

WASTE TANK VAPOR PROGRAM

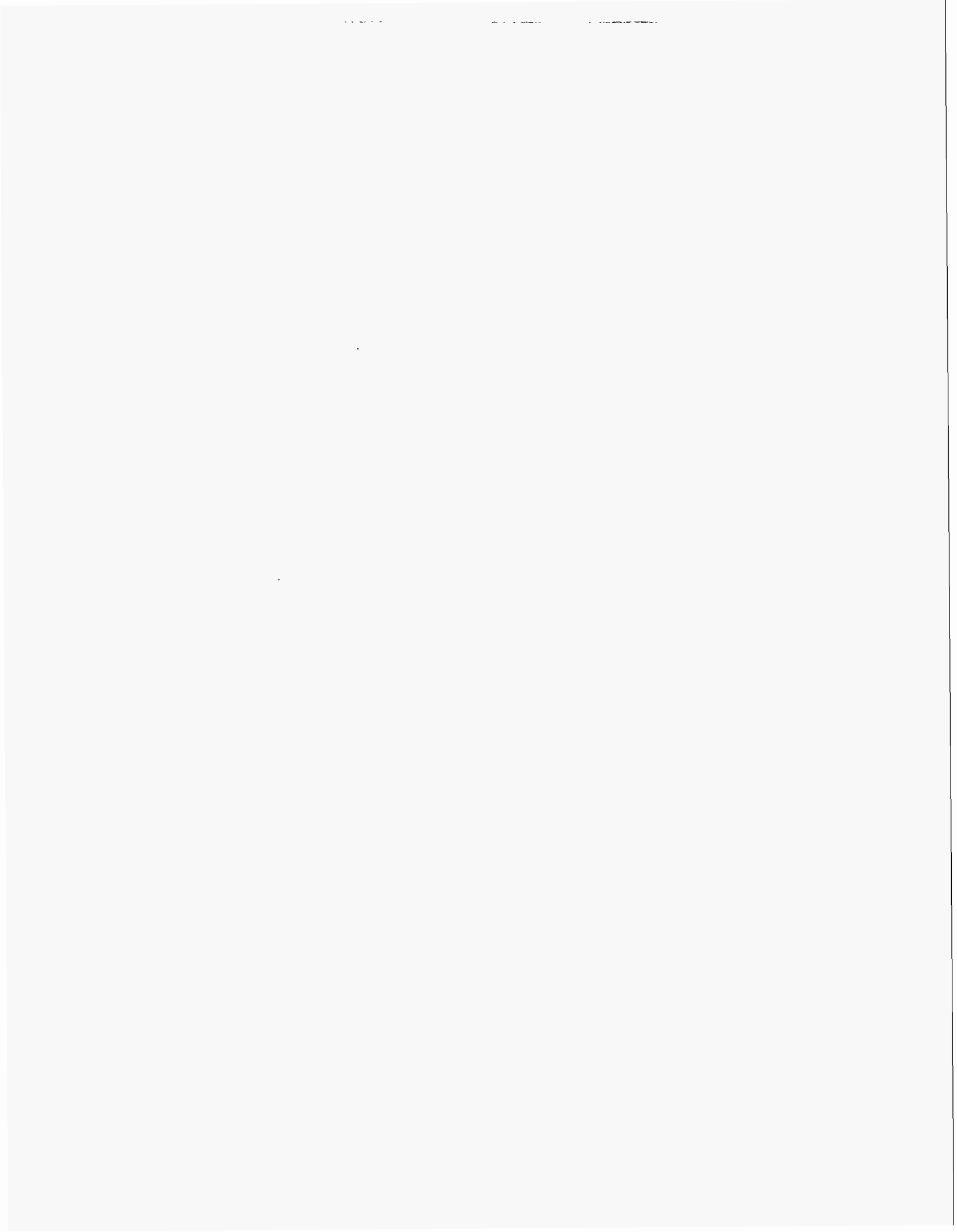
Vapor Space Characterization of Waste Tank 241-TY-103: Results from Samples Collected on 4/11/95

M. W. Ligothke
T. W. Clauss
K. H. Pool
R. B. Lucke
B. D. McVeety
G. S. Klinger
K. B. Olsen
O. P. Bredt
J. S. Fruchter
S. C. Goheen

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Pacific Northwest Laboratory
Richland, Washington 99352



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Summary

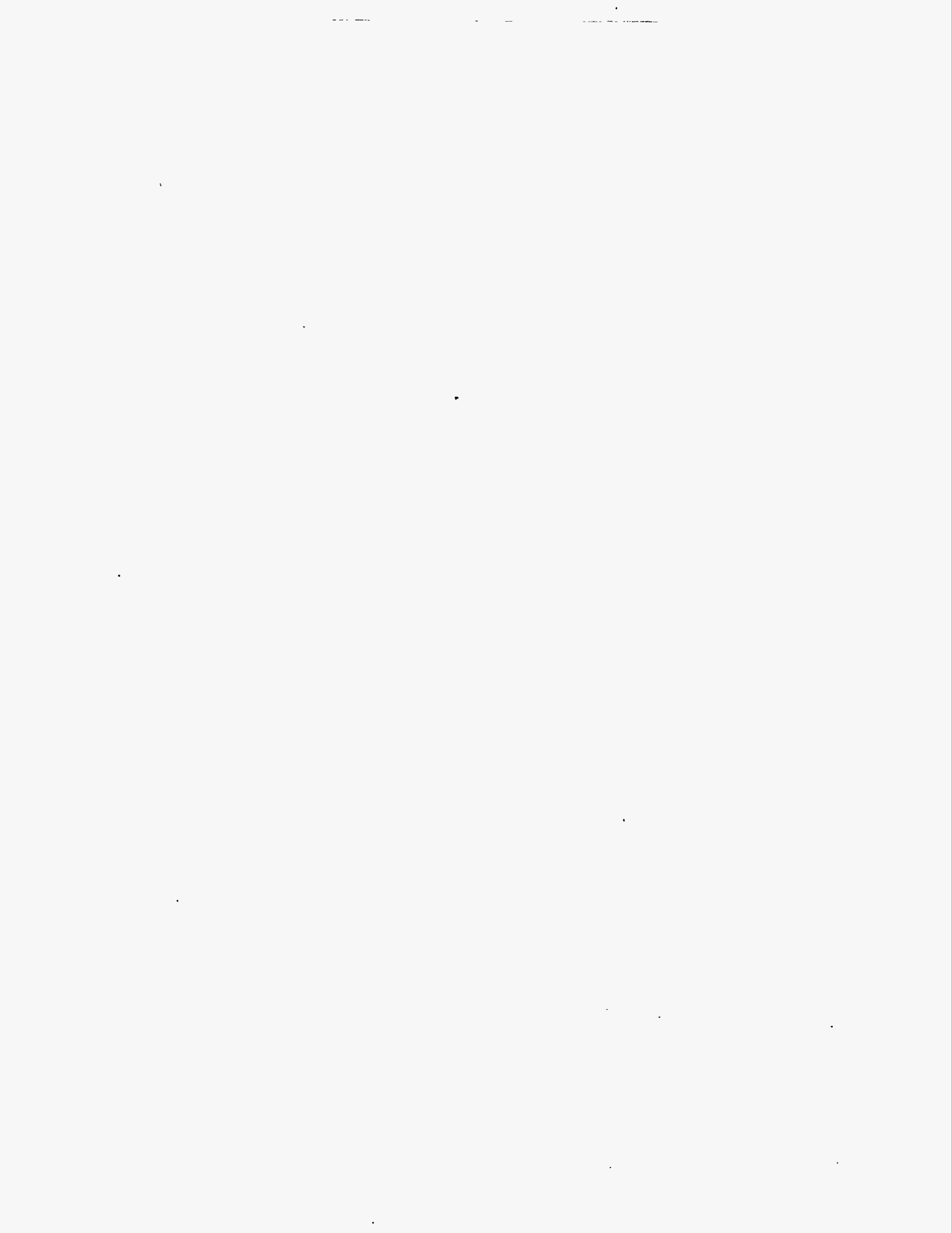
This report describes inorganic and organic analyses results from samples obtained from the headspace of the Hanford waste storage Tank 241-TY-103 (referred to as Tank TY-103). The results described here were obtained to support safety and toxicological evaluations. A summary of the results for inorganic and organic analytes is listed in Table 1. Detailed descriptions of the results appear in the text.

Quantitative results were obtained for the inorganic compounds ammonia (NH₃), nitrogen dioxide (NO₂), nitric oxide (NO), and water (H₂O). Sampling for hydrogen cyanide (HCN) and sulfur oxides (SO_x) was not requested. In addition, quantitative results were obtained for the 39 TO-14 compounds plus an additional 14 analytes. Of these, 16 were observed above the 5-ppbv reporting cutoff. Sixteen tentatively identified compounds (TICs) were observed above the reporting cutoff of (ca.) 10 ppbv and are reported with concentrations that are semiquantitative estimates based on internal-standard response factors. The 10 organic analytes with the highest estimated concentrations are listed in Table 1 and account for approximately 95% of the total organic components in Tank TY-103. Two permanent gases, carbon dioxide (CO₂) and nitrous oxide (N₂O), were also detected.

Table 1. Summary Results of Inorganic and Organic Samples
Collected from the Headspace of Tank TY-103 on 4/11/95

<u>Category</u>	<u>Analyte</u>	<u>Vapor^(a) Concentration</u>	<u>Units</u>
Inorganic	NH ₃	49 ± 1	ppmv
	NO ₂	≤ 0.06	ppmv
	NO	0.10 ± 0.02	ppmv
	H ₂ O	12.7 ± 0.1	mg/L
Organic	Tridecane	13.86	mg/m ³
	Dodecane	7.19	mg/m ³
	Tetradecane	5.20	mg/m ³
	Hexane	1.47	mg/m ³
	Trichlorofluoromethane	0.84	mg/m ³
	1-Butanol	0.55	mg/m ³
	Undecane	0.36	mg/m ³
	Acetone	0.33	mg/m ³
	Propane	0.23	mg/m ³
	Carbon Tetrachloride	0.20	mg/m ³
Permanent Gas	CO ₂	121	ppmv
	N ₂ O	159	ppmv

(a) Vapor concentrations were determined using sample-volume data provided by Westinghouse Hanford Company and are based on averaged data.

