Accepted Manuscript

Title: Determination of solvation descriptors for terpene hydrocarbons from chromatographic measurements

Author: Michael H Abraham Joelle R.M. Gola Javier Gil-Lostes William E. Acree Jr. J. Enrique Cometto- Muñiz



PII:S0021-9673(13)00551-7DOI:http://dx.doi.org/doi:10.1016/j.chroma.2013.03.068Reference:CHROMA 354222To appear in:Journal of Chromatography A

 Received date:
 15-2-2013

 Revised date:
 21-3-2013

 Accepted date:
 25-3-2013

Please cite this article as: M.H. Abraham, J.R.M. Gola, J. Gil-Lostes, W.E. Acree Jr., J.E.C.- Muñiz, Determination of solvation descriptors for terpene hydrocarbons from chromatographic measurements, *Journal of Chromatography A* (2013), http://dx.doi.org/10.1016/j.chroma.2013.03.068

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

CRI CCEPT **3** D P 1

Highlights 1

2 3 4

5

- Prediction of chromatographic rentention data for terpene hydrocarbons •
- Development of predictive linear free energy relationships •
- Calculation of solute descriptors for terpene hydrocarbons •

~

- 6 7
- 8

Determination of solvation descriptors for terpene hydrocarbons from 9 chromatographic measurements 10 Michael H Abraham, ^{a *} Joelle R. M. Gola, ^a Javier Gil-Lostes, ^a William E. Acree, Jr, ^b and 11 J. Enrique Cometto- Muñiz^c 12 ^a Department of Chemistry, University College London, 20 Gordon St, London WC1H 0AJ, 13 14 UK. Email: m.h.abraham@ucl.ac.uk ^b Department of Chemistry, 1155 Union Circle Drive #305070, University of North Texas, 15 16 Denton, TX 76203-5017, USA 17 ^c Chemosensory Perception Laboratory, Department of Surgery (Otolaryngology), University 18 of California, San Diego, La Jolla, CA 92093-0957 19 20 ABSTRACT 21 Gas chromatographic retention data on 107 terpene hydrocarbons from the literature together 22 with other data have been used to obtain a set of Abraham descriptors for these 107 23 compounds. For 88 aliphatic cyclic terpene hydrocarbons, a fragmentation scheme was 24 constructed that allows key descriptors to be estimated just from structure. The total set of 25 descriptors, including those estimated by the fragmentation schemes, were then used to predict water-octanol partition coefficients for the 88 compounds, there being good 26 27 agreement with values calculated from a number of well-known programs. For a small 28 number of terpene hydrocarbons, there was good agreement between predicted and 29 experimental values of nasal pungency thresholds, and predicted and experimental gas-blood, 30 gas-oil, and gas-water partition coefficients. It is suggested that the descriptors obtained for 31 the 107 terpene hydrocarbons can be used to predict water-solvent partition coefficients, gas-

- 32 solvent partition coefficients, and partition coefficients in a number of biological systems.
- 33

34

35 Keywords:

36 Terpenes

37 Gas liquid chromatography

38 Partition coefficient

39 Descriptors

40 Linear free energy relationships

41

42 **1. Introduction**

Terpenes are found in a wide variety of essential oils. Geraniol is the main constituent of geranium oil, limonene the major constituent of lemon oil, menthol the principal constituent of pepermint oil and α -pinene the main constituent of turpentine. Many terpenes are of industrial importance; Sell [1] lists 20 terpenes that are used on an industrial scale, for example 30,000 tons of myrcene per annum, and limonene has widespread use as a fragrance component.

49 In spite of their widespread occurrence and use, there is very little known as to the effect 50 of terpenes on humans, other than skin irritation and sensitization [2]. Cometto-Muñiz et al. 51 [3] determined odor detection thresholds (ODT), eye irritation thresholds (EIT) and nasal 52 pungency thresholds (NPT) for eleven terpenes. Nagata [4] included only three terpenes in 53 his extensive investigation on ODTs, and Rodriguez et al. [5] only seven terpenes in ODT 54 values for 100 compounds. There is an enormous number of terpenes likely to be encountered 55 in everyday life, and some method of predicting their ODT, EIT, and NPT thresholds 56 thresholds is clearly of importance. Since very little data exists on physicochemical properties of terpenes, the ability to predict such properties would also be extremely useful. Our 57 58 method is based on the two linear free energy relationships, LFERs, Eq. (1) and Eq. (2) [6-8]. SP = c + e E + s S + a A + b B + v V59 (1)

60

$61 \qquad SP = c + e E + s S + a A + b B + l L$

The dependent variable, *SP*, is some property of a series of solutes in a given system. *SP* may be partition coefficients as, $\log P_s$, for a series of solutes in a given water-solvent system or may be $\log K_s$ for a series of solutes in a given gas-solvent system. The former are mostly used in Eq. (1) and the latter in Eq. (2). Other systems can also be used. For example *SP* in Eq. (2) can be a set of retention data for a series of solutes on a given gas liquid chromatographic, GLC, stationary phase.

The independent variables, or Abraham descriptors, in Eq. (1) and Eq. (2) are properties of solutes as follows [6-8]: *E* is an excess molar refraction in cm³ mol⁻¹/10, *S* is a combined dipolarity/polarizability descriptor, *A* is the overall solute hydrogen bond acidity, *B* is the overall solute hydrogen bond basicity, *V* is McGowan's [9] characteristic molecular volume in cm³ mol⁻¹/100 and *L* is the gas to hexadecane partition coefficient at 25^oC, as log K_s .

(2)

73 The constant c and the set of coefficients, e, s, a, b, v and l characterise the system and are determined by multiple linear regression analysis. These coefficients are not just fitting 74 75 coefficients, but represent the complementary properties of the system: e gives the 76 contribution in Eq. (1) and Eq. (2) of the solute-system general dispersion interaction, s is the 77 solvent dipolarity (plus some polarizability), a is the system hydrogen bond basicity (the 78 complimentary property to solute hydrogen bond acidity) and b is the system hydrogen bond 79 acidity (the complimentary property to solute hydrogen bond basicity). The coefficients in 80 Eq. (1) and Eq. (2) for a few processes that have been used to obtain descriptors for terpenes 81 are in Table 1 [8, 10-13].

Abraham et al. [14] set out an equation for NPT values, based on Eq. (2) and later incorporated a number of terpenes into the algorithm [15]. Algorithms that included a few terpenes were also constructed for EIT [15] and ODT [16] values. These equations required knowledge of the descriptors in Eq. 2 for the terpenes, but it has not been easy to obtain these for any substantial number.

