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Dual Role of the Metalloprotease FtsH in Biogenesis of the DrrAB Drug Transporter*

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*Running Title: *Role of FtsH in Biogenesis of the DrrAB Drug Transporter*

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Keywords: FtsH, DrrAB, ABC transporter, Drug efflux, AAA⁺ family

Background: Metalloprotease FtsH is involved in quality control of membrane proteins

Results: Simultaneous (or sequential) expression of misfolded DrrAB and FtsH results in significant recovery of DrrAB-mediated doxorubicin efflux function.

Conclusion: FtsH facilitates refolding of previously misassembled DrrAB.

Significance: This is the first study showing that FtsH contains both protease and refolding functions and plays a dual role in biogenesis of membrane proteins.

SUMMARY

This study provides the first direct evidence for the dual role of the metalloprotease FtsH in membrane protein biogenesis. Using the physiological substrate DrrAB, it is shown that FtsH is not only responsible for proteolysis of unassembled DrrB protein but it also plays a much broader role in biogenesis of the DrrAB complex. Previous studies showed that the stable expression of DrrB in the membrane depends on simultaneous expression of DrrA. Here we show that DrrB is proteolyzed by FtsH when it is expressed alone. Moreover, DrrA and DrrB proteins expressed together in a temp-sensitive *ftsH* mutant strain of *E. coli* were found to be non-functional due to their incorrect assembly. Simultaneous expression of wild-type FtsH in *trans* resulted in normal doxorubicin efflux. Strikingly, doxorubicin efflux could be restored in mutant cells irrespective of whether FtsH was expressed simultaneously with DrrAB or

expressed after these proteins had already accumulated in an inactive conformation, thus providing crucial evidence for the ability of FtsH to refold the misassembled proteins. Complementation experiments also showed that the catalytic AAA domain of FtsH contains a chaperone-like activity, however, unlike wild-type FtsH, it was unable to restore function. Our results therefore show for the first time that FtsH contains the protease as well as refolding functions, and both the AAA and the proteolytic domains of FtsH are required for each of these activities.

Membrane proteins play essential roles in cell physiology. They carry out import of nutrients, export of toxins, antibiotics and drugs, and play important roles in energy and signal transduction. Improper assembly of membrane proteins is known to result in various diseases. However, because of the complexity of the assembly process and the diverse array of factors involved, understanding mechanisms of membrane protein assembly poses a serious challenge. The bacterial DrrAB (doxorubicin resistance proteins A and B) system is an attractive model for studying assembly of ABC (ATP-Binding Cassette) transporters. In this system, the catalytic function (DrrA) and the transport function (DrrB) are present on separate subunits (1), which together form a tetrameric complex in the membrane (2) and carry out ATP-dependent efflux of the anti-cancer drugs doxorubicin (Dox) and daunorubicin (3). Previous studies from this lab suggested that for proper function of the DrrAB complex the DrrA and DrrB proteins may be required to co-

assemble (4). It was also shown that interaction between DrrA and DrrB is essential for stable maintenance of DrrB in the membrane so that the expression of DrrB is undetectable in the absence of a simultaneous expression of DrrA (2). Co-expression of DrrA in *cis* or *trans* restores the wild-type levels of DrrB expression, therefore suggesting that DrrA protects DrrB from proteolysis by a cellular protease. The nature of the protease and whether it plays a specific role in quality control and biogenesis of the DrrAB complex has so far remained uncharacterized. In this study, we examine the role of FtsH (filament temperature-sensitive protein H) in this process.

FtsH is a zinc-dependent metalloprotease which belongs to the AAA (ATPases Associated with Diverse Cellular Activities) family of proteins. Along with other proteases, such as ClpAP, ClpXP, HslUV, and Lon, these proteins form the large AAA⁺ superfamily of proteins, members of which share a similar AAA-ATPase domain (5). FtsH is evolutionarily conserved with more than 40% sequence identity observed between bacterial, yeast and human homologs (6). *E. coli* FtsH is the best studied of all known members, and it has been shown to be the only growth-essential protease in *E. coli*. Yeast cells lacking the three FtsH orthologs (two m-AAA and one i-AAA) were also found to be non-viable, demonstrating the essential function of this enzyme in eukaryotic cells (6,7). FtsH is unique in being embedded in the cell membrane in *E. coli*, and in the mitochondrial inner membrane in eukaryotes (8), where it forms homohexameric ring-like structures. The major role of FtsH is believed to be in the quality control of specific membrane proteins, such as degradation of the unassembled SecY and subunit 'a' of F₀ sector of the ATP synthase, in addition to modulating levels of some soluble regulatory proteins (8-10).

FtsH contains two transmembrane helices at the N terminus, followed by a cytoplasmic domain containing the catalytic AAA domain in the middle and the proteolytic domain at the C terminus. The AAA domain (residues 144-398) consists of the conserved Walker A, Walker B and SRH (second region of homology) motifs, which are essential for ATP binding and hydrolysis (8). The proteolytic domain contains the conserved Zn²⁺ binding motif H⁴¹⁷EXXH⁴²¹, the third Zn²⁺-ligand residue E⁴⁷⁹, and the coiled-coil leucine-

zipper sequence (8). FtsH carries out proteolysis of polypeptides in an ATP and Zn²⁺-dependent manner, while other AAA⁺ proteases, such as Lon and ClpA/P, are serine peptidases (11). To initiate proteolysis of a membrane protein, the putative polypeptide binding site in the catalytic AAA domain of FtsH is believed to capture the cytoplasmic tail (either at the N or the C terminus) of the membrane substrate, followed by dislocation and processive unfolding of the protein to an open structure (8). Therefore, both the catalytic AAA domain and the proteolytic domain are required for proteolysis.

Although the major function of the AAA⁺ proteases is in proteolysis, they also exhibit chaperone-like activities, which allow them to monitor the folding status of a protein, promote disassembly or unfolding (12), and specifically degrade non-native proteins (13). For a long time, it has been speculated that the AAA⁺ proteases may also have the ability to refold their substrate proteins (14), which remains an open question till date (15). FtsH was originally identified by Ito and co-workers in a screen to isolate factors which may assist in membrane protein assembly (16). They used SecY-PhoA fusion to screen for stop-transfer defect (Std) mutations, and found that such a mutation lies in the *ftsH* gene. Depletion of FtsH also resulted in significant export defects of β -lactamase and OmpA in *E. coli* in addition to causing a strong Std phenotype. Together, these studies indicated that FtsH may be involved in protein assembly into and through the membrane and may play a role in determining orientation of membrane proteins (16). In another study, FtsH orthologs Yta10 and Yta12 in yeast mitochondria were shown to be required for the formation of a 48 kDa assembly intermediate of the F₀ subunit 9 (17). Finally, *in vitro* studies showed that the purified AAA domain of Yme1, a yeast mitochondrial homolog of FtsH, suppresses aggregation of a model polypeptide (18). Despite these observations, however, no direct evidence for the role of FtsH or its homologs in functional assembly of membrane proteins has been obtained so far.

In this study, we provide the first direct evidence that FtsH is a dual function enzyme containing both the protease and assembly functions. We show that not only is FtsH responsible for removal of the unassembled DrrB

but that it is actually able to refold previously misassembled DrrAB proteins and restore Dox efflux function of the complex. Our results also show that while the AAA domain of FtsH provides recognition and specificity for binding of the substrate, both ATP hydrolysis and the proteolytic functions of FtsH are used concurrently for refolding of DrrAB and restoration of function. Our studies, therefore, not only shed light on the mechanism of assembly of the DrrAB complex but also further elucidate the function of the FtsH protease.

EXPERIMENTAL PROCEDURES

Strains and plasmids – The *E. coli* strains and plasmids are described in Table 1.

Media and growth conditions – *E. coli* cells were normally grown in LB medium at 30°C or 37 °C, unless indicated otherwise. Chloramphenicol, kanamycin, or ampicillin was added to a final concentration of 20, 30 or 100 µg/ml, respectively, where indicated. *E. coli* cells used for the Dox efflux assay were grown in TEA medium (3).

Site-directed mutagenesis of ftsH – Site-directed mutagenesis of the *ftsH* gene was carried out by a Stratagene QuickChange Multisite-directed mutagenesis kit (La Jolla, CA). Using pUC18/*ftsH* plasmid as the template, Lys¹⁹⁸, located in the conserved Walker A motif of the AAA domain, was changed to asparagine. The resulting plasmid was named pUC*ftsH*(K198N). Another mutant, named *ftsH*(HEH), was obtained by substituting His⁴¹⁷Glu⁴¹⁸His⁴²¹ in the conserved HEAGH motif present in the proteolytic domain to Ala⁴¹⁷Gln⁴¹⁸Ala⁴²¹. The primers for both substitutions are shown in Table S1.

Subcloning of the AAA domain of ftsH into pUC18 vector – To completely remove the proteolytic domain, a fragment of *ftsH* corresponding to the first 1194 base pairs was PCR-amplified and ligated into the pUC18 vector using the *EcoRI* and *HindIII* restriction enzymes resulting in pUC18(AAA). This construct is referred to as *ftsH*(AAA) in this article. The primers are shown in Table S1.

