

# O ORALOMA DA DIABETES MELITOS TIPO 1 VS DIABETES MELITOS TIPO 2 - UM ESTUDO COMPARATIVO

Dissertação apresentada à Universidade Católica Portuguesa para obtenção do grau de Mestre em Medicina Dentária

Por

Vítor Daniel Moreira Brás

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Sob a orientação do Professor Doutor Nuno Rosa

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-The empires of the future are the empires of the mind-Sir Winston Churchill

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(\*CONSTANT SITE - FACE MESIO-VESTIBULAR DO PRIMEIRO MOLAR SUPERIOR

DIREITO; DEEPEST SITE - LOCAL MAIS PROFUNDO À SONDAGEM)

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### Lista de abreviaturas

**DMT - Diabetes Melitos** 

EGF - Epidermal growth factor

**GLUT** - Glucose transporter type

IFN - Interferon

IL - Interleukin

IRS - Insulin receptor substrate

TNF - Tumour necrosis factor

Resumo

A Diabetes Melitos é uma doença de grande prevalência a nível

mundial e os seus mecanismos fisiopatológicos não são ainda totalmente

conhecidos.

Actualmente, a terapêutica seguida em casos de Diabetes Melitos é

eminentemente sintomática consistindo na administração de insulina nos

casos de Diabetes Melitos tipo 1 ou quando se verifica a falência das células

beta do pâncreas de pacientes com Diabetes Melitos tipo 2, ou no controlo da

glicemia nos casos de Diabetes Melitos tipo 2.

A hiperglicemia, fenótipo característico desta patologia, produz efeitos

negativos no organismo a longo prazo, levando à modificação estrutural e

funcional de vários proteínas, afectando, deste modo, diversos processos

moleculares em que intervêm estas proteínas.

Os dois tipos Diabetes Melitos influenciam, de forma diferenciada, a

expressão de diversas proteínas, produzindo efeitos ao nível das vias de

sinalização celular e nos processos biológicos celulares.

Consequentemente, conhecer as características moleculares dos dois

tipos de Diabetes Melitos permite o desenvolvimento de técnicas diagnósticas

e terapêuticas mais precoces e eficazes específicas para cada uma delas.

Tendo como base o proteoma da cavidade oral, denominado Oraloma,

este estudo pretende actualizar os dados provenientes de estudos de

proteómica da cavidade oral de pacientes com os dois tipos de Diabetes

Melitos e estudá-los com recurso a ferramentas bioinformáticas que permitem

a caracterização das proteínas indexadas.

Dado a cavidade oral ser um ecossistema, é indissociável dos

microrganismos que nele existem. Consequentemente, é objectivo deste

estudo anotar as estirpes de microrganismos identificados pela literatura

actual, por forma a caracterizar de forma mais completa este aspecto

molecular da cavidade oral.

Palavras-chave: diabetes, oraloma, proteómica, bioinformática

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**Abstract** 

Diabetes Mellitus is a disease with high prevalence worldwide with

pathophysiological mechanisms not yet fully understood.

Nowadays, the therapeutical approach of Diabetes Mellitus is mainly a

symptomatic one based on insulin administration to type 1 Diabetes Mellitus

patients and type 2 Diabetes Mellitus patients with no funtional pancreatic

beta-cells or the serum glucose concentration control in type 2 Diabetes

Mellitus patients.

Hyperglykemia is a carachteristic phenotype of this pathology, it brings

long-run negative effects on the organism due to its role on modifying proteins

structure and function affecting the organism's integrity.

Both Diabetes Mellitus types influence, on different ways, protein

expression hence leading to changes on the molecular signaling pathways

and biologic process of the cells.

Thereafter, to know the molecular aspects of both Diabetes Mellitus

types is key to develop better, more efficient and precocious diagnostic and

therapeutic tecniques.

Based on the oral proteome, namely the oralome, this study aims to

update data from the pre-existing oral proteomic studies on both types of

Diabetes Mellitus and to study them using bioinformatic tools which allow

these proteins carachterization.

Being an ecossystem, the oral cavity is inseparable of the

microrganisms which inhabit in it. Therefore, another aim of the present study

is to annotate the microrganisms species identified by the current scientific

literature, in order to charactherize in a more complete way this molecular

aspect of the oral cavity.

**Keywords:** Diabetes, oralome, proteomics, bioinformatics

ΧI

### 1 Introdução

### 1.1 Diabetes Melitos

De acordo com a Organização Mundial de Saúde, a Diabetes Melitos é uma patologia crónica que ocorre quando o pâncreas não produz insulina suficiente para suprir as necessidades do organismo ou quando, apesar de o pâncreas produzir insulina, o organismo não consegue utilizar eficazmente a insulina produzida. Como corolário desta definição, é correto afirmar que a Diabetes Melitos é um grupo de distúrbios metabólicos que compartilham o fenótipo de hiperglicemia.

A principal causa de morte em pacientes diabéticos são complicações das artérias coronárias, geralmente o enfarte do miocárdio<sup>1</sup>.

### 1.2 Epidemiologia da Diabetes Melitos

A prevalência desta patologia atingiu proporções epidémicas, traduzindo-se, segundo estimativa da Organização Mundial de Saúde em 2008, em cerca de 200 milhões de indivíduos afetados. Se se mantiverem os padrões atuais, estima-se que até 2050 hajam 500 000 000 de indivíduos diabéticos.

Existem dois tipos mais prevalentes de Diabetes Melitos, o tipo 1 e o tipo 2.

A Diabetes Melitos tipo 2 é a mais prevalente, consistindo em cerca de 85 a 90% de todos os casos de Diabetes Melitos; sendo que a Diabetes Melitos tipo 1, uma doença autoimune em que são produzidos anticorpos para a enzima GAD (*glutamic acid decarboxylase*) acabando por atacar e destruir as células β pancreáticas produtoras de insulina, é menos comum.

Apesar de haver uma componente genética na incidência de Diabetes Melitos tanto de tipo 1 como de tipo 2, a Diabetes Melitos tipo 2 é influenciada, em cerca de 10% de indivíduos diabéticos deste tipo, por fatores individuais de risco tais como a obesidade e o estilo de vida sedentário.

A prevalência de Diabetes Melitos a nível mundial é crescente, mais marcada em países em desenvolvimento<sup>2,3</sup>.

### 1.2.1 Patogénese da Diabetes Melitos tipo 1

Uma redução no número de células  $\beta$  provoca um défice quase absoluto de insulina. Este tipo de Diabetes Melitos desenvolve-se normalmente na infância, tornando-se mais severo na puberdade<sup>4</sup>.

A Diabetes Melitos tipo 1 causa grandes distúrbios metabólicos e fisiológicos. O défice de insulina não permite a entrada de glucose do sangue para a célula, isto provoca um estado de inanição metabólica.<sup>4,5</sup> Consequentemente a baixa concentração de insulina sanguínea aliada a uma alta concentração de glucagão e epinefrina culminam na lipólise de triglicerídeos do tecido adiposo<sup>5</sup>. Os ácidos gordos libertados são transportados na corrente sanguínea pela albumina e sofrem β-oxidação a nível intracelular. Os resultados desta reacção são a acetil-coenzima A e enzimas reduzidas (NADH e FADH2), utilizadas no ciclo TCA e na fosforilação oxidativa nos tecidos. A quantidade de acetil-coenzima A produzida a partir de ácidos gordos é superior à que pode ser metabolizada, o excedente não-metabolizado é transformado em corpos cetónicos (acetoacetato, hidroxibutirato e acetona). A continuada formação de corpos cetónicos produz um estado de cetoacidose (acidose metabólica), que por sua vez provoca alteração no balanço electrolítico do organismo (diurese osmótica) levando o indivíduo a excretar volumes significativos de urina, devido à presença de glucose no fluido glomerular renal. Esta patologia pode ser fatal se não tratada, consistindo o tratamento na administração de insulina 4,5.

Existe também uma susceptibilidade genética na Diabetes Melitos tipo1, cerca de 95% dos pacientes brancos com Diabetes tipo 1 possuem tanto o alelo HLA-DR3 ou HLA-DR4 ou mesmo ambos<sup>4</sup>. É sugerido que alterações na molécula HLA classe II pode produzir efeitos ao nível do reconhecimento nos receptores de células T e modificar também a apresentação de antigénios.

Ainda que não estejam definidos quais, existe sugestão da possibilidade de certos factores ambientais despoletarem os fenómenos de autoimunidade da Diabetes tipo 1, tais como vírus, retrovirus e toxinas <sup>4</sup>.

### 1.2.2 Patogénese da Diabetes Melitos tipo 2

Na patogénese da Diabetes Melitos tipo 2, não existe evidência de fenómenos autoimunes como na Diabetes Melitos tipo 1, embora o estilo de vida e a predisposição genética sejam mais preponderantes no desenvolvimento da doença.

Na Diabetes Melitos tipo 2 existem dois defeitos metabólicos característicos: diminuição da secreção de insulina pelas células  $\beta$  e na resistência dos tecidos à insulina.

No início da sua patogénese, não se verifica qualquer diminuição na secreção de insulina, contudo o padrão pulsátil e oscilatório da sua secreção é perdido. Isto demonstra alterações ao nível da resposta das células  $\beta$  à hiperglicemia, ainda não ocorrendo modificações na síntese de insulina.

A contínua exposição a elevados níveis de glucose e de ácidos gordos livres leva a uma hipersensibilização das células  $\beta$  do pâncreas culminando na sua apoptose por consequência do stress causado pelos ácidos gordos livres ao nível do retículo endoplasmático (processo que depende de NF- $\kappa$ B e Óxido nítrico) $^6$ .

A resistência à insulina resulta de dois fatores, diminuição do número de receptores de insulina na membrana celular e a cascata de sinalização intracelular após ligação da insulina no receptor estar comprometida<sup>4</sup>.

### 1.2.3 Patogénese das complicações da Diabetes Melitos

Ambos os tipos de Diabetes Melitos possuem as mesmas morbilidades, nomeadamente microangiopatia, retinopatia, nefropatia e neuropatia. Á luz da literatura actual, estas complicações advêm da hiperglicemia. Quando a hiperglicemia está controlada, estas complicações são menos frequentes.

Duas razões principais estão na génese destas complicações em pacientes diabéticos, a glicosilação não-enzimática e a hiperglicemia intracelular e os seus efeitos na via de sinalização dos polióis.

No que concerne a glicosilação não enzimática, esta define-se como a ligação química reversível que ocorre entre a glucose e o grupo amina de

uma proteína sem o auxílio de enzimas. Os produtos de reacção da glicosilação não-enzimática envolvem proteínas como o glicogénio e outras da matriz extracelular que posteriormente sofrerão modificações estruturais tornando a ligação proteína-glicose irreversível. Quando a glicosilação da proteína se torna irreversível origina os denominados produtos avançados de glicosilação ou AGE's (*advanced glycosilation end-produts*) que dado as suas características bioquímicas (a adição de açúcar à sua estrutura altera-a e com isso a sua função) são potencialmente patogénicos<sup>7</sup>.

Os AGE's ocorrem não apenas com proteínas mas também com lípidos e ácidos nucleicos<sup>7</sup>. Quando ocorrem com proteínas como o glicogénio, formam ligações cruzadas entre os polipéptidos da molécula de glicogénio e no processo aprisionam proteínas plasmáticas ou intersticiais<sup>7</sup>. Quando tal ocorre em grandes vasos sanguíneos, por exemplo, aprisionam lipoproteínas de baixa densidade que leva à deposição de colesterol promovendo a aterogénese<sup>7</sup>. Em capilares como os glomerulares renais, a albumina liga-se à membrana basal glicosilada provocando o seu espessamento, característico em microangiopatia diabética<sup>7</sup>. Como os AGE's resistem à digestão proteolítica, dificultam a remoção de proteínas e as ligações cruzadas que formam aumentam a deposição de proteínas<sup>7</sup>. De facto, a ligação cruzada provocada pelos AGE's entre as proteínas da matriz extracelular impede a interacção destas proteínas com outros componentes da matriz extracelular provocando defeitos funcionais e estruturais<sup>7</sup>.

Os AGE's possuem a capacidade de se ligar aos receptores de vários tipos de células tais como:

- células endoteliais,
- monócitos
- linfócitos,
- macrófagos
- células mesangiais.

Esta ligação provoca efeitos biológicos dependendo do tipo de célula. Nos monócitos promove a sua migração, nos macrófagos a secreção de factores de crescimento e citocinas. Também aumentam a permeabilidade das células endoteliais, aumentam a actividade procoagulante de macrófagos

e células endoteliais e provocam o aumento de produção de matriz extracelular por parte dos fibroblastos e do músculo liso.

A hiperglicemia intracelular ocorre em tecidos que não necessitam de insulina para captar glicose do meio extracelular, como tecidos nervosos, tecidos da córnea, capilares, rins. Em situações de hiperglicemia, grandes quantidades de glicose entram para o meio intracelular e são modificados enzimaticamente formando sorbitol e posteriormente frutose. Os elevados níveis de sorbitol e frutose intracelular promovem um influxo aumentado de água levando à turgescência celular provocando lesões celulares osmóticas. Em tecidos como os da córnea, esta turgescência provoca opacidade e um aumento de volume. Os níveis intracelulares elevados de sorbitol comprometem a função das bombas de iões lesando as células de *Schwann* e os perícitos dos capilares da retina, resultando em neuropatia periférica e em microaneurismas da retina. O tratamento com inibidores da enzima que modifica a glicose em sorbitol, a aldose redutase, parece inibir o desenvolvimento de neuropatias e cataratas<sup>4</sup>.

### 1.2.4 Sintomatologia da Diabetes Melitos

A sintomatologia da Diabetes Melitos é semelhante para os dois tipos, contudo no caso da Diabetes Melitos tipo 1 os sintomas podem surgir subitamente ao passo que na Diabetes Melitos tipo 2 podem aparecer mais tardiamente sendo o diagnóstico efectuado anos após o *onset*.

Os sintomas desta patologia podem compreender alterações na visão, poliúria, polidipsia, fome, perda de peso e fadiga<sup>1</sup>.

#### 1.2.5 Tratamento da Diabetes Melitos

A administração de insulina é o tratamento preferencial em casos de Diabetes Melitos tipo 1. Está clinicamente comprovado que o controlo dos níveis séricos de glicose em pacientes com Diabetes Melitos tipo 1 reduz a incidência das principais complicações desta patologia, tais como a retinopatia, nefropatia e a neuropatia<sup>8</sup>.

No que concerne o tratamento da Diabetes Melitos tipo 2 esta consiste fundamentalmente no controlo da dieta e no exercício físico, não sendo tão

complexa como nos casos de Diabetes Melitos tipo 1<sup>8</sup>. Neste tipo de Diabetes Melitos são utilizados hipoglicemiantes orais para controlar os níveis séricos de glicose, combatendo a resistência à insulina por parte das células do organismo do paciente diabético; são também administradas estatinas de modo a prevenir complicações ateromatosas<sup>8</sup>. A diminuição da pressão sanguínea demonstra melhorar a eficácia do tratamento da Diabetes Melitos tipo 2<sup>8</sup>.

Nenhum dos tratamentos atualmente em vigor vai além do controlo glicémico por parte do paciente, não tendo em conta as redes de interação moleculares que conduzem às complicações a longo prazo, responsáveis pela maior morbilidade destas patologias. Urge, pois, esclarecer os mecanismos moleculares de desenvolvimento dstas complicações.

## 1.3 Interesse e desafios da utilização da saliva como fluido de diagnóstico

A saliva tem-se destacado como um potencial fluido de diagnóstico nos últimos anos dado apresentar certas vantagens relativamente a outros testes e exames de diagnóstico. O diagnóstico utilizando saliva não é invasivo, requer pouca ou nenhuma qualificação (podendo a sua colecção ser feita pelo próprio paciente), a saliva possui marcadores presentes igualmente no sangue, pode vir a ser uma alternativa em pacientes pouco colaborantes ou em que a colheita de sangue seja mais complicada, pode ser utilizada para determinar concentrações farmacológicas em pacientes e a presença de substâncias ilícitas no organismo de um indivíduo<sup>9,10</sup>.

Contudo, apesar das vantagens enumeradas, certas moléculas difundem-se mais facilmente na saliva que outras dado as suas características fisico-químicas; moléculas lipofílicas difundem-se mais rapidamente que moléculas lipofóbicas<sup>9</sup>. Outro obstáculo que a utilização da saliva enfrenta reside no facto de o seu fluxo ser variável de indivíduo para indivíduo e no mesmo indivíduo devido a múltiplos factores. Pacientes com doenças sistémicas, sujeitos a radioterapia e/ou polimedicados apresentam diferentes taxas de secreção por comprometimento das glândulas salivares<sup>9</sup>.

No que concerne a variabilidade salivar, estudos recentes de proteómica salivar demonstram que a variabilidade salivar se deve mais às diferenças inter-individuais que às diferenças intra-individuais<sup>11</sup>. Outro factor que pode ter influência é o momento de colheita (altura do dia), existem estudos que defendem que a quantidade relativa de proteínas se mantém constante durante o dia<sup>11</sup> e outros estudos que afirmam que a concentração relativa de proteínas na saliva varia de forma constante e padronizada ao longo do dia<sup>12</sup>.

### 1.4 O oraloma da Diabetes Melitos tipo 1 e 2

O oraloma, enquanto fisioma da cavidade oral, é o conjunto de proteínas presentes nesta cavidade e das suas características moleculares. 13,14

Estabelecer o oraloma de uma determinada patologia passa por esclarecer os mecanismos moleculares alterados como consequência dessa situação.. Para descrever o oraloma é necessário recorrer às ciências ómicas, proteómica, transcriptómica, metabolómica, genómica, interatómica, entre outros.

Actualmente são ainda escassos os estudos de proteómica aplicada à caracterização do oraloma da Diabetes Melitos tipo 1, havendo mais estudos nesta área a descrever o oraloma da Diabetes Melitos tipo 2.

#### 1.4.1 O Oraloma da Diabetes Melitos tipo 1 e 2 - estado da arte

Até à data do início do presente estudo, estavam indexadas 15 proteínas<sup>15</sup> no oraloma da Diabetes Melitos tipo 1 e 445 proteínas referentes ao oraloma da Diabetes Melitos tipo 2<sup>16,17</sup>.

