melitensis*. *J. Bacteriol.* **66**:294–299.
67. **Skamene, E., P. Gros, P. A. Kongshavn, C. St. Charles, and B. A. Taylor.** 1982. Genetic regulation of resistance to intracellular pathogens. *Nature* **297**:506–509.
68. **Sriranganathan, N., S. M. Boyle, G. Schurig, and H. Misra.** 1991. Superoxide dismutases of virulent and avirulent strains of *Brucella abortus*. *Vet. Microbiol.* **26**:359–366.
69. **Stadtman, E. R., B. S. Berlett, and P. B. Chock.** 1990. Manganese-dependent disproportionation of hydrogen peroxide in bicarbonate buffer. *Proc. Natl. Acad. Sci. USA* **87**:384–388.
70. **Wu, H.-J., K. L. Seib, Y. N. Srikhanta, S. P. Kidd, J. L. Edwards, T. L. Maguire, S. M. Grimmond, M. A. Apicella, A. G. McEwan, and M. P. Jennings.** 2006. PerR controls Mn-dependent resistance to oxidative stress in *Neisseria gonorrhoeae*. *Mol. Microbiol.* **60**:401–416.
71. **Yang, X., T. Becker, N. Walters, and D. W. Pascual.** 2006. Deletion of *znuA* virulence factor attenuates *Brucella abortus* and confers protection against wild-type challenge. *Infect. Immun.* **74**:3874–3879.
72. **Zaharik, M. L., V. L. Cullen, A. M. Fung, S. J. Libby, S. L. K. Choy, B. Coburn, D. G. Kehres, M. E. Maguire, F. C. Fang, and B. B. Finlay.** 2004. The *Salmonella enterica* serovar Typhimurium divalent cation transport systems MntH and SitABCD are essential for virulence in an *Nramp1*<sup>G169</sup> murine typhoid model. *Infect. Immun.* **72**:5522–5525.

---

Editor: A. J. Bäumlner