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5 Title: Morphological assessment of the *Octopus vulgaris* species-complex evaluated in  
6 light of molecular-based phylogenetic inferences.

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11

12 Running title: Morphological variation in the *vulgaris* complex

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14

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5 Cryptic species are common in the ocean, particularly among marine invertebrates  
6 such as octopuses. Delineating cryptic species is particularly problematic in octopus  
7 taxonomy where the plasticity recorded among taxonomic characters often results in  
8 low resolution at the species level. This study investigated the morphological  
9 relationships among seven phylogenetic clades (identified using cytochrome *c* oxidase  
10 subunit I) of the broadly distributed This study investigated the morphological  
11 relationships among seven phylogenetic clades of the broadly distributed *Octopus*  
12 *vulgaris* species-complex and close relatives. Morphological analyses in the present  
13 study were successful in delimiting *Octopus sinensis* d'Orbigny, 1841, Brazilian *O.*  
14 *vulgaris* and *O. vulgaris* sensu stricto, which was congruent with the molecular findings  
15 of this study. Male morphology was successful in distinguishing 14 of 15 total pairwise  
16 comparisons, and proved to be a more reliable indicator of species species-level  
17 relationships in comparison to female morphology. The majority of characters with the  
18 greatest discriminatory power were male sexual traits. Significant morphological  
19 differences were also recorded among sampling localities of conspecifics, with  
20 phenotype showing correlation with local environmental data. The findings of this study  
21 support the hypothesis that multiple *O. vulgaris*-like species are currently being  
22 incorrectly treated under a single species name *O. vulgaris*. Octopuses being exported  
23 globally under the name *O. vulgaris* are of extremely high fisheries market value and  
24 profile. Our findings have potentially significant implications for the naming and  
25 conservation of commercially harvested members of this species complex throughout  
26 their ranges.

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## 1 Introduction

2

3 The marine environment has traditionally been thought of as a large continuous system  
4 with relatively few barriers to dispersal. Organisms with an effective dispersal capability  
5 may therefore have the potential to maintain global genetic homogeneity (Waples,  
6 1987). However, dispersal distances of pelagic larvae are influenced by several  
7 physiological and biological factors (Hohenlohe, 2004) and are often unknown  
8 (Knowlton, 1993). A number of examples exist of organisms once thought to be  
9 cosmopolitan in distribution, that are now understood to represent morphologically  
10 similar yet genetically distinct cryptic species with relatively restricted distributions  
11 (Knowlton, 1993; Klautau *et al.*, 1999; Bickford *et al.*, 2007). Cryptic species are  
12 common among marine invertebrates (Knowlton, 1993), many of which lack identifiable  
13 delineating morphological traits (Klautau *et al.*, 1999). This results in cryptic taxa being  
14 'lumped' into single morphospecies, despite being genetically distinguishable. Cryptic  
15 diversity is often missed due to an inability to recognise distinguishing morphological  
16 traits, distortion of specimens through preservation, and/or an inability to quantify the  
17 chemical recognition/communication systems that delineate species.

18 One marine group where cryptic species are common are the cephalopods, including  
19 squids and octopuses (Norman *et al.*, 2014a; Norman *et al.*, 2014b). In recent years,  
20 much attention has been focussed on the taxonomy (Norman & Hochberg, 2005;  
21 Norman *et al.*, 2014b) and phylogenetic relationships (Carlini *et al.*, 2001; Guzik *et al.*,  
22 2005; Strugnell *et al.*, 2008a; Strugnell *et al.*, 2008b; Kaneko *et al.*, 2011; Acosta-Jofré  
23 *et al.*, 2012; Strugnell *et al.*, 2013) within the benthic octopuses and several cryptic  
24 species have been identified (Pickford & McConnaughey, 1949; Söller *et al.*, 2000;  
25 Allcock, 2005; Leite *et al.*, 2008; Allcock *et al.*, 2011; Amor *et al.*, 2014; Reid & Wilson,  
26 2015). The difficulties in identifying octopuses and understanding their evolutionary  
27 relationships are well illustrated by the current uncertainty and confusion surrounding  
28 the phylogeny and taxonomy of genus *Octopus* Cuvier, 1797 (type genus of the family  
29 Octopodidae d'Orbigny, 1839). *Octopus* has long been considered a 'catch all' genus  
30 (e.g., Nesis, 1998), with few morphological characters available for distinguishing  
31 among closely related taxa, but it has recently been characterised as octopuses with a  
32 well-defined 'patch-and-groove' skin topology, robust muscular arms with 200–  
33 350 prominent suckers in two columns down each arm, and arms two and three longer  
34 than arms one and four by a margin of around one mantle length (Gleadall, 2016).

1 Distinguishing octopus species is also hindered by their morphological plasticity  
2 (Robson, 1929; Pickford, 1945; Voight, 1994; O'Shea, 1999) since their characteristic  
3 soft body has few hard structures (Bookstein *et al.*, 1985) and is subject to distortion  
4 upon preservation (Pickford, 1964; Burgess, 1966; Voight, 2001). This means that  
5 using morphological characters to distinguish closely related species is particularly  
6 difficult (e.g., Norman & Kubodera, 2006) but recent morphology-based studies  
7 suggest that benthic octopuses can be distinguished based on discrete phenotypic  
8 differences (Gleadall *et al.*, 2010; Gleadall, 2013; Amor *et al.*, 2014; Gleadall, 2016).  
9 Recent taxonomic revisions (O'Shea, 1999; Norman *et al.*, 2014a) and molecular-  
10 based phylogenetic studies (Guzik *et al.*, 2005; Kaneko *et al.*, 2011; Acosta-Jofré *et al.*,  
11 2012; Lü *et al.*, 2013) have confirmed that genus *Octopus* as used previously was a  
12 polyphyletic assemblage of species groups comprising a number of different genera..  
13 The species group closest in morphology and behaviour to the type species of the  
14 genus (*Octopus vulgaris* Cuvier, 1797) has been identified as the '*O. vulgaris* species-  
15 group,' based on general similarities in size, mantle shape, arm length and skin  
16 sculpture (Robson, 1929). Species in this group are now considered to comprise the  
17 genus *Octopus* sensu stricto (O'Shea, 1999).