Subcloning of ftsH and the groESL genes– Wild-type *ftsH*, *ftsH*(K198N), *ftsH*(HEH) and *ftsH*(AAA) genes were subcloned into pET28a vector using *SnaBI* and *HindIII* restriction enzymes. The resulting plasmids were named

pET*ftsH*, pET*ftsH*(K198N), pET*ftsH*(HEH) and pET*ftsH*(AAA), respectively. The *groES/L* genes were subcloned from pKY326 (19) into pUC18 vector using *EcoRI* and *SmaI* restriction enzymes resulting in pUC*groESL*

Cloning of ftsH into the pBAD vector– Using pBAD/HisA (Invitrogen) as a template, a 3.9kb fragment between *NcoI* and *HindIII* sites was amplified. The *NcoI* site was substituted with the *XhoI* site in the primers used for amplification (Table S1). This resulted in deletion of the multiple cloning sites and the polyhistidine region. The *ftsH* gene was amplified from pUC*ftsH* using primers containing *XhoI* and *HindIII* sites and ligated to the above fragment.

Growth and Protein expression – *E. coli* TG1, AR796, AR797, *E. coli* SG1110, and *E. coli* SG1126 cells containing the indicated plasmids were grown at 30 °C or 37 °C to mid-log phase ($A_{600nm}=0.6$). The proteins were induced by addition of 0.1 or 0.25 mM IPTG, and the incubation was continued at 30 °C, 37 °C, or 42 °C for 3 h. The cells were spun down, resuspended in 5 ml lysis buffer (2 mM Tris-Cl, pH 7.5, 20% glycerol, 2mM EDTA, 1 mM DTT) and lysed by a single passage through a French pressure cell at 20,000 p.s.i. After centrifugation at 10,000 X *g* for 15 min, the pellet represented the inclusion body fraction. The supernatant was centrifuged at 100,000 X *g* for 1 h to separate the supernatant (cytosolic fraction) and the pellet (membrane fraction). The membrane, cytosol and the inclusion body fractions were analyzed by 12% SDS-PAGE, followed by Western-blot analysis using anti-DrrA or anti-DrrB antibodies (2).

Purification of the FtsH protein – FtsH protein was purified according to published protocols (9) with modifications. *E. coli* HMS174(DE3) cells containing the pET*ftsH* plasmid or its variants were inoculated in 1liter LB medium supplemented with 30 µg/ml kanamycin at 37 °C. The cells were grown to mid-log phase and induced with 0.25 mM IPTG at 20 °C overnight. The membrane fraction was prepared as described earlier (2). 5 mg of the membrane fraction was solubilized with 5 ml solubilization buffer (50 mM Tris-Cl, pH7.5, 500 mM KCl, 0.5 % (w/v) Nonidet P-40, 15% (w/v) Glycerol, 2.9 mM 2-mercaptoethanol). The solubilized protein was purified using Ni²⁺-NTA-agarose column and eluted with a gradient of 50 to 500 mM imidazole.

Fractions containing FtsH were collected and dialyzed against 2 liters of the dialysis buffer (10 mM Tris-Cl, pH7.5, 15% (w/v) glycerol, 50 mM KCl, 0.5% (w/v) Nonidet P-40, 5mM MgCl₂, 1 mM dithiothreitol) for 12 hours. The protein was stored at -80 °C until used.

ATPase activity assay – The ATPase activity of FtsH was detected by the malachite green-ammonium molybdate colorimetric assay (9).

In vivo FtsH Proteolytic assay - Membrane-bound DrrAB proteins were used as a substrate to determine the proteolytic activity of FtsH. pBAD vector or the pBAD*ftsH* plasmid was transformed into AR797 cells containing pDX101 (pSU2718*drrAB*). The cells were grown in LB medium to mid-log phase and the DrrAB proteins were induced with 0.25 mM IPTG at 42 °C for 1 h. To terminate the synthesis of DrrAB, 1000 µg/ml chloramphenicol was added to the cell culture and incubated at 42 °C for 30 min. The cells were washed extensively to remove chloramphenicol and IPTG. After resuspending the cells in fresh medium, synthesis of FtsH from the pBAD*ftsH* plasmid was induced by addition of 0.2% arabinose at 42 °C. An aliquot of cell culture was taken out at 0, 30, 60, 90, 120 and 180 min. Membrane fractions were prepared as described above, and 20 µg total membrane protein was loaded onto 12% SDS-PAGE, followed by Western-blot analysis using anti-DrrA, anti-DrrB or anti-FtsH antibodies.

In vitro FtsH Proteolytic assay - α -Casein was used as a substrate *in vitro* to demonstrate the proteolytic activity of purified FtsH and the FtsH(HEH) protein (20). 40 µg of α -casein (Sigma Aldrich) was mixed with 40 µg purified FtsH or FtsH (HEH) in 150 µl protease buffer (50 mM Tris-Cl, pH8.0, 20 mM KCl, 5 mM MgCl₂, 12.5 µM Zn(OAc)₂, 0.5 % (w/v) NP-40, 10 % (w/v) glycerol, 1 mM dithiothreitol) at 42 °C. The reaction was initiated by adding 8 mM ATP, and an aliquot (20 µl) of the sample was taken out at the indicated time points. The reaction was terminated by adding 7 µl of 4×SDS sample buffer. The samples were analyzed by 12% SDS-PAGE, followed by Coomassie Brilliant blue staining.

Whole cell Dox efflux assay – The whole cell Dox efflux assay was carried out according to the protocol published previously (3). The fluorescence spectra were recorded on an Alphascan-2 spectrofluorometer (Photon

Technology International, London, Ontario, Canada). The slope of the Dox efflux curve of the positive control (the first sample) in each panel was designated as 1.0. The efficiency of Dox efflux of each sample within one panel was calculated by dividing the slope of the efflux curve by the slope of sample 1. The average data obtained from three independent experiments were plotted in the histograms

RESULTS

FtsH is responsible for the proteolysis of unassembled DrrB – Previous studies from this laboratory showed that DrrB is undetectable in wild-type *E. coli* membranes in the absence of simultaneous expression of DrrA ((2), Fig 1A, lane 2), however stable expression of DrrB is seen when both DrrA and DrrB are expressed together (lane 1). These results suggest that DrrB is completely degraded when not complexed with DrrA. Review of recent literature suggested that proteins of the AAA⁺ family, especially FtsH, may be involved in the quality control of membrane proteins (13). To determine if this is true for DrrB, three proteases, including Lon, ClpA/ClpP and FtsH, were investigated. If any of these proteases is responsible for degradation of unassembled DrrB, stable expression of DrrB will be observed in cells deficient in that protease as compared to the wild-type cells. The FtsH-deficient *E. coli* AR797 strain (Table 1) used in this study contains a temperature sensitive mutation in the *ftsH* gene, therefore it was grown at 30 °C but the temperature was switched to 42 °C to inactivate FtsH. The Lon⁻ (SG1110) and the ClpA⁻ (SG1126) cells were grown normally at 30 °C or 37 °C. The isogenic *E. coli* AR796 parent strain was used as a control. Of the 3 protease-deficient strains tested, only the FtsH-deficient *E. coli* cells showed stable expression of DrrB in the absence of DrrA (Fig. 1B, lanes 1-3) (Fig. 1C, lane 2), while the Lon⁻ and the ClpA⁻ cells showed no effect on the stability of DrrB (Fig. 1C, lanes 3-4). Since DrrB was not seen in AR796 (wild-type) cells at either 30 °C or 42 °C (Fig. 1B, lanes 1 and 2), but it was stably expressed in the 797 (FtsH^{ts}) cells at 42 °C (lane 3), these results show that FtsH degrades unassembled DrrB. In contrast to the expression of DrrB alone, when both DrrA and DrrB were expressed together in wild-type 796 cells at 30 °C or 42 °C, stable expression of DrrB was seen (Fig. 1B, lanes

4 and 5), confirming that DrrA protects DrrB against FtsH proteolysis. Note that the amount of DrrA and DrrB in the wild-type cells was less at 42 °C as compared to at 30 °C (Fig. 1B, lanes 4-5), suggesting that the DrrAB complex acquires a more open conformation at higher temperature resulting in partial proteolysis by endogenous FtsH. Protection of DrrB from FtsH proteolysis by DrrA was also seen in the ClpA⁻ and Lon⁻ backgrounds (Fig. 1C, lanes 5-6). Together, the data in Fig. 1 show that FtsH plays an important role in quality control of the DrrB protein when DrrA is absent. To rule out the possibility that DrrA or DrrB may aggregate when their expression is induced at 42 °C, the membrane, cytosol, and inclusion body fractions were prepared from both wild type and FtsH^{ts} cells and analyzed by Western blotting using anti-DrrA and anti-DrrB antibodies. The data in Fig. 2 show that although some DrrB protein is present in the inclusion body fraction in both wild type (lower panel, lane 3) and the FtsH^{ts} cells (lane 9) at 30 °C, the induction of either strain at 42 °C did not result in any increase in the amount of inclusion body formation (lower panel, lanes 6 and 12). Moreover, no aggregated DrrAB proteins were seen in the stacking region of the gel in any of the fractions of wild type or FtsH^{ts} cells, indicating absence of any significant aggregation under the conditions used in these experiments. Note that the anti-DrrB antibody is an anti-peptide antibody, therefore it shows some cross-reactivity with epitopes in some other *E. coli* proteins, as explained in a previous publication (1).