As 15 proteínas quantificadas no oraloma da Diabetes Melitos tipo 1 envolvem maioritariamente processos biológicos imunológicos, envolvendo as vias de sinalização de citocinas.

Ao contrário do que se verifica na Diabetes Melitos tipo 1 onde ainda são relativamente poucos os estudos de proteómica efectuados, existem 445 proteínas indexadas como alteradas no oraloma da diabetes Melitos tipo 2. Deste universo de proteínas, os processos biológicos mais afectados são os processos metabólicos da glicose (havendo uma notória sobre-regulação das proteínas agonistas da gluconeogénese) e os processos biológicos de resposta a estímulos dos quais resulta uma sinalização intra-celular alterada, coroborando o facto de as vias de sinalização em que a insulina intervem estarem comprometidas.

### 1.5 Ferramentas bioinformáticas para o estudo do oraloma

Os dados gerados pelos vários estudos de proteómica que descrevem o oraloma de pacientes diabéticos necessitam, à semelhança de outros estudos aplicados a outras patologias, de ser integrados e sistematizados de modo a poderem ser modelados e interpretados, possibilitando outras inferências. O que se verifica é que existe uma grande quantidade de resultados dispersos na literatura carecendo de integração e interpretação como um todo e não em separado.

Nesse sentido, existe o OralCard<sup>18</sup>, uma ferramenta bioinformática que consiste num repositório que indexa os dados de diversos estudos de proteómica da cavidade oral a partir de anotações manuais.

Os dados obtidos através da revisão da literatura podem, posteriormente, ser interpretados com recurso a outras ferramentas bioinformáticas como o PANTHER<sup>19</sup> e o STRING<sup>20</sup>

### 2 Objectivos

O presente trabalho pretende, após revisão da literatura, reunir e integrar numa só plataforma os resultados dos diversos estudos de proteómica da cavidade oral referentes ao oraloma do paciente diabético, contribuindo através do estudo e análise *in silico* de tal informação para um melhor conhecimento dos mecanismos patogénicos dos dois tipos de Diabetes Melitos. Deste modo, foram objectivos deste trabalho:

- rever a literatura actual de estudos de proteómica da cavidade oral do paciente diabético;
- anotar manualmente a informação proveniente da revisão de artigos científicos, considerada essencial para interpretar os dados de proteómica;
- atualizar o OralCard com a informação anotada;
- analisar in silico os dados obtidos através de ferramentas bioinformáticas;
- registar e analisar as diferenças entre os dois tipos de Diabetes
   Melitos verificadas nas diversas ontologias génicas;
- correlacionar os dados obtidos e estudados com os aspectos clínicos da patogénese e patofisiologia da Diabetes Melitos
- anotar todos os microrganismos identificados pela literatura científica presentes na cavidade oral de pacientes com os dois tipos de Diabetes Melitos

### 3 Material e Métodos

Por forma a alcançar os objectivos propostos foi seguida a metodologia explicada neste capítulo.

Iniciou-se pela revisão da literatura e recolha de dados específicos quanto a proteínas alteradas nos casos de Diabetes Melitos tipo 1 e 2.

Numa segunda fase, a informação obtida foi analizada e interpretada com recurso a ferramentas bioinformáticas.

### 3.1 Atualização do Oraloma da DMT1 e DMT2

O Oraloma, que se encontra reunido no OralCard, espelha a fisiologia molecular da cavidade oral e tem por base a integração de informação proveniente das ciências ómicas tais como a genómica, transcriptómica, proteómica, interactómica e metabolómica.

Apesar do Oraloma já integrar informação referente aos dois tipos de Diabetes Melitos, tal carece de revisão e atualização constante uma vez que desde a última revisão surgiram mais estudos de proteómica salivar em pacientes diabéticos.

Sabendo que proteínas se encontram alteradas em cada tipo de Diabetes Melitos é possível comparar que vias de sinalização podem estar alteradas e que influência podem estas ter no ecossistema oral. Assim, foi efectuada uma revisão da literatura existente, de modo a atualizar o Oraloma de acordo com os mais recentes estudos de proteómica da cavidade oral.

Desde a última revisão do Oraloma<sup>13</sup>, mais estudos de proteómica da cavidade oral foram efectuados.

De modo a atualizar o Oraloma, recorreu-se ao arquivo digital de artigos científicos PubMed<sup>21</sup> com a seguinte fórmula de pesquisa:

((("DMT1" OR "DMT 1" OR "diabetes Melitos Type 1") AND (saliva\* OR oral) AND (protein OR proteomics) NOT treatment AND ("last 10 years"[PDat]))) AND ("last 10 years"[PDat]) cingida a estudos respeitantes a seres humanos e efectuados nos últimos 10 anos, para pesquisar literatura referente à Diabetes Melitus tipo 1. Foram considerados resultados até Janeiro de 2013,

inclusive.

Foram obtidos 119 resultados dos quais foram selecionados 14 artigos científicos<sup>15,22–34</sup> contendo informações relevantes quanto às proteínas encontradas alteradas em pacientes com Diabetes Melitos tipo 1.

A mesma metodologia foi utilizada para pesquisar informação molecular oral sobre pacientes com Diabetes Melitos tipo 2. Foi utilizada a seguinte fórmula de pesquisa:

(DMT2) OR (DMT 2) OR (diabetes Melitos type 2) OR (type 2 diabetes Melitos) OR (non insulin dependent diabetes Melitos) AND (SALIV\* OR oral) AND (proteome OR peptide OR protein OR proteomic) restringida a estudos efectuados entre 1/1/1997 e 31/01/2013 (dia em que foi efectuada a pesquisa) em seres humanos.

Foram obtidos 4323 resultados, dos quais foram selecionados 19 artigos científicos<sup>2,16,17,23,35–49</sup>.

Os artigos selecionados foram revistos e a informação contida foi anotada em tabelas de Microsoft Excel de modo a organizar a informação quanto à identificação das proteínas identificadas, proveniência histológica da amostra, quantificação da alteração registada, desenho do estudo e respectivos critérios de inclusão/exclusão de indivíduos e hábitos sociais da população estudada.

### 3.2 Caracterização do Oraloma da DMT1 e DMT2

Após a revisão da literatura científica mais atual quanto às proteínas que se encontram alteradas na cavidade oral de um paciente diabético, foi necessário anotar, catalogar e integrar essa informação de modo a possibilitar o estudo *in silico* do oraloma do paciente diabético.

Foram utilizadas técnicas algébricas e computacionais para estudar o conjunto das proteínas encontradas e assim comparar de que forma, as mesmas, podem influenciar o ecossistema oral nos dois tipos de diabetes.

### 3.2.1 Anotação dos dados obtidos na revisão da literatura

Muita informação relevante, proveniente de estudos de proteómica, referente ao oraloma do paciente diabético, encontra-se dispersa. Sendo um dos objectivos do presente trabalho reunir e integrar esses dados por forma a conseguir estudá-los *in silico* e compreender em que nível o ecossistema oral espelha os mecanismos e condições patofisológicas dos dois tipos de Diabetes Melitos.

Consequentemente, para efetuar este registo foi utilizado o software Microsoft Excel versão 14.0. Este software permite a realização de cálculos na própria folha de registo e possui uma vasta biblioteca de funções algébricas que podem ser utilizadas para o estudo dos valores observados.

O conjunto das proteínas em estudo foi organizado em tabela, onde se discriminaram diferentes características da proteína e do estudo que a identificou.

A informação anotada é idêntica à que é utilizada noutros trabalhos semelhantes por forma a estandardizar a metodologia de recolha de informação e permitir a sua posterior inclusão no repositório do Oraloma humano, o OralCard.

Para registar os dados provenientes da revisão da literatura, utilizou-se o programa Microsoft Excel 14.0. Foi preenchida uma tabela-base de Microsoft Excel cujas colunas correspondem a entradas de formulário para preenchimento conforme a informação obtida.

Na figura 1 estão enumeradas as colunas utilizadas.

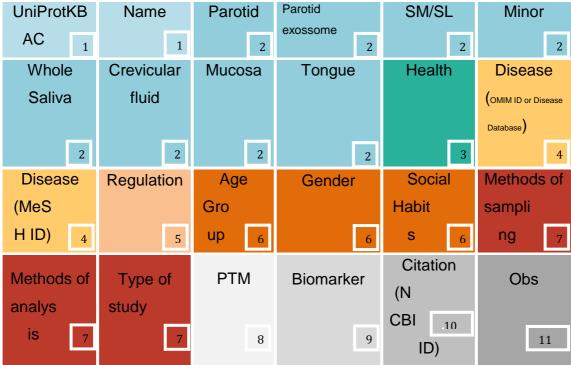


Figura 1 - - Campos de informação anotada manualmente para cada uma das proteínas identificadas. 1-identificação da proteína; 2-proveniência da proteína; 3-proteína presente em ind. saudáveis; 4- identificação da doença; 5-Regulação; 6- Caracterização da população estudada; 7- Metodologia do estudo; 8- proteína com modicações pós-transcripcionais; 9- proteína identificada como biomarcador da patologia; 10 - Código NCBI do artigo, 11- Observações, informações adicionais.

Durante a revisão da literatura sempre que surge uma proteína digna de registo são preenchidos os campos representados na Fig. 1.

Inicia-se a anotação por identificar a proteína através do seu código unívoco, o código Uniprot KB<sup>50</sup> e o seu nome (Fig.1-1); regista-se igualmente a proveniência da amostra onde a proteína foi identificada e refere-se se existe informação se a proteína anotada se encontra em indivíduos saudáveis(Fig.1-2 e 3). De modo a corresponder a proteína anotada a uma patologia em específico preenchem-se ambos os códigos OMIM e MESH da doença(Fig.1-4). Se houverem dados concretos quanto à quantidade da proteína na amostra, anota-se o valor da regulação, cujo cálculo se encontra explicado no ponto seguinte (Fig.1-5). Interessa igualmente registar o tipo de população estudado no artigo de onde se retirou informação da proteína (Fig.1-6), assim como a sua metodologia de colheita e análise de amostras (Fig.1-7). É possível registar se a proteína anotada apresenta modificações póstraducionais (Fig.1-8) ou se foi se foi sugerida como biomarcador para a

doença em estudo (Fig.1-9). Por último anota-se o código NCBI do artigo a partir do qual culminou a anotação (Fig.1-10) e, se necessário regista-se algum tipo de informação adicional não contemplado pelos campos anteriores, preenchendo-se o campo destinado a observações (Fig.1-11).

### 3.2.1.1 Cálculo da regulação da proteína

Nos estudos selecionados, cada proteína, na maior parte dos casos, foi analisada e quantificada tanto em diabéticos como no grupo controlo. De modo a quantificar a sua alteração, ou seja, a sua regulação, recorreu-se ao cálculo de "fold change". Este cálculo permite obter a variação de um valor inicial para um final, neste caso de um controlo saudável para um diabético tipo 1 ou tipo 2. Quando o resultado do "fold change" é inferior a 1, utiliza-se o negativo do seu inverso ou seja:

Se o controlo for 80 e o paciente diabético tiver 20 o "fold change" será:

20/80= 0,25 o inverso do negativo significa que:

-(1/(20/80)=-4

ou seja, 4 vezes menos. Este cálculo facilita a perceção da variação da quantidade (ou atividade) de uma proteína na doença em relação ao controlo saudável.

Este cálculo foi aplicado a todas as proteínas cujos estudos mencionavam valores do paciente diabético e do controlo.

O "fold change" permite então a percepção e quantificação da alteração de uma proteína entre o controlo saudável e o paciente diabético, facilitando a interpretação das alterações moleculares observadas entre os estados de saúde e de doença.

### 3.2.2 Caracterização segundo ontologias

As proteínas estudadas foram classificadas quanto à sua função molecular, processos biológicos em que intervêm e vias de sinalização afectadas, através da ferramenta PANTHER<sup>19,51</sup> (http://www.pantherdb.org). Esta ferramenta bioinformática possibilita a classificação de genes e proteínas de acordo com as funções que desempenham e a sua relação evolutiva, baseando-se em dados científicos experimentais publicados e pela inferência de revisores especializados, o que permite prever funções mesmo quando não haja evidência experimental direta.

Para tornar estas inferências o mais atualizadas e exatas possível, o PANTHER utiliza dados computacionais e revisão manual à medida que mais dados são gerados e acumulados.

O *input* de dados no PANTHER faz-se do respetivo código UniprotKB<sup>50</sup>. É igualmente possível introduzir uma lista de proteínas introduzindo os respetivos códigos UniprotKB em simultâneo, estudando as proteínas por grupos (figura 2). Para este estudo foram criadas sete listas no PANTHER, uma com o total das proteínas anotadas no Oraloma e duas outras listas com as proteínas alteradas em pacientes com Diabetes tipo 1 e 2 (e respectivas listas das proteínas sub e sobre reguladas em cada tipo de Diabetes Melitos).



Figura 2 - Campo de introdução de dados por grupo no PANTHER.

## 3.2.2.1 Classificação segundo processos biológicos, funções moleculares e vias de sinalização

Após introdução dos códigos UniprotKB e as listas de cada tipo de Diabetes Melitos criadas recorre-se ao PANTHER para classificar cada grupo de proteínas (Oralome, Diabetes Melitos tipo 1, Diabetes Melitos tipo 2) de acordo com o processo biológico em que participam e com as funções moleculares que desempenham.

O mesmo procedimento foi utilizado para as 3 listas e começando-se por selecionar uma das listas. Tomemos como exemplo a Diabetes Melitos tipo 1. Após clicar em submeter, e de acordo com os critérios selecionados, o PANTHER irá gerar uma lista com todos os genes de onde as proteínas em estudo são provenientes e a informação quanto a funções moleculares e processos biológicos em colunas.

Nesta lista (Figura 3) existe a opção de mostrar a informação em gráfico circular e de acordo com várias ontologias (função molecular, processo biológico, classificação proteica, localização celular e via de

sinalização).

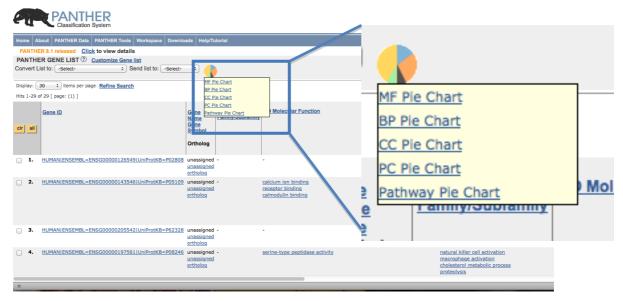


Figura 3 - Metodologia para criar gráficos circulares. MF- Molecular Funtion, BP - Biological Process, CC - Celular Component, PC - Protein Class, utilizando a ferramenta PANTHER.

Quando já se encontra numa vista de gráfico circular pode-se alterar a ontologia como explicado na figura 4:

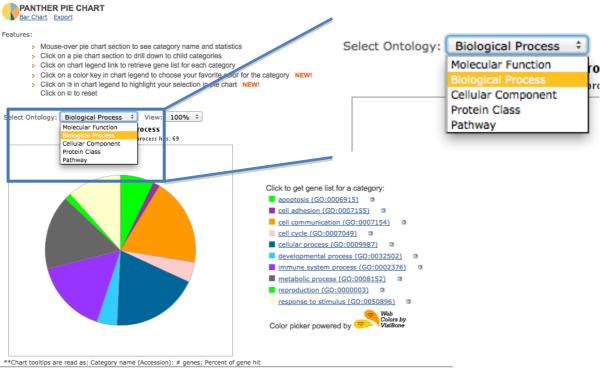


Figura 4 - Gráfico circular representando os processos biológicos onde estão presentes as proteínas alteradas nos pacientes com Diabetes Melitos tipo 1, segundo a ferramenta PANTHER.

Após gerar as listas no

PANTHER é possível exportar a informação relativa a cada uma das ontologias em ficheiro .txt; o conteúdo do ficheiro exportado é importado para uma folha de cálculo do Excel para posterior estudo.

### 3.2.3 Interpretação da Interactómica das proteínas alteradas

Após registar que proteínas se encontram alteradas nos dois tipos de Diabetes Melitos, em que funções moleculares e processos biológicos intervêm, interessa interpretrar a rede interatómica formada por essas proteínas.

A interatómica é a ciência que estuda a relação e interação de biomoléculas entre si, neste caso: proteínas.

Existe uma ferramenta bioinformática, nomeadamente o STRING<sup>20,52–54</sup>(http://string-db.org/), que permite inserir todos os códigos UniprotKB de cada tipo de Diabetes Melitos em simultâneo e gerar uma visualização esquemática da rede interatómica das proteínas em estudo.

O STRING (Figura 5) permite gerar estas redes de interatómica baseando-se em quatro fontes de informação:

- dados de genómica
- dados experimentais directos
- co-expressão
- text mining

Actualmente o STRING encontra-se na sua versão 9.05 e possui informação quanto a 5 214 234 proteínas provenientes de 1133 organismos.

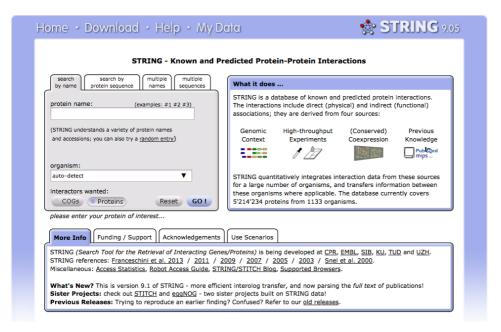


Figura 5 - Página de rosto da ferramenta bioinformática STRING, para estudos de interatómica

No separador "Multiple Names" (Figura 6) colocam-se todos os códigos Uniprot de um grupo de proteínas a estudar, neste caso a Diabetes Melitos 1 ou 2; introduzidos os códigos UniprotKB clica-se em "GO!".

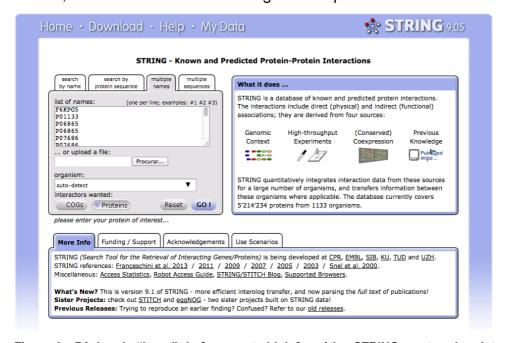


Figura 6 - Página de "input" da ferramenta bioinformática STRING mostrando a introdução das proteínas alteradas em Diabetes melitos Tipo 2.

