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Social control of brain morphology in a eusocial mammal

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Social status impacts reproductive behavior in diverse vertebrate species, but little is known about how it affects brain morphology. We explore this in the naked mole-rat, a species with the most rigidly organized reproductive hierarchy among mammals. Naked mole-rats live in large, subterranean colonies where breeding is restricted to a single female and small number of males. All other members of the colony, known as subordinates, are reproductively suppressed. Subordinates can become breeders if removed from the colony and placed with an opposite sex partner, but in nature most individuals never attain reproductive status. We examined the brains of breeding and subordinate naked mole-rats of both sexes, including several regions linked to reproduction and shown to be sexually dimorphic in other mammals. Stereological analyses revealed that neural morphology depends on status, such that breeders, regardless of sex, had more cells than subordinates in the ventromedial nucleus of the hypothalamus and a larger volume of the bed nucleus of the stria terminalis, paraventricular nucleus, and medial amygdala. Several other brain regions examined were unaffected. Surprisingly, males and females did not differ on any measure. These findings provide evidence that a change in social status triggers considerable neural remodeling and indicate that status, rather than sex, has a predominant role in determining neural structure in this remarkably social mammal.

eusociality | neuroplasticity | reproductive strategy | sex difference | social status

Sexual dimorphisms have been identified in the nervous systems of all vertebrate classes (1) and presumably arise and persist because they contribute in some way to the reproductive success of the organism. Males and females evolve different reproductive strategies, necessitating different morphologies and behaviors (2). In mammals, sex differences in the central nervous system often can be traced to developmental actions of gonadal steroid hormones (1). However, the study of sexual differentiation of the mammalian nervous system has focused on a limited number of relatively nonsocial species in which reproductive success is largely obtained through direct reproductive efforts.

In social species, reproduction may depend on status within the group, with members ranking lower in the hierarchy forgoing reproduction as well as sex-specific roles associated with reproduction (3). Naked mole-rats (*Heterocephalus glaber*) exhibit the strictest reproductive hierarchy known to mammals and the closest mammalian equivalent of eusociality. These small rodents, native to Africa, live in underground colonies averaging 60–80 individuals, including a single breeding female (the queen), one to three breeding males, and numerous nonreproductive adults, known as subordinates (4). Subordinates exhibit no mating behavior but assist in foraging, colony defense, maintenance of the tunnel system, and care of the young (4–7). Subordinates can become breeders if a breeding member of the colony dies or if they are removed from their colony and housed with a mate (5, 8, 9). Once established, however, the breeding pair is rarely overthrown, and it is estimated that in nature <5% of all naked mole-rats ever attain reproductive status (10). These animals therefore provide a rare opportunity for

testing hypotheses about how social status affects the mammalian brain, particularly regions involved in reproduction.

A change in status within a reproductive hierarchy alters neuronal gene expression, activity, and cell size in an African cichlid fish (11–16). It is not known whether the transition from subordinate to breeder similarly affects the brains of naked mole-rats. Studies of the naked mole-rat brain have been conducted only very recently and have focused on the organization of somatosensory cortex and the visual system (e.g., 17, 18). We recently described the distribution of vasopressin, a neuropeptide associated with social behaviors, in the naked mole-rat brain (19). Morphometric comparisons between the brains of breeders and subordinates, however, have not been made.

The degree to which neural sex differences are present in naked mole-rats also is not known, although subordinates display a relative lack of sex differences in anatomy and behavior. Mean body size does not differ, and the external genitalia and anogenital distance are very similar in male and female subordinates (7, 20, 21). Subordinates of both sexes participate equally in vocalizing, food retrieval, dominance behaviors, colony defense, and pup care (6, 22). Perhaps most significant, we previously found no sex differences in the morphology of the perineal muscles (which attach to the phallus and control penile reflexes in other mammals) or in cell size or number of the innervating motoneurons in the spinal cord (21, 23). Naked mole-rats are, to date, the only mammal that fails to exhibit a sex difference in morphology of this neuromuscular system.

