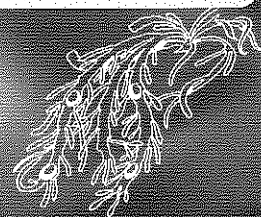
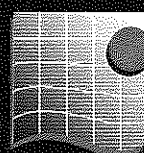


Strain selection in the edible brown seaweed *Alaria esculenta*: Genetic fingerprinting and hybridization studies under laboratory conditions

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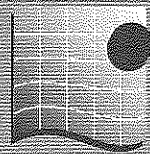
S. Krann & M.D. Guiry



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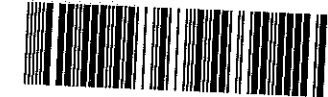
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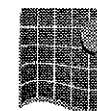
Strain selection in the edible brown seaweed *Alaria esculenta*: Genetic fingerprinting and hybridization studies under laboratory conditions

Contract No. IR.95.MR.011

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1. INTRODUCTION

The genus *Alaria* presently includes 12 species, 11 of which are located in the cold temperate North Pacific and only one is found in the North Atlantic (Widdowson, 1971). The North Atlantic species *Alaria esculenta* has two northern forms, *A. esculenta* forma *grandifolia* and forma *pylaii* (Lüning, 1990). The study presented here will concentrate on *Alaria esculenta*, the most common North Atlantic species.

1.1 Biogeography

The brown alga *Alaria esculenta*, which literally means 'edible wings', is a large brown seaweed belonging to the family *Alariaceae* of the order Laminariales (Kelp), and can reach a length up to 6 m (Guiry, 1997). Its short stipe gives rise to a long blade with a well-defined rib (Fig. 1). This seaweed forms the main canopy in exposed areas near or below the low-water mark to a depth of 35 m (Lüning, 1990). *Alaria esculenta* is present in the North Pacific and North Atlantic, where it is located North as far as the winter sea ice and as far south as the 16° C summer isotherm, in the European North Atlantic represented by the French coast of Brittany (Lüning, 1990); Fig. 2. Its absence in the southern North Sea and English Channel is due to high summer water temperatures of over 16° C, which it cannot survive (Munda & Lüning, 1977). The species is found all around the Irish coast, where rocky shores as a substratum are available (Sundene, 1962; Widdowson, 1971).

1.2 Utilisation

Alaria esculenta was used in the past in both Scotland and Ireland for human consumption and fodder. It was also gathered and spread on infertile land and used as fertiliser (Newton, 1931; Guiry and Hession, 1996; Guiry, 1997).

It is rich in sugars, proteins, vitamins and other trace metals and contains up to 42% alginic acid (Levring *et al.*, 1969; Indergaard & Minsaas, 1991; Lewallen & Lewallen, 1996). The species can be used for a variety of purposes from human consumption and alginate production to fodder and bodycare products (Guiry & Blunden, 1991; Guiry, 1997). Especially in North America *Alaria esculenta* and *Alaria marginata* are rapidly gaining popularity in the natural foods market (Lewallen & Lewallen, 1996). It also has potential as a foodstuff in aquaculture for herbivorous molluscs, e.g., abalone (Mai *et al.*, 1996).

Young *Alaria esculenta* (the only native species in Ireland) can be used as a substitute for *Undaria pinnatifida* (Wakame), a very popular seaweed in Asian countries with numerous applications and a yield over 300,000 t fresh weight per annum (Nisizawa *et al.*, 1987; Indergaard & Minsaas, 1991; Druehl, 1988; Yamanaka & Akiyama, 1993; Lewallen & Lewallen, 1996).

Experiments have shown that within 3 months a harvest of 9 t per hectare is possible (Kain & Dawes, 1987). *Alaria esculenta* is a fast-growing seaweed and can grow at up to 10 cm a day. Commercial potential for *Alaria esculenta* is considerable (Kain & Dawes, 1987; Kain, Holt & Dawes, 1990).

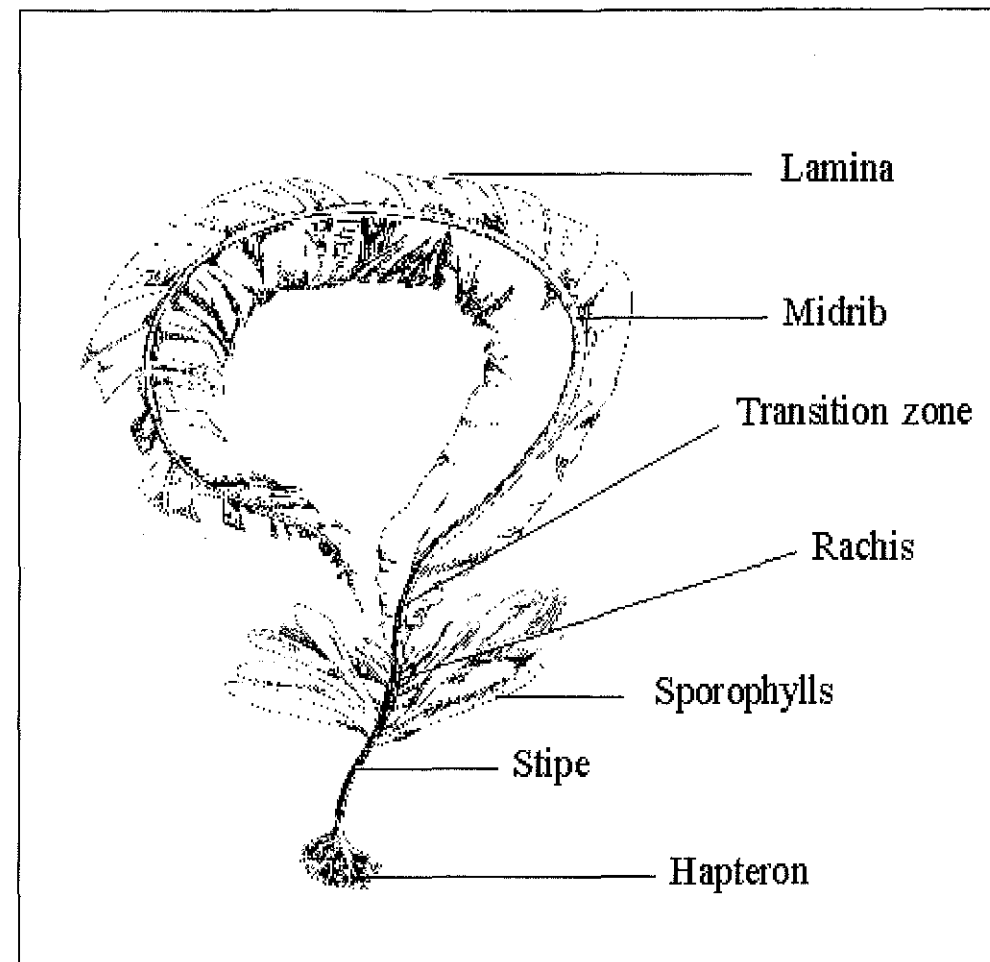


FIG. 1. HABIT OF *ALARIA ESCULENTA* SHOWING SOME MORPHOLOGICAL CHARACTERS.

1.3 Life history

Only during spring and in lesser amount during autumn two rows of ligulate sporophylls form in the upper parts of the stipe, the rachis (Widdowson, 1971). The sporophylls produce haploid spores by meiosis that germinate to form the haploid or gametophytic phase (male and female). Gametophytes produce the gametes (sperm and eggs) which fuse after fertilization to form a zygote. The zygote germinates to form diploid plantlets, the sporophytic phase. Thus the life history is that the large seaweed alternates with a microscopic filamentic phase (van den Hoek, 1996; Fig 3). *Alaria esculenta* is dioecious and has a heteromorphic diplohaplontic life history.

1.4 Hybridization experiments

Members of the Laminariaceae are able to cross inter- and intraspecific (tom Dieck, 1992; Egan *et al.*, 1990), and even intergeneric (Migita, 1984). Bolton *et al.* (1983)

produced hybrids between Atlantic and Pacific *Laminaria* species with different growth rates and morphological characteristics. They concluded that the ability to produce these characters appeared to be genetically determined. Lüning *et al.* (1978) also produced crosses between *Laminaria* species from both sides of the Atlantic. They suggested that blade width in hybrid offspring could be sex linked to the male gametophytes, which is in agreement with the results of a study by Egan *et al.* (1990).

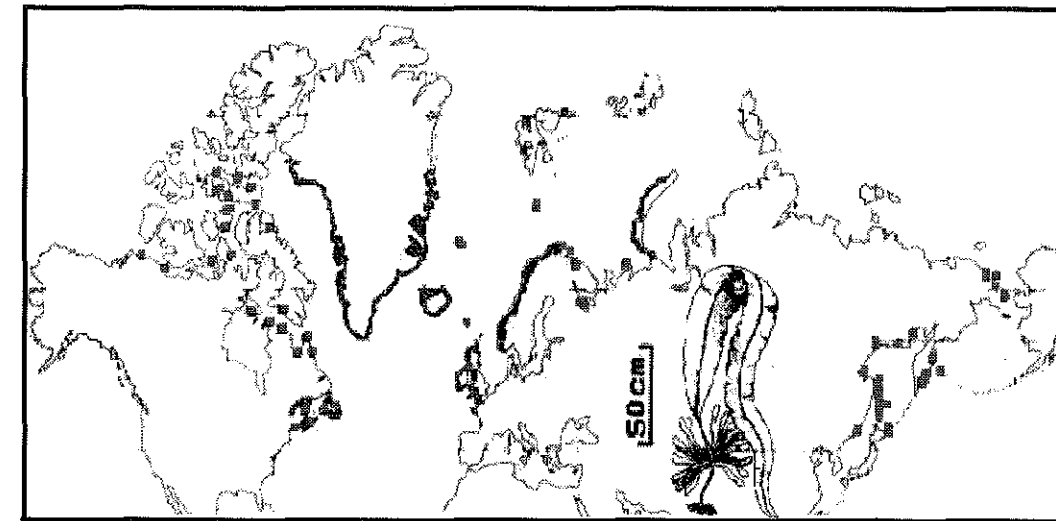


FIG. 2. THE DISTRIBUTION OF *ALARIA ESCULENTA* IN THE NORTH ATLANTIC AND THE NORTH PACIFIC OCEAN (AFTER LÜNING, 1990).

The temperature at which hybridization experiments are carried out plays an important role in the success of the crosses, as temperature will determine the production and release of gametes as well as the healthy growth of the sporophytes (tom Dieck, 1992). It can make a vast difference from which species the female gametophyte is provided as a successful cross may depend on this. Therefore it is important in hybridization studies to perform crosses and reciprocal crosses from every species (Egan *et al.*, 1990).

Lewis *et al.* (1986) produced interspecific hybrids of three *Macrocystis* species. They showed significantly different growth rates in different genetic groups. One of the self-crossed individuals showed the lowest growth rate. They suggested that the observed differences in growth rate between genetic groups was due to genetic adaptation to the conditions of the locality of growth of the parental plants.

Crossing experiments within the Laminariales are plagued with uncertainty because of the ability of female gametophytes to produce parthenogenic sporophytes. Parthenogenesis is a common feature of the members of the Laminariales (Nakahara & Nakamura, 1973; Lüning *et al.*, 1978; tom Dieck, 1992). Although most parthenogenic sporophytes have an abnormal morphology, become stunted or irregular and do not survive more than a few mm in length (Bolton *et al.*, 1983).

