SANDIA REPORT

SAND2010-7141 Unlimited Release Printed September 2010

Development of Efficient, Integrated Cellulosic Biorefineries

LDRD Final Report

Christopher R. Shaddix, Ethan Hecht, Kwee- Yan Teh, George Buffleben, Dean Dibble, and Andrew E. Lutz

Prepared by Sandia National Laboratories Albuquerque, New Mexico 87185 and Livermore, California 94550 Sandia National Laboratories is a multi-program laboratory managed and operated by Sandia Corporation, a wholly owned subsidiary of Lockheed Martin Corporation, for the U.S. Department of Energy's National Nuclear Security Administration under contract DE-AC04-94AL85000.



Issued by Sandia National Laboratorie s, operated f or the United States Department of Energy by Sandia Corporation.

NOTICE: This report was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government, nor any agency thereof, nor any of their employ ees, nor any of their contractors, subcontractors, or their employ ees, make any warranty, express or im plied, or assume any legal liability or responsibility for the accuracy , completeness, or usefulness of any information, apparatus, produc t, or process disclosed, or represent that its use would not infringe privatel y owned rights. Reference herein to any specific commercial product, process, or service by trad e name, trademark, manufacturer, or otherwise, does not necessarily constitute or im ply its e ndorsement, recommendation, or favoring by the United States Government, any agency thereof, or any of their contractors or subcontractors. The views and opinions expressed herein do not necessarily state or reflect thos e of the United S tates Government, any agency thereof, or any of their contractors.

Printed in the United States of Am erica. This report has been reproduced directly from the best available copy.

Available to DOE and DOE contractors from U.S. Department of Energy Office of Scientific and Technical Information P.O. Box 62 Oak Ridge, TN 37831

Telephone: (865)576-8401 Facsimile: (865)576-5728 E-Mail: <u>reports@adonis.osti.gov</u> Online ordering: <u>http://www.doe.gov/bridge</u>

Available to the public from U.S. Department of Commerce National Technical Information Service 5285 Port Royal Rd Springfield, VA 22161

> Telephone: (800)553-6847 Facsimile: (703)605-6900 E-Mail: <u>orders@ntis.fedworld.gov</u> Online order: <u>http://www.ntis.gov/help/ordermethods.asp?loc=7-4-0#online</u>



SAND2010-7141 Unlimited Release Printed September 2010

DEVELOPMENT OF EFFICIENT, INTEGRATED CELLULOSIC BIOREFINERIES

LDRD Final Report

Christopher R. Shaddix, Ethan Hecht, Kwee-Yan Teh, George Buffleben, Dean Dibble, and Andrew E. Lutz

> Sandia National Laboratories Livermore, CA 94550

Abstract

Cellulosic ethanol, generated from lignocellulosic biomass sources such as grasses and trees, is a promising alternative to conventional starch- and sugar-based ethanol production in term s of potential production quantities, CO ₂ im pact, and econom ic com petitiveness. In addition, cellulosic ethanol can be generated (at least in principle) without com peting with food production. However, approximately 1/3 of the lignocellulosic biomass material (including all of the lignin) cannot be converted to ethanol thr ough biochemical means and must be extracted at some point in the biochem ical process. In this project we gathered basic inform ation on the prospects for utilizing this lignin residue m aterial in therm ochemical conversion processes to improve the overall energy efficiency or li quid fuel production capacity of cellulosic biorefineries. Two existing pretreatm ent approaches, soaking in aqueous am monia (SAA) and the Arkenol (strong sulfuric acid) process, were implemented at Sandia and used to generated suitable quantities of residue material from corn stover and eucalyptus feedstocks for subsequent thermochemical research. A third, novel technique, using ionic liquids (IL) was investigated by Sandia researchers at the Joint Bioenergy Institute (JBEI), but was not successful in isolating sufficient lignin residue. Additional residue m aterial for therm ochemical research was supplied from the dilute-acid sim ultaneous saccharification/fermentation (SSF) pilot-scale process at the National Renewable Energy Laboratory (NREL). The high-temperature volatiles yields of the different residues were m easured, as were the char combustion reactivities. The residue chars showed slightly lower reactivity than raw biom ass char, except for the SSF residue, which had substantially lower reactivity. Exergy analysis was applied to the NREL standard process design model for therm ochemical ethanol production a nd from a prototypical dedicated biochem ical process, with process data supplied by a r ecent report from the National Research Council (NRC). The therm ochemical system analysis reveal ed that m ost of the system inefficiency is associated with the gasification process and subs equent tar reforming step. For the biochem ical process, the steam generation from residue combustion, providing the requisite heating for the conventional pretreatment and alcohol distillation processes, was shown to dom inate the exergy loss. An overall energy balance with dif ferent potential distillation energy requirem ents shows that as much as 30% of the biom ass energy content may be available in the future as a feedstock for thermochemical production of liquid fuels.

This page left intentionally blank.

Abstract
Table of Contents
List of Figures
List of Tables
Acknowledgments
Introduction11
Project Structure
Biomass Feedstocks and Pretreatment Strategies
Ionic Liquid (IL) Pretreatment14
Soaking in Aqueous Ammonia (SAA) Pretreatment
Arkenol Pretreatment
Dilute-Acid SSF Process Residue
Thermochemical Evaluation of Lignin Residues
Residue Processing and Feeding
Initial Combustion Characterization
Improved Combustion Characterization
System Analysis
Thermochemical Ethanol System
Biochemical Ethanol System
Baker's Yeast Fermentation
Conclusions
List of References
Appendix A: LDRD Project Presentations and Publications
Appendix B: Arkenol Pretreatment Process Steps

Table of Contents

List of	Figures
---------	---------

Page

Figure 1.	Schematic of typical current design layout for a biochemical ethanol biorefinery	13
Figure 2.	Schematic of a potential future design layout for an ethanol biorefinery with primarily biochemical conversion and gasification of lignin residues	13
Figure 3.	Schematic of a potential future design layout for an ethanol biorefinery with primarily biochemical conversion and fast pyrolysis of lignin residues.	14
Figure 4.	Fractionation yields and product compositions from IL processing of corn stover.	16
Figure 5.	Photographs of corn stover and ionic liquid, (top left), stover in solution in heated IL (top right), and glucan-rich product of fractionation, adjacent to raw stover (bottom)	16
Figure 6.	2-D NMR spectra of aromatic entities in corn stover and in its IL- prepared fractions. The circles identify particular structural elements corresponding to guaiacyl ("G") and p-coumarate ("PCA") units, with particular structures indicated schematically. Syringyl structures ("S") appear at the top of the spectra	17
Figure 7.	Photographs of dried, chopped corn stover (left), dried solids remaining after SAA treatment of the corn stover (middle), and lignin residue collected by drying the ammonia effluent (right)	18
Figure 8.	Schematic of the Arkenol pretreatment process, as appeared in Yamada et al., 2002	19
Figure 9.	Photographs of chopped eucalyptus (left) and processed solids remaining after a single hydrolysis step using 77% sulfuric acid	19
Figure 10.	Photograph of custom-designed fine particle feeder for dry coal and biomass samples. For feeding of particularly sticky biomass samples, steel shot can be added to the test tube to help break-up particle aggregates	20
Figure 11.	Schematic of the Sandia's optical entrained flow reactor facility, with a Hencken burner providing a high-temperature, high flow furnace environment and a particle-sizing pyrometry diagnostic measuring the temperature, size, and velocity of individual reacting particles	21

Figure 12.	Photograph of a devolatilizing stream of lignin residue produced by SAA pretreatment of corn stover. Furnace temperature of 1500 K	21
Figure 13.	Photographs taken through a microscope of SAA-generated lignin residue from corn stover knife-milled and sieved into different size classes: (a) fines, $< 37 \mu m$, (b) 75-106 μm , (c) 106-180 μm	22
Figure 14.	Photographs of 75-106 µm corn stover SAA lignin residue burning in a 1200 K furnace with 12%, 24%, 36%, and 60% O ₂ , respectively	23
Figure 15.	Photographs of 106-180 μ m particles of different lignin residues burning in 36%, O ₂ in a 1700 K furnace. The leftmost two photos show two different corn stover SAA pretreatment residues and the rightmost photo shows a eucalyptus SAA process residue	23
Figure 16.	Schematic of Sandia's Pressure-Capable Entrained Flow Reactor (PCEFR)	24
Figure 17.	Photographs of combustion of chars generated from raw and Arkenol- pretreated eucalyptus and of direct combustion of residues from pretreated corn stover in the optical EFR at 1700 K. "%" labels refer to oxygen content of flow. "Arkenol-1" refers to single-step hydrolysis and "Arkenol-2" refers to dual-step hydrolysis	25
Figure 18.	Mean corn stover residue char combustion particle temperatures measured as a function of height in Sandia's optical EFR when operating at 1700 K.	26
Figure 19.	Mean eucalyptus residue char combustion particle temperatures measured as a function of height in Sandia's optical EFR when operating at 1700 K.	27
Figure 20.	Mean corn stover residue char combustion particle temperatures measured as a function of height in Sandia's optical EFR when operating at 1700 K.	27
Figure 21.	Simplified block flow diagram of NREL's canonical thermochemical ethanol process design	29
Figure 22.	Energy inputs and outputs (left pie chart) and exergy inputs and outputs (right pie chart) for the 2012 design case thermochemical ethanol process	29
		29

Figure 23.	(Left) The dual-bed, indirect steam gasifier model overlaid on the NREL process flow diagram. (Right) Gasifier system exergy inputs and outputs	30
Figure 24.	Schematic of the modeled gasifier block system (left) and the calculated exergy loss of the system as a function of the excess air supplied to the char combustor (right)	31
Figure 25.	(Left) The alcohol synthesis reactor overlaid on the NREL process flow diagram. (Right) Synthesis reactor exergy inputs and outputs	31
Figure 26.	Schematic of the modeled wood dryer block system (left) and the calculated exergy loss of the system as a function of the inlet air temperature (right)	32
Figure 27.	Exergy budget for the thermochemical ethanol design case using 2007 and 2008 "state of technology" performance assumptions and the target 2012 design case	33
Figure 28.	Schematic of a prototypical lignocellulosic biochemical ethanol plant, utilizing separate enzymatic hydrolysis and fermentation steps	34
Figure 29.	Energy inputs and outputs (left pie chart) and exergy inputs and outputs (right pie chart) for the "low-performance" biochemical process design case	34
Figure 30.	Exergy budget for the prototypical biochemical conversion process based on NRC technology performance scenarios	35
Figure 31.	Energy consumption for ethanol distillation as a function of the ethanol in the feed to the distillation process and the technology employed (data from Madson and Lococo, 2000; Vane, 2008; Cote et al., 2008)	36
Figure 32.	Trade-off between electricity and thermochemical biofuel production using lignin-rich residue from a prototypical biochemical conversion process	36
Figure 33.	Schematic of modeled system of continuous culture of baker's yeast fermenting glucose under anaerobic, glucose-limited conditions	37
Figure 34.	Exergetic efficiencies of fermentation process, as a function of the cellular ATP consumption	37

List of Tables	Page
Table 1. Assumptions for NRC Low-, Medium-, and High-Performance Processes	22
110005505	

Acknowledgments

Manfred Geier, a postdoctoral research associate, assisted with som e of the experiments on lignin residue devolatilization and char combustion.

