

UNCLASSIFIED

**NABIR P.I. Meeting, April, 2005**

# **Bacterial Biotransformations for the *In situ* Stabilization of Plutonium**

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## Overview

**Plutonium contamination in the environment is generally low-level and may be present and transported in a range of forms (IV, V, VI).**

**Current remediation strategies are costly, financially and in terms of increased exposure risk to people and the environment. In situ bacterial biostabilization is a promising alternative.**

### Aqueous Speciation Related to Environmental Conditions

- Plutonium(VI) vs Uranium(VI) Hydrolysis
- Plutonium(IV/III) EDTA Speciation and Stability
- Siderophore Stabilization of Plutonium(IV)
- New Reduction Potentials

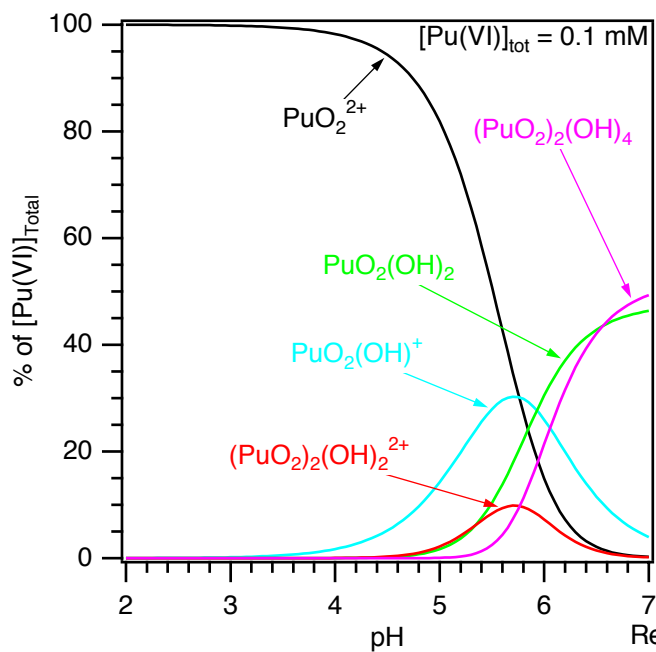
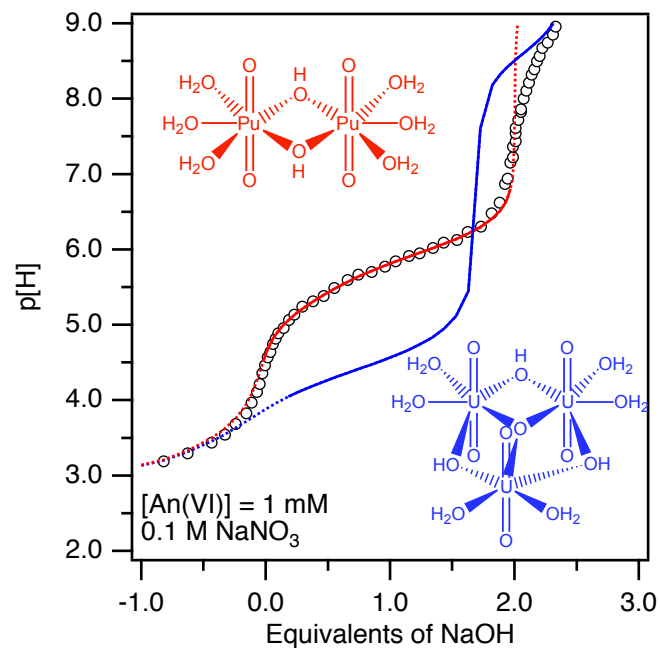
### Bacterial Biotransformations

- Siderophore-mediated Accumulation by Aerobic Bacteria
- EPS and Cell Adsorption by Aerobic Bacteria
- Reduction by DMRB

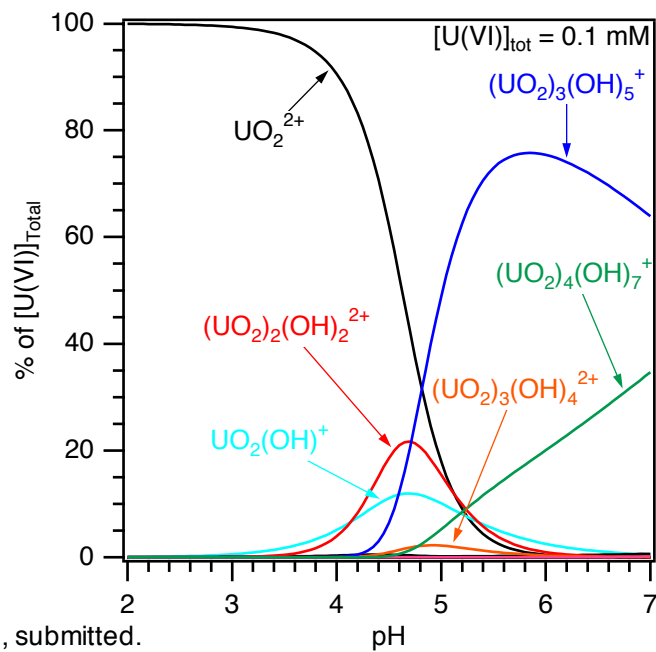
# Pu (VI) Hydrolysis

Why biostabilization methods being developed for U, e.g. reduction by DMRB, may or may not work for Pu

~600 papers on U(VI) hydrolysis  
5 papers on Pu(VI) hydrolysis  
2 papers on Pu(V) hydrolysis

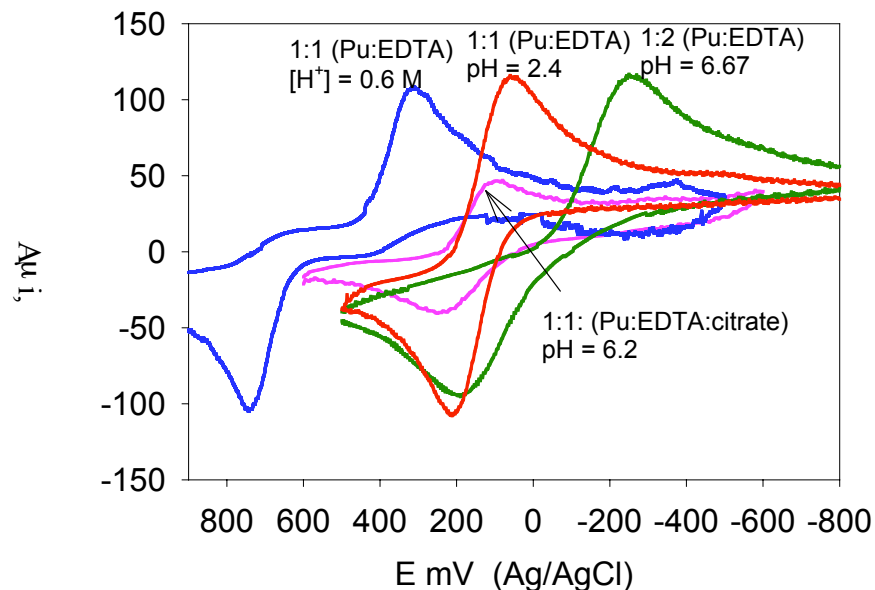


Reilly, Neu, *Inorg. Chem.*, submitted.