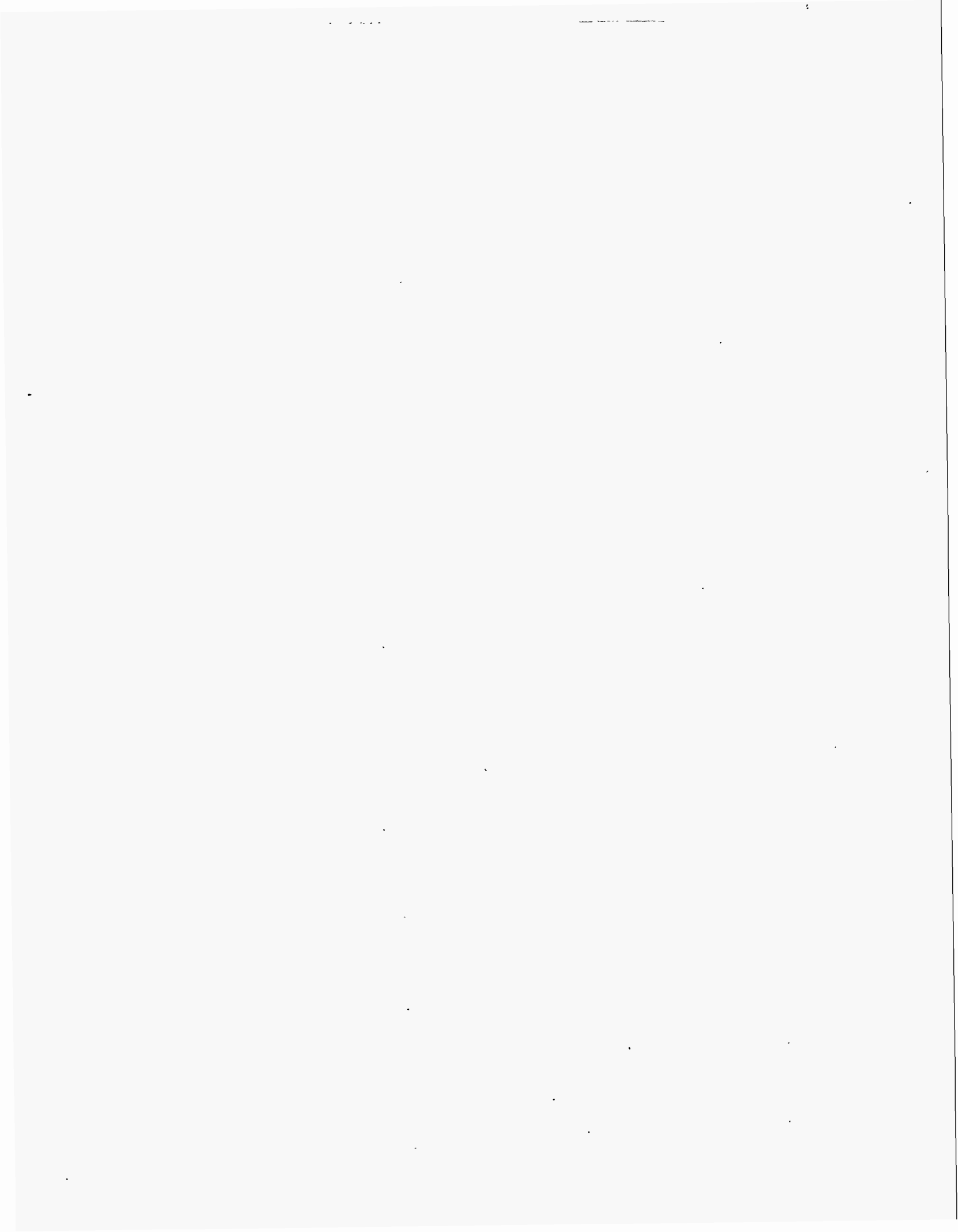


Acknowledgments

The authors gratefully acknowledge the support of other project staff at Pacific Northwest Laboratory who contributed to the successful completion of this sampling and analysis activity. Jeff Edwards served as the PNL single-point-of-contact and coordinated sample handling and communications with Westinghouse Hanford Company. Sally Slate, May-Lin Thomas, and Karen Schielke analyzed inorganic samples, and Gary Dennis prepared the solid-sorbent sample trains. Brenda M. Thornton provided word processing support.

Abbreviations

CAS	Chemical Abstracts Service
COC	chain of custody
C _v	concentration by volume
DIW	deionized water
emf	electromotive force
EPA	U.S. Environmental Protection Agency
GC/MS	gas chromatography/mass spectrometry
GC/TCD	gas chromatography/thermal conductivity detection
HP	Hewlett Packard
IC	ion chromatography
IL	impact level
IS	internal standard
MDL	minimum detection limit
NIST	National Institute for Standards and Technology
OSHA	Occupational Safety and Health Administration
PFA	perfluoroalkoxy
PNL	Pacific Northwest Laboratory
ppbv	part per billion by volume
ppmv	part per million by volume
QA	quality assurance
REL	recommended exposure limit
RPD	relative percent difference
SCIC	suppressed-conductivity ion chromatography
SIE	selective ion electrode
SRM	standard reference material
STP	standard temperature and pressure
TEA	triethanolamine
TIC	tentatively identified compound
VSS	vapor sampling system
WHC	Westinghouse Hanford Company



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1.0 Introduction

This report describes results of the analyses of tank-headspace samples taken from the Hanford waste Tank 241-TY-103 (referred to as Tank TY-103). Pacific Northwest Laboratory (PNL)^(a) contracted with Westinghouse Hanford Company (WHC) to provide sampling devices and to analyze inorganic and organic analytes collected from the tank headspace and ambient air near the tank. The organic analytes for TO-14 compounds were extended to include 14 analytes identified by the Toxicological Review Panel for Tank C-103 and reported by Mahlum et al. (1994). The WHC program management included these analytes for future tank analyses as identified in the fiscal year work plan. This plan is attached to a letter addressed to the WHC Tank Characterization Program Manager^(b). The plan also required PNL to analyze for selected permanent gases. The sample job was designated S4089, and samples were collected by WHC on December 30, 1994, using the vapor sampling system (VSS). Guidance specific to the sample job was provided in the tank characterization plan by Carpenter (1994) and was based in part on the data quality objectives by Osborne et al. (1994).

Sampling devices, including six sorbent trains (for inorganic analyses), and five SUMMA™ canisters (for organic analyses) were supplied to the WHC sampling staff on March 31. Samples were taken (by WHC) from the tank headspace on April 11 and were returned to PNL from the field on April 13. Inorganic (sorbent trap) samples were delivered to PNL on chain-of-custody (COC) 008887 (see Figure 1.1a). The SUMMA™ canisters were delivered on COC 008886 (see Figure 1.1b).

Project work at PNL was governed by an approved QA plan^(c). The samples were inspected upon delivery to the 326/23B laboratory and logged into PNL record book 55408 as described in PNL Technical Procedure PNL-TVP-07^(d). Custody of the sorbent traps was transferred to PNL personnel performing the inorganic analysis and stored at refrigerated ($\leq 10^{\circ}\text{C}$) temperature until the time of analysis. The canister was stored in the 326/23B laboratory at ambient (25°C) temperature until the time of analysis. Access to the 326/23B laboratory is limited to PNL personnel working on the waste-tank safety program. Analyses described in this report were performed at PNL in the 300 area of the Hanford Reservation. Analytical methods that were used are described in the text. In summary, sorbent traps for inorganic analyses containing sample materials were either weighed (for water analysis) or weighed and desorbed with the appropriate aqueous solutions (for ammonia, nitrogen dioxide, and nitric oxide analyses). The aqueous extracts were analyzed by either selective electrode or ion chromatography (IC). Organic analyses were performed using cryogenic preconcentration followed by gas chromatography/mass spectrometry (GC/MS). Permanent gas analysis was performed using gas chromatography/thermal conductivity detection (GC/TCD).

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- (a) Pacific Northwest Laboratory is operated for the U. S. Department of Energy by Battelle Memorial Institute under Contract DE-AC06-76RLO 1830.
 - (b) Letter from Mr. P. J. Melliger (PNL) to Mr. T. J. Kelley (WHC), September 30, 1994, Multi-Year Work Plan for PNL Support of TWRS Characterization for Fiscal Years 1995, 1996, and 1997. TWRS Characterization Project, Pacific Northwest Laboratory, Richland, Washington.
 - (c) PNL Quality Assurance Plan, PNL-MCS-027, Rev. 4. August 1994. TWRS Waste Tank Safety Program, Tank Vapor Project, Richland, Washington.
 - (d) PNL-TVP-07, Rev. 0, October 1994, *Sample Shipping and Receiving Procedure for PNL Waste Tank Samples*, PNL-Technical Procedure, Tank Vapor Project, Richland, Washington.

Custody Form Initiator J. A. Edwards - PNL Telephone (509) 373-0141
Page 85-3009 / FAX 376-0418

Company Contact R. D. Mahon - WHC Telephone (509) 373-2891
Page 85-3152 / FAX 373-3793

Project Designation/Sampling Locations 200 West Tank Farm Collection date 04 - 11 - 95
241-TY-103 Tank Vapor Sample SAF S5-020 Preparation date 03 -30 - 95
(VSS Truck)

Ice Chest No. Field Logbook No. WHC- ~~N-447-10~~ ⁶⁴⁷
~~724~~ ₄₁₃

Bill of Lading/Airbill No. N/A Offsite Property No. N/A

Method of Shipment Government Truck

Shipped to PNL

Possible Sample Hazards/Remarks Unknown at time of sampling

Sample Identification

S5-020 - A23 . 66U -	NH ₃ /NO _x /H ₂ O (Sorbent Trap # 1)	Line # 9
S5-020 - A24 . 67U -	NH ₃ /NO _x /H ₂ O (Sorbent Trap # 2)	Line # 10
S5-020 - A25 . 68U -	NH ₃ /NO _x /H ₂ O (Sorbent Trap # 3)	Line # 8
S5-020 - A26 . 69U -	NH ₃ /NO _x /H ₂ O (Sorbent Trap # 4)	Line # 10
S5-020 - A27 . 70U -	NH ₃ /NO _x /H ₂ O (Sorbent Trap # 5)	Line # 9
S5-020 - A28 . 71U -	NH ₃ /NO _x /H ₂ O (Sorbent Trap # 6)	Line # 10
S5-020 - A29 . 72U -	NH ₃ /NO _x /H ₂ O (Trap Trip Blank # 1)	
S5-020 - A30 . 73U -	NH ₃ /NO _x /H ₂ O (Trap Trip Blank # 2)	
S5-020 - A31 . 74U -	NH ₃ /NO _x /H ₂ O (Trap Trip Blank # 3)	
S5-020 - A32 . 75U -	NH ₃ /NO _x /H ₂ O (Field Blank # 1)	Line # 8
S5-020 - A33 . 76U -	NH ₃ /NO _x /H ₂ O (Field Blank # 2)	Line # 9
S5-020 - A34 . 77U -	NH ₃ /NO _x /H ₂ O (Field Blank # 3)	Line # 10

[] Field Transfer of Custody		[-] Chain of Possession		(Sign and Print Names)	
Relinquished By	Date	Time	Received By	Date	Time
G W Dennis <i>[Signature]</i>	03-31-95	1030	J A Edwards <i>[Signature]</i>	03-31-95	1030
J A Edwards <i>[Signature]</i>	03-31-95	1340	T B Utecht <i>[Signature]</i>	03-31-95	1340
T B Utecht <i>[Signature]</i>	4-13-95	1245	J A Edwards <i>[Signature]</i>	4-13-95	1245
J A Edwards <i>[Signature]</i>	4-14-95	1015	G.W. Dennis <i>[Signature]</i>	4-14-95	1015
G.W. Dennis <i>[Signature]</i>	4-17-95	0845	K.H. Pool <i>[Signature]</i>	4-17-95	0845

Final Sample Disposition

Comments:

PNL (only) Checklist	Pick-up / Delivery	Comments:
<input type="checkbox"/> Media labeled and checked?	<input checked="" type="checkbox"/> Y	
<input type="checkbox"/> Letter of instruction?	<input checked="" type="checkbox"/> Y	
<input type="checkbox"/> Media in good condition?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> COC info/signatures complete?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> Sorbents shipped on ice?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> Rad release stickers on samples?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> Activity report from 222S?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> COC copy for LRB, RIDS filed?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> COC copy for sorbent follow-on?	<input checked="" type="checkbox"/> Y / <input checked="" type="checkbox"/> N/A	
POC <i>[Signature]</i>	POC <i>[Signature]</i>	

(Revised 10/17/94 PNL)