Introduction

This report documents the results of a Laborator y Directed Research and Developm ent (LDRD) project with the aim of improving the design and performance of future biorefineries that convert lignocellulosic biom ass sources into ethanol and specialty chem icals. Dwindling supplies of conventional oil and increasing concern over greenhouse gas em issions have m otivated the search for renewable liquid fuels. Biom ass-derived liquid fuels are prom ising candidates. particularly in the U.S., with its large capace ity for generating biom ass (Perlack et al., 2005). Currently, farm and ethanol subsidies and a gua ranteed ethanol m arket have fostered the production of a large corn ethanol m arket. However, corn-based ethanol production engenders significant CO₂ production and has lim ited potential to offset U.S. oil consum ption. Far greater production potential and im proved reductions of CO ₂ are possible when converting lignocellulosic biom ass (grasses, trees, crop re sidues, etc.) into ethanol. This prom ising technology has seen substantial research invest ments for several years, prim arily focused on purely biochemical processing of the biom ass to yield ethanol and specialty chem icals and on purely therm ochemical processing of the biom ass to yield sim ilar products. Indeed, the U.S. Department of Energy office that oversees most of this research, the Office of Biomass Programs (OBP), has divided its biom ass conversion program into these two focal areas for a num ber of years.

One of the primary advantages of the biochemical conversion process is the ease of handling and feeding biomass feedstocks into the process (at the pretreatment stage) and the limited sensitivity to the biomass moisture level, because of the aqueous nature of the conversion process itself. For the thermochemical process, higher ethanol fuel yields are possible and the process shows low sensitivity to the type of biom ass feedstock, b ecause all of the biom ass can be effectively gasified. Biochemical processing can typically only convert 60-70% of the biom ass feedstock to sugars, for subsequent ferm entation to ethanol, because of the existence of several com ponents of the biomass, particularly lignin, protein, and extractives, that do not contain sugar polymers. At present, the limitations on ethanol yield of the dedicated biochemical process are not seen as particularly restrictive, because the pretr eatment and ethanol distillation steps require a substantial quantity of steam, which can just be generated by burning all of the lignin-rich residue from the process, as indicated in Fig. 1. However, efforts are being made to improve the performance of the biochem ical process, partic ularly to reduce the heat requirem ents of the pretreatment process (by operation at a lower temperature) and the distillation process (by incorporating m embrane-based separation/dis tillation, for exam ple). Progress in reducing the steam requirem ents of these processes will then "liberate" the energy content of the lignin residue for potential use in a therm ochemical "bottoming cycle," as suggested in the diagram s shown in Figures 2 and 3. The prospect for su ch hybrid biochem ical/thermochemical ethanol biorefinery designs motivates the research conducted in this LDRD project.

Project Structure

This project com bined an investigation of stat e-of-the-art biom ass pretreatm ent technologies with assessments of the com bustion and gasifica tion potential of realistic lignin extracts. In addition, energy and exergy balances were computed for benchm ark therm ochemical and biochemical conversion processes, to illuminate those process components that have the most

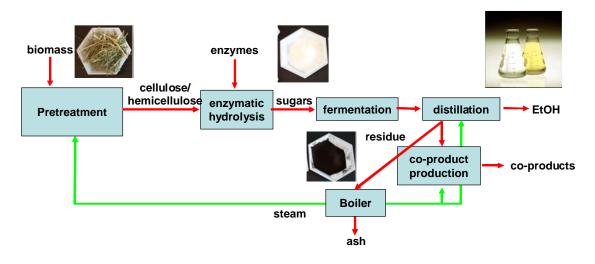


Figure 1. Schematic of typical curre nt design layout for a biochem ical ethanol biorefinery.

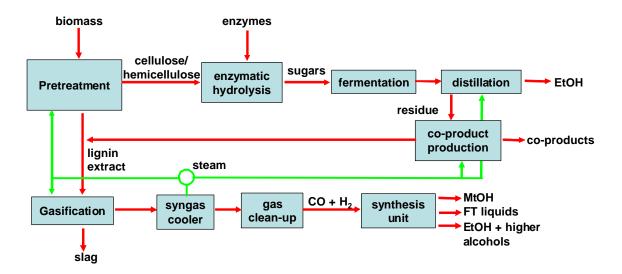


Figure 2. Schematic of a potential future design layout for an ethanol biorefinery with prime arily biocheme ical conversion and gasification of lignin residues.

promise for yielding efficiency im provements. A particular focus of the project was on sim ple, low-cost pretreatm ent processes that directly separate lignin f rom the cellulose in the pretreatment step. These pretreatm ents have not been widely investigated, because from the point-of-view of a dedicated biochem ical etha nol process, m aximizing the conversion of both hemicellulose and cellulose to ferm entable sugars is key to effective ethanol production. However, when considering hybrid biochem ical/thermochemical ethanol biorefineries, a lowcost (and, especially, low-tem perature) pretreat ment process that extracts som e hem icellulose with the lignin m ay be beneficial to the overall process efficiency and ethanol product yield in comparison to dilute acid pretreatm ent and other approaches that optim ize biochem ical conversion of cellulose and hem icellulose. In formation on the com bustion, gasification, or pyrolysis behavior of lignin-rich residues from biochemical conversion processes is virtually non-existent in the literature (see Blunk and Je nkins, 2000; Eriksson et al., 2004; Öhm an et al., 2006; and Li et al., 2008, for the only known exam ples), so work on this project should prove to be a vital source of information for designing appropriate thermochemical conversion devices.

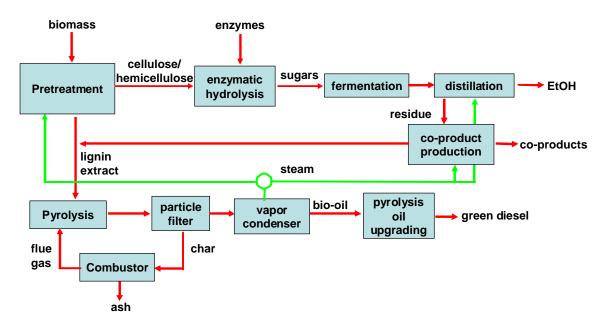


Figure 3. Schematic of a potential fu ture design layout for an ethanol biorefinery with prim arily bi ochemical conversion and fast pyrolysis of lignin residues.

Biomass Feedstocks and Pretreatment Strategies

A wide variety of biom ass pretreatm ent appr oaches have been developed to augm ent the biochemical conversion of different biomass sources to ethanol (Kumar et al., 2009; Alvira et al., 2010). The general goals of biom ass pretreatm ent for application to subsequent enzym atic hydrolysis are to increase the physical access to the cellulose and hem icellulose, to reduce the crystallinity of the cellulose, and to separate the lignin f rom the cellulose and hem icellulose (lignin blocks surface sites on the cellulose and he micellulose from enzymatic attack). Different pretreatment approaches achieve these goals to different extents and with differing financial and energy costs. Acid-based pretreatm ent approaches chem ically attack cellulose, whereas basic pretreatments directly attack the lignin. For this project, focus was given to im plementing pretreatment approaches that would directly vield a lignin-rich residue, both because these approaches have not been widely investigated and because these approaches could m ost easily yield the 100's of g of residue that was requi red for subsequent therm ochemical conversion analysis. The dilute sulfuric acid pretreatm ent, ammonia fiber expansion (AFEX) approach, and hydrothermal (steam expansion) approaches yield a residue at the end of the biochemical process and therefore were not investigated in this project. The three techniques that were im plemented

and investigated during the course of this pr technique known as "soaking in aqueous am process research, which occurred at the Joint Bi oenergy Institute (JBEI), was highly speculative and in the end did not yield sufficient usable characterization. The other two approaches were implemented at the SNL/CA site and did yield substantial lignin residue that was subsequently investigated for thermochemical processing.

The biom ass f eedstocks that were utilized during this research were all derived f rom larger feedstock samples that were acquired for research at JBEI. These feedstocks included corn stover (a representative and widely investigated U.S. agricultural residue), switchgrass (a widely investigated herbaceous U.S. energy crop), and eucalyptus (a prom ising fast-growing softwood tree).

Ionic Liquid (IL) Pretreatment

In contrast to m ost proposed biom ass pretr eatment technologies, whose basic properties are reasonably well characterized, the ionic liquid pr etreatment approach is a new and possibly revolutionary approach and thus has become the focus of pretreatment research at JBEI. The promise of ionic liquids is to elim inate the need for a separate pretreatm ent step and to rapidly hydrolyze the biom ass polysaccharides (cellulose and hem icellulose) under m ild conditions in which the depolymerization is highly selective (Sievers et al., 2009). Ionic liquids are essentially degree of dissociation into liquid-phase ions. liquid-phase salts, and therefore retain a strong This quality makes them powerful solvents. The first application of ionic liquids to dissolution of cellulose was reported in 2002 (Swatloski et al.), but this required a tem perature of ~ 100 °C. Subsequent research developed ILs that can effectively dissolve cellulose at near room temperatures (Ohno and Fukaya, 2007). Dissolution of whole biom ass is substantially m ore challenging than dissolution of cellulose, however, particularly because of the lignin component of lignocellulosic biomass. During the course of research into ionic liquid pretreatment as part of this project, a novel ionic liquid processing sche me for fractionating biomass was developed and has resulted in an expected patent filing (see Appendix). A couple of technical presentations have been made on this research (see Appendix) and a journal publication on the IL pretreatment has also being submitted (Dibble et al., 2010). Figure 4 shows a flow diagram that indicates the type of biom ass fractionation that results from the IL process that has been developed. Fig. 5 shows photographs of ionic liquid treatm ent of corn stover and the glucan-rich product stream from this process.