Após clicar em "GO!" aparecerá a lista de todas as proteínas a integrar a rede de interatómica. Em caso de ambiguidade, terá de se proceder à identificação manual das proteínas.

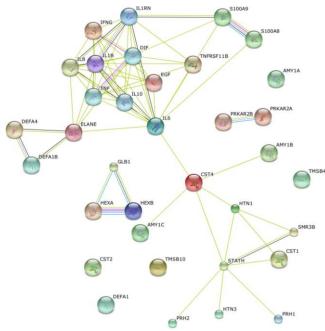


Figura 7 - Rede interatómica gerada no STRING, as diferentes cores das linhas representam as diferentes fontes de informação

Nas redes de interatómica geradas no STRING (Figura 7), as linhas que unem as proteínas podem ser alteradas de acordo com vários parâmetros:

- Confiança as linhas que traduzam interacções de maior confiança surgem mais espessas que as de menor grau de confiança,
- Evidência existe mais
   que uma linha para cada

interacção, cada cor das linhas corresponde a uma fonte de informação,

 Acção - cada cor de cada linha é um tipo de acção entre as duas proteínas.

O STRING disponibiliza funcionalidades que facilitam a interpretação dos dados de interatómica.

É possível evidenciar grupos de proteínas de acordo com ontologias tais como: função molecular, processo biológico, componente celular e via de sinalização. Outra funcionalidade disponível consiste no *clustering* que agrupa as proteínas de acordo com o seu *STRING global score*, ou seja, proteínas cujas interacções aparentam ser mais fidedignas. Os *clusters* ou grupos de proteínas podem ser modelados através de dois algoritmos, KMEANS e MCL, sendo que foi utilizado o algoritmo MCL que agrupa as interacções por "inflação" um parâmetro relativo à precisão do *clustering*.

# 3.2.4 Comparação entre os Oralomas da Diabetes Melitos tipo 1 e 2 e o total de anotações do Oralome

Após recolha dos dados referentes às 3 principais ontologias, funções moleculares, processos biológicos e vias de sinalização, é importante compará-los ao total de proteínas anotadas no Oralome.

Esta comparação permite determinar, com um intervalo de confiança específico, os valores obtidos com valores expectáveis se se verificasse uma proporção directa relativamente ao total de proteínas anotadas no Oralome e entre os dois tipos de Diabetes Melitos entre si. Esta comparação foi efectuada de igual forma tanto para a Diabetes Melitos tipo 1 como para a Diabetes Melitos tipo 2.

Para esta comparação anotaram-se os valores obtidos no PANTHER de cada item de cada ontologia em Microsoft Excel. Para cada um destes colocou-se o valor do total de anotações do Oralome e do verificado na DMT1 e na DMT2 lado a lado. Por cálculo de proporção directa, foi inferido o valor que seria de esperar para cada item se houvesse proporção directa entre as entidades comparadas.

Para quantificar a diferença entre os valores observados e os valores esperados para cada item, calculou-se a diferença fraccional que consiste em:

$$\mathbf{DF} = \left(\frac{\mathbf{Z}_{\mathsf{t}} - \hat{\mathbf{Z}}_{\mathsf{t}}}{\hat{\mathbf{Z}}_{\mathsf{t}}}\right)$$

Em que  $Z_t$  representa o valor observado  $\hat{Z}_t$  o valor esperado e DF a diferença fraccional.

A diferença fraccional é, então, o valor cuja multiplicação pelo valor esperado traduz quanto deveria ser adicionado ou subtraído ao valor observado para igualar o valor teoricamente esperado.

Recorrendo a uma distribuição binomial para calcular a probabilidade de se observar um resultado tão extremo que contradiga o que se inferiu, calculou-se o *p-value*.

Quanto menor o p-value menor probabilidade do resultado apresentado ser devido à aleatoridade. Consequentemente um resultado é tanto mais significativo quanto menor for o seu *p-value*.

#### 3.2.5 Comparação da microflora oral da Diabetes Melitos tipo 1 e 2

Para comparar os microrganismos presentes na cavidade oral de pacientes com DMT1 e DMT2, começou-se por efectuar uma revisão bibliográfica de estudos de microbiologia oral.

Utilizou-se o repositório online de artigos científicos PubMed no dia 29/6/2013 com a seguinte fórmula de pesquisa:

((Micro\*)OR(bacteria)) AND ((Oral) OR (saliv\*)) AND ((type 1 diabetes mellitus) OR (DMT1)) - para a DMT1, obtendo 64 resultados;

((Micro\*)OR(bacteria)) AND ((Oral) OR (saliv\*)) AND ((type 2 diabetes mellitus) OR (DMT2))- para a DMT2, obtendo 186 resultados.

A informação seleccionada foi anotada em Microsof Excel 14.0 cujas colunas correspondem a entradas de formulário para preenchimento conforme a informação obtida. As colunas agruparam a informação por:

- NCBI Taxon ID
- Organism Name
- All Body subsites
- Regulation
- Citation (NCBI ID)
- Obs.

## 4 Resultados e Discussão

Da aplicação das metodologias descritas na secção Material e Métodos, obtiveram-se os resultados que apresentamos e discutimos nesta secção.

## 4.1 Revisão de literatura e anotação manual dos dados obtidos

A atualização do Oraloma da diabetes (secção 3.1) resultou na anotação de 29 novas proteínas no oraloma da Diabetes Melitos tipo 1<sup>23–34</sup> e 58 novas proteínas no oraloma da Diabetes Melitos tipo 2<sup>17,23,35,37,38,46,47,55</sup> (Anexos - Tabela 1 e 2); estes valores não coincidem com os valores analisados na caracterização segundo ontologias uma vez que de todas as proteínas anotadas no oraloma do paciente diabético existem certas proteínas que ainda não constam na base de dados do PANTHER.

Enquanto na Diabetes Melitos tipo 1 todos os estudos referidos utilizaram métodos de análise de amostras diferentes, na Diabetes Melitos tipo 2, 3 estudos quantificaram as proteínas com recurso ao teste bioquímico ELISA. Esta realidade evidencia a necessidade da elaboração de metodologias estandardizadas no que concerce a análise de saliva e tecidos orais de modo a não prejudicar a validade dos dados obtidos quando relacionados entre estudos.

Tabela 1 - Proteínas que os oralomas dos dois tipos de Diabetes Melitos têm em comum

Cód. UniprotKB/Nome da	Diabetes	Diabetes					
proteína	Melitos tipo 1	Melitos tipo 2					
P01133 Pro-epidermal growth	-1,2607	1,21					
factor							
P07686 Beta-hexosaminidase	-1,125 (non-	não quantificado					
subunit beta	smokers)						
	3,0 (smokers)						
P04745 alpha-amylase	3,1407	4,02					
O00300 Tumor necrosis factor	-2,5691	+					
receptor superfamily member							
11B							
P05231 Interleukin-6	29,1667	0,00					
P08246 Neutrophil elastase	1,0635	Não quantificado					
P01036 Cystatin-S	Não quantificado	-2,44					
P01037 Cystatin-SN	Não quantificado	-4,27					
P02808 Statherin	-3,11	Não quantificado					
diphosphorylated							
P02810 Salivary acidic proline-	-2,21	Não quantificado					
rich phosphoprotein							
P06702 Protein S100-A9	1,56	+					

Na tabela 1 é possível verificar que apesar dos dois tipos de Diabetes Melitos possuírem proteínas em comum, estas podem apresentar regulações diferentes. O facto de possuírem regulações dispares pode consistir *per se* numa diferença, contudo é necessário ter em conta que existem outras variáveis que influenciaram estes valores, os estudos não seguem uma metodologia de colheita e análise comum dado não existir qualquer estandardização metodológica para a colheita e análise de amostras de tecidos e fluídos da cavidade oral.

#### 4.1.1 OralOma da Diabetes Melitos tipo 1

Antes do presente estudo haviam sido anotadas no Oralome 14 proteínas, a revisão de literatura efectuada possibilitou o registo de mais 19

proteínas (em 22 anotações), perfazendo um total de 29 proteínas anotadas. Deste conjunto de proteínas, 21 provêm da saliva total, 8 de amostras da mucosa, 3 de amostras da glândula parótida e 1 oriunda do fluído crevicular (figura 8).

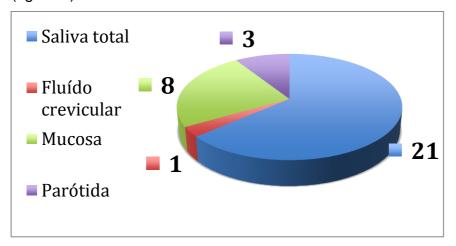


Figura 8 - Proveniência histológica das proteínas alteradas em casos de Diabetes Melitos tipo 1

#### 4.1.2 OralOma da Diabetes Melitos tipo 2

Foram adicionadas 58 proteínas ao oraloma do paciente com Diabetes Melitos tipo 2 que antes deste estudo já contava com 445 proteínas anotadas no Oralome. O universo de proteínas do oraloma da Diabetes Melitos tipo 2 é composto por 493 proteínas oriundas de amostras de saliva, 6 proteínas são provenientes de amostras da glândula parótida e 4 das glândulas submandibulares/sublinguais (figura 9).

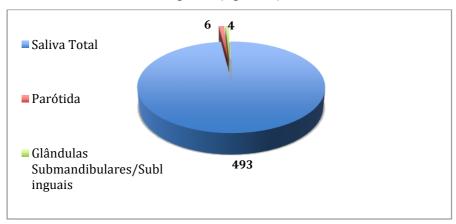


Figura 9 - proveniência das proteínas alteradas em casos de Diabetes Melitos tipo 2

A maioria das proteínas anotadas provêm de estudos de proteómica que utilizaram a saliva como amostra. A saliva acaba por conter proteínas originárias de diversos tecidos e fluídos uma vez que contacta com todas as estruturas orais e possui o contributo do fluído crevicular que sofre, por sua vez, influência directa do sangue por intermédio dos exsudados da microcirculação do sulco gengival conferindo à saliva capacidade de reproduzir doenças sistémicas e não apenas da cavidade oral.

#### 4.1.3 Metodologia dos estudos seleccionados na revisão da literatura

Das 33 anotações referentes à Diabetes Melitos tipo 1, 14 são resultado de estudos efectuados em crianças (sem especificar um intervalo de idades), 4 anotações provêm de estudos efectuados em indivíduos entre os 40 e os 58 anos de idade; o artigo que quantificou os níveis de elastase compreendeu indivíduos entre os 26 e os 51 anos; os estudos dos quais resultaram as restantes anotações envolveram indivíduos com idade inferior a 20 anos. A faixa etária tendencialmente mais jovem dos indivíduos estudados por cada autor justifica-se dado o *onset* da Diabetes Melitos tipo 1 ocorrer nessas mesmas idades.

No que concerne a regulação das proteínas anotadas em casos de Diabetes 1, verificam-se, no caso da hexosaminidase (subunidade A e B) e da Beta-Galactosidase valores de regulação diferentes sejam os indivíduos estudados fumadores ou não<sup>26</sup>. A regulação que registou o valor mais elevado foi referente à Interleucina-8 (regulação: 35)<sup>27</sup> seguida da Interleucina-6 (regulação: 29)<sup>27</sup>; a anotação com o valor mais baixo em termos de regulação pertence à Histatina-1 (Regulação: -3,14738)<sup>15</sup>.

À excepção de um estudo que compreendeu indivíduos com Diabetes Melitos tipo 2 dos 18 aos 66 anos, todos os estudos que quantificaram as proteínas alteradas em casos de Diabetes Melitos tipo 2 utilizaram populações de indivíduos com idades não inferiores a 30 anos. Este facto corrobora a informação que estabelece que o *onset* e diagnóstico da Diabetes Melitos tipo 2 ocorre geralmente na idade adulta.

A proteína Elongation factor G, mitochondrial (Q6CRY5, regulação: 702,46)<sup>55</sup> foi a proteína com a regulação mais elevada, ao passo que a proteína FAM92A1 registou o valor mais baixo de regulação (regulação: -91,53)<sup>55</sup>.

Outro dos obstáculos à obtenção, por meio de estudo *in silico*, de dados estatísticos com significância, é a quantidade de proteínas anotadas em Diabetes Melitos tipo 1 (33 proteínas) ser diminuto quando comparado ao Oralome total (4376 proteínas) e mesmo ao Oraloma de Diabetes Melitos tipo 2 que compreende 503 anotações. Urge, pois, desenvolver mais estudos de proteómica no sentido de identificar as proteínas orais alteradas em DMT1 relativamente a indivíduos saudáveis.

## 4.2 Caracterização do Oraloma do paciente diabético

## 4.2.1 Caracterização segundo Ontologias génicas

Por forma a analisar que funções moleculares possuem, em que processos biológicos intervêm e que vias de sinalização afectam, as proteínas alteradas em ambos os tipos de Diabetes foram caracterizadas de acordo com ontologias génicas. As ontologias génicas não irão ser traduzidas ao longo do presente trabalho de modo a evitar deturpar o seu significado biológico.

Para melhor comparar os dois tipos de Diabetes Melitos, recorreu-se a um cálculo de diferença fraccional, tendo por base a Diabetes Melitos tipo 2 dado esta possuir um maior número de proteínas anotadas. Este cálculo traduz a diferença entre quantas proteínas do Oraloma da Diabetes Melitos tipo 1 seriam expectáveis e as que foram anotadas na realidade em cada categoria ontológica, caso exista proporcionalidade directa entre a Diabetes Melitos tipo 1. Com recurso a uma distribuição binomial foi calculado o *p-value* em cada diferença fraccional de modo a poder atribuir um grau de significância estatística a cada resultado (Anexo 3)

#### 4.2.1.1 Funções Moleculares

Os oralomas dos dois tipos de Diabetes Melitos apresentam 4 tipos de funções moleculares comuns (Figura 10).

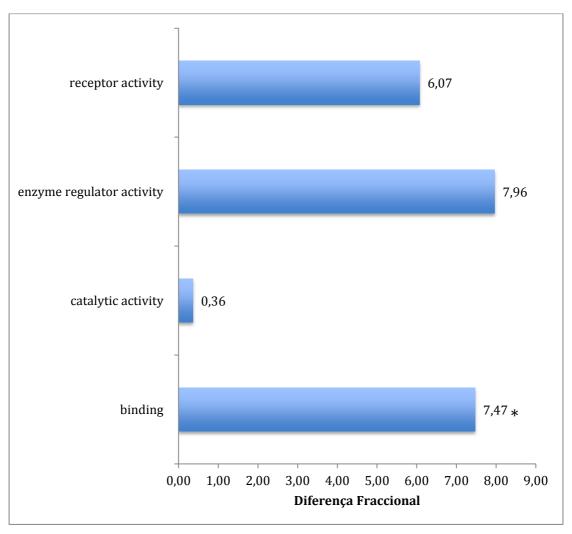


Figura 10 - Diferença fraccional do oraloma da DMT1 em relação ao oraloma da DMT2, estão apenas representadas as funções moleculares comuns entre os dois oralomas (\* valores com *p-value*<0,05)

A função molecular *binding* é uma função vasta, que pode englobar proteínas que com alguma probabilidade não se enquadram apenas nesta função molecular. Contudo, analisando as suas sub-funções, é possível denotar diferenças entre as proteínas implicadas na função de *binding* entre os dois oralomas (Figura 11). As 15 proteínas envolvidas em *binding* no oraloma da diabetes Melitos tipo 1 são essencialmente citocinas e proteínas envolvidas na ligação a cálcio, ao passo que a função molecular de *binding* compreende 125 proteínas do oraloma da Diabetes Melitos tipo 2 que consistem não só em citocinas como também em proteínas de sinalização intracelular. No entanto, tendo em conta a redundância desta função molecular e não obstante o *p-value* da sua diferença fraccional entre os

oralomas dos dois tipos de Diabetes Melitos ser inferior a 0,05, esta é pouco informativa como parâmetro comparativo entre estas patologias.

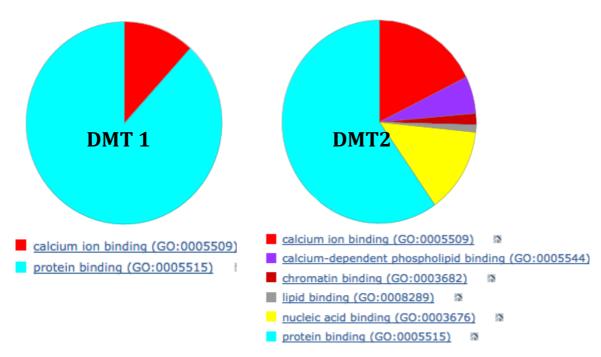


Figura 11 - Funções moleculares contidas na função molecular de *binding* dos oralomas dos dois tipos de Diabetes Melitos (gráfico adaptado de http://www.pantherdb.com).

Quanto à função molecular de *catalytic activity* no oraloma de Diabetes Melitos tipo 1 apenas é representada por 3 proteínas enquanto que no oraloma da Diabetes Melitos tipo 2 é desempenhada por 159 proteínas.

Outra função molecular comum entre os oralomas dos dois tipos de Diabetes Melitos é a de *enzyme regulator activity*. A DMT1 apresenta mais proteínas envolvidas nesta função molecular do que seria de esperar caso houvesse proporção à DMT2, isso traduz-se numa diferença fraccional de 7,96. A função molecular de *enzime regulator activity* nos oralomas dos dois tipos de Diabetes Melitos consiste em funções de fosforilação de substratos por transferência de grupos fosfato a partir de ATP e noutras funções que advêm da gluconeogénese ou glicogenólise<sup>7</sup>.

À semelhança do verificado na função de *enzyme regulator activity*, existem igualmente mais proteínas no oraloma de Diabetes Melitos tipo 1 a desempenhar a função de *receptor activity* do que seria expectável caso se verificasse proporcionalidade entre os oralomas dos dois tipos de Diabetes Melitos. No oraloma de Diabetes Melitos tipo 1 existem 3 proteínas a

desempenhar esta função molecular, uma moduladora de receptor de factores de necrose tumoral, um factor de necrose tumoral e um factor de crescimento epidermal; no oraloma de Diabetes Melitos tipo 2 são 30 as proteínas que desempenham esta função as quais compreendem processos de regulação de receptores, processos imunológicos e de regulação da cascata complemento.