We speculated that sexual differentiation might be “on hold” in subordinate naked mole-rats, and that sex differences might emerge only in those few individuals that become breeders. Alternatively, naked mole-rats might stand out from other mammals in having a sexually monomorphic brain throughout life, irrespective of reproductive status. To address these questions, we performed a stereological analysis of volume, cell number, and cell size in the naked mole-rat brain, using age-matched animals randomly assigned to become breeders or to remain subordinate. All breeders had been paired for at least 4 years and produced at least one litter. We selected brain regions that are related to reproduction and display robust sex differences in other animals (e.g., 24–27): the principal nucleus of the bed nucleus of the stria terminalis (BSTp), the paraventricular nucleus of the hypothalamus (PVN), the medial

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Abbreviations: BSTp, principal nucleus of the bed nucleus of the stria terminalis; PVN, paraventricular nucleus of the hypothalamus; MeA, medial nucleus of the amygdala; VMH, ventromedial nucleus of the hypothalamus; SCN, suprachiasmatic nucleus; ACo, anterior cortical amygdaloid nucleus.

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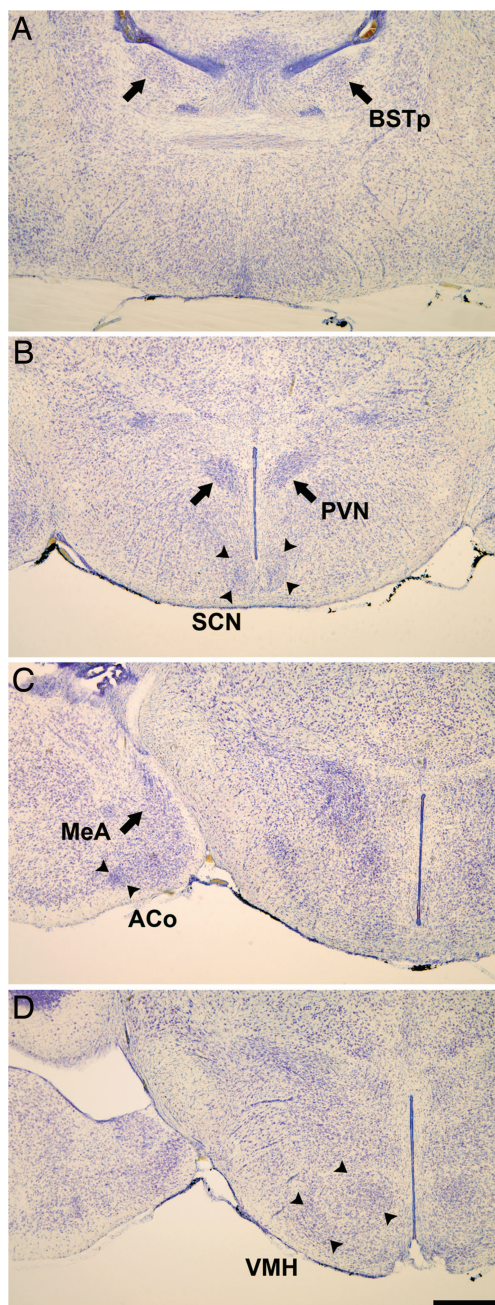


Fig. 3. Photomicrographs illustrate the location of nuclei in thionin-stained coronal sections of the naked mole-rat brain. Shown are BSTp (A); the PVN and SCN (B); MeA and ACo (C); and VMH (D). (Scale bar: 500 μm .)

“positive” social cues arising from the new mate may also play a role.

Effects of social cues and reproductive status on the brain have been particularly well studied in fish. For example, changes in the social environment trigger changes in reproductive status and alterations in neural morphology and neuropeptide gene expression in males of the species *Astatotilapia burtoni* (11–15). The behavioral and neural changes are reversible, as *A. burtoni* males may switch status both rapidly and repeatedly (46). Increases in immediate-early gene expression are observed in gonadotrophin-releasing hormone (GnRH) neurons within minutes of a relevant social stimulus, suggesting that this is a very early event in a molecular cascade presumably leading to longer-term changes (16). Breeding

status is also reflected in altered neuropeptide expression in naked mole-rats, with subordinates exhibiting less vasopressin in the dorsomedial hypothalamus (19). GnRH neurons have not yet been examined in naked mole-rats, but subordinates release less pituitary luteinizing hormone in response to a GnRH pulse than do breeders (47).