18 *Octopus vulgaris* was long considered to be a cosmopolitan species. First reported  
19 from the Mediterranean Sea and eastern North Atlantic, *O. vulgaris* has been reported  
20 from around the world. However, recent analyses (Söller *et al.*, 2000; Leite *et al.*, 2008;  
21 Amor *et al.*, 2014; Amor *et al.*, 2015; Gleadall, 2016) suggest that populations  
22 previously treated as *O. vulgaris* comprise a complex of morphologically similar but  
23 genetically distinct *vulgaris*-like species, the '*O. vulgaris* species-complex'. *Octopus*  
24 *vulgaris* sensu stricto (s. s.) occurs in the Mediterranean and eastern North Atlantic.  
25 Other members of this species-complex include several species 'Types,' which have  
26 been recognised based on geographic isolation and lack of plausible gene flow  
27 mechanisms (Norman *et al.*, 2014a). Type I occurs in the Caribbean and Gulf of  
28 Mexico; Type II in the western South Atlantic along the coast of Brazil; and Type III  
29 occurs in the eastern South Atlantic and the Indian Ocean, along the coast of South  
30 Africa. *Octopus sinensis* d'Orbigny, 1841, occurs in the western North Pacific (Gleadall,  
31 2016). Recent analyses based on molecular sequencing support the hypothesis that *O.*  
32 *vulgaris* s. s., *O. sinensis* and *O. vulgaris* Type II represent distinct species within the  
33 *O. vulgaris* species-complex (Amor *et al.*, 2015). However, the only recent  
34 morphological comparison undertaken to investigate the taxonomic relationships  
35 among members of the *O. vulgaris* species-complex are those between *O. vulgaris* s. s.

1 and *O. insularis* Leite & Haimovici, 2008 (in Leite *et al.*, 2008) and *O. sinensis* (Gleadall,  
2 2016). The present study employs the first ever global scale sampling strategy to  
3 investigate morphological variation and determine the validity of morphologically based  
4 identifications among members and close relatives of the *O. vulgaris* species-complex.  
5 Analyses are combined for conventional morphological traits and a more extensive  
6 data set. Phylogenetic analyses based on the mitochondrial 'barcode of life' gene *COI*  
7 are also used to provide insights into taxonomic resolution among taxa currently  
8 included within the species *O. vulgaris*.

9

## 10 Materials and methods

11

### 12 *Sampling*

13

14 Whole specimens and tissue samples of *O. vulgaris* species-group individuals were  
15 obtained from museums, university collections and fish markets from the continental  
16 shelves of the Atlantic, Indian and Pacific oceans and the Mediterranean Sea (Fig. 1,  
17 Table 1). Tissue samples were removed from fresh or frozen specimens and stored in  
18 70-90% ethanol. Specimens were then fixed in 10% formalin following methods  
19 outlined in Roper and Voss (1983), washed in tap water and later preserved in 70%  
20 ethanol.

21

22 [Insert Fig.1]

23

24 [Insert Table 1]

25

### 26 *Molecular analyses*

27

1 *Sequencing*: Genomic DNA was extracted from mantle or arm tissue samples of 1-2  
2 mm<sup>3</sup> (after first trimming away skin where possible) using a QIAGEN DNeasy Blood &  
3 Tissue Kit according to the manufacturer's instructions. Partial cytochrome c oxidase  
4 subunit I (*COI*) sequences were amplified via PCR using the universal primers  
5 LCO1490 and HCO2198 (Folmer *et al.*, 1994). PCR solutions (25 µL) were composed  
6 of 0.5 µL forward primer (10 µM), 0.5 µL reverse primer (10 µM), 12.5 µL MyTaq Red  
7 Mix (*Bioline*), 9.5 µL H<sub>2</sub>O and 2 µL DNA (5-10 ng total concentration). PCR cycle  
8 conditions were as follows: a single initial denaturing step (two minutes at 95°C); 35  
9 cycles of denaturing (30 seconds at 95°C); annealing (30 seconds at 48°C); and  
10 extension (30 seconds at 72°C); and a single final extension step (five minutes at  
11 72°C). PCR products were sequenced by Macrogen Inc (Seoul, Korea). *COI*  
12 sequences generated in this study were deposited in GenBank under accession  
13 numbers KU525758-KU525769. Additional sequences from previously published work  
14 were obtained from GenBank (Table S1). *Octopus cyanea* was selected as the  
15 outgroup to root the phylogenetic tree (Amor *et al.*, 2015). Multiple sequence alignment  
16 of the 482 base pair partial *COI* fragments was performed using *Geneious 7.1.3*  
17 (created by Biomatters; available from <http://www.geneious.com/>) and the 'Muscle  
18 Alignment' feature (Larkin *et al.*, 2007).

19 *Molecular-based phylogenetic analyses*: *jModelTest* v0.1.1 (Posada, 2008) was used  
20 to select the best-fit models of nucleotide substitution of the *COI* alignment. The  
21 appropriate model (GTR+G) was chosen based on 'goodness of fit' via the Akaike  
22 Information Criterion (AIC; Akaike, 1974). Maximum likelihood (ML) topologies were  
23 constructed using *RAxML* v8.0.19 (Stamatakis, 2014). Strength of support for internal  
24 nodes of ML construction was measured using 1000 rapid bootstrap replicates.  
25 Bayesian inference (BI) marginal posterior probabilities were calculated using *MrBayes*  
26 v3.2 (Ronquist & Huelsenbeck, 2003). Model parameter values were treated as  
27 unknown and were estimated. Random starting trees were used and the analysis was  
28 run for fifteen million generations, sampling the Markov chain every 1,000 generations.  
29 A mean standard deviation of split frequencies of <0.01 was used as a guide to ensure  
30 the two independent analyses had converged. The program *Tracer* v1.3 (Rambaut &  
31 Drummond, 2003) was then used to ensure Markov chains had reached stationarity,  
32 and to determine the correct 'burn-in' for the analysis.