# Pu(IV)EDTA Aqueous Speciation

## Cyclic Voltammetry



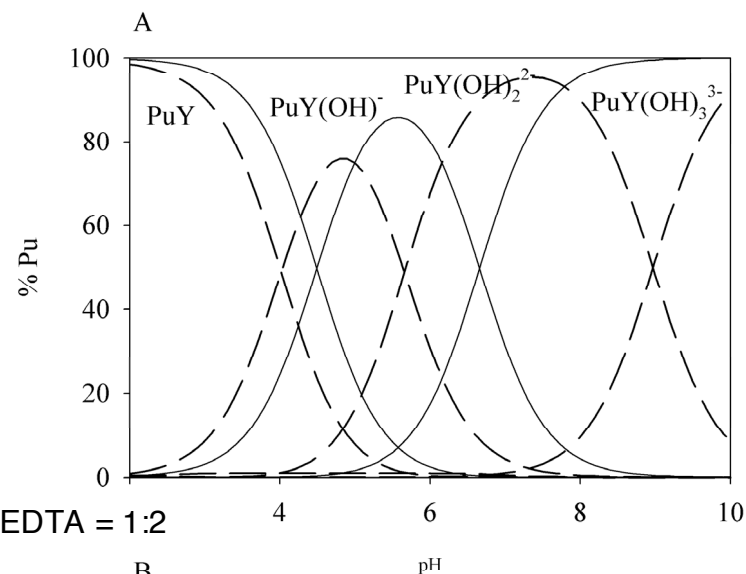
**New species are identified including  $Pu(EDTA)_2$ ,  $\log \beta = 35.43$**

**Stability of Pu(IV) is enhanced at environmentally relevant pH by the formation  $Pu(IV)$ -EDTA-hydroxo and  $Pu(IV)$ -EDTA-L mixed complexes.**

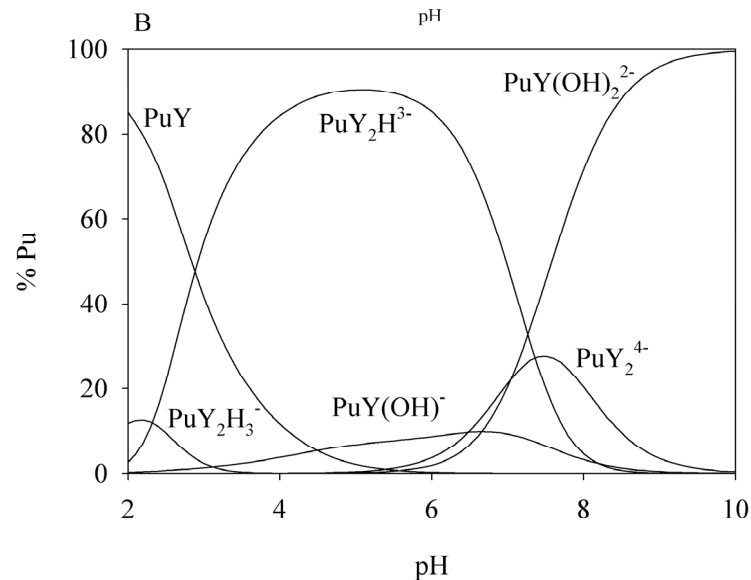
**Unusual speciation due to high charge and large coordination sphere**