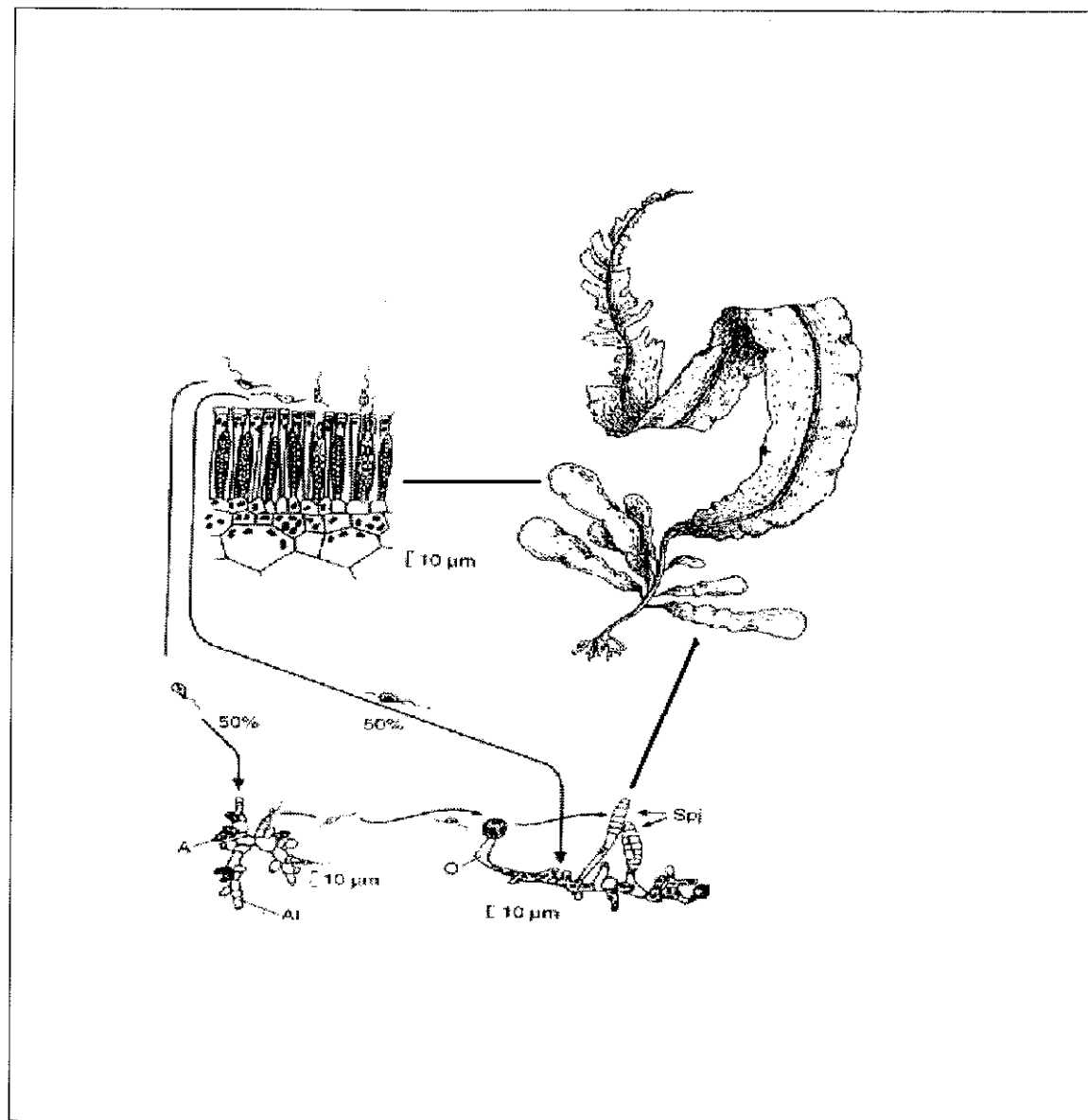


FIG. 3. LIFE CYCLE OF *ALARIA ESCULENTA*. THE SPOROPHYTE (A) PRODUCES SORI ON THE SPOROPHYLLS (B). SPORES (D) ARE PRODUCED IN THE SORI AFTER MEIOSIS IN UNILOCULAR SPORANGIA (C). SPORES (D) CONSISTING OF 50% MALE AND 50% FEMALE SETTLE AND GROW INTO MALE (E) AND FEMALE (F) GAMETOPHYTES RESPECTIVELY. MALE GAMETOPHYTES FORM SPERMS AND FERTILISE THE EGG FORMED ON THE OOGONIUM OF THE FEMALE GAMETOPHYTE. THE FERTILIZED EGG DEVELOPS INTO A ZYGOTE, WHICH DEVELOPS INTO YOUNG SPOROPHYTES (SPJ). AFTER VAN DEN HOEK *ET AL.*, (1996).

Hybridization experiments offer a means of assessing differentiation when correlated to specific environmental differences. However, interpretation of such experiments is not straightforward. Genetic differentiation may evolve without affecting compatibility and intersterility does not necessarily indicate accumulation of a large number of genetic differences (Innes, 1984).

Increased incompatibility is often associated with greater geographical separation (Rueness, 1973; Guiry and West, 1983) but there are numerous examples where genetic differentiation occurs among populations separated by only short distances (Lewis *et al.*, 1986; Coyer *et al.*, 1997). Hybridization experiments may add valuable information to hypotheses of phylogenetic relationships (Lüning, 1990).

In China, extensive studies on hybridization and inbreeding has led to stable and improved strains of *Laminaria japonica* with higher production and iodine contents. Large-scale production experiments showed that the new varieties yield up to 40% more biomass and iodine contents of 20-58% higher than the control plants. These new varieties are introduced to *Laminaria* farms and grown in large quantities in North China (Chaoyuan & Guangheng, 1987).

1.5 Population genetics

Morphology of lamina and stipe in *Alaria esculenta* can vary widely between specimens mainly, it seems, due to wave exposure, with broader lamina and longer stipes in more sheltered areas (Sundene, 1962; Widdowson, 1971; pers. observ.). Phenotypic plasticity is a common feature of members of the Laminariales (Norton *et al.*, 1982). Chapman (1974) found significant genetic differentiation between populations of *Laminaria* for stipe morphology. By contrast, in a study on the Pacific kelp *Costaria costata* no genetic differences were found between two morphologically distinct populations using restriction fragment length polymorphisms (RFLP) (Bhattacharya & Druehl, 1989). However, it is not known for the kelp *A. esculenta* if there is one variety in the Atlantic ocean (gene flow between populations, no founder effects) or more different varieties (no gene flow between populations). More varieties might indicate varieties with different optimal characteristics suitable for aquaculture.

RFLP is a common applied molecular tool to distinguish morphological similar populations (Hillis *et al.*, 1996). It has been used in numerous algal studies (e.g., Rice & Bird, 1990; Bhattacharya & Druehl, 1989; Bhattacharya *et al.*, 1991; Lehman and Manhart, 1997). To demonstrate possible genetic differences in *Alaria* species with the RFLP technique two non-coding regions in the DNA were amplified. These are the Rubisco spacer which separates the large and small subunits of the ribulose-1,5-biphosphate carboxylase/oxygenase genes and the nuclear ribosomal DNA (rDNA) internal transcribed spacer regions (ITS1 and ITS2). These non-coding spacers are considered to be more variable than coding genes, because they are under the least degree of functional constraint (Saunders and Druehl, 1993). These spacers have been shown to provide good resolution in examining relationships among and within populations of green algae (Kooistra, 1992; Bakker, 1992) and red algae (van Oppen *et al.*, 1995).

1.6 Kelp evolution and divergence

The outward morphological diversity of kelps suggests that kelps are an ancient assemblage, however, recent evidence suggest that kelp divergence started in the late

Miocene about (10-15 Ma) or even as late as late Pliocene (3-5 Ma) ago from a common ancestor (Estes & Steinberg, 1988; Saunders & Druehl, 1992). Stam *et al.* (1988) calculated that five *Laminaria* species diverged from a common ancestor 15-19 Ma ago most probably originating in the North Pacific and then invaded the North Atlantic after inundation of the Bering Land Bridge 3.5 Ma ago. This recent divergence event implies that North Atlantic *Laminaria* species probably have close relatives in the Pacific Ocean. Hybridisation experiments have shown this close relationship between Pacific and Atlantic *Laminaria* species (Bolton *et al.* 1983; tom Dieck, 1992). Stam *et al.* (1988) identified five Pacific *Laminaria* sister species for five Atlantic *Laminaria* species. Relationships between Pacific and Atlantic *Alaria* species are unknown.

OBJECTIVES

There were three main objectives for this study:

- 1) To isolate, hybridize and grow strains of *Alaria esculenta* and other *Alaria* species from the Atlantic and Pacific to develop varieties with optimal characteristics for aquaculture in Ireland.
- 2) To identify genetic variation among *A. esculenta* strains from the Atlantic Ocean using the Restriction Fragment Length Polymorphism (RFLP) technique.
- 3) To obtain phylogenetic information about the relationships between North Atlantic and North Pacific species of the genus *Alaria* by hybridization experiments and fingerprinting

2. MATERIALS AND METHODS

2.1 Cultivation and hybridization techniques

Five individual plants with mature sori at the sporophylls were sampled from the geographical locations given in Table 1. Male and female gametophyte cultures from locations marked with an asterisk were sent by the author mentioned.

Zoospores from mature sori were released using techniques described by South (1970) and Nakahara & Nakamura (1973). Seawater with zoospores was divided in sterile replidishes and kept under long-day conditions at 10° C and 20 μmol photons. m⁻² s⁻¹. After about 10 d zoospores germinated into gametophytes and were visible under a microscope. After 2 - 3 weeks the gametophytes were separated in male and female gametophytes. Within 2 - 3 months full male and female cultures were obtained from the geographical locations listed in Table 1. The gametophytes had been maintained in the vegetative state in red fluorescent light at an irradiance of 10 μmol photons. m⁻².s⁻¹ in glass dishes containing sterile enriched seawater (Von Stoch 0.25 strength; Guiry & Cunningham, 1984) which was changed every month. Gametophyte cultures were grown

under red light as this prevents the gametophytes from maturing and hence forming eggs and sperm. This was necessary to allow for conducting cross experiments (Bolton *et al.* 1983).

Crosses were made using individual male and female gametophytes previously isolated from spores released by parent sporophytes collected in the sites listed in Table 1. Cross experiments were carried out in two different ways:

- By placing a female gametophyte (about 10 cells long) of a species of *Alaria* with one male gametophyte from another species of *Alaria* and vice-versa in Replidishes.
- By gently grinding the male and female gametophytes of the cross of interest in a mortar to produce a suspension of male and female filaments of 1-10 cells long. The fragments were then poured in Replidishes or small 20 ml Petri-dishes.

The crosses were incubated at 10, 15, 20, and 25 °C, long-day conditions (16:8 h) and 30 μmol photons.m⁻².s⁻¹ white light to initiate the formation of eggs and sperm (Lüning 1990). As soon as sporophytes appeared they were measured with an binocular. After two months viable healthy sporophytes with a length of 2 cm were transferred to 20 ml universal containers on a shaker. Daylength conditions were changed to longday 13:11 h to mimic spring or autumn conditions and initiate fast growth of the developing sporophytes. The medium was changed weekly or fortnightly. Sporophytes outgrowing the containers were transferred to small tanks with an aeration system under the same conditions.

In addition, self crosses and isolated male and female gametophytes were incubated parallel to each crossing experiment to observe possible parthenogenesis or apogamy (Nakahara & Nakamura, 1973). The plants were examined and measured in length and width weekly in the raply dishes, fortnightly in the sterile containers and once or twice a month in the tank.

2.2 Relative Growth Rates

The relative growth rate of the self crosses and hybrid sporophytes of the different crosses was calculated using the formula for relative growth rate (RGR)

$$RGR = \frac{L_n(L_2) - L_n(L_1)}{t_2 - t_1}$$

Where L2 is the length at time t2 and L1 the length at time t1 in increments per day. The final length and width of the crosses were determined with image analysing equipment.

2.3 Detection of genetic variation

DNA extraction and purification

Plants from the sites listed in Table 1 were transported to the laboratory in a coolbox wrapped in plastics bags on cooling elements or sent in a similar way by mail. On

arrival, plants were cleaned and directly processed for DNA extraction or quick-frozen in liquid nitrogen and stored at -70° C.

TABLE 1. ORIGIN OF *ALARIA* GAMETOPHYTE CULTURES USED IN THE PRESENT INVESTIGATION.

Species	Code	sex	origin	Collector
<i>Alaria esculenta</i>	IRL	M & F	Ireland (Inis Oírr)	S. Kraan
<i>Alaria esculenta</i>	ACH	M & F	Achill Island Co. Mayo	S. Kraan
<i>Alaria esculenta</i>	CBH	M & F	Corbet Head Co. Down	S. Kraan
<i>Alaria esculenta</i>	SLH	M & F	Slea Head Co. Kerry	S. Kraan
<i>Alaria esculenta</i>	SJP	M & F	John's Point Co. Donegal	S. Kraan
<i>Alaria esculenta</i>	ICL	M & F	Iceland (Tjörnes)	K. Lüning*
<i>Alaria esculenta</i>	NOR	M & F	Norway (Ålesund)	J. Rueness *
<i>Alaria esculenta</i>	HFX	M & F	Canada (Halifax)	C. Bird *
<i>Alaria praelonga</i>	AP	M & F	Japan (Hokkaido)	M. Masuda*
<i>Alaria crassifolia</i>	AC	M & F	Japan (Hokkaido)	M. Masuda*
<i>Alaria tenuifolia</i>	AT	F	Canada (Vancouver)	L. Druehl
<i>Alaria nana</i>	AN	M & F	Canada (Vancouver)	L. Druehl
<i>Alaria marginata</i>	AM	M & F	Canada (Vancouver)	L. Druehl

DNA was extracted from ca. 1 g of blotted wet weight of healthy plant material or ca. 0.5 g of -70°C stored blade material. Extraction as follows: Grind material in liquid nitrogen, add powder to a 2 mL eppendorf tube with 2 µL β-mercapto-ethanol and 900 µL extraction buffer (2% CTAB (v/v), 4M NaCl, 0.5 M EDTA pH 8 and 1M TRIS-HCL pH 8). Incubate for 1 h under slow agitation, add 900 µL CIA (chloroform:isoamylalcohol, 24:1 v/v) and mix gently for 3 min. Centrifuge at 14,000 g at 10° C. Transfer upper aqueous phase to sterile new tube, repeat CIA extraction until the interphase has disappeared. Transfer upper phase to fresh sterile tube, precipitate polysaccharides by adding 0.1 vol 5M K-acetate, mix, 0.25 vol 96% Ethanol, mix, 1 vol CIA, mix, centrifuge as before and collect upper phase in a new tube. Add 1 vol cold (-

20° C) isopropanol, mix gently, and leave to settle for at least 1 h at 4° C. To precipitate the crude DNA, centrifuge as before for 30 min. Rinse pellet 3 times in 70% Ethanol, air dry and dissolve in 100 µL 0.1 x TE buffer. Add 10 units Dnase-free Rnase (Boehringer Mannheim) and incubate at 37° C for 30 min. Precipitate DNA as described above. Redissolve in 100 µL 0.1 x TE buffer. Average yields were 10-100 µg of high molecular weight DNA.

PCR Amplification

Double stranded amplifications were performed in a Hybaid Omn-E thermal cycler with an initial denaturation step of 95° C for 3 min followed by 35 cycles with the following temperature profile: 1 min 95° C, 2 min 55° C and 2 min 74° C and ended with one extension step of 72° C for 5 min. The reaction volume was 100 µL comprised of 10-100 ng genomic DNA, 50 µM of each dATP, dNTP, dCTP and dGTP, ca. 0.5 µM of each primer, 10 µL of 10x reaction buffer (Sigma), 6 µL 25 mM MgCl₂ (Sigma) and 2.5 units of Taq (Sigma). Primer pair P1 and G4 were used to amplify the region in the ribosomal cistron from position 1542 in the 18S across the ITS1, the 5.8S, ITS2 and to position 42 in the 26S (Saunders and Druehl, 1993). Primer pair s130s and 11105f were used to amplify a small part of the Large subunit across the Rubisco spacer into the small subunit, starting at position 1105 in the Large subunit to position 172 in the small subunit (Valetin and Zetche, 1990).

