Science in China Series C: Life Sciences



www.scichina.com life.scichina.com www.springerlink.com

# Toward a general evaluation model for soil respiration (GEMSR)

ZHOU GuangSheng<sup>†</sup>, JIA BingRui, HAN GuangXuan & ZHOU Li

Key Laboratory of Vegetation and Environmental Change, Institute of Botany, Chinese Academy of Sciences, Beijing 100093, China

Soil respiration is an important component of terrestrial carbon budget. Its accurate evaluation is essential to the study of terrestrial carbon source/sink. Studies on soil respiration at present mostly focus on the temporal variations and the controlling factors of soil respiration, but its spatial variations and controlling factors draw less attention. Moreover, the evaluation models for soil respiration at present include only the effects of water and heat factors, while the biological and soil factors controlling soil respiration and their interactions with water and heat factors have not been considered yet. These models are not able to accurately evaluate soil respiration in different vegetation/terrestrial ecosystems at different temporal and spatial scales. Thus, a general evaluation model for soil respiration (GEMSR) including the interacting meteorological (water and heat factors), soil nutrient and biological factors is suggested in this paper, and the basic procedure developing GEMSR and the research tasks of soil respiration in the future are also discussed.

soil respiration, controlling factors, general evaluation model

Accurately evaluating global carbon budget is a key not only for estimating atmospheric CO<sub>2</sub> concentration and forecasting climate change in the future, but also for implementing the responsibilities from United Nations Framework Convention on Climate Change (UNFCCC) and Kyoto Protocol. Therefore, the international joint programme, Global Carbon Project (GCP), was sponsored by International Geosphere-Biosphere Program (IGBP), International Human Dimensions Program on Global Environment Change (IHDP) and World Climate Research Program (WCRP) in 2001. The scientific aim of GCP is to accurately evaluate the temporal and spatial distributions of global carbon sources and sinks and their trends in the future.

There are various terrestrial ecosystems in China.  $CO_2$  emission of China is secondary in the world at present, only lower than the amount of the United States of America (USA). So how to strengthen the management of terrestrial ecosystems for reducing its carbon emission or increasing its carbon sink becomes an urgent problem. To find the answer, we should correctly understand the

effects of climate change and human activities on the processes and mechanisms of terrestrial carbon cycle and its interaction with environment, and accurately evaluate terrestrial carbon budget.

Soil respiration is a major flux between atmosphere and land, mainly including microbial and root respirations. Soil respiration is estimated to be 68-100 Gt C/a, being the secondary flux in the global carbon dioxide exchange, higher than net primary productivity (NPP, 50-60 Gt C/a) and lower than gross primary productivity (GPP, 100-120 Gt C/a)<sup>[1]</sup>. Studies on soil respiration and its related controlling factors will help further understand the terrestrial carbon cycle<sup>[2]</sup> and develop a general evaluation model for soil respiration (GEMSR).

Considering the importance of soil respiration in the global carbon cycle, much work has been done on the

Received April 24, 2007; accepted November 15, 2007

doi: 10.1007/s11427-008-0030-z

<sup>&</sup>lt;sup>†</sup>Corresponding author (email: gszhou@ibcas.ac.cn)

Supported by the National Natural Science Foundation of China (Grant No. 40625015 and 30770413) and the Knowledge Innovation Program of the Chinese Academy of Sciences (Grant No. KZCX2-YW-432)

observation methods, involved processes and influencing mechanisms of soil respiration, as well as quantitative evaluation of soil respiration, but no GEMSR has been developed for accurately evaluating the carbon sources and sinks of terrestrial ecosystems. Here we discuss the possibility developing GEMSR based on recent findings and development in soil respiration observation and its controlling factors.

#### 1 Soil respiration measurements

Soil respiration has been measured with different methods, mainly including alkali absorption<sup>[3]</sup>, infrared gas analysis (IRGA)<sup>[4]</sup>, gas chromatography<sup>[5]</sup>, atmospheric composition monitoring method (measurements of CO<sub>2</sub> and  $\delta^{13}CO_2$  or  $O_2/N_2)^{[6]}$ , and eddy covariance<sup>[7]</sup>. However, these methods are different in sampling times and sampling areas, which increase the difficulties in comparing and synthetically analyzing soil respiration at different temporal and spatial scales. The sampling time is on the diurnal scale for alkali absorption method and on the half hour and even longer for the others. The measurement area is a circle of 25 cm in diameter for alkali absorption method, a square of 50 cm×50 cm or 100 cm×100 cm for IRGA and gas chromatography, and even larger (e.g., ecosystem scale) for atmospheric composition monitoring method and eddy covariance, by which the footprint of soil respiration depends on the meteorological factors and the land surface characteristics. Thus, different methods should be selected carefully based on soil respirations at different temporal and spatial scales, in order to accurately describe the soil respiration rates and their controlling factors<sup>[8]</sup>.

#### 2 Controlling factors of soil respiration

Soil respiration is the maximum flux in terrestrial carbon exchange except for plant canopy photosynthesis. For accurate evaluation and prediction of carbon dioxide exchange between atmosphere and land, it is necessary to further understand the processes and controlling factors of soil respiration at different temporal and spatial scales<sup>[2]</sup>. Soil respiration originates mainly from biotic metabolism in soil. Thus, the factors affecting biotic activities would influence soil respiration rate, including climate factors, soil factors, plants and litter fall, etc.

#### 2.1 Moisture and temperature related factors

Moisture and temperature are usually taken as the main

factors controlling soil respiration. Soil respiration rate increases with increasing temperature<sup>[4,9]</sup>. When physiological temperature exceeds a certain threshold, the enzyme activities related to respiration in root and microorganism would reduce<sup>[4]</sup>, resulting in a decline in the sensitivity of soil respiration to temperature. Meanwhile, long-term stresses at a high temperature would also increase the diffusional resistance of cellular membrane to oxygen and decrease soil respiration rate by the inhibition of plant growth<sup>[10]</sup>.