Todas as funções moleculares desempenhadas pelo Oraloma da Diabetes Melitos tipo 1 também são efectuadas pelo oraloma da diabetes Melitos tipo 2 sendo que este possui outras funções moleculares tais como antioxidant activity, ion channel activity, structural molecule activity, transcription regulator activity, translator regulator activity e transporter activity.

Apesar das diferenças descritas para esta ontologia é preciso ter em conta o facto que em proporção o oraloma descrito para a Diabetes Melitos tipo 2 ser substancialmente maior do que o descrito para a Diabetes Melitos tipo 1.

#### 4.2.1.2 Processos Biológicos

A ontologia referente aos processos biológicos classifica os oralomas dos dois tipos de Diabetes Mellitus quanto aos processos biológicos por eles influenciados sendo que consiste na ontologia que providencia mais pontos de comparação entre os oralomas dos dois tipos de Diabetes Mellitus.

O oraloma da DMT1 contém proteínas que participam em 10 processos biológicos ao passo que o oraloma da DMT2 inclui proteínas que participam em 16 processos biológicos.

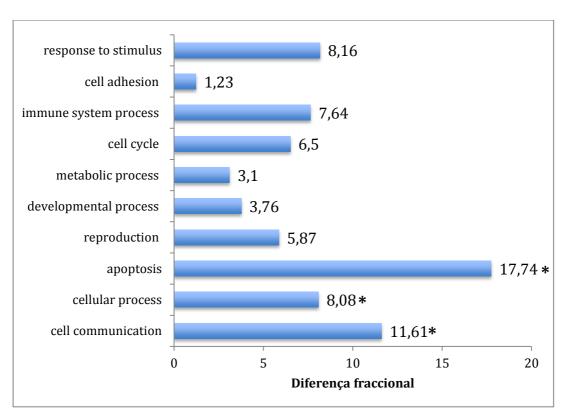


Figura 12 - Diferença fraccional do oraloma da DMT1 em relação ao oraloma da DMT2, estão apenas representadas os processos biológicos comuns entre os dois oralomas (\* valores com *p-value<0,05*)

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Quando comparados, os dois oralomas influenciam 10 processos biológicos em comum, ou seja, todos os processos influenciados pelo oraloma da DMT1 são igualmente influenciados pelo oraloma da DMT2 (Figura 12). Contudo, a comparação entre os processos biológicos influenciados é estatisticamente significativa, ou seja, apresenta um *p-value* inferior a 0,05 em apenas 3 processos biológicos.

Todos os processos biológicos possíveis de comparar entre os dois oralomas demonstraram que, face ao valor esperado tomando o oraloma da DMT2 como referência e partindo do princípio que existe proporcionalidade entre eles, o oraloma da DMT1 apresenta mais proteínas a influenciar cada processo biológico.

O processo biológico de cell communication apresenta uma diferença fraccional de 11,61 e p-value inferior a 0,05. Este processo biológico é principalmente influenciado pelas citocinas do oraloma da DMT1 que influenciam, entre outros, dois sub-processos biológicos como a intracellular signaling cascade, que por sua vez compreende a JAK-STAT cascade e calcium-mediated signaling, e a cascata de sinalização JNK pertencente à sinalização por MAP-cinases que é um agonista da apoptose<sup>56,57</sup>. Os níveis elevados de glicose culminam na produção de espécies de oxigénio reactivas que activam a cascata de sinalização JAK-STAT; esta via de sinalização induz a produção de *Tumor Growth Factor-beta* e de proteína da matriz extra-celular como a fibronectin e collagen IV<sup>58</sup>. As proteínas JAK são activadoras dos receptores membranares de citocinas e, na presença do ligando, são fosforiladas, desencadeando as STAT's que por sua vez aumentarão a transcrição de certas sequências de ADN que influenciarão a apoptose, proliferação e crescimento celular. O oraloma da DMT2 influencia este processo biológico também por influenciar, entre outras cascatas de sinalização, a cascata de sinalização JAK-STAT e MAPKKK<sup>58,59</sup>.

O processo biológico *cellular process* abrange os processos do ciclo de vida celular como a mitose, motilidade, adesão, citocinese e a acima referida comunicação celular. Por abranger este último processo, a

comunicação celular, que por si também se encontra aumentado no oraloma da DMT1 face ao oraloma da DMT2, é plausível que o processo biológico cellular process apresente também uma diferença fraccional positiva de 8,08 com p-value inferior a 0,05. Contudo este processo biológico é redundante dado abranger outros processos biológicos do mesmo nível como a cell communication, cell adhesion e cell cycle, e consequentemente não se apresenta como um bom parâmetro comparativo entre os oralomas da DMT1 e DMT2.

O valor mais elevado de diferença fraccional entre os oralomas dos dois tipos de Diabetes Melitos corresponde ao processo biológico de *apoptosis* que apresenta uma diferença fraccional de 17,74. Este valor de diferença fraccionnal é estatisticamente significativo uma vez que apresenta um *p-value* inferior a 0,05. É expectável que o processo biológico de *apoptosis* apresente tal diferença fraccional dado a DMT1 ter características autoimunes, indutoras de apoptose a células do organismo, maioritariamente às células β pancreáticas<sup>6,60</sup>.

Dos processos biológicos comparados, 7 comparações não apresentam significância estatística. Na base de tal facto é provável que esteja a reduzida dimensão do oraloma da DMT1.

Os processos biológicos *immune system process* e *response to stimulus* são maioritariamente representados pelas citocinas e factores de necrose tumoral dos oralomas dos dois tipos de Diabetes Melitos.

# 4.2.1.2.1 Processos biológicos afectados exclusivamente pelo oraloma da DMT2

Todos os processos biológicos representados no oraloma da DMT1 estão também representados no oraloma da DMT2. Por outro lado, o oraloma da DMT2 inclui proteínas envolvidas em 5 processos biológicos de forma exclusiva, nomeadamente os processos de *localization, transport, system process, homeostatic process* e *generation of precursor metabolites and energy.* Tal pode ser explicado pelo reduzido oraloma da DMT1, dado que os processos biológicos enumerados estarão, de acordo com a patofisiologia da Diabetes Melitos, alterados em ambos os tipos de Diabetes Melitos<sup>7</sup>.

## 4.2.1.3 Caracterização das vias de sinalização afectadas

Por último, os oralomas dos dois tipos de Diabetes Melitos foram comparados na ontologia de vias de sinalização (Figura 13). A este nível o oraloma da DMT1 abrange 21 vias de sinalização, ao passo que o oraloma da DMT2 compreende 62 vias de sinalização, entre os dois oralomas existem 9 vias de sinalização em comum que, quando comparadas desta forma, 7 apresentam um *p-value* inferior a 0,05.

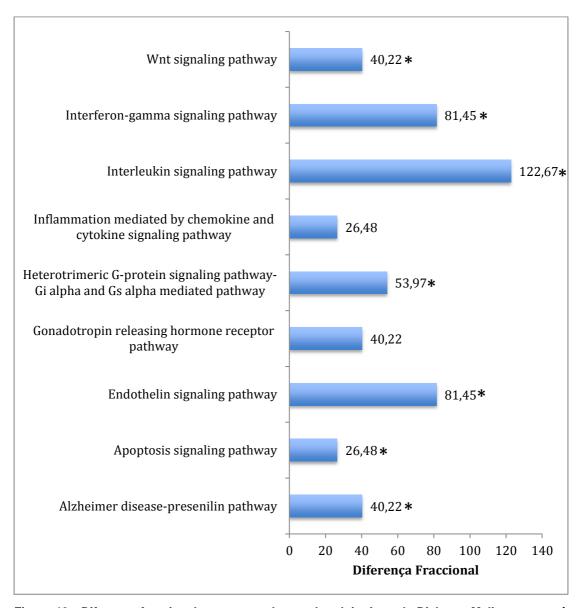


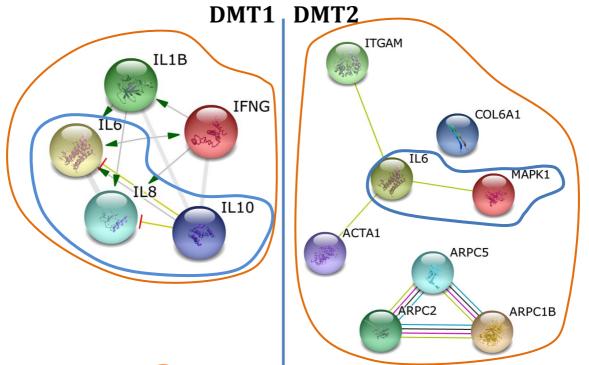
Figura 13 - Diferença fraccional entre os oralomas dos dois tipos de Diabetes Melitos quanto à ontologia de vias de sinalização (\* valores com *p-value<0,05*)

A via de sinalização *Wnt signaling pathway* pode subdividir-se em três tipos que regulam desde a transcrição génica, a forma da célula e os níveis de cálcio intracelulares. Não obstante, o factor de transcrição TCF7L2, componente desta via de sinalização, foi identificado como factor de risco para a Diabetes Melitos tipo 2, sendo que esta via de sinalização influencia o desenvolvimento do pâncreas endócrino e modula a secreção de insulina, a sobrevivência e a proliferação das células-β do pâncreas e a adipogénese<sup>61–64</sup>, a diferença fraccional apresentada (40,22) pode sugerir uma sobre-expressão das proteínas desta via por forma a colmatar a perda de células-β do pâncreas.

O interferão-gama é uma citocina pleiotrófica que desempenha funções cruciais ao nível da imunidade intracelular contra patógenos e tumores. Esta citocina é maioritariamente produzida por células *Natural Killer* e *Natural Killer T* no domínio na imunidade inata, contudo esta molécula desempenha funções na imunidade adquirida sendo secretada por células Th1 CD4 e CD8<sup>65</sup>. O interferão gama produz efeitos tanto por activação das cinases da cascata de sinalização JAK-STAT como por outras vias de sinalização, o balanço entre o efeitos desta citocina entre a via de sinalização dependente ou independente da STAT1 é que determina a resposta biológica ao interferão gama<sup>59</sup>. A sinalização mediada por interferão gama está, comparativamente ao oraloma da DMT2, consideravelmente aumentada no oraloma da DMT1, dado expectável uma vez tratar-se de uma doença auto-imune e o interferão gamma ser uma citocina sobre-regulada neste tipo de doenças<sup>66</sup>.

Da mesma forma que a sinalização mediada por interferão a, mais abrangente, sinalização mediada por interleucinas é a via de sinalização mais aumentada no oraloma da DMT1 face ao oraloma da DMT2.

Atendendo ao facto destas vias de sinalização posuírem proteínas em comum entre si, representaram-se num mapa de interatómica como na figura 14.



Conjunto delimitado por - *Inflammation mediated by chemokine and cytokine signaling pathway*Conjunto delimitado por - *Interleukin signaling pathway* 

Figura 14 - Rede interatómica das proteínas envolvidas nas vias de sinalização de *Inflammation mediated by chemokine and cytokine signaling pathway* e *interleukin signaling pathway*, nos oralomas dos dois tipos de Diabetes Melitos. Rede determinada com recurso à ferramenta STRING. Confidence>0,7

A *Interleukin-6* é uma citocina com variadas implicações fisiológicas entre as quais a resistência à insulina, derivado dos seus efeitos inibitórios na transcrição génica de *IRS-1* e *GLUT-4*, receptores de insulina<sup>67</sup>. Consequentemente, é crucial referir que no oraloma da DMT2 o único estudo que quantificou a expressão de *Interleukin-6* registou uma regulação de 0<sup>17</sup>, ao passo que o estudo que quantificou a *interleukin-6* no oraloma da DMT1 registou uma variação de 29,17<sup>27</sup>. A literatura actual não é consensual quanto à IL-6, havendo autores que quantificaram a IL-6 sérica como normal ou diminuída<sup>68,69</sup>, embora hajam outros autores a quantificar a mesma como aumentada e relacionada com a prévia hiperglicemia e a progressão de complicações microvasculares diabéticas<sup>70,71</sup>.

A via de sinalização da apoptose é influenciada apenas pelo Tumour Necrosis Factor-alpha no oraloma da DMT1 e por 3 proteínas (Cathepsin G, MAPK1 e uma subunidade da Hsp70). Apesar de ser uma comparação com p-value inferior a 0,05 a reduzida quantidade de proteínas nos oralomas dos

dois tipos de Diabetes Melitos, não permite inferir nada não sendo um termo de comparação válido entre a DMT1 e DMT2 até haverem mais estudos de proteómica que quantifiquem mais proteínas nesta via de sinalização, o mesmo se aplica às vias de sinalização *Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pathway*, endothelin signaling pathway, alzheimer disease-presenilin pathway e à Gonadotropin releasing hormone receptor pathway, embora esta última com p-value superior a 0,05.

# 4.2.2 Comparação do oraloma do paciente diabético com o total de anotações do Oralome

Neste estudo comparativo, interessa relacionar o observado, ou seja, as proteínas anotadas nos dois tipos de Diabetes Melitos com o total de anotações em pacientes saudáveis (oraloma normal). Para tal foi utilizado uma vez mais o cálculo de diferença fraccional (Anexo 4 e 5).

### 4.2.2.1 Processos Biológicos

Do estudo comparativo da ontologia génica de processos biológicos, verifica-se que apenas 3 resultados em 9 no oraloma da DMT1 e 4 resultados em 16 no oraloma da DMT2 apresentam significância estatística (Fig. 15).

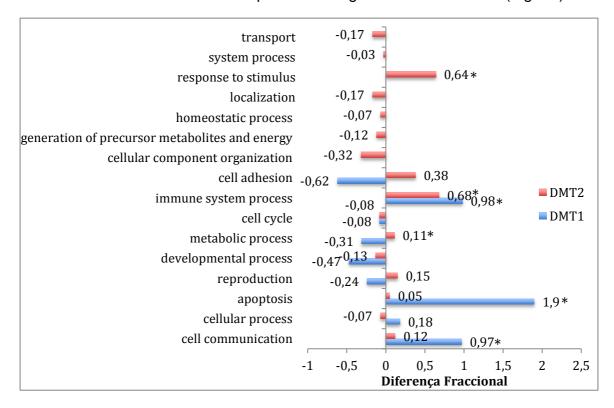


Figura 15 - Diferença fraccional entre os oralomas dos dois tipos de Diabetes Melitos e o de indivíduos saudáveis, quanto à ontologia de processos biológicos (\* valores com *p-value<0,05*)

Do estudo comparativo da ontologia génica de processos biológicos, verifica-se que apenas 3 resultados em 9 no oraloma da DMT1 e 4 resultados em 16 no oraloma da DMT2 apresentam significância estatística (Fig. 15).

Considerando apenas os processos biológicos que apresentam *p-value* inferior a 0,05, temos que os processos biológicos mais alterados no oraloma da DMT1 são os de *cell communication, apoptosis* e *immune system process;* ao passo que no oraloma da DMT2 consistem nos processos biológicos de *cell adhesion, immune system process, metabolic process* e *response to stimulus* (este último sendo mais vago uma vez que engloba, entre outros, o já referido processo biológico de *immune system process*).

A comparação entre o total de proteínas anotados em indivíduos saudáveis e o oraloma dos dois tipos de Diabetes Melitos ao nível da ontologia de processos biológicos, permite concluir que os processos biológicos alterados que apresentam significância estatística corroboram certos aspectos da patofisiologia dos dois tipos de Diabetes Melitos e distinguem-nos das restantes patologias anotadas no Oralome. Alguns processos biológicos estão aumentados num dos tipos de Diabetes Melitos e diminuídos noutro, como é o caso dos processos biológicos:

- Metabolic process, com uma diferença fraccional de 0,11 na DMT2 e
   -0,31 na DMT1
- apoptosis, que apresenta uma diferença fraccional de 0,05 na DMT2 e
   1,9 na DMT1
- cell communication, que regista uma diferença fraccional de 0,12 na DMT2 e 0,97 na DMT1

considerando apenas os processos biológicos em que pelo menos uma das diferenças fraccionais apresenta *p-value*<0,05.

#### 4.2.2.2 Funções Moleculares

As funções moleculares desempenhadas pelas proteínas do oraloma dos dois tipos de Diabetes Melitos estão todas alteradas em relação ao total de proteínas anotadas em individuos saudáveis (oraloma normal) (Fig. 16).

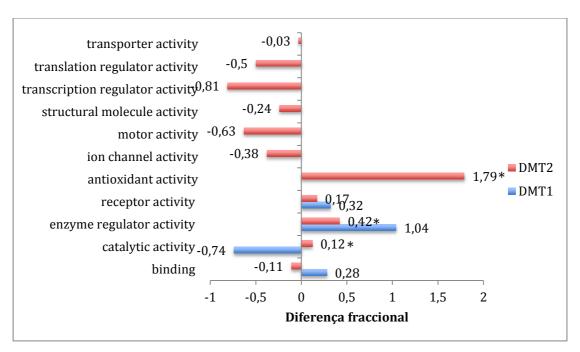


Figura 16 - Diferença fraccional entre os oralomas dos dois tipos de Diabetes Melitos e o de indivíduos saudáveis, quanto à ontologia de funções moleculares (\* valores com *p-value<0,05*)

As funções moleculares desempenhadas pelas proteínas do oraloma dos dois tipos de Diabetes Melitos estão todas alteradas em relação ao total de proteínas anotadas em individuos saudáveis (oraloma normal) (Fig. 16).

Ao nível da ontologia génica de funções moleculares, a comparação estatística entre o total de proteínas anotadas em individuos saudáveis e na DMT1, não é estatísticamente significativa dado a comparação de nenhuma das funções moleculares apresentar um *p-value* inferior a 0,05. Não obstante, é possível verificar que alguns destes valores reflectem aspectos já descritos, como a função molecular de *catalytic activity*. Esta encontra-se diminuída face ao estado de saúde, sendo constituída por *serine proteases*, a coagulação sanguínea dá-se por uma reacção em cadeia destas moléculas; seria, portanto, de esperar que esta função estivesse exacerbada em relação aos individuos saudáveis, mas por outro lado tem um *p-value* de 0,99. Como corolário, a comparação efectuada ao nível da ontologia génica de funções moleculares demonstra a reduzida dimensão do oraloma da DMT1, sendo os resultados a este nível mais aleatórios do que representativos dos padrões desta patologia.