Expression of DrrB alone or DrrAB together in FtsH-deficient cells results in growth inhibition- The growth analysis of the wild type and FtsH^{ts} cells expressing DrrB or DrrAB at 30 °C or 42 °C was carried out (Figs. 3A and 3B). The relative growth of various strains at the four hour time point was plotted in a histogram (Fig. 3C). The data in Fig. 3A show that the growth of mutant FtsH^{ts} cells is compromised at 42 °C (Fig. 3A; 797, open diamonds; Fig. 3C, Column 4) as compared to the growth of wild-type cells under similar conditions (Figs. 3A, 796, lines; Fig. 3C, Column 2). This result is not surprising due to the essential nature of *E. coli* FtsH. Interestingly, the expression of DrrB alone in FtsH^{ts} cells at 42 °C further inhibited the growth of these cells (Fig. 3A, compare 797, open diamonds with 797/B, filled triangles; Fig. 3C, Column 7). However, growth

inhibition was not seen when DrrB was expressed in wild-type cells at 42 °C (compare 796, lines with 796/B, open rectangles; Fig. 3C, column 6). Since DrrB accumulates in FtsH^{ts} cells but is proteolysed in wild-type cells (Fig. 1B), it allows us to conclude that the growth defect seen in mutant cells is caused by the accumulation of unassembled DrrB protein. Surprisingly, however, growth inhibition was also seen in the FtsH^{ts} cells expressing DrrA and DrrB together at 42 °C (Fig. 3B, 797/AB, filled triangles; Fig. 3C, Column 7). This effect was unexpected because DrrA and DrrB can be expressed together in the wild-type cells at 42 °C without any negative effect on their growth (Fig. 3B, 796/AB, open rectangles; Fig. 3C, Column 6). These data indicate that the DrrAB proteins expressed in FtsH^{ts} cells at 42 °C may be misfolded, and the accumulation of misfolded membrane proteins results in growth inhibition. It was also observed that if the FtsH^{ts} cells initially induced at 42 °C were shifted down to 30 °C, the cell growth resumed, albeit slowly. After 43 hours of temperature shift-down, the final growth was about half as compared to the cells induced and maintained at 30 °C (data not shown), indicating that the growth inhibition of these cells is quite severe. In summary, the data in Fig. 3 suggest that FtsH is not only responsible for removing unassembled DrrB (in the absence of DrrA) but it may also be critical for proper assembly of the DrrAB complex in the membrane.

The growth defect of the FtsH^{ts} cells expressing DrrB or DrrAB at 42 °C could be rescued by overexpression of FtsH in *trans* (Figs. 3A-B, open triangles; Fig. 3C, Column 8), indicating that the absence of functional FtsH was solely responsible for this defect. The growth defect in each case was also suppressed by overexpression of the chaperone GroESL (Figs. 3A-B, filled circles; Fig. 3C, Column 9) but not to the same extent as seen with FtsH. Western blot analysis of the membrane fractions (prepared from the four hour cultures of samples # 6-9 in Figs. 3A-B) showed that while FtsH overexpression resulted in significant proteolysis of DrrAB (Fig. 3D, lanes 3 and 7), overexpression of GroESL did not (lanes 4 and 8). This might imply that FtsH restores growth by simply removing misfolded DrrAB proteins, while GroESL is able to alter their conformation, thus alleviating growth inhibition.

FtsH preferentially proteolyzes misfolded DrrAB - To determine if FtsH discriminates between properly assembled and misfolded DrrAB, the effect of overexpression of FtsH proteolysis was compared in wild type or FtsH^{ts} cells at 30 °C or 42 °C. Interestingly, overexpression of FtsH in either wild type (Figs. 4A-B, compare lanes 1 and 3) or FtsH^{ts} cells (lanes 2 and 4) produced no significant proteolysis of DrrA and DrrB expressed at 30 °C, showing that FtsH does not proteolyze properly assembled DrrAB. However, when the DrrAB proteins were expressed in wild-type cells at 42 °C, simultaneous overexpression of FtsH resulted in significant proteolysis (Figs. 4A-B, compare lanes 3 and 7). These data suggest that the DrrAB proteins must acquire a partially unfolded conformation at a higher temperature (as also seen in Fig. 1B), thus making them more susceptible to proteolysis by over-expressed FtsH. As expected, overexpression of FtsH in FtsH^{ts} cells also showed significant proteolysis of DrrAB expressed at 42 °C (Figs. 4A-B, lanes 4 and 8). This is consistent with the data in Fig. 3, which suggested that the DrrAB proteins expressed at 42 °C in FtsH^{ts} cells are misfolded. (Please note that the conformation of the DrrAB proteins in wild-type cells at 42 °C is completely different from the DrrAB expressed in mutant FtsH^{ts} cells at 42 °C even though they are both sensitive to overexpressed FtsH. In a later experiment in Fig. 8, it is shown that the DrrAB proteins expressed in wild type cells at 42 °C retain normal function, while the DrrAB expressed in FtsH^{ts} cells are inactive due to their misfolding)

The rate of proteolysis of misfolded DrrAB by FtsH was analyzed in a separate time-course experiment. The misfolded DrrAB proteins were first allowed to accumulate in the membranes of FtsH^{ts} cells by induction with IPTG for 1 hour at 42 °C. Chloramphenicol was added to stop further synthesis, as described under Methods. The synthesis of FtsH from pBAD_ftsH was induced by addition of arabinose, and the proteolysis of DrrA and DrrB by FtsH was determined by Western blot analysis. The data in Figs. 4C and 4D (filled circles) show that synthesis of FtsH (Fig. 4E, lower panel) resulted in increasing proteolysis of misfolded DrrA and DrrB from the membrane. At 120 minutes after addition of arabinose, about 75-80% of DrrA and DrrB were removed from the membrane. These observations are in agreement

with the dislocation model proposed previously for the activity of FtsH (8). No significant proteolysis of DrrAB was seen in the absence of FtsH synthesis (Figs. 4C and 4D, filled rectangles).

The AAA domain of FtsH contains a chaperone-like activity, but it is not sufficient by itself to restore the Dox efflux function - The growth experiment in Fig. 3B suggested that FtsH may be critical for the assembly of the DrrAB complex. Further support for this idea was obtained by comparing the rate of assembly of the DrrAB complex in wild type and FtsH^{ts} cells. The data in Fig. 5 show that the assembly of DrrA and DrrB in the membrane of FtsH^{ts} cells is significantly compromised already at 30 °C as compared to in the wild type cells. A significant difference in the amounts of DrrAB in the membrane of wild type and mutant cells was seen at all time points tested (Fig. 5, compare lanes 1-3 with 7-9). However, this difference is most evident at the early time points, which suggests that the rate of assembly of DrrAB is affected by FtsH. This is most likely due to the partial defect of FtsH function in FtsH^{ts} cells already at 30 °C, leading to the low efficiency of the DrrAB complex formation.

To determine if the ability to promote assembly of DrrAB resides in the AAA domain of FtsH, variants containing mutations in the Walker A motif of the AAA domain (K198N mutation) or the conserved amino acids in the proteolytic domain (the HEH mutation and the AAA subclone are described under Methods) were created. The K198N allele contained a defective AAA domain, while the HEH mutant and the AAA subclone contained an intact AAA domain. As expected, the K198N mutation resulted in a significantly reduced ATPase activity, however the HEH mutant and the AAA subclone were unaffected (Table 2). The *in vitro* proteolytic activity assay showed that while the wild-type FtsH completely proteolyzed α -casein in one hour, no significant reduction in the α -casein level was seen with the HEH mutant even after two hours of incubation (Fig. 6). These analyses confirmed that the AAA and the proteolytic domain mutants behave as expected. Therefore, they were used in two different complementation experiments (described below) to determine if the AAA domain by itself is sufficient for promoting assembly of the DrrAB complex.

The expression of the DrrAB proteins in FtsH^{ts} cells was previously shown to result in severe growth inhibition (Fig. 3B, filled triangles). This inhibition was reversed by simultaneous expression of FtsH (Fig. 3B, open triangles). In the next experiment, we asked whether co-expression of the HEH allele or the AAA subclone can rescue FtsH^{ts} cells from the growth inhibition resulting from DrrAB expression. The data in Fig. 7 show that the simultaneous expression of either the HEH mutant (Fig. 7A, open circles) or the AAA subclone (Fig. 7A, filled triangles) with DrrAB can complement the growth defect of FtsH^{ts} cells, indicating that the AAA domain of FtsH indeed contains a chaperone-like activity. Interestingly, the HEH mutant showed much better complementation of the growth defect as compared to the AAA subclone, perhaps due to a more native conformation of the full-length HEH protein as compared to the AAA subclone. The K198N mutation, on the other hand, showed no growth complementation effect (Fig. 7A, open triangles), showing that the ATPase activity associated with the AAA domain is important for the chaperone function of FtsH. Western blot analysis showed that the DrrAB complex is membrane-associated in the FtsH(HEH), FtsH(AAA), and FtsH (K198N)-containing strains (Figs. 7B and 7C, lanes 5-7), and the amounts of DrrA and DrrB in these cells were comparable to the levels in their absence (lane 3). Therefore the restoration of growth by the HEH and AAA clones must result from a change in conformation of the DrrAB proteins brought about by the functional AAA domain present in these two clones. Whether the HEH and AAA variants of FtsH can also restore function of the DrrAB complex is addressed in the next experiment.

