

***IN SITU* COMMUNITY CONTROL OF THE STABILITY OF BIOREDUCED URANIUM**

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NABIR



We could not do this work without
our excellent collaborators!

**Kelly P. Nevin, Regina T. O'Neil, Helen Vrionis, Irene Ortiz Bernad,
and Derek R. Lovley**

UMASS

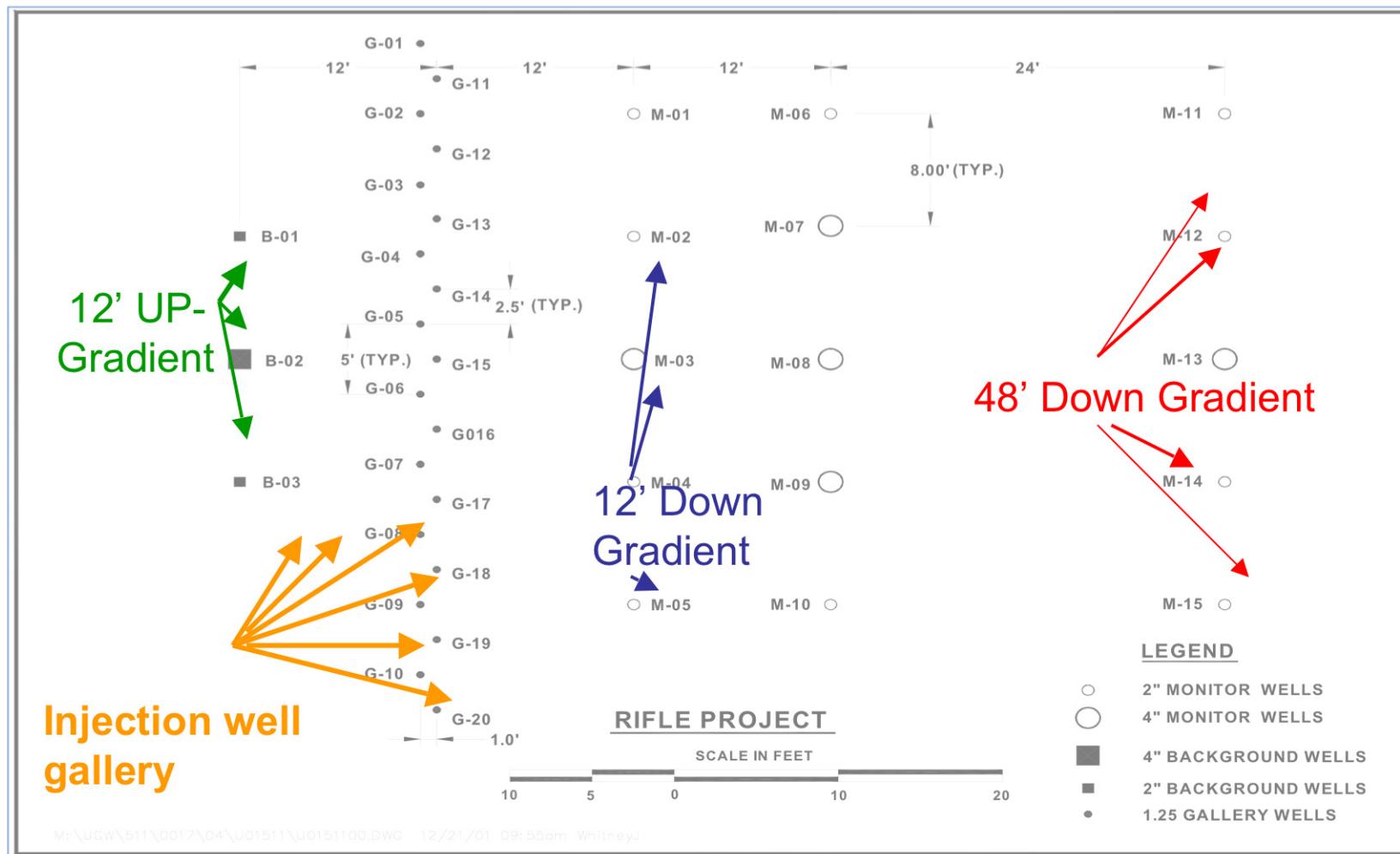
Richard Dayvault
S.M. Stoller Corp.