Two-dimensional nuclear m agnetic resonance (2-D NMR) im aging was perform ed at UC Berkeley on the different fractions produced by the IL processing as a way to track the disposition of lignin, per the technique reporented by Kim et al. (2009, 2010). Fig. 6 shows the aromatic regions of the structure, indicative of lignin moieties. Particular structural entities are highlighted with circles and are defined belo w the figure. Sim ilar NMR im aging has been conducted on both raw biom ass and different pretre eated sam ples, to com pare the effects of pretreatment on the lignin bonding. This will be presented later in this report.

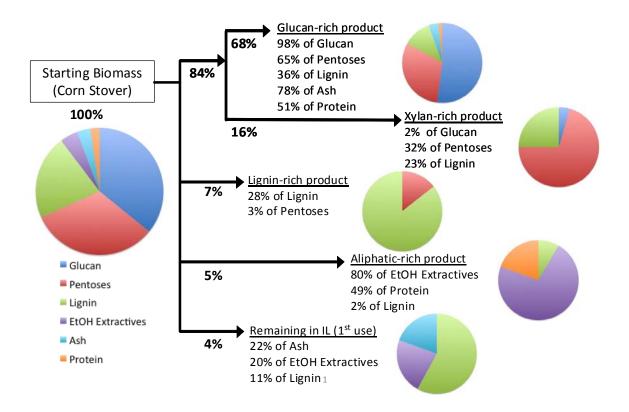


Figure 4. Fractionation yields and product compositions from IL processing of corn stover.

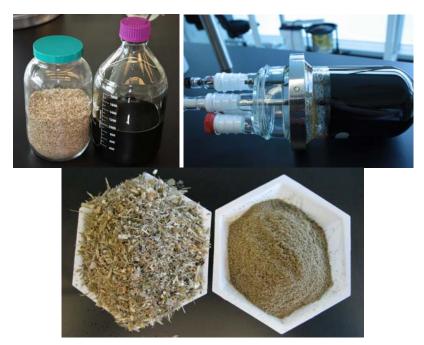


Figure 5. Photographs of corn stover a nd ionic liquid, (top left), stover in solution in heated IL (top right), and glucan-rich product of fractionation, adjacent to raw stover (bottom).

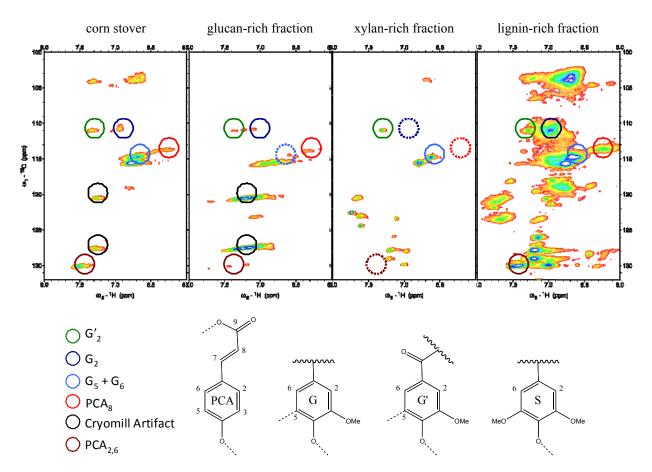


Figure 6. 2-D NMR spectra of arom atic entities in corn stover and in its IL-prepared fractions. The circles identify particular structural elem ents corresponding to guaiacyl ("G") and p-coum arate ("PCA") units, with particular structures indicated schem atically. Syringyl structur es ("S") appear at the top of the spectra.

Soaking in Aqueous Ammonia (SAA) Pretreatment

A simple, low-energy pretreatment technique known as "soaking in aqueous am monia" (SAA) that yields lignin residues directly from the pretreatm ent process was im plemented, as this process is easy to scale-up to the sizable proce ssing runs required to generate sufficient residues for thermal processing. This technique was developed by Prof. Yoon Lee of Auburn University (Yoon et al., 1995; Kim et al., 2003; Kim and Lee, 2005, 2007) and has been m ost recently applied by his form er student, T.H. Kim (form erly of the USDA and currently at Iowa State University) (Kim et al., 2008; Li et al., 2010). R ecently this technique was applied to the pilotscale pretreatment of switchgrass in 4 kg increments in a 75 L vessel (Himmelsbach et al., 2009). In this process, the lignin is solubilized in the ammonia solution, together with a portion of the hemicellulose (linked C5 and C6 sugars). Subs equent drying of the solution yields a lignin residue. For optim ized conditions with a 60 $^{\circ}$ C processing tem perature, Kim and Lee (2007) report 62% lignin removal and 15% xylan removal from corn stover. Xylan removal was $\sim 30\%$ and lignin removal $\sim 60\%$ when applying SAA to barley hulls at 75°C (Kim et al., 2008). While these removal fractions of hem icellulose make SAA somewhat disfavored for application in a dedicated biochem ical biorefinery, wherein the so lubilized hem icellulose is not available f or subsequent fermentation to ethanol, this low-co st, low-energy pretreatment approach may prove to be advantageous in bioref ineries that u tilize integrated biochem ical and therm ochemical processing, wherein hemicellulose solubilization does not represent a conversion loss.

In this project, the m oderate-temperature SAA process was refined and successfully applied to two different samples of corn stover (Midwest and California source material) and to eucalyptus wood. A 1:5 m ixture ratio of dry biom ass relative to 15% NH ₄OH solution was used, and the mixture was heated to 60 °C and allowed to r eact for 37 hr. Chem ical analysis of the solids derived from the SAA effluent from the two corn stover process treatm ents show that it is composed of ~ 50% lignin, with the remainder a mixture of hemicellulose sugars and inorganics. Fig. 7 shows photographs of the initial chopped corn stover feedstock, processed solids, and dried lignin residue (recovered from the ammonia wash).



Figure 7. Photographs of dried, chopped corn SAA treatm ent of the corn stover drying the ammonia effluent (right). stover (left), dried solids rem aining after (middle), and lignin residue collected by

Arkenol Pretreatment

The Arkenol pretreatm ent process, which uses st rong sulfuric acid to separate lignin from the cellulose and hem icellulose, was developed in the m id-1990's and was im mediately patented (Farone and Cuzens, 1997, 1998; Cuzens and Mille r, 1997). Currently, BlueFire Renewables owns the rights to the Arkenol process and is in the process of building com mercial plants in Lancaster, CA, and in Fulton, MS, to pr oduce ethanol from MSW and wood residues, respectively. A schematic of the Arkenol pretreat ment process is shown in Fig. 8. In the strong acid process, the cellulose and hem icellulose are hydrolyzed and go into solution with the acid, leaving the lignin behind. Residual acid in the sugar solution is neutralized with lim e, yielding solid gypsum as a byproduct. In this project, the Arkenol process was applied to the eucalyptus sample to provide a com parison with the resi due derived from eucalyptus by applying the SAA process. The detailed procedure used for applying the Arkenol process is included as Appendix B. As a process summary, the raw biomass was reacted with 77% sulfuric acid and heated to 100 °C, then allowed to react for one hour. As shown in Fig. 8, in com mercial applications the hydrolysis process is generally applied in two stages, where the second stage provides additional removal of the carbohydrates from the lignin. In this project, samples of the lignin residue from both one-step hydrolysis and two-step hydrolysis were produced for experiments on residue

thermochemical conversion. Fig. 9 shows photogr aphs of the raw eucalyptus wood and the lignin-containing processed solids after a single hydrolysis step.

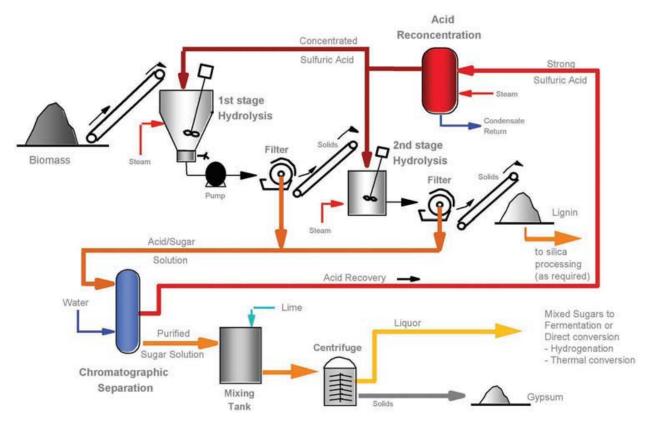


Figure 8. Schematic of the Arkenol pretreatm ent process, as appeared in Yam ada et al., 2002.



Figure 9. Photographs of chopped eucalyptus (left) and processed solids remaining after a single hydrolysis step using 77% sulfuric acid.

Dilute-Acid SSF Process Residue

The most widely used lignocellulose pretreatment approach is undoubtedly dilute sulfuric acid pretreatment, which does not effectively hydr olyze the carbohydrates and therefore requires

subsequent enzym atic hydrolysis (e.g. as s detoxification, to com plete the hydrolysis "saccharification"). The National Renewable configuration that has been investigating and optimizing dilute sulfuric acid pretreatment of corn stover followed by sim ultaneous saccharifica tion and ferm entation (SSF), a cost-reducing intensification of the process path. After distillation of the ethanol, the lignin-containing process dregs are rem oved. For this project, NREL research ers provided us with a sam ple of the lignin residue from their SSF process, when processing corn stover.

Thermochemical Evaluation of Lignin Residues

The goals of this portion of the project were to (a) evaluate m ethodologies for handling and feeding the lignin residues, because biom ass particle feeding is generally an im portant practical consideration in both laboratory and com mercial therm ochemical processes, (b) evaluate the high-temperature, high-heating-rate volatiles content of the residues, because this is an important parameter in determining carbon conversion and minimum reactor size needed for combustion or gasification, and (c) measure the combustion/gasification reactivity of the residue char particles, for comparison to the existing experimental database of coals of various ranks.