We previously showed that the wild-type DrrA and DrrB proteins together carry out ATP-dependent efflux of the anticancer drug doxorubicin (3). Here, we investigated whether co-expression of the AAA subclone or the HEH allele can restore the Dox efflux function of the misassembled DrrAB proteins expressed in FtsH^{ts} cells at 42 °C (Fig. 8). The data in Fig 8A indicate that the rate of DrrAB-mediated Dox efflux at 30 °C in the wild-type 796 and FtsH^{ts} 797 cells is comparable (Columns 1 and 3). Induction of wild-type cells at 42 °C showed only a slight reduction in Dox efflux (Column 2), which is

likely due to the destabilization effect produced by high temperature on DrrAB, as seen earlier in Figs. 1 and 4. By contrast, the FtsH^{ts} cells induced at 42 °C showed very little or no DrrAB-mediated Dox efflux (Fig. 8A, column 4), which was comparable to the efflux seen with control cells containing empty vector (Fig. 8A, Columns 5 and 6). These data confirm that the DrrAB proteins are misfolded in the absence of functional FtsH. Simultaneous overexpression of the FtsH variants, K198N, HEH or the AAA subclone, in FtsH^{ts} cells did not restore function of misassembled DrrAB (Fig. 8B, columns 4-6), even though restoration of growth by HEH and the AAA subclone was earlier seen in Fig. 7A. Similarly, overexpression of GroESL also did not complement the DrrAB-mediated Dox efflux in FtsH^{ts} cells (Fig. 8B, column 7). One possible explanation for these data could be that even though the AAA domain of FtsH (or GroESL) is able to alter the conformation of the DrrAB proteins and relieve growth inhibition, it is not sufficient by itself to restore proper conformation required for full function of the complex.

Wild-type FtsH can refold previously misassembled DrrAB and restore function - Surprisingly, co-expression of wild-type FtsH restored the Dox efflux function of the misassembled proteins expressed in FtsH^{ts} cells at 42 °C resulting in a significant recovery (about 45%) of Dox efflux by the DrrAB complex (Fig. 8B; compare columns 2 and 3). Since this could only have resulted if FtsH expressed in *trans* facilitated assembly of the complex, these results imply that the AAA and the proteolytic domains of FtsH must work hand-in-hand to bring about functional assembly of the DrrAB complex. In this experiment, however, FtsH and the DrrAB proteins were expressed simultaneously by IPTG induction, therefore it was not possible to determine if FtsH assists only the newly synthesized DrrAB to achieve proper conformation, or if it can also bring about refolding of the DrrAB proteins that have already been misfolded.

Therefore, in the next experiment, DrrAB and wild-type FtsH were expressed in FtsH^{ts} cells in a sequential manner. The *ftsH* gene subcloned under the control of the *araBAD* promoter was induced by arabinose, while the *drrAB* genes remained under the control of the *lac* promoter

induced by IPTG. The FtsH^{ts} cells containing both the plasmids (797/AB/pBADftsH) were grown at 30 °C to mid-log phase. The DrrAB proteins were first induced with IPTG at 42 °C for 1 hour (this condition inactivates FtsH and renders DrrAB inactive as seen in Figs. 8A and 8B). The cells were then washed several times to remove extracellular IPTG and stop further synthesis of DrrAB. The expression of FtsH was induced by arabinose for 1 hour at 42 °C, and the cells were subjected to the Dox efflux assay. To maintain the chromosomally-encoded FtsH in an inactive conformation, the temperature was maintained at 42 °C throughout the duration of the experiment. As previously seen in Fig. 8B, simultaneous expression of DrrAB and FtsH resulted in restoration of Dox efflux (Fig. 8C; 797/AB/pUCftsH, column 3). More interestingly, however, even greater restoration of the DrrAB-mediated Dox efflux was seen when FtsH was induced after DrrAB proteins had been pre-synthesized in these cells (Fig. 8C, 797/AB/pBADftsH+ara, column 5). In the absence of arabinose induction of FtsH, much lower restoration of Dox efflux was seen (Fig. 8C, 797/AB/pBADftsH – ara, column 4). These data, therefore, show that the sequential expression of DrrAB and FtsH can still restore the function of the previously misfolded DrrAB proteins to the same (or even higher) extent as seen with simultaneous expression.

Finally, the effect of over-expression of FtsH on Dox efflux function of DrrAB expressed at different temperatures was investigated. Irrespective of whether the DrrAB proteins were induced at 30 °C, 37 °C, or 42 °C in FtsH^{ts} cells, simultaneous expression of FtsH resulted in a very similar final Dox efflux efficiency (Fig. 8D). At 42 °C, co-expression of FtsH enhanced Dox efflux of misfolded DrrAB 5-fold (compare columns 3 and 6), yielding about 45% Dox efflux efficiency. Interestingly, over-expression of FtsH at 30 °C or 37 °C reduced the efficiency by about half (compare columns 1-2 with 4-5), once again yielding final Dox efflux efficiency of about 45%. These results imply that FtsH produces an optimal level of functional complexes in the membrane perhaps by exerting both proteolytic and refolding effects concurrently.

DISCUSSION

Non-native proteins, especially unassembled membrane proteins, interfere with cellular processes and are known to become toxic to the cells. Therefore, quality control systems, consisting of chaperones and proteases, play essential roles by monitoring their folding and either refolding or degrading misfolded proteins (21-23). Hsp60 (GroEL/ES) and Hsp70 (DnaK) proteins provide classical examples of ATP-dependent chaperones which prevent aggregation of newly translated proteins and promote their refolding (22). A special class of chaperones (e.g. ClpB in bacteria and its homologs Hsp78 and Hsp104 in eukaryotes) is known to resolubilize protein aggregates and, in cooperation with the Hsp70 chaperones (specifically DnaKJE), can result in regaining function of the affected protein (15). On the other end of the spectrum are proteins classically defined as proteases, for example Lon, ClpA/P, ClpX/P and FtsH, whose major function is considered to be removal of irreversibly damaged proteins from the cell (6,14,24). Despite their differences, however, both classical chaperones and proteases share common features. For example, both have the ability to recognize and bind non-native polypeptides and both bring about unfolding of their substrates, which are subsequently refolded (by a chaperone) or degraded (by a protease)(12,22,25). Because of the ATP-dependent unfolding function of AAA⁺ proteases, it has been speculated that they may also have the ability to refold substrate proteins and may participate in protein biogenesis. However, very little direct evidence is available for the role of FtsH or other AAA⁺ proteases in biogenesis, especially of membrane protein complexes. In this study, we provide clear evidence that the *E. coli* FtsH is able to both degrade and refold misassembled DrrAB proteins, resulting in regaining the Dox efflux function of the membrane complex.

We show that in the absence of the DrrA protein, DrrB acquires an FtsH-sensitive conformation and is completely proteolyzed. However, in the absence of functional FtsH, the DrrB protein accumulates even in the absence of DrrA confirming that FtsH monitors the folding status of DrrB and removes it if it is improperly assembled. The molecular details of proteolysis of

DrrB by FtsH are currently unknown, however based on the prevalent model for its action (8,26) we assume that FtsH could initiate proteolysis of DrrB either at the N- or the C-terminal end (both of which are found in the cytoplasm, (27)). Cross-linking studies previously showed that the N terminus of DrrB is the major site of interaction with DrrA (3,27); therefore we propose that proteolysis of DrrB initiates at its N-terminal tail, and binding of DrrA to this region of DrrB protects it from proteolysis by FtsH.

Interestingly, we found that the function of FtsH is not limited to proteolysis of unassembled DrrB, but it also plays an essential role in folding and assembly of the DrrAB complex. This conclusion is supported by several lines of evidence presented in this paper. First, the expression of DrrA and DrrB together in FtsH^{ts} cells at 42 °C (which results in inactivation of FtsH) was found to be growth inhibitory (Fig. 3B) suggesting that the complex is improperly assembled in the absence of a functional FtsH. Second, the rate of assembly of the DrrAB complex in the FtsH^{ts} cells at 30 °C was found to be significantly reduced as compared to the wild-type cells (Fig. 5). Third, functional analysis showed complete absence of the DrrAB-mediated Dox efflux in FtsH^{ts} cells under conditions of FtsH inactivation, suggesting that most or all of the DrrAB proteins expressed in these cells at 42 °C are misassembled. By contrast, the DrrAB proteins expressed in wild-type cells at 42 °C retained on average 85-90% of the Dox efflux activity (Fig. 8A). Finally, co-expression of FtsH in *trans* in FtsH^{ts} cells restored the ability of DrrAB to carry out Dox efflux, confirming that FtsH facilitates assembly of the DrrAB complex (Fig. 8B). Nevertheless, this result was surprising because FtsH contains a functional proteolytic domain. It's overexpression in FtsH^{ts} cells results in proteolysis of the misfolded DrrAB proteins (as seen in Fig. 4), however the data in Fig. 8B show that FtsH also facilitated some folding resulting in about 45% recovery of the Dox efflux activity. Either the AAA domain by itself or the GroESL chaperone was unable to complement the Dox efflux function of DrrAB in FtsH^{ts} cells, even though each was able to alleviate the growth defect. Therefore, together these data suggest that both the AAA and the protease domains of FtsH

are essential for promoting functional assembly of DrrAB.