Darrell Chandler

ANL

Shift of viable biomass & community composition by acetate addition to groundwater ~UMTRA site

Well field

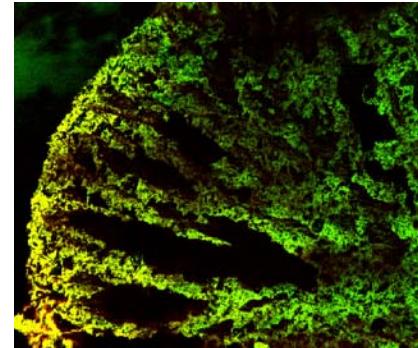


UMTRA Old Rifle site

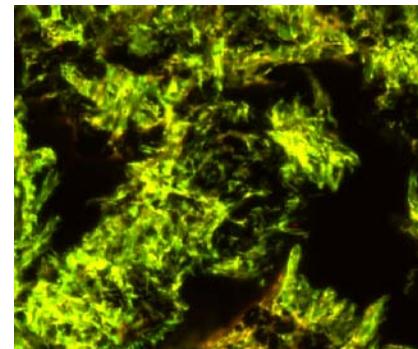


Sampler
with
Beads

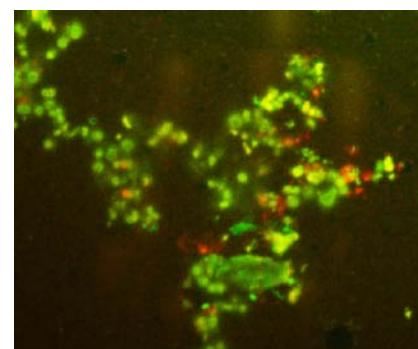
Confocal Laser Scanning Microscopy of beads



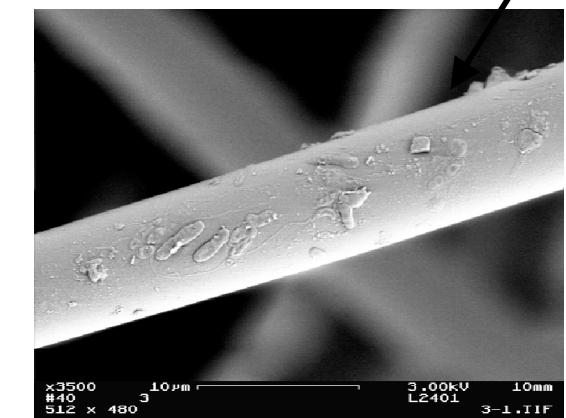
CLSM cross section of
bead showing surface
area for colonization



CLSM close-up of
cross
section



CLSM image
of microbial
colonization
live/dead stain



SEM of microbial
colonization of glass wool

Acetate Infusion induced

A. Changes in the mineralogy  $\text{FeS}_{0.9}$

B. Changes in the microbial community

Program is to test hypotheses as to **How changes in A & B Maintained the decrease in U(VI) in the groundwater**

TEST

Monitor U(VI) loss in-well sediment incubators
different mineral & microbiological amendments