## New Species Distributions (---)

Pu:EDTA = 1:1



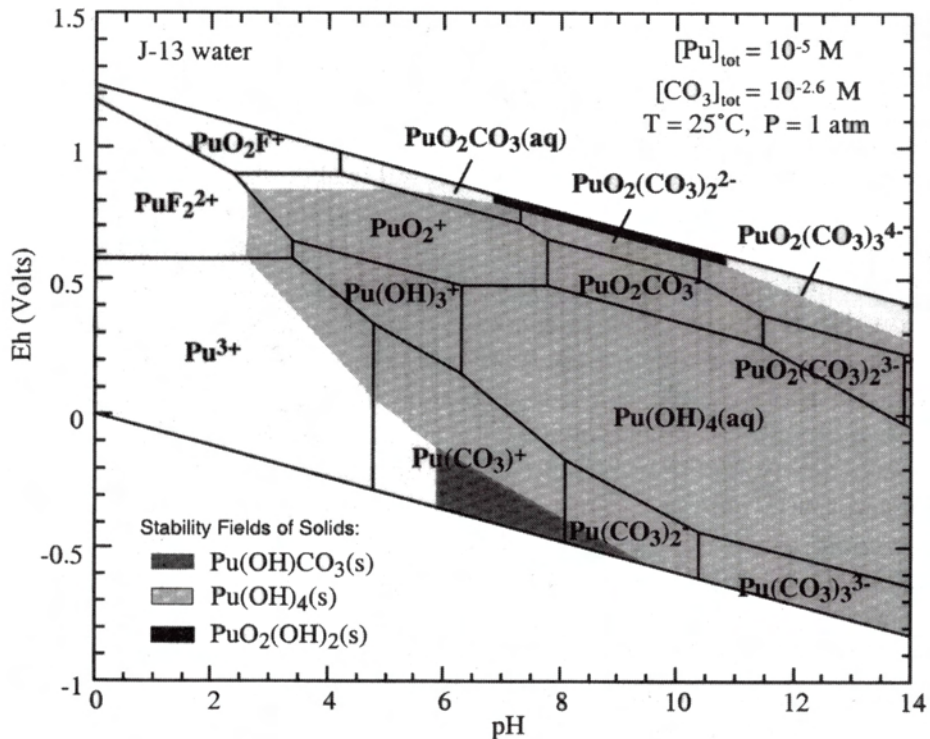
Pu:EDTA = 1:2



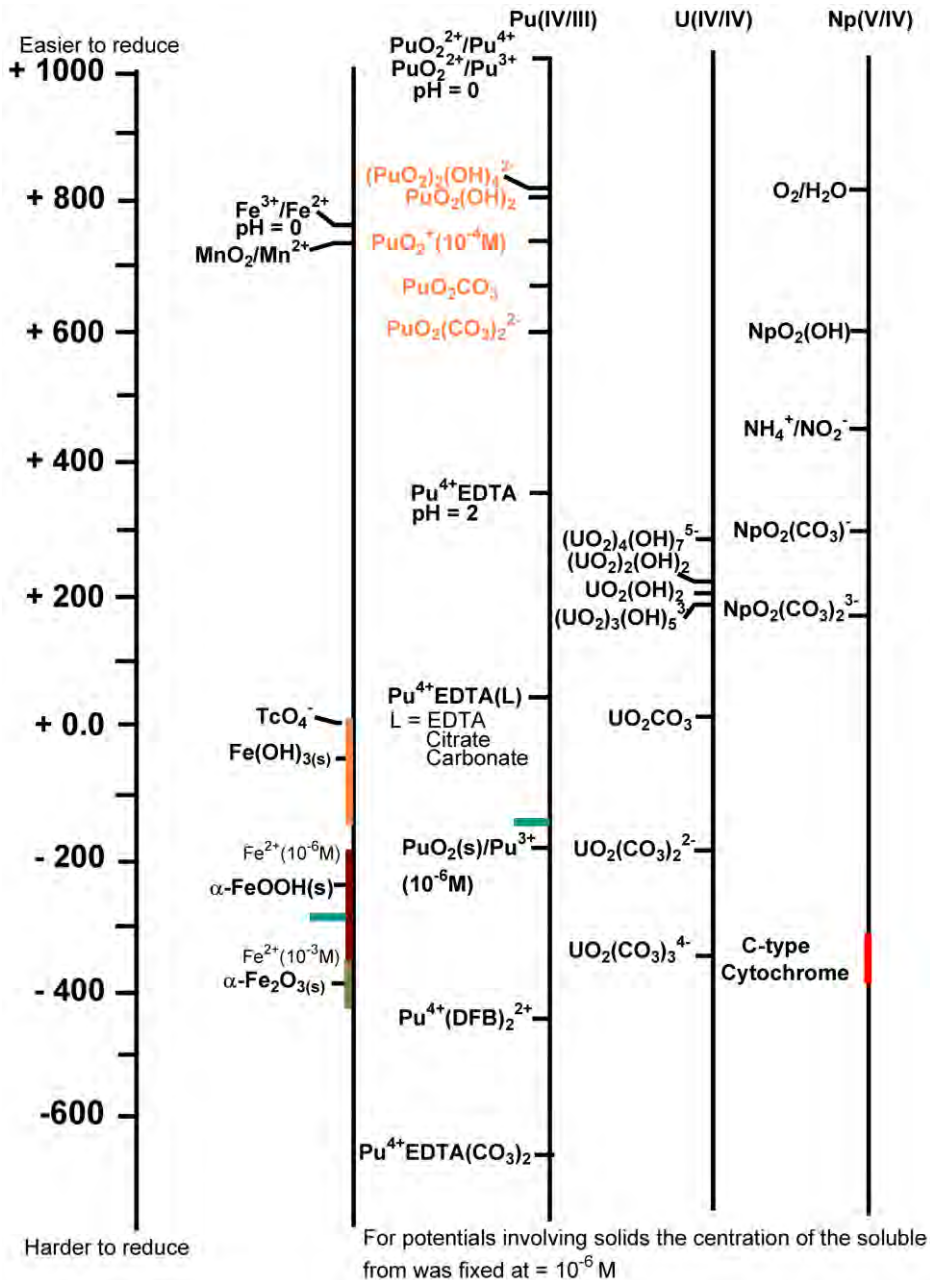
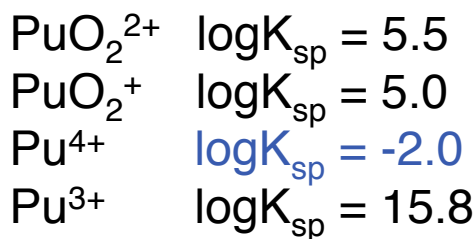
Boukhalfa, Reilly, Neu, *Inorg. Chem.*,  
2004, 43(19), 5816.