Amplifications were checked for correct length, purity and yield on 1.5% agarose TAE gels stained with EtBr according the methods of Sambrook et al. (1989). Excess primers, salts and nucleotides were removed from PCR products using the PCR purification kit according the manufacturer's instructions (Boeringer Mannheim). One restriction enzyme reaction could be performed with 15 µL of purified product.

Restriction endonuclease digestion of PCR fragments

Fifteen µL of the PCR amplified Rubisco spacer or rDNA spacers of all isolates listed in table 1 were completely digested with 2-4 units of restriction endonuclease for 4 h as follows: Fil 1.5 ml Eppendorf tube with 15 µL PCR product (cleaned), 3 µL 10x incubation buffer and 2 µL restriction enzyme (Boeringer Mannheim). Mix this 20 µL and spin down for 10 sec. The 8 enzymes used were RSA1, HindIII, CFO1, DRA1, XBA, Dpn1, PST1 and SMA Restriction fragments were run on 3% TAE agarose gels (1% agarose/ 2% MS agarose, Boeringer Mannheim) at 20 V for 20 h and stained afterwards with EtBr. The stained restriction fragments were visualized by UV-fluorescence, captured with a video camera and printed with a thermic printer. Fragment seize was determined according standard methods (Hillis et al. 1996) All digestions were performed twice so accurate calculation of the seize of the fragments was possible.

2.4 Data analysis

The data sets of relative growth rates, width, length and biomass showed a departure of a normal distribution and heteroscedasticity was pronounced. Therefore we applied the Kruskal-Wallis non-parametric analysis of variance (with tied ranks and unequal sample

seize) to examine the null hypothesis: similarity between RGR of Atlantic crosses = similarity in RGR between Atlantic with Pacific crosses = similarity between Pacific crosses. Multiple comparisons were made as a posteriori testing using Dunn's test if the null hypothesis was rejected (Zar, 1996).

Presence and absence of restriction fragments were scored manually from the thermic prints and a presence/absence matrix was constructed for all individuals. Pairwise similarities were calculated using Dice (Jacquard) coefficient (Sd) = two times the number of bands shared by two individuals, divided by the number of bands displayed by each individual. Sd does not allow for negative matches which might cause false similarities among those taxa sharing many negative matches (Coyer *et al.*, 1997). From the bandsharing data a pairwise distance matrix was generated where in the number 1 corresponds to two individuals being identical and the value 0 corresponds to two individuals that are entirely dissimilar. No further statistical analysis was performed on the restriction fragments because of the low amount of fragments generated.

3. RESULTS

3.1 Hybridization experiments

All successful crosses between Atlantic x Atlantic, Atlantic x Pacific and Pacific x Pacific *Alaria* species are listed in table 2 and 3. Successful crosses were produced at 10° C only. All the (self) crosses and reciprocal crosses between *Alaria esculenta* strains of Atlantic populations proved to be successful except for the Male Iceland x Female Halifax cross. This cross did not produce any viable healthy sporophytes. Although the products of some crosses were small, like the Iceland and Halifax crosses, they looked like perfect healthy sporophytes and were not parthenogenetic. Parthenogenetic sporophytes in the female parallel cultures were easily identified because of their round cell clump-like appearance without haptera. The Irish self cross, Irish x Norway crosses and female Irish x male Halifax cross produced reproductive structures (sporophylls) in culture.

Between crosses of Atlantic *A. esculenta* with other species of the Pacific only the *A. praelonga* x *A. esculenta* and reciprocal cross produced viable healthy sporophytes which formed sporophylls in culture. From *A. crassifolia* only male gametophytes hybridized with *A. esculenta*. The reciprocal cross produced small (± 1 cm long) stunted sporophytes which resembled the parthenogenetic sporophytes from the control. From *A. tenuifolia* only female gametophytes hybridized with male *A. esculenta* and produced sporophylls. The reciprocal cross was not tested because no male *A. tenuifolia* gametophytes were available. Crosses and reciprocal crosses between *A. nana* and *A. marginata* did not produce any sporophytes.

The control cultures with female or male gametophytes alone produced in some cases parthenogenetically or apogamous derived sporophytes. These sporophytes showed a high mortality rate and never grew >1 cm. It was therefore very easy to recognize healthy

and viable hybrids from parthenogenic or apogamous sporophytes. In some experiments the parthenogenic or apogamous sporophytes survived up to a year without growth development.

TABLE 2. HYBRIDIZATION WITHIN 4 NORTH ATLANTIC STRAINS OF *ALARIA ESCULENTA* AND THE DEVELOPMENT OF THE HYBRIDS INDICATED AS AGE, MORPHOLOGY AND MAXIMUM LENGTH THAT WAS ACHIEVED DURING CULTIVATION AT 10° C, 30 μ MOL PHOTONS.M⁻².S⁻¹ AND LD (13:11). (-) = NO DEVELOPMENT OF SPOROPHYTES. HAPTERA + = WELL DEVELOPED, +/- = HAPTERA OF HALF THE SPOROPHYTES IS WELL DEVELOPED, - = POOR HAPTERA DEVELOPMENT. SP = SPOROPHYLLS FORMED, NT = CROSSING NOT TESTED. FOR ORIGIN CODES SEE TABLE 1

Female	MALE			
	IRL	ICL	NOR	HFX
IRL	Normal, sp 43 cm, Haptera + 7 months	Small/broad 6.5 cm, Haptera - 6 months	Normal, 46 cm, Haptera +/- 6 months	Normal, sp 42 cm, Haptera + 6 months
ICL	Small/broad 8 cm, Haptera +/- 6 months	Nt	Small, 3 cm, Haptera +/- 16 months	- 12 months
NOR	Long/thin, sp 27 cm, Haptera +/- 6 months	Normal, sp 70 cm, Haptera + 9 months	Nt	Small, 21 cm, Haptera +/- 9 months
HFX	Normal/ Missing midribs, 13 cm, Haptera + 6 months	Small, 18 cm, Haptera + 9 months	Very small, 2.5 cm, Haptera - 9 months	Nt

TABLE 3. HYBRIDIZATION OF NORTH ATLANTIC *ALARIA ESCULENTA* WITH PACIFIC *ALARIA* SPECIES AND WITHIN PACIFIC SPECIES AND THE DEVELOPMENT OF THE HYBRIDS INDICATED AS MORPHOLOGY AND MAXIMUM LENGTH THAT WAS ACHIEVED DURING CULTIVATION AT 10 °C, 30 μ MOL PHOTONS.M⁻².S⁻¹ AND LD (13:11). (-) = NO DEVELOPMENT OF SPOROPHYTES. HAPTERA + = WELL DEVELOPED, +/- = HAPTERA OF HALF THE SPOROPHYTES IS WELL DEVELOPED, - = POOR HAPTERA DEVELOPMENT. SP = SPOROPHYLLS FORMED, NT = CROSSING NOT TESTED. FOR ORIGIN CODES SEE TABLE 1

Female	MALE				
	IRL	AP	AC	AN	AM
IRL	Normal, sp 43 cm, Haptera + 7 months	Normal, sp 21 cm, Haptera + 6 months	Normal 6 cm, Haptera - 7 months	Few elongated stipes 8 cm Haptera - 6 months	Few elongated stipes 8 cm Haptera - 7 months
AP	Normal, sp 11 cm, Haptera + 6 months	Normal/long, thin, 13 cm Haptera +/- 8 months	Clumps <1cm Haptera - 9 months	Clumps <1cm Haptera - 12 months	Clumps <1cm Haptera - 9 months
AC	Stunted <1cm Bleached Haptera - 7 months	Normal, sp 25 cm Haptera + 9 months	Nt	Stunted 1cm Bleached Haptera - 7 months	Clumps <1cm Haptera - 12 months
AN	Few elongated stipes 6 cm Haptera - 6 months	Normal 5 cm Haptera +/- 6 months	Normal 6 cm Haptera +/- 6 months	Nt	- 20 months
AM	Clumps <1cm Haptera - 14 months	Normal, sp 60 cm, Haptera + 10 months	Clumps <1cm Haptera - 12 months	- 20 months	Long, thin 22 cm Haptera + 8 months
AT	Normal, sp 61 cm, Haptera + 6 months	Normal, 12.5 cm, Haptera + 9 months	Stunted <1cm Haptera - 6 months	Stunted <1cm Haptera - 6 months	Normal 56 cm Haptera + 9 months

Within the Pacific crosses *A. praelonga* male gametophytes hybridized with all other Pacific *Alaria* species. *Alaria tenuifolia* female gametophytes hybridized with *A. praelonga* and *A. marginata* male gametophytes, the latter also producing sporophylls.

3.2 Relative growth rates

All hybrids and selfcrosses showed a similar growth pattern of rapid growth during the first 4 to 5 weeks and a slower growth during the rest of the period. In this report, these growth rates will be called "settlement relative growth rate" (RGRset) and "Relative growth rate" (RGR) respectively. The mean RGRset and RGR for length and width increments per day are presented in figure 4 to 7.

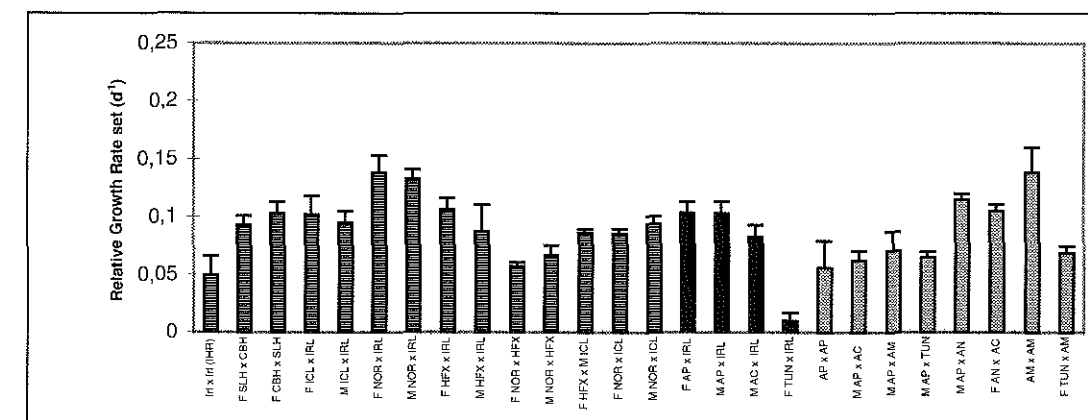


FIG. 4. AVERAGE SETTLEMENT RELATIVE GROWTH RATES FOR PLANT LENGTH (D⁻¹ WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE 1.

For Fig. 4 the Kruskal-Wallis test showed significant differences between crosses ($H=38.96535$, $X^2_{0.05,25}=37.652$). The null hypothesis was rejected; the settlement relative growth rates for length are different between crosses. Multiple comparisons are given in Appendix I. Significant differences in Appendix I are marked with patterns, vertical striped for Atlantic, black for Atlantic x Pacific and grey for Pacific.

The Irish self crosses and Irish x Norway crosses are in most cases significantly different from other crosses in length elongation.

For Fig. 5 the Kruskal-Wallis test showed significant differences between crosses ($H=82.47678$, $X^2_{0.05,25}=37.652$). The null hypothesis was again rejected. The relative growth rates for length are different between crosses. Multiple comparisons are presented in Appendix II. Significant differences in Appendix II are marked with patterns, horizontal striped for Atlantic, black for Atlantic x Pacific and grey for Pacific. The Irish x *tenuifolia*, Irish x Norway and *A. marginata* self cross are in most cases significantly different from the other crosses.

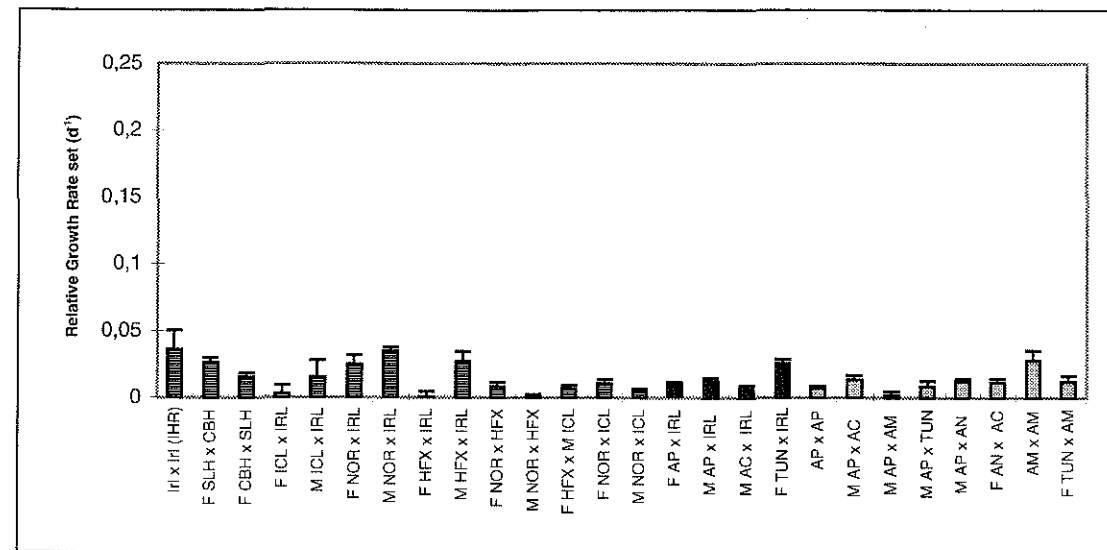


FIG. 5. AVERAGE RELATIVE GROWTH RATES FOR PLANT LENGTH (D^{-1} WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE I.