The most crucial evidence for the refolding function of FtsH, however, came from the sequential expression studies. Irrespective of whether FtsH was expressed simultaneously with DrrAB or expressed after the non-functional DrrA and DrrB proteins had already accumulated in FtsH^{ts} cells, it was able to restore the function of the complex (Fig. 8C), thus showing conclusively that FtsH not only facilitates assembly of the DrrAB complex but it is also actively involved in refolding previously misassembled DrrAB proteins. Interestingly, we also found that the sequential expression of DrrAB and FtsH resulted in a significantly higher recovery of the Dox efflux function of DrrAB as compared to simultaneous expression (Fig. 8C). This finding suggests that FtsH treats its substrate differently during its synthesis as compared to after it has already been synthesized.

In summary, our studies confirm that the AAA domain of FtsH can recognize and bind substrates and change their conformation, which is in agreement with the previous studies (18). However, we also show that the two activities (ATPase and proteolytic) of FtsH must be present simultaneously and occur in a coordinated manner to facilitate assembly and refolding of DrrAB. Much more extensive analysis will be required in the future to understand the nature of the molecular processes involved in refolding of DrrA and DrrB and to determine if other factors also play a role in the assembly of the DrrAB complex. Further studies will also provide clues about how degradation and assembly of multi-subunit complexes are regulated, and whether other AAA+ proteases may also contain chaperone activity. This study raises intriguing questions about the distinction between classical chaperones like GroESL (that can prevent aggregation of many proteins) and the classical proteases like FtsH that not only carry out proteolysis but also actively participate in refolding of their specific substrates, as shown in this study. Bukau and Mogk previously (22) coined four terms to describe the various activities of chaperones and proteases: Holders (small heat shock proteins, Hsps), Folders (GroESL and DnaK), Unfolders (ClpA, ClpX, and ClpB), and Proteases (Lon, ClpP, and FtsH). In light of the findings reported in this article, we

propose a new term 'Specific Refolder' to describe the function of FtsH and possibly other AAA+

proteases that may be shown in the future to contain such an activity.

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FOOTNOTES

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The abbreviations used are: Dox, doxorubicin; AAA, ATPase Associated with diverse cellular Activities; ABC, ATP-Binding Cassette

FIGURE LEGENDS

FIGURE 1. Role of DrrA and FtsH in stable maintenance of DrrB. (A) DrrA is required for stable expression of DrrB. Wild-type *E. coli* (TG1) cells expressing DrrAB (pDX101) or DrrB alone (pDX103) were grown at 37° C to mid-log phase ($A_{600nm}=0.6$) and induced with 0.25 mM IPTG for 3 h. 30 µg membrane proteins were loaded onto 12% SDS-PAGE gels, followed by Western blotting against anti-DrrA (upper panel) and anti-DrrB (lower panel) antibodies. **(B) FtsH is responsible for proteolysis of unassembled DrrB.** Wild-type *E. coli* (796) or the *E. coli* 797 (*ftsH*^{ts}) cells expressing DrrAB (pDX101) or DrrB alone (pDX103) were grown at 30 °C to mid-log phase and induced with 0.25 mM IPTG at either 30 °C or 42 °C for 3 h. Western blot analysis of the membrane fraction was carried out as in Fig. 1A. **(C) ClpA/P or the Lon protease is not involved in quality control of DrrB.** Wild-type *E. coli* (796), *E. coli* 797 (*ftsH*^{ts}), *E. coli* SG1110 (Lon⁻), and *E. coli* SG1126 (ClpA⁻) cells expressing DrrAB (pDX101) or DrrB alone (pDX103) were grown at 30 °C or 37 °C to mid-log phase and induced with 0.25 mM IPTG at either 42 °C or 37 °C for 3 h, as indicated. Analysis was carried as described in (B) above.

FIGURE 2. Western blot analysis of the cytosol, membrane, and inclusion body fractions of *E. coli* 796 or 797 cells expressing DrrAB at 30 °C or 42 °C. Wild-type *E. coli* 796 or the *E. coli* 797 (*ftsH*^{ts}) cells expressing DrrAB (pDX101) were grown at 30 °C to mid-log phase and induced with 0.25 mM IPTG at either 30 °C or 42 °C for 3 h. Cell fractions were prepared, as described under Methods. 20 µg of each fraction was loaded onto 12% SDS-PAGE gels, followed by Western blotting against anti-DrrA (upper panel) and anti-DrrB (lower panel) antibodies. (m, membrane; c, cytosol; IB, inclusion body). Note that the anti-DrrB antibody is an anti-peptide antibody, therefore it shows some cross-reactivity with epitopes in other *E. coli* proteins, as explained in a previous publication (1).

FIGURE 3. Growth inhibition resulting from expression of DrrB alone or DrrAB together in *E. coli* 797 cells can be relieved by the overexpression of FtsH or GroESL. (A) Effect of overexpression of FtsH or GroESL on growth of 797 cells expressing DrrB. 797 cells containing pDX103 (DrrB) and pUC*ftsH* or pUC*groESL* were grown at 30 °C and induced at 42 °C, as in Fig. 1B. The growth was monitored at O.D. 600 nm for four hours after induction. **(B) Effect of overexpression of FtsH or GroESL on growth of 797 cells expressing DrrAB.** 797 cells containing pDX101 (DrrAB) and pUC*ftsH* or pUC*groESL* were analyzed, as described in (A) above. **(C) Quantitation of the growth of 796 or 797 cells expressing DrrB or DrrAB at 4 hours after induction.** The growth of the sample #1 at 4 hours in Fig. 3A or 3B was designated as 1.0. The relative growth of each culture at 4 hours was calculated. The histogram represents an average of 3 experiments. Columns 1-9 correspond to the cultures 1-9 in Figs 3A (filled) and 3B (open). **(D) Western blot analysis.** The membrane fractions were

prepared from sample #s 6-9 collected at the 4 hour time point in Fig. 3A and 3B. Western blot analysis was carried out as in Fig. 1(A).

FIGURE 4. FtsH preferentially degrades misfolded DrrAB. (A) *E. coli* 796 or 797 cells containing pDX101(DrrAB) and pUC*ftsH* were grown and analyzed by Western blotting, as in Fig. 1A. (B) **Quantitative analysis of the amounts of DrrA and DrrB.** The intensity of the bands on the nitrocellulose membrane in Fig. 4A was determined by densitometric scanning. The intensity of DrrA and DrrB in lane 1 was designated as 1. The data represent an average of 3 experiments. Columns 1-8 correspond to the lanes 1-8 in Fig. 4A. (C), (D) and (E) **In vivo FtsH proteolytic assay.** The assay was carried out as described under Methods. The membrane fractions were prepared from the cells collected at the indicated time points after addition of arabinose. 20 μ g total membrane protein was loaded onto 12% SDS-PAGE, followed by Western blot with anti-DrrA, anti-DrrB, or anti-FtsH antibodies. The intensity of the bands on the nitrocellulose membrane was determined by densitometric scanning. The intensity of DrrA and DrrB on the membrane at 0 min was designated as 1.0. (C) Quantitation of DrrA in the membrane at the indicated time points after addition of arabinose. The data shown represent an average of 3 experiments. (D) Quantitation of DrrB in the membrane. (E) Western blot analysis showing amount of FtsH present in the membrane at various times after addition of arabinose. Top panel: cells containing pBAD vector only. Bottom panel: cells containing the plasmid pBAD*ftsH*.

FIGURE 5. FtsH promotes assembly of the DrrAB complex. (A) *E. coli* 796 or 797 cells expressing DrrAB were grown at 30 °C to mid-log phase and the culture was divided into two halves. One half was induced with 0.25 mM IPTG at 30 °C (lanes 1-3 for 796 and lanes 7-9 for 797) and the other half was induced at 42 °C (lanes 4-6 for 796 and lanes 10-12 for 797). Aliquots of each sample were collected at 30 min intervals. Hours of induction are shown at the top of the gel. 0 hour represents $A_{600nm} = 0.6$ at which time IPTG was added. Western blot analysis was carried out as in Fig. 1A. (B) **The quantitative analysis of DrrA and DrrB.** The intensity of bands on the nitrocellulose membrane was determined by densitometric scanning. The intensity of DrrA and DrrB in lane 1 was designated as 1. The data represent an average of 3 experiments.