Figure 1.1a Chain-of-Custody for Inorganic Samples for Tank TY-103

Westinghouse Hanford Company	CHAIN OF CUSTODY	WHC 008886
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Custody Form Initiator	J. A. Edwards - PNL	Telephone (509) 373-0141 Page 85-3009 / FAX 376-0418
Company Contact	R. D. Mahon - WHC	Telephone (509) 373-2891 Page 85-3152 / FAX 373-3793
Project Designation/Sampling Locations	200 West Tank Farm 241-TY-103 Tank Vapor Sample SAF S5-020 (VSS Truck)	Collection date 04 - <u>11</u> - 95 Preparation date 03 - 30 - 95
Ice Chest No.		Field Logbook No. WHC- ⁶⁴⁷ N-447-10 _{TP1}
Bill of Lading/Airbill No.	N/A	Offsite Property No. N/A ⁴³⁵⁸
Method of Shipment	Government Truck	
Shipped to	PNL	
Possible Sample Hazards/Remarks	Unknown at time of sampling	

Sample Identification

S5-020 - A01 . 029	Ambient Air SUMMA #1 Upwind of TY-103
S5-020 - A02 . 067	Ambient Air SUMMA #2 Through Port 15
S5-020 - A04 . 080	SUMMA #3 Port 11
S5-020 - A05 . 081	SUMMA #4 Port 13
S5-020 - A06 . 097	SUMMA #5 Port 15

Field Transfer of Custody		Chain of Possession (Sign and Print Names)			
Relinquished By	Date	Time	Received By	Date	Time
J A Edwards / <i>J A Edwards</i>	03-31-95	1340	T B Utecht / <i>T B Utecht</i>	03-31-95	1340
<i>E B Wright</i> / <i>E B Wright</i>	4-13-95	1245	<i>J A Edwards</i> / <i>J A Edwards</i>	4-13-95	1245

Final Sample Disposition

Comments:

PNL (only) Checklist	Pick-up / Delivery	Comments:
<input type="checkbox"/> Media labeled and checked?	<input checked="" type="checkbox"/> Y / <input type="checkbox"/> N	
<input type="checkbox"/> Letter of instruction?	<input checked="" type="checkbox"/> Y / <input type="checkbox"/> N	
<input type="checkbox"/> Media in good condition?	<input checked="" type="checkbox"/> Y / <input type="checkbox"/> N	
<input type="checkbox"/> COC info/signatures complete?	<input checked="" type="checkbox"/> Y / <input type="checkbox"/> N	
<input type="checkbox"/> Rad release stickers on samples?	<input type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> Activity report from 222S?	<input type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> COC copy for LRB, RIDS filed?	<input type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
<input type="checkbox"/> COC copy for sorbent follow-on?	<input type="checkbox"/> Y / <input checked="" type="checkbox"/> N	
	POC <i>(Signature)</i>	POC <i>(Signature)</i>

(Revised 10/17/94 PNL)

Figure 1.1b Chain-of-Custody for Organic Samples for Tank TY-103



2.0 Inorganic

Solid sorbent traps, prepared in multi-trap sampling trains, were supplied to WHC for sampling the tank headspace using the VSS. Blanks, spiked blanks (when requested), and exposed samples were returned to PNL for analysis. Analyses were performed to provide information on the tank-headspace concentration of the following analytes: ammonia (NH₃), nitrogen dioxide (NO₂), nitric oxide (NO), and water (H₂O). Procedures were similar to those developed previously during sample jobs performed with the VSS connected to the headspace of Tank C-103 (Ligotke et al. 1994). During those sample jobs, control samples provided validation that samples were effectively trapping NH₃ and mass. Sample preparation, handling, and disassembly were performed as described in Technical Procedure PNL-TVP-09^(a). Analytical accuracy was estimated based on procedures used. Sample preparation and analyses were performed following PNL quality assurance (QA) impact level (IL) II requirements.

2.1 Standard Sampling Methodology

Standard glass tubes containing sorbent materials to trap vapors of selected analytes of NH₃, NO, NO₂, and H₂O (SKC Inc., Eighty Four, Pennsylvania) were obtained, prepared, and submitted for use by WHC. The sorbent traps were selected based on their use by the Occupational Safety and Health Administration (OSHA) to perform workplace monitoring, and because of available procedures and verification results associated with that particular application. The typical sorbent traps used consisted of a glass tube containing a sorbent material specific to the compound of interest. In general, the tubes contained two sorbent layers, or sections; the first layer was the primary trap, and the second layer provided an indication of breakthrough. In the tubes, sorbent layers are generally held in packed layers separated by glass wool. The sorbent traps, having glass-sealed ends, were received from the vendor.

The type and nominal quantity of sorbent material varied by application. Sorbent traps were selected for the tank sample job and included the following products. The NH₃ sorbent traps contained carbon beads impregnated with sulfuric acid; nominally, 500 mg were contained in the primary and 250 mg in the breakthrough sections. The NH₃ was chemisorbed as ammonium sulfate {(NH₄)₂SO₄}. The NO₂ traps contained a zeolite impregnated with triethanolamine (TEA), with 400 mg in the primary and 200 mg in the breakthrough sections. The NO₂ was absorbed and disproportionated to equi-molar quantities of nitrite ions (NO₂⁻) and nitrate ions (NO₃⁻). Glass tubes containing 800 mg of an oxidant such as chromate were used to convert NO to NO₂. The converted NO was then collected as nitrite and nitrate in an NO₂ trap. The water traps contained 300 mg of silica gel in the primary and 150 mg in the breakthrough sections.

Samples provided by PNL to trap inorganic compounds include all or some of the following: samples, spiked samples, spares, single trap blanks, and spiked blanks. The samples of each were prepared from same-lot batches, with the oxidizer sections of the NO_x sorbent trains having been stored previously in a freezer. After sample preparation, all samples, spiked samples, blanks, and spiked blanks were stored in a freezer, primarily because of handling recommendations for the oxidizer tubes attached to some samples. After receipt of exposed and radiologically cleared samples

(a) Pacific Northwest Laboratory. 10/94. *Sorbent Trap Preparation for sampling and Analysis: Waste Tank Inorganic Vapor Samples*, PNL-TVP-09 (Rev.0), PNL Technical Procedure, Richland, Washington.

from WHC and disassembly of the sorbent trains, samples were provided to the analytical laboratory at ambient temperature, and selected oxidizer sections were returned to a freezer until completion of analyses.

The sorbent traps were prepared in multi-trap sorbent trains configured so sample flow passed in order through the traps, targeting specific analytes, and then through a desiccant trap. The specific order of traps within the various sorbent trains is described in Section 2.4. The ends of the glass-tube traps were broken, and the traps were weighed and then connected to each other using uniform lengths of 3/8-in. perfluoroalkoxy (PFA)-grade Teflon® tubing. The tubing was heated in hot air and forced over the open ends of the traps to form a tight seal. The inlets of the sorbent trains each consist of a short section of tubing having a 3/8-in. stainless steel Swagelok® nut, sealed using a Swagelok® cap. The trailing ends of the sorbent trains (the downstream end of the traps containing silica gel) were each sealed with red-plastic end caps provided by the manufacturer. The sorbent-tube trains remained sealed other than during the actual sampling periods. C-Flex® tubing was provided by WHC to connect the downstream ends of the sorbent trains to the sampling exhaust manifold connections.

2.1.1 Concentration Calculations. The concentrations of target compounds in the tank headspace were determined from sample results, assuming effective sample transport to the sorbent traps. Concentration, in parts per million by volume (ppmv), was determined by dividing the mass of the compound, in μmol , by the volume of the dried tank air sampled in mol. The micromolar sample mass was determined by dividing the compound mass, in μg , by the molecular weight of the compound, in g/mol. The molar sample volume was determined, excluding water vapor, by dividing the standard sample volume (at 0°C and 760 torr), in L, by 22.4 L/mol. For example, the concentration (C_v) of a 3.00-L sample containing 75.0 μg of NH_3 equals

$$C_v = \frac{75.0 \mu\text{g}}{17 \text{ g/mol}} \left(\frac{3.00 \text{ L}}{22.4 \text{ L/mol}} \right)^{-1} = 32.9 \text{ ppmv} \quad (2.1)$$

This calculational method produces concentration results that are slightly conservative (greater than actual) because the volume of water vapor in the sample stream is neglected. The volume of water vapor is not included in the measured sampled volume because of its removal in desiccant traps upstream of the mass flowmeter. However, the bias is generally expected to be small. For a tank-headspace temperature of 35°C, the magnitude of the bias would be about 1 to 6%, assuming tank-headspace relative humidities of 20 to 100%, respectively. The concentration of mass (determined gravimetrically) was also per dry-gas volume at standard conditions.

2.2 Analytical Procedures

The compounds of interest were trapped using solid sorbents and chemisorption (adsorption of water vapor). Analytical results were based on extraction and analysis of selected ions. Analytical procedures used are specified in the text. All were compiled in PNL-MA-599.

2.2.1 Ammonia Analysis. The sorbent material from the NH_3 -selective sorbent traps was placed into labeled 20-mL glass scintillation vials. Vials containing front-, or primary-, section sorbent material were treated with 10.0 mL of deionized water (DIW), and vials containing back-up-section

sorbent material were treated with 5.0 mL of DIW. After extraction, the NH_3 sorbent traps were analyzed using the selective ion electrode (SIE) procedure PNL-ALO-226^(a). Briefly, this method includes 1) preparing a 1000- $\mu\text{g}/\text{mL}$ (ppm) NH_3 stock standard solution from dried reagent-grade NH_4Cl and DIW on the day analyses are performed; 2) preparing 0.1-, 0.5-, 1.0-, 10-, and 100-ppm NH_3 working calibration standards by serial dilution of the freshly made stock standard; 3) generating an initial calibration curve from the measured electromotive force (emf) signal versus NH_3 concentration data obtained for the set of working standards; 4) performing a calibration-verification check, using one of the midrange standards, after analyzing every four or five samples; 5) continuing this sequence until all samples of the batch have been measured, including duplicates and spiked samples; and 6) remeasuring the complete set of calibration standards (at the end of the session). Emf signal measurements obtained for samples are compared to those for standards, either graphically or algebraically (using linear regression) to determine NH_3 concentration in the samples.

2.2.2 Nitrite Analysis. The sorbent traps for NO_2 and NO were desorbed in an aqueous TEA and n-butanol solution and analyzed by suppressed-conductivity ion chromatography (SCIC) for nitrite according to PNL-ALO-212, Rev. 1^(b) modified to obviate interferences by concentrations of non-target analytes. Specifically, the modifications used were 1) eluent 1.44 mM Na_2CO_3 + 1.8 mM NaHCO_3 at 2.0 mL/min, 2) one guard column (AG4A) and two separator columns (AS4A) in series instead of just one separator column, and 3) all standards, samples, and blanks injected into the IC sample loop through 0.45- μm syringe filters.

For the analysis, the sorbent materials were placed into labeled 20-mL glass scintillation vials. To each vial, 3.0 mL of desorbing solution (15 g TEA + 1 mL n-butanol in 1.0 L DIW) was added. Primary sorbent-tube sample materials and back-up (breakthrough) sorbent-trap materials were analyzed separately using identical procedures. Each analytical session was conducted as follows. Working nitrite standards (0, 0.1, 0.25, and 0.5 ppm) were prepared by diluting a stock nitrite standard with desorbing solution. An initial calibration curve was prepared from the instrument response (chromatographic peak height) versus nitrite standard concentration data for the set of working standards. A calibration verification check using one of the midrange standards was performed after the analysis of every six samples. If the instrument response indicated that sample nitrite concentration was outside the calibration range (> 0.5 ppm nitrite), the sample was diluted with desorbing solution and reanalyzed. After all samples of a batch were analyzed, the complete set of calibration standards was remeasured to verify consistent instrument response, and the analytical session was terminated.

Instrument responses (peak height) observed for samples were compared to those for standards to determine the nitrite concentration of the samples. Because NO_2 and NO converted to NO_2 were collected on the sorbent as equal quantities of nitrite and nitrate, and the analysis was specific for nitrite, the molar masses of NO_2 and NO were determined by doubling the analytically determined molar mass of nitrite.