Residue Processing and Feeding

The SAA corn stover lignin residues were dried and then initially were pulverized using an "Attritor" stirred-ball m ill. This m aterial was found to be feedable, with som e plugging difficulties, in our new pulverized fuel feeder system for combustion analysis, shown in Fig. 10.

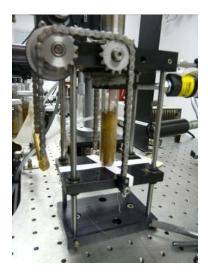


Figure 10. Photograph of custom -designed fine particle feeder for dry coal and biom ass samples. For feeding of particularly sticky biom ass samples, steel shot can be added to the test tube to help break-up particle aggregates. With use of this feeder, high-tem perature devolatilization experiments were conducted by operating our optical entrained flow reactor with a slightly fuel-rich fuel-oxidizer feed to the Hencken burner (see Figs. 11-12).

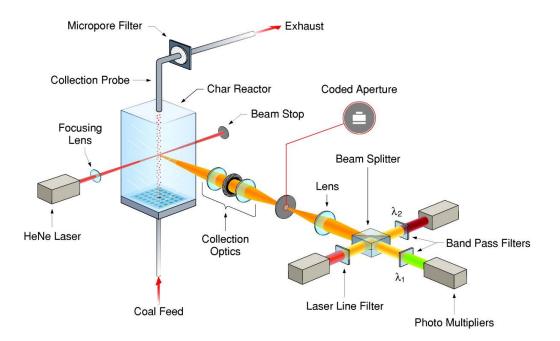


Figure 11. Schematic of the Sandia's optical entrained flow reactor facility, with a Hencken burner providing a high-tem perature, high flow furnace environm ent and a particle-sizing pyrom etry diagnostic m easuring the tem perature, size, and velocity of individual reacting particles.



Figure 12. Photograph of a devolatilizing stream of lignin residue produced by SAA pretreatment of corn stover. Furnace temperature of 1500 K.

The particle size distribution of the residue processed with the Attritor was analyzed using an optical microscope, revealing that the overwhelming preponderance of particles were $< 10 \ \mu m$ in size and tended to form large particle agglomerates (i.e. the material behaved as fine dust). Both feeding and chem ical kinetic analysis of combustion and gasification rates is improved for somewhat larger particles (on the order of 100-150 μm in size), so the lignin residues were resolidified and then m illed with a W iley knife mill, fitted with a custom -designed screen to provide an appropriately sized product output. The knife-milled residue was then sieved into three size classes and examined under a microscope, as shown in Fig. 13. The microscopy revealed that the particles were roughly sphenical in shape (as opposed to the long fibrous particles generally produced from milling raw biomass) and segregated well in the sieving process.

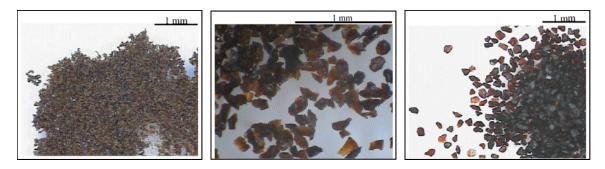


Figure 13. Photographs taken through a m icroscope of SAA-generated lignin residue from corn stover knife-milled and sieved into different size classes: (a) fines, < 37 μm, (b) 75-106 μm, (c) 106-180 μm.

Initial Combustion Characterization

Initial combustion experiments were performed at a low furnace tem perature of 1200 K and oxygen concentrations of 12-60 vol-%. Previous m easurements in our laboratory have demonstrated that high oxygen concentrations in the flow are conducive to determ ination of the char combustion kinetic rates, because low oxyge n concentrations can lead to oxygen diffusive control of the particle burning rate (Murphy a nd Shaddix, 2006). Lignin residue particles in the 75-106 µm size class were used for these experiments, because this particle size class has proven to be the optim al size for determination of the char combustion kinetics of coal. Fig. 14 shows time-lapse photographs of the lignin particle s burning in the furnace at different oxygen concentrations. As the oxygen concentration incr eases, the particles appear to burn hotter (as denoted by the color of the visible em ission from the particles) and burn out m ore quickly (as evidenced by the length of the visible particle str eaks). In addition, the particles ignite earlier for a higher oxygen content of the flow. Attem pts at perform ing single-particle particle-sizing pyrometry on the lignin residue particles proved di fficult, because the particles would ignite at different heights in the furnace and then burnout relatively quickly. In addition, the optical signals were weak and the deduced particle sizes were too sm all (~ 60 μ m) to be reliably measured with the coded aperture optical technique. It was deduced that the sm all apparent char particle sizes resulted from extensive devolatilization (small biomass particles can lose upwards of 90% of their mass to volatilization when heated rapidly to high tem peratures) (Bharadwaj et al., 2004). To overcom e these difficulties, it was deci ded to feed a larger size fraction of m illed residue (106-180 μ m) and to use a high-temperature furnace environment of 1700 K, to minimize variations in the ignition height of different residue particles from a given sample and to provide a direct com parison with an extensive database of coal char reactivity inform ation available at this temperature. Photographs of the com bustion of these particles are shown in Fig. 15. W hile some optical particle m easurements were possibl e, the variation in the ignition height of the particles, their rapid burnout, variable particle feed rates (because of difficulty feeding), and particle dispersion within the furnace (because of their high volatiles loss and the jetting action of volatiles) m ade the m easurements inef ficient, with consum ption of m uch of the lim ited samples yielding only a few good op tical measurements. The optical m easurements did show a mean char particle size of ~ 90 μ m, validating the choice of particle sizes in the feed.

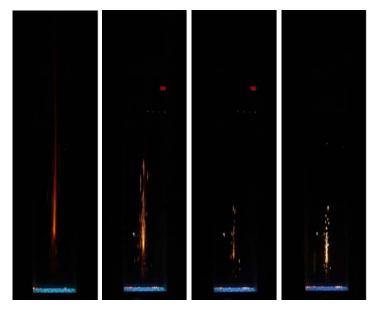


Figure 14. Photographs of 75-106 μm corn stover SAA lignin residue burning in a 1200 K furnace with 12%, 24%, 36%, and 60% O₂, respectively.

Improved Combustion Characterization

To im prove upon both the quality and quantity of optical data collected, we decided to devolatilize the residue particles in an electrica lly heated, downflow, enclosed reactor known as the Pressure-Capable Entrained Flow Reactor (PCEFR), shown in Fig. 16. These devolatilized particles were then fed into the optical entrai ned flow reactor for characterization of their combustion reactivity. By devolatilizing the particles before feeding into the combustion reactor, the particles tend to stay at the reactor centerlin e, leading to m ore efficient capture of optical data. Also, as dem onstrated recently in a sepa rately funded project evaluating coal particle combustion, pre-generation of char particles removes the effect of variations in the devolatilization process in the optical entrained flow reactor when raw particles are fed into environments with different oxygen concentra tions. For the lignin residues (and for their associated raw biomass feedstock), a particle size range of 75-180 µm was fed into the PCEFR

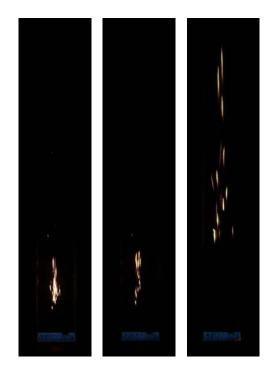


Figure 15. Photographs of 106-180 μm particles of different lignin residues burning in 36%, O₂ in a 1700 K furnace. The leftm ost two photos show two different corn stover SAA pretreatm ent residues and the ri ghtmost photo shows a eucalyptus SAA process residue.

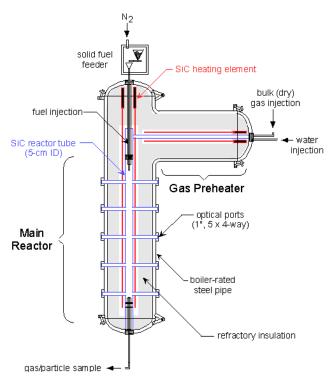


Figure 16. Schematic of Sandia's Pr Reactor (PCEFR).

essure-Capable Entrained Flow

when flowing N $_2$ at 1200 °C, and the collected char particles were received to 75-106 μ m for feeding into the optical EFR.

Analysis of the volatile yield f rom the lignin residue particles is awaiting com pletion of commercial lab analysis of the ash content of the char sam ples and will be reported in a subsequent journal article on this work. Si milarly, we are awaiting the results of f ull proximate/ultimate analysis of the residues them selves. Photographs of the com bustion of the eucalyptus char and those of its Arkenol residues in the optical EFR at 1700 K are shown in Fig. 17, together with photographs of corn stove r residues directly fed into the furnace. Unfortunately, there was insufficient corn stover residue sam ple to generate enough char material for optical characterization of the co mbustion of the char feed. The differences in apparent burnout height of the different char materials probably reflect differences in densities of the chars, because the char sam ples were sieved to be in the sam e initial size range. A denser particle of the same size as another will inherently take longer to burn out, simply because there is more carbonaceous m ass to consum e. On this basis, it appears that the raw eucalyptus char particles are substantially lighter than those produced from the Arkenol-pretreated eucalyptus. The SAA pretreated corn stover particles bur n out very quickly, while those from the SSF process take substantially longer to burn, possible y reflecting the $\sim 16\%$ as h in these particles (typ. analysis provided by NREL). The raw eucalypt us char and, especially, the SSF residue, show substantial vellow emission in a wide band extending outside of the burning particles. This

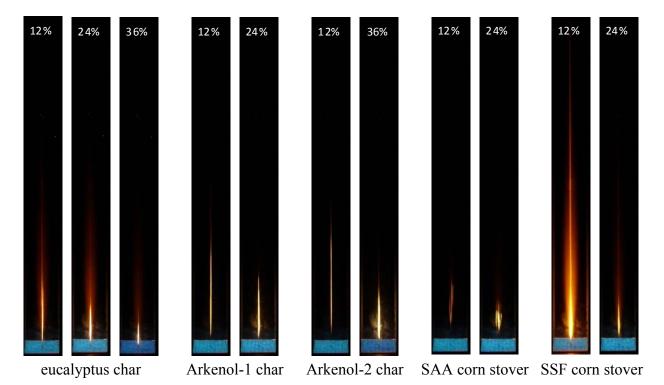


Figure 17. Photographs of combustion of chars generated from raw and Arkenol-pretreated eucalyptus and of direct com bustion of re sidues from pretreated corn stover in the optical EFR at 1700 K. "%" labels refer to oxygen content of flow. "Arkenol-1" refers to single-step hydrolysis and "Arkenol-2" refers to dual-step hydrolysis.

observation in the optical EFR is typically associ ated with Na release f rom the particles. The results of ash analysis of the feedstocks and the different residues should confirm the presence of significant sodium in the eucalyptus and in the SSF corn stover residue. The strong sulfuric acid Arkenol pretreatment undoubtedly removes alkali metals from the eucalyptus and carries them in the acid-sugar solution. In the SAA pretreatment, the alkali metals probably remain with the solids and thus are segregated from the lignin residue that is produced from the ammonia wash.