# Pu Speciation Under Environmental Conditions

$E_{red}$  (mV) vs NHE, pH = 7



## Solubility of primary (hydr)oxide phases

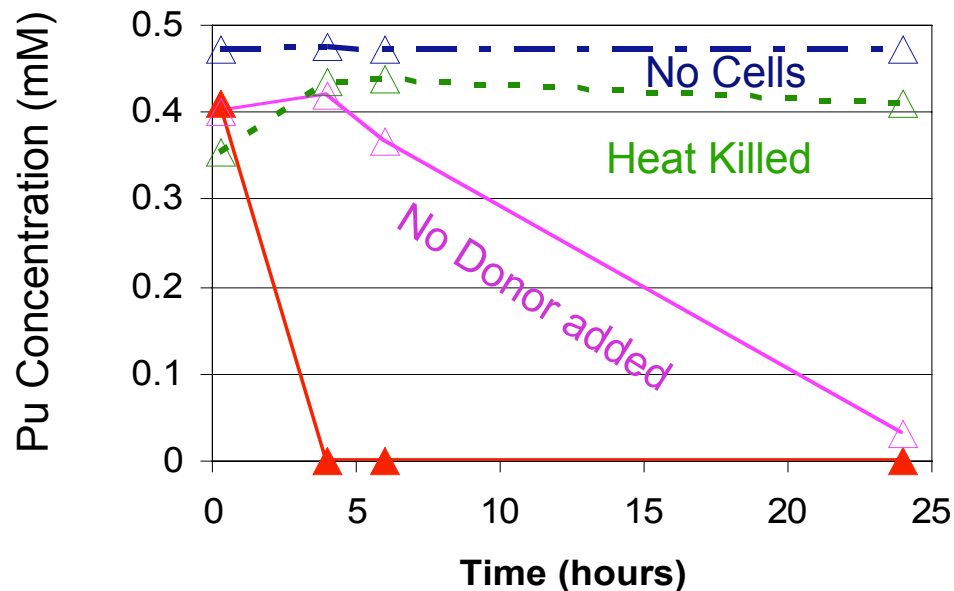


# Pu(VI) as a Terminal Electron Acceptor for DMRB

*Shewanella oneidensis* MR1

10 mM Lactate

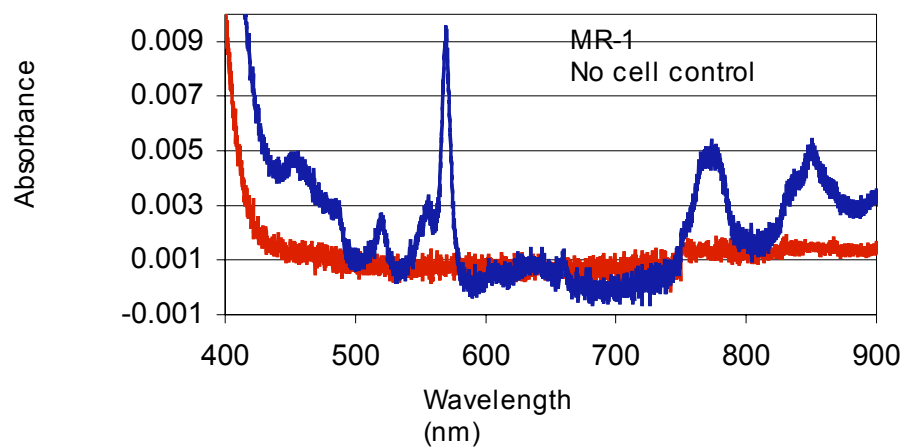
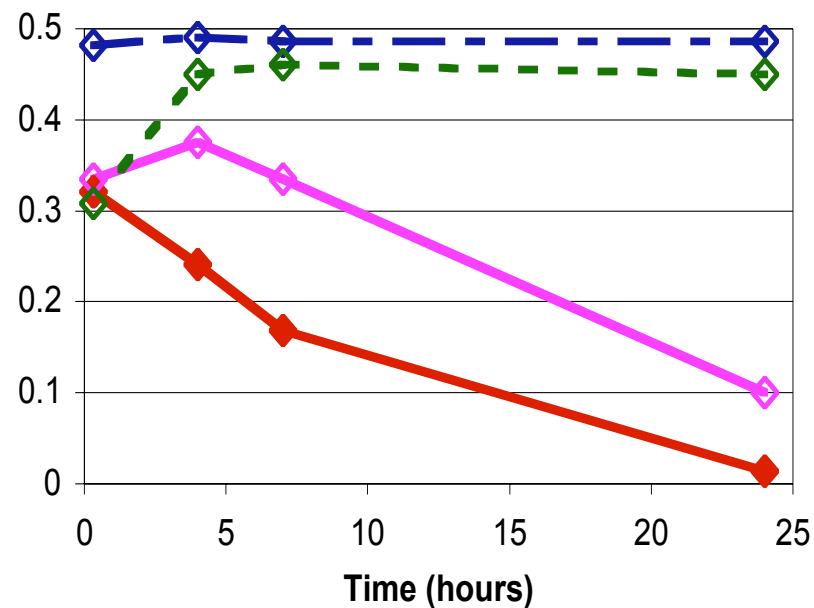
$5 \times 10^8$  Cells/mL



*Geobacter metallireducens* GS15

10 mM Acetate

$5 \times 10^8$  Cells/mL



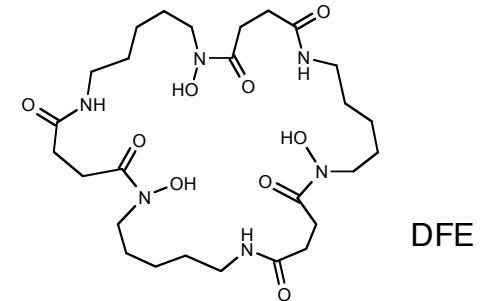
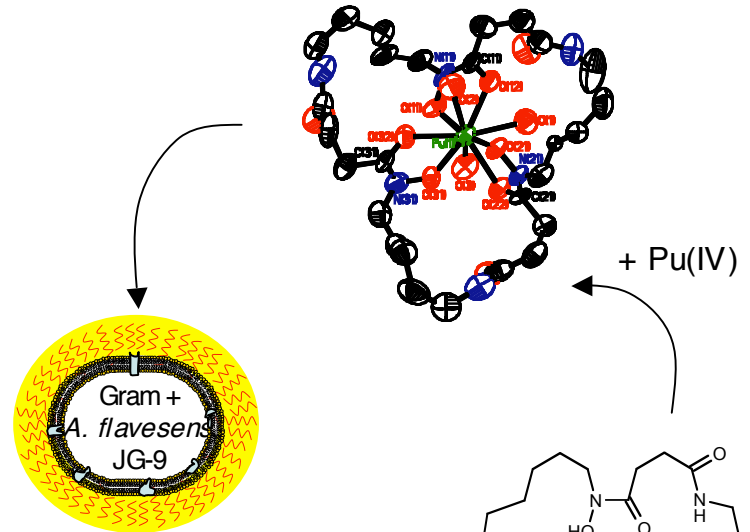
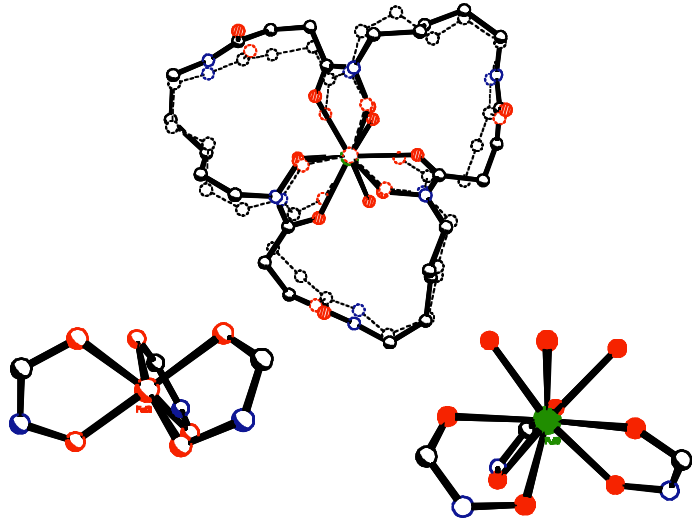
DMRB reduce Pu(VI) and Pu(V)

Product appears to be  $\text{PuO}_2$  (hyd)

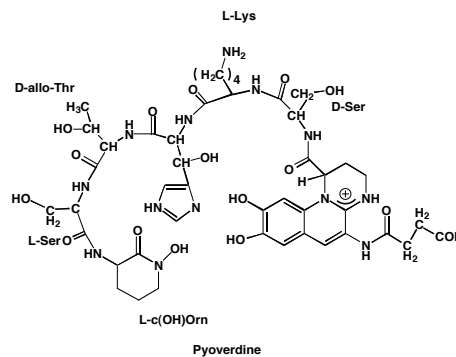
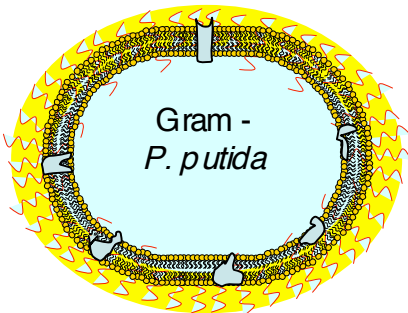
Characterization and dissolution studies in progress