The increase of precipitation (or soil water content) would generally accelerate soil respiration rate<sup>[11]</sup>. When soil water content exceeds a certain threshold, the high precipitation (or soil water content) would increase the diffusional resistance of CO<sub>2</sub> in soil and decrease soil respiration rate<sup>[12–13]</sup>. Moreover, soil respiration rate will decrease with increasing drought stress, which could inhibit root growth<sup>[14]</sup> and ion uptake<sup>[15]</sup>, and reduce maintenance consumption<sup>[16]</sup> and transportation of photosynthetic production<sup>[17]</sup>, etc.

Usually, soil respiration rate increases with increasing temperature. However, it would be inhibited by the limit of water under the conditions of higher temperature in arid and semiarid regions<sup>[11]</sup>. Furthermore, drought stress would result in decreasing root respiration rate more significantly in the high temperature zone than that in the low temperature zone<sup>[18]</sup>.

#### 2.2 Biotic factors

The effects of biotic factors on soil respiration rate include the respirations from roots and microorganisms. Root respiration rate is influenced by the root-to-shoot ratios and chemical compositions of plant species. Root respiration rate fluctuates seasonally due to the differences in quantity and quality of root secretion and the activities of rhizosphere microorganisms resulting from the seasonal changes in the biomass and the allocation of plant photosynthetic products<sup>[19]</sup>. The activities of soil microorganisms depend on the soil organic matter inputted from plant aboveground biomass and root<sup>[20]</sup>. Aboveground biomass of terrestrial ecosystems is correlated positively with the activities, quantity and biomass of soil microorganisms and enzymes<sup>[3]</sup>. The seasonal pattern of soil respiration rate could be usually expressed as a one-humped curve, with the maximum value appearing in late spring or early summer and the minimum in winter<sup>[21-23]</sup></sup>. Microbial respiration rate has a similar seasonal dynamics<sup>[24]</sup>, and its contribution to the total soil respiration rate changes with seasons<sup>[25]</sup>. Meanwhile, soil respiration rate is also controlled by aboveground biomass. For example, the removal shoots of spring wheat and soybean reduced root respiration rate<sup>[26]</sup>.

#### 2.3 Soil nutrients

Soil nutrients influence root growth and soil organic matter, which control soil respiration rate. The study on the effects of soil nitrogen and phosphorus on root respiration rate of rice seedlings indicates that soil respiration rate increases with increasing concentrations of soil nitrogen and phosphorus<sup>[27]</sup>. However, when the concentrations of soil nitrogen and phosphorus exceed 4.28 and 0.2 mmol/L, respectively, root respiration rate decreases with the increasing concentrations of soil nitrogen and phosphorus because of the inhibition of photosynthesis.

Microbial respiration rate is correlated with the quantity and the composition of soil organic matter (SOM)<sup>[28]</sup>. Generally, SOM could be divided into active, slow and passive pools based on the decomposition rate, and 80% of the microbial respiration rate comes from the active pool and 20% from the slow pool<sup>[29]</sup>.

Besides the above-mentioned factors, soil respiration is also affected by soil pH value<sup>[30]</sup>, atmospheric CO<sub>2</sub> concentration<sup>[31]</sup>, grazing<sup>[32–33]</sup>, deforest<sup>[34]</sup> and fertilization<sup>[35–37]</sup>, which could indirectly influence soil respiration by changing moisture, temperature, biotic factors or soil nutrients.

#### 3 Quantitative evaluation of soil respiration

In recent years, many studies on soil respiration have been done to evaluate carbon sources or sinks at the regional and global scales. A lot of soil respiration models have been established based on temperature, moisture or temperature-moisture interaction (Tables 1-3). Relationship between soil respiration and temperature could be expressed by linear, quadratic, power, exponential and Arrhenius models (Table 1). Among them, an exponential model is most commonly used. Soil respiration rate could be fitted better with the exponential model at low temperature, but worse at high tempera- ture<sup>[25,38]</sup>. Relationship between soil respiration and soil water content could also be expressed by linear, quad- ratic, cubic, logarithmic and exponential models (Table 2). Table 3 lists the relationships between soil respiration and temperature-moisture interaction, including linear, exponential, combination of exponential and power or Arrhenius models.