In a given com bustion environment, sim ilar-size particles will burn at dif ferent tem peratures according to the rate at which they are reacting. The oxidati on of carbon and hydrogen components of the char particle releases energy, su ch that the more quickly a particle burns, the greater its com bustion tem perature (Glassm an and Yetter, 2009). This fact can be used to evaluate the relative reactivity of different fuels, based on their com bustion temperatures in a given environm ent, or, with a detailed energy ba lance, can be used to derive overall char combustion kinetic rate parameters (Murphy and Shaddix, 2006). Mean char particle temperature measurements for the various residues are show n in Figs. 18 and 19, as a function of height above the burner in the optical EFR. Fig. 18 show s that the SSF residue has substantially lower reactivity than the SAA residue., which might be attributed, at least in part, to the significant ash content of the SSF residue. Fig. 19 shows that the raw eucalyptus has slightly higher reactivity than the Arkenol residues, as one m ight expect. Somewhat surprisingly, the one-pass and twopass Arkenol residues show nearly identical pa rticle combustion temperatures, suggesting little chemical difference in the chars form ed from these residues with different degrees of hydrolytic removal of carbohydrates.

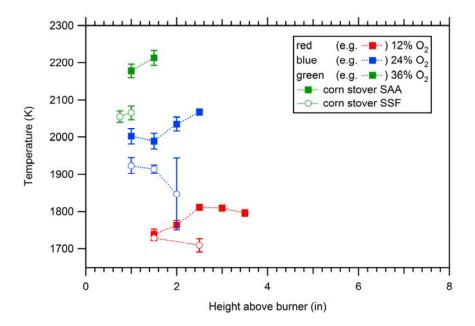


Figure 18. Mean corn stover residue char combustion particle temperatures measured as a function of height in Sandia's optical EFR when operating at 1700 K.

From the profiles shown in Figs. 18 and 19, the peak mean particle temperatures can be used as a measure of the characteristic particle reactivity, specifically relative to oxygen as an oxidizer, but

also more generally towards any gasifying reactant, whether it is oxygen, carbon dioxide, or steam. In this way, the reactivity of these resi dues in comparison to typical coals used in combustors and gasifiers can be evaluated. Fig. 20 shows a comparison of the peak mean particle temperatures from the biomass residue chars in comparison to those previously measured for the same furnace conditions for the full rank spectru m of pulverized coal, ranging from the m ost reactive coals, lignite, to the least reactive, anthracite (Shaddix et al., 2009). Because of the similarity in peak m ean temperatures for the one-pass and two-pass Arkenol residues, a single profile for Arkenol residues is shown in Fig. 20.

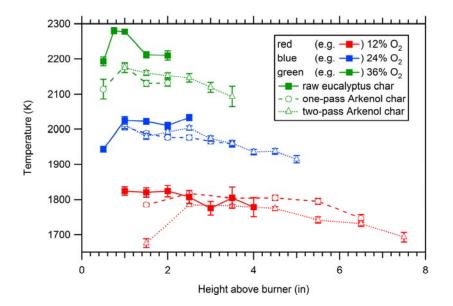


Figure 19. Mean eucalyptus residue char com bustion particle temperatures measured as a function of height in Sandia's optical EFR when operating at 1700 K.

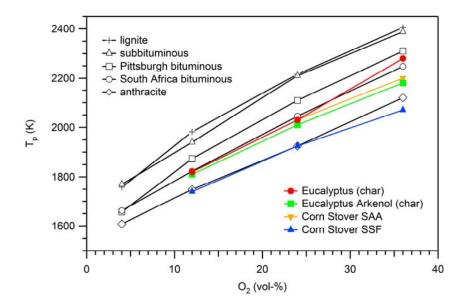


Figure 20. Mean corn stover residue char combustion particle temperatures measured as a function of height in Sandia's optical EFR when operating at 1700 K.

Not unexpectedly, the char formed from the raw eucalyptus has the highest reactivity of all of the biomass samples investigated. The SAA treated corn stover and the Arkenol process eucalyptus char particles have very sim ilar reactivities, while the SSF process corn stover residue has a substantially lower reactivity. The overall rank position of the biom ass chars *is* surprising. Normally one would expect raw biom ass to have at least as great a reactivity as lignite. However, the eucalyptus chars were prepared *ex situ*, in the PCEFR, with a residence tim e of over 1 second, which m ay decrease the reactivity com pared to an *in situ* formed char. Also, in support of the findings here, W ornat et al. (1996) measured the com bustion reactivity of two different biom ass chars (from Southern pine a nd switchgrass) that were produced from flash pyrolysis of biom ass at 625 °C and found them to be equivalent to *in situ* formed high-volatile bituminous coal chars (such as the Pittsburgh and South Africa coals shown in Fig. 20). Of course, one caveat to this comparison of apparent reactivities of the biomass chars and coal chars is that the coal chars tend to be largely spheri cal, whereas the biomass-derived chars (for all but the SAA-treated corn stover) tend to have disp arate aspect ratios along the plant grain, which facilitates heat transfer from the particle to the surrounding gas and thereby lowers their apparent reactivity relative to coal chars. This consider ation requires further study to help quantify this effect for the different biom ass sam ples. Furtherm ore, a calibration check perform ed after completion of the biomass sample measurements shows that the two-color pyrometer calibration constant needs to be adjusted upward for the biom ass data - this correction is in the process of being fully quantified.

System Analysis

Work on this portion of the project began by conducting an exergy analysis of a canonical thermochemical process for lignocellulosic ethanol production that has been evaluated by NREL using ASPEN Plus software. After this, an exergy and enthalpy analysis was conducted on a canonical biochemical process for cellulosic etha nol production. In addition, a detailed analysis was conducted of the biochem ical fermentation process with baker's yeast, a commonly used fermentation organism.

Thermochemical Ethanol System

NREL researchers analyzed the hypothetical perf ormance of a lignocellulosic ethanol plant based on indirect gasification of wood chips with subsequent tar reform ing, syngas cleanup, syngas compression, and finally m ixed alcohol production over a catalyst (Phillips et al., 2007; Phillips 2007). A schematic of this system is shown in Fig. 21. In the LDRD project, an exergy balance for the NREL process m odel for therm ochemical biom ass-to-ethanol conversion was constructed, in collaboration with NREL researchers, from the input and output process variables in the ASPEN Plus m odel of the system that was developed at NREL. NREL has conducted "state-of-the-art" analyses of the system based on evaluations of current technology in 2007 and 2008, as well as projected improvements that are targeted to be available by 2012. The projected, 2012 performance is considered the default design case. A com parison of the overall energy budget and overall exergy budget for the 2012 desi gn case is shown in Fig. 22. The exergy balance is based on an evaluation of the ability of the system to "do work" and thereby is a more meaningful measure of system efficiency than can be gleaned from an energy balance, which

does not discriminate between low-grade heat energy and electricity, for exam ple (Dincer 2002; Rosen et al., 2008; Rosen and Bulucea, 2009). The results in Fig. 22 show that the ethanol and higher alcohol products of the system only represent 42% of the input exergy, com pared to representing 59% of the input energy. Thus, the overall process efficiency is not as large as one might believe based on the energy balance. In addition, whereas the energy balance implicates compressor heat loss and the air-cooled heat exchanger as being the dominant inefficiencies in

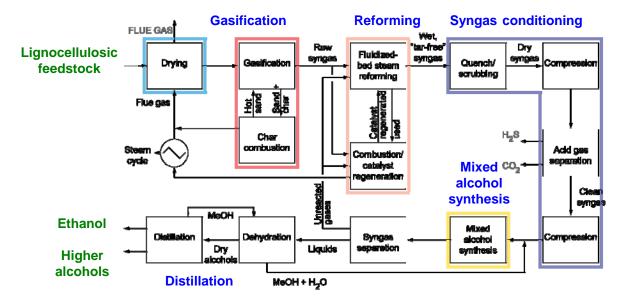


Figure 21. Simplified block flow diagram of NREL's canonical thermochemical ethanol process design.

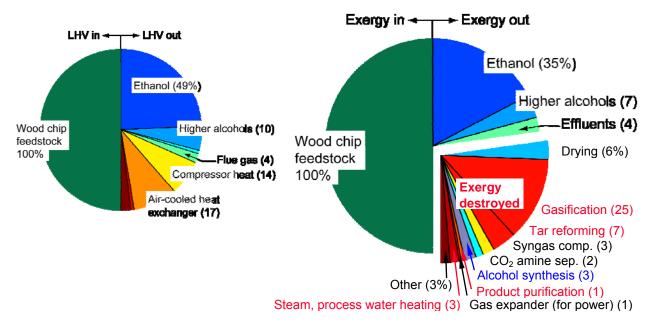


Figure 22. Energy inputs and outputs (left pi e chart) and exergy inputs and outputs (right pie chart) for the 2012 design case thermochemical ethanol process.

the system, the exergy analysis shows that the ga sification process and the tar reforming step are the primary sources of inefficiency, totaling over r 30% of the input exergy and over 50% of the exergy 'loss' in the system. In contrast, the heat transfer losses that dominate the energy losses in the system only amount to 6% of the exergy budge t, because of the preponderance of low-grade heat that has limited ability to do work.

Further insights are gained by looking at the energy and exergy budgets of the individual process steps. The low-temperature, low-pressure (1270 K, 1.7 atm) indirect steam gasifier was found to have an exergetic efficiency of 71%, as shown in Fig. 23. In comparison, the NREL study determined that the gasifier first-law efficiency was 76%. The exergy analysis showed that approximately one-half of the exergy loss was associated with the gasifier itself and half with the char combustor. A significant am ount of the input exergy (9%) is contained in the flue gas, so there exists the opportunity to improve the efficiency of this process with additional heat recovery from the flue. The extensive production of hydrocarbons in this low-temperature gasifier represents a significant fraction of the system exergy ($\sim 1/3$ of the input exergy).