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- (a) Procedure entitled "Ammonia (Nitrogen) in Aqueous Samples," PNL-ALO-226, in the *Analytical Chemistry Laboratory (ACL) Procedure Compendium*, Vol. 3: Inorganic Instrumental Methods. Pacific Northwest Laboratory, Richland, Washington.
- (b) Procedure entitled "Determination of Inorganic Anions by Ion Chromatography," PNL-ALO-212, in the *Analytical Chemistry Laboratory (ACL) Procedure Compendium*, Vol. 3: Inorganic Instrumental Methods. Pacific Northwest Laboratory, Richland, Washington.

2.2.3 Mass (Water) Analysis. Sorbent traps used to make each sample train were weighed using a semi-micro mass balance, after labeling and breaking the glass tube ends, without plastic end caps. After receipt of exposed samples, the sorbent traps were again weighed to determine the change in mass. Records of the measurements were documented on sample-preparation data sheets. The mass concentration, generally roughly equal to the concentration of water, was determined by dividing the combined change in mass from all traps in a sorbent train by the actual volume of gas sampled. Blanks and spiked blanks were included to provide information on uncertainty.

2.3 Quality Assurance/Quality Control

Analytical work was performed according to quality levels identified in the project QA plan and several PNL documents. The samples were analyzed following PNL IL II. The PNL documents include PNL-MA-70 (Part 2), PNL-ALO-212, PNL-ALO-226, and MCS-046. A summary of the analysis procedures and limits for the target inorganic compounds is provided in Table 2.1. From the table, it can be seen that the minimum detection limit (MDL) required to resolve the analyte at one-tenth of the recommended exposure limit (REL) for each of the target analytes is achieved using current procedures and with a vapor-sample volume of 3 L and a desorption-solution volume of 3 mL (10 mL for NH₃).

Table 2.1 Analysis Procedures and Typical Detection Limits of Target Inorganic Analytes

<u>Analyte</u>	<u>Formula</u>	<u>Procedure</u>	REL ^(a) (ppmv)	0.1 x REL ^(a) (ppmv)	MDL ^(b) (ppmv)
Ammonia	NH ₃	PNL-ALO-226	25	2.5	0.5
Nitrogen Dioxide	NO ₂	PNL-ALO-212	1	0.1	0.02
Nitric oxide	NO	PNL-ALO-212	25	2.5	0.02
Mass (water) ^(c)	n/a	n/a	n/a	n/a	n/a

(a) Target analytical limits are equal to one-tenth of the REL.

(b) MDL is defined as the vapor concentration that can be detected with an uncertainty equal to about the magnitude of the measurement. The uncertainty is expected to reduce to about one-quarter of the magnitude of the measurement at a concentration of four times the MDL. The MDLs were based on the assumption that 3 L of vapor are sampled; if greater volumes of vapor are sampled, correspondingly smaller MDLs can be achieved. The MDLs were also based on desorbing-solution volumes of 10 mL for NH₃ and 3 mL for the other analytes.

(c) The vapor-mass concentration, thought to be largely water vapor, is determined gravimetrically.

The accuracy of concentration measurements depends on errors associated with both sampling and analysis (see Section 2.4). Sampling information, including sample volumes, was provided by WHC; sample volume uncertainty was not provided. The accuracy of analytical results depends on the method used. For NH₃ analyses, the accuracy of laboratory measurements by SIE was estimated to be ± 5% relative, independent of concentration at 1 µg/mL or greater levels. The uncertainty includes preparation of standards, purity of the ammonium salt used to prepare standards, potential operator bias, ambient temperature variations, etc. Unfortunately, no known National Institute for Standards and Technology (NIST)-traceable standard reference material (SRM) is available against which to compare working standards. Similarly, no known NIST SRM is available for nitrite analysis (for NO₂ and NO). Based on experience in comparing nitrite working standards prepared from

several different sources and factors mentioned for NH₃ above, the estimated maximum bias for samples derived from sampling for NO₂ is $\pm 10\%$, and for samples derived from sampling for NO, it is $\pm 5\%$ relative. The accuracy of measurements of sample mass is ± 0.05 mg, or much less than 1% of the mass changes of most samples. The analytical accuracy of measurements of the change in mass of sorbent trains is typically about ± 2 mg per 5-trap sorbent train.

2.4 Inorganic Sample Results

Samples were obtained by WHC from the headspace of Tank TY-103 on 4/11/95 using the VSS. The sample job designation number was S5020. Samples were prepared, submitted to WHC, returned to PNL, and then analyzed to provide information on the concentrations of NH₃, NO₂, NO, and mass (largely H₂O). Sampling and analysis for hydrogen cyanide (HCN) and sulfur oxides (SO_x) was not requested. The inorganic samples were received from WHC on 4/13/95; the sample-volume information was received on 4/17/95.

A list of samples, sampling information, sample volumes, and gravimetric results is shown in Table 2.2. The types of sample trains used and the order of sorbent traps within each train are also shown in the table. For example, the sorbent train NH₃/NO_x/H₂O contained an NH₃ trap at the inlet end, an NO_x series in the middle (Section 2.4.2), and a desiccant trap at the outlet end. Analytical mass and concentration results are shown in Table 2.3. Sample volumes were provided by WHC; sample-volume uncertainty was not provided. Tank-headspace concentration results (Table 2.3) are based on this information, and the listed uncertainties equal plus-or-minus one standard deviation of the individual results from each set of samples. Where analytical results from samples were nearly indistinguishable from those of blanks, indicating very low vapor concentrations of the analyte, the concentration results (Table 2.3) are listed as "less than or equal to" a probable maximum value determined by subtracting the average of the blanks less one standard deviation from the average of the samples plus one standard deviation. Results of control samples, such as spiked blanks, are discussed in this section. Spiked blanks, when used, were transported to the field but not opened. Spiked samples, when used, were opened in the field and used to collect tank vapors. Sample results were not corrected for the percentage recoveries of spiked blanks.

2.4.1 Ammonia Results. The concentration of NH₃ was 49 ± 1 ppmv, based on all six samples. The blank-corrected NH₃ quantities in the sorbent traps ranged from 6.3 to 6.8 μmol in front sections and were negligible in back sections. Blank corrections were applied because the quantities of NH₃ in three of three field blanks (0.08 μmol in front and 0.05 μmol in back sections) were $\sim 1\%$ of the quantities in samples. Three of three trip blanks were also analyzed and found to contain 0.08 μmol in front and 0.04 μmol in back sections. Although spiked blanks were not tested, the percentage recoveries of three sets of blanks spiked with 12.2, 22.3, and 46.4 μmol of NH₃ were $101 \pm 4\%$, $109 \pm 2\%$, and $104 \pm 1\%$, respectively, during related sample jobs (Clauss et al. 1994; Ligothke et al. 1994). The analysis of one sample was duplicated and yielded a repeatability of $\pm 2\%$. One sample leachate was spiked after initial analysis with roughly the quantity of NH₃ in the sample and yielded a percentage recovery of 115%. The reason for the poor recovery was not determined; however, two control spikes were also prepared and yielded recoveries of 104% and 106%. A 5-point calibration was performed over an NH₃ range of 0.1 to 1000 $\mu\text{g/mL}$.

Table 2.2 List of PNL Inorganic Samples, Controls, and Gravimetric Results Obtained from a Heated Tube Inserted into the Headspace of Tank TY-103 on 4/11/95

Sample Number	Sorbent Train Type	Sample Port and Volume Information ^(a)				
		Sample Port	Flow Rate (mL/min)	Duration (min)	Volume (L)	Mass Gain (g)
<u>Samples:</u>						
S5020-A23-66U	NH ₃ /NO _x /H ₂ O Train	9	200.4	15.0	3.01	0.0406
S5020-A24-67U	NH ₃ /NO _x /H ₂ O Train	10	200.4	15.0	3.01	0.0406
S5020-A25-68U	NH ₃ /NO _x /H ₂ O Train	8	200.4	15.0	3.01	0.0414
S5020-A26-69U	NH ₃ /NO _x /H ₂ O Train	10	200.4	15.0	3.01	0.0414
S5020-A27-70U	NH ₃ /NO _x /H ₂ O Train	9	200.4	15.0	3.01	0.0409
S5020-A28-71U	NH ₃ /NO _x /H ₂ O Train	10	200.4	15.0	3.01	0.0405
<u>Controls:</u>						
S5020-A29-72U	NH ₃ /NO _x /H ₂ O Trip Blank	n/a ^(b)	n/a	n/a	n/a	0.0020
S5020-A30-73U	NH ₃ /NO _x /H ₂ O Trip Blank	n/a	n/a	n/a	n/a	0.0026
S5020-A31-74U	NH ₃ /NO _x /H ₂ O Trip Blank	n/a	n/a	n/a	n/a	0.0027
S5020-A32-75U	NH ₃ /NO _x /H ₂ O Field Blank	n/a	n/a	n/a	n/a	0.0028
S5020-A33-76U	NH ₃ /NO _x /H ₂ O Field Blank	n/a	n/a	n/a	n/a	0.0028
S5020-A34-77U	NH ₃ /NO _x /H ₂ O Field Blank	n/a	n/a	n/a	n/a	0.0026

(a) Sampling information and dry-gas sample volumes, corrected to 0°C and 760 torr, were provided by WHC. Uncertainty values were not provided with sample-volume results.

(b) n/a = not applicable.

2.4.2 Nitrogen Oxides Results. Measurements of NO₂ and NO were made using six 5-segment NH₃/NO_x/H₂O sorbent-trap trains (the NO_x trains consisted of NO₂ trap, oxidizer, and NO₂ trap). Related sample jobs, performed using the VSS in Tanks BY-104, -105, and -106 both with and without NO_x trains protected by a leading NH₃ trap (e.g., Clauss et al. 1994), indicated that the presence of the upstream NH₃ traps resulted in NO concentrations that were about 1.3- to 1.6-fold less than those from unprotected NO₂ traps. The NO₂ concentrations were also potentially less following an NH₃ trap.