However, in contrast to the changes in *A. burtoni*, the rise to breeding status of a former subordinate in a naked mole-rat colony is a protracted process; many months may elapse before a new breeding pair is established after the death or removal of the former breeders (7). The transition to breeding status also does not appear to be reversible; naked mole-rats are the longest-lived rodents, routinely surviving >20 years in captivity, and breeders generally maintain their status until death (20, 48). Although we do not know how long an animal must be a breeder before measurable neural changes occur, we suggest that the increases in neuron number and overall size of reproductive brain regions seen here may represent a relatively stable, long-term outcome of events occurring much more rapidly. Currently, the earliest physiological marker heralding a change in status in naked mole-rats is the increase in reproductive hormones seen 5–8 days after separating a subordinate from its colony (8, 42, 49).

The endocrine changes that accompany a change in breeding status (50) suggest that hormones must be considered as possible factors mediating effects of social status on brain morphology. Each of the reproductive brain regions we examined expresses gonadal steroid hormone receptors in other rodents (51, 52), and our own preliminary observations confirm that the four regions showing effects of status also exhibit androgen receptor immunoreactivity in naked mole-rats (M.M.H., unpublished work). Behavioral observations, however, suggest that some effects of breeding status on neural morphology may prove to be independent of the gonads in naked mole-rats. In a colony setting, only the breeders display genital nuzzling and they do so at all times of the queen’s ovulatory cycle, during pregnancies, and after gonadectomy of both members of the breeding pair (6, 53). Similarly, breeding pairs maintain their status and continue to suppress reproduction of other colony members over long periods of time, even when not producing offspring and in the absence of gonads (53). Some effects of social status on the brain and behavior are also likely to be independent of the gonads in fish (16, 54).

We do not know whether the brain changes seen here would be maintained following gonadectomy of the breeders. If so, then gonadal steroids could be ruled out as necessary for maintaining these neural differences. It would remain possible that gonadal steroids are required for initiating the changes or that steroids from a nongonadal source (e.g., neurosteroids or adrenal steroids) are responsible. Adrenal glucocorticoids are associated with social stress and/or social rank and also can have effects on neural morphology (55), but evidence as to whether cortisol levels relate to social status in naked mole-rat colonies has been contradictory (45, 56). The recent demonstration that steroid hormone receptors can be activated by neurotransmitters, in the absence of hormone (so-called ligand-independent activation of receptors), suggests another pathway by which the environment may influence reproductive brain regions (57). More generally, the activation of neural gene expression either by hormones or neurotransmitters provides a framework for understanding how social stimuli may affect brain morphology.

Naked mole-rats lie at one extreme of the spectrum of sociality displayed by mammals, with larger colony sizes, greater interrelatedness of colony members, and more marked behavioral specializations than seen in any other social mammal (38). Nonetheless, many mammalian species (particularly among the rodents, canids, and primates) exhibit a reproductive division of labor, in which some members of the social group reproduce whereas others are reproductively suppressed and assist in rearing young that are not their own (58). Although effects of social status on the brain may

be more pronounced and therefore easier to detect in naked mole-rats, the phenomenon is likely to apply much more broadly.

Materials and Methods

Animals and Tissue Collection. Housing conditions and animal history and care have been reported (23). Brains from 8 breeding females, 8 breeding males, 10 subordinate females, and 8 subordinate males were collected. Animals were anesthetized and rapidly decapitated, and brains were removed and immersion fixed in 5% acrolein in phosphate buffer for 4 h. Brains were then transferred to 30% sucrose in phosphate buffer and frozen-sectioned at 30 μm in the coronal plane. Alternate sections were mounted onto slides and stained with thionin. All procedures adhered to institutional and federal guidelines.

Tissue Analyses. All measures were performed on slides coded to conceal the sex and status of the animals. Stereological analyses of the BSTp, SCN, PVN, ACo, MeA, and VMH (Fig. 3) were performed by using StereoInvestigator software (MicroBrightfield, Williston, VT).

The VMH was initially subdivided into three regions (dorsomedial, central, and ventrolateral), which were analyzed separately. Because VMH subregion did not significantly interact with either sex or status in statistical analyses, subregions were combined in the data presented here. To calculate overall volumes, outlines of each region were traced in each section, and the summed areas were multiplied by section thickness. Unbiased estimates of cell number within each region were obtained by using the optical disector method, with parameters adjusted for each brain region. Counting frames varied from 16 \times 16 μm to 20 \times 20 μm , and sampling grid

size was varied from 80 \times 80 μm to 150 \times 150 μm depending on the size of the region analyzed. For a given measure, parameters were held constant for all animals. Mean cell size in each region was determined by randomly placing a grid in each section through the rostral-caudal extent of each brain region and tracing every in-focus cell with a neuronal morphology that fell within the grid. At least 50 cells per animal were traced for each region. All volume and cell count data reflect unilateral estimates.