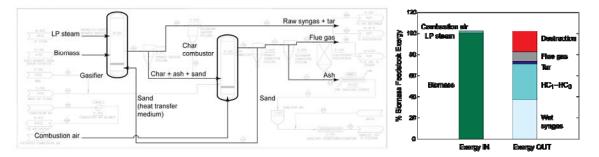


Figure 23. (Left) The dual-bed, indirect st eam gasifier m odel overlaid on the NREL process flow diagram. (Right) Gasifier system exergy inputs and outputs.

An analysis was also conducted of the influence of the indirect gasifier operating temperature on exergy loss. For this analysis the presum ed amount of char com bustion excess air was varied (from the 20% excess assum ed in the NREL analys is) and the fuel feed and steam inputs were assumed constant. NREL correlations for syngas a nd char composition versus temperature were used, and the two reactors were assum ed adiabatic. The results of this analysis are shown on the right side of Fig. 24. Reducing the com bustion air supply from 20% to 10% in excess of the stoichiometric amount for com plete char com bustion raises the gasification tem perature from 1605 °F to 1620 °F (874 °C to 882 °C), and adiabatic char combustion temperature from 2190 °F to 2220 °F (1200 °C to 1215 °C). Correspondingly, the exergy destruction decreases from 9% to 8.5% of feedstock exergy. Lowering the steam -to-feedstock ratio (set at 0.4 kg/kg in NREL studies) serves the sam e purpose of raisi ng operating tem perature and reducing exergy destruction. In fact, for high tem perature entr ained flow gasifiers, for which the syngas approaches chem ical equilibrium (Kovacik et al., 1990), one can show that the gasification process has to be carried out at the highest possi ble energy state (highest tem perature, pressure) to minimize exergy destruction. The analytical proof was solved for internal combustion engines (Teh et al., 2008), but the results m ay also be extended to gasification. However, in the latter case, one needs to weigh the exergy destruction minimization/efficiency maximization objective of the gasification process without losing sight of its primary function, which is to supply syngas

of certain desired characteristics (e.g., H₂-to-CO ratio) for the synthesis step downstream . Also, high temperature gasification systems for producing syngas generally require the use of oxygen as a reactant, and the im pact of oxygen gene ration on the overall process exergy budget would need to be evaluated.

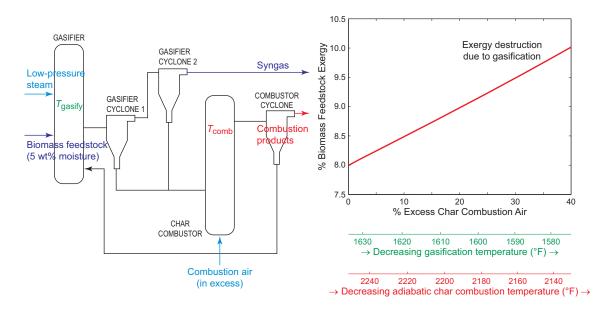


Figure 24. Schematic of the m odeled gasifier block system (left) and the calculated exergy loss of the system as a function of the excess air supplied to the char combustor (right).

The alcohol synthesis reactor was modeled as a high-P, isothermal (570 K, 67 atm) process akin to the Fischer-Tropsch process, but using a sulfided m olybdenum catalyst (MoS ₂). Using the catalyst performance targets specified by NREL, the synthesis process was found to be nearly reversible: only 2% exergy input is destroyed and the process has an overall exergetic efficiency of 95% (see Fig. 25). The NREL perform ance targets specify 60% CO conversion (*cf.* state-oftechnology: 10–40%) and 90% alcohol selectivity (*cf.* state-of technology: 70–80%). Given the aggressive performance targets and the proximity of the resultant process to an ideal, reversible

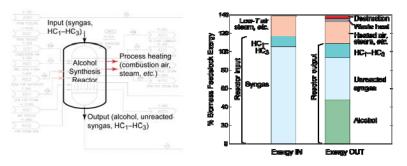


Figure 25. (Left) The alcohol synthesis r eactor overlaid on the NREL process flow diagram. (Right) Synthesis reactor exergy inputs and outputs.

process, it appears that the NREL perform achievable.

Figure 22 shows that feedstock drying represents a 6% exergy loss from the system. To evaluate the potential for reducing this inefficiency, calculations were perform ed for the wood chip dryer as a function of the moisture content of the feed and the inlet air tem perature to the dryer, while holding the dried wood moisture content (5%) and dried wood and air exit tem peratures (220 °F and 230 °F) constant (see Fig. 26). The system was assumed to be adiabatic (i.e. no heat losses through the walls). With these assumptions, the exergy loss is seen to be a strong function of the moisture content of the raw feedstock. Lowering raw feedstock moisture level from 50 wt%, per NREL studies, to 40 wt%, for instance, would reduce exergy destruction by a third. At a 20 wt% moisture content—near the level for air-dried lumber—the exergy destroyed drops to about 1% of biom ass feedstock exergy. The drying process efficiency also increases, albeit to a m uch smaller extent, with decreasing inlet air te mperature, i.e., accom plished using sensible energy/thermal resource of lower quality—but at a higher quantity. At 50 wt% m oisture level, decreasing the inlet air temperature from 1200 °F to 900 °F (650 °C to 480 °C), while increasing the flow rate accordingly, reduces the exergy de stroyed by 10%. The trend dim inishes at lower moisture levels.

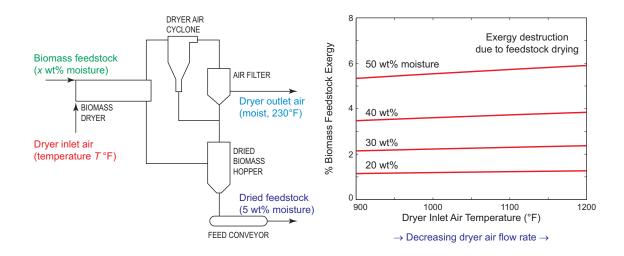


Figure 26. Schematic of the modeled wood dryer block system (left) and the calculated exergy loss of the system as a function of the inlet air temperature (right).

In addition to this detailed exam ination of the 2012 thermochemical process design case, overall evaluations were also perform ed of the 2007 a nd 2008 "State of Technology" evaluations that were performed by NREL using ASPEN Plus. This analysis allowed a quantitative assessment of the impact of the thermochemical process performance improvements projected by NREL on the production of ethanol and higher alcohols and is summarized in Fig. 27. By reducing exergy losses, the exergetic efficiency of producing etha nol and higher alcohol is seen to progress from 24% in 2007 to 31% in 2008 and 42% in 2012. From 2007 to 2008 there is a significant drop in the exergy losses associated with CO ₂ separation and process heating. From 2008 to 2012,

significant reductions in exergy loss associated with tar reform ing and syngas com pression are forecast, based on improvements in catalyst activity and selectivity.

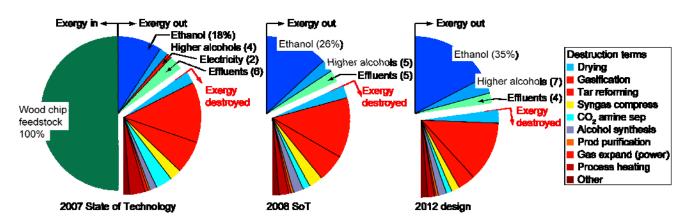


Figure 27. Exergy budget for the therm ochemical ethanol design case using 2007 and 2008 "state of technology" perform ance assumptions and the target 2012 design case.

Biochemical Ethanol System

We analyzed the efficiency of biom ass-to-ethanol fuel production based on prototypical biochemical conversion technologies (using either hot water or dilute acid pretreatment followed by enzymatic hydrolysis - see Fig. 28). W e considered three representative scenarios developed for a 2009 National Research Council study on altern ative liquid transportation fuels (National Research Council, 2009). The critical assum ptions used for the three different scenarios are shown in Table 1. The "low-perform ance" scenar io represents current technology; "m ediumperformance" represents where the technology is likely to be after reasonable, evolutionary advancements (by 2020); "high-perform ance" represents the m ost optim istic technology advancements (as a lim iting case). Thus, the lo w-performance and high-perform ance scenarios provide realistic bounds on the expected behavior of a dedicated biochemical ethanol plant.

Table 1:	Assumptions for	NRC Low-, Medium-	-, and High-Performa	nce Processes
----------	-----------------	-------------------	----------------------	---------------

Variable	Low	Medium	High
Solids loading	18%	21%	25%
Pretreatment yield	80%	85%	95%
Saccharification extent	90%	95%	95%
Fermentation yield, glucose	85%	90%	90%
Fermentation yield, xylose	75%	81%	81%

The results of our energy and exergy analysis of this process for the low-performance scenario are shown in Figure 29. The process efficiency desirable outputs) is 38% based on an energy Heat

(considering ethanol and electricity as the balance and 33% based on the exergy balance.

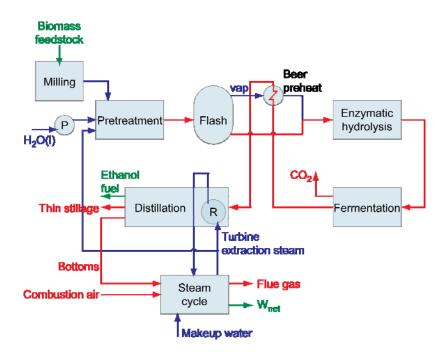


Figure 28. Schematic of a prototypical lignocellulosic biochem ical ethanol plant, utilizing separate enzymatic hydrolysis and fermentation steps.

loss is the principal source of energy loss, wher eas the steam cycle is the prim ary source of exergy loss (i.e. is the prim ary source of ineffici ency in the system). At first glance, it would appear that exergy destruction is low for a ll the biochem ical sub-processes (pretreatm ent, enzymatic hydrolysis, and ferm entation), as well as ethanol distillation. However, it should be noted that the exergetically inefficient steam cycle serves to meet the steam and power demands of the pretreatment, distillation, and feedstock size reduction steps.