87 Descriptors for 30 terpenes have been obtained [17], mostly from GLC retention data 88 on various stationary phases. It was pointed out [17] that none of the GLC stationary phases possessed any hydrogen bond acidity, and hence that the important *B*-descriptor could not be 89 90 obtained from GLC data. Abraham et al. [17] managed to obtain the B-descriptor for a few terpenes from known water-octanol partition coefficients, as log P_{oct} , and from their own 91 92 measurements of HPLC capacity factors but for most of the terpenes they were only able to 93 estimate B-descriptors. Ahmed and Poole [12] listed descriptors for a large number of 94 compounds, including 23 terpenes. They gave no reference for the origin of the descriptors 95 but by inspection those for 19 of the terpenes were taken from the paper by Abraham et al. 96 [17]. Thus the *B*-descriptor for these 19 terpenes is subject to the same uncertainty as 97 mentioned by Abraham et al.[17]. In a later paper, Karunasekara and Poole [18] obtained 98 descriptors for 15 terpenes using a combination of GLC data with values of $\log P_{oct}$ for nine 99 terpenes and hexane-acetonitrile partition coefficients, as log $P_{\text{hex-me}}$. The *b*-coefficient for 100 the water-octanol system is numerically very large (-3.460) and so can lead to reasonably 101 accurate values of the *B*-descriptor; the *b*-coefficient in the hexane-acetonitrile system is 102 numerically much smaller (-0.966) and might be expected to lead to less accurate values. We 103 can roughly estimate the possible error in the *B*-descriptor, through the term SD(reg)/b where 104 SD(reg) is the standard deviation of the particular regression equation used, and b is the b-105 coefficient. For the octanol-water system the term is 0.116/3.46 = 0.034 and for the hexane-

106 acetonitrile system the term is 0.117/0.966 = 0.121, very much larger than the estimated error for the octanol-water system. Thus, as pointed out before [17], unless a water-octanol 107 108 partition coefficient is available (or very rarely another water-solvent system with a large b-109 coefficient), it is very difficult to obtain a reliable *B*-descriptor. Both Abraham et al [17] and 110 Karunasekara and Poole [18] used experimental values of the gas-water partition coefficient, K_w , in their descriptor determinations. These experimental values are invariably obtained 111 112 from the terpene solubility in water and the terpene vapour pressure. Unfortunately, variation 113 in reported experimental values suggest that there is likely to be considerable uncertainty in literature values of log K_w . For example, log K_w for limonene at 25°C is reported as -0.373 114 [19], -0.144 [20] and + 0.233 [21] so that extreme caution is necessary if log K_w values are 115 116 used.

In the present work we concentrate on terpene hydrocarbons, and collect in Table 2 the Abraham descriptors for the few terpene hydrocarbons that have been obtained through the use of GLC retention data coupled with an experimental value of log P_{oct} [12, 16]. Although the terpenes in Table 2 are structurally quite simple, there are sometimes quite large differences in the values of the descriptors for a given compound. Note that for all the terpene hydrocarbons we shall deal with, the *A*-descriptor is zero, and so we omit the zero *A*-values from Table 2.

We therefore set out to obtain a coherent set of descriptors for terpene hydrocarbons in the hope that we would have enough reliable values for the *B*-descriptor to be able to construct an algorithm for the prediction of *B*-values.

127

128 **2. Methods**

129 Our method for the determination of descriptors for a given terpene is based on Eq. (1) 130 and Eq. (2). The 'unknown' descriptors are usually E, S, A, B, V and L but for the terpene hydrocarbons the unknown descriptors are E, S, B, V and L. Values of E were obtained from 131 known experimental refractive indices at 20° C [22-25], exactly as described before [6, 7], and 132 133 V can simply be calculated from the number of atoms and rings in a molecule [6, 26]. If a water-solvent partition coefficient, P_s , is available, this can be converted into the 134 corresponding gas-solvent partition coefficient, K_s , through Eq. (3). This requires a 135 knowledge of K_w ; as we have seen, literature values of K_w (or log K_w) are not very reliable, 136 and we prefer to use $\log K_w$ as an extra descriptor to be determined. 137

138	
139	P = K / K
140	$\Gamma_{\mathcal{S}} = \Pi_{\mathcal{S}} / \Pi_{\mathcal{W}}$
141	Then the 'unknown' descriptors are S, B, L and log K_w . There is little problem in obtaining
142	equations for GLC retention data, following Eq. (2), and we have some 90 such equations
143	constructed mostly from retention indices on the constituents of natural oils and waxes. In a
144	very useful and important paper, Babushok et al. [27] surveyed the literature on retention
145	indices, I, on three particular stationary phases, viz.: dimethylsilicone (DIMS),
146	dimethylsilicone with 5% phenyl groups (DIMSP) and polyethylene glycol (PEG). Since
147	their data base covers a wide range of compounds, we thought that our equations for I-values
148	might be of use to other workers who wish to obtain descriptors from GLC data.
149	
150 151	I(DIMS) = 39.1 - 39.2 E + 107.5 S + 128.9 A + 205.0 L (4)
151 152 153 154	N = 174, SD = 15.9, R ² = 0.998, F =17338.6, PRESS = 45930, Q ² = 0.997, PSD = 16.5
155	I(DIMSP) = 45.9 - 42.3 E + 135.9 S + 126.9 A + 204.0 L (5)
157	N = 173, SD = 13.8, R^2 = 0.998, F =22907.9, PRESS = 34605, Q^2 = 0.998, PSD = 14.4
158	
159	I(PEG) = -62.2 + 91.8 E + 652.2 S + 1038.5 A + 213.3 L (6)
160	N = 172, SD = 35.3, R^2 = 0.993, F =5723.7, PRESS = 226094, Q^2 = 0.992, PSD = 36.8
161	
162	In some instances, the number of compounds for which we had descriptors was very small,
163	the large majority of compounds being terpenes, and so it was not possible to obtain an
164	equation for the I-values. We then waited until we had determined descriptors for enough
165	terpenes (through other equations) to construct an equation. An example is the set of retention
166	indices obtained by Darjazi et al. [28] using a DB-5 fused silica column.
167 168 169	I(DB-5) = 62.7 - 16.0 E + 124.2 S + 62.5 A + 200.5 L (7)
170 171 172 173	N = 55, SD = 10.3, R^2 = 0.998, F = 5994.4, PRESS = 8679.4, Q^2 = 0.997, PSD = 13.2

In Eqs. (5)-(7), N is the number of data points, that is compounds, R is the correlation coefficient, F is the Fisher F-statistic, PRESS and Q^2 are the leave-one-out statistics and PSD is the predictive standard deviation [29]. The rather better statistics of Eq. (7) is probably due to the data of Babushok et al. [27] being derived from several different sources. As is invariably the case for GLC retention data, none of the equations has a statistically significant term in *B*.

180 For a few terpene hydrocarbons, experimental values of log P_{oct} are listed in the EPI 181 software program, available through ChemSpider [20] and in the BioLoom software program [30]. We also calculated log P_{oct} using a number of the most widely used programes, viz the 182 EPI and the ACD programes [20], the ClogP program [30], the AlogPS program [31] and 183 184 SPARC [32]. Griffin et al. [33] have measured HPLC capacity factors on a C-18 stationary 185 phase using a number of water-methanol eluants for a large number of terpenes, including 10 186 terpene hydrocarbons. Zenkevich and Kushakova [13] have determined perfluorodecalin-187 acetonitrile partition coefficients for a large number of compounds, including some terpenes. 188 We have used their data to obtain equation coefficients as shown in Table 1. The b-189 coefficient is very small, 0.150, and so the equation is not at all useful in the determination of 190 the solute *B*-descriptor.

The set of equations on the lines of Eq. (1) and Eq (2) together with the corresponding dependent variables were then solved for the unknown descriptors *S*, *B*, *L* and log K_w by the trial-and-error procedure 'Solver' in Microsoft Excel. The equations that we used are in Table 1, together with a large number of equations for GLC retention data that are of little intrinsic value.