For each animal, cortical thickness was measured in three sections chosen to correspond to the level of (i) the BSTp (most anterior), (ii) SCN and PVN, and (iii) ACo, MeA and VMH (most posterior). Three measurements were taken from each hemisphere in each of the chosen sections, resulting in an average cortical thickness based on a total of 18 measurements per animal. Measures were taken from the lower boundary of layer I to the beginning of the white matter below layer VI. The first measure was taken immediately lateral to the elevation of the corpus callosum, and second and third measures were taken 150 and 300 μm lateral, respectively (as in ref. 59).

Dependent variables were analyzed by using two-way ANOVAs (sex-by-status). Tissue artifact caused some animals to be removed from analyses for specific brain regions. There was always a minimum of five animals per group per brain region.

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- Cooke B, Hegstrom CD, Villeneuve LS, Breedlove SM (1998) *Front Neuroendo* 19:323–362.
- Darwin C (1871) *The Descent of Man, and Selection in Relation to Sex* (Murray, London).
- Abbot DH (1987) *J Zool* 213:455–470.
- Jarvis JUM (1981) *Science* 212:571–573.
- Brett RA (1991) in *The Biology of the Naked Mole-Rat*, eds Sherman PW, Jarvis JUM, Alexander RD (Princeton Univ Press, Princeton), pp 97–136.
- Lacey EA, Alexander RD, Braude SH, Sherman PW, Jarvis JUM (1991) in *The Biology of the Naked Mole-Rat*, eds Sherman PW, Jarvis JUM, Alexander RD (Princeton Univ Press, Princeton), pp 209–242.
- Lacey EA, Sherman PW (1991) in *The Biology of the Naked Mole-Rat*, eds Sherman PW, Jarvis JUM, Alexander RD (Princeton Univ Press, Princeton), pp 275–336.
- Faulkes CG, Abbott DH, Jarvis JUM (1990) *J Reprod Fertil* 88:559–568.
- Margulis SW, Saltzman W, Abbott DH (1995) *Horm Behav* 29:227–247.
- Jarvis JUM, O’Riain MJ, Bennett NC, Sherman PW (1994) *Trends Ecol Evol* 9:47–51.
- Davis MR, Fernald RD (1990) *J Neurobiol* 21:1180–1188.
- Francis RC, Soma K, Fernald RD (1993) *Proc Natl Acad Sci USA* 90:7794–7798.
- Hofmann HA, Fernald RD (2000) *J Neurosci* 20:4740–4744.
- White SA, Nguyen T, Fernald RD (2002) *J Exp Biol* 205:2567–2581.
- Greenwood AK, Fernald RD (2004) *Biol Reprod* 71:909–918.
- Burmeister SS, Jarvis ED, Fernald RD (2005) *PLoS Biol* 3:1996–2004.
- Catania KC, Remple MS (2002) *Proc Natl Acad Sci USA* 99:5692–5697.
- Xiao J, Levitt JB, Buffenstein R (2006) *Brain Res* 1077:81–89.
- Rosen GJ, De Vries GJ, Goldman SL, Goldman BD, Forger NG (2007) *J Comp Neurol* 500:1093–1105.
- Jarvis JUM (1991) in *The Biology of the Naked Mole-Rat*, eds Sherman PW, Jarvis JUM, Alexander RD (Princeton Univ Press, Princeton), pp 384–425.
- Peroulakis ME, Goldman B, Forger NG (2002) *J Neurobiol* 51:33–42.
- Pepper JW, Braude SH, Lacey EA, Sherman PW (1991) in *The Biology of the Naked Mole-Rat*, eds Sherman PW, Jarvis JUM, Alexander RD (Princeton Univ Press, Princeton), pp 243–274.