A mesma análise estatística efectuada para o oraloma da DMT2 regista 3 funções moleculares com *p-values* inferiores a 0,05, nomeadamente

as funções moleculares de *antioxidant activity, catalytic activity* e *enzyme regulator activity.* A função de *antioxidant activity* é a que está mais aumentada face ao total de proteínas anotadas em indivíduos saudáveis, justificando-se este valor atendendo ao elevado stress oxidativo em casos de Diabetes Melitos tipo 2<sup>72</sup>.

Atendendo à patofisiologia da DMT2, que leva à ativação de vias enzimáticas alternativas e clivagem de moléculas que não a glicose para a obtenção de ATP's, é expectável que as funções moleculares de *catalytic* activity e enzyme regulator activity estejam exacerbadas face ao total de anotações do Oralome<sup>7</sup>.

## 4.2.2.3 Vias de Sinalização

A comparação estatística por diferença fraccional foi também efectuada para a ontologia de vias de sinalização. Devido aos dois tipos de Diabetes Melitos afectarem numerosas vias de sinalização, apenas foram representadas as que possuem um *p-value* inferior a 0,05 (Fig. 17).

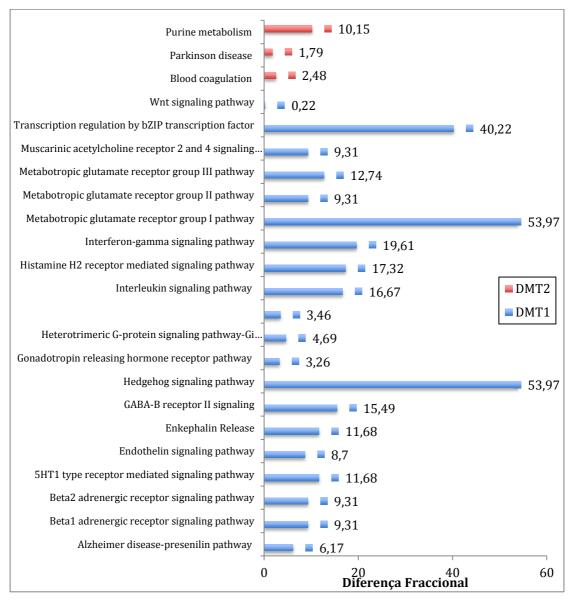


Figura 17 - Diferença fraccional entre os oralomas dos dois tipos de Diabetes Melitos e o de indivíduos saudáveis, quanto à ontologia de vias de sinalização (todos os valores apresentados possuem *p-value<0,05*)

O oraloma da DMT1 compreende 23 vias de sinalização e quando comparadas com o oraloma normal, 21 destas vias de sinalização apresentou um *p-value* inferior a 0,05, o que confere significância estatística a estas comparações. A via de sinalização mais aumentada é a *hedgehog signaling pathway* e a *Metabotropic glutamate receptor pathway*.

No caso do oraloma da DMT2, que engloba 65 vias de sinalização, a comparação com o oraloma normal apenas apresenta significância estatística relativamente a 3 vias de sinalização. Uma dessas vias de sinalização é a de *blood coagulation pathway* que se apresenta aumentada face aos individuos saudáveis, como de resto seria de esperar. As outras duas vias de sinalização com *p-value* inferior a 0,05 consistem nas vias de *parkinson disease* e *purine metabolism*.

# **4.2.3** Caracterização da Interatómica das proteínas alteradas no oraloma de pacientes diabéticos

A interatómica, área que estuda as interações entre biomoléculas, neste caso proteínas, pode ser estudada com recurso à ferramenta bioinformática STRING. Os mapas de interatómica gerados permitem visualizar de forma mais directa as relações proteicas do oraloma da Diabetes Melitos tipo 1 e 2.

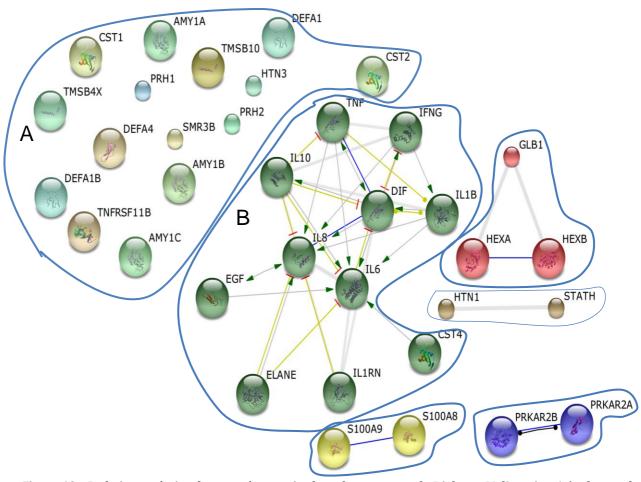


Figura 18 - Rede interatómica das proteínas orais alteradas em casos de Diabetes Melitos tipo 1 (rede gerada com recurso à ferramenta STRING; Clustering: MCL; Confidence > 0,7)

Na figura 18 está representada a rede interatómica das proteínas alteradas em casos de Diabetes Melitos tipo 1. No grupo A estão agrupadas as proteínas que não interagem com nenhuma outra ou o nível de confiança dessa interação encontra-se abaixo do valor estipulado como limite de

confiança: 0,7. Nos restantes grupos de proteínas encontram-se as proteínas que interagem com uma confiança superior a 0,7.

O grupo B representa as interacções entre as citocinas, um processo fundamental devido à DMT1 ser uma doença autoimune. É possível verificar que as interações representadas favorecem os processos inflamatórios.

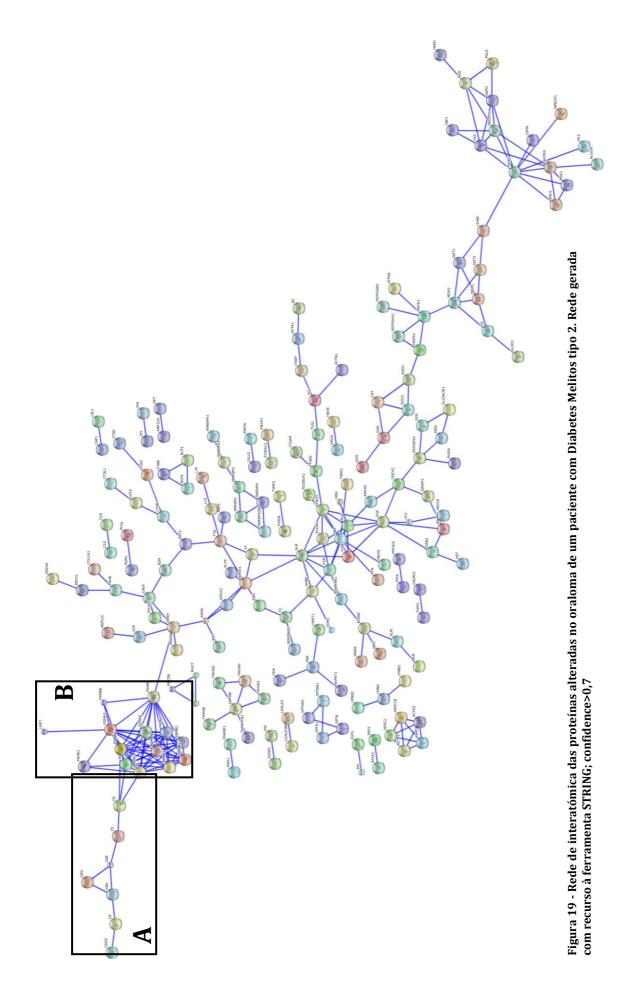
Nos estudos que quantificaram as subunidades A e B da betahexosaminidase (HEXA e HEXB, respectivamente) verificou-se que a sua regulação era superior em indivíduos fumadores, o mesmo se constata em estudos de proteómica da urina excretada por pacientes diabéticos<sup>73</sup>; esta alteração, tendo em conta a interatómica que demonstra a reação da betahexosaminidase e a beta-galactosidase (GLB1), pode inferir-se que existem efeitos na matriz extracelular provocados pela sobre-expressão desta proteína.

As proteínas *histatin-1* e *statherin* (HST1 e STATH, respectivamente) encontram-se ambas sub-reguladas. A primeira (*histatin-1*) é uma precursora da camada protectora proteica formada na superfície dentária e também exibe propriedades anti-fúngicas e anti-bacterianas<sup>39,40</sup>. Já a *statherin* estabiliza a saliva supersaturada com sais de cálcio, inibindo a sua precipitação. Esta proteína influencia também a formação dos cristais de hidroxiapatite na superfície dentária<sup>39,40</sup>. Estudos comprovam que a Diabetes Melitos afecta negativamente a expressão de proteínas salivares como as apresentadas<sup>39,39,45</sup>. Os níveis diminuídos de *statherin* podem mesmo estar relacionados com um aumento da prevalência de lesões da cavidade oral em pacientes diabéticos<sup>40</sup>.

As proteínas S100A8/9 formam um complexo denominado *calprotectin* e são proteínas pro-inflamatórias que desempenham papéis importantes na quimiotaxia<sup>74</sup>. Sendo uma doença autoimune, verifica-se que a regulação destas proteínas encontra-se aumentada no oraloma de pacientes com Diabetes Melitos tipo 1.

A via de sinalização da Proteína Cinase A (dependente de AMP cíclico) é referente à sinalização intracelular, promovendo os processos de gluconeogénese<sup>75</sup>. É, então, expectável que esta via de sinalização esteja exacerbada face à falta de glicose intracelular para produção de energia (ATP) nos casos de Diabetes Melitos tipo 1. Isto é confirmado pelos dados

disponíveis que mostram que estas duas subunidades, (PRKAR2A/B) estão sobre-reguladas (regulação:1,3532) no oraloma deste tipo de Diabetes, contudo mais uma vez se conclui que seria preciso dispor de mais proteínas anotadas que participassem nesta via de sinalização para poder afirmar que a mesma se encontra alterada positivamente.



O oraloma da Diabetes Melitos tipo 2 é numeroso consistindo em 503 proteínas, o que tem como consequência uma rede interatómica bastante complexa (Figura 19).

Na figura 19-A estão patentes parte das proteínas do sistema complemento, envolvido na coagulação sanguínea. Já na figura 19-B é representado o proteassoma, um complexo proteico responsável por degradar proteínas sinalizadas como dispensáveis por ubiquitinas.

## 4.2.4 Comparação da microflora oral da Diabetes Melitos tipo 1 e 2

A cavidade oral é um ecossistema composto não só por tecidos e moléculas de origem humana como também pelas bactérias que nele se incluem, consequentemente existe uma reciprocidade de alterações no indivíduo e na microflora oral consoante as condições do meio, pH, temperatura, patologia do indivíduo, entre outros. Como corolário, a quantificação das diferentes espécies de microrganismos, pode ser característica de uma patologia que, alterando as condições orais do indivíduo, influi a prevalência e proliferação/diminuição de uma ou outra espécie microbiana.

Na Diabetes Melitos tipo 2 foram anotadas 146 entradas correspondentes a um total de 61 microrganismos dos domínios eucariota e bateriano provenientes de 16 artigos científicos<sup>76–91</sup>. Estes analisaram faixas etárias diferentes sendo que as idades estão compreendidas entre os 23 e os 88 anos, não havendo distinção quanto ao género dos indivíduos. 72 anotações correspondem a indivíduos não fumadores, 32 correspondem a fumadores e as restantes não possuem informações quanto aos hábitos sociais da população estudada.

Na Diabetes Melitos tipo 1 foram anotados 4 microrganismos a partir de 3 artigos científicos<sup>92–94</sup> que não mencionavam hábitos sociais sendo que de todos nenhum utilizou indivíduos com mais de 18 anos.

Todos os microrganismos anotados na DMT1 estão também anotados na DMT2. Disto resulta que à luz da literatura actual apenas 4 espécies de microrganismos estão identificados como alterados na cavidade oral de ambos os tipos de Diabetes, nomeadamente: A. Actinomycetemcomitans, prevotella intermedia, Capnocytophaga spp., candida sp..

Tabela 2 - Microrganismos comuns anotados que foram quantificados na cavidade oral de pacientes com ambos os tipos de Diabetes. (\*Constant site - Face mesio-vestibular do primeiro molar superior direito; Deepest site - local mais profundo à sondagem)

		A. Actinomycetem		Capnocytophaga		Candida
		comitans	Prevotella	spp.	₽	sp.
	Article ID		Intermedi		Article ID	
	Ā		а		Arti	
		С	onstant site*			Mouth
Diabetes Melitos tipo 1		S O NO SAIN S O O				Rinse
	10	1,56		1,46	0.1	12,08
	7562335	.,		1,12	1619012	-,-,-
					161	
		Deepest site*				
S M		-5,6	0	1		
cete		Constant site*				
Diat	10	-6,5	-1,02	1,48		
	9526910		eepest site*	.,		
	92		-1,92	-1,35		
		-4,9				Oaline tatal
		Placa bacteriana supragengival				Saliva total
		-1,02	1,58		~	aumentada
	43	·	•		17488443	
	17488443				748	
		Diagraphy	4	an ab cal	<del>-</del>	
		Placa bacteriana subgengival				
tipo 2		1,07   1,43				
	_	Bolsas subgengivais				
	15677809					
litos		Alterações não signifi	cativas de			
Diabetes Melitos tipo 2		acordo com o estudo				
	- 86	Bolsas subgengivais				
	16579698		1.00			
		-1,84	-1,09			
	`	Bolsas gengivais saudáveis				
	18380556	aumentada	aumentada	diminuída		
		Polege subge	ngivais com n	periodontita		
	183	Bolsas subgengivais com periodontite				
			aumentada	diminuída		

A comparação do microbiota oral dos dois tipos de Diabetes Melitos encara diversos vieses que acabam por impossibilitar a inferência de factos prováveis. Nem todos os estudos que quantificam as mesmas bactérias em ambos os tipos de Diabetes o fazem de forma absoluta (indicando um valor específico para a concentração de certa estirpe bacteriana), os locais anatómicos de recolha da amostra não são os mesmos e a população estudada na Diabetes Melitos tipo 2 tem mais idade e certos hábitos, como fumar que não se verificam na população (na sua maioria) adolescente estudada na Diabetes Melitos tipo 1.

Conhecer o proteoma dos microrganismos característicos dos dois tipos de Diabetes Melitos e a interacção de tais proteínas com as proteínas do hospedeiro pode clarificar aspectos importantes destas patologias, nomeadamente a sua etiologia e os seus efeitos na cavidade oral. O grupo de investigação do Instituto de Ciências da Saúde da Universidade Católica Portuguesa - Centro Regional das Beiras está no presente a desenvolver uma ferramenta que permitirá efectuar esta análise.

## 5 Conclusão

A primeira conclusão possível após elaboração deste estudo é a necessidade de mais estudos de proteómica oral em pacientes com Diabetes Melitos tipo 1. Esta patologia conta com um reduzido número de proteínas anotadas comparativamente à Diabetes Melitos tipo 2. Deste facto advém que as possíveis inferências a retirar de uma comparação entre os oralomas dos dois tipos de Diabetes Melitos estão limitados pela dimensão dos mesmos.

É igualmente necessário estandardizar os métodos de colheita e análise em estudos de proteómica da cavidade oral, tendo em vista uma maior proporcionalidade entre os valores obtidos pelos diversos estudos, eliminando desta forma vieses.

Molecularmente, após a análise destas duas entidades patológicas, a Diabetes Melitos tipo 1 e tipo 2, conclui-se que tanto diferenças como semelhanças se refletem no conteúdo proteíco da cavidade oral. A dificuldade em estabelecer diferenças moleculares entre os dois tipos de Diabetes Melitos prende-se com o facto de no caso da Diabetes Melitos tipo 2 esta só ser diagnosticada tardiamente quando as complicações decorrentes da Diabetes crónica estão já instaladas e os estudos de Diabetes Melitos tipo 1 estudarem indivíduos com uma média de idades inferior a 20 anos perto do onset da doença.

Uma das principais conclusões deste estudo é que existem duas fases na Diabetes Melitos tipo 2 que afectam os dados de proteómica gerados. Uma primeira fase em que se dá o *onset* da patologia e uma segunda fase em que os efeitos no oraloma se devem ao estado hiperglicémico crónico e à acumulação de AGE's, sendo esta segunda fase um elemento de semelhança em relação à Diabetes Melitos tipo 1.

Actualmente, à luz dos dados disponíveis, verifica-se que no oraloma da Diabetes Melitos tipo 1 são mais abundantes proteínas com funções imunológicas e de sinalização celular, ao passo que no oraloma da Diabetes Melitos tipo 2 verifica-se uma maior concentração de proteínas catalíticas.

Após este estudo comparativo é possível afirmar que a saliva reflecte, pelo menos, parcialmente as implicações sistémicas da patofisiologia da Diabetes Melitos tipo 1 e 2, como é reflectido pela comparação ao oraloma normal em que, por exemplo, os processos biológicos de *immune system process* se mostraram aumentados.

Entre as duas doenças, o cálculo de diferença fraccional mostrou sempre que os parâmetros ontológicos da DMT1 se encontram aumentados face aos da DMT2, o que pode ser atribuido à diferença de dimensão entre os dois oralomas.

O presente trabalho científico contribuiu para a actualização da base de dados do Oralome, interpretando a informação obtida com ferramentas bioinformáticas e cálculo de diferença fraccional. Culminou na constatação de que certas proteínas séricas caracteristicamente alteradas nos dois tipos de Diabetes Melitos estão também alteradas na saliva podendo esta ser utilizada como fluído de diagnóstico quando mais informação estiver disponível neste campo.