Assessing Subsurface Microbiota

Collecting Microbes

Bead Coupons

Must colonize from water

Actively Growing

^{13}C in DNA & PLFA

Groundwater

mRNA

+ Non Growing

Sediment

Disruptive

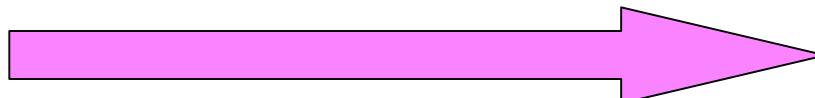
Includes slow & non

Growing

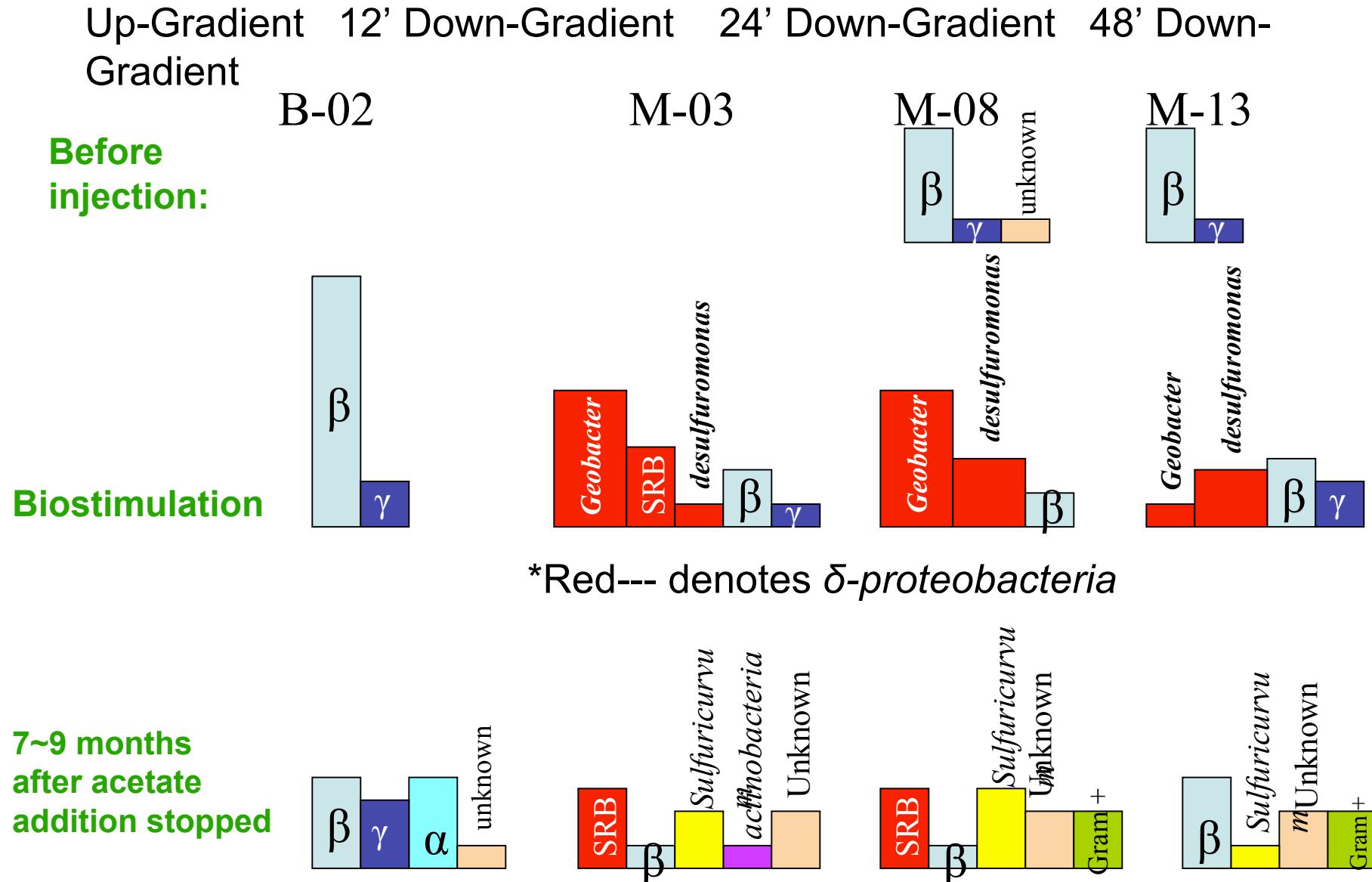
Biomass Intermediate Lowest

Highest

Complexity Increases



Why did U(VI) decrease in groundwater persist 7-9 months after acetate infection stopped? (Bead Coupons)



Preliminary data (DGGE from Bead Coupons) suggests:

Up gradient & before injection stable community dominated by
β-proteobacteria (*Hydrogenophaga*, *Dechloromonas*, *Rhodoferax*,
Ferribacterium, *Rhodocyclus*, *Methylophilus*, *Azoarcus*,
γ-Proteobacteria *Pseudomonas*