- Seney ML, Goldman BD, Forger NG (2006) *J Neurobiol* 66:1354–1364.
- Matsumoto A, Arai Y (1983) *Endocrinol Jpn* 30:277–280.
- Cooke BM, Tabibnia G, Breedlove SM (1999) *Proc Natl Acad Sci USA* 96:7538–7540.
- Forger NG, Rosen GJ, Waters EM, Jacob D, Simerly RB, de Vries GJ (2004) *Proc Natl Acad Sci USA* 101:13666–13671.
- Viau V, Bingham B, Davis J, Lee P, Wong M (2005) *Endocrinology* 146:137–146.
- De Vries GJ, Simerly RB (2002) in *Hormones, Brain, and Behavior, Volume IV*, eds Pfaff DW, Arnold AP, Etgen AM, Fahrbach SE, Moss RL, Rubin RT (Academic, San Diego), pp 137–191.
- Simerly RB (2002) *Annu Rev Neurosci* 25:507–536.
- Abizaid A, Mezei G, Sotonyi P, Horvath TL (2004) *Eur J Neurosci* 19:2488–2496.
- Madeira MD, Sousa N, Santer RM, Paula-Barbosa MM, Gundersen HJG (1995) *J Comp Neurol* 361:585–601.
- Gorski RA, Gordon JH, Shryne JE, Southam AM (1978) *Brain Res* 148:333–346.
- Swaab DF, Hofman MA (1988) *Brain Res Dev Brain Res* 44:314–318.
- Vandenbergh JG (1983) *Proc Life Sci* 342–349.
- Foster DL, Ebling FJ, Claypool LE (1988) *Reprod Nutr Dev* 28:349–364.
- Bennett NC, Faulkes CG (2000) *African Mole-Rats: Ecology and Eusociality* (Cambridge Univ Press, Cambridge, UK).
- Kleiman DG (1977) *Quart Rev Biol* 52:39–69.
- Alexander RD, Hoogland JL, Howard RD, Noonan KM, Sherman PW (1979) in *Evolutionary Biology and Human Social Behavior: An Anthropological Perspective*, eds Chagnon NA, Irons W (Duxbury, Belmont, CA), pp 402–435.
- Morris JA, Jordan CL, Breedlove SM (2004) *Nat Neurosci* 7:1034–1039.
- Forger NG (2006) *Neuroscience* 138:929–938.
- Fowler CD, Liu Y, Ouimet C, Wang Z (2002) *J Neurobiol* 51:115–128.
- Faulkes CG, Abbott DH (1993) *J Reprod Fertil* 99:225–230.
- Smith TE, Faulkes CG, Abbott DH (1997) *Horm Behav* 31:277–288.
- Reeve HK (1992) *Nature* 358:147–149.
- Clarke FM, Faulkes CG (1998) *Proc R Soc Lond B* 265:1391–1399.
- Hofmann HA, Benson ME, Fernald RD (1999) *Proc Natl Acad Sci USA* 96:14171–14176.
- Faulkes CG, Abbott DH, Jarvis JU, Sherriff FE (1990) *J Reprod Fertil* 89:317–323.
- Buffenstein R (2005) *J Gerontol A Biol Sci Med Sci* 60:1369–1377.
- Faulkes CG, Abbott DH (1991) *J Reprod Fertil* 93:427–435.
- Faulkes CG, Abbott DH, Jarvis JU (1991) *J Reprod Fertil* 91:593–604.
- Simerly RB, Chang C, Muramatsu M, Swanson LW (1990) *J Comp Neurol* 294:76–95.
- Shughrue PJ, Lane MV, Merchenthaler I (1997) *J Comp Neurol* 388:507–525.
- Goldman SL, Forger NG, Goldman BD (2006) *Horm Behav* 50:77–84.
- Godwin J, Crews D, Warner RR (1996) *Proc R Soc Lond B* 263:1683–1688.
- Tamashiro KL, Nguyen MM, Sakai RR (2005) *Front Neuroendocrinol* 26:27–40.
- Clarke FM, Faulkes CG (1997) *Proc R Soc Lond B* 264:993–1000.
- Blaustein JD (2003) *Ann N Y Acad Sci* 1007:238–250.
- Solomon NG, French JA (1997) in *Cooperative Breeding in Mammals*, eds Solomon NG, French JA (Cambridge Univ Press, New York), pp 1–10.
- Diamond MC, Krech D, Rosenzweig MR (1964) *J Comp Neurol* 123:111–120.