The concentrations of NO₂ and NO were ≤ 0.06 and 0.10 ± 0.02 ppmv, respectively, based on all six samples. Blank-corrected NO₂⁻ quantities in the sorbent traps averaged ≤ 0.0039 μmol (NO₂ samples) and 0.0070 μmol (NO samples). The level of NO₂⁻ in field blanks was 0.0092 ± 0.0007 μmol in front (three of six blanks analyzed) and 0.0046 ± 0.0004 μmol in back sections (two of six blanks analyzed); the field blank results were used to correct sample data. Nitrite trip blank levels were 0.0089 ± 0.0003 μmol in front (four of six blanks analyzed) and 0.0052 ± 0.0003 μmol in back sections (two of six blanks analyzed). Although spiked blanks were not tested, blanks spiked with 0.0064, 0.047, 0.11, and 0.74 μmol of NO₂⁻ during related sample jobs yielded percentage recoveries of $153 \pm 14\%$, $103 \pm 4\%$, $106 \pm 8\%$, and $111 \pm 7\%$, respectively (Clauss et al. 1994; Ligothke et al. 1994). The analyses of two samples were duplicated and yielded repeatabilities of

Table 2.3 Inorganic Vapor Sample Results Obtained from a Heated Tube Inserted into the Headspace of Tank TY-103 on 4/11/95

Sample	Analytical Results (μmol)			Sample Volume (L)	Vapor ^(a) Concentration (ppmv)
	Front Section	Back Section	Total ^(b) Blank-Corrected		
<u>NH₃ Samples:</u>			<u>6.57^(c)</u>	<u>3.01^(c)</u>	<u>49 ± 1^(c)</u>
S5020-A23-66U	6.63	0.03	6.55	3.01	49
S5020-A24-67U	6.52	NA ^(d)	6.44	3.01	48
S5020-A25-68U	6.34	0.04	6.26	3.01	47
S5020-A26-69U	6.87	NA	6.79	3.01	51
S5020-A27-70U	6.76	NA	6.68	3.01	50
S5020-A28-71U	6.79	NA	6.71	3.01	50
<u>NO₂ Samples:</u>			<u>≤0.0039</u>	<u>3.01</u>	<u>≤ 0.06</u>
S5020-A23-66U	0.0133	0.0044	n/a ^(d)	3.01	n/a
S5020-A24-67U	0.0120	0.0056	n/a	3.01	n/a
S5020-A25-68U	0.0098	NA	n/a	3.01	n/a
S5020-A26-69U	0.0106	NA	n/a	3.01	n/a
S5020-A27-70U	0.0099	NA	n/a	3.01	n/a
S5020-A28-71U	0.0101	NA	n/a	3.01	n/a
<u>NO Samples:</u>			<u>0.0070</u>	<u>3.01</u>	<u>0.10 ± 0.02</u>
S5020-A23-66U	0.0147	NA	0.0055	3.01	0.08
S5020-A24-67U	0.0172	NA	0.0080	3.01	0.12
S5020-A25-68U	0.0157	NA	0.0065	3.01	0.10
S5020-A26-69U	0.0172	NA	0.0080	3.01	0.12
S5020-A27-70U	0.0157	0.0044	0.0065	3.01	0.10
S5020-A28-71U	0.0164	0.0051	0.0072	3.01	0.11
<u>Gravimetric Samples:</u>			<u>38.2 mg</u>	<u>3.01</u>	<u>12.7 ± 0.1 mg/L</u>
S5020-A23-66U	n/a	n/a	37.9	3.01	12.6
S5020-A24-67U	n/a	n/a	37.9	3.01	12.6
S5020-A25-68U	n/a	n/a	38.7	3.01	12.9
S5020-A26-69U	n/a	n/a	38.7	3.01	12.9
S5020-A27-70U	n/a	n/a	38.2	3.01	12.7
S5020-A28-71U	n/a	n/a	37.8	3.01	12.6

- (a) Blank-corrected vapor concentrations were calculated using WHC-reported dry-air sample volumes (corrected to 0°C and 760 torr). In the calculation for concentration, the nitrite values (listed) were doubled to account for unanalyzed nitrate. Sample results were not corrected for percentage recovery of spiked samples or spiked blanks.
- (b) Total blank-corrected analyte masses (nitrite for NO₂ and NO) were determined, when significant, by subtracting the quantity of analyte found in blanks from that found in samples. The level of analytes found in blanks is described in the subsections of Section 2.4.
- (c) Underlined values represent the average of the set samples. Concentration uncertainty equals ± 1 standard deviation (absolute) for each set of samples. The use of "≤" is defined in Section 2.4.
- (d) NA = not analyzed; n/a = not applicable. Only selected back sorbent sections were analyzed. Past results have shown back sections of NH₃ samples to contain insignificant quantities of the analyte.

$\pm 0\%$ and $\pm 4\%$. One sample leachate was spiked with 0.25 ppm NO_2 and yielded a percentage recovery of 96%. A 4-point calibration was performed over a concentration range of 0 to 0.5 $\mu\text{g NO}_2^-$ per mL in the desorbing matrix.

2.4.3 Gravimetric Results. The mass concentration of material collected in the 5-trap sorbent trains, believed to be primarily water vapor, was 12.7 ± 0.1 mg/L. The result was based on an average mass gain of 38.2 mg from all six $\text{NH}_3/\text{NO}_2/\text{H}_2\text{O}$ sample trains. The blank correction applied to the results was -2.7 mg per sample train, based on a mass gain of 2.7 ± 0.1 mg per three field blank 5-trap sorbent trains. Three trip blanks gained 2.4 ± 0.4 mg per train; the use of trip blanks to correct sample data would have resulted in a vapor mass concentration 1% greater than the actual result. Although no spiked blanks were tested, the percentage recovery of mass from three blank H_2O traps spiked with 51 mg of water was $103 \pm 2\%$ during a related sample job (Clauss et al. 1994).

3.0 Organic

3.1 SUMMA™ Canister Preparation

Before sending SUMMA™ canisters out to the field for sampling, the canisters are cleaned and verified contaminant free according to PNL Technical Procedure PNL-TVP-02^(a). The cleaning procedure uses an EnTech 3000 cleaning system that controls 1) filling the canisters with purified humid air and 2) evacuating, for several cycles with applied heat, before allowing the canister to evacuate overnight. The canister is filled a final time with purified humid air for analysis by PNL Technical Procedure PNL-TVP-01^(b), which is a modification of U.S. Environmental Protection Agency (EPA) compendium Method TO-14. If the canister is verified as clean, free of TO-14 and unknown contaminants to a level of 5 parts per billion by volume (ppbv), the canister is evacuated to 5 mtorr, tagged, and stored for use in the field. Before sending the canisters out to the field for sampling, the canister vacuum is measured to determine if any leakage has occurred. If the vacuum has remained constant during storage, the canisters are prehumidified with 100 μ L of distilled water and labeled with a field-sampling identification. Canisters stored more than 30 but less than 60 days are re-evacuated and rehumidified before use. If stored more than 60 days, the canisters are recleaned and validated before use.

3.2 Sample Analysis Methods

The SUMMA™ canister sample was analyzed according to PNL Technical Procedure PNL-TVP-03^(c), which is a modified version of EPA compendium Method TO-14. The method uses an EnTech cryoconcentration system interfaced with either a 5971 or a 5972 Hewlett Packard (HP) benchtop GC/MS. The EnTech concentrator is used to pull a metered volume of sample air from the SUMMA™ canister, cryogenically concentrate the air volume, then transfer the volume to the GC/MS for analysis. A 100-mL volume of sample is measured and analyzed from the tank headspace. The organic components in the sampled air are separated on an analytical column, J&W Scientific DB-1 phase, 60-m by 0.32-mm internal diameter with 3- μ m film thickness. The GC oven is programmed to run a temperature gradient beginning at 40°C, holding for 5 min, and ramping at 4°C per min to a final temperature of 260°C, with a 5-min hold. Twenty-four hours before the analysis, the SUMMA™ canister samples were pressurized with purified air (Aadco Instruments, Inc., 1920 Sherwood St., Clearwater, Florida 34625). The starting pressure was first measured using a calibrated diaphragm gauge (Cole Parmer), then pressurized to a level exactly twice the original pressure. For example, if the canister had a starting pressure of 740 torr, it was pressurized to 1480 torr. This dilution was an effort to improve the precision of the analysis. The sample dilution was taken into account when calculating the analysis results.

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- (a) Pacific Northwest Laboratory. 8/94. *Cleaning SUMMA™ Canisters and the Validation of the Cleaning Process*, PNL-TVP-02 (Rev. 0), PNL Technical Procedure, Richland, Washington.
- (b) Pacific Northwest Laboratory. 8/94. *Determination of TO-14 Volatile Organic Compounds in Ambient Air Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometric Analysis*, PNL-TVP-01 (Rev. 0). PNL Technical Procedure, Richland, Washington.
- (c) Pacific Northwest Laboratory. 8/94. *Determination of TO-14 Volatile Organic Compounds in Hanford Tank Headspace Samples Using SUMMA™ Passivated Canister Sampling and Gas Chromatographic-Mass Spectrometric Analysis*, PNL-TVP-03 (Rev. 0), PNL Technical Procedure, Richland, Washington.

The instrument calibration mixture for the TO-14 analysis consists of the standard 39 organic analytes with an additional 14 tank-related compounds. Together, these 53 compounds that are directly quantified in this analysis make up the target analyte list (these 53 compounds will be referred to as target analytes). The calibration mixture was prepared by blending a commercially prepared 39-compound TO-14 calibration mixture with a 14-compound mixture created using a Kin-Tek® permeation-tube standard generation system. The operation of the permeation tube system follows the method detailed in PNL Technical Procedure PNL-TVP-06^(a). The standard calibration mix was analyzed using four aliquot sizes ranging from 30 mL to 200 mL, and a response factor for each compound was calculated. The GC/MS response for these compounds has been previously determined to be linearly related to concentration. Currently, 1-butanol is not being measured in the samples as a calibrated analyte. It is being quantified as a tentatively identified compound (TIC). Once the appropriate permeation tube has been obtained, 1-butanol will be measured as a calibrated compound. Performance-based detection limits for the target analytes will be developed as a pool of calibration data becomes available. Currently, the nominal detection limit of 5 ppbv is met.

The SUMMA™ canister samples were analyzed for permanent gases according to PNL Technical Procedure PNL-TVP-05^(b) with the exceptions listed in the following text and in the Quality Assurance/Quality Control section of this report. This method was developed in-house for the analysis of permanent gases defined as hydrogen (H₂), carbon dioxide (CO₂), carbon monoxide (CO), methane (CH₄), and nitrous oxide (N₂O) by GC/TCD and is not validated in any other laboratory. No previous work up of the sample canister is necessary before permanent gas analysis. Aliquots of sampled air are manually drawn from each canister into a 5-mL gas-tight syringe and directly injected into a GC/TCD fitted with a 1.0-mL injection loop. An aliquot of 5 mL is used so that the injection loop is flushed and filled with sample air, ensuring that no dilution of the sample takes place within the injection loop. One set of GC conditions is used to analyze for CO, CO₂, N₂O, and CH₄ using Helium (He) as the carrier gas. A second GC analysis is performed for H₂ (using nitrogen as the carrier gas) to enhance the signal sensitivity and lower the detection limit for this analyte. A total of 30 to 60 mL, depending upon the number of repeat analyses performed, may be drawn from each 6-L canister.

3.3 Quality Assurance/Quality Control

Before the tank sample was analyzed, a diagnostic check was performed on the GC/MS instrument by running an instrument "high-sensitivity tune," as described in PNL-TVP-03. Upon satisfactory completion of the instrument diagnostic check, a blank volume of purified nitrogen was analyzed to check the cleanliness of the system. The instrument was then calibrated using a standard gas mixture containing 39 volatile organic compounds listed in EPA compendium Method TO-14 and an additional 14 tank-related compounds. A gas mixture containing bromochloromethane, 1,4-difluorobenzene, and chlorobenzene-d₃ was used as an internal standard (IS) for all blank, calibration standard, and sample analyses. Analyte responses from sample components, ISs, and standards were obtained from the extracted ion plot from their selected mass ion. The calibration was generated by calculating the relative response ratios of the IS to calibration standard responses and

(a) Pacific Northwest Laboratory. 8/94. *Preparation of TO-14 Volatile Organic Compounds Gas Standards*, PNL-TVP-06 (Rev. 0). PNL Technical Procedure, Richland, Washington.

(b) Pacific Northwest Laboratory. 8/94. *Analysis Method for the Determination of Permanent Gases in Hanford Waste Tank Vapor Samples Collected in SUMMA™ Passivated Stainless Steel Canisters*, PNL-TVP-05 (Rev. 0). PNL Technical Procedure, Richland, Washington.

plotting the ratios against the ratio of the calibration-standard concentration (in ppbv) to the IS concentration. Once it is determined that the relative response is linear with increasing concentration, an average response factor is calculated for each target analyte and used to determine the concentration of target compounds in each sample.