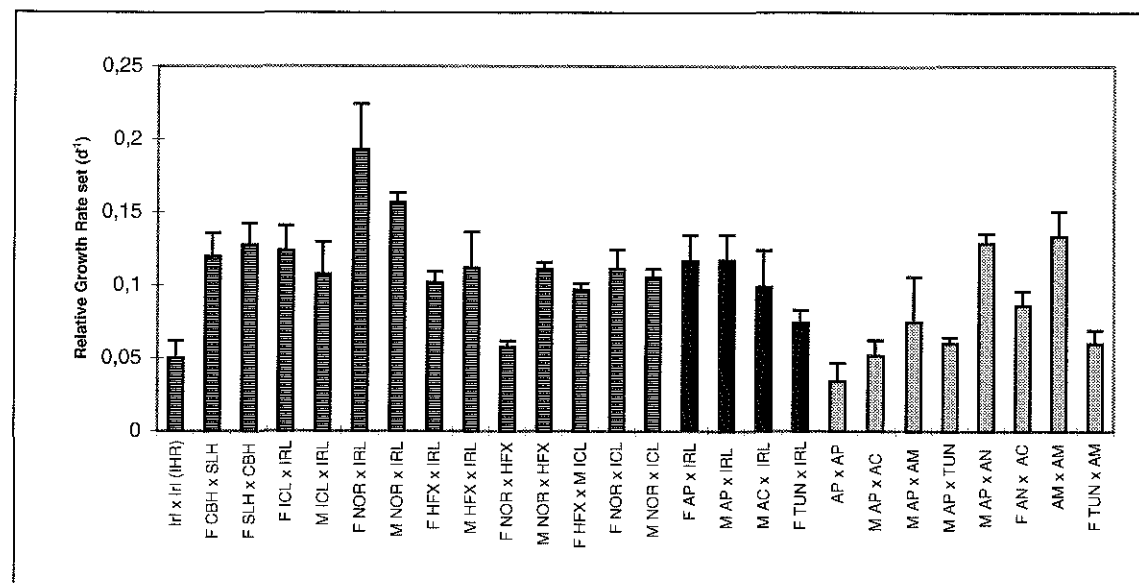


FIG. 6. AVERAGE SETTLEMENT RELATIVE GROWTH RATES FOR PLANT WIDTH (D^{-1} WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE I.

For Fig. 6 the Kruskal-Wallis test showed significant differences between crosses ($H=90.85175$, $X^2_{0.05,25}=37.652$). The null hypothesis was again rejected. The settlement relative growth rates for width are different between crosses. Multiple comparisons are presented in Appendix III. Significant differences in Appendix III are marked with patterns, striped for Atlantic, black for Atlantic x Pacific and grey for Pacific. Almost all

slow growing Pacific crosses were significantly different from Atlantic crosses.

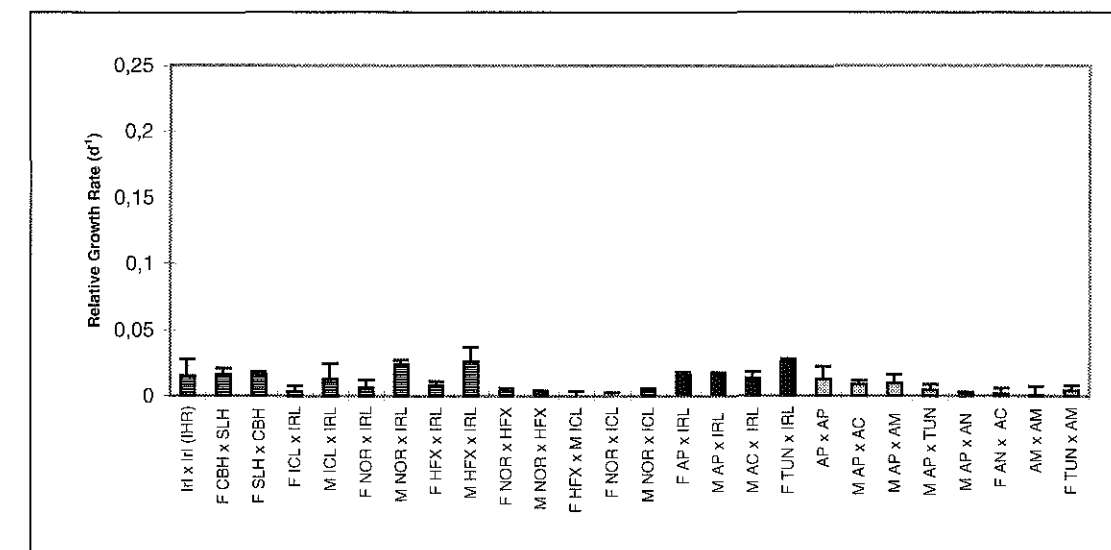


FIG. 7. AVERAGE RELATIVE GROWTH RATES FOR PLANT WIDTH (D^{-1} WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE I.

For Fig. 7 the Kruskal-Wallis test showed significant differences between crosses ($H=50.97903$, $X^2_{0.05,25}=37.652$). The null hypothesis was again rejected. The relative growth rates for width are significant different between crosses. Multiple comparisons are presented in Appendix IV. Significant differences in Appendix IV are marked with patterns, striped for Atlantic, black for Atlantic x Pacific and grey for Pacific. The female Irish x male Norway, female Irish x male Halifax and female *tenuifolia* x male Irish crosses are in most cases significantly different from all other crosses.

3.3 Length, width and length:width ratio

For reasons of equal comparisons all length and width measurements were taken after 120 (+/- 5) d. The results are presented in Figs. 8 and 9.

The ratio length:width presented in Figure 10 reveals the shape of the sporophytes, where a high number indicates long and thin sporophytes and a small number broad and small sporophytes.

Irish self-crosses from Inis Oírr formed a lot of sporophytes with very long thin elongated stipes which resulted in the large ratio figure. Irish crosses between Sleah Head and Corbet Head, Halifax x Ireland, Norway x Ireland, *tunefolia* x Ireland and *A. marginata* self cross produced the longest sporophytes with broad leaves. Most Pacific crosses produced small sporophytes. Crosses of Iceland with other strains did not perform well resulting in small/broad sporophytes.

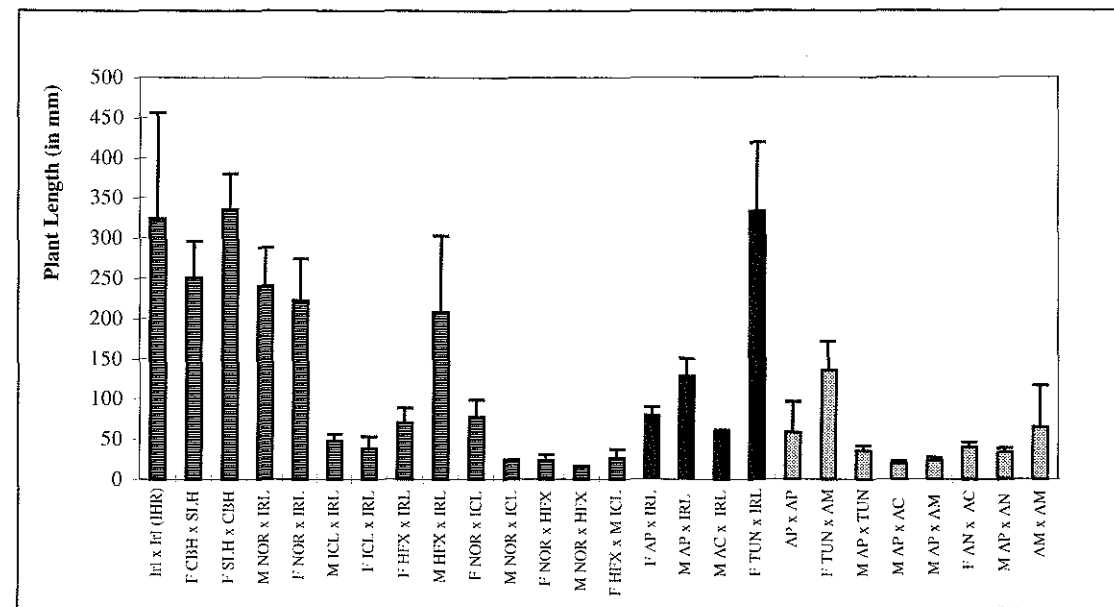


FIG. 8. AVERAGE PLANT LENGTHS (IN MM WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE 1.

For Fig. 8 the Kruskal-Wallis test showed significant differences between crosses ($H=100.6577$, $X^2_{0.05,25}=37.652$). The null hypothesis was rejected. The average plant lengths after 120 d are different between crosses. Multiple comparisons are presented in Appendix V. Significant differences in Appendix V are marked as patterns, striped for

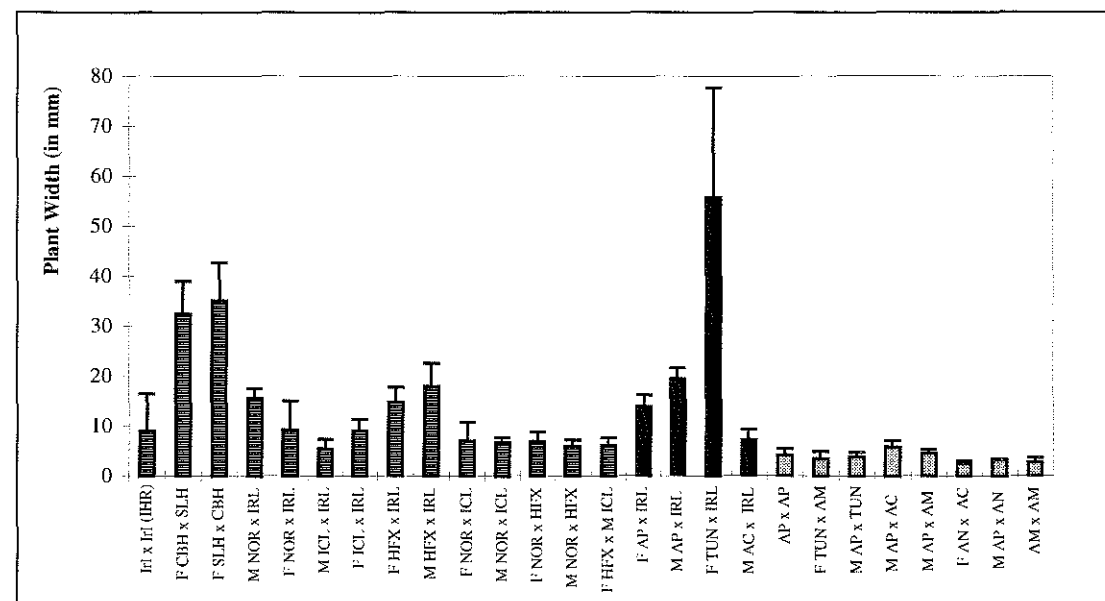


FIG. 9. AVERAGE PLANT WIDTHS (IN MM WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE 1.

Atlantic, black for Atlantic x Pacific and grey for Pacific. The Irish x *tenuifolia*, *marginata* x *tenuifolia* Irish x Norway and the Irish self crosses are significantly different from the other crosses but do not differ significantly amongst each other.

For Fig. 9 the Kruskal-Wallis test showed significant differences between crosses ($H=90.85175$, $X^2_{0.05,25}=37.652$). The null hypothesis was again rejected. The average plant widths after 120 d are different between crosses. Multiple comparisons are presented in Appendix VI. Significant differences in Appendix VI are marked with patterns, striped for Atlantic, black for Atlantic x Pacific and grey for Pacific. Most Atlantic and Atlantic x Pacific crosses differ significantly from Pacific crosses in width. Almost no differences were found between Atlantic and Atlantic x Pacific crosses.

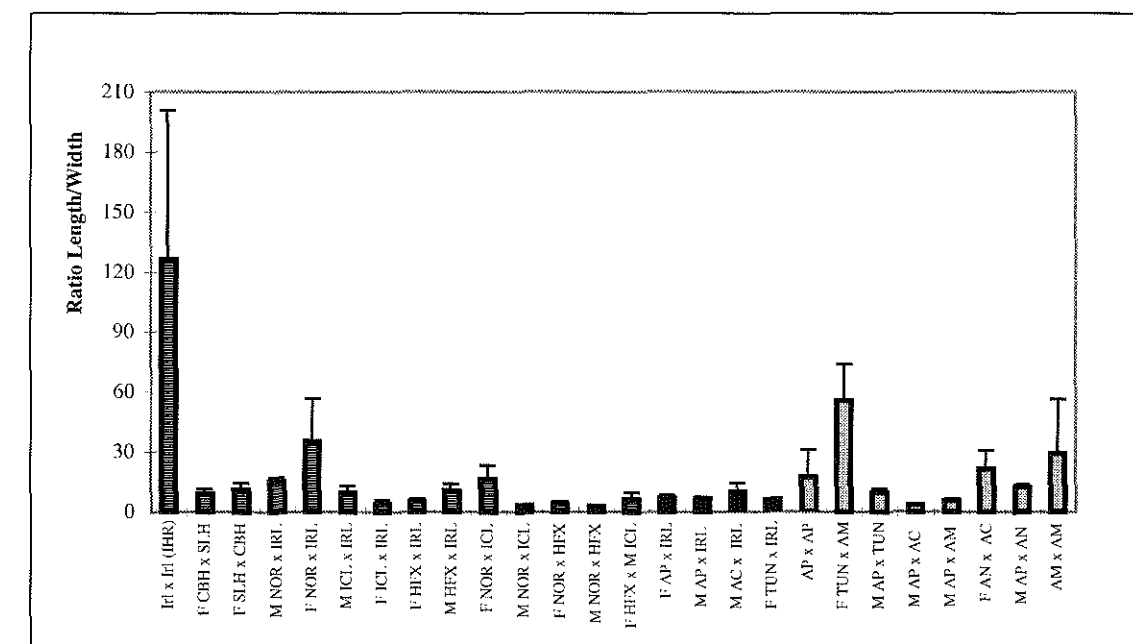


FIG. 10. RATIO LENGTH:WIDTH OF THE EXPERIMENTAL SPOROPHYTES (WITH 95% CONFIDENCE LIMITS) WITHIN NORTH ATLANTIC *ALARIA ESCULENTA* CROSSES (STRIPED) WITHIN PACIFIC *ALARIA* CROSSES (GREY) AND BETWEEN PACIFIC AND ATLANTIC *ALARIA* CROSSES (BLACK). FOR ORIGINAL CODES SEE TABLE 1.