FIGURE 6. In vitro digestion of α -casein by purified FtsH or FtsH(HEH). (A) α -casein was mixed with purified wild-type FtsH or FtsH (HEH) protein in the protease buffer as described in the Methods. The samples were analyzed by 12% SDS-PAGE, followed by Coomassie Brilliant blue staining. (B) **Quantitative analysis of the amounts of α -casein.** The intensity of the bands on the nitrocellulose membrane was determined by densitometric scanning. The intensity of α -casein in lane 1 was designated as 1. The intensities represent an average of 3 experiments.

FIGURE 7. The AAA domain of FtsH is sufficient to complement the growth defect resulting from the expression of DrrAB in 797 cells. *E. coli* 797 cells containing the *drrAB* genes (pDX101) and the wild-type or a mutated version (K198N, HEH, or AAA subclone) of the *ftsH* gene on a compatible plasmid were grown at 30 °C and induced at 42 °C, as in Fig. 1B. (A) **Growth analysis at 600 nm.** (B) **Western blot analysis.** Membranes prepared from the cells collected at the 3 hour time point were analyzed, as in Fig. 1A. (C) **Quantitative analysis of DrrA and DrrB.** The intensity of the bands on the nitrocellulose membrane in Fig. 7B was determined by densitometric scanning. The intensity of DrrA and DrrB in lane 1 was designated as 1.0. The data shown represent an average of 3 experiments. The columns 1-8 correspond to samples 1-8 in Fig. 7B.

FIGURE 8. DrrAB-mediated Dox efflux. (A) **Dox efflux in *E. coli* 796 or 797 cells.** The cells containing pDX101 (DrrAB) or the empty vector were grown at 30 °C to mid-log phase and induced with 0.1 mM IPTG at either 30 °C or 42 °C for 1 h. The washed cells were loaded with Dox, and efflux was initiated by addition of glucose and detected fluorometrically. The slope of the efflux curve of sample 1 in each panel was designated as 1.0. The efficiency of Dox efflux was then calculated by dividing the

slope of each efflux curve by the slope of sample 1. The average data obtained from three independent experiments are shown in the histograms. **(B) Complementation of the DrrAB-mediated Dox efflux in 797 cells by simultaneous overexpression of wild-type FtsH or its variants.** *E. coli* 797 cells containing pDX101(DrrAB) and pUC*ftsH* (wild-type FtsH), pUC*ftsH*(K198N), pUC*ftsH*(HEH), pUC*ftsH*(AAA) or pUC*groESL*, were analyzed as described under (A) above. **(C) Effect of sequential expression of DrrAB and FtsH on complementation of Dox efflux.** *E. coli* 797 cells containing pDX101 (DrrAB) and pBAD*ftsH* were grown at 30 °C to mid-log phase and induced with 0.1mM IPTG for 1 h at 42 °C. The cells were washed extensively to remove IPTG and divided into two halves. One half was kept at 42 °C for 1 h without any induction (-ara, 42 °C) and the other half was induced by 1% arabinose at 42 °C (+ara, 42 °C) for 1 h. As controls, 796/AB, 797/AB and 797/AB/pUC*ftsH* were grown at 30 °C to mid-log phase and induced with 0.1 mM IPTG at 42 °C for 2 h. The cells were loaded with Dox, and efflux was measured. The data were analyzed as described under (A) above. **(D) Effect of simultaneous over-expression of FtsH on Dox efflux by DrrAB proteins expressed at different temperatures.** 797 cells containing pDX101 (DrrAB) and pUC*ftsH* were grown at 30 °C to O.D. 0.6, induced with 0.1 mM IPTG at 30 °C, 37 °C or 42 °C for 1 h, and analyzed as in (A) above.

TABLE 1. Bacterial strains, plasmids, and antisera

Name	Description	Reference or Source
Bacterial Strains		
TG1	K-12 $\Delta(lac-pro)$ <i>supE thi hsdD5/F' tra36 proA⁺B⁺ lacI^q lacZΔM15</i>	(28)
HMS174(DE3)	F ⁻ <i>recA1 hsdR (rK12- mK12+)</i> (DE3) (Rif ^R)	(28)
AR796	F ⁻ , <i>zhd-33::Tn10, araD139, (argF-lac)U169, rpsL150, rpsE, relA1, flbB5301, deoC1, ptsF25, rbsR, zgj3198::Tn10kan.</i>	(29)
AR797	F ⁻ , <i>zhd-33::Tn10, araD139, (argF-lac)U169, rpsL150, rpsE, relA1, flbB5301, deoC1, ptsF25, rbsR, zgj3198::Tn10kan. ftsH1</i>	(29)
SG1110	$\Delta lon-510, zba-1091:: \Delta Tn10(\lambda)$	(30)
SG1126	$\Delta arg, clpA319:: \Delta kan$	(30)
Plasmids		
pSU2718	Cloning vector, pACYC184 derivative, Cm ^r	(1)
pKY326	<i>groES/groEL</i> in pHSG575, Cm ^r	(19)
pDX101	<i>drrAB</i> in pSU2718, Cm ^r	(1)
pDX103	<i>drrB</i> in pSU2718, Cm ^r	(1)
pUC$ftsH$	<i>ftsH</i> in pUC18, Amp ^r , restriction sites: <i>EcoRI, HindIII</i>	This study
pUC$ftsH$(K198N)	pUC18 containing <i>ftsH</i> with mutation of Lys ¹⁹⁸ to Asn, Amp ^r	This study
pUC$ftsH$(HEH)	pUC18 containing <i>ftsH</i> with mutations of His ⁴¹⁷ Glu ⁴¹⁸ His ⁴²¹ to Ala ⁴¹⁷ Gln ⁴¹⁸ Ala ⁴²¹ , Amp ^r	This study
pUC$ftsH$(AAA)	pUC18 containing <i>ftsH</i> with deletion of proteolytic domain, Amp ^r	This study
pUC$groESL$	<i>groES/groEL</i> in pUC18, Amp ^r , restriction sites: <i>NdeI, HindIII</i>	This study
pET$ftsH$	<i>ftsH</i> in pET28a with C-terminal histag, Kan ^r	This study
pETF$ftsH$(K198N)	<i>ftsH</i> (K198N) in pET28a with C-terminal histag, Kan ^r	This study
pETF$ftsH$(HEH)	<i>ftsH</i> (HEH) in pET28a with C-terminal histag, Kan ^r	This study
pETF$ftsH$(AAA)	<i>ftsH</i> (AAA) in pET28a with C-terminal histag, Kan ^r	This study
pBAD$ftsH$	<i>ftsH</i> in pBAD/HisA, Amp ^r	This study

TABLE2. The ATPase activity of purified FtsH and its variants

The proteins were purified, and their ATPase activity was determined as described in Methods.

FtsH Variant	ATPase Activity (nmol Pi/min/mg)
Wild-type FtsH	139.5
FtsH(K198N)	18.1
FtsH(HEH)	147.8
FtsH(AAA)	155.6