196

197 **3. Results**

198 By far the main set of terpene hydrocarbons are aliphatic cyclic compounds with a wide 199 variety of number and sizes of rings. For several of the terpene hydrocarbons we had a large 200 number of equations for GLC retention data, 65 in the case of α -pinene. Many of the 201 equations have large values of the s-coefficient, see for example Eq. (6), so that there is little 202 problem in obtaining the corresponding S-descriptor. We also had experimental values of log 203 P_{oct} [20, 30], HPLC capacity factors [33], and hexane-acetonitrile partition coefficients [12] 204 for a number of terpenes that helped to yield the total set of descriptors, including the B-205 descriptor. We then set out a fragmentation scheme for the estimation of B, using the number 206 and type of ethylenic double bonds and the number and type of rings in the terpene as the

207 fragments. The fragments were defined manually and an Excel spreadsheet was used for the calculations. As we dealt with more and more terpenes, we updated the scheme until we 208 209 reached an equation with the coefficients shown in Table 3. Since we had all the data 210 available, we set out similar fragmentation schemes for E and S, as shown also in Table 3. It 211 is very important to note that these schemes are intended to apply only to aliphatic cyclic 212 terpene hydrocarbons. Furthermore, they are purely empirical schemes. However, the 213 fragment scheme for B, especially, should be of some value in cases where a given set of 214 equations cannot yield a reliable value. We also used the same set of fragments to set out a 215 scheme for L, although we had to delete values for the saturated cyclic terpenes. Values of E, S, B and L obtained through the fragment schemes given in Table 3 are intended to be used in 216 217 Eq. (1) and Eq (2).

218

The final set of descriptors for the 88 aliphatic cyclic terpene hydrocarbons that we have studied is given in Table 4; in all cases the *A*-descriptor is zero. Also included in Table 4 are the molecular formula of the terpene and the number of equations for GLC retention data used in the analysis, n(GLC). Since our analysis yields log K_w as a matter of course, we include this very difficult to measure parameter as well. Where descriptors could be obtained from experimental data, for example *E* from a refractive index or *B* from a water-octanol partition coefficient these were taken as the preferred values.

226

We also studied a much smaller number, twelve, of aliphatic acyclic terpene hydrocarbons. These were easier to deal with than the aliphatic cyclic hydrocarbons because both *S* and *B* could be estimated by analogy from data on other aliphatic acyclic hydrocarbons, and *S* could also be established from equations for GLC retention data. In the event, there was good agreement between the two sets of S-values, and the final set of descriptors is in Table 4.

There are a few terpene hydrocarbons that are derivatives of benzene or of 1,2,3,4tetrahydronaphthalene, and we examined seven aromatic hydrocarbons, given as the final entries in Table 4. In these cases, there was only poor agreement between the *S*-descriptor obtained from the GLC equations, and the *S*-descriptor obtained by analogy with values for known aromatic hydrocarbons. In all cases, the *S*-descriptor from the GLC equations was smaller than the values estimated from known aromatic hydrocarbons. Fortunately, calculated values of log P_{oct} using several methods give reasonably consistent results, see

240 later, and so we selected *S* and *B* values that were consistent with results from the GLC 241 equations, with estimations using known values for aromatic hydrocarbons, and with the 242 calculated log P_{oct} values. This is a quite different analysis from that used for the aliphatic 243 terpene hydrocarbons, but we found it to be the only way that we could estimate descriptors. 244 Our selected values are in Table 4.

245

246 **4. Discussion**

As explained, above, for the 88 aliphatic cyclic hydrocarbons and the 12 aliphatic acyclic 247 248 hydrocarbons, the key descriptor S was obtained from the GLC equations or from the 249 fragmentation scheme in Table 3, and for most of the compounds the 'difficult' descriptor B 250 was obtained from the corresponding fragmentation scheme. We can check if our set of 251 descriptors is reasonable or not by a comparison of $\log P_{oct}$ calculated from the descriptors in Table 4 and log P_{oct} calculated by standard literature methods. We therefore assemble in 252 Table 5 values of log Poct calculated using the BioLoom ClogP program [30], the Advanced 253 Chemistry Development program (ACD) and the EPI program as implemented in 254 255 ChemSpider software [20], the AlogPS program [31] and the SPARC program [32]. These five methods are probably the most used calculations for log P_{oct} . We have shown [11] that 256 257 experimental values of log Poct are well represented through Eq. 8. Both the s-coefficient and especially the b-coefficient in Eq. (8) are numerically large and so estimations of log P_{oct} 258 259 should provide a quite rigorous test of our descriptors.

260

$$Log P_{oct} = 0.088 + 0.562 E - 1.054 S + 0.034 A - 3.460 B + 3.814 V$$
(8)

Results of the log P_{oct} calculations are in Table 5. It is not our intention to compare the 262 various literature methods for these calculations, but we note that the AlogPS method usually 263 gives log P_{oct} values smaller than the average, and that the SPARC method usually gives 264 265 values larger than the average. The differences in the various calculations can become very 266 large, so that for α -cubebene they amount to over four log units. The log P_{oct} values obtained from our descriptors, that include the S- and B-descriptors, are invariably within the upper 267 268 and lower limits of the calculated descriptors, and are usually close to the average value of 269 the ClogP, ACD, and EPI calculations. We therefore conclude that the fragment calculations 270 for S and B do yield reasonable values, so that the total set of descriptors then leads to

reasonable values for log P_{oct} . There is then every reason to think that the descriptors set out in Table 4 will yield good estimates of log P and log K values for the very large number of water-solvent and gas-solvent systems for which we have the required equations [8]. In a similar vein, the descriptors can be used to estimate nasal pungency thresholds. The calculations of log P, log K and NPT are completely trivial – all that is needed is to combine the descriptors with the corresponding equation coefficients.

There are a few cases in which our predictions can be compared to experiment. The most recent equation for NPT values [34] is Eq. (9), and our predicted values using the descriptors in Table 4 are compared with the observed values [17] in Table 6. The agreement is excellent and although we have only two sets of data, there seems no reason why the descriptors in Table 4 should not yield reasonable predictions of nasal pungency thresholds.

282

283
$$Log(1/NPT) = -7.770 + 1.543 S + 3.296 A + 0.876 B + 0.816 L$$
 (9)

284

A more extensive set of experimental results are those of Falk et al [35] on gas-blood, gas-water and gas-olive oil partitions at 37° C. We have equations for gas-blood partitions [36], gas-olive oil partitions [37] and gas-water partitions [38], Eq. (10) – Eq. (12), all of which refer to 37° C, and we compare our predictions with experimental values in Table 6.

289
$$\log K_{blood} (37^{\circ}C) = -1.062 + 0.460 E + 1.067 S + 3.777 A + 2.556 B + 0.375 L$$
 (10)

290
$$\log K_{olive oil} (37^{\circ}C) = -0.156 - 0.254 E + 0.859 S + 1.656 A + 0.873 L$$
 (11)

291
$$\log K_w (37^{\circ}\text{C}) = -1.035 + 0.575 E + 2.482 S + 3.601 A + 4.561 B - 0.973 V$$
 (12)

Agreement is generally very good, even for equations that have a substantial *b*-coefficient, so for the four terpenes studied by Falk et al. [35] the *B*-descriptor that is compatible with the coefficients in Table 3 seems to lead to reasonable values of the gas-solvent partition coefficients.

We note that the descriptors in Table 4 can be used for the prediction of gas-water partition coefficients from 0° C to 100° C, in combination with the equation coefficients we have already listed [38].