Standards for the permanent gas analysis were blended from commercially prepared and certified standards for each of the analytes reported in Table 3.5. The instrument was calibrated over three data points for CO, CO₂, N₂O, and CH₄ using He as a carrier gas, and then the samples were analyzed. The carrier was changed to N₂, the calibration was performed for H₂ only, and the samples were reanalyzed. A least-squares linear-regression routine was applied to the data set to generate the best fit for each compound. The quantitation for each analyte was performed by direct comparison of sample analyte peaks to the plot generated for the compound. The lowest calibration standard for each analyte is reported as the method detection limit. An MDL for the instrument has not been determined. An N₂ reagent blank, ambient air sample collected ~10 m upwind of TY-103, and the ambient air collected through the VSS were used as method blanks and used to determine the potential for analyte interferences in the samples. Continuing calibration standards for this sample set fell within ± 25% of the expected concentrations for the analytes reported.

3.3.1 Quantitation Results of Target Analytes. The quantitative-analysis results for the target analytes were calculated directly from the calibration curve generated using the IS method described above and in PNL-TVP-03. The conversion from ppbv to mg/m³ assumes standard temperature and pressure (STP) conditions of 760 torr and 273K and was calculated directly from the following equation:

$$\text{mg/m}^3 = \frac{(\text{ppbv}/1000) \times \text{g mol wt of compound}}{22.4 \text{ L/mol}} \quad (3.1)$$

3.3.2 Identification and Quantitation of Tentatively Identified Compounds. The TICs are determined by mass-spectral interpretation and comparison of the spectra with the EPA/NIST/WILEY Library, which is a part of the HP 5971/5972 instrument operating system. Chromatographic peaks with an area count greater than, or equal to, one tenth of the total area count of the nearest eluting IS are tentatively identified and quantitatively estimated. The quality of the mass-spectral searches was then reviewed by the principal investigators before the identification was assigned to each chromatographic peak.

The concentration of each TIC was estimated using a relative response factor calculated using the total peak area for the nearest eluting IS. The IS peak area was used to calculate a response factor using the IS concentration in mg/m³:

$$\text{Response Factor} = \frac{\text{IS conc. (mg/m}^3\text{)}}{\text{IS peak area}} \quad (3.2)$$

The calculated response factor was then multiplied by the TIC peak area to give an estimated concentration for that compound.

The ppbv concentrations are calculated from mg/m^3 and the molecular weight of the analyte.

$$\text{TIC in ppbv} = \frac{\text{TIC (mg/m}^3) \times 22.4 \text{ L/mol} \times 1000}{\text{TIC g mol wt}} \quad (3.3)$$

The IS level added to all blank, standard, and sample injections was 104 ppbv for bromochloromethane, 101 ppbv for 1,4-difluorobenzene, and 98.5 ppbv for chlorobenzene- d_5 . The IS concentrations were converted from ppbv to mg/m^3 at STP using a molecular weight of 129.39 (g/mol) for bromochloromethane, 114.09 for 1,4-difluorobenzene, and 117.6 for chlorobenzene- d_5 . All sample concentrations were multiplied by a factor of two to account for the dilution step described in Section 3.2.

3.4 Analysis Results

The results from the GC/MS analysis of the tank-headspace samples are presented in Tables 3.1 and 3.2. The results of replicate analyses on a single SUMMA™ canister are presented in Tables 3.3 and 3.4. The result of GC/MS analysis of the ambient-air sample collected upwind of Tank TY-103 and through the VSS near Tank TY-103 are presented in Tables 3.5 and 3.6. The GC/TCD analysis results for permanent gases for both the ambient samples and the tank headspace are presented in Table 3.7. A representative total ion chromatogram showing the identity of major constituents is given in Figure 3.1.

Table 3.1 lists the quantitative results for target analytes. Sixteen target analytes above the 5-ppbv reporting cutoff were detected in the tank-headspace samples. Hexane (1.47 mg/m^3), trichlorofluoromethane (0.84 mg/m^3), and acetone (0.33 mg/m^3) accounted for 75% of the target analyte concentration and 8% of the total concentration identified by both the target and TIC analyses. Hexane was 42% of the target analyte concentration and 5% of the total concentration identified by both analyses. The total target analyte concentration was measured to be 3.53 mg/m^3 or 11% of the total concentration identified by both the target and TIC analyses.

Table 3.2 lists the semi-quantitative results for the TICs. A total of 18 TICs was identified. However, 16 of the 18 TICs were identified in two or more of the SUMMA™ canisters. Three TICs were identified as unknown without any associated molecular weight. The predominant species observed in these samples were tridecane (13.86 mg/m^3), dodecane (7.19 mg/m^3), and tetradecane (5.20 mg/m^3). Tridecane, the highest concentration TIC, accounted for 49% of the TIC concentration and 44% of the total concentration identified by both analyses. The total concentration of the TICs was found to be 28.33 mg/m^3 or 89% of the total concentration identified by both the target and TIC analyses. Quantitation of TICs is based on the total ion response of the closest eluting internal standard. Hexane, a target compound, coelutes with the first internal standard (bromochloromethane). Since hexane was found in the tank samples at a level of approximately 380 ppbv, the total ion response of bromochloromethane will include the contribution from hexane. Therefore, all TICs measured against the bromochloromethane response will be biased low. This consists of TICs eluting in the retention time range of approximately 4 to 17 minutes.

SUMMA™ canister PNL 81 was analyzed in replicate for target analytes and TICs to determine analytical precision. The relative percent difference (RPD) results are presented in Tables 3.3 and 3.4. The RPD was calculated for analytes detected above the detection limit and found in both replicates. Ten of the 16 target analytes and 12 of 15 TICs had an RPD of less than 10%.

Tables 3.5 and 3.6 list the compounds identified in the ambient air collected upwind of Tank TY-103 and ambient air through the VSS sample. One target analyte, acetone (0.03 mg/m^3), was identified in the ambient air through the VSS sample. One TIC, 3-butene-2-one (0.07 mg/m^3), was identified in the upwind ambient-air sample.

Table 3.7 lists results of the permanent gas analysis from samples collected from the headspace of TY-103, ambient air collected $\sim 10 \text{ m}$ upwind of the tank, and ambient air collected through the VSS. Permanent gases identified in the headspace were CO_2 (121 ppmv) and N_2O (159 ppmv). Carbon dioxide in the headspace, at 121 ppmv, was at a lower concentration than in ambient air. Nitrous oxide was not detected in either the ambient-air sample collected $\sim 10 \text{ m}$ upwind of the tank or the ambient-air sample collected through the VSS. A replicate analysis was performed on SUMMA™ canister PNL 81 (see Table 3.7 footnote c); however, only the results from the first analysis are included in the average concentration of the tank-headspace samples.

4.0 Conclusions

The concentrations of selected inorganic and organic compounds were determined from samples of the headspace of Tank TY-103 on 4/11/95. Sampling and analysis methods followed those described by Ligothke et al. (1994) and Clauss et al. (1994) for samples obtained from C-103, a tank containing a relatively complex headspace composition. Method-validation measurements during that study did appear to validate the trapping and analysis of NH_3 , but did not eliminate the possibility of interferences that could affect NO_x results. It is recommended that additional control samples be obtained if a tank is discovered in the future to contain significant quantities of NO_x . In the current sample job, NO_x samples were obtained after first passing the sample flow through an NH_3 trap. The average and standard deviation of the concentration results from inorganic sorbent trains were 49 ± 1 ppmv (NH_3), ≤ 0.06 ppmv (NO_2), 0.10 ± 0.02 ppmv (NO), and 12.7 ± 0.1 mg/L (vapor-mass concentration). The vapor-mass concentration is expected to consist largely of water vapor. Field blanks were used to correct data. All analytical results were within the target criteria $\{\pm 25\%$ precision, 70 to 130% accuracy (Carpenter 1994)} for inorganic analytes found at concentrations exceeding the lower target analytical limits (Table 2.1).

Organic analysis of the tank-headspace samples from Tank TY-103 identified 16 target analytes above the 5-ppbv reporting cutoff and 18 TICs above the 10-ppbv reporting cutoff. However, only 16 of the 18 TICs were observed in two or more of the SUMMA™ canisters. Hexane, the highest concentration target analyte, accounted for 42% of the total concentration of target analytes and 5% of the total concentration identified by both analyses. The total concentration of the target analytes accounted for 11% of the total concentration identified by both the target and TIC analyses. Tentatively identified compounds accounted for 89% of the total concentration identified by both the target and TIC analyses. Tridecane, the highest concentration TIC, accounted for 49% of the TIC concentration and 44% of the total concentration identified by both analyses. Results of replicate analysis on a single SUMMA™ canister observed 10 of 16 target analytes and 12 of 15 TICs having an RPD of less than 10%. One target analyte, acetone, was identified in the ambient air through the VSS sample. One TIC, 3-butene-2-one, was identified in the upwind ambient-air sample. Two permanent gases, CO_2 and N_2O , were detected in the tank-headspace samples.

5.0 References

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6.0 Further Reading

Pacific Northwest Laboratory. Analytical Laboratory Procedure Compendium. Procedures PNL-ALO-271. PNL-MA-599, Richland, Washington.

Pacific Northwest Laboratory. Quality Assurance Manual, Part 2: Good Practices Standard. PNL-MA-70, Part 2, Richland, Washington.

Pacific Northwest Laboratory. PNL TWRS Tank Vapor Program. MCS-046, Richland, Washington.

Table 3.1 Positively Identified and Quantitated Target Analytes^(a) of Samples Collected from the Headspace of Tank TY-103 in SUMMATM Canisters on 4/11/95

Analyte	CAS No.	Mol Wt	S5020-A04.080 ^(b)		S5020-A05.081 ^{(b)(c)}		S5020-A06.097 ^(b)		Means and Standard Deviations			
			PNL 80 ^(c)		PNL 81 ^{(c)(d)}		PNL 97 ^(c)		(mg/m ³)	St Dev	(ppbv)	St Dev
			(mg/m ³)	(ppbv)	(mg/m ³)	(ppbv)	(mg/m ³)	(ppbv)				
Dichlorodifluoromethane	75-71-8	120.9	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Chloromethane	74-87-3	50.5	<0.01	<5	<0.01	<5	<0.01	<5	(e)	(e)	(e)	(e)
1,2-Dichloro-1,1,2,2-tetrafluoroethane	76-14-2	170.9	<0.04	<5	<0.04	<5	<0.04	<5	(e)	(e)	(e)	(e)
Vinyl Chloride	75-01-4	62.5	<0.01	<5	<0.01	<5	<0.01	<5	(e)	(e)	(e)	(e)
Bromomethane	74-83-9	94.9	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Chloroethane	75-00-3	64.5	<0.01	<5	<0.01	<5	<0.01	<5	(e)	(e)	(e)	(e)
Acetonitrile	75-05-8	41.1	0.14	78	0.14	75	0.13	72	0.14	0.01	75	3.0
Acetone	67-64-1	58.1	0.32	125	0.32	124	0.36	140	0.33	0.02	129	9.4
Trichlorofluoromethane	75-69-4	137.4	0.79	128	0.82	133	0.92	149	0.84	0.07	137	11.0
1,1-Dichloroethene	75-35-4	96.9	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Methylene Chloride	75-09-2	84.9	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
1,1,2-Trichloro-1,2,2-trifluoroethane	76-13-1	187.4	0.05	6	0.05	6	0.06	7	0.05	0.01	7	0.6
Propanenitrile	107-12-0	55.1	0.05	20	0.05	20	0.02	10	0.04	0.02	17	6.1
Propanol	71-23-8	60.1	0.07	27	0.07	27	0.04	15	0.06	0.02	23	6.9
1,1-Dichloroethane	75-34-3	99.0	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
2-Butanone	78-93-3	72.1	0.04	12	0.04	14	0.08	24	0.05	0.02	17	6.7
cis-1,2-Dichloroethene	156-59-2	96.9	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Hexane	110-54-3	86.2	1.42	369	1.54	400	1.46	379	1.47	0.06	383	16.0
Chloroform	67-66-3	119.4	0.04	8	0.04	8	0.04	8	0.04	0.00	8	0.2
Tetrahydrofuran	109-99-9	72.1	0.07	21	0.07	23	0.07	21	0.07	0.00	22	0.8
1,2-Dichloroethane	107-06-2	99.0	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
1,1,1-Trichloroethane	71-55-6	133.4	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Butanenitrile	109-74-0	69.1	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Benzene	71-43-2	78.1	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Carbon Tetrachloride	56-23-5	153.8	0.19	28	0.21	31	0.19	28	0.20	0.01	29	1.6
Cyclohexane	110-82-7	84.2	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
1,2-Dichloropropane	78-87-5	113.0	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Trichloroethene	79-01-6	131.4	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Heptane	142-82-5	100.2	0.03	6	0.03	7	0.03	6	0.03	0.00	6	0.4
cis-1,3-Dichloropropene	10061-01-5	111.0	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
4-Methyl-2-Pentanone	108-10-1	100.2	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Pyridine	110-86-1	79.1	0.07	21	0.03	10	0.02	6	0.04	0.03	12	8.0
trans-1,3-Dichloropropene	10061-02-6	111.0	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)