Table 1 Relationships between soil respiration and temperature

Equation	Example	Vegetation	Ref.
Linear	$F (g \text{ C} \cdot \text{m}^{-2} \cdot \text{yr}^{-1}) = 265.9 + 27.7T, R^2 = 0.83$	Peat lands	[39]
	<i>T</i> : mean annual air temperature ( $^{\circ}$ C)		
	$F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 57.626 - 3.544\text{T}, R^2 = 0.99$	Stipa baicalensis meadow steppe	[40]
	<i>T</i> : soil temperature at 10 cm depth ( $^{\circ}$ C)		
Quadratic	$F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 89.78 + 1.54T + 5T^2, R^2 = 0.83$	Tundra	[41]
	<i>T</i> : mean soil temperature at the depths of 0, 5, 10 and 20 cm ( $^{\circ}C$ )		
Power	$\ln F(\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = -1.66 + 2.20 \ln (T + 10), R^2 = 0.89$	Tall grass prairie	[42]
	<i>T</i> : soil temperature at 5 cm depth ( $^{\circ}$ C)		
	Farm: $F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = 1.66 \times 10^{-8} (T + 26.5)^{4.19}, R^2 = 0.90$	Farm and forest	[4]
	Forest: $F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = 2.41 \times 10^{-8} (T+13.4)^{4.34}, R^2 = 0.86$		
	<i>T</i> : mean soil temperature at the depths of 5 and 30 cm ( $^{\circ}$ C)		
Exponential	$F (\text{mg C} \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 21.13 \text{e}^{0.1371T}, R^2 = 0.80$	Mixed hardwood forest	[43]
	<i>T</i> : soil temperature at 10 cm depth ( $^{\circ}$ C)		
	$F (g CO_2 \cdot m^{-2} \cdot h^{-1}) = 0.14 e^{0.113T}, R^2 = 0.75$	Mixed hardwood forest	[44]
	<i>T</i> : soil temperature at 10 cm depth ( $^{\circ}$ C)		
	$F (\mu \text{mol CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = 0.375 \text{e}^{0.066T}, R^2 = 0.46$	Peat lands	[39]
	<i>T</i> : air temperature (°C)		
	$\ln F (g \operatorname{CO}_2 \cdot \mathrm{m}^{-2} \cdot \mathrm{d}^{-1}) = 7.069 + 0.133T - 0.002T^2, R^2 = 0.66$	Winter wheat	[45]
	<i>T</i> : soil temperature at 10 cm depth ( $^{\circ}$ C)		
Arrhenius	$F \ (\mu \text{mol CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = a \ \exp(-308.56/(T-227.13)), R^2 = 0.79$	Different ecosystems	[46]
	T: air temperature (K); a: ecosystem-dependent variable		

 Table 2
 Relationships between soil respiration and soil water content

Equation	Example	Vegetation	Ref.
Linear	$W < 0.12 \text{ cm}^3 \cdot \text{cm}^{-3}$ , $F(\text{mg C} \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 2852W - 128$ , $R^2 = 0.48$	Mixed hardwood forest	[43]
	$W > 0.12 \text{ cm}^3 \cdot \text{cm}^{-3}$ , $F(\text{mg C} \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = -198W + 201$ , $R^2 = 0.22$		
	<i>W</i> : volumetric water content at $0-15$ cm depth (cm <sup>3</sup> ·cm <sup>-3</sup> )		
	$F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 58.15 \text{W} - 105.88, R^2 = 0.85$	Leymus chinensis steppe	[9]
	W: volumetric water content at $0-10$ cm depth (%)		
	$F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 87.94 \text{W} - 642.66, R^2 = 0.85$	Stipa baicalensis meadow steppe	[40]
	W: gravimetric water content at $10-20$ cm depth (%)		
Quadratic	$F (\mu \text{mol CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = -7487.7 + 34365W - 39391W^2, R^2 = 0.40$	Tropical forest	[47]
	<i>W</i> : volumetric water content at $0-100$ cm depth (%)		
Cubic	Forest: $F (g \text{ C} \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 1.90 W^3 + 0.14, R^2 = 0.30$	Forest and pasture	[48]
	Pasture: $F(g \text{ C} \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 3.46 W^3 + 0.09, R^2 = 0.54$		
	<i>W</i> : volumetric water content at $0-30$ cm depth (cm <sup>3</sup> ·cm <sup>-3</sup> )		
Logarithmic	Forest: $F$ (g C·m <sup>-2</sup> ·h <sup>-1</sup> )=-0.043 lg (- $\psi$ )+0.16, $R^2$ =0.31	Forest and pasture	[48]
	Pasture: $F(g \text{ C} \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = -0.047 \text{lg} (-\psi) + 0.19, R^2 = 0.62$		
	$\Psi$ : matric potential at 0–30 cm depth (MPa)		
	$F (g \text{ C} \cdot \text{m}^{-2} \cdot \text{d}^{-1}) = 3.467 \text{ lg} W = 2.053, R^2 = 0.92$	Stipa grandis steppe	[3]
	W: gravimetric water content at $0-20$ cm depth (%)		
Exponential	$T \le 10^{\circ}$ C, there was no significant relationship with $W_{\rm S}$ or $W_{\rm L}$	Eucalyptus Pauciflora forest	[11]
	$T \ge 10^{\circ}$ C, ln F (mg CO <sub>2</sub> ·m <sup>-2</sup> ·h <sup>-1</sup> )=-0.019W <sub>S</sub> +5.31, R <sup>2</sup> =0.71		
	$\ln F(\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = -0.005 W_{\text{L}} + 5.76, R^2 = 0.76$		
	<i>T</i> : air temperature (°C); $W_{\rm S}$ : gravimetric water content at 0–20 cm depth (%);		
	$W_{\rm L}$ : litter water content (%)		

## 4 Toward a general soil respiration evaluation model

Up to now, many studies have been done on soil respiration and its controlling factors (e.g., moisture, temperature, soil nutrients, and biological mechanisms, etc.), and lots of soil respiration evaluation models have been established based on moisture, temperature or moisture-temperature interaction. However, these relationships are based on only the average soil respiration rate and its controlling factors in the same vegetation type or ecosystem, and they do not include the effects of biotic factors (e.g., root biomass), soil characteristics (e.g., soil nutrients) and spatial heterogeneity of soil water contents. Meanwhile, the differences of soil respiration rates and their main controlling factors at different temporal scales are not taken into account from the current different measurement methods, e.g., instantaneous soil respiration rate measured by LI-6400-09, half hour values of soil respiration by gas chromatography and average values of 24 hours by alkali absorption method. The interactive processes and mechanisms among moisture, temperature, soil nutrients and biological factors are still unclear at present. Furthermore, no GEMSR including the interacting moisture, temperature, biologic processes, and soil nutrients has been established at different spatial and temporal scales. The reasons are listed as follows.