299

300 It is now possible to use the coefficients in Table 3 to predict *E*, *S* and *B* for aliphatic 301 cyclic terpene hydrocarbons. Then if GLC retention data are known for a few systems for 302 which equations are available, the entire set of descriptors *E*, *S*, *B*, *V* and *L* can be obtained. 303 We use the retention data of Babushok et al [27] for the systems characterized through Eq.

(4), (5) and (6) and calculate E, S, B, V as suggested. These are given in Table 7 for three 304 terpenes listed by Babushok et al [27]. As before, we can use the obtained set of descriptors 305 to estimate log P_{oct} through Eq. (8) and can compare our estimated values with literature 306 307 calculations, as shown in Table 7. There is good agreement between the log P_{oct} values predicted from our descriptors and those calculated by literature methods. Experimental data 308 309 on GLC retention on just a few characterized phases are all that is needed to determine the L-310 descriptor. Then partition coefficients can be estimated for transfer from the gas phase to 311 numerous solvents, as well as a large number of biological properties, including the important 312 nasal pungency thresholds. We give predictions for the latter in Table 7, with values of NPT 313 in ppm. Both α -bulnesene and β -sesquiphellandrene are quite potent vapours, of about the 314 same potency as that of hexanoic acid, which has $\log (1/\text{NPT}) = -1.30$ [14].

315

316 We carried out two final checks on our fragment scheme shown in Table 3. First we calculate E, S and B from the fragmentation scheme for the terpenes for which experimental 317 values of log P_{oct} are known, see Table 5. Then knowing V from simple calculation of atom 318 319 fragments [9] we have all the fragment data necessary to calculate log P_{oct} using Eq. (8). 320 Details are in Table 8, where the last column gives the average experimental values of log 321 Poct. Comparison of the calculated and average experimental values gives an average error, AE, (calculated – experimental) of -0.08, an average absolute error, AAE, of 0.12 and a 322 323 standard deviation, SD, of 0.17 log units. The latter compares favorably with the error in the 324 experimental values of $\log P_{oct}$.

Second, we calculate the values of *E*, *S*, *B* and *L* from the fragmentation scheme for the terpenes and use these to calculate the first ten values in Table 6 (these refer to the equations in *L*). For the ten sets of data the preferred descriptors from Table 4 yield AE = -0.12, AAE = 0.19 and SD = 0.22 log units. If we use the descriptors as calculated by the fragmentation scheme we find AE = 0.13, AAE = 0.20 and SD = 0.24 log units, not significantly different to the errors found with the preferred descriptors.

We suggest that this indicates that the fragmentation scheme in Table 3 can, indeed, be used reliably to estimate *E*, *S*, *B* and *L*. Knowledge of these descriptors, plus *V*, is enough to be able to predict log P_{oct} and a large number of other physicochemical and biological values for aliphatic cyclic terpene hydrocarbons.

335	References
336	
337	[1] C. S. Sell, The Chemistry of Fragrances, 2 nd Ed., RSC Publishing, Cambridge, 2006.
338	[2] S. Meakins, The safety and toxicology of fragrances, in C. S. Sell, Ed. The Chemistry of
339	Fragrances, 2 nd Ed., RSC Publishing, Cambridge, 2006.
340	[3] J. E. Cometto- Muñiz, W. S. Cain, M. H. Abraham, R. Kumarsingh, Pharmacol.
341	Biochem. Behavior, 60 (1998) 765-770
342	[4] Y. Nagata, Measurement of odor threshold by triangle odor bag method. In Odor
343	measurement review. Office of Odor, Noise and Vibration. Environmental Management
344	Bureau, Ministry of Environment, Tokyo, pp. 118-127, 2003
345	[5] O. Rodriguez, M. A. Teixeira, A. E. Rodrigues, Flavour and Fragr. J. (2011)
346	Wileyonlinelibrary.com, DOI 10.1002/ffj.2076
347	[6] M. H. Abraham, Chem. Soc. Revs., 22 (1993) 73-83.
348	[7] M. H. Abraham, A. Ibrahim, A. M. Zissimos, J. Chromatogr.A, 1037 (2004)
349	29-47.
350	[8] M. H. Abraham, R. E. Smith, R. Luchtefeld, A. J. Boorem, R. Luo, W. E. Acree,
351	Jr., J. Pharm. Sci., 99 (2010) 1500-1515.
352	[9] M. H. Abraham, J. C. McGowan, Chromatographia, 23 (1987) 243-246.
353	[10] M. H. Abraham, J. Andonian-Haftvan, G. S. Whiting, A. Leo, R. W. Taft,
354	J. Chem. Soc. Perkin Trans. 2, (1994) 1777-1791.
355	[11] M. H. Abraham, H. S. Chadha, G. S. Whiting, R. C. Mitchell, J. Pharm. Sci., 83 (1994)
356	1085-1100.
357	[12] H. Ahmed, C. F. Poole, J. Chromatogr. A., 1104 (2006) 82-90.
358	[13] I. G. Zenkevich and A. S. Kushakova, Russ. J. Gen. Chem., 81 (2011) 337-344.
359	[14] M. H. Abraham, R. Kumarsingh, J. E. Cometto- Muñiz, W. S. Cain, Archives Toxicol.,
360	72 (1998) 227-232.
361	[15] M. H. Abraham, J. M. R. Gola, J. E. Cometto- Muñiz, W. S. Cain, Indoor Built Environ.,
362	10 (2001) 252-257.
363	[16] M. H. Abraham, R. Sánchez-Moreno, J. E. Cometto-Muñiz, W. S. Cain, Chem.
364	Senses, 37 (2012) 207-218.
365	[17] M. H. Abraham, R. Kumarsingh, J. E. Cometto- Muñiz, W. S. Cain, M. Roses,
366	E. Bosch, M. L. Diaz, J. Chem. Soc., Perkin Trans.2, (1998) 2405-2411.
367	[18] T. Karunasekara. C. F. Poole, J. Chromatogr. A., 1235 (2012) 159-165.

368 [19] J. Li, E. M. Perdue, S.G.Paviostathis, R. Araujo, Environ. Int., 1998, 24 (1998),