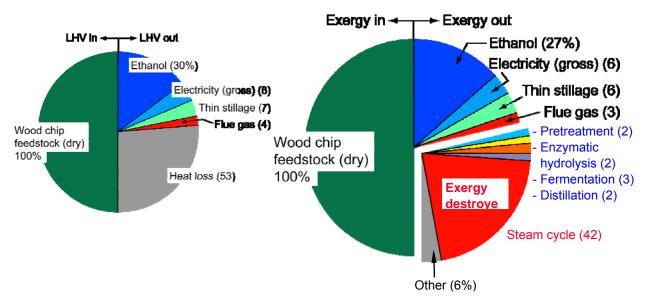


Figure 29. Energy inputs and outputs (left pi e chart) and exergy inputs and outputs (right pie chart) for the "low-performance" biochemical process design case.

Figure 30 shows the exergy budget im plications of the expected evolution of the current biochemical process technology to future and id ealized process states. The overall exergetic efficiency improves from 33% (using current "l ow" technology) to 38%, with a possible further improvement to 42% (under the m ost optimistic "high" technology assum ption). The prim ary source of this efficiency improvement lies in reduced steam generation requirements.

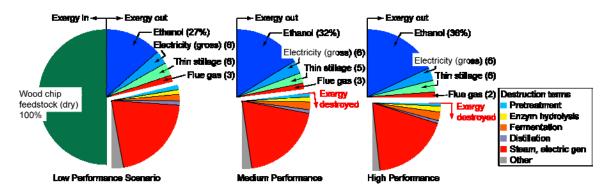


Figure 30. Exergy budget for the prototypical biochemical conversion process based on NRC technology performance scenarios.

The large exergy loss associated with the st eam cycle has im plication f or the potential integration of the bio- and therm o-chemical approaches to produce lignocellulosic ethanol, since the process steam and electricity needs reduce the availability of lignin-rich residue (f rom the biochemical conversion process) as feedstock for subsequent thermochemical conversion to fuel. The steam requirements for ethanol distillation may decrease significantly in the future, with the commercial adoption of m embrane vapor permeation technology. The projected energy consumption of this distillation technology is a pproximately one-third the energy consum ption of conventional, low-capital cost distillation and dehydration technology, as shown in Fig. 31.

The heating value of the available lignin residue provides a quantitative m easure to investigate the feasibility of an integrated bio-/therm o-chemical biofuel production concept. Figure 32 illustrates the trade-off between available residue heating value and gross electricity production. after steam requirem ents f or pretreatm ent a nd distillation processes have been satisf ied. Estimates of electricity use during biochem ical fuel production vary widely, depending on the assumed system configuration and scale, feedstock, etc. For instance, two NREL process designs specified electricity consumption per gallon ethanol production to be, respectively, 1.4 kW h and 5.3 kWh (equivalent to 0.5 MJ and 1.4 MJ per kg dr y biomass feedstock). As shown in Figure 32, our analysis indicates that electricity use at the high end would leave little excess lignin-rich residue for therm ochemical fuel production, assum ing that electricity is co-generated (with steam) via com bustion of the residue. Available, excess residue increases by 3% to 4% of original feedstock LHV if the process steam requirement for ethanol separation decreases based on more-advanced membrane vapor perm eation technologies. On the other hand, advances in biochemical fuel production (based on NRC t echnology outlook) would m ean better utilization of feedstock sugars, leaving sm aller quantities of recalcitrant (unhydrolyzed) cellulose and hemicellulose in the residue for additional ther mochemical production of biofuel. Assuming that the USDA or lower NREL estimate for electricity requirement is accurate, residue with around

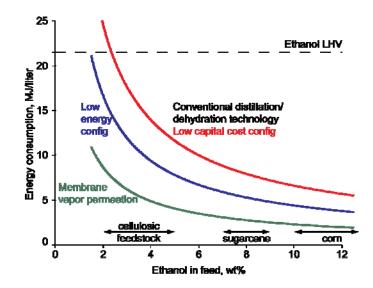


Figure 31. Energy consum ption for ethanol distillation as a function of the ethanol in the f eed to the distillation process and the technology em ployed (data from Madson and Lococo, 2000; Va ne, 2008; Cote et al., 2008).

20–30% of the original biom ass feedstock LHV content m ay be diverted from electricity generation and m ade available for therm ochemical biofuel production. Furtherm ore, if improvements are m ade in the pretreatm ent process that reduce the pressure and quantity of steam required for pretreatm ent even larger am ounts of the feedstock LHV will be available for thermochemical processing.

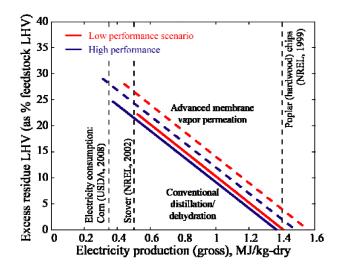


Figure 32. Trade-off between electricity and therm ochemical biofuel production using lignin-rich residue from a prototypical biochemical conversion process.

Baker's Yeast Fermentation

We analyzed the thermodynamics of microbial growth with respect to fermentation of glucose to ethanol by baker's yeast (*S. cerevisiae*), a model fermentation organism with a well-understood metabolism. The m odel couples a detailed cellula r m etabolic flux analysis with bulk changes across a fermentation reactor, as shown schematically in Fig. 33. Results of this work have been published by Teh and Lutz (2010) and will ther efore only be briefly sum marized here. The exergy analysis showed that the anaerobic m etabolism of yeast is very efficient, retaining m ore than 90% of the exergy of the growth m edium. However, a portion of the metabolism is devoted to cellular production and to the generation of m etabolic by-products other than ethanol, such that the glucose-to-ethanol efficiency is less than 75%. Fig. 34 shows these results as a function of ATP consumption, which reflects the extent of cell stress/activity.

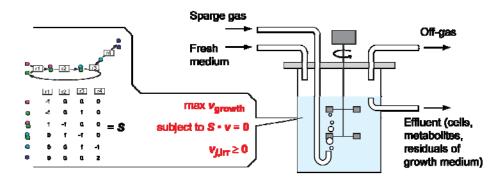


Figure 33. Schematic of modeled system of continuous culture of baker's yeast fermenting glucose under anaerobic, glucose-limited conditions.

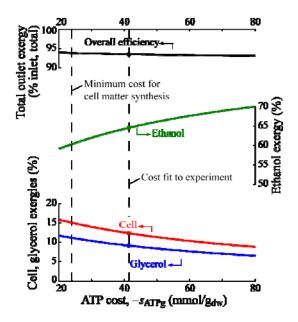


Figure 34. Exergetic efficiencies of fermentation process, as a function of the cellular ATP consumption.

Conclusions

In this LDRD project, research was perform ed on pretreatment strategies to rem ove lignin from lignocellulosic feedstocks, on the therm ochemical conversion characteristics of such lignin-rich residues, and on using exergy analysis to iden tify inefficiencies in therm ochemical and biochemical approaches to cellulosic ethanol production. The overall goal of this work was to lay the foundation for m ore efficient and m ore productive conversion of lignocellulosic feedstocks to ethanol via an integrated process wherein excess lignin-rich residues from the base biochemical process are processed therm ochemically to yield additional liquid fuel or high-efficiency electricity.

Two existing pretreatm ent approaches, soaki ng in aqueous am monia (SAA) and the Arkenol process (utilizing strong sulfuric acid), were im plemented and used to generated sufficient residue m aterial from corn stover and eucalyptus feedstocks for subsequent therm ochemical research. Ionic liquid (IL) processing of biom ass was investigated by Sandia researchers at the Joint Bioenergy Institute (JBEI), but was not succe ssful in isolating sufficient lignin residue for thermochemical characterization. Additional residue material for therm ochemical research was supplied from the dilute-acid sim ultaneous saccharification/ferm entation (SSF) pilot-scale process at the National Renewable Energy Labor atory (NREL). The high-tem perature volatiles yields of the different residues were m easured, as well as the char com bustion reactivities. The residue chars showed slightly lower reactivity than raw biomass char, except for the SSF residue, which had substantially lower reactivity.

Exergy analysis was applied to the NREL standa rd process design m odel for cellulosic ethanol production from a dedicated therm ochemical process (utilizing an indirect gasifier) and from a prototypical dedicated biochem ical process, with process data supplied by a recent report from the National Research Council (NRC). The therm ochemical system analysis revealed that m ost of the system inefficiency is associated with the gasification process and subsequent tar reforming step. For the biochem ical process, the steam generation from residue com bustion, providing the requisite heating for the conven tional pretreatm ent and alcohol distillation processes, was shown to dom inate the exergy lo ss. An overall energy balance with different potential distillation energy requirem ents shows that as m uch as 30% of the biom ass energy content may be available in the future as a feedstock for therm ochemical production of liquid fuels.

List of References

- P. Alvira, E. Tom ás-Pejó, M. Ballesteros, M. J. Negero, "Pretreatm ent technologies for an efficient bioethanol production process ba se don enzym atic hydrolysis: a review," *Biores. Technol.* 101:4851-4861 (2010).
- A. Bharadwaj, L.L. Baxter, A.L. Robinson, "Eff ects of intraparticle heat and m ass transfer on biomass devolatilization: experim ental results and m odel predictions," *Energy Fuels* 18:1021-1031 (2004).
- S.L. Blunk, B.M. Jenkins, "Com bustion propertie s of lignin residue from lignocellulosic fermentation," Report prepared for Na tional Renewable Energy Laboratory, NREL Subcontract ACG-8-18021-01, April 2000.
- P. Cote, G. Noel, S. Kroll, M. Schwartz, M. Kazmir, "From design to operation of a 2 Mgal/y membrane-based ethanol dewatering system," Fuel Ethanol Workshop, 2008.
- J.C. Cuzens, J.R. Miller, "Acid hydrolysis of bagasse for ethanol production," *Renewable Energy* 10:285-290 (1997).
- D.C. Dibble, C. Li, L. Sun, A. George, A. Cheng, Ö.P. Çetinkol, P. Benke, S. Singh, B.A. Simmons, "Ionic liquid pretreatm ent with gel- free precipitation and IL recycling utilizing a phase-switchable solvent system," submitted to *Green Chemistry* (2010).
- I. Dincer, "The role of exergy in energy policy making," Energy Policy 30:137-149 (2002).
- G. Eriksson, G. Kjellström, B Lundqvist, S. Paulrud, "Combustion of wood hydrolysis residue in a 150 kW powder burner," *Fuel* 83:1635-1641 (2004).
- W.A. Farone, J.E. Cuzens, U.S. patent no. 5,620,877 (1997).
- W.A. Farone, J.E. Cuzens, U.S. patent no. 5,726,046 (1998).
- I. Glassman, R.A. Yetter, Combustion, 4th Ed., Academic Press: Amsterdam, 2008, pp. 531-542.
- J.N. Himmelsbach, A. Isci, D.R. Raman, R.P. Anex, "Design and testing of a pilot-scale aqueous ammonia soaking biomass pretreatment system," *Appl. Engin. Agric.* 25:953-959 (2009).
- H. Kim, J. Ralph, "Solution-state 2D NM R of ball-milled plant cell walls in DMSO-*d*₆/pyridine-*d*₅," *Org. Biomol. Chem.* 8:576-591 (2010).
- H. Kim, J. Ralph, T. Akiyam a, "Solution-st ate 2D NMR of ball-m illed plant cell walls in DMSO-*d*₆," *Bioenergy Res.* 1:56-66 (2009).
- T.H. Kim, J.S. Kim, C. Sunwoo, Y.Y. Lee, "P retreatment of corn stover by aqueous am monia," *Biores. Technol.* 90:39-47 (2003).