33

34 *Morphological analyses*

1

2 Standard morphological characters were measured using digital callipers following  
3 Roper and Voss (1983) and Norman and Sweeney (1997): dorsal mantle length (MLd),  
4 ventral mantle length (MLv), mantle width (MW), head width (HW), funnel length (FL),  
5 free funnel length (FFL), gill length (GL) and length of the male hectocotylus (third right,  
6 R3). Enlarged sucker diameter (SDe), non-enlarged sucker diameter (SDn),  
7 specialisations at the tip of the hectocotylus (ligula length, LL; calamus length, CL), the  
8 length of the male reproductive tract terminal organ length (TOL) and arm width (AW)  
9 were all recorded to the nearest 0.1 mm. Web depth (WD) was measured from the  
10 beak opening to the mid-point of the web sector; and the length of the arms on the left  
11 (ALL1-4) and right (ALR1-4) side from the beak opening to the arm tip, were measured  
12 to the nearest 1 mm using stretch-resistant cord. The number of suckers on the left  
13 third arm (SCL) and the right third arm (SCR; which for males is the sucker count of the  
14 hectocotylised arm, HASC) were counted with the aid of a dissecting microscope. Arm  
15 lengths and sucker counts were excluded where damage to an arm was perceived to  
16 inhibit growth, suckers appeared damaged and no scars/remnants were visible, or arm  
17 regeneration was evident (Tables S2 and S3). All missing data due to these exclusions  
18 were replaced with the 'local' mean of that trait across the geographic location as  
19 missing data was not permitted in analyses.

20 Morphological datasets were recorded only for mature males and females. To account  
21 for differences attributed to variation in overall size, and to allow for investigation of size  
22 free trait variation, all morphometric and meristic traits (with the exception of SC, FFL,  
23 LL and DL) were transformed to indices, dividing each trait by the specimen's dorsal  
24 mantle length (a proxy for body size). The remaining indices were obtained as follows:  
25 Sucker counts of each arm were divided by the respective arm length, FFL was divided  
26 by FL, LL was divided by CL, and DL was divided by TOL. Morphological relationships  
27 were investigated using the complete set of traits recorded during the present study (25  
28 traits for males; 20 traits for females; Tables S2 and S3, respectively). For comparison  
29 with published data, a reduced number of traits was also analysed independently (12  
30 traits for males; 8 traits for females; see traits marked with '\*' in Tables S2 and S3,  
31 respectively). The reduced set of traits were MLd, MW, HW, FL, FFL, WD, ALL3/R3,  
32 SDn, SCL3/R3 (HASC, males only), LL (males only) and CL (males only). Analyses of  
33 reduced and complete trait data sets were performed on males and females separately



1 to enable the inclusion of male specific reproductive characters in morphological  
2 analyses.

3 Morphological indices of both males and females were mean scale transformed  
4 (Berner, 2011), and normalised using the 'normalise variables' function in PRIMER E+  
5 v6 and PERMANOVA+ (Anderson *et al.*, 2008) to enable comparisons of traits despite  
6 differing scales of measurement. All morphological analyses were performed using  
7 PRIMER E+ v6 (Clarke & Gorley, 2006) and PERMANOVA+ (Anderson *et al.*, 2008).  
8 Collinearity and redundancy of morphological traits was investigated via Principal  
9 Component Analysis (PCA) vector plots, Draftsman plots and Spearman correlation  
10 matrices as detailed in the user manual (Anderson *et al.*, 2008). Highly correlated  
11 variables ( $R^2 \geq 85\%$ ) were considered redundant. The effect of within-clade multivariate  
12 dispersion (i.e. the significance of within-clade variation contributing to between-clade  
13 differences) was investigated via permutational distance-based tests for homogeneity  
14 of multivariate dispersions (PERMDISP). Differences in morphological traits among  
15 sampled individuals were analysed via permutational multivariate ANOVA  
16 (PERMANOVA). A resemblance matrix based on Euclidean distance was calculated.  
17 To visualise the relationships among locations, PCA was performed using the *COI*-  
18 based phylogenetic clade as an independent factor to group individuals into  
19 taxonomically informative entities. Variable contributions to variation were investigated  
20 via Similarity Percentages (SIMPER) analysis (Clarke, 1993). In order to evaluate the  
21 discriminative power of the morphological traits used, estimates of group assignment  
22 were performed using Canonical Analysis of Principal Components (CAP).

23

#### 24 *Comparative analyses*

25

26 Environmental data were incorporated to estimate correlations between morphological  
27 variation and each environmental predictor variable. Mean annual (1900-1997) sea  
28 surface temperature (SST), sea bottom temperature (SBT) and salinity were obtained  
29 from NOAA (2014). A distance based linear model (Anderson *et al.*, 2008) was used to  
30 perform a marginal test on each environmental variable to determine the overall  
31 morphological variation explained. To quantify the variability in the morphological

1 resemblance matrix that was explained by environmental variables, a step-wise  
2 sequential test was performed using the AIC to select the model of best fit.

3

## 4 Results

5

### 6 *Phylogenetic relationships*

7

8 Topologies resulting from molecular-based ML and BI analyses showed a highly  
9 supported monophyletic clade containing *O. insularis*, *O. mimus* Gould, 1852, *O.*  
10 *bimaculoides* Pickford and McConnaughey, 1949, and *O. maya* Voss and Solís  
11 Ramírez, 1966 (bootstrap value [BS] = 95, posterior probability [PP] = 1; Fig. 2). This  
12 clade was sister taxon to (1) a clade containing *O. hummelincki* Adam, 1936, and (2) a  
13 clade containing the *O. vulgaris* species-complex, *O. tetricus* Gould, 1852, and *O. cf.*  
14 *tetricus* of Australasia (BS = 64, PP = 0.66). All members of the *O. vulgaris* species-  
15 complex formed a highly supported monophyletic clade which also included *O. tetricus*  
16 and *O. cf. tetricus* (BS = 95, PP = 1; *O. vulgaris* group). The *O. vulgaris* species-  
17 complex formed three distinct monophyletic clades, which corresponded to three of the  
18 *O. vulgaris* 'Types' described in Norman *et al.*, (2014a): Clade 9, *O. sinensis*  
19 (Kermadec Is and Asia; BS = 75, PP = 1); Clade 10, *O. vulgaris* Type II (southern  
20 Brazil: BS = 69, PP = 0.83); and Clade 11, *O. vulgaris* s. s. and *O. vulgaris* Type III  
21 (South Africa: BS = 88, PP = 1), which also included a single individual from southern  
22 Brazil.

23

24 [Insert Fig. 2]

25

### 26 *Morphological relationships*

27

28 *Comparison of complete and reduced trait datasets*: PERMANOVA comparisons and  
29 assignment of individuals to their *a priori* molecular-based phylogenetic clade via CAP

1 were more successful using male and female complete trait datasets (Tables 2-3 and  
2 S8-S11). Analyses based on the reduced trait datasets are presented in online  
3 supplementary data associated with this manuscript. Analyses based on the complete  
4 trait data sets are presented below.