- 369 353-358.
- 370 [20] ChemSpider, the EPI suite and the ACD suite.
- 371 [21] I. Fichan, C. Larroche, J. B. Gros, J. Chem. Eng. Data, 44 (1999) 56-62.
- 372 [22] J. L. Simonsen, The Terpenes, vol. I, second Ed., Cambridge University Press, 1947.
- 373 [23] J. Simonsen and L. N. Owen, The Terpenes, vol. II, second Ed., Cambridge University
 374 Press, 1949.
- 375 [24] J. Simonsen and D. H. R. Barton, The Terpenes, vol. III, Cambridge University Press,
 376 1952.
- 377 [25] www.cas.org/products/scifinder
- 378 [26] M. H. Abraham, J. C. McGowan, Chromatographia, 23 (1987) 243-246.
- 379 [27] V. I. Babushok, P. J. Linstrom, I. G. Zenkevich, J. Phys. Chem. Ref. Data, 40 (2011)
 380 043101-1 043101-46.
- 381 [28] B. B. Darjazi, A. Rustaiyan, A. Talaei, A. Khalighi, K. Larijani, B. Golein, E.
- Hayatbakhsh, R. Taghizad, Iran J. Chem. Chem. Eng., 28 (2009) 99-111.
- [29] M. H. Abraham, W. E. Acree, Jr., A. J. Leo, D. Hoekman, New J. Chem., 33 (2009)
 568-573.
- 385 [30] BioLoom, BioByte Corp, 201 W. 4th Street, #204 Claremont, CA 91711-4707,
 386 USA.
- 387 [31] www.vcclab.org/lab/alogps/
- 388 [32] archemcalc.com/sparc/pKa/
- 389 [33] S. Griffin, S. G. Wyllie, J. Markham, J. Chromatogr.A, 864 (1999) 221-228.
- 390 [34] M. H. Abraham, R. Sánchez-Moreno, J. Gil-Lostes, J. E. Cometto-Muñiz, W. S.Cain.
- 391 Physicochemical Modeling of Sensory Irritation in Humans and Experimental Animals,
- 392 Chapter 25, pp 376-389, in 'Toxicology of the Nose and Upper Airways, Ed. J. B.
- 393 Morris, D. J. Shusterman. Informa Healthcare USA, New York, 2010.
- 394 [35] A. Falk, E. Gullstrand, A. Lof, E. Wigaeus-Hjelm, Brit. J. Ind. Med. 47 (1990) 62-64.
- 395 [36] M. H. Abraham, A. Ibrahim, W. E. Acree, Jr., Chem. Res. Toxicol. 18 (2005) 904-911.
- 396 [37] M. H. Abraham, A. Ibrahim, J. Chem. Inf. Model. 46 (2006) 1735-1741.
- 397 [38] M. H. Abraham, W. E. Acree, Jr., Phys. Chem. Chem. Phys., 14 (2012) 7433-7440.
- 398

399 Table 1

System	С	e	S	a	b	v	Ref
Gas-water	-0.994	0.577	2.549	3.813	4.841	-0.869	10
Water-octan-1-ol ^a	0.088	0.562	-1.054	0.034	-3.460	3.814	11
Water-hexadecane	0.087	0.667	-1.617	-3.587	-4.869	4.433	11
Hexane-acetonitrile	0.097	0.189	-1.332	-1.649	-0.966	0.773	12
Perfluorodecalin-	-0.357	0.477	-2.360	-5.637	0.150	0.167	13 ^b
acetonitrile					(
System	С	е	S	a	b	l	
Gas-water	-1.271	0.822	2.743	3.904	4.814	-0.213	10
Gas-octan-1-ol ^a	-0.198	0.002	0.709	3.519	1.429	0.858	8
Gas-hexadecane	0.000	0.000	0.000	0.000	0.000	1.000	8

400 Coefficients in Eq. (1) and Eq. (2), at 25° C.

401 ^a Wet octan-1-ol. ^b Data from ref. 13, equation coefficients calculated in this work.

K CON

402 **Table 2**

403 Descriptors for some terpene hydrocarbons

.

م م م

Terpene	Ε	S	В	V	L	Ref
α-Pinene	0.446	0.14	0.12	1.2574	4.308	17
α-Pinene	0.441	0.19	0.22	1.2574	4.348	18
β-Pinene	0.530	0.24	0.19	1.2574	4.394	17
β-Pinene	0.525	0.23	0.20	1.2574	4.584	18
Limonene	0.488	0.28	0.21	1.3230	4.725	17
Limonene	0.497	0.34	0.17	1.3230	4.693	18

404

16

405 **Table 3**

406 Fragment schemes for the estimation of E, S, B and L for aliphatic cyclic terpene

407 hydrocarbons.

Fragment	E	S	В	L
Constant term	-0.0108	0.2194	-0.1183	-2.6575
Number of internal double bonds, C-C=C-C	0.1619	0.0575	0.1317	2.4783
Number of external double bonds, C=CH ₂	0.1704	0.0715	0.1136	2.4276
Number of -C=C-C=C- groups	-0.0091	-0.0449	-0.0368	-0.0668
Number of -C=C-C=C-C=C- groups	0.2089	0.0771	-0.0619	-2.0175
Total number of rings	0.2702	0.0001	0.1112	2.4445
Number of 3-membered rings	-0.1409	-0.0159	-0.0060	-0.1166
Number of 4-membered rings	-0.1696	-0.0547	-0.0101	-0.3862
Number of 5-membered rings	-0.1136	-0.0458	-0.0474	-0.1566
Number of 6-membered rings	-0.0486	-0.0430	-0.0080	-0.0158
Number of 7-membered rings	-0.0133	-0.0009	-0.0024	-0.0664
Number of 10-membered rings	0.0183	0.0393	-0.0818	-0.1531
Number of 11-membered rings	0.0228	-0.1219	-0.0680	-0.4200
Number of compounds	88	88	88	80
SD	0.046	0.005	0.024	0.1653

408

409 **Table 4**

410 Descriptors for terpene hydrocarbons

Terpene	MF	n(GLC)	Ε	S	В	V	L	log Kw
Santene	C9H14	5	0.459	0.19	0.17	1.1165	3.898	-0.39
1,3,8-p-Menthatriene	C10H14	3	0.914	0.44	0.30	1.2800	5.138	0.99
Thuja-2,4(10)-diene	C10H14	4	0.638	0.24	0.24	1.2144	4.448	0.09
Tricyclene	C10H16	15	0.500	0.11	0.17	1.1918	4.280	-0.64
α-Pinene	C10H16	65	0.438	0.20	0.14	1.2574	4.256	-0.64
β-Pinene	C10H16	57	0.515	0.19	0.15	1.2574	4.515	-0.57
Limonene	C10H16	84	0.501	0.31	0.23	1.3230	4.688	0.05
α-Terpinene	C10H16	40	0.526	0.25	0.23	1.3230	4.699	-0.09
β-Terpinene	C10H16	48	0.522	0.29	0.22	1.3230	4.869	-0.04
γ-Terpinene	C10H16	53	0.522	0.29	0.22	1.3230	4.840	0.02
Camphene	C10H16	36	0.520	0.20	0.17	1.2574	4.353	-0.45
α-Thujene	C10H16	40	0.376	0.21	0.20	1.2574	4.223	-0.37
Sabinene	C10H16	31	0.457	0.23	0.18	1.2574	4.483	-0.36
2-Carene	C10H16	4	0.489	0.22	0.14	1.2574	4.659	-0.57
Δ -3-Carene	C10H16	50	0.492	0.22	0.14	1.2574	4.679	-0.58
α-Phellandrene	C10H16	40	0.497	0.25	0.22	1.3230	4.643	-0.15
β-Phellandrene	C10H16	39	0.529	0.26	0.20	1.3230	4.759	-0.19
Terpinolene	C10H16	53	0.564	0.29	0.20	1.3230	5.029	-0.12
α-Fenchene	C10H16	24	0.486	0.19	0.19	1.2574	4.330	-0.38
β-Fenchene	C10H16	11	0.451	0.20	0.17	1.2574	4.276	-0.47
Pinane, Z, cis	C10H18	2	0.421	0.12	0.13	1.3004	4.534	-0.95
Pinane, E, trans	C10H18	2	0.421	0.12	0.13	1.3004	4.487	-0.95
Thujane	C10H18	4	0.304	0.16	0.14	1.3004	4.480	-0.83
β-Acoradiene	C15H24	3	0.802	0.26	0.28	1.9189	6.993	-0.18
Aromadendrene	C15H24	16	0.700	0.23	0.29	1.8533	6.839	-0.21
Alloaromadendrene	C15H24	17	0.722	0.23	0.29	1.8533	6.973	-0.20
α-Amorphene	C15H24	4	0.821	0.25	0.36	1.9189	7.074	0.19
α-Bisabolene, E,trans	C15H24	3	0.668	0.35	0.39	1.9845	7.273	0.45
α-Bisabolene, Z, cis	C15H24	5	0.668	0.35	0.39	1.9845	7.069	0.45
β-Bisabolene	C15H24	3	0.656	0.36	0.37	1.9845	7.044	0.37
γ-Bisabolene, E, trans	C15H24	3	0.668	0.35	0.39	1.9845	7.166	0.45
γ-Bisabolene, Z, cis	C15H24	4	0.668	0.35	0.39	1.9845	7.122	0.45
β-Bourbonene	C15H24	19	0.627	0.14	0.20	1.8533	6.645	-0.87
Bicyclogermacrene	C15H24	9	0.800	0.29	0.38	1.9189	7.105	0.38
α -Bergamotene, E, trans	C15H24	13	0.695	0.28	0.38	1.9189	6.711	0.29
α -Bergamotene, Z, cis	C15H24	5	0.695	0.28	0.38	1.9189	6.608	0.29
Cadina-1(2),4-diene, Z, cis	C15H24	7	0.823	0.25	0.36	1.9189	7.358	0.19
α-Cadinene	C15H24	13	0.809	0.29	0.36	1.9189	7.307	0.29
β-Cadinene	C15H24	1	0.791	0.29	0.36	1.9189	6.950	0.28
δ-Cadinene	C15H24	36	0.817	0.29	0.36	1.9189	7.220	0.29
γ-Cadinene	C15H24	23	0.811	0.31	0.34	1.9189	7.162	0.30
α-Caryophyllene	C15H24	13	0.772	0.27	0.32	1.9845	6.708	-0.04
β-Caryophyllene, E, trans	C15H24	31	0.724	0.33	0.26	1.9189	6.613	-0.14
β-Caryophyllene, Z, cis	C15H24	9	0.724	0.33	0.26	1.9189	6.617	-0.14
g-Caryophyllene	C15H24	1	0.699	0.33	0.26	1.9189	6.529	-0.16
α-Cedrene	C15H24	12	0.711	0.18	0.24	1.8533	6.764	-0.57