A Gram-negative, heterotrophic, carbon-limited, (high
cyclo/monoenoic PLFA), facultative-anaerobic, oxygenated
(UQ/MK~2), utilizing refractory organics

Infusion *Geobacter*, *Desulfuromonas/Pelobacter* DIRB, δ-
Proteobacteria *Desulfobacter*, β-proteobacteria *Ferribacterium*, γ-
Proteobacteria *Pseudomonos*,

Rapid Biostimulation DIRB,SRB goes anaerobic & growth
stimulated

7-9 months later , *Sulfuricurvum*, β-& γ-Proteobacteria, Gram-positive
Clostridia, δ-Proteobacteria SRB

Sulfur oxidizing bacteria use sulfide to → reduce Nitrate/ Oxygen
maintain anaerobic status UQ/MK ~06-0.2 & SRB form U(IV)?

Hypotheses:

- 1) After stimulation Fe(III) terminal electron acceptor ↓ non sulfate-reducing DIRB will be leave or be out-competed by more versatile microbes ? SRB
- 2) SRB & Sulfur oxidizing bacteria play a critical role in the post-treatment maintenance of bio-reduced uranium by directly reducing U(VI), generating H_2S , HS^- and/or $\text{FeS}_{0.9}$ → oxygen sinks maintaining U(IV).
- 3) Bioprecipitated amorphous $\text{FeS}_{0.9}$ in sediments will maintain low U(IV) reoxidation rates under conditions of low biomass, especially in presence **of sulfur oxidizing bacteria**
but $\text{FeS}_{0.9}$ by itself is not sufficient to remove U(VI) from groundwater by abiotic reduction

Conceptual Model

Start Acetate to C-starved β -proteobacter, oxygenated \rightarrow
Anaerobic \rightarrow Geobacter + DIRB \rightarrow reduce Fe(III) \rightarrow great \uparrow
increase biomass \rightarrow Lo \downarrow cal Fe(III) surfaces all reduced \rightarrow wave
Geobacter (+ DIRB) moves distally

Continued acetate \rightarrow SRB increase in diversity (DSR) & biomass
 \rightarrow produce HS⁻ \rightarrow HS⁻ + Fe(II) \rightarrow FeS_{0.9}

Stop Acetate Biomass \downarrow Cell lysis feeds Heterotrophs

Gram-positive Clostridia + Desulfotomaculum (+ SRB) \rightarrow reduce U(VI) & produce acid \downarrow carbonate & U(VI) complex

Sulfur Oxidizing (Sulfuricurvum) use HS⁻ to reduce NO₃ & O₂

Without NO₃ & O₂ Dechloromonas & Geobacter not reoxidize U(IV)

Does Cigar Lake U mine deposit have high FeS_{0.9} + SOB & low UQ/MK?