5 *Analyses of male specimens:* Male arm lengths (L2, L3, L4 and R2) displayed  $\geq 85\%$   
6 correlation with each other. Arm length data was most complete for arm L3, therefore  
7 ALL3 was retained whilst the remaining correlated arm lengths were considered  
8 redundant and excluded from analyses. Within-clade variation had no significant impact  
9 among clade analyses ( $p = >0.05$ ). A significant difference was recorded among the six  
10 molecular-based phylogenetic clades investigated (Pseudo-F = 5.2805,  $df = 5$ ,  $p =$   
11 0.001). Pairwise comparisons among these six phylogenetic clades showed 14/15  
12 (93%) significant differences (Table 2). All members of the *O. vulgaris* species-complex  
13 were distinguished based on morphological analyses ( $p = <0.02$ ). *Octopus vulgaris* s.  
14 s. and *O. sinensis* were distinguished primarily by differences in GL and ALR4.  
15 *Octopus vulgaris* s. s. was distinguished from *O. vulgaris* Type II primarily by SDe.  
16 *Octopus sinensis* was distinguished from *O. vulgaris* Type II by significantly longer gills  
17 (GL).

18 *Octopus insularis* specimens were found to be morphologically distinct from all other  
19 taxa in the *O. vulgaris* species-complex ( $p = <0.002$ ). The greatest sources of variation  
20 between *O. vulgaris* s. s. and *O. insularis* were attributed to differences in ALR3 and  
21 HASC. *Octopus vulgaris* Type II and *O. insularis* were primarily distinguished by DL  
22 and HASC. *Octopus sinensis* and *O. insularis* were distinguished by variations in GL  
23 and TOL. *Octopus tetricus* and *O. cf. tetricus* differed significantly from each other  
24 ( $p=0.012$ ), particularly through differences in SCL3 and DL. No morphological  
25 differences were found between *Octopus vulgaris* s. s. and *O. cf. tetricus* ( $p = 0.095$ ).

26

27 [Insert Table 2]

28

29 Based on the principal component biplot for males (Fig. 3a), *O. vulgaris* s. s. and *O.*  
30 *vulgaris* Type II males showed the greatest morphological variability in comparison to  
31 other taxa, as demonstrated by their occupation of highly positive and highly negative  
32 PC1 and PC2 spaces. *Octopus vulgaris* s. s., *O. sinensis* and *O. vulgaris* Type II

1 showed the least discrimination, although *O. vulgaris* s. s. and *O. vulgaris* Type II  
2 individuals had relatively longer arms than *O. sinensis* (PC1). *Octopus vulgaris* Type II  
3 individuals had relatively fewer suckers on the third arm pair than *O. vulgaris* s. s. and  
4 *O. sinensis* (PC2). *Octopus tetricus*, *O. cf. tetricus* and *O. insularis* demonstrated  
5 negative PC2 loadings attributed to high sucker numbers. *Octopus tetricus* and *O.*  
6 *insularis* showed the least overlap with other taxa included in the analysis but *O. cf.*  
7 *tetricus* overlapped with all members of the *O. vulgaris* species-complex.

8

9 [Insert Fig. 3]

10

11 Of the 68 male individuals analysed, 54 (79%) were correctly assigned to their *a priori*  
12 group via CAP (Table 2). For *O. vulgaris* s. s., 16 individuals (84%) were correctly  
13 classified: the remainder were misclassified as *O. sinensis* (n = 3). Twelve *O. sinensis*  
14 individuals (75%) were correctly assigned to their *a priori* group, with the remaining  
15 individuals being misclassified as *O. vulgaris* s. s. (n = 1), Brazilian Type II (n = 1), *O.*  
16 *insularis* (n = 1) or *O. cf. tetricus* (n = 1). Nine *O. vulgaris* Type II individuals (82%)  
17 were correctly classified whilst the remaining individuals were misclassified as *O.*  
18 *vulgaris* s. s. (n = 1) and *O. insularis* (n = 1). Eight *O. insularis* individuals were  
19 correctly assigned (67%), with the remaining individuals misclassified as *O. vulgaris* s.  
20 s. (n = 1), *O. tetricus* (n = 1) or *O. cf. tetricus* (n = 2). Four *O. tetricus* individuals (80%)  
21 were correctly assigned, with the remaining individual being misclassified as *O.*  
22 *sinensis*. All *O. cf. tetricus* individuals (n = 5) were correctly assigned to their respective  
23 *a priori* group.

24 *Analysis of female specimens:* Significant within-clade variation was recorded for *O.*  
25 *vulgaris* s. s. and *O. insularis* females ( $p = 0.03$ ). The main-effects model showed  
26 significant morphological differences among the six molecular-based phylogenetic  
27 clades of female individuals (Pseudo-F = 3.8184,  $df = 5$ ,  $p = 0.001$ ). Pairwise  
28 comparisons showed that 10/15 (67%) comparisons had significant morphology-based  
29 differences (Table 3). All members of the *O. vulgaris* species-complex (*O. vulgaris* s. s.,  
30 *O. sinensis* and south Brazilian Type II) were successfully distinguished based on  
31 multivariate morphological analyses ( $p = \leq 0.01$ ). *Octopus vulgaris* s. s. and *O. sinensis*  
32 were distinguished primarily by arm length (L3) and sucker diameter. Arm width was

1 the primary source of variation between *O. vulgaris* s. s. and *O. vulgaris* Type II.  
2 *Octopus sinensis* and *O. vulgaris* Type II were found to differ in gill length and arm  
3 width. All members of the *O. vulgaris* species-complex could be distinguished from *O.*  
4 *insularis* ( $p = \leq 0.003$ ). Variation between *O. vulgaris* s. s. and *O. insularis* was primarily  
5 attributed to differences in the number of suckers on the third arm pair, which was also  
6 the greatest source of variation between *O. vulgaris* Type II and *O. insularis*. *Octopus*  
7 *sinensis* and *O. insularis* were best delineated by the variation in sucker numbers on  
8 the third left arm. No morphological distinctions were detected between *O. tetricus* and  
9 *O. cf. tetricus* ( $p = 0.3$ ).