β-Cedrene	C15H24	7	0.757	0.20	0.22	1.8533	6.785	-0.59
α-Copaene	C15H24	16	0.624	0.14	0.32	1.8533	6.567	-0.34
β-Copaene	C15H24	9	0.624	0.15	0.30	1.8533	6.849	-0.41
α-Cubebene	C15H24	25	0.616	0.17	0.29	1.8533	6.320	-0.41
β-Cubebene	C15H24	19	0.616	0.19	0.27	1.8533	6.624	-0.46
Cyclosativene	C15H24	6	0.613	0.08	0.24	1.7877	6.659	-0.83
Cyperene	C15H24	4	0.720	0.18	0.24	1.8533	6.637	-0.57
β-Elemene	C15H24	20	0.714	0.40	0.33	1.9845	6.411	0.31
δ-Elemene	C15H24	11	0.714	0.38	0.34	1.9845	6.163	0.31
g-Elemene	C15H24	7	0.714	0.38	0.33	1.9845	6.690	0.26
Germacrene A	C15H24	9	0.764	0.45	0.29	1.9845	6.965	0.27
Germacrene B	C15H24	11	0.764	0.45	0.29	1.9845	7.189	0.27
Germacrene D	C15H24	25	0.764	0.40	0.24	1.9845	6.869	-0.10
α-Guaiene	C15H24	6	0.695	0.30	0.31	1.9189	6.821	0.01
β-Guaiene, E, trans	C15H24	3	0.695	0.30	0.31	1.9189	7.047	0.01
β-Guaeine, Z, cis	C15H24	4	0.695	0.30	0.31	1.9189	6.972	0.01
α-Gurjunene	C15H24	10	0.737	0.21	0.28	1.8533	6.659	-0.29
β-Gurjunene	C15H24	7	0.775	0.23	0.26	1.8533	6.792	-0.31
y-Gurjunene	C15H26	6	0.734	0.30	0.29	1.9619	6.950	-0.11
α-Humulene	C15H24	38	0.764	0.27	0.32	1.9845	6.896	-0.04
Isolongifolene	C15H24	3	0.728	0.14	0.27	1.8533	6.647	-0.52
Longicyclene	C15H24	6	0.656	0.11	0.22	1.7877	6.600	-0.82
Longifolene	C15H24	8	0.757	0.20	0.22	1.8533	6.677	-0.59
α-Longipinene	C15H24	8	0.665	0.18	0.32	1.8533	6.464	-0.21
α-Muurolene	C15H24	19	0.802	0.25	0.36	1.9189	7.138	0.18
y-Muurolene	C15H24	22	0.802	0.26	0.34	1.9189	7.062	0.11
α-Patchoulene	C15H24	6	0.720	0.18	0.24	1.8533	6.999	-0.57
β-Patchoulene	C15H24	4	0.720	0.18	0.24	1.8533	6.518	-0.57
γ-Patchoulene	C15H24	5	0.720	0.22	0.20	1.8533	7.003	-0.61
α-Santalene	C15H24	5	0.635	0.17	0.29	1.8533	6.801	-0.40
β-Santalene	C15H24	4	0.668	0.26	0.28	1.9189	6.902	-0.26
α-Selinene	C15H24	4	0.823	0.26	0.34	1.9189	6.895	0.12
β-Selinene	C15H24	20	0.765	0.28	0.32	1.9189	7.067	0.04
7-epi-α-Selinene	C15H24	4	0.823	0.26	0.34	1.9189	7.266	0.12
Selena-4,11-diene	C15H24	4	0.823	0.26	0.34	1.9189	6.895	0.12
Selena-3,7(11)-diene	C15H24	6	0.823	0.25	0.36	1.9189	7.416	0.19
Thujopsene	C15H24	10	0.742	0.17	0.29	1.8533	6.898	-0.34
Valencene	C15H24	11	0.807	0.26	0.34	1.9189	7.145	0.11
α-Ylangene	C15H24	16	0.640	0.13	0.33	1.8533	6.557	-0.31
β-Ylangene	C15H24	3	0.640	0.15	0.31	1.8533	6.835	-0.35
α-Zingberene	C15H24	5	0.680	0.30	0.34	1.9845	7.053	0.08
Abietadiene	C20H32	2	0.950	0.20	0.45	2.5148	9.991	0.06
Murcene	C10H16	10	0 192	0.20	0 22	1 3896	1 512	0.27
a-Ocimene E trans	C10H16	40	0.405	0.29	0.32	1 3886	<u>л.313</u> <u>Д</u> 707	0.37
β_{-} Ocimene Z cis	C10H16	42	0.559	0.31	0.31	1.3000	1.121	0.40
trans_Allo_ogimene	C10H16	0	0.501	0.29	0.31	1.3000	5 170	0.37
cis_Allo_ocimene	C10H16	7 10	0.394	0.39	0.20	1 3886	5 1 8 0	0.39
a-Farnesene (F F)	C15H24	10	0.000	0.39	0.20	2 0501	7 030	0.40
α -Farnesene (7 E)	C15H24	10	0.711	0.30	0.45	2.0301	6 956	0.78
B-Farnesene trans F	C15H24	12	0.711	0.30	0.45	2.0301	6 760	0.78
p-ramesene, uans, E	0131124	12	0./11	0.37	0.40	2.0301	0.709	0.00