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# 7 Anexos

7.1 Anexo 1 - Tabela do oraloma da DMT1

7.2 Anexo 2 - Tabela do oraloma da DMT2

# 7.3 Anexo 3 - Comparação DMT1-DMT2

Biological Process	DMT2	DMT1 esperado	DMT1 observado	Diferença fracional	P value
cell communication	85	1,031	13	11,61	0,002462245
cellular process	118	1,431	13	8,08	0,040874835
apoptosis	22	0,267	5	17,74	0,016482194
reproduction	12	0,146	1	5,87	0,569442458
developmental process	52	0,631	3	3,76	0,715785571
metabolic process	221	2,680	11	3,10	0,96307574
cell cycle	33	0,400	3	6,50	0,402741419
immune system process	105	1,274	11	7,64	0,086762533
cell adhesion	37	0,449	1	1,23	0,931509418
response to stimulus	72	0,873	8	8,16	0,111240069
Total Oraloma normal	419				
Total Diabetes 1	29				

Função Molecular	DMT2	DMT1 esperado	DMT1 observado	Diferença fracional	p-value
binding	146	1,770807194	15	7,47	0,045866893
catalytic activity	182	2,207444584	3	0,36	0,999982504
enzyme regulator activity	46	0,557925554	5	7,96	0,207615796
receptor activity	35	0,424508574	3	6,07	0,440893433

Pathway	DMT2	DMT1 esperado	DMT1 observado	Diferença fracional	
Alzheimer disease-presenilin pathway	4	0,05	2	40,22	0,03
Apoptosis signaling pathway	3	0,04	1	26,48	0,19
Endothelin signaling pathway	2	0,02	2	81,45	0,01
Gonadotropin releasing hormone receptor pathway  Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha	6	0,07	3	40,22	0,01
mediated pathway	3	0,04	2	53,97	0,02
Inflammation mediated by chemokine and cytokine signaling pathway	12	0,15	4	26,48	0,01
Interleukin signaling pathway	2	0,02	3	122,67	0,00
Interferon-gamma signaling pathway	1	0,01	1	81,45	0,07
Wnt signaling pathway	2	0,02	1	40,22	0,13

#### 7.4 Anexo 4 - DMT1 comparado ao OralOma normal

Dialogical Dragons	Oraloma	Diabetes	Diabetes	Diferença	Duralus
Biological Process	Normal	esperado	observado	fracional	P value
cell communication	544	6,598	13	0,97	0,007013492
cellular process	912	11,061	13	0,18	0,287905016
apoptosis	142	1,722	5	1,90	0,026555207
reproduction	109	1,322	1	-0,24	0,741568258
developmental process	469	5,688	3	-0,47	0,942642667
metabolic process	1307	15,852	11	-0,31	0,977046763
cell cycle	269	3,263	3	-0,08	0,648398234
immune system process	459	5,567	11	0,98	0,01466063
cell adhesion	219	2,656	1	-0,62	0,938322452
Total Oraloma normal	2391				
Total Diabetes 1	29				

Função Molecular	Oraloma Normal	Diabetes esperado	Diabetes observado	Diferença fracional	
binding	965	11,70	15	0,28	0,15
catalytic activity enzyme regulator	946	11,47	3	-0,74	1,00
activity	202	2,45	5	1,04	0,09
receptor activity	188	2,28	3	0,32	0,40

Pathway	Oraloma Normal	Diabetes esperado	Diabetes observado	Diferença fracional	
Alzheimer disease-presenilin pathway	23	0,28	2	6,17	0,03
Apoptosis signaling pathway	23	0,28	1	2,58	0,24
Beta1 adrenergic receptor signaling pathway	16	0,19	2	9,31	0,02
Beta2 adrenergic receptor signaling pathway	16	0,19	2	9,31	0,02
5HT1 type receptor mediated signaling pathway	13	0,16	2	11,68	0,01
Endothelin signaling pathway	17	0,21	2	8,70	0,02
Enkephalin Release	13	0,16	2	11,68	0,01
GABA-B receptor II signaling	10	0,12	2	15,49	0,01
Hedgehog signaling pathway	3	0,04	2	53,97	0,00
Gonadotropin releasing hormone receptor pathway	58	0,70	3	3,26	0,03
Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pathway	29	0,35	2	4,69	0,05
Inflammation mediated by chemokine and cytokine signaling pathway	74	0,90	4	3,46	0,01
Interleukin signaling pathway	14	0,17	3	16,67	0,00
Histamine H2 receptor mediated signaling pathway	9	0,11	2	17,32	0,01
Interferon-gamma signaling pathway	4	0,05	1	19,61	0,05
Metabotropic glutamate receptor group I pathway	3	0,04	2	53,97	0,00
Metabotropic glutamate receptor group II pathway	16	0,19	2	9,31	0,02
Metabotropic glutamate receptor group III pathway	12	0,15	2	12,74	0,01

Muscarinic acetylcholine receptor 2 and 4 signaling pathway	16	0,19	2	9,31	0,02
Transcription regulation by bZIP transcription factor	4	0,05	2	40,22	0,00
Wnt signaling pathway	43	0,52	1	0,92	0,41

### 7.5 Anexo 5 - DMT2 comparada ao oraloma normal

				_	
	Oraloma	Diabetes	Diabetes	Diferença	Р
Biological Process	Normal	esperado	observado	fracional	value
apoptosis	127	22,780	24	0,05	0,43
cell adhesion	194	34,797	48	0,38	0,02
cell communication	503	90,221	101	0,12	0,11
cell cycle	237	42,510	39	-0,08	0,74
cellular component organization	304	54,527	37	-0,32	1,00
cellular process	843	151,206	141	-0,07	0,86
developmental process	411	73,720	64	-0,13	0,91
generation of precursor metabolites and					
energy	89	15,964	14	-0,12	0,73
homeostatic process	30	5,381	5	-0,07	0,63
immune system process	398	71,388	120	0,68	0,00
localization	27	4,843	4	-0,17	0,71
metabolic process	1261	226,181	250	0,11	0,01
reproduction	68	12,197	14	0,15	0,34
response to stimulus	286	51,299	84	0,64	0,00
system process	281	50,402	49	-0,03	0,61
transport	459	82,329	68	-0,17	0,97
Total Oraloma normal	2336				
Total Diabetes	419				

	Oralom	Diabetes	Diabetes	Diferenç	
	a	esperad	observad	а	Р
Função Molecular	Normal	0	0	fracional	value
antioxidant activity	16	2,87	8	1,79	0,01
binding	910	163,22	146	-0,11	0,96
catalytic activity	909	163,04	182	0,12	0,03
enzyme regulator activity	181	32,47	46	0,42	0,01
ion channel activity	27	4,84	3	-0,38	0,86
motor activity	30	5,38	2	-0,63	0,97
receptor activity	167	29,95	35	0,17	0,19
structural molecule activity	426	76,41	58	-0,24	0,99
transcription regulator activity	86	15,43	3	-0,81	1,00
translation regulator activity	45	8,07	4	-0,50	0,96
transporter activity	149	26,73	26	-0,03	0,59

Total Oraloma normal	2336
Total Diabetes	419

		1		1	
Pathway	Oraloma Normal	Diabetes esperado	Diabetes observado	Diferença fracional	P value
5-Hydroxytryptamine degredation	10	1,79	1,00	-0,44	0,83
Adenine and hypoxanthine salvage pathway	3	0,54	2,00	2,72	0,10
Alzheimer disease-amyloid secretase pathway	9	1,61	2,00	0,24	0,48
Alzheimer disease-presenilin pathway	22	3,95	4,00	0,01	0,56
Angiogenesis	28	5,02	3,00	-0,40	0,88
Angiotensin II-stimulated signaling through G proteins and beta-arrestin	15	2,69	3,00	0,12	0,50
Apoptosis signaling pathway	25	4,48	3,00	-0,33	0,83
Asparagine and aspartate biosynthesis	2	0,36	2,00	4,58	0,05
ATP synthesis	5	0,90	1,00	0,12	0,59
Axon guidance mediated by netrin	7	1,26	2,00	0,59	0,36
Axon guidance mediated by semaphorins	8	1,43	1,00	-0,30	0,76
Axon guidance mediated by Slit/Robo	8	1,43	1,00	-0,30	0,76
B cell activation	16	2,87	3,00	0,05	0,55
Blood coagulation	16	2,87	10,00	2,48	0,00
Cadherin signaling pathway	19	3,41	2,00	-0,41	0,86
Cytoskeletal regulation by Rho GTPase	50	8,97	7,00	-0,22	0,79
De novo purine biosynthesis	12	2,15	1,00	-0,54	0,88
Dopamine receptor mediated signaling pathway	25	4,48	1,00	-0,78	0,99
EGF receptor signaling pathway	35	6,28	8,00	0,27	0,29
Endothelin signaling pathway	16	2,87	2,00	-0,30	0,78
FAS signaling pathway	9	1,61	3,00	0,86	0,22
FGF signaling pathway	31	5,56	9,00	0,62	0,11
Fructose galactose metabolism	4	0,72	1,00	0,39	0,51
Glutamine glutamate conversion	3	0,54	1,00	0,86	0,42
Glycolysis	17	3,05	5,00	0,64	0,19
Gonadotropin releasing hormone receptor pathway	55	9,87	6,00	-0,39	0,93
Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pathway	27	4,84	3,00	-0,38	0,86
Heterotrimeric G-protein signaling pathway-Gq alpha and Go alpha mediated pathway	21	3,77	2,00	-0,47	0,89
	10	1,79	1,00	-0,44	0,83
Heterotrimeric G-protein signaling pathway-rod outer segment phototransduction  Huntington disease	59	10,58	9,00	-0,15	0,73
Inflammation mediated by chemokine and cytokine signaling pathway	74		12,00		0,68
Insulin/IGF pathway-mitogen activated protein kinase kinase/MAP kinase cascade	9	13,27	1,00	-0,10 -0,38	0,80
Integrin signalling pathway	84	1,61 15,07	16,00	0,06	0,80
Interferon-gamma signaling pathway	3		1,00		0,44
Interleukin signaling pathway	13	0,54	2,00	0,86	0,42
N-acetylglucosamine metabolism	2	2,33		-0,14	0,30
		0,36	1,00	1,79	
Nicotine pharmacodynamics pathway	14	2,51	1,00	-0,60	0,92
Nicotinic acetylcholine receptor signaling pathway	36	6,46	1,00	-0,85	1,00
p38 MAPK pathway	9	1,61	2,00	0,24	0,48
p53 pathway	15	2,69	1,00	-0,63	0,93
Parkinson disease	36	6,46	18,00	1,79	0,00
PDGF signaling pathway	18	3,23	1,00	-0,69	0,96
Pentose phosphate pathway	5	0,90	2,00	1,23	0,23
Phenylalanine biosynthesis	2	0,36	2,00	4,58	0,05
P13 kinase pathway	14	2,51	1,00	-0,60	0,92
Plasminogen activating cascade	8	1,43	4,00	1,79	0,06
Purine metabolism	1	0,18	2,00	10,15	0,01

Pyridoxal phosphate salvage pathway	1	0,18	1,00	4,58	0,16
Pyrimidine Metabolism	6	1,08	1,00	-0,07	0,66
Pyruvate metabolism	7	1,26	3,00	1,39	0,13
Ras Pathway	20	3,59	3,00	-0,16	0,70
Salvage pyrimidine deoxyribonucleotides	2	0,36	1,00	1,79	0,30
Salvage pyrimidine ribonucleotides	6	1,08	2,00	0,86	0,29
Synaptic_vesicle_trafficking	8	1,43	1,00	-0,30	0,76
T cell activation	20	3,59	5,00	0,39	0,29
TCA cycle	6	1,08	2,00	0,86	0,29
TGF-beta signaling pathway	9	1,61	2,00	0,24	0,48
Toll receptor signaling pathway	10	1,79	3,00	0,67	0,27
Tyrosine biosynthesis	2	0,36	2,00	4,58	0,05
Ubiquitin proteasome pathway	24	4,30	4,00	-0,07	0,62
Vasopressin synthesis	5	0,90	2,00	1,23	0,23
VEGF signaling pathway	12	2,15	2,00	-0,07	0,63
Vitamin B6 metabolism	1	0,18	1,00	4,58	0,16
Wnt signaling pathway	41	7,35	2,00	-0,73	0,99
Xanthine and guanine salvage pathway	2	0,36	2,00	4,58	0,05

Total Oraloma normal	2336
Total Diabetes	419

UniProtKBAC	Name	Parotid	Whole Saliva	Crevicular Fluid	Mucosa	Health	Disease (MeSH ID)	Regulation	Age group	Gender*	Social Habits*	Methods of Sampling**	Methods of Analysis***	Type of Study
F6KPG5	Albumin		X			х	68003922	-0,3	4-17	MF	otherwise healthy children, healthy children in the control		Protein-dye binding	Cross-sectional
000300	Tumor necrosis factor receptor superfamily member 118 (Osteoclastogenesis inhibitory factor) (Osteoprotegerin)				x	х	68003922	-1,6		MF	non-smokers. healthy individuals with no medication which could influence periodontitis except for diabetes type 1 medication, not pregnant womenwithout using any contraceptive treatment or lactant. All non-smoking individuals	gingival tissue samples	RNA extraction and measured by biophotometer	Proteomics
P01036	Cystatin-S (Cystatin-4) (Cystatin-SA-III) (Salivary acidic protein 1)		х			x	68003922		Children					
P01037	Cystatin-SN (Cystain-SA-I) (Cystatin-1) (Salivary cystatin-SA-1)		х			х	68003922		Children					
P01133	Pro-epidermal growth factor (EGF) [Cleaved into: Epidermal growth factor (Urogastrone)]		х			х	68003922	-0,3	48,7	MF	periodontally healthy, excluded: patients under pharmacological treatment, hormonal contraception, pregnants.		Quantikine Human EGF Immunoassay	
P01375	Tumor necrosis factor				x	x	68003922	-0,4		MF	non-smokers. healthy individuals with no medication which could influence periodontitis except for diabetes type 1 medication, not pregnant womenwithout using any contraceptive treatment or lactant. All non-smoking individuals	gingival tissue samples	RNA extraction and measured by biophotometer	Proteomics
P01579	Interferon gamma				х	x	68003922	2,3		MF	non-smokers. healthy individuals with no medication which could influence periodontitis except for diabetes type 1 medication, not pregnant womenwithout using any contraceptive treatment or lactant. All non-smoking individuals	gingival tissue samples	RNA extraction and measured by biophotometer	Proteomics
P01584	Interleukin-1 beta (IL-1 beta) (Catabolin)				х	x	68003922	5		MF	non-smokers. healthy individuals with no medication which could influence periodontitis except for diabetes type 1 medication, not pregnant womenwithout using any contraceptive treatment or lactant. All non-smoking individuals	gingival tissue samples	RNA extraction and measured by biophotometer	Proteomics
P01876	Ig alpha-1 chain C region		х			х	68003922	-0,3	10-19	MF	orherwise Healthy	gg	direct enzyme-linked immunosorbent assay	non-Proteomics
P02808	Statherin		х			х	68003922	-2,1	Children					
P02810	Salivary acidic proline-rich phosphoprotein 1/2 (Db-s)		x			х	68003922	-1,2	Children					
P02814	Submaxillary gland androgen- regulated protein 3B		х			х	68003922	-1,3	Children					
P04745	Alpha-amylase 1	Х				Х	68003922	3,2	42-68	MF		tissue samples	Immunohistochemistry	non-Proteomics
P05109	Protein S100-A8		х		x	x	68003922 68003922	29,2	Children	MF	non-smokers. healthy individuals with no medication which could influence periodontitis except for diabetes type 1 medication, not pregnant womenwithout using any contraceptive treatment or lactant. All non-smoking individuals	gingival tissue samples	RNA extraction and measured by biophotometer	Proteomics
P06702	Protein S100-A9		х			х	68003922	1,6	Children					
P06865	Beta-hexosaminidase subunit alpha		х			х	68003922	1,3	20-35	MF	non-smokers		Chatteriee et al. Method (1975)	Proteomics
P06865	Beta-hexosaminidase subunit alpha		х			х	68003922	2,3	20-35	MF	Smokers. periodontally healthy, excluded: patients under pharmacological treatment, hormonal contraception, pregnants.		Chatteriee et al. Method (1975)	Proteomics
P07686	Beta-hexosaminidase subunit beta		х			x	68003922	-0,12	20-35	MF	non-smokers. periodontally healthy, excluded: patients under pharmacological treatment, hormonal contraception, pregnants.		Chatteriee et al. Method (1975)	Proteomics
P07686	Beta-hexosaminidase subunit beta		х			х	68003922	3	20-35	MF	smokers. periodontally healthy, excluded: patients under pharmacological treatment, hormonal contraception, pregnants.		Chatteriee et al. Method (1975)	Proteomics
P08246	Neutrophil elastase			х		х	68003922	1,1	26-51	MF	average was a smoker for 6,5+-3,2 yrs. dmt-1 patients for 10,03+-5,73 yrs		ELISA	non-Proteomics
P09228	Cystatin-SA (Cystatin-2)		Х			х	68003922		Children					
P10145	Interleukin-8 (IL-8)				x	x	68003922	35		MF	non-smokers. healthy individuals with no medication which could influence periodontitis except for diabetes type 1 medication, not pregnant womenwithout using any contraceptive treatment or lactant. All non-smoking individuals	gingival tissue samples	RNA extraction and measured by biophotometer	Proteomics
P12838	Neutrophil defensin 4 (Defensin, alpha 4) (HNP-4) (HP-4)		x			х	68003922	1,9	Children					
	/\ /		•	•								•	•	•

	Taken I I I I I I I	1	1		1	1	1		1				1	I .
D4 20C4	cAMP-dependent protein kinase type	x				x	68003922	1,4	42-68	MF		Air		Dti
P13861 P15515	II-alpha regulatory subunit Histatin-1				1		68003922	-2.2	Children			tissue samples	Immunohistochemistry	non-Proteomics
			х		ļ	х								
P15516	Histatin-3		x			x	68003922	-1,1	Children					
											non-smokers. periodontally healthy, excluded: patients under			
			х			х	68003922	-0,3	20-35	MF	pharmacological treatment, hormonal contraception,		Chatteriee et al. Method	
P16278	Beta-galactosidase										pregnants.		(1975)	Proteomics
											smokers. periodontally healthy, excluded: patients under			
			×			×	68003922	-0,23	20-35	MF	pharmacological treatment, hormonal contraception,		Chatteriee et al. Method	
P16278	Beta-galactosidase										pregnants.		(1975)	Proteomics
											non-smokers. healthy individuals with no medication which		,	
											could influence periodontitis except for diabetes type 1			
					×	x	68003922	1.7		MF	medication, not pregnant womenwithout using any		RNA extraction and	
	Interleukin-1 receptor antagonist							-,-			contraceptive treatment or lactant. All non-smoking		measured by	
P18510	protein (IL-1RN)										individuals	gingival tissue samples	biophotometer	Proteomics
F 10310	protein (it-1kiv)										non-smokers. healthy individuals with no medication which	girigival cissue samples	biopriotoinetei	FIOLEOIIIICS
											could influence periodontitis except for diabetes type 1			
					×	х	68003922	-1,8		MF	medication, not pregnant womenwithout using any		RNA extraction and	
											contraceptive treatment or lactant. All non-smoking		measured by	
P22301	Interleukin-10 (IL-10)										individuals	gingival tissue samples	biophotometer	Proteomics
	cAMP-dependent protein kinase type					.,	68003922	1.4	42-68	MF				
P31323	II-beta	×				X	00003922	1,4	42-06	IVIF		tissue samples	Immunohistochemistry	non-Proteomics
P59665	Neutrophil defensin 1		х			х	68003922	1,9	Children					
P59666	Neutrophil defensin 3		х			х	68003922	+	Children					
P62328	Thymosin beta-4		х			х	68003922		Children					
P63313	Thymosin beta-10		Х		1	х	68003922		Children					
		ĺ		1	1								direct enzyme-linked	
Q9Y298	IgG VH protein		х			х	68003922	2,4	10-19	MF	otherwise Healthy		immunosorbent assay	non-Proteomics