# Siderophore-Mediated Pu Accumulation

Fe(III) and Pu(IV) siderophore structures

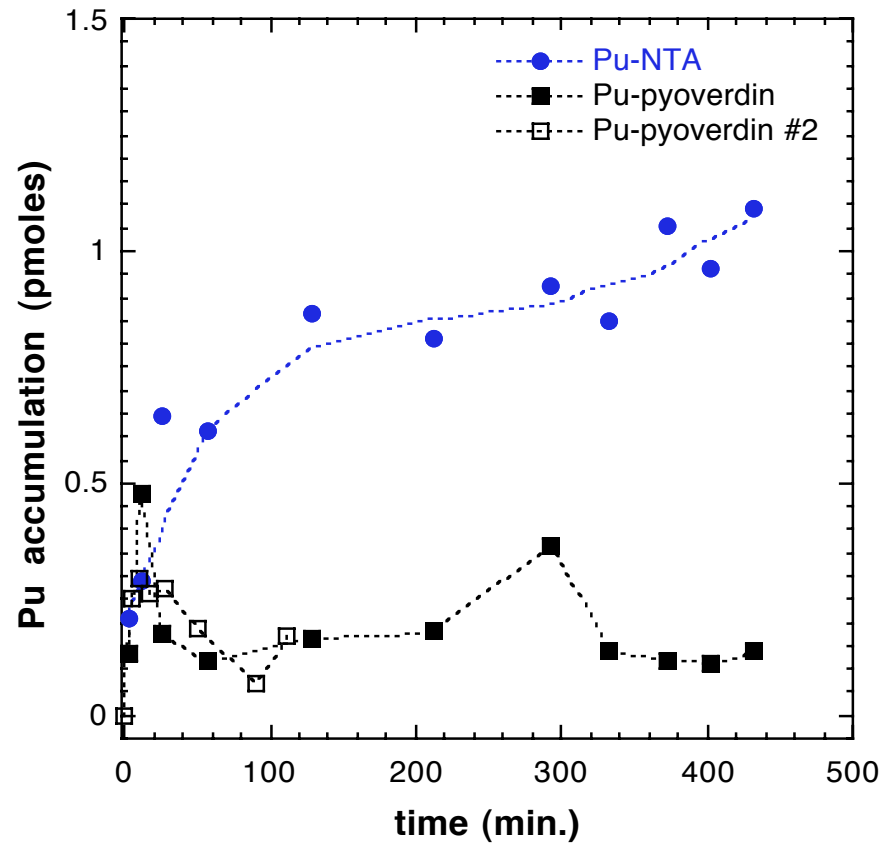
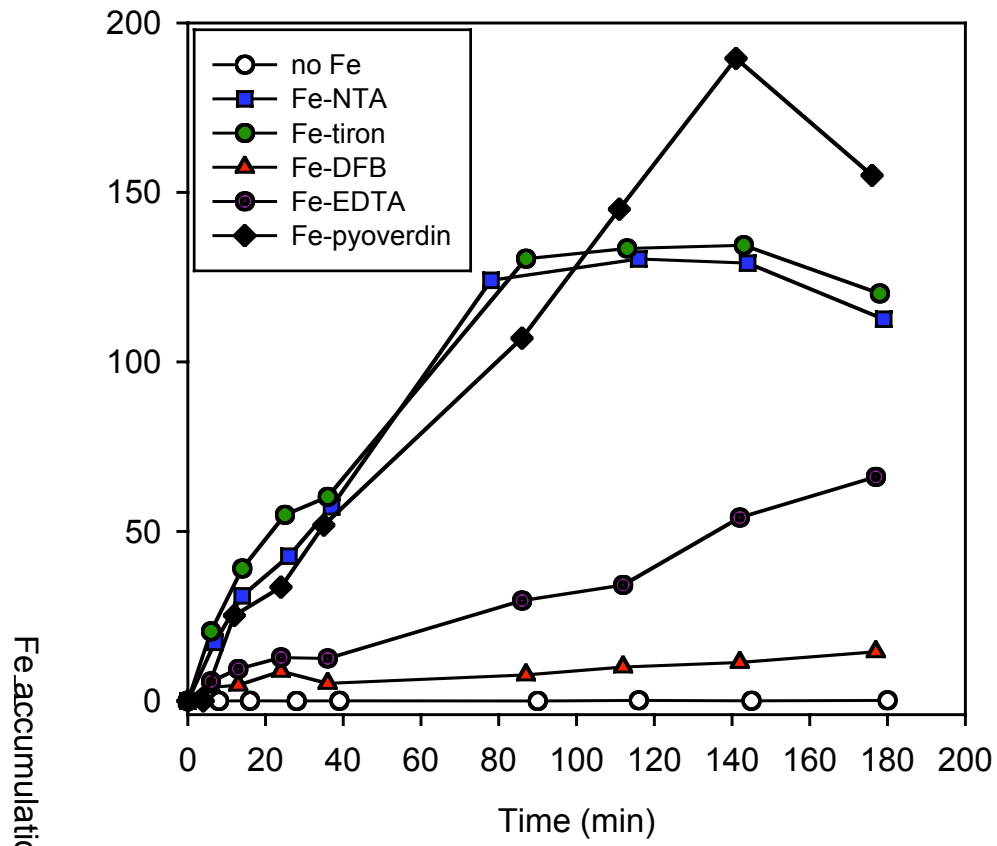


+ Pu(IV)  
+ Fe(III)  
???



**Can Pu be accumulated  
by other bacteria  
via other types of siderophores?**

## *P. putida* Metal Acquisition from Multiple Chelates



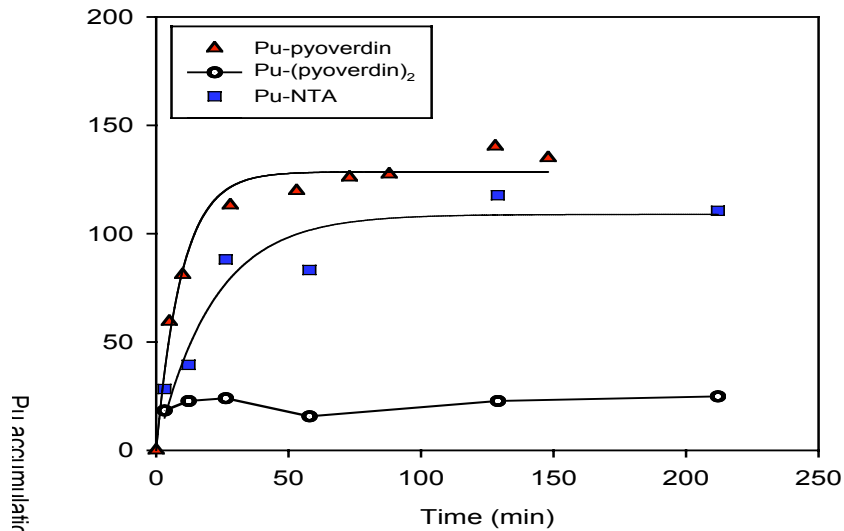
Uptake inversely proportional to Fe-L complexes stability.