β-Farnesene, cis, Z	C15H24	4	0.711	0.36	0.46	2.0501	6.756	0.78
Phyt-1-ene	C20H40	1	0.010	0.10	0.10	2.8836	8.517	-2.75
Phyt-2-ene	C20H40	4	0.050	0.10	0.10	2.8836	8.842	-2.73
Phytane	C20H42	1	0.000	0.00	0.00	2.9266	8.727	-3.54
Cadalene	C15H18	4	1.423	0.92	0.25	1.7899	7.674	1.83
α-Calacorene	C15H20	13	0.930	0.65	0.25	1.8329	7.079	0.82
β-Calacorene	C15H20	5	0.930	0.64	0.25	1.8329	7.174	0.79
Calamenene, E, trans	C15H22	16	0.860	0.56	0.20	1.8759	6.932	0.27
Calamenene, Z, cis	C15H22	13	0.860	0.56	0.20	1.8759	6.975	0.27
Cuparene	C15H22	8	0.770	0.54	0.25	1.8759	6.934	0.45
Abietatriene	C20H30	3	0.950	0.65	0.25	2.4718	9.593	0.27

411

412

-92

412 **Table 5**

413 Comparison of log P_{oct} calculated from the descriptors in Table 4 (taken) and log P_{oct}

414 calculated from literature software.

	Taken	ClogP	ACD	EPI	ALOGPS	SPARC	Experimental
Santene	3.82	4.18	3.82	3.94	3.35	4.03	
1,3,8-p-Menthatriene	3.98	3.96	4.03	4.74	4.15	4.34	
Thuja-2,4(10)-diene	3.99	3.61	3.60	4.47	3.50	5.38	X
Tricyclene	4.21	4.38	3.98	4.13	3.43	5.30	
α-Pinene	4.48	4.18	4.31	4.27	3.66	4.72	4.44[20], 4.48 [30], 4.83 [20]
β-Pinene	4.45	4.70	4.24	4.35	3.94	4.84	
Limonene	4.30	4.35	4.55	4.83	4.50	4.71	4.38 [30], 4.50 [20], 4.57 [19]
α-Terpinene	4.35	4.41	4.29	4.75	4.51	4.65	4.25 [30]
β-Terpinene	4.36	4.35	4.25	4.75	4.30	4.70	4.35 [30], 4.50 [20]
γ-Terpinene	4.35	4.35	4.25	4.75	4.36	4.59	4.35 [30], 4.50 [20]
Camphene	4.38	4.70	4.24	4.35	4.56	4.80	4.22 [30]. 4.37 [30]
α-Thujene	4.18	4.19	4.02	4.61	4.07	5.11	
Sabinene	4.27	4.03	3.94	4.69	3.04	5.22	
2-Carene	4.44	4.44	4.32	4.61	3.26	5.05	4.44 [20]
D-3-Carene	4.44	4.44	4.32	4.61	3.64	5.05	4.38 [20, 30]
α-Phellandrene	4.39	4.41	4.41	4.62	4.29	4.79	
β-Phellandrene	4.46	4.41	4.39	4.70	3.98	4.91	4.16 [30]
Terpinolene	4.47	4.35	4.21	4.88	3.82	4.69	4.24 [30], 4.47 [20]
α-Fenchene	4.30	4.70	4.24	4.35	4.05	4.88	
β-Fenchene	4.34	4.70	4.19	4.17	4.09	4.84	
Pinane, Z, cis	4.71	5.19	4.69	4.35	4.04	5.11	
Pinane, E, trans	4.73	5.19	4.69	4.35	4.04	5.11	
Thujane	4.57	4.52	4.39	4.70	3.63	5.44	
β-Acoradiene	6.61	6.30	6.56	6.99	5.87	7.10	
Aromadendrene	6.30	6.39	6.41	6.13	3.70	7.55	
Alloaromadendrene	6.32	6.39	6.43	6.13	3.70	7.55	
α-Amorphene	6.36	6.30	6.56	6.19	5.00	7.25	
α-Bisabolene, E,trans	6.31	6.38	6.61	7.05	5.95	7.09	
α-Bisabolene, Z, cis	6.31	6.38	6.61	7.05	5.95	7.09	
β-Bisabolene	6.37	6.38	6.43	7.12	6.01	7.18	
γ-Bisabolene, E, trans	6.31	6.38	6.70	7.18	5.88	7.17	
γ-Bisabolene, Z, cis	6.31	6.38	6.70	7.18	5.88	7.17	
β-Bourbonene	6.67	5.98	6.13	5.44	3.47	7.44	
Bicyclogermacrene	6.24	6.45	6.50	6.22	6.10	7.07	
α -Bergamotene, E, trans	6.19	6.45	6.54	6.57	6.17	7.13	
α-Bergamotene, Z, cis	6.19	6.45	7.03	6.57	6.17	7.13	
Cadina-1(2),4-diene, Z, cis	6.36	6.30	6.34	6.19	5.33	7.13	
α-Cadinene	6.31	6.30	6.56	6.19	5.00	7.25	
β-Cadinene	6.30	6.30	6.43	6.19	4.92	7.16	
δ-Cadinene	6.32	6.30	6.27	6.32	4.92	7.16	
γ-Cadinene	6.36	6.30	6.35	6.27	5.23	7.34	
α-Caryophyllene	6.70	5.74	6.59	6.95	6.07	6.78	
β-Caryophyllene, E, trans	6.57	6.45	6.78	6.30	5.35	6.97	
β -Caryophyllene, Z, cis	6.57	6.45	6.78	6.30	5.35	6.97	
g-Caryophyllene	6.55	6.45	6.42	6.30	5.35	6.97	
α-Cedrene	6.54	6.91	6.38	6.43	5.18	7.64	
	1	1	1		-	1	1