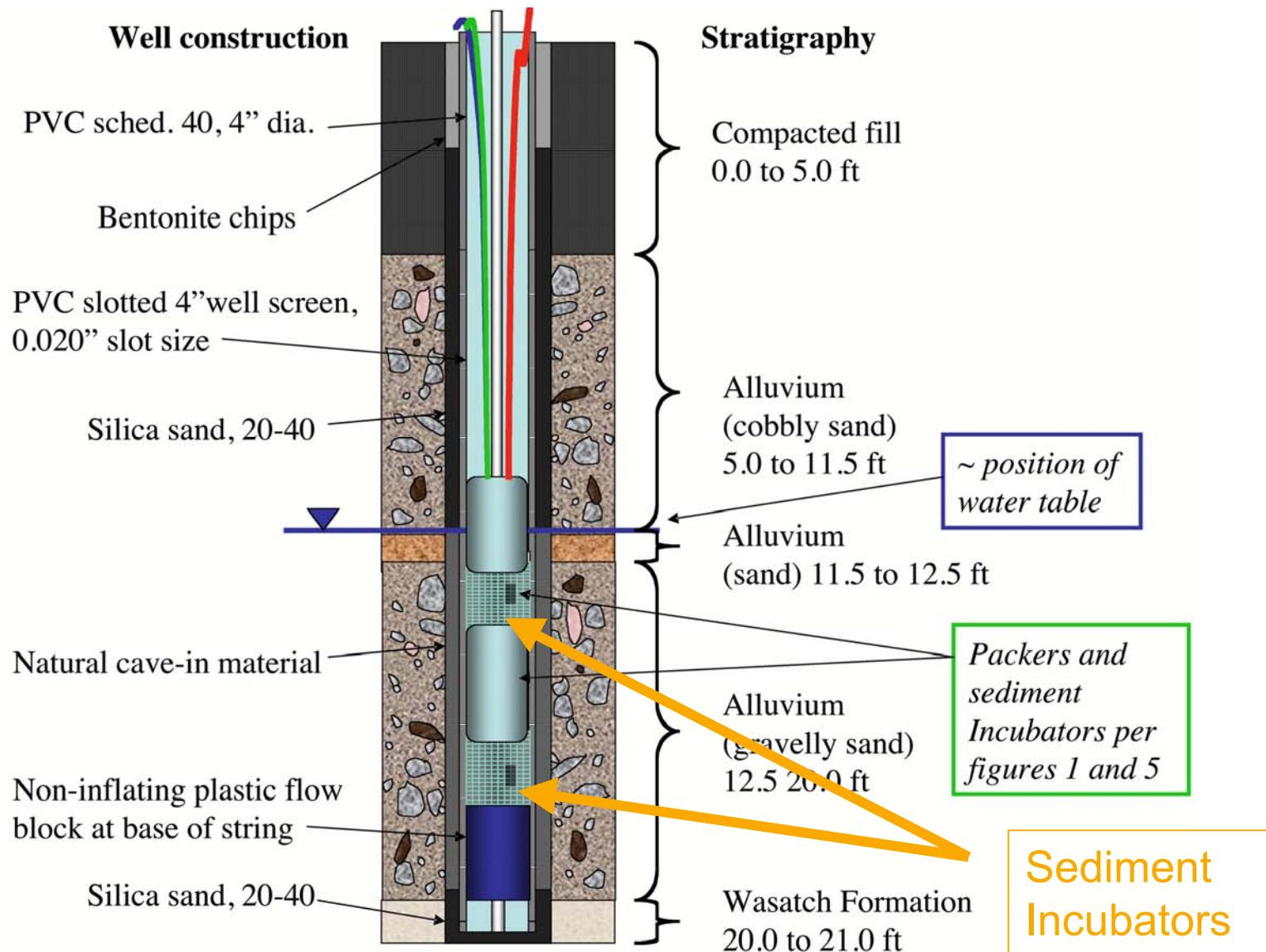


Figure 4. Proposed emplacement of two sediment incubators in a typical 4" well (B-02, Old Rifle UMTRA Site).