10

11 [Insert Table 3]

12

13 The principal component biplot for females (Fig. 3b) showed that *O. vulgaris* s. s. and  
14 *O. sinensis* have the most morphological variability, with highly positive and negative  
15 PC1 and PC2 loadings. *Octopus vulgaris* Type II was characterised by positive PC2  
16 loadings (low SCL/R3). *Octopus insularis* individuals formed a distinct group  
17 characterised by positive PC1 and negative PC2 loadings (low arm lengths and high  
18 sucker counts).

19 Overall, 41 of the 62 analysed female individuals (66%) were correctly assigned via  
20 CAP (Table 3). Sixteen *O. vulgaris* s. s. individuals (76%) were correctly classified,  
21 whilst four individuals were misclassified as *O. sinensis* and a single individual as *O.*  
22 *tetricus*. Ten *O. sinensis* individuals (50%) were correctly assigned to their *a priori*  
23 group, with the remaining individuals being misclassified as *O. vulgaris* s. s. ( $n = 5$ ), *O.*  
24 *insularis* ( $n = 1$ ), *O. tetricus* ( $n = 2$ ) and *O. cf. tetricus* ( $n = 2$ ). Five *O. vulgaris* Type II  
25 individuals (71%) were correctly assigned, with a single individual misclassified as *O.*  
26 *vulgaris* s. s., *O. sinensis* and *O. tetricus*. All *O. insularis* individuals ( $n = 6$ ) were  
27 correctly assigned, whilst 75% of *O. tetricus* and 50% of *O. cf. tetricus* individuals were  
28 assigned correctly.

29 *Reduced trait analyses of male O. vulgaris* s. s.: Significant differences were recorded  
30 among Galician, Mediterranean and Mauritanian males ( $p = 0.001$ ), with the pairwise  
31 multivariate model showing a significant difference among the three localities ( $p =$   
32  $\leq 0.004$ ; Table 4).

1 [Insert Table 4]

2

3 A PC biplot (Fig. 4a) showed that, basically, each sampling locality for *O. vulgaris* s. s.  
4 males could be distinguished, although a small number of individuals overlapped.  
5 Individuals from the Mediterranean were found to have more suckers (L3, R3) than  
6 Galician and Mauritanian (eastern North Atlantic) individuals. Galician males were  
7 distinct from Mauritanian males along PC1, Galician males having longer arms (L3,  
8 R3).

9

10 [Insert Fig. 4]

11

12 Based on the CAP, 24 of the 27 *O. vulgaris* s. s. males (89%) were correctly assigned  
13 (Table 4). All individuals from Mauritania (n = 8) were successfully assigned to their  
14 correct sampling locality: eight of the nine Mediterranean individuals (89%) were  
15 correctly assigned, with a single individual being misclassified as Galician; and eight of  
16 the ten Galician individuals (80%) were correctly assigned, with the remaining two  
17 individuals misclassified as Mauritanian.

18 Variation attributable to environmental data explained 31.4% of the variation in male  
19 morphology ( $R^2 = 0.31354$ ). Investigating each trait independently via marginal tests,  
20 SST explained 21.3% ( $p = 0.001$ ) and SBT 21.2 % ( $p = 0.001$ ) of the variation.  
21 Sequential tests revealed that SST accounted for 21.3% of the variation seen in the  
22 morphological data ( $p = 0.002$ ). Once SST was accounted for, SBT explained a further  
23 10% of the variation ( $p = 0.002$ ). Latitude, longitude and depth did not explain any  
24 further variation, although each was found to explain a significant amount of the  
25 variation in morphology when analysed independently ( $p = 0.001$ ,  $p = 0.005$  and  $p =$   
26  $0.023$ , respectively),

27 *Reduced trait analyses of female O. vulgaris* s. s.: A significant difference was recorded  
28 among Galician, Mediterranean and Mauritanian females ( $p = 0.001$ ), with the pairwise  
29 multivariate model showing a significant difference among the three localities ( $p =$   
30  $\leq 0.002$ ; Table 5)

1

2 [Insert Table 5]

3

4 A PC biplot (Fig. 4b) distinguished *O. vulgaris* s. s. females by locality. Individuals from  
5 the eastern North Atlantic (Galicia and Mauritania) were more similar to each other  
6 than they were to Mediterranean females, which have longer funnels (FL). Individuals  
7 from the eastern North Atlantic differed, with Galician males possessing more suckers  
8 (SCL/SCR) and a larger head (HW).

9 Of 27 female *O. vulgaris* s. s. individuals, 26 (96%) were correctly assigned to their *a*  
10 *priori* group (Table 5). Individuals from Mauritania and the Mediterranean (France)  
11 were all assigned with 100% accuracy, and nine of the ten Galician individuals were  
12 assigned correctly (90%), with the remaining individual misclassified as Mediterranean.

13 Of the overall variation in female morphology, 33.9% was explained by variation in  
14 environmental data ( $R^2 = 0.33854$ ). Investigating each trait independently via marginal  
15 tests showed that latitude explained 20.8% ( $p = 0.001$ ) and SST 18.8% ( $p = 0.001$ ) of  
16 the variation. In sequential tests, latitude accounted for 20.8% of the morphological  
17 variation ( $p = 0.001$ ). With latitude accounted for, SST explained a further 13% of the  
18 variation ( $p = 0.002$ ); and once both latitude and SST were accounted for, SBT,  
19 longitude and depth explained no further variation (although a significant amount of  
20 variation in morphology was explained when these parameters were analysed  
21 independently:  $p = 0.002$ ,  $p = 0.001$  and  $p = 0.001$ , respectively).

22

23 Discussion

24

25 Molecular-based phylogenetic analyses of *O. vulgaris* species-group individuals in the  
26 present study support the presence globally of six distinct clades, which were used as  
27 a discriminant factor in morphological analyses. Conventional morphological traits were  
28 successful in distinguishing the majority of these clades. Multivariate morphological  
29 analyses support the hypothesis of species distinctions within the *O. vulgaris* species-  
30 complex (*O. vulgaris* s. s., *O. vulgaris* Type II and *O. sinensis*). Although each of these

1 species was successfully distinguished, further distinctions were detected among the  
2 sampling localities of *O. vulgaris* s. s., suggesting a requirement for broad sampling of  
3 species-level morphology across the known distribution for each species to ensure  
4 robust future morphological analyses of this group.