Fe uptake profile suggests that exogenous ligands release iron to the siderophore either in solution or at the membrane surface.

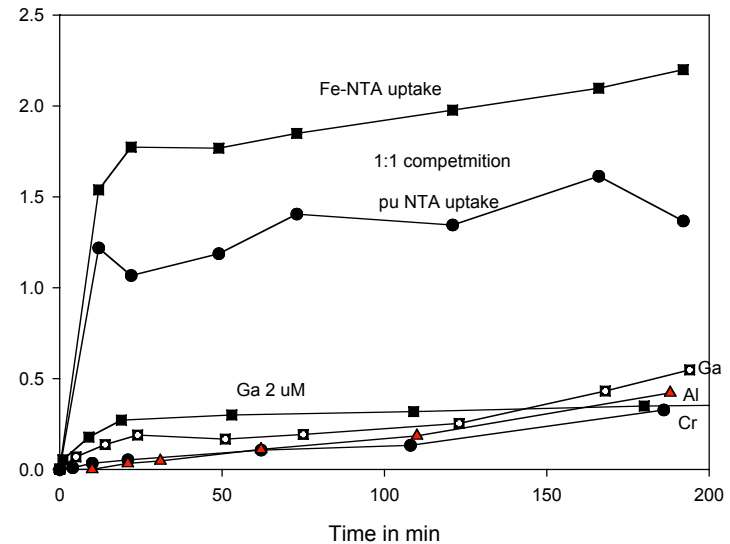


# Requirements for Metal–Siderophore Uptake

Siderophore binding, membrane protein recognition, metal shuttle, intra-cellular release thought to require specific physico-chemical characteristics.



*P. Putida* cells take up NTA and pyoverdin complexes, both in the presence and absence of Fe.



*P. Putida* cells pre-incubated with 2  $\mu$ M of pyoverdin complexes of Cr(III), Ga(III) and Al(III) unable to acquire Fe from NTA, EDTA or pyoverdin complexes.

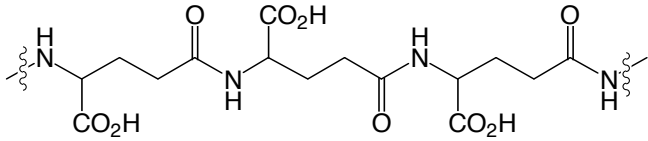
Characteristics required for complete translocation?

specific radii, trivalent charge, specific molecular conformation, neutral molecular charge, metal reduction, ligand exchange (solution or membrane)....

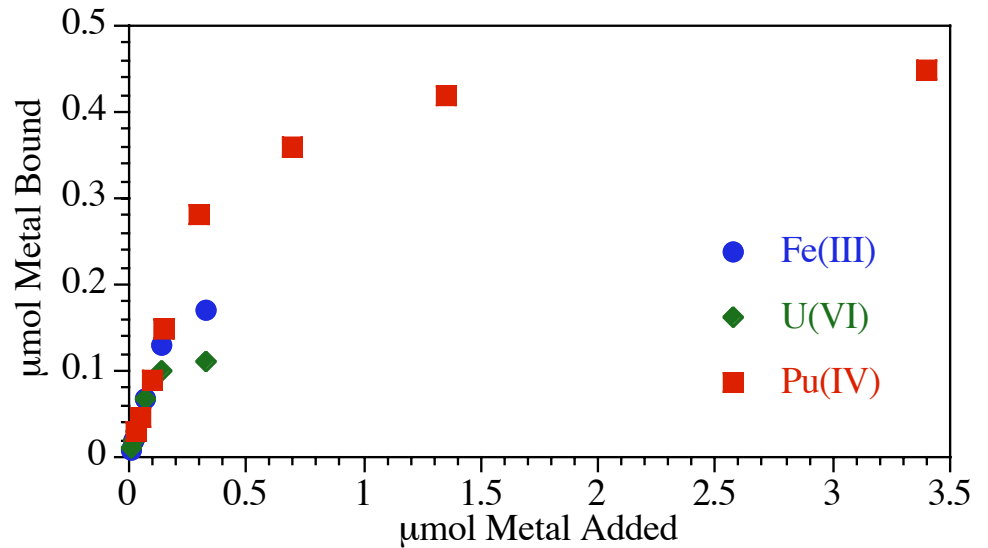
Pu(IV) and Th(IV) uptake reveal combination of ligand exchange and reduction is key

# Metal Binding of Microbial Extracellular Polymers

## PGA of *B. licheniformis*



~800 kDa, forms soluble metal complexes, generally >10:1 glu to M

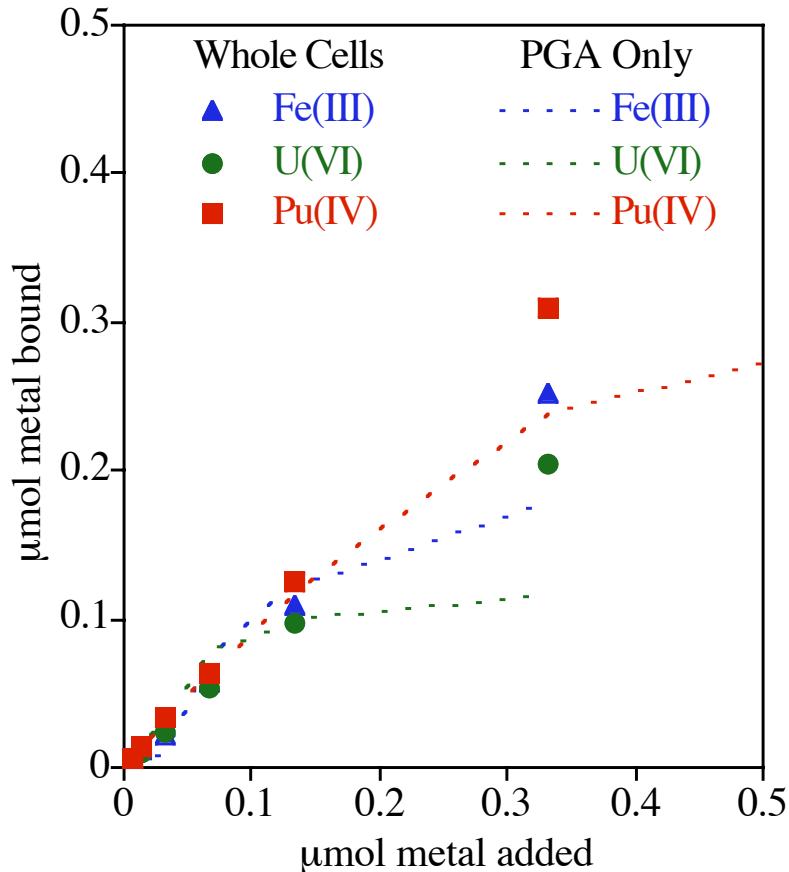


~0.12 mmol metal bound per mg PGA (alone)