(i) Spatial heterogeneity of soil respiration and its controlling factors

Most of the current studies on soil respiration focus on a certain vegetation type or ecosystem. Spatial heterogeneity of soil respiration is avoided by stochastic samplings and calculating the mean value. The controlling factors mainly include temperature and moisture, but biotic factors, especially root biomass and soil nutrients are seldom included. Meanwhile, the standardized sampling depth and observation identifier are also not defined for temperatures (e.g., air temperature, soil temperature, etc.) and moistures (e.g., gravimetric water content, volumetric water content, water potential, and water level, etc.). Therefore, most of evaluation models for soil respiration are developed in a uniform vegetation type or ecosystem based on limited observations of moisture and temperature. These models are difficult to be applied in other ecosystems or regions because of ignoring the effects of spatial heterogeneity on soil respiration rate and the statistical relationships.

(ii) Short-term soil respiration measurement and singularity of vegetation type

Currently, most of the soil respiration studies are a certain short-term soil respiration measurement, ignor-

Table 3	Relationships	between so	il respiration	and tem	perature-	moisture	interact	tion
---------	---------------	------------	----------------	---------	-----------	----------	----------	------

Equation	Example	Vegetation	Ref.
Linear	$F (g \text{ C} \cdot \text{m}^{-2} \cdot \text{d}^{-1}) = 0.88 + 0.013 TW, R^2 = 0.83$	Arid shrub steppe	[49]
	<i>T</i> : soil temperature at 10 cm depth (°C); <i>W</i> : volumetric water content at 10 cm depth ( $\text{cm}^3$ ·cm <sup>-3</sup> )		
	$F (g \text{ CO}_2 \cdot \text{m}^{-2} \cdot \text{d}^{-1}) = 0.715 + 0.210T + 0.285P_{1-3} + 0.083P_{4-7}, R^2 = 0.64$	Pine forest	[50]
	<i>T</i> : air temperature (°C), $P_{1-3}$ , $P_{4-7}$ : precipitation of 3 days and $4-7$ days before experiment, respectively (cm)		
	W < 7.5%, there was no significant relationship with $W$ ;	Leymus chinensis steppe	[51]
	$W > 7.5\%$ , $F(\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = -147.7 + 5.1W + 6.0T + 1.2WT$ , $R^2 = 0.81$		
	<i>T</i> : soil temperature at 5 cm depth (°C);		
	<i>W</i> : gravimetric water content at $0-10$ cm depth (%)	_	
Exponential	$\ln F(\text{g CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = -2.63 + 0.11T + 0.04MI, R^2 = 0.90$	Temperate forest	[52]
	<i>T</i> : mean soil temperature at the depths of 5 and 15 cm ( $^{\circ}$ C); <i>MI</i> : moisture index		
	ln F (g C·m <sup>-2</sup> ·d <sup>-1</sup> )=0.087T+0.025W-0.264, $R^2$ =0.69	Riparian buffers and crop fields	[53]
	<i>T</i> : soil temperature at 5 cm depth (°C); <i>W</i> : gravimetric water content at $0-5$ cm depth (%)		
	ln $F(\text{mg C}\cdot\text{m}^{-2}\cdot\text{d}^{-1})=5.86+0.01W+0.04T+0.01WT$ , $R^2=0.71$	Leymus chinensis steppe	[25]
	<i>T</i> : air temperature (°C); <i>W</i> : gravimetric water content at $0-20$ cm depth (%)		
Exponential-power	$W < 19\%$ , $F(\mu mol CO_2 \cdot m^{-2} \cdot s^{-1}) = 0.33 W^{0.69} e^{0.042T}$ , $R^2 = 0.76$	Ponderosa pine	[54]
	$W > 19\%$ , $F(\mu \text{mol CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = 26.17 W^{-0.82} \text{e}^{0.047\text{T}}$ , $R^2 = 0.95$		
	<i>T</i> : soil temperature at 10 cm depth ( $^{\circ}$ C);		
	<i>W</i> : volumetric water content at $0-30$ cm depth (%)		
	$F (\text{mg C} \cdot \text{m}^{-2} \cdot \text{d}^{-1}) = 5911.65 W^{0.91} \text{e}^{0.04T}, R^2 = 0.86$	Degraded steppe	[55]
	<i>T</i> : air temperature (°C); <i>W</i> : gravimetric water content at $10-20$ cm depth (%)		
	$F (\text{mg CO}_2 \cdot \text{m}^{-2} \cdot \text{h}^{-1}) = 1.97 \times 10^{-5} \text{e}^{0.045T} (W - 21.42) (58.54 - W)^{4.46}, R^2 = 0.96$	Deciduous broadleaved forest	[56]
	<i>T</i> : soil temperature at 10 cm depth ( $^{\circ}$ C);		
	<i>W</i> : volumetric water content at $0-15$ cm depth (%)		
Exponential-Arrhenius	$F(\mu \text{mol } \text{CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}) = C \times e^{(-E/RT)} \times e^{Swt}$	Tundra	[57]
	C: constant( $\mu$ mol·m <sup>-2</sup> ·s <sup>-1</sup> ); <i>T</i> : soil temperature at 1 cm depth (K); <i>R</i> : gas constant (8.31 J·mol <sup>-1</sup> ·K <sup>-1</sup> ); <i>E</i> : apparent activation energy (J·mol <sup>-1</sup> ); <i>Swt</i> = <i>AW</i> /( <i>W</i> + <i>B</i> ); <i>W</i> : depth to water table below soil surface(cm); <i>A</i> and <i>B</i> : fitted parameters		

ing the difference at temporal scales. For example, the differences of the controlling factors at different time scales during one day and during a period of every ten days. Meanwhile, many studies are limited in a specific vegetation type or ecosystem. It is difficult to include some factors changing insignificantly with time (e.g., soil organic matter). Thus, those insufficiencies make soil respiration models only suitable at specific spatial and temporal scales, but not suitable for other ecosystem types or regions. Therefore, it is urgent to develop an integrated measurement for soil respiration and its controlling factors in various vegetation types and ecosystems, in order to acquire these parameters of GEMSR including the interacting moisture, temperature, soil nutrient factors, and biologic characteristics.