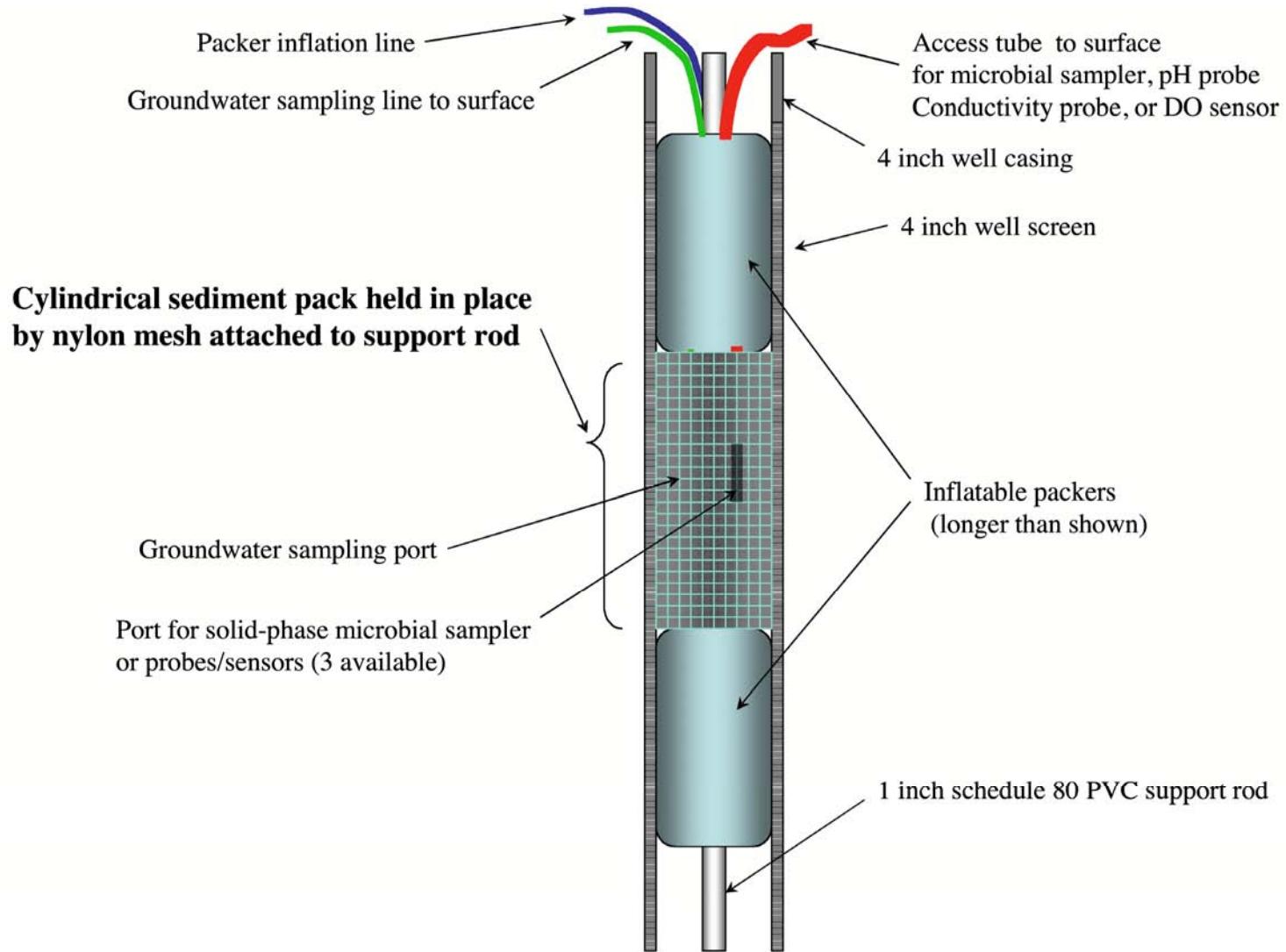


Figure 1. Conceptual design of sediment incubator

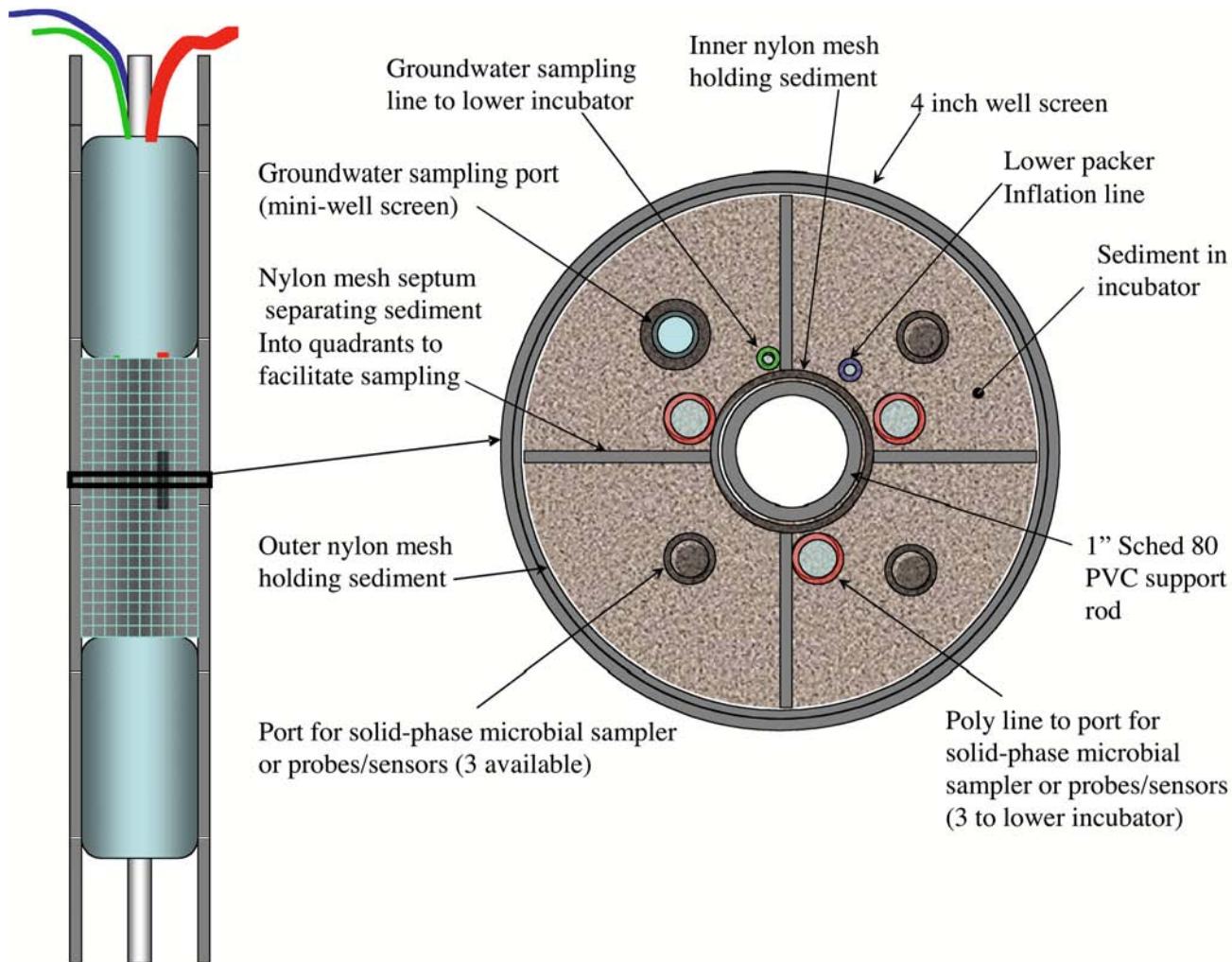


Figure 5. Top view (section) of sediment incubator.

Sediment for the in situ sediment incubators

Rifle sediment Cobles removed & mixed 1-3 mm sieved for higher permeability sand-silt

U(VI) 0.17 mg/L from groundwater during perfusion

Sediment + DIRB + Lactate \rightarrow SRB \uparrow DIRB \downarrow [Hypothesis 1]

Sediment + Lactate + DIRB vs Sediment + Lactate + SRB measure bioreduced U(IV) [Hypothesis 2]

Sediment + FeS_{0.9} Sterile (short time) vs SRB + acetate [Hypothesis 3]

Model: Sediment + FeS_{0.9} + Lactate SRB + SOB

Assessing subsurface microbiota

From Sediment Samplers

DNA 16S rRNA, rDNA,
Genes
DSR (SRB),
DIRB ? NADPH-iron reductase?
soxA sulfite oxidase

by DGGE, Q-PCR, T-RFLP

RNA D. Chandler

Lipids PLFA, Respiratory Quinones, PHA, DMA (Clostridia) ? Spores (DPA)

Better Respiratory Quinone Assessment

Problem present at mmol/mol PLFA

HPLC/electrochemical cell/electrospray/ MS/MS

HPLC separates components so greater duty cycle

Electrochemical cell → Reduces only Quinones at the specific E_o potential : + 112 mV Ubiquinone; + 36 mV Desmethyl Menaquinone;
- 74 mV Menaquinone

100% ionized with - 2 charge → ideal for electrospray ionization

Compare to atmospheric pressure chemical ionization (APCI)
~inefficient H⁺ charge transfer from activated gas

MS/MS search for progenitors ions at products: UQ m/z – 197, - 98.5
DMK m/z - 173, -86.5; MK m/z -187 -93.5

Greatly increase sensitivity → greatly increase localization