Pu(IV) remains associated with PGA during repeated pH cycling 2-12

Whole cells (in culture media) take up more Pu per mass than does polyglutamate

Siderophores and EPS associate Pu with cells



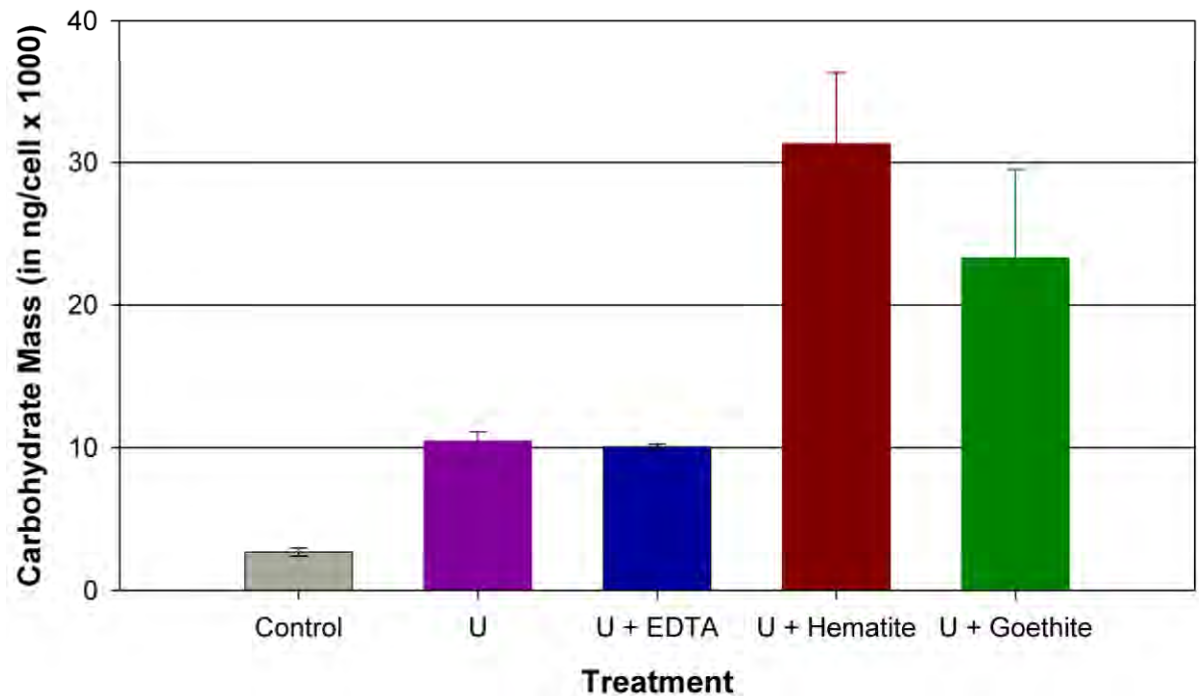
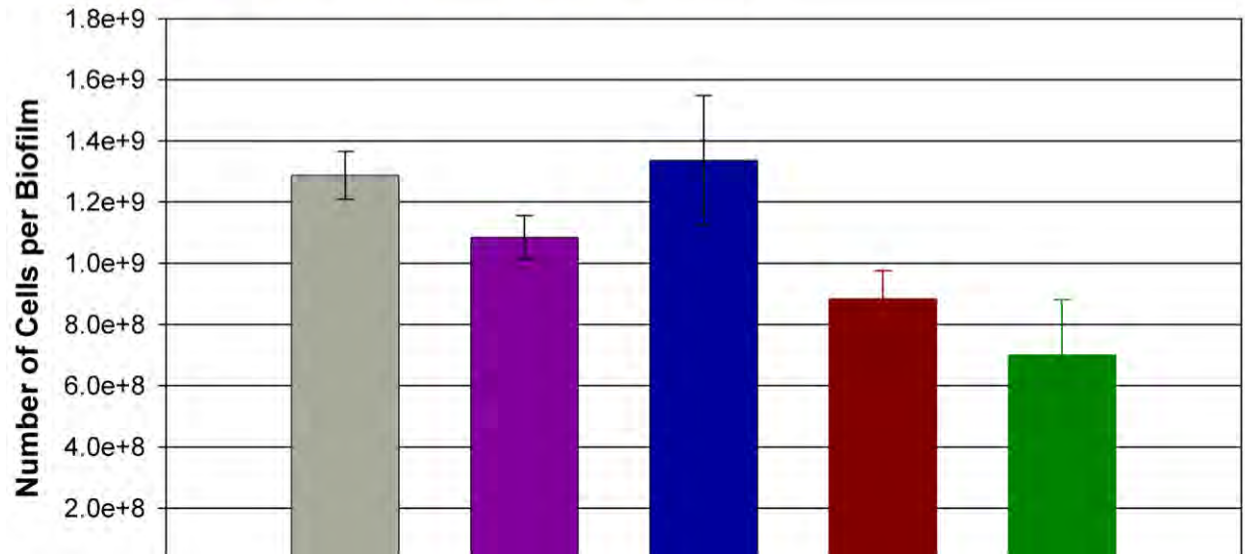
## Effect of U(VI) on *P. putida* Biofilms