Citation (NCBI ID)

UniProtK B AC	Parotid	SM/SL	Whole Saliva	Health	Regulation	Age group	Gender	Social Habits	Methods of Sampling	Methods of Analysis	PTM	Biomarker	Citation (NCBI ID)
			Juliva					Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			()
			x	x	-2,3	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			1
P62258		1					-	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	<u> </u>		19118452
			x	x	-3	36-62	M/F	alcoholic: no	Onsumurated salva (2011). 5 nether, at a a.m. (Defore or leakings), hie subjects were asked to mise their modulis full origing with water, then to until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at 4.80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
Q04917							, i	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis			19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			í
P61981			х	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			19118452
P61981						+		pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and		+	19118452
			x	x		36-62	M/F	alcoholic: no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P31947								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis			19118452
						00.00		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			1
P63104			x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis			19118452
103104								Non-smoker: Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			13110432
			x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P05387								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis			19118452
			×			36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			1
P52209			*			30-02	IVI/F	pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80. C until	spectrometry (LC-MS/MS) analysis			19118452
						1		Non-smoker; Non-	Grieniaus. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			
			x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
095336								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and			19118452
			x	v		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P11021	l	1	*			30-02	IVI/F	alconolic; no pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	1		19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			
			х	x	3,3	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P39687		$\vdash$				1	<b>—</b>	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	1	+	19118452
			×	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P68133			^	~		30 02	,.	pregnancy	analysis	spectrometry (LC-MS/MS) analysis			19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			í
			х	x	3,4	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			f
P61160								pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and			19118452
			x	x		36-62	M/F	alcoholic: no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
015143			^	~		30 02	,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis			19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			í
015144			х	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis			19118452
015144						-		pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and		_	19118452
			x	x	6.1	36-62	M/F	alcoholic: no	Onsumurated salva (2011). 5 nether, at a a.m. (Defore or leakings), hie subjects were asked to mise their modulis full origing with water, then to until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at 4.80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
015511								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis			19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			ł
P61158			х	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis			19118452
r01130								pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			19110432
			x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P07108								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis			19118452
						25.52		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass			ł
P23526			x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis			19118452
LJJLO								Non-smoker; Non-	Justimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			15110452
			x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
Q01518						1		pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	<b> </b>		19118452
	l	1	×	×		36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their based forward and allow saliva to flow the act and a stable container for E-min. There consider a water immediately frage and stored at 80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		i
Q10588	l	1	*	×		50-02	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DEC) and EC-tandem mass spectrometry (LC-MS/MS) analysis	1		19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	1		
	l	1	x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
P43652		1						pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	<del>                                     </del>		19118452
	l	1 1	×			36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
P14550	l	1	*			30-02	IVI/F	alconolic; no pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	1		19118452
						1		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	1		
			x	×	-2,2	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
P30838		1						pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	<del> </del>	+	19118452
			v		-3.3	36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
060218			^	^	-5,5	30-02	141/1	pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until analysis.	spectrometry (LC-MS/MS) analysis	1		19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			i
	l	1	x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
Q5STX8		1				1	-	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	1	+	19118452
			v			36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
P02763	l	1	*			30-02	IVI/F	alconolic; no pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	1		19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and			
	l	1	x	x	3,2	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1 .
P01009		$\vdash \vdash \vdash$				1	<b>—</b>	pregnancy Non smokers Non	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	1	+	19118452
	l	1	×	×		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	1		1
P04217	l	1		^		-502	,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	1		19118452
PU421/								p			•	_	-

				- 1		1		Non-smoker; Non-		
			х	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid formatography (2-DLC) and LC-tandem mass speciments (LC-MS/MS) analysis	
P02765								pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m., (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till.  In this control of the	19118452
			×	x	2,2	36-62	M/F	alcoholic; no	bisinitiated same continued and allowed the case of the continued and allowed the case of the continued and allowed the case of the case of the continued and allowed the case of the continued and allowed the case of the ca	
P01023								pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
						36-62	M/F	Non-smoker; Non-		
A8K2U0		-	*	x		30-02	IVI/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	10 in a 1/2	
		:	x	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P12814								pregnancy Non-smoker; Non-	analysis.  [Instrimulated salva (20m1). Briefly, at 8 a.m. (hefore hreakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit.  Five salivas samples each from the control and DM groups were pooled separately and	19118452
			×	x		36-62	M/F	alcoholic: no	Unstimulated saliva (20m.), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at 8.0°C until sliguid chromatopal liquid chromatoparphy (2-bLC) and LC-tandem mass	
P06733							ŕ	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five Saliva samples each from the control and DM groups were pooled separately and	
P54802		:	x	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid formatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
134002		-						Non-smoker; Non-	aliasyss. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lill. Five salivas assings each from the control and DM groups were pooled separately and	15110452
		:	×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P01019								pregnancy	analysis.  Illustimulated salika (20mi) Briefly at 8.3 m. (hefore breakfast) the subjects were asked to risse their mouths thoroughly with water than in til.  Fig. 381/88 samples seek from the control and DM groups were gooled separately and	19118452
			,	×	-3,6	36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20m.), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These seceimens were immediately frozen and stored at -80°C und to two-dimensional liquid chromatography (2-LOL) and LC-tandem mass	
P04083			^	^	-5,0	30-02	141/1	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	
		:	×	х	-4,3	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applied to two-dimensional liquid chromatography (2-DLC) and LC-tandem m	
P07355		-	-	-				pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were pooled separately and the subjects were provided by the subject with the subject were provided by the subject win	19118452
			×	x	-2,7	36-62	M/F	alcoholic: no	Unsurfaceted salves (conting, or term), a count, in conting, a count, in conting, a count, in conting, a count, in conting, a count, in count, a	
P12429					,			pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20m.L). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	
P03973		:	×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen at -80 °C until subjected to two-d	19118452
PU39/3			-					pregnancy Non-smoker: Non-	Janaisss.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till five saliva samples each from the control and DM groups were pooled separately and	19110432
			×	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P01008								pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
						36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20m1). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt before saliva (20m1). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with vater, then to tilt before the saliva samples seach from the control and DM groups were pooled separately and their heads from your and allows affixed to two-dimensional liquid chromatography (2-DLC) and CL-tandem mass	
Q10567			*	x		36-62	IVI/F	alcoholic; no pregnancy	their neads forward and allow saliva to now into a sterile container for 5 min. Tinese specimens were immediately frozen and stored at -80 °C, until specimens to the control of the contr	19118452
								Non-smoker; Non-	olimits.  Univasimulated saliva (20mt). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
		:	x	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P02647								pregnancy	analysis.  [Instrimulated salva (20m1). Briefly, at 8 a.m. (hefore hreakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit.  Five salivas samples each from the control and DM groups were pooled separately and	19118452
				x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20m.), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit her heads forward and allows salive to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at 8.0°C until liquid thromatography (2-bLt) garagety (2-bLt) and L-Crandem mass	
P02652			^			30 02	,.	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	
P04114		:	×	х	-4,1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid formatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P04114			_					pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till.  In this water is the subject of the subject is the su	19118452
			×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P05090								pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
						25.52	/=	Non-smoker; Non-	Unstimulated saliva (20m). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse there mouths thoroughly with water, then to tilt  which heads forward and allow refulls to few ultra a perilacentaries for 5 min. These considerable from a soft and the saliva samples seach from the control and DM groups were pooled separately and  table heads forward and allow refulls to few ultra a perilacentaries for 5 min. These considerable from and cut forward and silver saliva samples seach from the control and DM groups were pooled separately and  table heads forward and allow refulls to 16 min the saliva strong and cut forward and saliva strong and saliva strong and cut forward and saliva strong and saliva strong and cut forward and saliv	
095445			×	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass speciment (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	olimits.  Univasimulated saliva (20mt). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
		:	x	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P08519								pregnancy	analysis.  Instrinciatoria (aliva (20m.)). Briefly, at 8.a.m. (hefore heakfast), the subjects were asked to risse their mouths thoroughly with water, then to till.  Five sallivas samples each from the control and DM groups were pooled separately and	19118452
			×	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20m.). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit her heads forward and allows allive 10 mil not a sterile container for 5 min. These seceimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and C-trandem mass	
Q9ULZ3							,.	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  five saliva samples each from the control and DM groups were pooled separately and	
P05089			×	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid formatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
ru5089								pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till.  In this control and DM groups were pooled separately and the subjects were asked to rinse their mouths thoroughly with water, then to till.	19118452
			×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P17174			_					pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
	1 7		_			20.00	20.00	Non-smoker; Non-	Unstimulated saliva (20m1). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt help the saliva samples seach from the control and DM groups were pooled separately and their heads fromward and allow saliva to few into a stella container for 5 min. These concinence were immediately frome and the control and DM groups were pooled separately and their heads fromward and allow saliva to few into a stella container for 5 min. These concinence were immediately frome and the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples seach from the control and DM groups were pooled separately and the saliva samples sa	
			x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis.	19118452
P00505		l l					-	Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till be a control of the contro	15110432
P00505									their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	
			x	x		36-62	M/F	alcoholic; no		
P00505 P06576			x	x		36-62	M/F	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
		:					,	pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	19118452
				×		36-62 36-62	M/F	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
P06576		:				36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and subjects on the control and DM groups were pooled separately and Five saliva samples each from the control and DM groups were pooled separately and Five saliva samples each from the control and DM groups were pooled separately and	
P06576 O75882		:					,	pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately froze and stored at 8.80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately froze and stored at 8.80 °C until 5 Five saliva samples each from the control and DM groups were pooled separately and 5 Five saliva samples sach from the control and DM groups were pooled separately and 5 subjected to 4 two-dimensional liquid chromatography (2-DLC) and LC-tandem mass 1 subjects were asked to rinse their mouths thoroughly with water, then to tilt 5 five saliva samples each from the control and DM groups were pooled separately and 5 subjected to 4 two-dimensional liquid chromatography (2-DLC) and LC-tandem mass 1 subjects were asked to rinse their mouths thoroughly with water, then to tilt 5 five saliva samples each from the control and DM groups were pooled separately and 5 subjects were asked to rinse their mouths thoroughly with water, then to tilt 5 five saliva samples each from the control and DM groups were pooled separately and 5 five saliva samples each from the control and DM groups were pooled separately and 5 five saliva samples each from the control and DM groups were pooled separately and 5 subjects were asked to rinse their mouths thoroughly with water, then to tilt 5 five saliva samples each from the control and DM groups were pooled separately and 5 subjects were asked to rinse their mouths thoroughly with water, then to tilt 5 five saliva samples each from the control and DM groups were pooled separately and 5 subjects were asked to rinse their mouths thoroughly with water, then to tilt 5 five saliva sample	19118452
P06576		:				36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non- alcoholic; no pregnancy	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit where heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis subjected to two-dimensional liquid prompts were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	
P06576 O75882		:	x		5.4	36-62	M/F	pregnancy Non-smoker; Non-alcoholic; no pregnancy Non-smoker; Non-alcoholic; no pregnancy Non-smoker;	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until standards aliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until standards. Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis spectrometry (LC-MS/MS)	19118452
P06576 O75882		:	x	x x	5,4	36-62 36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non- alcoholic; no pregnancy	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until spectrometry (LC-MS/MS) analysis  Septrometry (LC-MS/MS) analysis  Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Septrometry (LC-MS/MS) analysis  Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P06576 O75882 P20160		:	x	x x	5,4	36-62 36-62	M/F M/F	pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non- alcoholic; no pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  [Institution of the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  [Institution of the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appears to the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appears to two dimensional liquid chromatography (2-DLC) and LC-tandem mass appears to the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appears and the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appears and the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  [Institution of the control and DM groups were pooled se	19118452
P06576  075882  P20160  Q7M4Q5		:	x	x x	5,4	36-62 36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452 19118452 19118452
P06576 O75882 P20160		:	x	x x	5,4	36-62 36-62	M/F M/F	pregnancy Non-smoker; Non- alcoholic; no pregnancy	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452 19118452
P06576  075882  P20160  Q7M4Q5		:	x	x x	5,4	36-62 36-62	M/F M/F	pregnancy Non-smoker; Non- alcoholic; no alcoholic; no	Instimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt will be a subjected to two-dimensional liqui	19118452 19118452 19118452

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	х	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass socitometry (LC-MS/MS) analysis	19118452
	х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt wheir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens. The specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens until chromatography (2-DLC) and LC-tandem mass specimens	19118452
	x	x	5,5	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt water in the saliva samples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
	х	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Jalanays. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt being heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till Five salivas samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	x	-2,2	36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C units subjected to two dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	x	×	-7.7	36-62	M/F	pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
	v					pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
	^					pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
	^					pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
						pregnancy Non-smoker; Non-	analysis.  Spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	19118452
	^					pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
	x					pregnancy Non-smoker; Non-	analysis.  Spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	19118452
	х		3,8			pregnancy Non-smoker; Non-	analysis.  Spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	19118452
	x					pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	19118452
	х	x		36-62	M/F	alcoholic; no pregnancy Non-smoker; Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass analysis.  Spectrometry (LC-MS/MS) analysis  Pursually a samples each from the control and DM groups were pooled separately and	19118452
	х	х		36-62	M/F	alcoholic; no pregnancy Non-smoker; Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass analysis.  Journal of the stored of the st	19118452
	х	x		36-62	M/F	alcoholic; no pregnancy Non-smoker; Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were easked to rinse their mouths thoroughly with water, then to til.  Fire salivas amples each from the control and DM groups were pooled separately and	19118452
	х	x		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were easked to rinse their mouths thoroughly with water, then to till. Five salivas amples each from the control and DM groups were pooled separately and	19118452
	х	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	х	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	х	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Brietly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  spectrometry (LC-MS/MS) analysis	19118452
			X	x       x         x	x       x       x       36-62         x       x       x       x         x       x       x       36-62         x	x       x       x       36-62       M/F         x       x       x       5,5       36-62       M/F         x       x       x       x       36-62       M/F <td< td=""><td>  X</td><td>  1</td></td<>	X	1