3.4 Genetic variation

Restriction fragment length polymorphisms on the ITS1, 5.8S and ITS2 and the RuBisCo spacer revealed 123 scorable bands of which 23 were variable. The absence and presence of bands are presented in a matrix in Appendix VII. Genetic distances are calculated with the Dice(Jaquard) coefficient index (S_j) and are presented in Table 4. From Table 4 it is clear that *Alaria esculenta* from Ireland, Scotland, Halifax, France and *A. marginata* from Canada are identical in genetic composition at the spacer level. *Alaria esculenta* from Iceland and Norway are less identical with the above described species because of absence or presence of restriction sites, i.e., the restriction enzyme PST1 recognized a unique restriction site in the ITS spacers of Norway and Iceland not shared with one of

the other species. Enzyme XBA revealed a unique site for *A. crassifolia* and enzyme DRA showed one site only present in *A. praelonga*. All *Alaria* species are least related to *Undaria pinnatifida*, a member of the family Alariaceae but not belonging to the genus *Alaria* and chosen as an outgroup here.

TABLE 4. DISTANCE MATRIX FOR PAIRWISE COMPARISONS OF rDNA SPACERS AND RUBISCO SPACER RESTRICTION FRAGMENTS OF *ALARIA* SPECIES BASED ON DIGESTIONS WITH 8 RESTRICTION ENZYMES. PAIRWISE SIMILARITIES WERE CALCULATED ACCORDING TO THE DICE (JACQUARD) COEFFICIENT. PAIRWISE DISTANCES BETWEEN SPECIES ARE LISTED ABOVE THE DIAGONAL. THE TOTAL NUMBER OF RESTRICTION FRAGMENTS GENERATED IN A SPECIES IS LISTED BETWEEN PARENTHESES ON THE DIAGONAL. THE NUMBER OF RESTRICTION FRAGMENTS SHARED BETWEEN SPECIES IS LISTED BELOW THE DIAGONAL. SCL=SCOTLAND, FRC=FRANCE. FOR OTHER CODES SEE TABLE 1.

	IRL	SCL	NOR	ICL	HFX	FRC	AM	AN	AC	AP	UND
IRL	(11)	1	0.77	0.95	1	1	1	0.82	0.92	0.82	0.45
SCL	10	(11)	0.77	0.95	1	1	1	0.82	0.92	0.82	0.45
NOR	5	5	(8)	0.86	0.77	0.77	0.77	0.67	0.71	0.67	0.29
ICL	10	10	6	(12)	0.95	0.95	0.92	0.82	0.86	0.78	0.43
HFX	10	10	5	10	(11)	1	1	0.82	0.92	0.82	0.45
FRC	10	10	5	10	10	(11)	1	0.82	0.92	0.82	0.45
AM	6	6	5	6	6	6	(7)	0.86	0.92	0.86	0.31
AN	9	9	5	9	9	9	6	(13)	0.8	0.83	0.42
AC	6	6	5	6	6	6	6	6	(8)	0.8	0.29
AP	9	9	5	9	9	9	6	10	6	(13)	0.42
UND	5	5	2	5	5	5	2	5	2	5	(13)

4. DISCUSSION AND CONCLUSIONS

4.1 Atlantic cross experiments

Intraspecific hybridization

The results presented in this study for crosses between *A. esculenta* from different geographical locations in the North Atlantic show that the four strains are interfertile except for the male Halifax x female Iceland cross. Müller (1979) demonstrated in *Ectocarpus siliculosus* that sterility barriers do occur within species from different geographical locations.

The significant morphological best and fastest growing crosses were produced by the female Iceland x male Norway, female Ireland x male Norway, female Ireland x male Halifax and the Irish selfcross. All except the female Ireland x male Norway cross produced well developed haptera and sporophylls and are therefore most suitable for rope cultivation in Atlantic Irish waters. Slower growing but morphologically healthy crosses were produced between female Halifax x male Iceland, female Norway x male Halifax and female Norway x male Ireland. Similar results were found in crosses of *Laminaria digitata* from Halifax and Helgoland (Tom Dieck, 1993).

All female Iceland crosses produced poor or no results. Female Halifax crosses produced hybrids that did not develop midribs. The poor hybridization results from Icelandic plants might be caused by ecotypic variation, i.e., different temperature tolerance of the gametophyte and/or sporophyte and temperature responses of growth (Breeman, 1988). However, these results are laboratory results under controlled conditions and do not reflect field conditions.

The winter isotherms are the same for Iceland, Halifax and Norway (0 - 5°C). Only the summer isotherm is different for Iceland (5 - 10°C v.s. 10 - 15°C for Ireland, Norway and Halifax (Lüning 1990). Fertilisation, zygote and young sporophyte development in all crosses was induced at 10°C, which is the upper summer isotherm boundary for Iceland. This relatively high temperature for crosses with Iceland gametophytes might prove unfavourable resulting in the observed poor results in Iceland crosses. Tom Dieck & de Oliveira (1992) showed differences in fertility of female gametophytes of five species of *Laminaria* at three different temperatures.

Lüning & Freshwater (1988) showed that little or no variation exists in the upper temperature tolerance of *Chondrus crispus* and *Scytosiphon lomentaria* ranging from Iceland to Spain and of isolates of *Laminaria saccharina* and *Laminaria longicuris* from various locations, although only a few detailed studies are available. There is however a possibility that local and genetically fixed temperature ecotypes have evolved in *Alaria esculenta*.

Genetic analysis

The calculated pairwise distances from the restriction fragment analysis between *Alaria esculenta* isolates from Ireland, Scotland, Halifax and France (Table 4) showed that they were genetically identical. Norway and Iceland showed differences at the DNA level resulting in more or less restriction fragments.

The differences encountered in restriction fragments in respect of the DNA spacers examined suggest that the Icelandic and Norwegian *A. esculenta* populations are more isolated than the other Atlantic *Alaria* species. Probably little or no gene flow occurs between the Icelandic and Norwegian populations and between other populations. The different genotype and therefore genetic make-up of the Icelandic strain might account for the existing fertility barrier and the poor cross results in the Icelandic crosses. However, these genetic results are only preliminary results and serve to illustrate possible existing genetic variation between *A. esculenta* populations rather than to establish conclusive facts regarding the nuclear genome of *A. esculenta*.

Coyer *et al.* (1997) showed distinguishable biogeographic populations along the northeast Pacific coast of the kelp *Postelsia palmaeformis*. They showed decreasing genetic relatedness with increasing distance between populations assessed with M13 Fingerprinting and RAPDs. Bhattacharya *et al.* (1990) showed with RFLPs 8 distinct populations of the kelp *Costaria costata* over a range of 400 km. These populations appeared to be discreet breeding groups.

4.2 Pacific x Atlantic and Pacific Cross experiments

Interspecific hybridization

From the results presented in Table 3 it is clear that *A. esculenta* from the Atlantic does hybridize with other Pacific *Alaria* species, showing a close relationship between *Alaria* species throughout the northern hemisphere. In fact, the best cross produced was that between Atlantic *A. esculenta* and Pacific *A. tenuifolia*, giving rise to the largest and best-developed plants. This cross would be most suitable to be used in closed-tank cultivation.

Crosses between *A. esculenta* and *A. praelonga* produced healthy and well developed sporophytes. Crosses between female *A. esculenta* and male *A. crassifolia* produced slow-growing offspring, whereas the reciprocal cross did not. Crosses and reciprocal crosses of *A. esculenta* with *A. marginata* and *A. nana* were not successful, resulting in a few elongated stipes, indicating that various fertility barriers are active. Similar close relationships and fertility barriers have been found in other Atlantic and Pacific kelp species (Bolton *et al.*, 1983; Lewis *et al.*, 1986; tom Dieck & de Oliveira, 1993; tom Dieck, 1993).

Crosses among Pacific species showed that they hybridize in many cases, producing the largest plants in *A. tenuifolia* x *A. marginata* crosses. Only *A. nana* x *A. marginata* crosses did not produce any sporophytes which is in contrast with the observations of Widdowson (1971). He found plants that appear to be hybrids between *A. nana* and *A. marginata* in the wild. Widdowson (1971) considered that *A. marginata* and *A. tenuifolia* intergrade with each other through a series of populations that appear to be ecotypes. The different cross results shown in Table 3 however, support the hypothesis that *A. marginata* and *A. tenuifolia* are two genetically different species.

Fertilisation in *Alaria* species is facilitated by the sex pheromone lamoxirene, which can be found in all species of the Alariaceae, Laminariaceae and Lessionaceae

(Müller *et al.*, 1985), but incompatibility was expressed between several species of the genus *Alaria*. In three crosses (two inter-specific and one intra-specific) no microscopic sporophytes were formed. This in contrast with Laminariales from the North Atlantic and Pacific where microscopic sporophytes were produced in all crosses but became irregular and stunted during later development (tom Dieck, 1992). Yarish *et al.* (1990) postulated that the sharing of a common sex pheromone is sufficient reason to expect interfertility. This is not true for the genus *Alaria*. Reproductive isolation develops gradually by different processes that might lead to fertility barriers. There are several examples in the Laminariales of gradual reproductive isolation. In several *Undaria* species the successful formation of hybrids was expressed differently in reciprocal crosses (tom Dieck, 1992). The same results were encountered in the crosses between *A. crassifolia* and *A. esculenta* during the present investigation.

Species belonging to the order Laminariales are able to produce sporophytes from unfertilized eggs (parthenogenic sporophytes), although most parthenogenic sporophytes have an abnormal morphology (Nakahara & Nakamura, 1973; tom Dieck, 1992). In the present study parthenogenic sporophytes did not survive or were clearly recognized as cell clumps without haptera or half-bleached stunted sporophytes up to 1 cm long, resembling the parthenogenic sporophytes in female control cultures.

Genetic analysis

Druehl (1990) found that environmental factors caused distinctive species morphotypes in *A. nana*, *A. marginata* and *A. tenuifolia*. Using restriction enzyme digestion of the small subunit rDNA and intergeneric spacers he found three restriction map variants indicating distinctive breeding populations of the three species which is in contrast with the findings of Widdowson (1971). Druehl & Saunders (1990) concluded that there is no consistent relationship between morphological and rDNA variation in *Alaria*. After sequencing of the entire 18S ribosomal gene for *A. nana* and *A. marginata* they found a nucleotide divergence of only 0.05%, indicating almost negligible genetic divergence. In this study *A. marginata* and *A. nana* showed different restriction fragment patterns at the intertranscribed spacer level. The differences in results compared with the study of Druehl & Saunders (1990) probably is caused due to the use of the 18S slow-evolving ribosomal gene instead of the fast-evolving spacer regions applied in this study. Cross experiments showed that *A. nana* and *A. marginata* do not hybridise, showing a fertility barrier which is an indication of a large genetic distance between the two species.

In this study *Alaria marginata* is identical to four *A. esculenta* isolates in its restriction patterns. Remarkably *A. marginata* did not hybridize with *A. esculenta* in this study despite of the similarities in genotype of the ITS 1 and ITS 2. In Table 4, *A. nana* and *A. praelonga* share the same distances, whereas *A. crassifolia* is more closely related to *A. esculenta*. This is also inconsistent with the hybridisation experiments. These results show clearly that eight restriction enzymes, of which five are informative, are not sufficient to solve genetic relationships in the genus *Alaria*. Clearly more enzymes have to be applied to generate a reliable data set as only 2.56% of the entire ITS1 and ITS2 is sampled with the five informative restriction enzymes.

Chromosomes

If we compare chromosome numbers (n) in the genus *Alaria* we can distinguish two groups. One group with n=14 viz., *A. nana*, *A. marginata*, *A. tenuifolia*, *A. fistulosa*, *A.*

taeniata and one group with 22-28 viz., *A. praelonga*, *A. crassifolia*, *A. grandifolia* (subspecies of *A. esculenta* according to Lüning, 1990) and *A. esculenta* (Robinson and Cole, 1971; Lewis, 1996).

Lewis (1996) suggested that the evolution of most taxa of brown algae was accompanied by polyploidy. Nakahara & Nakamura (1973) obtained diploid gametophytes in several members of the Laminariales. When these were crossed with haploid or diploid gametophytes, triploid and tetraploid sporophytes were obtained respectively. They even managed to cross diploid with triploid gametophytes of *A. crassifolia* resulting in pentaploid sporophytes. These laboratory investigations illustrate hypothetical mechanisms by which polyploidy could have arisen in the genus *Alaria*. The existence of polyploids suggests one mechanism by which speciation has probably occurred (Lewis, 1996).