(iii) The estimation of soil respiration is mostly based on the statistical correlations with moisture and temperature, and few soil nutrients and biotic factors are included. Those kinds of soil respiration models are devoid of generality, and only suitable for each special vegetation type or site. The spatial heterogeneity and the change of biotic factors are not reflected in the soil respiration models at present, and it is difficult to get upscaled to the regional or global scales. Meanwhile, no sampling standardizations have been made for temperatures (e.g., mean annual air temperature, mean diurnal air temperature and soil temperature at 10 cm depth, etc.), moistures (e.g., gravimetric water content, volumetric water content, water potential, water level, etc.), depths (e.g., 0-5, 0-10, 0-15, 0-20 cm, etc.), and sampling times (e.g., instantaneous soil respiration rate by LI-6400-09, half hour values by gas chromatography and average values of 24 h by alkali absorption method). It was difficult to compare these soil respiration models.

Therefore, to clarify the processes and mechanisms of soil respiration responding to its main controlling factors

and to establish GEMSR, we must use the similar instruments with high time resolution to observe the spatial heterogeneity and controlling factors of soil respiration in various vegetation types or ecosystems. Thus, the long-term comprehensive observation data can be obtained including soil respiration rate, moisture, temperature, soil nutrient and biological factors.

Currently, LI-6400-09 (Li-cor, Lincoln, NE, USA) has been extensively applied to measure instantaneous soil respiration rate based on infrared gas analysis method. The instrument is portable and quick in measuring (usually a measurement takes only 5-10 min), and the outputs include temperature, relative humidity and soil temperature, etc. The long-term soil respiration rate, moisture, temperature, soil nutrients and biologic characteristics could also be obtained with a set of instruments, including LI-6400-09, soil moisture measurement system (Hydrosense, Campell scientific Australia pty. Ltd.), and the corresponding measurements of above- and below-ground biomass and soil properties.

We have observed a lot of soil respiration rates with Li-6400-09 in typical grassland in Inner Mongolia (4 years)<sup>[51,58-59]</sup>, maize field in Jinzhou (2 years)<sup>[60-61]</sup>,</sup></sup> reed wetland in Panjin (2 years)<sup>[62-63]</sup>, different forestland use practices of broad-leaved Korean pine forest in Changbai Mountain (3 years) including primary broadleaved Korean pine forest, secondary mixed of Betula platyphlla and Populus davidiana, cropland, larch plantation forest, clear-cut broad-leaved Korean pine forest<sup>[64–67]</sup> since 2002. Partitioning soil respiration into root and microbial respirations, soil respiration measurements at Leymus chinensis populations with five planting densities (0, 30, 60, 90 and 120 plants/ $0.25 \text{ m}^2$ ) were made in 2003 in a greenhouse at Institute of Botany, the Chinese Academy of Sciences<sup>[69]</sup>. Many data were obtained from these studies, including soil respiration rate, air temperature and humidity, soil temperature and moisture at different soil depths, above- and below- ground biomass and soil physicochemical properties, etc. Those studies in maize field and typical grassland showed that there were linear relationships between soil respiration rate and root biomass under the conditions of constant moisture, temperature and soil nutrients<sup>[60,68-69]</sup>. The statistical soil respiration models were established based on the main controlling factors of soil moisture and temperature in the broad-leaved Korean pine forest in

Changbai Mountain and in fenced and grazing typical grasslands<sup>[51,59,64]</sup>. There are no studies on the effects of soil nutrients on soil respiration rate and integrated analysis of moisture, temperature, soil nutrients, and biological characteristics yet. However, these studies provide basal data and methods for further understanding the controlling mechanisms of the main factors and establishing GEMSR including the interacting moisture, temperature, soil nutrient, soil nutrient and biological factors.

To establish GEMSR, we must understand how to acquire the necessary observation data. We should choose various vegetation types to study the effects of soil nutrients (mainly soil carbon and soil nitrogen) on soil respiration rate. Long-term observation data of soil respiration and the related controlling factors are essential for the development of soil respiration evaluation model for soil respiration. To quantitatively describe the interactive effects of moisture, temperature, soil nutrient and biological factors (mainly soil carbon and nitrogen) on soil respiration rate, the effects of these factors at different temporal scales need to be partitioned. Firstly, the effects of biotic factors on soil respiration rate could be included based on lots of soil respiration measurements in different sampling plots at a certain time with similar environment conditions including moisture, temperature and soil nutrients. Secondly, the gradually interactive effects of biotic factors with temperature, moisture and soil nutrients on soil respiration rate will be analyzed, disclosing the processes and mechanisms of the interactive effects of belowground biomass, temperature, moisture and soil nutrients on soil respiration rate. Therefore, the cropland ecosystem (e.g. maize field) as a good experiment field with uniform soil nutrients and smaller changing among years, and others as assistant ecosystems, GEMSR including soil nutrients could be established.

Above all, GEMSR including the interacting moisture, temperature, soil nutrient and biological factors would be established as the following procedures (Figure 1):

(i) Based on the data in many measurement plots at the same time during the growing season in a certain ecosystem, keeping soil nutrients, temperature and moisture in the plots as the same, the processes and mechanisms of root biomass controlling soil respiration rate will be analyzed at different growth phases. The relationship of soil respiration rate with root biomass will be established.