Figure 1

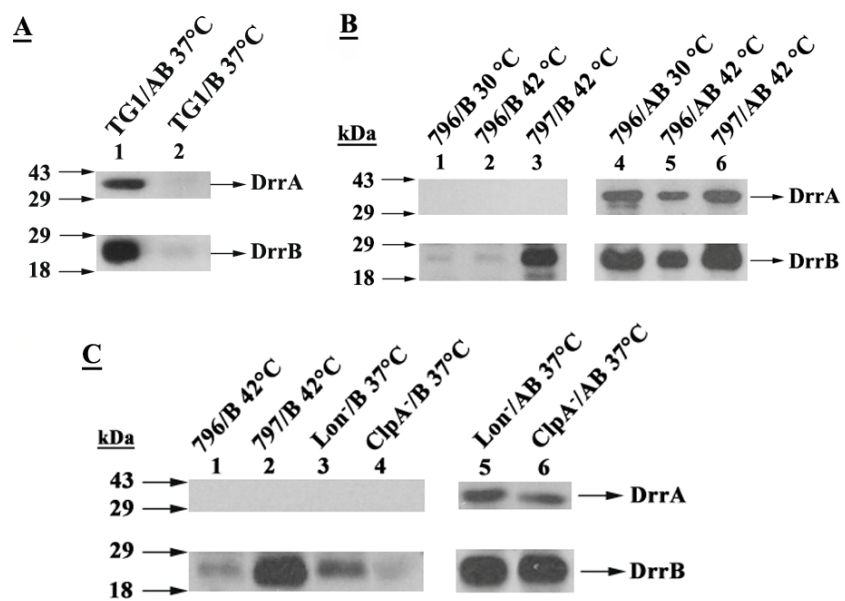


Figure 2

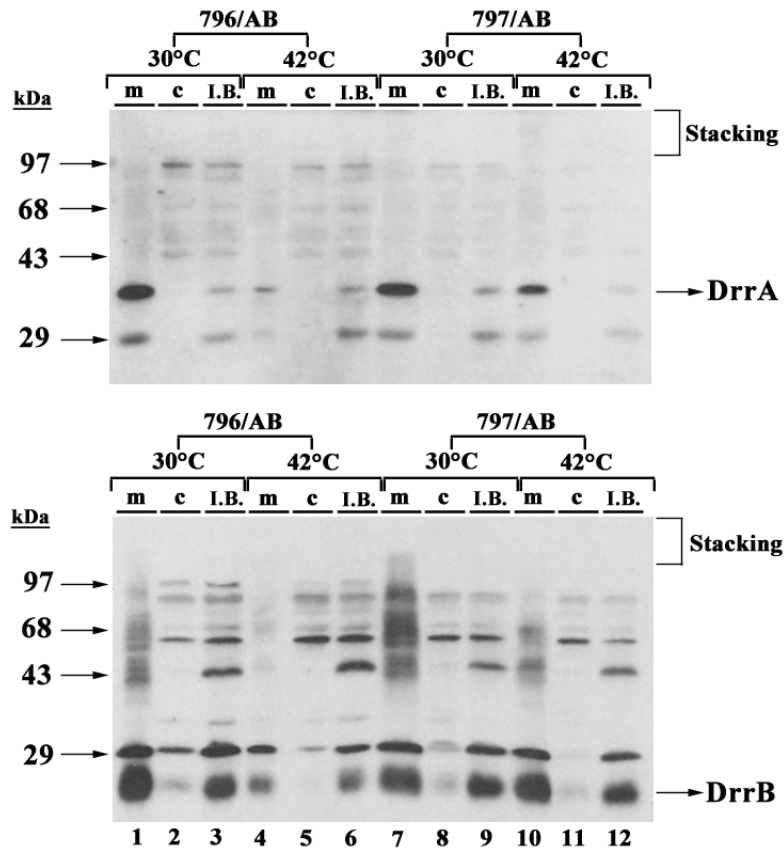


Figure 3

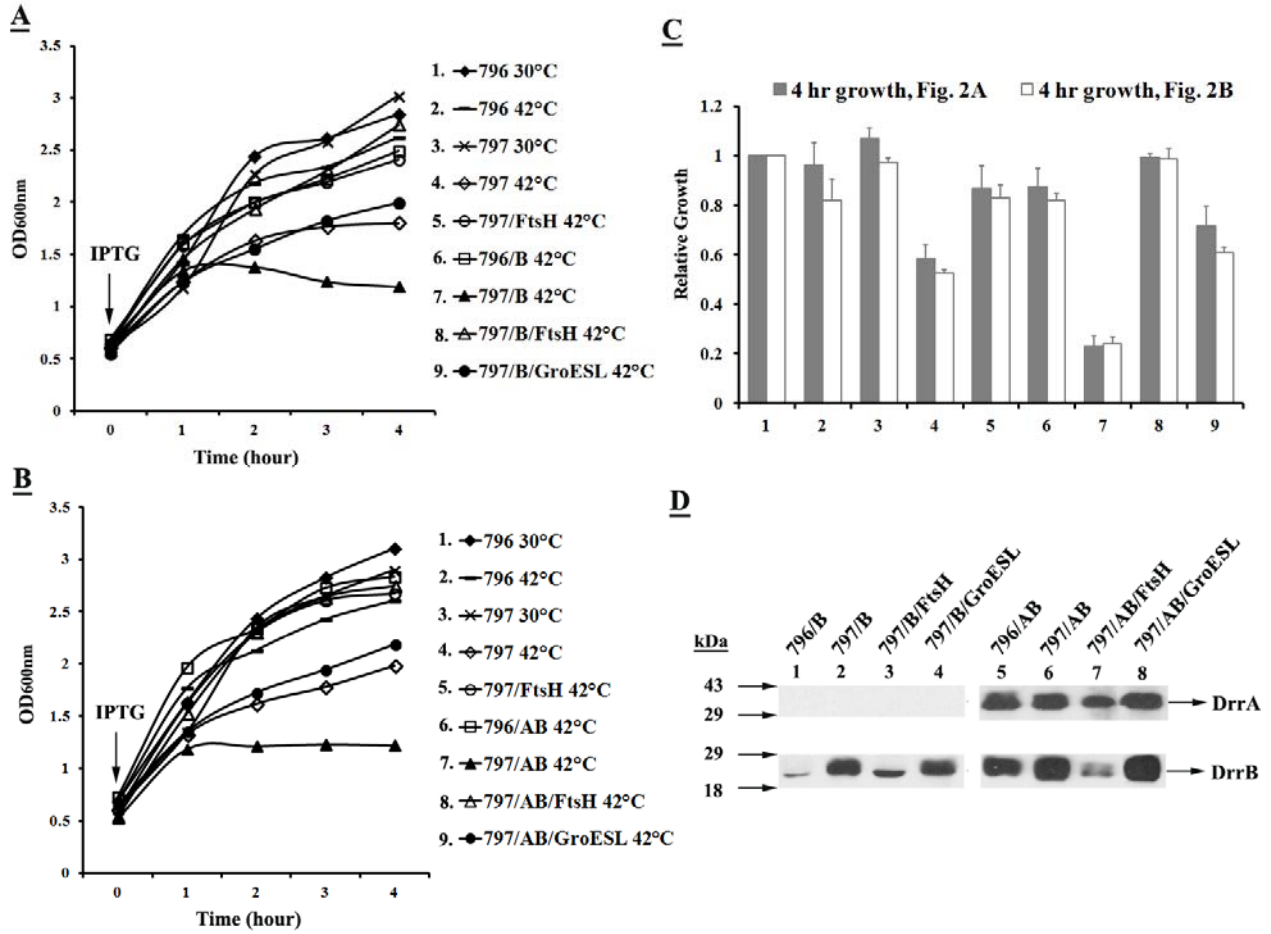


Figure 4

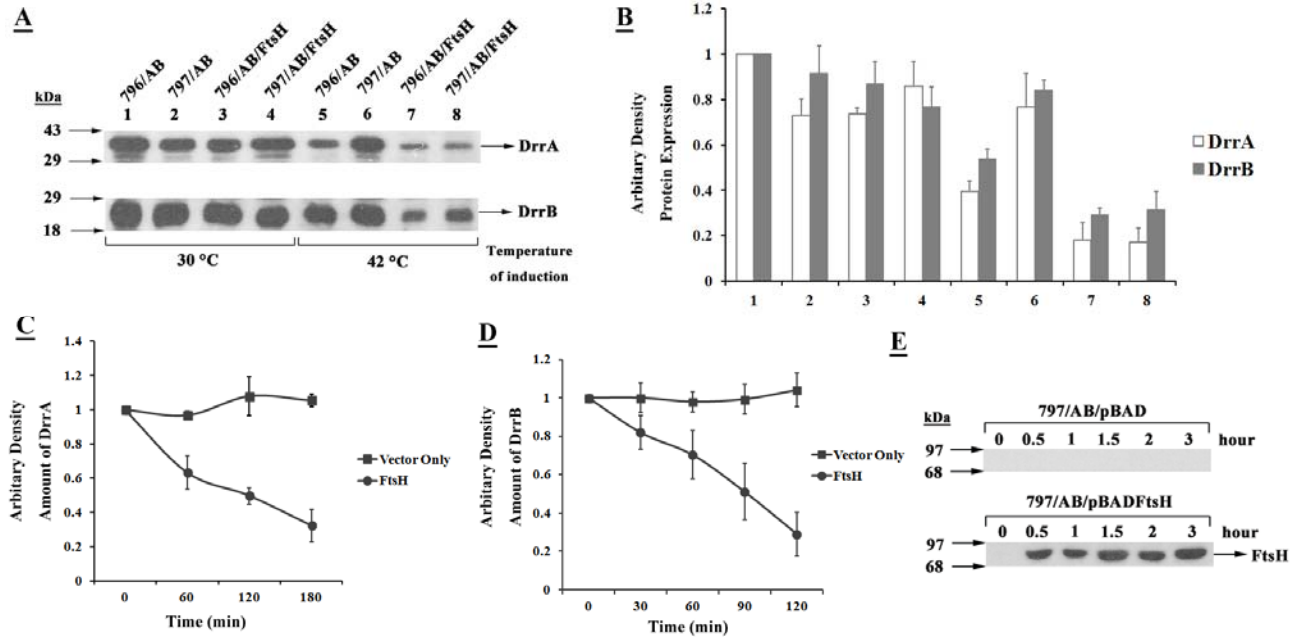


Figure 5

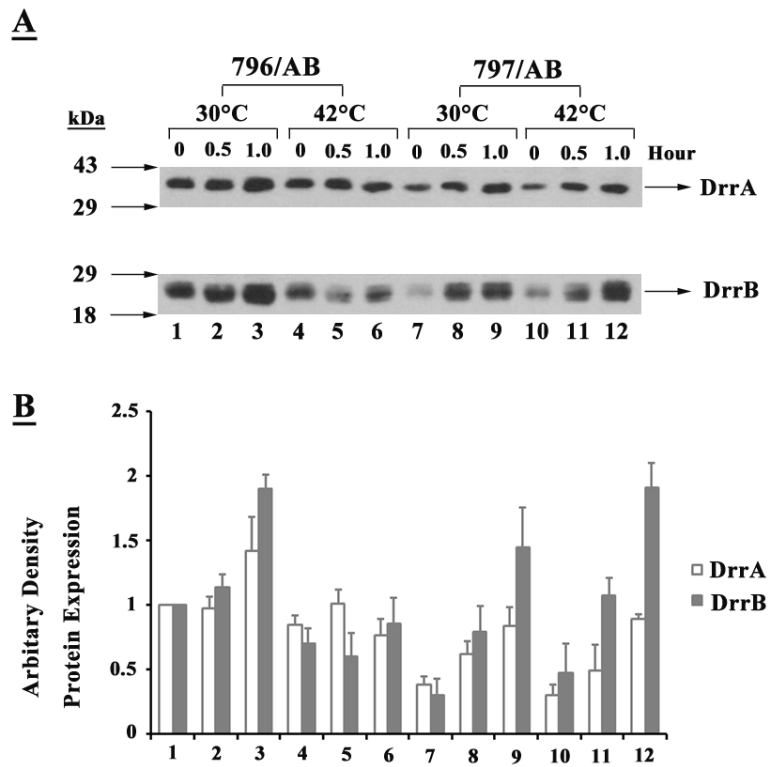
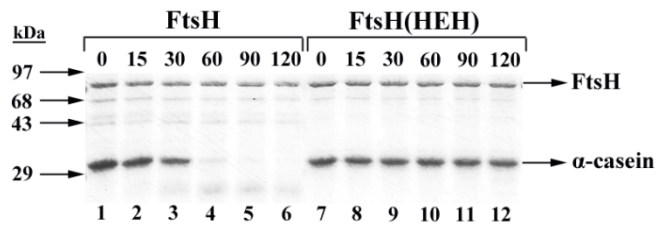