In hybridization, two sets of chromosomes from different sources are combined. The success of hybridization between species by sexual means initially depends on the compatibility of the gametes and then on the compatibility of the genomes. The results presented here (Table 3) show that the *Alaria* species with matching chromosome numbers, i.e., $n=22-28$, hybridized without difficulty, except for *A. crassifolia* which produced in two different reciprocal crosses stunted and abnormal sporophytes. This might be an indication of gradual reproductive isolation. Species with $n=14$ produced only two healthy viable crosses with $n=22-28$ species. Lewis (1996) showed that there are few exceptions in brown algae that it is not necessary for the chromosome number to match to obtain hybrids. Crosses among $n=14$ species showed even more incompatibility. No viable crosses were produced except for the *A. marginata* x *A. tenuifolia* cross. In $n=14$ crosses incompatibility is probably caused by incompatibility of the gametes or perhaps by chromosomal mispairing at meiosis. Hybridization results indicate that *A. esculenta*, *A. praelonga* and *A. crassifolia* are most closely related, and to a lesser extent, to *A. tenuifolia* and *A. marginata* and least related to *A. nana*. These findings are in contrast with the restriction patterns which are probably caused by the few informative restriction enzymes (see discussion on genetic analysis).

Kelp evolution and divergence

The ability of the Atlantic *A. esculenta* to hybridise with several Pacific *Alaria* species indicates a close relationship between the species in the two oceans and is further support of the original hypothesis of Stam *et al.*, (1988) that species of laminariales diverged from a common ancestor 15-19 Ma ago, most probably in the north Pacific, and then invaded the North Atlantic after inundation of the Bering Land Bridge 3.5 Ma ago. It is possible that *Alaria* species with $n=14$ chromosomes diverged first from a common ancestor of the genus *Alaria*, representing the oldest assemblage of *Alaria* species at the beginning of the Miocene 27 Ma ago. Probably during the mid-Miocene steepening of the temperature gradient between high and low latitudes extensive radiation in North Pacific Laminariales took place (Stam *et al.*, 1988). This major cooling step in the Tertiary leading to glaciation of the higher latitudes was possibly the driving force of speciation in Laminariales (Lüning, 1990). The climatic disturbance coupled with the continuously changing configuration of the archipelagos in the North Pacific due to plate tectonics and

sea level fluctuation could provide the ecological opportunity in which polyploids (amphiploids) can exploit their inherent advantage (Grant, 1981). Climatic changes in general bring about changes in the distribution of the species, altering genetic isolation. New contacts are made, natural hybridization occurs, and hybrid polyploids are formed (Grant, 1981). It therefore is possible that $n=22-28$ *Alaria* species are polyploid *Alaria* species and diverged from hybrids of $n=14$ species and radiated in several new species during the mid-Miocene cooling period. Possible donors for the polyploid hybrids might be *A. tenuifolia* and *A. marginata* because of the ability of these donors to hybridize with the postulated polyploid *Alaria* species (see Table 3). Not until the opening of the Bering Land Bridge were *Alaria* species and Laminariales in general able to invade the Atlantic Ocean. Because polyploids often exhibit superior vigor, homeostatic buffering and environmental adaptability as compared with their diploid partners they are likely to be the most suitable candidates to invade the Atlantic and adapt to their new environment (Grant, 1981). With our limited knowledge of chromosome numbers in the genus *Alaria*, *A. praelonga*, *A. crassifolia* and *A. esculenta* are the most likely candidates from which only *A. esculenta* has really invaded the Atlantic. There is a possibility that other species might have invaded the Atlantic but did not survive the severe Pleistocene glaciation conditions.

Remarkably, nowadays the species with $n=22-28$ all occur at the North West Pacific coast/polar circle area or in the case of *A. esculenta* in the Atlantic Ocean as well. The $n=14$ species are mainly found at the north eastern Pacific coast (Widdowson, 1971). In the geological past, different types of temperature stress have existed on both sites of the Pacific, with more extreme temperature ranges on the West Pacific coasts and more colder gradual ranges on the East Pacific coasts. Nowadays the Asiatic side of the Pacific has severely compressed isotherms and a wide annual span of seawater temperatures (Lüning, 1990). This supports the idea that the hypothesised polyploid species are found in the more extreme temperature ranges of the West Pacific coast/polar circle area (polar light regime) due to their possible superior adaptability.

4.3 Summary

Intraspecific crosses of the genus *Alaria esculenta*

The significant morphological best and fastest growing crosses were produced by the female Iceland x male Norway, female Ireland x male Norway, female Ireland x male Halifax and the Irish self cross. All except the female Ireland x male Norway cross produced well developed haptera and sporophylls and are therefore most suitable for rope cultivation in Atlantic Irish waters.

The restriction fragment length polymorphism technique distinguishes genetic different populations of the genus *A. esculenta* in the North Atlantic Ocean.

Interspecific crosses of *Alaria* species

The significant morphological and fastest growing cross was produced between Atlantic *A. esculenta* and Pacific *A. tenuifolia*, producing the largest and best developed plants. This cross would be most suitable to be used in closed tank cultivation.

There is a close relationship between Atlantic and Pacific *Alaria* species indicating a recent species radiation in the genus *Alaria*.

5. REFERENCES

- Bakker, F.T., Olsen J.L., Stam W.T. & van den Hoek C. (1992). Nuclear ribosomal DNA internal transcribed spacer regions (ITS1 and ITS2) define discrete biogeographic groups in *Cladophora albida* (Chlorophyta). *J. Phycol.* **28**: 839-845.
- Bhattacharya, D. & Druehl L.D. (1987). Molecular genetic analysis of variation in *Costaria costata* (Turner) Saunders. *Hydrobiologia* **151/152**: 63-67.
- Bhattacharya, D. & Druehl L.D. (1988). Phylogenetic comparison of the small-subunit ribosomal DNA sequence of *Costaria costata* (Phaeophyta) with those of other algae, vascular plants and oomycetes. *J. Phycol.* **24**: 539-543.
- Bhattacharya, D. & Druehl L.D. (1989). Morphological and DNA sequence variation in the kelp *Costaria costata* (Phaeophyta). *Mar. Biol.* **102**: 15-23.
- Bhattacharya, D., Mayes C. and Druehl L.D. (1991). Restriction endonuclease analysis of ribosomal DNA sequence variations in *Laminaria* (Phaeophyta). *J. Phycol.* **27**: 624-628.
- Bolton, J.J., Germann I. & Lüning K. (1983). Hybridization between Atlantic and Pacific representatives of the simplices section of *Laminaria* (Phaeophyta). *Phycologia* **22**: 133-140.
- Breeman, A.M. (1988). Relative importance of temperature and other factors in determining geographic boundaries of seaweeds: experimental and phenological evidence. *Helgoländer Meeresunters.* **42**: 199-241.
- Chapman, A.R.O. (1974). The genetic basis of morphological differentiation in some *Laminaria* populations. *Mar. Biol.* **24**: 85-91.
- Coyer, J.A., Olsen J.L. and Stam W.T. (1997). Genetic variability and spatial separation in the sea palm kelp *Postelsia palmaeformis* (Phaeophyceae) as assessed with M13 fingerprints and RAPDS. *J. Phycol.* **33**: 561-568.
- Dixon, P.S. and Irvine L.M. (1977). *Seaweeds of the British Isles*. Vol 1 Rhodophyta part 1. Introduction, Nemaliales and Gigartinales. British Museum (Natural History), London.
- Druehl, L.D. (1988). *Cultivated edible kelp*. In Lembi, C.A. and J.B. Waaland (eds.), *Algae and human affairs*. Cambridge University press, Cambridge pp. 119-134.
- Egan, B., Garcia-Ezquivel Z., Brinkhuis B.H. and Yarish C. (1990). Genetics of morphology and growth in *Laminaria* from the North Atlantic Ocean: implications for biogeography. In Garbary, D.J. and G.R. South (eds.), *Evolutionary biogeography of the marine algae of the North Atlantic*. NATO ASI Series, Vol. G22. Springer-Verlag, Berlin pp. 147-171.
- Fletcher, R.L. (1987). *Seaweeds of the British Isles*. Vol 3 Fucophyceae (Phaeophyceae) part 1. British Museum (Natural History), London.
- Grant, V. (1981) *Plant speciation* Ed. 2. Columbia University Press, New York, 552 pp.
- Guiry, M.D. and Blunden G. (1991). *Seaweed resources in Europe: use and potential*.

- John Wiley & sons, Chichester.
- Guiry, M.D. and Cunningham E.M. (1984). Photoperiodic responses in the reproduction of north-eastern Atlantic *Gigartina acicularis* (Rhodophyta: Gigartinales). *Phycologia* **23**: 357-367.
- Guiry, M.D. and Hession C. (1996). *Eat up your seaweed!* Ireland of the Welcomes **45**: 22-25.
- Guiry, M.D. and West, J.A. (1983). Lie history and hybridization studies on *Gigartina stellata* and *Petrocelis cruenta* (Rhodophyta) in the North Atlantic. *J. Phycol.* **19**: 474-494.
- Guiry, M.D. (1997). *Research and development of a sustainable Irish seaweed industry*. Occasional papers in Irish science and technology. No. 14 Went Memorial Lecture 1996. Royal Dublin Society 11 pp.
- Hillis, D.M., Moritz C. & Mable B. K. (1996). *Molecular Systematics* Ed 2. Sinauer Assoc., Sunderland, MA, USA 653 pp.
- Indergaard, M. and Minsaas J. (1991). *Animal and human nutrition*. In Guiry, M.D. and G. Blunden (eds.), *Seaweed resources in Europe: use and potential*. John Wiley & sons, Chichester 21-63 pp.
- Innes, D.J. (1984). Genetic differentiation among populations of marine algae. *Helgol. Meeresunters.* **38**: 401-417.
- Kain J.M. and Dawes C.P. (1987). Useful European seaweeds: past hopes and present cultivation. *Hydrobiologia* **151/152**: 173-181.
- Kain, J.M., Holt T.J. & Dawes C.P. (1990). *European Laminariales and their cultivation*. In Yarish, C., C.A. Penniman and P. van Patten (eds.), *Economically important marine plants of the Atlantic: their biology and cultivation*. Groton: Connecticut Sea Grant College Program 158 pp.
- Kain J.M. (1991). *Cultivation of attached seaweeds*. In Guiry, M.D. and G. Blunden (eds.), *Seaweed resources in Europe: use and potential*. John Wiley & sons, Chichester pp. 334-377.
- Kooistra, W.H.F.C., Stam W.T., Olsen J.L. & van den Hoek C. (1992). Biogeography of *Cladophoropsis membranaceae* (Chlorophyta) based on comparisons of nuclear rDNA ITS sequences. *J. Phycol.* **28**: 660-668.
- Lehman, R.L. & Manhart J.R. (1997). A preliminary comparison of restriction fragment patterns in the genus *Caulerpa* (Chlorophyta) and the unique structure of the chloroplast genome of *Caulerpa sertularioides*. *J. Phycol.* **33**: 1055-1062.
- Levring, Hoppe & Schmid (1969) *Marine algae* pp. 144-146.
- Lewallen and Lewallen (1996). *Sea vegetable gourmet cookbook and wildcrafter's guide*. Mendocino Sea Vegetable Company, CA, USA 128 pp.
- Lewis, J.L., Neushul M. & Harger B.W.W. (1986). Interspecific hybridization of the species of *Macrocystis* in California. *Aquaculture* **57**: 203-210.
- Lewis, J.L. (1996). Chromosomes of the brown algae. *Phycologia* **35**: 19-40.
- Lüning, K., Chapman A.R.O. & Mann K.H. (1978). Crossing experiments in the non-digitate complex of *Laminaria* from both sides of the Atlantic. *Phycologia* **17**: 293-298.
- Lüning, K., and Freshwater, W. (1988). Temperature tolerance of Northeast Pacific marine algae. *J. Phycol.* **24**: 310-315.