5 Previous molecular sequence-based phylogenetic analyses using five mitochondrial  
6 genes placed *O. sinensis* into a well-supported monophyletic clade, distinct from all  
7 other members of the *O. vulgaris* species-complex (Amor *et al.*, 2014). Reid and  
8 Wilson (2015) considered mitochondrial-based differences to warrant the distinction of  
9 Kermadec Island individuals from *O. vulgaris* s. s., establishing the name *O. jollyorum*  
10 for this clade, which also encompassed Asian Type IV *vulgaris* individuals.  
11 Subsequently, this clade was renamed *O. sinensis*, which was recently redescribed  
12 and distinguished morphologically from *O. vulgaris* s. s. (the former having shorter  
13 arms with fewer suckers; Gleadall, 2016). Although, individuals from Asia and the  
14 Kermadec Islands are currently understood to comprise a single species, the  
15 substantial geographic distance between these two geographic regions warrants  
16 further investigation into their species-level diversity.

17 Vidal *et al.*, (2010) compared the morphology and chromatophore patterns of *O.*  
18 *vulgaris* paralarvae from the eastern North Atlantic (Galicia, Spain; *O. vulgaris* s. s.)  
19 and the western South Atlantic (southern Brazil; *O. vulgaris* Type II), noting  
20 considerable differences in chromatophore numbers. These differences support the  
21 hypothesis that *O. vulgaris* Type II is distinct from *O. vulgaris* s. s. Phylogenetic  
22 analyses and differences in adult morphology in the present study strongly support this  
23 hypothesis, showing that individuals from southern Brazil form a monophyletic clade,  
24 distinct from *O. vulgaris* s. s. and *O. sinensis*.

25 Superficial morphological similarity among species in the *O. vulgaris* species-complex  
26 had resulted in the assumption that *O. vulgaris* is a single cosmopolitan species.  
27 Despite estimates of 3-15 million years divergence between Australasian/Asian taxa  
28 (Amor *et al.*, 2014) and 19-41 million years divergence between *O. insularis* and other  
29 members of the *O. vulgaris* species-group (Amor *et al.*, 2015), principal component  
30 plots show that the morphology of these taxa is relatively conservative. Within the *O.*  
31 *vulgaris* species-complex, the clades considered to be different at the species level  
32 have allopatric distributions, so the selective pressures for strong phenotypic  
33 adaptations through interspecific competition may have been low. Differentiation in  
34 morphological traits is often most extreme where closely related species occur in



1 sympatry (Brown & Wilson, 1956), as a means to reduce resource overlap and to limit  
2 interspecific competition, allowing otherwise directly competing taxa to co-exist. Such  
3 'ecological character displacement' appears to be a common strategy among closely  
4 related taxa and has been documented in a number of plant, reptile, mammal, bird, fish  
5 and snail taxa (Dayan & Simberloff, 2005). One exception within the *O. vulgaris*  
6 species-group is the parapatric distribution of *O. vulgaris* Type II (sub-tropical southern  
7 Brazil) and *O. insularis* (mid-Atlantic islands and tropical northern Brazil). Although  
8 these two taxa are relatively distantly related, they are very similar in morphology,  
9 which may represent a unique opportunity to investigate the extent of this phenomenon  
10 within the *O. vulgaris* species complex. The closely related sibling pair of ocellate  
11 species in southern California (*O. bimaculatus* Verrill, 1883, and *O. bimaculoides*  
12 Pickford & McConnaughey, 1949) have used a different strategy to maintain sympatry:  
13 *O. bimaculoides* undergoes direct benthic development, while *O. bimaculatus*  
14 undergoes the meroplanktonic paralarval development typical of this group of  
15 octopuses (Pickford & McConnaughey, 1949).

16 The sexual traits of male individuals were found to be important characters for  
17 morphology-based discrimination of species in the *O. vulgaris* species-complex,  
18 confirming the utility of male sexual traits in cephalopod systematics in line with similar  
19 findings associated with other animal groups that sexual traits are more variable than  
20 non-sexual traits (Pomiankowski & Moller, 1995), and are often the only reliable way to  
21 identify closely related species (Arnqvist, 1998). However, a study of a different  
22 octopus genus, *Pareledone*, found that morphological traits (including three sexual  
23 traits) were unsuccessful in distinguishing among species, although they were well-  
24 defined traits characteristic of *Pareledone* at the generic level (Allcock *et al.*, 2008).

25 Amor *et al.*, (2014) used 17 morphological characters (five of which were sexual traits)  
26 to distinguish *O. tetricus* (from New Zealand and the eastern coast of Australia) and *O.*  
27 *cf. tetricus* (western Australia). HASC was found to be the primary source of variation  
28 between the two species, with significantly greater values for *O. cf. tetricus*. The utility  
29 of HASC has been demonstrated previously for distinguishing among octopus species  
30 (Toll, 1988; Gleadall, 2016). Among 12 species, Toll (1988) reported intraspecific  
31 HASC values to be relatively fixed. In contrast, the present study found HASC values  
32 for *O. vulgaris* s.s. correlated significantly for sampling localities of similar latitude.  
33 Individuals from the Mediterranean (France) and the eastern North Atlantic (Spain) had  
34 overlapping but significantly differing HASC values (144-168 and 156-183,

1 respectively). Mauritanian specimens were found to have significantly lower HASC  
2 values (114-150) than those for both France and Spain. The significant differences in  
3 HASC values reported within *O. vulgaris* s. s. are considered to represent population-  
4 level differences. Alternatively, since specimens from Mauritania display minimal  
5 overlap in this character compared with those from France and Spain, this may indicate  
6 the development of further species-level diversity within *O. vulgaris* s.s. as currently  
7 recognized (cf. also the findings of Cabranes *et al.*,(2008)). Voight (2012) cited wide  
8 variation in HASC as a potential problem for species-level inferences, concluding that  
9 variation in sucker numbers of  $\leq 15\%$  between potential species should be interpreted  
10 with caution. Concerning HASC values among Australasian members of the *O. vulgaris*  
11 species-group, those of western Australian *O. cf. tetricus* are almost 40% greater than  
12 those for eastern Australian *O. tetricus*. Such a wide range in HASC values within *O.*  
13 *vulgaris* s. s. therefore suggests the need for caution in basing species within this  
14 group on discrimination between HASC values.