β-Cedrene	6.61	6.91	6.23	6.51	4.38	7.74	
α-Copaene	6.25	7.02	6.21	5.36	3.75	7.15	
β-Copaene	6.31	7.02	6.13	5.44	3.38	7.25	
α-Cubebene	6.32	6.50	6.26	6.73	3.49	7.65	
β-Cubebene	6.37	6.50	6.18	6.81	3.49	7.73	
Cyclosativene	5.72	6.70	5.84	5.57	5.33	7.21	
Cyperene	6.54	6.91	6.19	6.21	5.51	7.60	
β-Elemene	6.49	6.36	5.77	7.04	5.40	7.26	
δ-Elemene	6.48	6.36	6.59	6.96	5.39	7.17	
g-Elemene	6.52	6.36	6.49	7.09	5.23	7.27	
Germacrene A	6.61	6.62	6.86	7.12	6.02	6.81	
Germacrene B	6.61	6.62	6.39	7.18	5.87	6.79	
Germacrene D	6.83	6.68	6.57	6.99	5.64	7.00	
α-Guaiene	6.41	6.30	6.60	6.51	5.80	7.04	
β-Guaiene, E, trans	6.41	6.30	6.58	6.51	5.80	7.04	
β-Guaeine, Z, cis	6.41	6.30	6.33	6.56	5.52	6.97	
α-Gurjunene	6.38	5.87	6.45	6.18	4.42	7.31	
β-Gurjunene	6.45	6.39	6.43	6.13	3.75	7.59	
y-Gurjunene	6.66	6.79	6.80	6.40	5.62	7.10	
α-Humulene	6.69	6.77	6.59	6.95	6.07	6.78	
Isolongifolene	6.48	6.91	6.15	6.12	6.04	7.61	
Longicvclene	6.40	6.85	5.91	5.60	5.64	8.09	
Longifolene	6.61	7.17	6.17	5.48	4.65	7.26	
α-Longipinene	6.13	7.17	6.40	5.40	4.45	7.17	
α-Muurolene	6 35	6 30	6 44	6 19	5.00	7 25	
v-Muurolene	6.41	6.30	6.54	6.27	4.48	7.34	
α-Patchoulene	6.54	6.91	6.31	6.43	5.72	7.63	
B-Patchoulene	6 54	6.65	6.13	5.87	5.68	7 54	
v-Patchoulene	6.59	6.91	6 37	5 48	5 35	7 72	
α-Santalene	6.33	6.91	6.02	6.43	4 96	8.02	
B-Santalene	6 54	6.73	6.15	6.64	5 56	7.28	
α-Selinene	6.42	6 30	6.41	6.30	5.86	7.20	
ß-Selinene	6.43	6.30	6.33	6.38	4 97	7.23	
7-eni-a-Selinene	6.42	6.30	6.41	6.30	5.86	7.25	
Selena-4 11-diene	6.42	6.30	6.63	6.43	5.60	7.23	
Selena_3 7(11)-diene	6.36	6.30	6.73	6 35	5.01	7.10	
Thuionsene	6 39	6.65	6.05	6.12	5.70	7.24	
Valencene	6.41	6.30	6.29	6.30	5.86	7.43	
a Vlangene	6.24	7.02	6.21	5.36	3.88	7.21	
B Vlangene	6.24	7.02	6.13	5.30	3.88	7.13	
p-Tialigene	6.55	6.44	6.60	6.02	5.38	7.23	
Abietediene	0.33	0.44	0.00	0.92	7.79	0.72	
Abletadiene	8.44	8.23	0.42		1.28	9.75	
Muraana	1 17	4.22	1 25	1 00	4 22	102	4 17 [20]
Myrcene	4.17	4.33	4.25	4.88	4.52	4.83	4.17 [30]
a-Ocimene, E, trans	4.1/	4.55	4.14	4.88	4.25	4.82	4.1/[20]
p-Ocimene, Z, cis	4.55	4.33	4.20	4.80	4.25	4.82	4.57 [20]
trans-Allo-ocimene	4.40	4.36	4.39	4.72	4.36	4.98	4.36 [20]
cis-Allo-ocimene	4.41	4.36	4.39	4.72	4.36	4.98	
α-Farnesene, (E, E)	6.35	6.36	6.30	7.10	5.70	7.18	
α -Farnesene, (Z, E)	6.35	6.36	6.14	7.17	5.70	7.18	
β-Farnesene, trans, E	6.32	6.36	6.14	7.17	5.70	7.27	
β-Farnesene, cis, Z	6.34	6.36	6.14	7.17	5.70	7.27	

Phyt-1-ene	10.64	10.27	9.87	9.73	8.82	11.38	
Phyt-2-ene	10.66	10.27	9.93	9.78	8.76	11.39	
Phytane	11.25	10.75	9.87		9.09	11.59	
Cadalene	5.88	5.74	5.70	5.72	5.70	5.60	
α-Calacorene	6.05	5.88	6.22	6.16	5.47	5.54	
β-Calacorene	6.06	5.88	6.05	6.24	5.45	5.68	
Calamenene, E, trans	6.44	6.18	6.02	6.25	5.60	5.63	
Calamenene, Z, cis	6.44	6.18	6.02	6.25	5.60	5.63	
Cuparene	6.24	6.26	5.89	6.19	6.06	5.74	
Abietatriene	8.50	8.13	8.20	7.76	7.07	7.55	

415

416

Table 6 416

- Comparison of predicted and experimental values of $\log (1/NPT)$ and of $\log K$ for some gas-417
- solvent partitions at 37°C 418

Terpene	Pred ^a	Pred ^b	Obs
log (1/NPT)			
Δ -3-Carene	-3.49	-3.50	-3.21
α-Terpinene	-3.35	-3.34	-3.30
log K(gas-blood)			
a-Pinene	1.31	1.52	1.18 ± 0.11
β-Pinene	1.45	1.47	1.36 ± 0.11
Δ -3-Carene	1.51	1.68	1.51 ± 0.09
Limonene	1.85	1.86	1.62 ± 0.08
log K(gas-oil)			
a-Pinene	3.62	3.64	3.46 ± 0.07
β-Pinene	3.82	3.60	3.63 ± 0.07
Δ -3-Carene	3.99	3.90	3.70 ± 0.10
Limonene	4.08	4.05	3.76 ± 0.11
log <i>K</i> (gas-water)		U.	
a-Pinene	-0.87	-0.55	-0.92 ± 1.30
β-Pinene	-0.81	-0.59	-0.92 ± 0.98
Δ -3-Carene	-0.79	-0.42	-0.39 ± 0.61
Limonene	-0.22	-0.20	0.20 ± 0.36

scheme, Table 3, see later. 420

421

^a Using the descriptors in Table 4. ^b Using descriptors calculated by the fragmentation 419

24

422 **Table 7**

423 Estimation of descriptors through coefficients in Table 3, and derived estimated values of log

424 Poct

Descriptors	MF	n(L)	Ε	S	В	V	L	log Kw
Verbenene	C10H14	3	0.635	0.25	0.29	1.2144	4.467	0.36
α-Bulnesene	C15H24	2	0.735	0.30	0.30	1.9189	7.107	-0.02
β-Sesquiphellandrene	C15H24	3	0.696	0.36	0.33	1.9845	7.161	0.20
Log Poct	Pred	ClogP	ACD	EPI	ALOGP	SPARC		log(1/NPT)
Verbenene	3.81	3.76	3.86	4.13	3.64	5.01		-3.49
α-Bulnesene	6.47	6.30	6.73	6.27	n/a	7.11		-1.25
β-Sesquiphellandrene	6.53	6.44	6.52	6.99	6.36	7.33		-1.08

425

426 **Table 8**

Terpene	Ε	S	В	V	Calc	Exp ^b	Av exp
α-Pinene	0.473	0.18	0.22	1.2547	4.20	4.44, 4.48, 4.83	4.58
Limonene	0.543	0.31	0.23	1.3230	4.32	4.38, 4.50, 4.57	4.48
α-Terpinene	0.526	0.29	0.21	1.3230	4.39	4.25	4.25
β-Terpinene	0.543	0.31	0.23	1.3230	4.32	4.35, 4.50	4.43
γ-Terpinene	0.535	0.29	0.25	1.3230	4.27	4.35, 4.50	4.43
Camphene	0.538	0.20	0.16	1.2574	4.41	4.22, 4.37	4.30
2-Carene	0.502	0.22	0.22	1.2574	4.17	4.44	4.44
D-3-Carene	0.502	0.22	0.22	1.2574	4.17	4.38	4.38
β-Phellandrene	0.534	0.30	0.19	1.3230	4.45	4.16	4.16
Terpinolene	0.535	0.29	0.25	1.3230	4.27	4.24, 4.47	4.35

427 Calculation of $\log P_{oct}$ through the fragmentation scheme in Table 3^a

Cox -

428 ^a Values of E, S and B obtained from the fragmentation scheme in Table 3. V calculated from

430