Figure 6

A



B

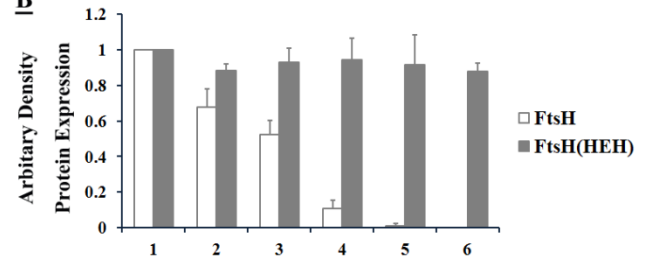


Figure 7

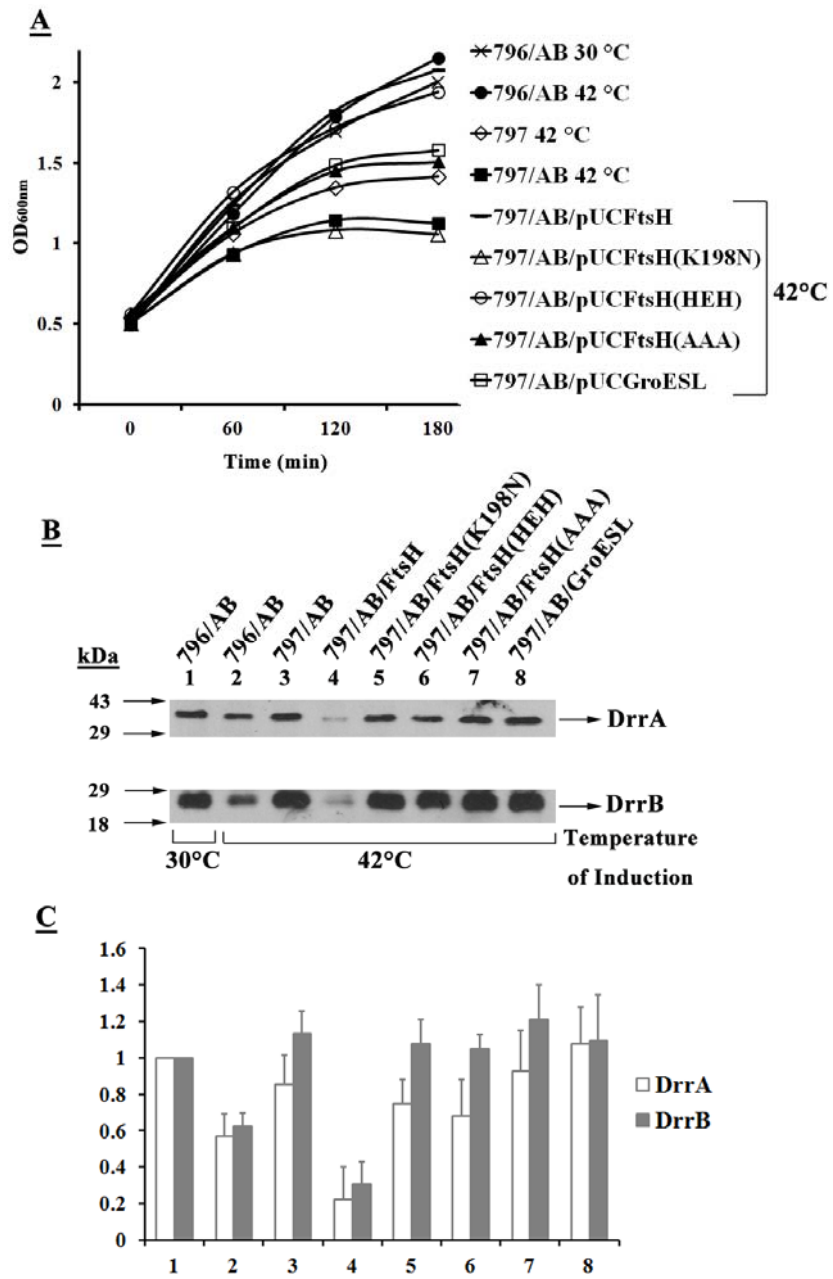
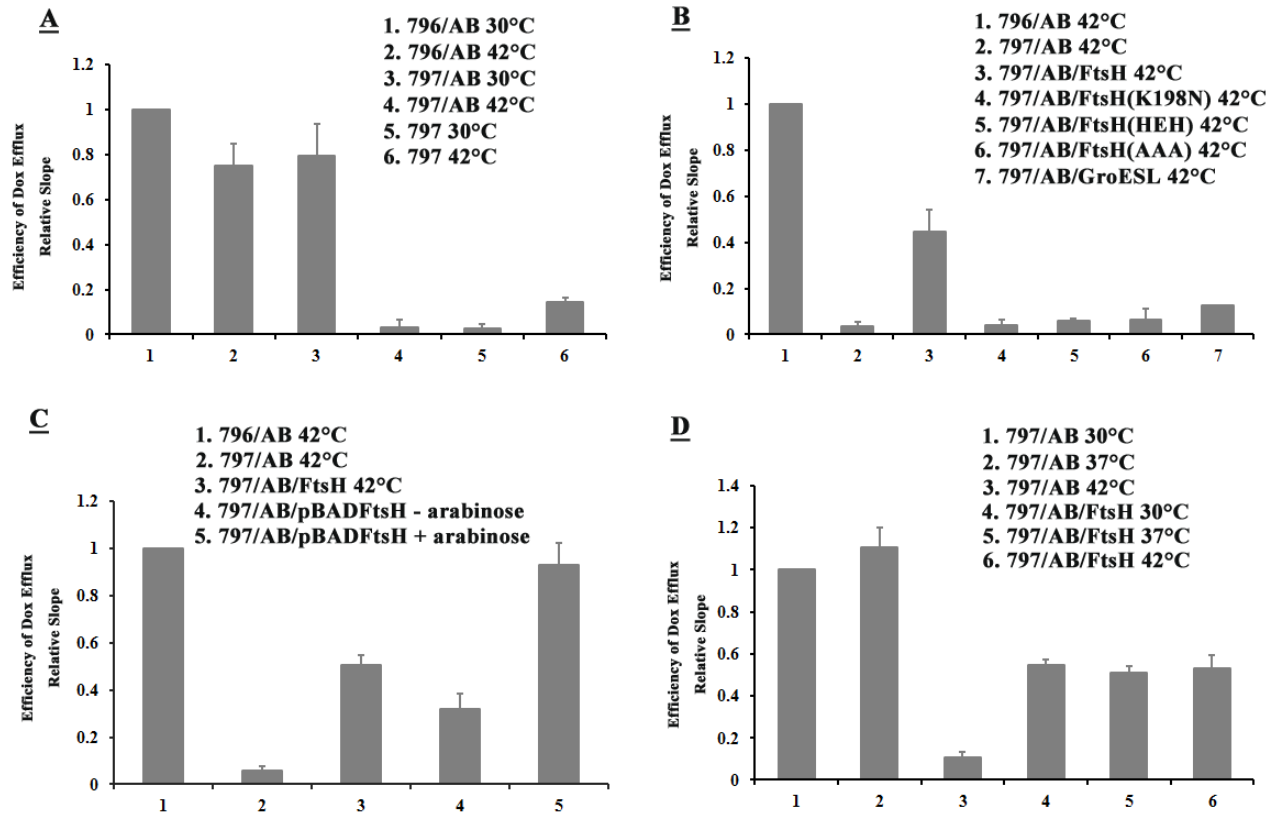


Figure 8



Dual Role of the Metalloprotease FtsH in Biogenesis of the DrrAB Drug Transporter

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