15 The discriminatory power of female based morphological analyses was weaker than  
16 that for males. In the complete and reduced trait datasets, more morphological traits  
17 are available for male reproductive characters and these traits were found to be  
18 important in distinguishing among molecular sequence-based phylogenetic clades on  
19 the basis of morphology. The most significant female traits for distinguishing among  
20 species were non-sexual. Sexual traits, particularly the hectocotylus, are also important  
21 distinguishing taxonomic characters for decabrachian cephalopods, including the  
22 Idiosepiidae Appellöf, 1898 (Norman & Lu, 1997; von Byern & Klepal, 2010),  
23 Loliginidae Lesueur, 1821 (Brakoniecki, 1996), Ommastrephidae Steenstrup, 1857  
24 (O'Dor & Lipinski, 1998) and Sepiolidae Leach, 1817 (Bello, 1995). In contrast to body  
25 size and shape traits (which are likely to be less phenotypically and genetically variable  
26 between species), sexual traits are often exaggerated and diverse among close  
27 relatives (Pomiankowski & Moller, 1995), making them ideal candidates for  
28 distinguishing among species. While sexual traits were the primary source of  
29 morphological variation in the present study, non-sexual traits for both male and female  
30 morphology were highly successful in distinguishing among sampling localities of *O.*  
31 *vulgaris* s. s. (Galicia, France and Mauritania).

32 The need for better species resolution within the family Octopodidae is particularly  
33 important in view of the growing global exploitation of octopuses as a commercial  
34 fisheries resource (Norman & Finn, 2014). Global production of octopuses exceeds

1 350,000 tonnes with a total export value of US\$1.07 billion, surpassing the catch and  
2 value of many fisheries for finfish (FAO, 2012). A major limitation of the global catch  
3 statistics reported by the FAO is the poor resolution of octopus species, with only five  
4 (*O. vulgaris*, *O. maya*, *Eledone cirrhosa*, *Eledone moschata* and *Enteroctopus dofleini*)  
5 of the estimated 100 species of commercially harvested octopus listed in global  
6 statistics (Norman & Finn, 2014). As the majority of octopus fisheries world-wide are in  
7 decline (Norman & Finn, 2014), this low species resolution highlights the requirement  
8 for more accurate species identification in order to develop more sustainable octopod  
9 fisheries practices. Octopuses being exported globally under the name *O. vulgaris* are  
10 of high market value and profile (Norman *et al.*, 2014a), with those of northwestern  
11 Africa the largest single-species octopus fishery in the world (FAO, 2012). Aquaculture  
12 and captive growing of wild caught juveniles are receiving increasing profile and  
13 funding, particularly in China, Japan, Mexico and Spain. Differences among  
14 geographical areas in hatchling features and paralarvae viability (Iglesias *et al.*, 2007;  
15 Iglesias *et al.*, 2014) may also be linked to taxonomic differences. The findings  
16 presented here support the hypothesis that multiple *O. vulgaris*-like species are  
17 currently being incorrectly treated under a single species name. Our findings therefore  
18 have significant implications for the naming, marketing, value, documentation and  
19 potentially conservation of commercially harvested members of this species complex  
20 throughout their ranges.

21

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11

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18



- 1 Table 1: Sample data for octopus species analysed in the present study. Sample type refers to the type of data used: whole = whole animals, tissue = tissue samples or data = existing data from the literature.

Species/Type	Location	Institution	Sample Type	Reference
<i>O. vulgaris</i> s. s.	Banyuls-sur-Mer, France	Santa Barbara Museum of Natural History	Data	
<i>O. vulgaris</i> s. s.	Galicia, Spain	Consejo Superior de Investigaciones Científicas (CSIC), Vigo	Whole/Tissue	
<i>O. vulgaris</i> s. s.	Mauritania	Instituto Español de Oceanografía (IEO), Tenerife	Whole/Tissue	
<i>O. sinensis</i>	China	Fisheries College, Ocean University of China, Qingdao	Whole/Tissue	Reid and Wilson (2015)
<i>O. sinensis</i>	Keelung / Da si, Taiwan	National Taiwan Ocean University, Keelung	Whole/Tissue	Reid and Wilson (2015)
<i>O. sinensis</i>	Kermadec Islands, New Zealand	Australian Museum, Sydney	Whole/Tissue	Reid and Wilson (2015)
<i>O. sinensis</i>	Kyushu / Sendai, Japan	Tohoku University, Sendai	Whole/Tissue	
Type II (Brazil)	Pontal do Paraná, Brazil	Universidade Federal do Paraná (UFPR)	Whole	
<i>O. insularis</i>	Rio Grande do Norte/Brazil	Universidade Federal do Rio Grande do Norte (UFRN)	Whole	
<i>O. insularis</i>	Saint Peter and Saint Paul Archipelago, Brazil	Universidade Federal do Rio Grande do Norte (UFRN)	Whole	
<i>O. insularis</i>	Trindade Island, Brazil	Universidade Federal do Rio Grande do Norte (UFRN)	Whole	
<i>O. mimus</i>	Tocapilla / Pisagua, Chile	Consejo Superior de Investigaciones Científicas (CSIC), Vigo	Data	Guerra <i>et al.</i> (1999)
<i>O. tetricus</i>	New South Wales, Australia	Museum Victoria	Whole/Tissue	
<i>O. tetricus</i>	Tasmania, Australia	Museum Victoria	Tissue	
<i>O. cf. tetricus</i>	Western Australia, Australia	Fisheries and Marine Research Laboratories, Western Australia Museum Victoria	Whole/Tissue	

1 Table 2: Pairwise comparisons of male *Octopus vulgaris* species-group and *O.*  
 2 *insularis* individuals based on 25 morphological traits. Lower left diagonal represents  
 3 PERMANOVA results with significant differences ( $p < 0.05$ ) highlighted in bold. Upper  
 4 right diagonal represents results of SIMPER analyses showing traits that contribute  
 5 most to variation between groups. SIMPER results are also shown in bold if  
 6 corresponding PERMANOVA showed a significant difference. Far right column  
 7 represents the percentage of individuals assigned to their *a priori* group via Canonical  
 8 Analysis of Principal Components (CAP) analysis (see Table S4 for full CAP analysis  
 9 table).