Figure 1 Procedures for developing GEMSR including the interacting moisture, temperature, soil nutrient and biological factors.

(ii) Based on the data in many measurement plots at different time scales in one day (e.g., different hours) during growing season in a certain ecosystem, maintaining soil nutrients and moisture in the plots as the same, the interactive effects of root biomass and temperature on soil respiration rate will be measured.

(iii) Based on the diurnal data in many measurement plots during the whole year in a certain ecosystem, the interactive effects of root biomass, temperature and moisture on soil respiration rate will be included. The dynamic soil respiration model including the effects of moisture, temperature and biotic factors would be established.

(iv) Based on the observation data in different time periods during an entire year in various ecosystems with different soil nutrients, the analysis would focus on the effects of soil nutrients on soil respiration rate. The dynamic soil respiration model including the interacting effects of moisture, temperature, soil nutrient, and biological factors would be obtained.

To develop GEMSR, the important studies in the future should be emphasized as follows:

(i) The responding processes and mechanisms of soil respiration to the interactions of moisture, temperature

and biotic factors. We should focus on the spatial heterogeneity of soil respiration and its mainly controlling factors in different ecosystems, and analyze the processes and mechanisms of the effects of the interactions of root biomass, temperature and moisture on soil respiration.

(ii) The responding processes and mechanisms of soil respiration to the interactions of moisture, temperature, soil nutrients and biological characteristics. We should focus on the differences of the processes and mechanisms of the effects from the interactions of moisture, temperature and biotic factors on soil respiration in different ecosystems, and clarify the processes and mechanisms of the effects of soil nutrients (mainly soil carbon and nitrogen) on soil respiration rate, and eventually disclose the interactive effects of moisture, temperature, root biomass and soil nutrients on soil respiration rate.

(iii) Study on GEMSR including the interacting moisture, temperature, soil nutrients and biological characteristics. We should focus on the effects of the interaction of different factors on the processes and mechanisms of soil respiration in different ecosystems, and clarify the processes and mechanisms of soil respiration in different land use practices, and eventually establish GEMSR. GEMSR could calculate total soil respiration including microbial and root respirations, and could also partition into two main components. The model could also evaluate soil respiration in different ecosystems and land use practices, and it could be coupled into the dynamic terres-

- Zhou G S, ed. Global carbon cycle. Beijing: China Meteorological Press, 2003
- 2 Craine J M, Wedin D A, Chapin F S. Predominance of ecophysiological controls on soil CO<sub>2</sub> flux in a Minnesota grassland. Plant Soil, 1999, 207: 77-86
- 3 Chen S Q, Cui X Y, Zhou G S, et al. Study on the CO<sub>2</sub>-release rate of soil respiration and litter decomposition in *Stipa grandis* Steppe in Xilin river basin, Inner Mongolia. Acta Botan Sin, 1999, 41: 645– 650
- 4 Fang C, Moncrieff J B. The dependence of soil CO<sub>2</sub> efflux on temperature. Soil Biol Biochem, 2001, 33: 155-165
- 5 Dong Y, Zhang S, Qi Y, et al. Fluxes of CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub> from a typical temperate grassland in Inner Mongolia and its daily variation. Chin Sci Bull, 2000, 45(17): 1590-1594
- 6 Keeling R F, Piper S C, Heimann M. Global and hemispheric CO<sub>2</sub> sinks deduced from changes in atmospheric O<sub>2</sub> concentration. Nature, 1996, 381: 218-221
- 7 Wofsy S C, Goulden M L, Munger J M, et al. Net exchange of CO<sub>2</sub> in a mid-latitude forest. Science, 1993, 260: 1314–1317
- 8 Zhou G S, Wang Y H, Jiang Y L, et al. Conversion of terrestrial ecosystems and carbon cycling. Acta Phytoecol Sin, 2002, 26(2): 250-254
- 9 Jia B R, Zhou G S, Wang F Y, et al. A comparative study on soil respiration between grazing and fenced typical *Leymus chinensis* steppe, Inner Mongolia. Chin J Appl Ecol, 2004, 15(9): 1611–1615
- 10 Atkin O K, Edwards E J, Loveys B R. Response of root respiration to changes in temperature and its relevance to global warming. New Phytologist, 2000, 147: 141-154
- 11 Keith H, Jacobsen K L, Raison R J. Effects of soil phosphorus availability, temperature and moisture on soil respiration in *Eucalyptus pauciflora* forest. Plant Soil, 1997, 190: 127-141
- 12 Cavelier J, Penuela M C. Soil respiration in the cloud forest and dry deciduous forest of Serrania of Macuira, Colombia. Biotropica, 1990, 22(4): 346-352
- 13 Rochette P, Desjardins R L, Pattey E. Spatial and temporal variability of soil respiration in agricultural fields. Can J Soil Sci, 1991, 71: 189-196
- 14 Espeleta J F, Eissenstat D M, Graham J H. Citrus root responses to localized drying soil: A new approach to studying mycorrhizal effects on the roots of mature trees. Plant Soil, 1998, 206(1): 1-10
- 15 Eissenstat D M, Whaley E L, Volder A, et al. Recovery of citrus surface roots following prolonged exposure to dry soil. J Exp Bot, 1999, 50: 1845-1854
- 16 Bouma T J, Bryla D R, Li Y, et al. Is maintenance respiration in roots a constant? In: Stokes A, eds. The Supporting Roots of Trees and Woody Plants: Form, Function and Physiology. Developments in Plant and Soil Sciences Series. Netherlands: Kluwer Academic Publishers, 2000. 87: 391-396

trial ecosystem productivity model to improve the accuracy of evaluating terrestrial ecosystem carbon budget.