TY-103 Table 3.1 (Contd)

Analyte	CAS No.	Mol Wt	S5020-A04.080 ^(b)		S5020-A05.081 ^{(b)(c)}		S5020-A06.097 ^(b)		Means and Standard Deviations			
			PNL 80 ^(c)		PNL 81 ^{(c)(d)}		PNL 97 ^(c)		(mg/m ³)	St Dev	(ppbv)	St Dev
			(mg/m ³)	(ppbv)	(mg/m ³)	(ppbv)	(mg/m ³)	(ppbv)	(mg/m ³)	St Dev	(ppbv)	St Dev
1,1,2-Trichloroethane	79-00-5	133.4	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Toluene	108-88-3	92.1	0.05	13	0.06	14	0.05	13	0.05	0.01	13	0.6
1,2-Dibromoethane	106-93-4	187.9	<0.04	<5	<0.04	<5	<0.04	<5	(e)	(e)	(e)	(e)
Tetrachloroethylene	127-18-4	165.8	0.07	10	0.08	11	0.07	10	0.07	0.01	10	0.4
Chlorobenzene	108-90-7	112.6	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Ethylbenzene	100-41-4	106.2	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
p/m-Xylene ^(f)	106-42-3	106.2	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Cyclohexanone	108-94-1	98.1	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
Styrene	100-42-5	104.2	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
1,1,2,2-Tetrachloroethane	79-34-5	167.9	<0.04	<5	<0.04	<5	<0.04	<5	(e)	(e)	(e)	(e)
o-Xylene	95-47-6	106.2	<0.02	<5	<0.02	<5	<0.02	<5	(e)	(e)	(e)	(e)
1,3,5-Trimethylbenzene	108-67-8	120.2	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
1,2,4-Trimethylbenzene	95-63-6	120.2	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
Decane	124-18-5	142.3	0.04	6	0.04	6	0.04	6	0.04	0.00	6	0.0
1,3-Dichlorobenzene	541-73-1	147.0	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
1,4-Dichlorobenzene	106-46-7	147.0	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
1,2-Dichlorobenzene	95-50-1	147.0	<0.03	<5	<0.03	<5	<0.03	<5	(e)	(e)	(e)	(e)
1,2,4-Trichlorobenzene	120-82-1	181.5	<0.04	<5	<0.04	<5	<0.04	<5	(e)	(e)	(e)	(e)
Hexachloro-1,3-butadiene	87-68-3	260.8	<0.06	<5	<0.06	<5	<0.06	<5	(e)	(e)	(e)	(e)

(a) TO-14 plus 14 additional target analytes.

(b) WHC sample identification number.

(c) PNL canister number.

(d) Replicates of this sample are found in Table 3.3

(e) Average and/or standard deviation are not meaningful for this analyte.

(f) m-Xylene and p-Xylene coelute; the reported concentration is the sum of these two compounds.

Table 3.2 Tentatively Identified Compounds and Estimated Concentrations^(a) of Samples from the Headspace of Tank TY-103 in SUMMATM Canisters Collected on 4/11/95

Tentatively Identified Compound ^(e)	CAS No. ^(e)	Mol Wt	Ret Time	S5020-A04.080 ^(b)		S5020-A05.081 ^{(b)(c)}		S5020-A06.097 ^(b)		Means and Standard Deviations			
				PNL 80 ^(d) (mg/m ³)	(ppbv)	PNL 81 ^(d) (mg/m ³)	(ppbv)	PNL 97 ^(d) (mg/m ³)	(ppbv)	(mg/m ³)	St Dev	(ppbv)	St Dev
Propane	74-98-6	44	4.9	0.26	130	0.21	106	0.24	121	0.23	0.02	119	12.1
Methyl Alcohol	67-56-1	32	5.8	0.16	113	0.16	114	0.18	127	0.17	0.01	118	8.1
Butane	106-97-8	58	6.4	0.08	29	0.08	32	0.08	31	0.08	0.00	31	1.6
Isopropyl Alcohol	67-63-0	60	9.1	0.06	21	<0.03	<10	0.06	23	0.06	(f)	22	(f)
3-Buten-2-one	78-94-4	70	12.8	<0.03	<10	<0.03	<10	<0.03	<10	(f)	(f)	(f)	(f)
1-Butanol	71-36-3	74	17.1	0.55	163	0.55	168	0.55	167	0.55	0.00	167	1.2
Pentanal	110-62-3	86	18.6	0.08	20	0.08	21	0.08	20	0.08	0.00	20	0.3
Hexanal	66-25-1	100	24.2	0.08	17	0.08	18	0.08	17	0.08	0.00	17	0.8
Nitric acid, butyl ester	928-45-0	119	25.4	0.05	10	0.05	10	0.05	9	0.05	0.00	10	0.3
Unknown Alcohol			28.0	(g)	(g)	(g)	(g)	0.08	(g)	(f)	(f)	(f)	(f)
Octanal	124-13-0	128	34.4	0.07	11	0.07	12	0.07	12	0.07	0.00	11.8	0.6
Undecane	1120-21-4	156	39.4	0.35	51	0.38	54	0.34	49	0.36	0.02	51	2.7
Dodecane	112-40-3	170	43.5	7.05	929	7.53	992	7.00	922	7.19	0.29	947.5	38.4
Undecane, 2,6-dimethyl-	17301-23-4	184	44.1	<0.08	<10	<0.08	<10	<0.08	<10	(f)	(f)	(f)	(f)
Unknown C12 Alkene/Cycloalkane		168	45.5	<0.08	<10	<0.08	<10	<0.08	<10	(f)	(f)	(f)	(f)
Unknown C14 Alkane		198	46.4	0.09	10	0.10	11	0.09	10	0.09	0.00	10.5	0.3
Tridecane	629-50-5	184	47.3	13.49	1642	14.68	1787	13.42	1634	13.86	0.71	1687.6	85.8
Unknown C12 Ketone		184	49.5	0.12	15	0.21	25	0.21	25	0.18	0.05	21.9	6.2
Unknown Alkane			50.1	(g)	(g)	(g)	(g)	0.05	(g)	(f)	(f)	(f)	(f)
Tetradecane	629-59-4	198	50.8	4.65	526	6.29	712	4.66	527	5.20	0.95	588.2	107.0
Unknown Ketone			52.9	0.07	(g)	0.14	(g)	0.05	(g)	0.09	0.05	(f)	(f)

- (a) Semi-quantitative estimate calculated using concentration of closest eluting IS.
 (b) WHC sample identification number.
 (c) Replicates of this sample are found in Table 3.4.
 (d) PNL SUMMATM canister number.
 (e) Obtained by mass spectral interpretation and comparison with the EPA/NIST/WILEY Library.
 (f) Mean and/or standard deviation are not meaningful for this analyte.
 (g) Molecular weight not available for this analyte.

Table 3.3 Positively Identified and Quantitated Target Analytes^(a) of Replicate Analyses of a Single SUMMA™ Canister Collected from the Headspace of Tank TY-103 on 4/11/95

Analyte	CAS No.	Mol Wt	S5020-A05.081 ^(b)		S5020-A05.081 ^(b)		Relative Percent Difference %
			PNL 81 ^(c)	(mg/m ³) (ppbv)	PNL 81 ^(c)	(mg/m ³) (ppbv)	
Dichlorodifluoromethane	75-71-8	120.9	<0.03	<5	<0.03	<5	
Chloromethane	74-87-3	50.5	<0.01	<5	<0.01	<5	
1,2-Dichloro-1,1,2,2-tetrafluoroethane	76-14-2	170.9	<0.04	<5	<0.04	<5	
Vinyl Chloride	75-01-4	62.5	<0.01	<5	<0.01	<5	
Bromomethane	74-83-9	94.9	<0.02	<5	<0.02	<5	
Chloroethane	75-00-3	64.5	<0.01	<5	<0.01	<5	
Acetonitrile	75-05-8	41.1	0.14	75	0.17	90	19.4
Acetone	67-64-1	58.1	0.32	124	0.33	129	3.1
Trichlorofluoromethane	75-69-4	137.4	0.82	133	0.85	139	3.6
1,1-Dichloroethene	75-35-4	96.9	<0.02	<5	<0.02	<5	
Methylene Chloride	75-09-2	84.9	<0.02	<5	<0.02	<5	
1,1,2-Trichloro-1,2,2-trifluoroethane	76-13-1	187.4	0.05	6	0.06	7	18.2
Propanenitrile	107-12-0	55.1	0.05	20	0.02	9	85.7
Propanol	71-23-8	60.1	0.07	27	0.05	18	33.3
1,1-Dichloroethane	75-34-3	99.0	<0.02	<5	<0.02	<5	
2-Butanone	78-93-3	72.1	0.04	14	0.07	21	54.5
cis-1,2-Dichloroethene	156-59-2	96.9	<0.02	<5	<0.02	<5	
Hexane	110-54-3	86.2	1.54	400	1.59	414	3.2
Chloroform	67-66-3	119.4	0.04	8	0.05	9	22.2
Tetrahydrofuran	109-99-9	72.1	0.07	23	0.07	23	0.0
1,2-Dichloroethane	107-06-2	99.0	<0.02	<5	<0.02	<5	
1,1,1-Trichloroethane	71-55-6	133.4	<0.03	<5	<0.03	<5	
Butanenitrile	109-74-0	69.1	<0.02	<5	<0.02	<5	
Benzene	71-43-2	78.1	<0.02	<5	<0.02	<5	
Carbon Tetrachloride	56-23-5	153.8	0.21	31	0.2	30	4.9
Cyclohexane	110-82-7	84.2	<0.02	<5	<0.02	<5	
1,2-Dichloropropane	78-87-5	113.0	<0.03	<5	<0.02	<5	
Trichloroethene	79-01-6	131.4	<0.03	<5	<0.02	<5	
Heptane	142-82-5	100.2	0.03	7	0.03	7	0.0
cis-1,3-Dichloropropene	10061-01-	111.0	<0.02	<5	<0.02	<5	
4-Methyl-2-Pentanone	108-10-1	100.2	<0.02	<5	<0.02	<5	
Pyridine	110-86-1	79.1	0.03	10	0.03	8	0.0

TY-103 Table 3.3 (Contd)