	<i>O. vulgaris</i> s. s.	<i>O. sinensis</i>	Type II (Brazil)	<i>O. insularis</i>	<i>O. tetricus</i>	<i>O. cf. tetricus</i>	Correct (%)
<i>O. vulgaris</i> s. s.	-	<b>GLL/ALR4</b>	<b>SDe</b>	<b>SCR3/ALR 3</b>	<b>SCL3/SDn</b>	SCL3/DL	84.2
<i>O. sinensis</i>	<b>0.003</b>	-	<b>GLL/GLR</b>	<b>TOL/GLL</b>	<b>SDn</b>	<b>SCL3/TOL</b>	75.0
Type II (Brazil)	<b>0.011</b>	<b>0.001</b>	-	<b>DL/SCR3</b>	<b>SCL3/AW</b>	<b>SCL3/DL</b>	81.8
<i>O. insularis</i>	<b>0.002</b>	<b>0.001</b>	<b>0.001</b>	-	<b>SDe</b>	<b>ALR3</b>	66.7
<i>O. tetricus</i>	<b>0.009</b>	<b>0.001</b>	<b>0.001</b>	<b>0.001</b>	-	<b>SCL3/DL</b>	80.0
<i>O. cf. tetricus</i>	0.095	<b>0.001</b>	<b>0.01</b>	<b>0.001</b>	<b>0.012</b>	-	100

10

11 Table 3: Pairwise comparisons of female *Octopus vulgaris* species-group and *O.*  
 12 *insularis* individuals based on 20 morphological traits. Lower left diagonal represents  
 13 PERMANOVA results with significant differences ( $p < 0.05$ ) highlighted in bold. Upper  
 14 right diagonal represents results of SIMPER analyses showing traits that contribute  
 15 most to variation between groups. SIMPER results are also shown in bold if  
 16 corresponding PERMANOVA showed a significant difference. Asterisks represent  
 17 pairwise comparisons effected by significant within clade variation. Far right column  
 18 represents the percentage of individuals assigned to their *a priori* group via CAP  
 19 analysis (see Table S6 for full CAP analysis table).

	<i>O. vulgaris</i> s. s.	<i>O. sinensis</i>	Type II (Brazil)	<i>O. insularis</i>	<i>O. tetricus</i>	<i>O. cf. tetricus</i>	Correct (%)
<i>O. vulgaris</i> s. s.	-	<b>ALL3/SDn</b>	<b>AW</b>	<b>SCR/L3*</b>	SCR3/HW	SCL/R3	76.2
<i>O. sinensis</i>	<b>0.004</b>	-	<b>GLL/AW</b>	<b>SCL3</b>	SCR3/FL	HW	50
Type II (Brazil)	<b>0.01</b>	<b>0.001</b>	-	<b>SCR/L3</b>	<b>SCR3/AW</b>	<b>AW</b>	71.4
<i>O. insularis</i>	<b>0.001*</b>	<b>0.001</b>	<b>0.003</b>	-	<b>SCL3/FL</b>	<b>ALL1/3</b>	100
<i>O. tetricus</i>	0.053	0.119	<b>0.039</b>	<b>0.004</b>	-	HW	75
<i>O. cf. tetricus</i>	0.181	0.05	<b>0.041</b>	<b>0.012</b>	0.114	-	50

20

1 Table 4: Pairwise comparisons of male *Octopus vulgaris* sensu stricto individuals  
 2 based on 12 morphological traits. Lower left diagonal represents PERMANOVA results  
 3 with significant differences ( $p < 0.05$ ) highlighted in bold. Upper right diagonal  
 4 represents results of SIMPER analyses showing traits that contribute most to variation  
 5 between groups. SIMPER results are also shown in bold if corresponding  
 6 PERMANOVA showed a significant difference. Far right column represents the  
 7 percentage of individuals assigned to their *a priori* group via CAP analysis (see Table  
 8 S5 for full CAP analysis table).

	Galicia	Mediterranean	Mauritania	Correct (%)
Galicia	-	ALR3/SCL3	ALR3	80
Mediterranean	<b>p=0.004</b>	-	<b>FFL/LL</b>	88.9
Mauritania	<b>p=0.001</b>	<b>p=0.003</b>	-	100

9

10 Table 5: Pairwise comparisons of the sampling locations of female *Octopus vulgaris*  
 11 sensu stricto individuals based on eight morphological traits. Lower left diagonal  
 12 represents PERMANOVA results with significant differences ( $p < 0.05$ ) highlighted in  
 13 bold. Upper right diagonal represents results of SIMPER analyses showing traits that  
 14 contribute most to variation between groups. SIMPER results are also shown in bold if  
 15 corresponding PERMANOVA showed a significant difference. Far right column  
 16 represents the percentage of individuals assigned to their *a priori* group via CAP  
 17 analysis (see Table S7 for full CAP analysis table).

	Galicia	Mauritania	Mediterranean	Correct (%)
Galicia	-	SCL3/HW	FFL/FL	90
Mauritania	<b>p=0.001</b>	-	<b>FFL/SCL3</b>	100
Mediterranean	<b>p=0.002</b>	<b>p=0.001</b>	-	100

18

19

1 Fig. 1: Sampling localities (triangles) for whole animals/tissue samples of members of  
2 the *Octopus vulgaris* species-group and close relatives. Distributions of *O. vulgaris*  
3 sensu stricto and species 'Types' are shaded in dark grey (Norman et al., 2014a).  
4 Distributions of non-*vulgaris* species are represented by dashed lines.. Externally  
5 sourced data (Banyuls-sur-Mer, France; Table 1) is represented by a circle.

6

7 Fig. 2: Bayesian topology depicting the relationships among members of the *Octopus*  
8 *vulgaris* species-group and close relatives *O. mimus* and *O. insularis*. Analyses are  
9 based on partial sequence of the mitochondrial *COI* gene, showing BI posterior  
10 probabilities above and ML bootstrap values below major nodes. Outgroup is *O.*  
11 *cyanea*. Node labels represent geographic locations represented. Clade number is also  
12 shown (C1-11). *Octopus vulgaris* 'Types' refer to; Mediterranean/NE Atlantic (*O.*  
13 *vulgaris* s. s.), southern Brazil (Type II), South Africa (Type III) and *O. sinensis*  
14 (Norman et al., 2014a). Haplotype characters in parentheses correspond to individuals  
15 in Table S1.

16

17 Fig. 3: Principal Component biplot of male (a) and female (b) *Octopus vulgaris* species-  
18 group and *O. insularis* individuals grouped by *COI* based phylogenetic clade. Analysis  
19 is based upon 25 and 20 morphological traits respectively.

20

21 Fig. 4: Principal Component biplot 27 *Octopus vulgaris* sensu stricto males (a) and  
22 females (b), grouped by locality. Analysis is based on 12 and 8 morphological traits  
23 respectively.