### We appreciate Assistant Professor, Dr. Xiongwen Chen from Alabama A & M University for his great contribution in language editing.

- 17 Yang G P, Wang S T. Effects of osmotic stress on the respiration of wheat roots. Acta Phytophysiol Sin, 1989, 15(2): 179-183
- 18 Bryla D R, Bouma T J, Hartmond U, et al. Influence of temperature and soil drying on respiration of individual roots in citrus: Integrating greenhouse observations into a predictive model for the field. Plant Cell Environ, 2001, 24: 781-790
- 19 Amthor J S. Plant respiratory responses to the environment and their effects on the carbon balance. In: Wilkinson R E, eds. Plant-Environment Interactions. New York: Marcel-Dekker, 1994. 501-554
- 20 Zhao J B, Yuan D X, Ma Z W. A study on the CO<sub>2</sub> release amount and its change from the soil in the Xi'An area. Carsologica Sinica, 2000, 19(4):309-313
- 21 Wang W, Guo J X. Contribution of CO<sub>2</sub> emission from soil respiration and from litter decomposition in *Leymus chinensis* community in Northeast Songnen Grassland. Acta Ecol Sin, 2002, 22(5):655-660
- 22 Chen G S, Yang YC, Wang X G, et al. Root respiration in a natural forest and two plantations in subtropical China: Seasonal dynamics and controlling factors. Acta Ecol Sin, 2005, 25(8): 1941–1947
- 23 Liu Y, Han S J, Hu Y L, et al. Effects of soil temperature and humidity on soil respiration rate under *Pinus sylvestriformis* forest. Chin J Appl Ecol, 2005,16(9): 1581-1585
- 24 Zhang C B, Yang J C. Preliminary study on respiration rate of soil microorganism under different vegetations on *Aneurolepidium chinensis* grassland of northeast China. Chin J Appl Ecol, 1996, 7(3): 293-298
- 25 Li L H, Wang Q B, Bai Y F, et al. Soil respiration of a *Leymus chinensis* grassland stand in the XiLin river basin as affected by over-grazing and climate. Acta Phytoecol Sin, 2000, 24(6): 680-686
- 26 Liu H S, Li F M, Jia Y. Effects of shoot removal and soil water content on root respiration of spring wheat and soybean. Environ Exp Bot, 2006, 56(1): 28–35
- 27 Cui K H, He Z C, Zhang J Y, et al. Effects of nitrogen and phosphorus in mimic wastewater on root respiration of rice seedlings. J Wuhan Bot Res, 1996, 14(4): 323-328
- 28 Townsend A R, Vitousek P M, Desmarais D J, et al. Soil carbon pool structure and temperature sensitivity inferred using CO<sub>2</sub> and <sup>13</sup>CO<sub>2</sub> incubation fluxes from five Hawaiian soils. Biogeochemistry, 1997, 38: 1–17
- 29 Schimel D S, Braswell B H, Holland E A, et al. Climatic, edaphic and biotic controls over storage and turnover of carbon in soils. Global Biogeochem Cycles, 1994, 8(3): 279-293
- 30 Bouma T J, Bryla D R. On the assessment of root and soil respiration for soils of different textures: Interactions with soil moisture content and soil CO<sub>2</sub>. Plant Soil, 2000, 227: 215-221
- 31 Berntson G M, Bazzaz F A. Belowground positive and negative feedbacks on CO<sub>2</sub> growth enhancement. Plant Soil, 1996, 187: 119-131
- 32 Northup B K, Brown J R, Holt J A. Grazing impacts on the spatial distribution of soil microbial biomass around tussock grasses in a

ZHOU GuangSheng et al. Sci China Ser C-Life Sci | Mar. 2008 | vol. 51 | no. 3 | 254-262

tropical grassland. Appl Soil Ecol, 1999, 13: 259-270

- 33 Reeder J D, Schuman G E. Influence of livestock grazing on C sequestration in semi-arid mixed-grass and short-grass rangelands. Environ Pollut, 2002, 116: 457-463
- 34 Yang Y S, Chen G S, Wang X G, et al. Effect of clear-cutting on soil respiration of Chinese fir plantation. Acta Pedol Sin, 2005, 42(4): 584-590
- 35 Meng L, Ding W X, Cai Z C, et al. Storage of soil organic C and soil respiration as effected by long-term quantitative fertilization. Adv Earth Sci, 2005, 20(6): 687-692
- 36 Yang L F, Cai Z C. Soil respiration during maize growth period affected by N application rates. Acta Pedol Sin, 2005, 42(1): 9–15
- Zhuge Y P, Zhang X D, Liu Q. Effect of long-term fertilization on respiration process of mollisols. Chin J Soil Sci, 2005, 36(3): 391-394
- 38 Chang Z Q, Shi Z M, Feng Q. Effect of temperature in different communities on soil respiration in Qilian Mountains. Chin J Agrometeorol, 2005, 26(2): 85–89
- 39 Chimner R A. Soil respiration rates of tropical peatlands in Micronesia and Hawaii. Wetlands, 2004, 24(1): 51-56
- 40 Dong Y S, Qi Y C, Liu J Y, et al. Variation characteristics of soil respiration fluxes in four types of grassland communities under different precipitation intensity. Chin Sci Bull, 2005, 50: 583–591
- 41 Peterson K M, Billings W D. Carbon dioxide flux from tundra soils and vegetation as related to temperature at Barrow, Alaska. Am Midland Naturalist, 1975, 94: 88-98
- 42 Kucera C L, Kirkham D R. Soil respiration studies in tallgrass prairie in Missouri. Ecology, 1971, 52(5): 912-915
- 43 Davidson E A, Belk E, Boone R D. Soil water content and temperature as independent or confounded factors controlling soil respiration in a temperate mixed hardwood forest. Global Change Biol, 1998, 4: 217-227
- Kang S, Doh S, Lee D S, et al. Topographic and climatic controls on soil respiration in six temperate mixed-hardwood forest slopes, Korea. Global Change Biol, 2003, 9: 1427-1437
- 45 Buyanovsky G A, Wagner G H, Gantzer C J. Soil respiration in a winter wheat ecosystem. Soil Sci Soc Am J, 1986, 50: 338-344
- 46 Lloyd J, Taylor J A. On the temperature dependence of soil respiration. Funct Ecol, 1994, 8: 315-323
- 47 Sotta E D, Meir P, Malhi Y, et al. Soil CO<sub>2</sub> efflux in a tropical forest in the central Amazon. Global Change Biol, 2004, 10: 601–617
- 48 Davidson E A, Verchot L V, Cattanio J H, et al. Effects of soil water content on soil respiration in forests and cattle pastures of eastern Amazonia. Biogeochemistry, 2000, 48: 53-69
- 49 Wildung R E, Garland T R, Buschbom R L. The interdependent effect of soil temperature and water content on soil respiration rate and plant root decomposition in arid grassland soils. Soil Biol Biochem, 1975, 7: 373-378
- 50 Reinke J J, Adriano D C, McLeod K W. Effects of litter alteration on carbon dioxide evolution from a South Carolina pine forest floor. Soil Sci Soc Am J, 1981, 45: 620-623
- 51 Wang F Y, Zhou G S, Jia B R, et al. Effects of heat and water factors on soil respiration of restoring *Leymus Chinensis* Steppe in degraded land. Acta Phytoecol Sin, 2003, 27(5): 644-649