*P. Putida* grown on membrane discs on U-containing agar with minimal nutrients

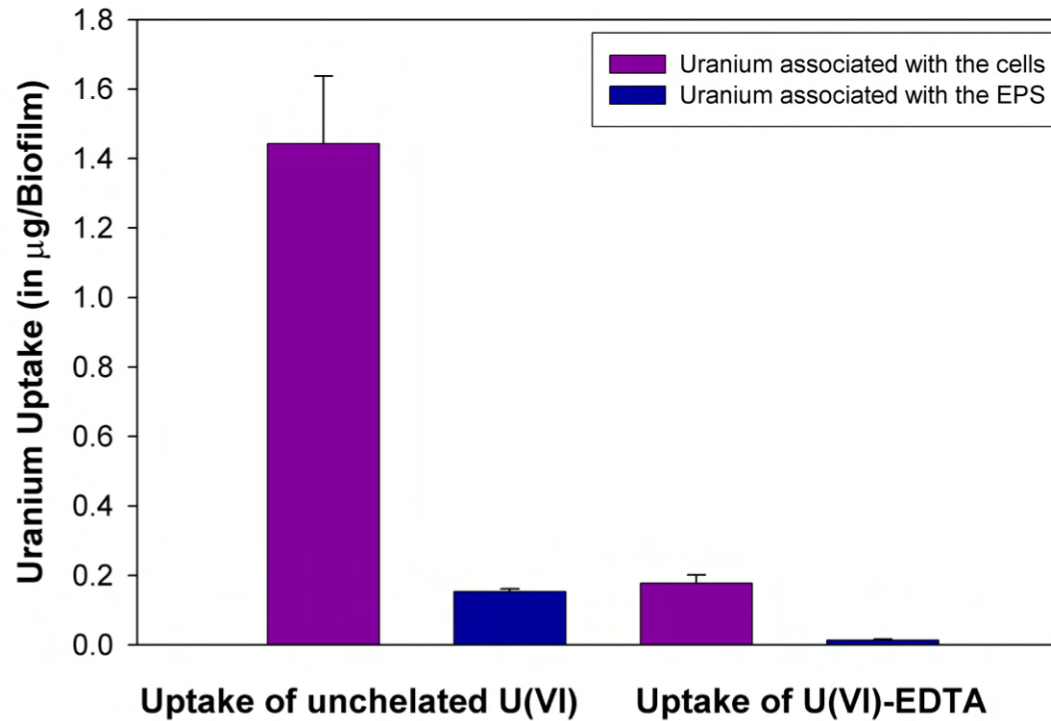
[U] =  $10^{-4}$  M  
or 50  $\mu\text{g/g}$   
72 hr growth  
Fractionation by cent.

EPS, but not cell growth,  
affected by U.

U(VI) adsorbed onto Fe(III) minerals increased EPS produced by *P. putida*



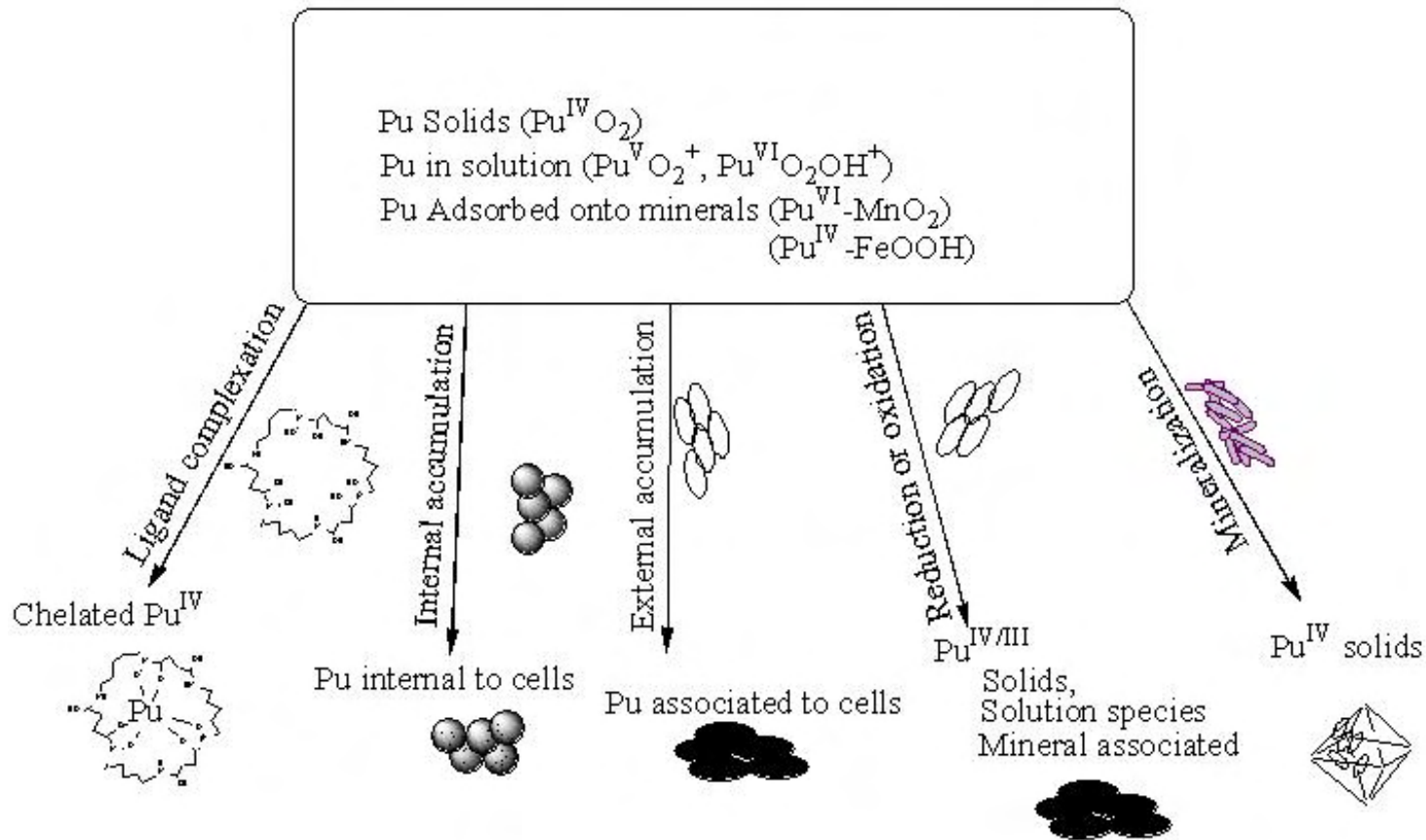
## Distribution of U(VI) within *P. putida* Biofilms



Less U removed from substrate by *P. putida* in the presence of EDTA

U preferentially associated with cell fraction

# Pu Biogeochemistry



## Advances in Pu aqueous geochemistry

Solubilization, biosorption, bioaccumulation, mineralization  
biotransformation mechanisms all affect Pu

Stability of products, rates of combined processes, ...?

# *Acknowledgements*

Dr. Hakim Boukhalfa  
Dr. Gary Icopini  
Mr. Sean Reilly

Dr. Larry Hersman  
Prof. Patricia Holden  
Dr. Cheryl Kuske



U.S. DOE, OSC, OBER, ERSD, NABIR