- Lüning, K. (1990). *Seaweeds. Their environment, Biogeography and Ecophysiology*. Eds. C. Yarish and H. Kirkman, John Wiley & Sons, Inc., New York 527 pp.
- Madlener, J.C. (1977). *The seaweeds book*. Clarkson N. Potter, New York.
- Mai, K.S., Mercer J.P. & Donlon J. (1996). Comparative studies on the nutrition of 2 species of *Abalone*, *Haliotis tuberculata* and *Haliotis discus* Hanni-Ino.5. The role of poly unsaturated fatty acids in macro algae in *Abalone* nutrition. *Aquaculture* **139**: 77-89.
- Migita, S. (1984). In: *Seaweeds. Their environment, Biogeography and Ecophysiology*, Eds. C. Yarish and H. Kirkman, John Wiley & Sons, Inc., New York, 1990 527 pp.
- Müller, D.G. (1979). Genetic affinity of *Ectocarpus siliculosus* (Dillw.) Lyngb. From the Mediterranean, North Atlantic and Australia. *Phycologia* **18**: 312-318.
- Müller, D.G., Maier I. & Gassmann, G. (1985). Survey on sexual pheromone specificity in Laminariales (Phaeophyceae). *Phycologia* **24**: 475-477.
- Munda, I.M. and Lüning K. (1977). Growth performance of *Alaria esculenta* off Helgoland. *Helgoländer wiss. Meeresunters.* **29**: 311-314.
- Nakahara, H. and Nakamura Y. (1973). Parthenogenesis, apogamy and apospory in *Alaria crassifolia* (Laminariales). *Mar. Biol* **18**: 327-332.
- Newton, L. (1931). *A handbook of the British seaweeds*. British Museum (Nat. Hist.), London.
- Nisizawa, K., Noda H., Kikuchi R. & Watanabe T. (1987). The main seaweed foods in Japan. *Hydrobiologia* **151/152**: 5-29.
- Norton, T.A, Mathieson A. & Neushul A.C. (1982). A review of some aspects of form and function in seaweeds. *Bot. Mar.* **25**: 501-510.
- Pérez, R., Kaas R. and Barbaroux O. (1984). Culture expérimentale de l'algue *Undaria pinnatifida* sur les côtes de France. *Sci. PLche* **343**: 3-15.
- Rice, E.L. and Bird C.J. (1990). Relationships among geographically distant populations of *Cracilaria verrucosa* (Gracilariales, Rhodophyta) and related species. *Phycologia* **29**: 501-510.
- Robinson, G.G.C. and Cole, K. (1971). Cytological investigations of some North American species of the genus *Alaria* Greville. *Bot. Mar.* **14**: 59-62.
- Rueness, J. (1973). Speciation in *Polysiphonia* (Rhodophyceae, Ceramiales) in view of hybridization experiments: *P. hemisphaerica* and *P. boldii*. *Phycologia* **12**: 107-109.
- Sambrook, J., E.F. Fritch, E.F. & Maniatis, T. (1989). *Molecular cloning: A laboratory manual* (Ed. 2) Cold Spring Harbor Laboratory Press, New York, 3 vols.
- Saunders, G.W. and Druehl L.D. (1993). Nucleotide sequence of the internal transcribed spacers and 5.8S rRNA genes from *Alaria marginata* and *Postelsia palmaeformis* (Phaeophyta; Laminariales). *Mar. Biol.* **115**: 347-352.
- Sundene, O. (1962). The implications of transplant and culture experiments on the growth and distribution of *Alaria esculenta*. *Nytt Mag. Bot.* **9**: 155-174.
- Tom Dieck, I. (1992). North Pacific and North Atlantic digitate *Laminaria* species (Phaeophyta): hybridisation experiments and temperature responses. *Phycologia* **31**: 147-163.
- Tom Dieck, I. (1993). Temperature tolerance and survival in darkness of kelp gametophytes (Laminariales, Phaeophyta): ecological and biogeographical implications. *Mar. Ecol. Prog. Ser.* **100**: 253-264.

- Tom Dieck, I., and de Oliveira E.C. (1993). The section digitatae of the genus *Laminaria* (Phaeophyta) in the northern and southern Atlantic: crossing experiments and temperature responses. *Mar. Biol.* **115**: 151-160.
- van den Hoek, C., Mann, D.G. & Jahns, H.M. 1995. *Algae: An Introduction to Phycology*. Cambridge University Press, Cambridge, 623 pp.
- van Oppen, M.J.H., Draisma, S.G.A., Olsen, J.L. & Stam W.T. (1995). Multiple trans-Arctic passages in the red alga *Phycodrys rubens*: evidence from nuclear rDNA ITS sequences. *Mar. Biol.* **123**: 179-188.
- Widdowson, T.B. (1971a). A statistical analysis of variation in the brown alga *Alaria*. *Syesis* **4**: 125-143.
- Widdowson, T.B. (1971b). A taxonomic revision of the genus *Alaria* Greville. *Syesis* **4**: 11-49.
- Wu, C.Y. and Guangheng L. (1987). Progress in the genetics and breeding of economic seaweeds in China. *Hydrobiologia* **151/152**: 57-61.
- Yamanaka, R. and Akiyama K. (1993). Cultivation and utilisation of *Undaria pinnatifida* (wakame) as food. *J. Appl. Phycol.* **5**: 249-253.
- Yoshida, T. (1979). *Streblonema* (Phaeophyceae) infection in the frond of cultivated *Undaria* (Phaeophyceae). *Proc. Int. Seaweed Symp.* **9**: 219-223.
- Zar, J.H. (1996). *Biostatistical analysis* (Ed. 3) Prentice Hall International Editions, London, 1974. By Prentice-Hall, Inc. 662 pp.

Appendices

- I Statistical results of the multiple comparisons using Dunn's test for relative growth rates settlement length.
- II Statistical results of the multiple comparisons using Dunn's test for relative growth rates length.
- III Statistical results of the multiple comparisons using Dunn's test for relative growth rates settlement width.
- IV Statistical results of the multiple comparisons using Dunn's test for relative growth rates width.
- V Statistical results of the multiple comparisons using Dunn's test for plant lengths.
- VI Statistical results of the multiple comparisons using Dunn's test for plant widths.
- VII Restriction fragment data matrix for the ITS and Rubisco spacer of *Alaria* species.

IRL	FCBH	FSLH	FIMR	FNOR	FIHR	FICL	FHFH	FIMR	FICL	FNOR	FNOR	FHFH	FHFH	FAP	FIHR	FIMR	FTUN	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM
IRL(self)	M SLH	M CBH	M NOR	M IMR	M ICL	M IHR	M IMR	M HFX	M NOR	M ICL	M HFX	M NOR	M ICL	M IHR	M AP	M AC	M IRL	AP	AM	FTUN	AC	AM	AC	AN	AM
IRL	36.39086	39.88327	35.70877	44.82493	46.65527	39.88327	38.00218	42.26135	42.26135	41.32654	43.40155	42.26135	44.82493	36.39086	35.56823	41.32654	39.88327	46.65527	40.54568	41.32654	39.88327	57.86874	39.88327	39.88327	44.82493
F CBH	4.32123	3.004309	6.620176	5.478456	2.768278	3.631132	4.254102	1.906504	3.336757	2.106913	0.041555	0.494654	2.453992	4.416783	4.299956	1.981086	3.865148	0.515942	0.803327	0.47116	0.413409	0.637732	4.406311	5.051945	5.172504
F SLH	1.243798	3.257975	2.426933	0.727507	0.413089	0.160123	2.31095	0.489344	2.194357	4.594036	4.109116	1.298492	0.137525	0.179005	2.356944	0.102958	3.448254	4.026245	4.307989	4.677395	2.331787	0.614221	1.469851	2.050073	
F IMR	29.26632	39.88327	41.92988	34.23561	32.02448	36.97968	36.97968	35.90661	38.27657	36.97868	39.88327	30.09478	29.09467	35.90661	34.23561	41.92988	35.00504	35.90661	34.23561	54.13125	34.23561	34.23561	34.23561	39.88327	
F NOR	3.983313	3.152951	0.222594	0.730234	1.306618	1.061422	0.573153	0.912088	3.177531	2.674965	0.246254	1.359342	1.138366	1.056909	1.002855	2.283574	2.492498	2.79475	3.0183	1.531771	1.63329	2.385431	2.80909		
F IHR	35.70877	37.98099	29.26632	26.64598	32.43245	32.43245	31.20457	33.90482	32.43245	35.70877	24.29273	23.04215	21.20457	29.26632	37.98099	30.16279	31.20457	29.26632	51.13348	29.26632	29.26632	29.26632	35.70877		
F ICL	0.256886	2.823612	3.129089	2.80467	4.804661	2.940958	4.785418	7.025607	6.644382	3.5397	3.114835	3.621908	4.95206	2.810179	5.590347	6.757562	6.951787	7.514106	3.901428	2.072698	1.192847	0.127173			
F HFX	34.94157	34.94157	33.80495	36.31234	34.94157	38.00218	27.55333	26.45733	33.80495	32.02448	40.14481	32.84575	33.80495	32.02448	40.14481	32.84575	33.80495	32.02448	52.76061	32.02448	32.02448	38.00218			
F IMR	2.320839	0.590967	2.134814	4.678346	4.028452	1.528332	0.025739	0.249856	2.099715	0.272577	5.200591	3.818782	4.440158	3.616335	3.798373	0.416297	1.243515	1.330386							
F ICL	39.53187	38.5309	40.74852	39.53187	42.26135	33.18195	32.27764	38.5309	36.97868	44.19798	37.69216	38.5309	36.97868	55.90651	36.97868	36.97868	42.26135								
FNOR	1.529005	0.164424	2.083762	1.548541	0.763765	2.080384	1.776024	0.032885	1.74115	1.702733	1.487098	1.58574	1.732981	0.987979	2.52484	3.138174	4.09116								
Fnor	1.400031	3.504899	2.947619	0.761153	0.498703	0.301679	1.49612	0.310896	3.524339	3.359739	3.154467	3.367556	2.355563	0.921205	1.569447	2.456585									
F HFX	39.77818	38.5309	41.32654	31.98286	31.04364	37.50323	35.90661	43.305	36.64097	37.50323	35.90661	55.20326	35.90661	35.90661	35.90661	35.90661	41.32654								
F HFX	2.234265	1.717236	0.554815	1.782368	1.5939	0.162587	2.097478	1.969805	1.755593	1.802512	1.965748	1.15845	2.419878	3.050849	4.032286										
F HFX	40.74852	43.40155	34.62246	33.75876	39.77818	38.27657	45.28945	38.96629	39.77818	38.27657	56.77333	38.27657	38.27657	43.40155											
F HFX	0.55728	2.743745	3.744891	4.469508	2.416784	4.504542	0.766513	1.018314	0.630244	0.459842	1.01128	3.920155	5.217116	5.874093											
F HFX	42.26135	33.18195	32.27764	38.5309	36.97868	44.19798	37.69216	38.5309	36.97868	55.90651	36.97868	36.97868	42.26135												
F HFX	2.108197	4.21391	3.979183	1.888821	3.458263	0.082185	0.302787	0.038761	0.119438	0.432682	3.503177	4.791005	5.474888												
F HFX	36.39086	35.56823	41.32654	39.88327	46.65527	40.54568	41.32654	39.88327	57.86874	39.88327	39.88327	44.82493													
F AP	1.394046	1.207308	0.680642	1.068436	2.154501	1.659589	1.940372	2.004316	1.802788	1.621334	2.256421	2.948641													
F IHR	0.323376	2.465667	0.218502	3.538285	3.318209	4.561561	4.509991	3.871629	0.469238	1.354307	2.363421														
F IMR	31.04364	29.09467	37.84889	29.99627	31.04364	29.09467	51.03543	29.09467	29.09467	51.03543	29.09467	35.56823													
F IMR	2.28937	0.04169	3.404866	4.012845	4.299443	4.395545	3.988257	0.446676	0.951227	1.546286															
FTUN	35.90661	43.305	36.64097	37.50323	35.90661	55.20326	35.90661	35.90661	35.90661	35.90661	41.32654														
FTUN	2.013093	1.334719	1.345488	1.663857	1.820928	0.814565	1.700383	3.331328	4.177106																
AP	41.92988	35.00504	35.90661	34.23561	54.13125	34.23561	34.23561	39.88327																	
FTUN	3.102402	3.473309	3.750935	4.021155	2.166032	0.630435	1.382576	2.269636																	
AP	42.56044	43.305	41.92988	59.29781	41.92988	41.92988	46.65527																		
FTUN	0.199716	0.106223	0.180857	0.216422	3.61715	4.23127	4.453639																		
MAP	36.64097	35.00504	54.62113	35.00504	35.00504	40.54568																			
MAP	0.357523	0.459458	0.079334	4.089887	4.825495	4.915092																			
MAP	35.90661	55.20326	35.90661	35.90661	41.32654																				
MAP	0.083086	0.315803	4.352031	5.069169	5.139208																				
MAP	54.13125	34.23561	34.23561	39.88327																					
FAN	0.37717	4.65159	5.403731	5.399984																					
MAP	54.13125	54.13125	57.86874																						
MAP	2.564754	3.04045	3.968672																						
MAP	34.23561	39.88327																							
MAP	0.752141	1.407082																							
MAP	39.88327																								
AM	0.761448																								

APPENDIX I (RGR-set-length)

IRL	F CBH	F SLH	F IMR	F NOR	F IHR	F ICL	F HFX	F IMR	F ICL	F NOR	Fnor	F HFX	F HFX	F AP	F IHR	F IMR	FTUN	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM																									
IRL(self)	M SLH	M CBH	M NOR	M IMR	M ICL	M IHR	M IMR	M HFX	M NOR	M ICL	M HFX	M NOR	M ICL	M IHR	M AP	M AC	M IRL	M AP	M AM	M FTUN	M AC	M AM	M AN	M AM																										
IRL	x	36.574	39.6088	35.463	44.5164	46.3342	39.6088	37.7406	41.9705	41.9705	41.0421	43.1028	41.9705	44.5164	36.1404	35.3234	41.0421	39.6088	46.3342	40.2666	41.0421	39.6088	57.4704	39.6088	39.6088	44.5164																								
F CBH	M SLH	x	30.4105	24.7703	36.574	38.7659	30.4105	27.9338	33.4285	33.4285	32.2552	34.8396	33.4285	36.574	25.7308	24.5701	32.2552	30.4105	38.7659	31.2625	32.2552	30.4105	51.5635	30.4105	30.4105	36.574																								
F SLH	M CBH		x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																								
F IMR	M NOR			x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																							
F NOR	M IMR				x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																						
F IHR	M ICL					x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																					
F ICL	M IHR						x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																				
F HFX	M IMR							x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																			
F IMR	M HFX								x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																		
F ICL	M NOR									x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																	
FNOR	M ICL										x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																
Fnor	M HFX											x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	35.6595	38.0131	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088															
F HFX	M NOR												x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																	
F HFX	M ICL													x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088																
F AP	M IHR														x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088															
F IHR	M AP															x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088														
F IMR	M AC																x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088													
FTUN	M IRL																	x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088												
AP	AP																		x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088											
FTUN	AM																			x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088										
MAP	FTUN																				x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088									
MAP	AC																						x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088							
MAP	AM																								x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088					
FAN	AC																										x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088			
MAP	AN																												x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088	
AM	AM																													x	29.0649	39.6088	41.6413	34	31.8041	36.7242	36.7242	39.6088	28.8944	28.8944	35.6595	34	41.6413	34.7641	35.6595	34	53.7587	34	34	39.6088