- 52 Reiners W A. Carbon dioxide evolution from the floor of three Minnesota Forests. Ecology, 1968, 49: 471-483
- 53 Tufekcioglu A, Raich J W, Isenhart T M, et al. Soil respiration within riparian buffers and adjacent crop fields. Plant Soil, 2001, 229: 117–124
- 54 Xu M, Qi Y. Soil-surface CO<sub>2</sub> efflux and its spatial and temporal variations in a young ponderosa pine plantation in northern California. Global Change Biol, 2001, 7: 667–677
- 55 Chen Q S, Li L H, Han X G, et al. Influence of temperature and soil moisture on soil respiration of a degraded steppe community in the Xilin river basin of Inner Mongolia. Acta Phytoecol Sin, 2003, 27: 202-209
- 56 Lee M S, Nakane K, Nakatsubo T, et al. Effects of rainfall events on soil CO<sub>2</sub> flux in a cool temperate deciduous broad-leaved forest. Ecol Res, 2002, 17: 401–409
- 57 Oberbauer S F, Gillespie C T, Cheng W, et al. Environmental effects on CO<sub>2</sub> efflux from riparian tundra in the northern foothills of the Brooks Range, Alaska, USA. Oecologia, 1992, 92: 568-577
- 58 Jia B R, Zhou G S, Wang F Y, et al. Affecting factors of soil microorganism and root respiration. Chin J Appl Ecol, 2005, 16(8): 1547– 1552
- 59 Jia B R, Zhou G S, Wang F Y, et al. Partitioning root and microbial contributions to soil respiration in *Leymus chinensis* populations. Soil Biol Biochem, 2006, 38: 653-660
- 60 Han G X, Zhou G S, Xu Z Z, et al. Biotic and abiotic factors controlling the spatial and temporal variation of soil respiration in an agricultural ecosystem. Soil Biol Biochem, 2007, 39: 418-425
- 61 Han G X, Zhou G S, Xu Z Z, et al. Responses of soil respiration to the coordinated effects of soil temperature and biotic factors in a maize field. Acta Phytoecol Sin, 2007, 31(3): 363-371
- 62 Xie Y B, Jia Q Y, Zhou L, et al. Soil respiration and its controlling factors at *Phragmites* communis wetland in Panjin. J Meteorol Environ, 2006, 22(4): 53-58
- 63 Lu G H, Zhou L, Zhao X L, et al. Vertical distribution of soil organic carbon and total nitrogen in reed wetland. Chin J Appl Ecol, 2006, 17(3): 384-389
- 64 Jiang Y L, Zhou G S, Zhao M, et al. Soil respiration in broad-leaved and korean pine forest ecosystems, Changbai Mountain, China. Acta Phytoecol Sin, 2005, 29(3): 411–414
- 65 Wand X, Zhou G S, Jiang Y L, et al. Comparison of soil respiration in broad-leaved korean pine forest and reclaimed cropland in Changbai Mountains, China. Acta Phytoecol Sin, 2006, 30(6): 887–893
- 66 Wang X, Zhou G S, Jiang Y L, et al. Soil respiration in natural mixed (*Betula platyphlla* and *Populus davidiana*) secondary and primary broad-leaved Korean pine forest. Acta Phytoecol Sin, 2007, 31(3): 348-354
- 67 Wang X, Zhou G S, Jiang Y L, et al. Soil respiration in a clear-cut broad-leaved Korean pine forest of Changbai Mountain. Acta Phytoecol Sin, 2007, 31(3): 355-362
- 68 Jia B R, Zhou G S, Wang Y H, et al. Effects of temperature and soil water content on soil respiration of degraded and restoring *Leymus chinensis* steppe, Inner Mongolia. J Arid Environ, 2006, 67: 60-76
- 69 Jia B R, Zhou G S, Yuan W P. Modeling and coupling of soil respiration and soil water content in fenced *Leymus chinensis* steppe, Inner Mongolia. Ecol Modelling, 2007, 201: 157–162