- T.H. Kim, Y.Y. Lee, "Pretreatm ent of corn stover by soaking in aqueous am monia," *Biores. Technol.* 121-124:1119-1132 (2005).
- T.H. Kim, Y.Y. Lee, "Pretreatm ent of corn st over by soaking in aqueous am monia at moderate temperatures," *Biores. Technol.* 136-140:81-92 (2007).
- T.H. Kim, F. Taylor, K. Hicks, "Bioethanol production from barley hull using SAA (soaking in aqueous ammonia) pretreatment," *Biores. Technol.* 99:5694-5702 (2008).
- G. Kovacik, M. Oguztoreli, A. Cham bers, B. Ozum, "Equilibrium Calculations in Coal Gasification," *Int. J. Hydrogen Energy* 15:125–131 (1990).
- P. Kumar, D.M. Barrett, M.J. Delwiche, P. Stroeve, "Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production," *Ind. Eng. Chem. Res.* 48:3713-3729 (2009).
- W.-Z. Li, Y.-J. Yan, T.-C. Li, Z.-W . Ren, M. Huang, J. W ang, M.-Q. Chen, Z.-C. Tan, "Preparation of hydrogen via catalytic gasification of residues from biomass hydrolysis with a novel high strength catalyst," *Energy Fuels* 22:1233-1238 (2008).
- X. Li, T.H. Kim, N.P. Nghiem, "Bioethanol production from corn stover using aqueous ammonia pretreatment and two-phase sim ultaneous saccharification and ferm entation (TPSSF)," *Biores. Technol.* 101:5910-5916 (2010).
- P.W. Madson, D.B. Lococo, "Recovery of volatile products from dilute high-fouling process streams," *Appl. Biochem. Biotech.* 84-86:1049-1061 (2000).
- J.J. Murphy, C.R. Shaddix, Combust. Flame 144:710-729 (2006).
- National Research Council, <u>Liquid Transportation Fuels from Coal and Biomass: Technological</u> <u>Status, Costs, and Environm ental Impacts</u>, National Academ ies Press, ISBN: 978-0-309-13712-6, 2009.
- M. Öhm an, C. Bom an, H. Hedm an, R. Ekl und, "Residential com bustion perform ance of pelletized hydrolysis residue from lignocellulosic ethanol production," *Energy Fuels* 20:1298-1304 (2006).
- H. Ohno, Y. Fukaya, "Task specific ionic liquids for cellulose technology," *Chem. Lett.* 38:2-7 (2009).
- R.D. Perlack, L.L. Wright, A.F. Turhollow, R.L. Graham, B.J. Stokes, D.C. Erbach, "Biomass as Feedstock for a Bioenergy and Bioproducts Indus try: The Technical Feasibility of a Billion-Ton Annual Supply," ORNL/TM-2005/66, April 2005.
- S.D. Phillips, "Technoeconom ic analysis of a lignocellulosic biom ass indirect gasification process to m ake ethanol via m ixed alcohol synthesis," *Ind. Eng. Chem. Res.* 46:8887-8897 (2007).

- S. Phillips, A. Aden, J. Jechura, D. Dayton, T. Eggeman, "Thermochemical ethanol via indirect gasification and m ixed alcohol synthesis of lignocellulosic biom ass," Technical Report, NREL/TP-510-41168, April 2007.
- M.A. Rosen, C.A. Bulucea, "Using exergy to understand and improve the efficiency of electrical power technologies," *Entropy* 11:820-835 (2009).
- M.A. Rosen, I. Dincer, M. Kanoglu, "Role of exer gy in increasing efficiency and sustainability and reducing environmental impact," *Energy Policy* 36:128-137 (2008).
- C.R. Shaddix, E.S. Hecht, S. Jimenez, S.M. Lee, "Evaluation of rank effects and gas temperature on coal char burning rates during oxy-fuel combustion," Proceedings of the 34th International Technical Conference on Coal Utilization and Fu el Systems, Clearwater FL, May 31 – June 4, 2009.
- C. Sievers, M.B. Valenzuela-Olarte, T. Marziale tti, I. Musin, P.K. Agrawal, C.W. Jones, "Ionicliquid-phase hydrolysis of pine wood," *Ind. Eng. Chem. Res.* 48:1277-1286 (2009).
- R.P. Swatloski, S.K. Spear, J.D. Holbrey, R.D. Rogers, "Dissolution of cellulose with ionic liquids," *J. Am. Chem. Soc.* 124:4974-4975 (2002).
- K.-Y. Teh, A.E. Lutz, "Therm odynamic analysis of ferm entation and anaerobic growth of baker's yeast for ethanol production," *J. Biotech.* 147:80-87 (2010).
- K.-Y. Teh, S.L. Miller, C.F. Edwards, "Therm odynamic requirem ents for m aximum internal combustion engine cycle efficiency. Part 1: optimal combustion strategy," *Int. J. Engine Res.* 9:449-465 (2008).
- L.M. Vane, "Separation technologies for the recovery and dehydration of alcohols from fermentation broths," *Biofuels Bioprod. Bioref.* 2:553-588 (2008).
- M.J. Wornat, R.H. Hurt, K.A. Davis, N.Y.C. Yang, "Single-particle combustion of two biom ass chars," *Proc. Combust. Instit.* 26:3075-3083 (1996).
- T. Yamada, M.A. Fatigati, M. Zh ang, "Performance of immobilized *Zymomonas mobilis* 31821 (pZB5) on actual hydrolysates produced by Arkenol technology," *Appl. Biochem. Biotech.* 98-100:899-907 (2002).
- H.H. Yoon, Z.W. Wu, Y.Y. Lee, "Am monia-recycled percolation process for pretreatm ent of biomass feedstock," *Appl. Biochem. Biotech.* 51/52:5-19 (1995).

Appendix A

LDRD Project Presentations and Publications

"Thermodynamic analysis of ferm entation and an aerobic growth of baker's yeast," K.-Y.Teh, *ASME International Mechanical Engineering Congress and Exposition (IMECE)*, Lake Buena Vista, FL, Nov. 2009.

"Fractionation and recovery of lignin, carbohydrates, and aliphatic components of biomass in an ionic liquid pretreatm ent process" Dean C. Dibble, Anthe George, Aurelia Cheng, Chenlin Li, Ozgul Persil, Seem a Singh, Blake Sim mons, ACS 2010 Spring Meeting, March 21-25, San Francisco CA.

"Comparative exergy analysis of NREL therm o-chemical biomass-to-ethanol process designs," K.-Y.Teh, A.E. Lutz, C.R. Shaddix, A. Dutta, M. Biddy and A. Aden, *ACS 2010 Spring Meeting*, March 21-25, 2010, San Francisco CA.

"Thermodynamic analysis of ferm entation and an aerobic growth of baker's yeast for ethanol production," K.-Y. Teh, A.E. Lutz, *Journal of Biotechnology* 147 (2010) 80-87.

"The role of therm ochemical processing in op timizing the perf ormance of future cellulosic ethanol biorefineries" C. Shaddix, E. Hecht, K.-W. Teh, A. Lutz, 3rd Annual World Congress of Industrial Biotechnology, July 25-27, 2010, Dalian, China.

"Combustion properties of biomass lignin residues" E.S. Hecht, C.R. Shaddix, 33rd International Combustion Symposium, Aug. 1-6, Beijing, China.

"Ionic liquid pretreatm ent with gel-f ree pr ecipitation and IL recycling utilizing a phaseswitchable solvent system ," Dean C. Dibble, Chenlin Li, Lan Sun, Anthe George, Aurelia Cheng, Özgül Persil Çetinkol, Peter Benke, Seema Singh, Blake A. Simmons, (2010) *in internal review prior to submission to Green Chemistry*.

"An Ionic liquid pretreatm ent process that in cludes IL recycling and a characterization of pretreatment fractions from corn stover" Dean C. Dibble, Chenlin Li, Lan Sun, Peter Benke, Anthe George, Seema Singh. Accepted for oral presentation at the AIChE Annual Meeting Nov. 7-12, Salt Lake City UT.

Several other journal papers are in preparation.

Appendix B

Arkenol Pretreatment Process Steps

- 1. prepare 77% sulfuric acid and cool overnight in refrigerator
- 2. slowly add 1.00 kg chopped biom ass to 3.25 kg cool 77% sulfuric acid (giving 2.5:1 mass ratio of acid/biomass) in bioreactor
- 3. soak biomass in acid for 3 hours
- 4. add 4.10 L D.I. water to reactor
- 5. stir and heat reactor until reaches 99.5° C.
- 6. maintain at 99.5° C for one hour, while continuing to stir
- 7. remove from heat and pass liquids through a cloth filter
- 8. wash separated solids with 2 L D.I. water; repeat
- 9. dry solids in oven at 45° C
- 10. neutralize all collected liquids with mixture of 2.2 kg CaOH and 2.5 L D.I. water
- 11. filter out gypsum created in step 10
- 12. wash gypsum with D.I. water; repeat
- 13. dry gypsum
- 14. add dried solids to 77% sulfuric acid to make 2.5:1 mass ratio
- 15. repeat steps 3 13