Analyte	CAS No.	Mol Wt	S5020-A05.081 ^(b)		S5020-A05.081 ^(b)		Relative Percent Difference %
			PNL 81 ^(c) (mg/m ³)	(ppbv)	PNL 81 ^(c) (mg/m ³)	(ppbv)	
trans-1,3-Dichloropropene	10061-02-	111.0	<0.02	<5	<0.02	<5	
1,1,2-Trichloroethane	79-00-5	133.4	<0.03	<5	<0.03	<5	
Toluene	108-88-3	92.1	0.06	14	0.06	14	0.0
1,2-Dibromoethane	106-93-4	187.9	<0.04	<5	<0.04	<5	
Tetrachloroethylene	127-18-4	165.8	0.08	11	0.08	11	0.0
Chlorobenzene	108-90-7	112.6	<0.03	<5	<0.03	<5	
Ethylbenzene	100-41-4	106.2	<0.02	<5	<0.02	<5	
p/m-Xylene ^(d)	106-42-3	106.2	<0.02	<5	<0.02	<5	
Cyclohexanone	108-94-1	98.1	<0.02	<5	<0.02	<5	
Styrene	100-42-5	104.2	<0.02	<5	<0.02	<5	
1,1,2,2-Tetrachloroethane	79-34-5	167.9	<0.04	<5	<0.04	<5	
o-Xylene	95-47-6	106.2	<0.02	<5	<0.02	<5	
1,3,5-Trimethylbenzene	108-67-8	120.2	<0.03	<5	<0.03	<5	
1,2,4-Trimethylbenzene	95-63-6	120.2	<0.03	<5	<0.03	<5	
Decane	124-18-5	142.3	0.04	6	0.04	6	0.0
1,3-Dichlorobenzene	541-73-1	147.0	<0.03	<5	<0.03	<5	
1,4-Dichlorobenzene	106-46-7	147.0	<0.03	<5	<0.03	<5	
1,2-Dichlorobenzene	95-50-1	147.0	<0.03	<5	<0.03	<5	
1,2,4-Trichlorobenzene	120-82-1	181.5	<0.04	<5	<0.04	<5	
Hexachloro-1,3-butadiene	87-68-3	260.8	<0.06	<5	<0.06	<5	

(a) TO-14 plus 14 additional target analytes.

(b) WHC sample identification number.

(c) PNL canister number.

(d) m-Xylene and p-Xylene coelute; the reported concentration is the sum of these two compounds.

Table 3.4 Tentatively Identified Compounds and Estimated Concentrations^(a) of Replicate Analyses of a Single SUMMATM Canister Collected from the Headspace of Tank TY-103 on 4/11/95

Tentatively Identified Compound ^(d)	CAS No. ^(d)	Mol Wt	Ret Time	S5020-A05.081 ^(b)		S5020-A05.081 ^(b)		Relative Percent Difference %
				PNL 81 ^(c) (mg/m ³)	(ppbv)	PNL 81 ^(c) (mg/m ³)	(ppbv)	
Propane	74-98-6	44	4.9	0.21	106	0.22	114	7.0
Methyl Alcohol	67-56-1	32	5.8	0.16	114	0.15	106	7.6
Butane	106-97-8	58	6.4	0.08	32	0.11	43	28.9
Isopropyl Alcohol	67-63-0	60	9.1	<0.03	<10	<0.03	<10	
3-Buten-2-one	78-94-4	70	12.8	<0.03	<10	<0.03	<10	
1-Butanol	71-36-3	74	17.1	0.55	168	0.54	163	2.7
Pentanal	110-62-3	86	18.6	0.08	21	0.08	20	5.2
Hexanal	66-25-1	100	24.2	0.08	18	0.08	18	1.2
Nitric acid, butyl ester	928-45-0	119	25.4	0.05	10	0.06	11	5.5
Unknown Alcohol			28.0	(e)	(e)	0.07	(e)	
Octanal	124-13-0	128	34.4	0.07	12	0.07	12	5.8
Undecane	1120-21-4	156	39.4	0.38	54	0.37	54	0.8
Dodecane	112-40-3	170	43.5	7.53	992	7.20	948	4.5
Undecane, 2,6-dimethyl-	17301-23-4	184	44.1	<0.08	<10	<0.08	<10	
Unknown C12 Alkene/Cycloalkane		168	45.5	<0.08	<10	<0.08	<10	
Unknown C14 Alkane		198	46.4	0.10	11	0.10	11	2.1
Tridecane	629-50-5	184	47.3	14.68	1787	14.47	1762	1.4
Unknown C12 Ketone		184	49.5	0.21	25	0.21	25	0.5
Unknown Alkane			50.1	(e)	(e)	0.05	(e)	
Tetradecane	629-59-4	198	50.8	6.29	712	5.47	619	14.0
Unknown Ketone			52.9	0.14	(e)	0.10	(e)	39.7

(a) Semi-quantitative estimate calculated using concentration of closest eluting IS.

(b) WIIC sample identification number.

(c) PNL SUMMATM canister number.

(d) Obtained by mass spectral interpretation and comparison with the EPA/NIST/WILEY Library.

(e) No molecular weight available for calculation.

Table 3.5 Positively Identified and Quantitated Target Analytes^(a) for Ambient Air and Ambient Air Through the VSS Collected Near Tank TY-103 in SUMMA™ Canisters on 4/11/95

Analyte	CAS No.	Mol Wt	Ambient Air Upwind S5020-A01.029 ^(b) PNL 29 ^(c)	Ambient Air Through VSS S5020-A02.067 ^(b) PNL 67 ^(c)
Acetone	67-64-1	58.08	(mg/m ³) (ppbv) <0.02 <5	(mg/m ³) (ppbv) 0.03 11.05

(a) TO-14 plus 14 additional analytes.

(b) WHIC sample identification number.

(c) PNL canister number.

Table 3.6 Tentatively Identified Compounds and Estimated Concentrations^(a) for Ambient Air and Ambient Air Through the VSS Collected Near Tank TY-103 in SUMMA™ Canisters on 4/11/95

Tentatively Identified Compound ^(d)	CAS No. ^(d)	Mol Wt	Ret Time	Ambient Air Upwind S5020-A01.029 ^(b) PNL 29 ^(c)		Ambient Air Through VSS S5020-A02.067 ^(b) PNL 67 ^(c)	
				(mg/m ³)	(ppbv)	(mg/m ³)	(ppbv)
3-Buten-2-one	78-94-4	70	12.8	0.07	23	<0.03	<10

(a) Semi-quantitative estimate calculated using concentration of closest eluting IS.

(b) WHC sample identification number.

(c) PNL canister number.

(d) Obtained by mass spectral interpretation and comparison with the EPA/NIST/WILEY Library.

Table 3.7 Permanent Gas Analysis Results for Samples Collected from the Headspace of Tank TY-103 and for Ambient Air and Ambient Air Through the VSS Collected near Tank TY-103 in SUMMA^(TM) Canisters on 4/11/95.

Permanent Gas Analyte	Ambient Air Upwind	Ambient Air Through VSS	Tank Samples				Average Concentration Tank Samples (ppmv)
	S5-020-A01.029 ^(a) PNL 029 ^(b) Concentration (ppmv)	S5-020-A02.067 ^(a) PNL 067 ^(b) Concentration (ppmv)	S5-020-A04.080 ^(a) PNL 080 ^(b) Concentration (ppmv)	S5-020-A05.081 ^(a) PNL 081 ^(b) Concentration (ppmv)	S5-020-A06.097 ^(a) PNL 097 ^(b) Concentration (ppmv)	S5-020-A05.081 ^(a) PNL 081 ^{(b)(c)} Concentration (ppmv)	
Hydrogen	< 93	< 93	< 93	< 93	< 93	< 93	< 93
Methane	< 12	< 12	< 12	< 12	< 12	< 12	< 12
Carbon Dioxide	357	356	127	124	113	122	121
Carbon Monoxide	< 12	< 12	< 12	< 12	< 12	< 12	< 12
Nitrous Oxide	< 12.6	< 12.6	130	146	201	195	159

(a) WHC sample identification number.

(b) PNL canister number.

(c) Replicate analysis for PNL 81; results are not included in the calculation of average concentrations.

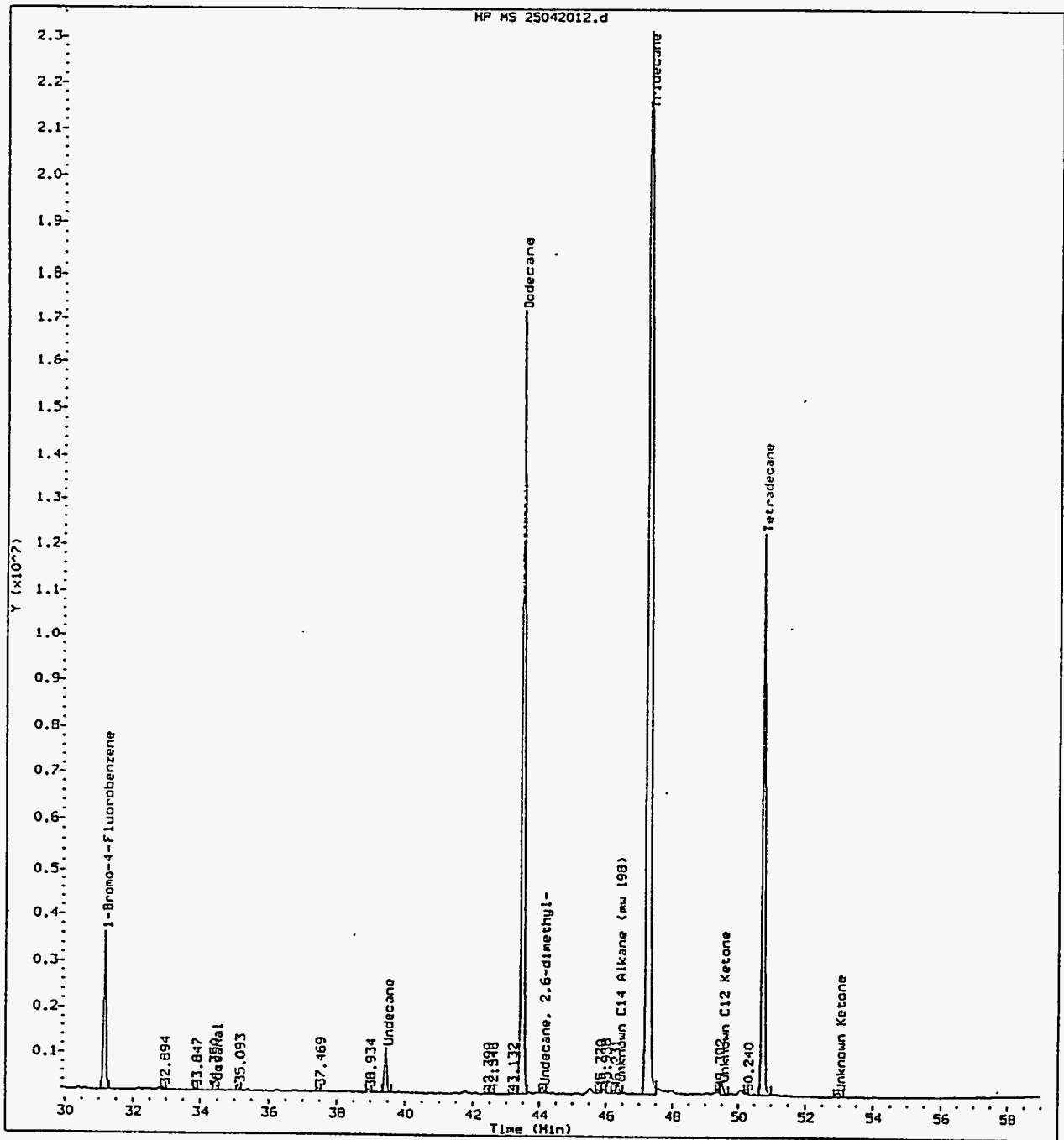


Figure 3.1b Total Ion Chromatogram (30 - 58 min) for Hanford Waste Tank TY-103 SUMMA™ Canister Sample S5020-A04-080 Collected on 4/11/95

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