APPENDIX II (RGR-length)

IRL	F CBH	F SLH	F IMR	F NOR	F IHR	F ICL	F HFX	F IMR	F ICL	F NOR	Fnor	F HFX	F HFX	F AP	F IHR	F IMR	FTUN	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM			
IRL(self)	M SLH	M CBH	M NOR	M IMR	M ICL	M IHR	M IMR	M HFX	M NOR	M ICL	M HFX	M NOR	M ICL	M IHR	M AP	M AC	M IRL	M AP	M AM	M FTUN	M AC	M AM	M AN	M AM				
IRL	x	34.96161	38.09853	34.11084	42.81906	44.56749	38.09853	36.30162	40.37019	40.37019	39.47722	41.45937	40.37019	42.81906	35.68255	35.17944	40.37019	38.09853	44.56749	38.73129	39.47722	38.09853	55.27916	38.09853	38.09853	42.81906		
F CBH	M SLH	x	28.98864	23.50302	34.96161	37.08239	28.98864	26.5829	31.91544	31.91544	30.77814	33.28246	31.91544	34.96161	25.73105	25.02871	31.91544	28.98864	37.08239	29.81536	30.77814	28.98864	49.44319	28.98864	28.98864	34.96161		
F SLH	M CBH		x	27.95668	38.09853	40.05356	32.70359	30.59141	35.32392	35.32392	34.29982	36.56373	35.32392	38.09853	29.85416	29.25098	35.32392	32.70359	40.05356	33.4386	34.29982	32.70359	51.70892	32.70359	32.70359	38.09853		
F IMR	M NOR			x	27.95668	38.09853	40.05356	30.59141	35.32392	35.32392	34.29982	36.56373	35.32392	38.09853	29.85416	29.25098	35.32392	32.70359	40.05356	33.4386	34.29982	32.70359	51.70892	32.70359	32.70359	38.09853		
F NOR	M IMR				x	27.95668	38.09853	40.05356	30.59141	35.32392	35.32392	34.29982	36.56373	35.32392	38.09853	29.85416	29.25098	35.32392	32.70359	40.05356	33.4386	34.29982	32.70359	51.70892	32.70359	32.70359	38.09853	
F IHR	M ICL					x	27.95668	38.09853	40.05356	30.59141	35.32392	35.32392	34.29982	36.56373	35.32392	38.09853	29.85416	29.25098	35.32392	32.70359	40.05356	33.4386	34.29982	32.70359	51.70892	32.70359	32.70359	38.09853
F ICL	M IHR						x	27.95668	38.09853	40.05356	30.59141	35.32392	35.32392	34.29982	36.56373	35.32392	38.09853	29.85416	29.25098	35.32392	32.70359	40.05356	33.4386	34.29982	32.70359	51.70892	32.70359	32.70359

IRL	F CBH	F SLH	F IMR	F NOR	F IHR	F ICL	F HFX	F IMR	F ICL	F NOR	Fnor	F HFX	F HFX	F AP	F IHR	F IMR	FTUN	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM																										
IRL(self)	M SLH	M CBH	M NOR	M IMR	M ICL	M IHR	M IMR	M HFX	M NOR	M ICL	M HFX	M NOR	M ICL	M IHR	M AP	M AC	M IRL	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM																										
IRL	IRL(self)	35.33958	38.51041	34.47961	43.28197	45.0493	38.51041	36.69407	40.80663	40.80663	39.904	41.90759	40.80663	43.28197	35.33958	35.5977	40.80663	38.51041	45.0493	39.15001	39.904	38.51041	55.87678	38.51041	38.51041	43.28197																									
F CBH	M SLH	0.850254	0.638672	2.448015	1.254234	0.328212	1.727733	0.867939	2.133173	1.441954	2.949221	1.838228	2.48209	2.425953	1.515905	1.260253	0.733619	2.714788	0.013742	1.997312	1.766382	0.639283	0.291458	2.218942	2.461301	0																									
F SLH	M CBH	0.076786	2.286114	2.386371	1.196089	3.296131	2.303505	1.766868	2.75535	4.746608	3.182999	4.071029	3.821427	0.941372	0.583679	0.003444	2.542466	0.818142	3.591596	3.231453	1.865627	0.92708	3.941706	4.260228	0.850254	0.076786																									
F IMR	M NOR	1.843983	2.24831	1.162937	2.989772	2.074431	1.533364	2.552495	4.325948	2.958232	3.741222	3.565208	0.726018	0.423329	0.066127	2.185609	0.813027	3.268997	2.964565	1.721766	0.929505	3.562013	3.844353	0.838672	1.843983	1.843983																									
F NOR	M IMR	4.022445	2.704732	5.341404	4.518475	0.084335	4.574262	6.707245	4.931381	5.929619	5.493293	1.297934	1.643965	1.739366	0.712732	2.318442	5.582972	5.140724	3.858097	2.039405	6.01081	6.341089	2.448015	4.022445	4.022445																										
F IHR	M ICL	0.876817	0.318096	0.611475	3.463489	0.111638	1.588813	0.542861	1.151774	1.171719	3.052021	2.786858	2.063935	4.124426	1.191287	0.610705	0.405974	0.770354	0.680068	0.809305	1.051664	1.254234	0.876817	0.876817	0.876817	0.876817																									
F ICL	M IHR	1.278202	0.440176	2.62708	1.032312	2.411158	1.488723	1.978025	2.113905	1.517385	1.590042	1.186651	2.796224	0.34991	1.356348	1.355382	0.235166	0.037049	1.234209	1.975964	0.3652	1.278202	1.278202	1.278202	1.278202	1.278202																									
F HFX	M IMR	3.73881	33.73881	32.64132	35.0624	33.73881	36.69407	26.87028	27.15922	33.73881	30.92213	38.76294	31.71514	32.64132	30.92213	50.94449	30.92213	30.92213	36.69407	3.73881	33.73881	32.64132	35.0624	33.73881	36.69407	26.87028	27.15922																								
F IMR	M HFX	38.17111	37.20459	39.34588	38.17111	40.80663	32.26047	32.50153	38.17111	35.7058	42.6766	36.39472	37.20459	35.7058	53.9821	35.7058	35.7058	40.80663	38.17111	38.17111	37.20459	39.34588	38.17111	40.80663	32.26047	32.50153	38.17111																								
F ICL	M NOR	3.821972	5.363568	4.298626	5.062099	5.161933	0.899796	1.073386	1.496187	0.428852	2.717464	5.084143	4.127031	3.12741	2.421312	4.736968	4.887389	2.437912	3.821972	3.821972	5.363568	4.298626	5.062099	5.161933	0.899796	1.073386	1.496187																								
FNOR	M ICL	1.581645	0.462423	1.078752	1.173153	2.944968	2.71555	2.325785	4.003979	1.804754	0.595465	0.305059	0.95845	0.97164	0.73118	0.266129	1.647947	1.581645	1.581645	0.462423	1.078752	1.173153	2.944968	2.71555	2.325785	4.003979	1.804754																								
Fnor	M HFX	38.40894	37.20459	39.904	31.11087	31.36077	37.20459	34.67063	41.81435	35.37971	36.21229	34.67063	53.30305	34.67063	39.904	1.058347	0.440806	0.317906	4.291728	4.072273	4.745036	7.143269	3.762886	1.259246	1.268661	2.684308	2.425005	0.911068	0.632382	3.994392																					
F HFX	M NOR	0.616329	0.710793	1.16551	3.621931	3.179696	5.397475	2.255943	0.034218	0.193367	1.332202	1.643713	0.192467	0.471761	2.005672	40.80663	32.26047	32.50153	38.17111	35.7058	42.6766	36.39472	37.20459	35.7058	53.9821	35.7058	35.7058	40.80663	38.17111	37.20459																					
F HFX	M ICL	0.091022	4.800213	4.528762	4.037417	5.392386	2.637248	0.604932	0.862605	2.147177	2.380566	0.371007	0.178597	2.722396	35.33958	35.5977	40.80663	38.51041	45.0493	39.15001	39.904	38.51041	55.87678	38.51041	43.28197	0.091022	0.091022	0.091022	0.091022																						
F AP	M IHR	4.487077	4.213028	3.30673	5.135137	2.71046	0.595019	0.766145	1.784289	2.266009	0.4993	0.260901	2.631315	0.346142	0.732628	1.739681	1.445724	3.515333	4.116345	2.513284	2.245425	4.668656	5.063033	1.82825	4.487077	4.487077	4.487077	4.487077	4.487077																						
F IHR	M AP	25.29929	32.26047	29.30203	37.48328	30.13769	31.11087	29.30203	49.97771	29.30203	29.30203	35.33958	0.346142	0.732628	1.739681	1.445724	3.515333	4.116345	2.513284	2.245425	4.668656	5.063033	1.82825	4.487077	4.487077	4.487077	4.487077	4.487077	4.487077	4.487077																					
F IMR	M AC	0.457756	2.020256	1.205418	4.046939	3.676568	2.214019	2.066478	2.598388	2.784557	0.893896	0.457756	2.020256	1.205418	4.046939	3.676568	2.214019	2.066478	2.598388	2.784557	0.893896	0.457756	2.020256	1.205418	4.046939	3.676568	2.214019	2.066478	2.598388	2.784557	0.893896																				
FTUN	M IRL	2.069607	0.715979	2.971071	2.699189	1.527919	0.856251	2.13754	3.493052	0.838421	2.069607	0.715979	2.971071	2.699189	1.527919	0.856251	2.13754	3.493052	0.838421	2.069607	0.715979	2.971071	2.699189	1.527919	0.856251	2.13754	3.493052	0.838421	2.069607	0.715979	2.971071	2.699189	1.527919	0.856251																	
AP	AP	41.09543	41.81435	40.48657	57.25666	40.48657	40.48657	45.0493	41.09543	41.81435	40.48657	57.25666	40.48657	40.48657	45.0493	41.09543	41.81435	40.48657	57.25666	40.48657	40.48657	40.48657	45.0493	41.09543	41.81435	40.48657	57.25666	40.48657	40.48657	40.48657	40.48657	40.48657	40.48657																		
FTUN	AM	1.887698	1.670878	0.592789	0.273622	2.095345	2.325874	0.013742	1.887698	1.670878	0.592789	0.273622	2.095345	2.325874	0.013742	1.887698	1.670878	0.592789	0.273622	2.095345	2.325874	0.013742	1.887698	1.670878	0.592789	0.273622	2.095345	2.325874	0.013742	1.887698	1.670878	0.592789	0.273622	2.095345	2.325874	0.013742															
MAP	FTUN	0.217896	1.585077	1.173833	0.214721	0.490854	1.997312	0.217896	1.585077	1.173833	0.214721	0.490854	1.997312	0.217896	1.585077	1.173833	0.214721	0.490854	1.997312	0.217896	1.585077	1.173833	0.214721	0.490854	1.997312	0.217896	1.585077	1.173833	0.214721	0.490854	1.997312	0.217896	1.585077	1.173833	0.214721	0.490854	1.997312														
MAP	AC	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382	1.322926	1.016827	0.431681	0.700881	1.766382										
MAP	AM	0.159435	1.840247	2.122587	0.639283	52.26794	52.26794	55.87678	0.159435	1.840247	2.122587	0.639283	52.26794	52.26794	55.87678	0.159435	1.840247	2.122587	0.639283	52.26794	52.26794	55.87678	0.159435	1.840247	2.122587	0.639283	52.26794	52.26794	55.87678	0.159435	1.840247	2.122587	0.639283	52.26794	52.26794	55.87678	0.159435	1.840247	2.122587	0.639283											
FAN	AC	1.32331	1.501877	0.291458	33.05715	38.51041	0.282339	2.218942	1.32331	1.501877	0.291458	33.05715	38.51041	0.282339	2.218942	1.32331	1.501877	0.291458	33.05715	38.51041	0.282339	2.218942	1.32331	1.501877	0.291458	33.05715	38.51041	0.282339	2.218942	1.32331	1.501877	0.291458	33.05715	38.51041	0.282339	2.218942	1.32331	1.501877	0.291458	33.05715	38.51041										
MAP	AN	38.51041	38.51041	2.461301	0	0	0	0	38.51041	38.51041	2.461301	0	0	0	0	0	0	38.51041	38.51041	2.461301	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0									
AM	AM	0.850254	0.638672	2.448015	1.254234	0.328212	1.727733	0.867939	2.133173	1.441954	2.949221	1.838228	2.48209	2.425953	1.515905	1.260253	0.733619	2.714788	0.013742	1.997312	1.766382	0.639283	0.291458	2.218942	2.461301	0	0.850254	0.638672	2.448015	1.254234	0.328212	1.727733	0.867939	2.133173	1.441954	2.949221	1.838228	2.48209	2.425953	1.515905	1.260253	0.733619	2.714788	0.013742	1.997312	1.766382	0.639283	0.291458	2.218942	2.461301	0

APPENDIX IV (RGR-Width)

IRL	F CBH	F SLH	F IMR	F NOR	F IHR	F ICL	F HFX	F IMR	F ICL	F NOR	Fnor	F HFX	F HFX	F AP	F IHR	F IMR	FTUN	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM
IRL(self)	M SLH	M CBH	M NOR	M IMR	M ICL	M IHR	M IMR	M HFX	M NOR	M ICL	M HFX	M NOR	M ICL	M IHR	M AP	M AC	M IRL	AP	FTUN	MAP	MAP	MAP	FAN	MAP	AM