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	Q9UBR2											19118452
1				v			26.62	N4/F				
Section   Process   Proc	ОбУНКЗ			×	×		30-02	IVI/F		their needs forward and anow salve to now into a stellar container for 5 min. These specimens were immediately note in an a stored at 50°C and		19118452
										Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt		
1				x	×		36-62	M/F				
1	Q1T7A3						<u> </u>			undryss.		19118452
1				×	×		36-62	M/F		onstandated stand (Lone), or one in the control of canada, the stable of white distribution in the control of the stable of of the stab		
1	043866			^	^		30 02	,.		analysis.	pectrometry (LC-MS/MS) analysis	19118452
1										onstitutated saliva (2011c). Driefly, at 6 a.m. (before breakfast), the subjects were asked to finse their models thoroughly with water, then to the		
	042007			х	x		36-62	M/F				40440453
1	P13987		<del>                                     </del>				<del> </del>	<del>                                     </del>		undiyaa.		19118452
				x	×		36-62	M/F		their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until		
1	Q8N274											19118452
1995   1995							26.62	24/5		onstandance salive (20112). Oneny, at o dam, (before breaklast), the subjects were asked to mise their mounts thoroughly with water, then to the		
1.   1.   1.   1.   1.   1.   1.   1.	067W64			×	×		30-02	IVI/F		their reads for ward and allow stated to now into a sterile container for 5 min. These specimens were immediately indicated and stored at 50°C and		19118452
1995   1995							ì			Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	ve saliva samples each from the control and DM groups were pooled separately and	
Section 1				x	x		36-62	M/F	alcoholic; no			
1	P29373											19118452
1				v			26-62	M/E				
1	Q9Y3E8	1		^	^		33-02	,.	,			19118452
1992   1992												1 1 /
Section processes from the common and coding open are posted against and section for the control of the control		1		x	х	2,7	36-62	M/F	,			
1	P36222							1		analysis.		19118452
		l		×	×		36-62	M/F				
March	Q15782	L						,.		analysis.	ectrometry (LC-MS/MS) analysis	19118452
1									Non-smoker; Non-			
				х	x		36-62	M/F				
1	000299						<b> </b>					19118452
Column   C				x	x		36-62	M/F				
14-04   MP   accordance for the Peach Consult and adoles and to the What of the County and adoles and the Peach of The Peach County and adoles and the Peach of The Peach County and adoles and the Peach of The Peach County and adoles and the Peach of The Peach County and adoles and the Peach of The Peach County and adoles and the Peach of The Peach County and adoles and the Peach Of The Peach County and adoles and the Peach Of The Peach County and adoles and the Peach Of The Peach County and adoles and to the What adoles and the Peach Of The Peach County and adoles and the Peach Of	Q13185									analysis.		19118452
Models and the section of the sectio	075300			x	x		36-62	M/F				10118452
Description	073390		t t				1				ve saliva samples each from the control and DM groups were pooled separately and	19118432
1960   1960				x	×		36-62	M/F	alcoholic; no			
## 1900   Section   Processing Companies of the processing Companies of the processing Companies of the processing Companies Companies of the processing Companies Com	Q00610										ectrometry (LC-MS/MS) analysis	19118452
1990   1							26.62	N4/E		onstandated stand (Lone), or one in the control of canada, the stable of white distribution in the control of the stable of of the stab		
Non-anders foo   Section	P10909			*			30-02	IVI/F				19118452
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Non-monder, No.   Discharded salve (2014), Enrish, at 8 am, (before breaking), the subjects were saled to more ther months throughly with water, these to six should be accorded to the programment of the subjects of the s				x	×		36-62	M/F				
2.3. 3.6.6.2 M/F according to the control and position of	Q14019		<b></b>									19118452
pressure of the pressure of th				×	×	2.1	36-62	M/F				
in abcolute, no pregnancy analysis.  In abcolute	P23528					-,-		,.		analysis.		19118452
pro2552												
Non-serolece, No.    Non-serolece, No.   Distributished salva (20mL), Breffly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit subjected to two dimensional liquid chromatography (2-DC) and C-Londem mass spectrometry (C.M.S/MS) analysis.   Non-serolece, No.   Society of the service of the	002452			х	x		36-62	M/F				10110153
prize	PU2452		<del>                                     </del>				1					19118452
programs of the modes from the control and DM groups were pooled separately and subjected to two-dimensional liquid dromatography (2-DLC) and LC-Landem mass pectrometry (LC-MSA)S analysis analysis.    Programs		1		×	×		36-62	M/F		their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	ibjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	i
before the first forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.    Nor smoker, Non-smoker, Non-smok	P12109	<u> </u>							pregnancy	analysis.	nectrometry (LC-MS/MS) analysis	19118452
personancy analysis somewhere the production of the program of the		1	1 T				25.52	/				
P06681	D00726	l		×	×		36-62	M/F				10110453
Accholic, no   pregnancy   p	r00/30		<del>                                     </del>				<b> </b>	<del>                                     </del>				19118452
pregnancy analysis.  Non-smoker, Non-smoke	1	1		×	x		36-62	M/F		their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	i
belief to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass analysis.  2	P06681									analysis.		19118452
post of the post o		1					25.52	/=		onstituated sainte (20112). Briefly, at 0 a.m. (before breaklasty, the subjects were asked to mise their models thoroughly with water, then to the		
Non-smoker; Non-shore and solid program of the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads for near darial allow aslava to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry. (IC-MS/MS) analysis  191184:  Non-smoker; Non-smoker; Non-shore, Non-smoker; Non-s	P01031	l		×	×		36-62	M/F		their reads for ward and allow salve to now into a sterile container for 5 min. These specimens were immediately indicated and stored at 50°C and		19118452
A	. 31031	1					1	1				13110432
Non-smoker, Non-	1	1		×	×	4,8	36-62	M/F		their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until Sul		i l
y control of the program of the prog	P13671									undryss.		19118452
presenancy x x 36-62 M/F  Non-smoker; Non-sinchis; non-senancy x x 36-62 M/F  P07357 X X 36-62 M/F  Non-smoker; Non-shore, Non-shore	1	1		Ų			26.62	NA/E		onstandated stand (Lone), or one in the control of canada, the stable of white distribution in the control of the stable of of the stab		i l
Non-smoker, Non-sm	P10643	1		^	*		50-02	IVI/F				19118452
p07357							1			Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	ve saliva samples each from the control and DM groups were pooled separately and	15110452
Non-smoker; Non-smoker, Non-sm		1		×	×		36-62	M/F				i
P07358	P07357	<b> </b>					1	-				19118452
P07358 pregnancy analysis.    P07358   pregnancy   pre	l	l		v	v		36-62	M/F				
Non-smoker; Non-alcoholic, no alcoholic, no by their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately froze and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (L-MS/MS) analysis.  191184:  Non-smoker; Non-alcoholic, no alcoholic, no alcoholic, no pregnancy Non-smoker; Non-alcoholic, no alcoholic, no alcoholic	P07358	l		^	^		30-02	141/1		their needs forward and anow salve to now into a stellar container for 5 min. These specimens were immediately note in an a stored at 50°C and		19118452
P07360 pregnancy analysis.  pregnancy analysis.  Non-smoker; Non- Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to risse their mouths thoroughly with water, then to tilt.  The subject of										Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	ve saliva samples each from the control and DM groups were pooled separately and	
Non-smoker; Non- Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt five saliva samples each from the control and DM groups were pooled separately and x x x 36-62 M/F alcoholic; no their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		1		x	х		36-62	M/F				
x x 36-62 M/F alcoholic; no their heads forward and allow saliw to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	P07360	<u> </u>	1				<b> </b>	<del>                                     </del>				19118452
	1	1		×	x		36-62	M/F				
programmy functions	P02748								pregnancy		pectrometry (LC-MS/MS) analysis	19118452

P00751			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	19118452
P08603			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimentry (LC-MS/MS) analysis	19118452
P05156			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass analysis.	19118452
075367			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
			х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Q9UBG3			x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	anahysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass control of the control of	
P31146			x	х	-2,4	36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	ulanays. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until their heads forward in the saliva saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until the first forward in the saliva saliva to the saliva saliva saliva to the saliva saliva to the saliva saliva saliva to the saliva saliva to the saliva sa	19118452
P01040	1		x	х	2,2	36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Septrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  five saliva samples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P01034			x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Septermentry (LC-MS/MS) analysis  Septermentry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P28325			×	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P54108				x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjects to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P32320						36-62	M/F	pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
P04839			x	х		****		alcoholic; no pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
P30046			х	х		36-62	M/F	alcoholic; no pregnancy Non-smoker; Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.    Comparison of the control of the contro	19118452
Q9UGM3	1		x	х		36-62	M/F	alcoholic; no pregnancy Non-smoker; Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sankyiss.    Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until sankyiss.   Clusting the control of the control of the subject of two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until sankyis (LC-MS/MS) analysis   Clusting the control of the control of the subject of the subject of the control of the subject of	19118452
Q02487			х	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sankyiss.  Joseph Comments (LC-MS/MS) analysis analysis.  Working and LC-sandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass appeared by the subjected t	19118452
Q02413	-		x	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimentry (LC-MS/MS) analysis (UC-MS/MS) analysis (UC-MS/M	19118452
P32926			х	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five salivas ampless each from the control and DM groups were pooled separately and	19118452
P15924			x	x	-5,9	36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P09417			×	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q01459			х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Section of the saliva samples seach from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P53634			×	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Section of the saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q9UHL4			×	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	19118452
P27487			х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Section of the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies seen the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies seen the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applies are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass are the control and DM groups were pooled separately and subject to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass are the control and DM groups were pooled separ	19118452
P27695			×	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass analysis.	19118452
04W4Y1			х	x		36-62	M/F	Non-smoker; Non- alcoholic; no	The saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q15075			х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Interiors. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q9BW34			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Institution and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC tandem mass spectrometry (LCMS/MS) analysis.	19118452
P19957			х	x		36-62	M/F	Non-smoker; Non- alcoholic; no	anahysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis.	19118452
			x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P26641	1	<u> </u>				1	L	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452

1										
Part	P13639		×	х		36-62	M/F		analysis. spectrometry (LC-MS/MS) analysis	19118452
Part	Q9NZ08		×	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Column	P30040		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
	000836		×	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
1			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Security for the property of t			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	aluanys. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
			×	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Secretary Secret			x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Secondary Company of the company of			×	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
1			x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Name   Section   Part   Section   Part   Section   Part   Part   Section   Part   Pa			x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Note the property of the prope			x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
No.   Section   Continued and section   Continued an			х	х	-2,6	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Section of the section of the control and DM progress were provided represently and section of the section of t			х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-monder, Your   Section(1977)   Policy   Po			х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
23142   X			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker, Non- programs  Non-smoker, Non- process  X			х	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker, Non-sm			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker, Non-sm			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker, Non- pressure,  Non-smoker, Non- pressure,  Non-smoker, Non- pressure,  Non- press			x	х	-6,1	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker; Non-sm			x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker, Non-sm			x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Non-smoker, Non-sm	P04075		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	19118452
x x 36-62 M/F alcoholic; no pregnancy  Non-smoker; Non-smoker; Non-smoker (Non-smoker)  X x 36-62 M/F alcoholic; no pregnancy  Non-smoker;	Q53XB4		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
x x 36-62 M/F alcoholic; no pregnancy   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass   Subjec	P16930		х	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
x x 36-62 M/F alcoholic; no pregnancy   Short Miles   Shor	P09958		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	Q08380		x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
	Q92820		x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452

P17900		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt help heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
095479		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P46926		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P14314		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt wheir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P00367		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P35754		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens (IC-MS/MS) analysis.	19118452
		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	aluanysa. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P00390		x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Five salivas anamples each from the control and DM groups were pooled separately and spectrometry (LC-MS/MS) analysis	19118452
		×	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P09211		x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	anahysis.  Instituted saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P48637		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Five Salivas anamples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P04406		х	х	3,3	36-62	M/F	Non-smoker; Non- alcoholic; no	anahasis.  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Five salivas amplies each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P06737		×	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	anahysis.  Instituted saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
		x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	anahysis.  Instituted saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q92896		x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	anahysis.  Instituted saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
Q8NBJ4 P28676		x	х	-7,1	36-62	M/F	Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilit heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Sepectrometry (LC-MS/MS) analysis  Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P28676		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectromenty (LC-MS/MS) analysis	19118452
Q6ICN0		x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no pregnancy	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectroment ('CL-MS/MS) analysis	19118452
P34932		×	х		36-62	M/F	Non-smoker; Non- alcoholic; no	altarysis. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (C-MS/MS) analysis	19118452
P04792		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Justines and the state of the s	19118452
P07900		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Justines and the state of the s	19118452
P08238		х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (L-WS/MS) analysis	19118452
Q9Y5Z4		х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt wheir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q14103		х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P31943		х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q53F48		х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P61978		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P22626		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q5U071		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
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P04196		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Sectometry (IC-MS/MS) analysis	19118452
Q4VB24		x	x	2,1	36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P04908		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q6FI13		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt wheir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q99877		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	unistimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q39877		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Justings. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens (I-C.MS/MS) analysis	19118452
P00492		x	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P11215		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens (I-C.MS/MS) analysis.	19118452
		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectromenty (L-MS/MS) analysis	19118452
P05107		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q8WWA0		х	х	3,2	36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis  Five salivas angines each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Spectrometry (LC-MS/MS) analysis	19118452
P19827		х	х	2,8	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
O14624		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Spectrometry (LC-MS/MS) analysis  Five flavors aliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q14824 Q12906		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P07476		х	х		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
P06870		х	х		36-62	M/F	Pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
043240		х	х	-5	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (IC-MS/MS) analysis	19118452
Q9UBX7		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	adustrass.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt where heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (CL-MS/MS) analysis	19118452
Q9UKR3		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	alutriss.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (IC-MS/MS) analysis	19118452
P01042		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt wheir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (IC-MS/MS) analysis	19118452
O5TCI8		х	х	-3,3	36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	undraka.  In their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P20700		х	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (C-WS/MS) analysis	19118452
Q03252		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt wheir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P02750		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P30740		х	х	2	36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P09960		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q6FGL0		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P00338		х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P07195		x	х		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt heir heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452

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Non-emoker, You Constituted stallway (20mil, Briefly, at 8 am., (before breakfast), the subjects were asked to rinse their mounts throroughly with water, then to till subjects were asked to rinse their mounts throroughly with water, then to till subjects were asked to rinse their mounts throroughly with water, then to till subjects were asked to rinse their mounts throroughly with water, then to till subject to the reads forward and allow salaws to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the reads forward and allow salaws to flow with to a sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the reads forward and allow salaws to flow with to a sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the reads forward and allow salaws to flow with to a sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the reads forward and allow salaws to flow with the sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the reads forward and allow salaws to flow with the sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the reads forward and allow salaws to flow with the sterile container for 5 min. These specimens were immediately frozen and stored at -80° C until subject to the dimensional liquid chromatography (2-DLC) and LC tandem mass spectrometry (LCM-MSAS) analysis.  2014 18 2	150			x	x		36-62	M/F			rapny (z-DEC) and EC-tandem mass	19118452
Actional control   Part   Pa	130						+				M groups were pooled separately and	13110432
Non-monker, Non-acknolic, no pregnancy acknolic, no pregnancy analysis.  **X				x	×		36-62	M/F		(		
P066660 My and the presentative of the present	218						<u> </u>			analysis. spectrometry (LC-MS/MS) analysis		19118452
pregnancy analysis.    Sectionmetry (LC MS/MS) analysis   Sectionmetry (LC MS/MS) anal		T						1 .				
Piggo   Section   Sectio		- 1		×	×		36-62	M/F			rapny (2-DLC) and LC-tandem mass	19118452
Section   Sect	DOU		-+				1	1			M groups were pooled separately and	19118452
pregnancy  2018		- 1		×	×		36-62	M/F				1
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pregnancy and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva (20mL). Briefly, at 8 a.m.												
Non-smoker, Non-				x	×		36-62	M/F	alcoholic; no		raphy (2-DLC) and LC-tandem mass	
before the safe forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until spectrometry (LC-MS/MS) analysis five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis.    Variable   Va	L88							<u> </u>		dilaiysis.	M groups were pooled separately and	19118452
pregnancy O5865  X  X  X  X  X  X  X  X  X  X  X  X  X		- 1					26.62	NA/E		onsultations suited (20112). Shelly, all outside shell associated in the distribution of the shell associated (20112), and shell associated (20112). Shelly, all of the shell associated (20112) and the shell associated (20112) and the shell associated (20112). Shelly, all of the shell associated (20112) and the shell associated (		1
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a clonbling in pregnancy operands of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.    Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DC) and LC-tandem mass analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subject to two-dimensional liquid chromatography (2-DC) and LC-tandem mass analysis.   Socious of the heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subject to two-dimen			- +								M groups were pooled separately and	13120432
Non-smoker; Non-smoker; Non-smoker and slow aslaw as forward and allow asla		- 1		×	×		36-62	M/F		their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	raphy (2-DLC) and LC-tandem mass	1
x x 36-62 M/F alcoholic; no pregnancy 316-62 M/F al	865									unuryas.		19118452
pregnancy analysis.  29/17/18/96   Description of the pregnancy analysis   Description		Г	T				00.77					
Non-smoker, Non-alcoholic; no alcoholic; no pregnancy  Non-smoker, Non-alcoholic; no alcoholic; no a	745			×	×		36-62	M/F			rapny (z-DLC) and LC-tandem mass	19118452
x x 36-62 M/F alcoholic; no preemancy    Mosmoder, Non-	/43		-+				+	<b>t</b>			M groups were pooled separately and	19118452
Q9U/70 pregnancy analysis. spectrometry (LC-MS/MS) analysis spectr		- 1		×	×		36-62	M/F		onstandance sunta (Lone), or our file or		1
x x 36-62 M/F alcoholic; no their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis.  Non-smoker; Non-alcoholic; no pregnancy alcoholic; no pregnancy analysis.  Non-smoker; Non-lide the subjects were asked to rinse their mouths thoroughly with water, then to tilt their beads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis  Non-smoker; Non-alcoholic; no pregnancy analysis.  Non-smoker; Non-alcoholic; no pregnancy anal	J70						<u> </u>			analysis. spectrometry (LC-MS/MS) analysis		19118452
P15S86 pregrancy analysis.  Non-smoker; Non-sunker; No		T						1 .				
Non-smoker; Non-slacoholic; no their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		- 1		×	×		36-62	M/F	1	the medal of ward and another the assemble for a finite medal of the m	rapny (2-DLC) and LC-tandem mass	
x x 36-62 M/F alcoholic; no their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	080	-+	-+				+	1			M groups were pooled separately and	19118452
		1		×	×		36-62	M/F		(		1
	PD5	J					1	,.	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	,,,	19118452
Non-smoker; Non- Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were a sked to rinse their mouths thoroughly with water, then to till Five saliva samples each from the control and DM groups were pooled separately and		j								Unstimulated saliva (20mt). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and E		
x x 36-62 M/F alcoholic; no their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until		- 1		x	x		36-62	M/F			raphy (2-DLC) and LC-tandem mass	
053H01 pregnancy analysis. spectrometry (LC-MS/MS) analysis	HU1						1	1	pregnancy	analysis. spectrometry (LC-MS/MS) analysis		19118452

	1	ı				1	Non-smoker: Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q5TD07							pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m., (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×	3,1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q09666							pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
				2,4	36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P22894		×	×	2,4	30-02	IVI/F	alconolic; no pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
							Non-smoker, no			
							intake of antibiotics in the			
		×	x	+	35-65	M/F	last 6 months, no	Non-stimulated whole expectorated saliva; Subjects refrained from eating, drinking, and oral hygiene.		
							pregnancy or			
P22894							lactation Non-smoker: Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	ELISA Five saliva samples each from the control and DM groups were pooled separately and	20192865
		×	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P14598							pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
					36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q15080		×	×		30-02	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
2-0-0-0							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P59666				1			pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P08246							pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
		,			36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P80188		×			30-02	WITE	alconolic; no pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
P43490		x	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P43490	<b> </b>		-	-			pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Spectrometry (LC-MS/MS) analysis  Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×		36-62	M/F	alcoholic: no	Their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P05204							pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
					36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P10153		×	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
110133				1			Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	13110431
		x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q9UNZ2			_	-	-		pregnancy Non-smoker: Non-	analysis.  Unstimulated saliya (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×	2,1	36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (ZUML). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P80303		"	-	_,_		,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
Q6UX06		x	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
цоолоо			1	1			Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	13110431
		x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P26022			_	-	-		pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×		36-62	M/F	alcoholic; no	Onsumulated salva (2011L), shelpy, at a a.m. (Detore breakast), me subjects were asked to mise their modula torroughly which water, then to the their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at ~80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
075594						,	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
					36-62		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P19021		×	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
		×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q0VDC6			-	+	+	1	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P62937							pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
	l		1		20.00	/-	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P23284		x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
		<u> </u>	1			1	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	25110452
		×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
O60664	<b> </b>		+	+	+	1	pregnancy Non-smoker: Non-	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×		36-62	M/F	alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
060437					1		pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
	l		1		20.00	/-	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
006830		x	x	-2,6	36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
~00000		<b></b>	1	1	1	1	Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	13110432
		×	x	1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
				-	-	<del> </del>	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
P30044	-		1		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P30044		×	v		30-02	,	pregnancy	their neads forward and allow saliva to now into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
P30044 P30086		×	×						Florest Control of Con	
		-	+		_		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	1 1
P30086		x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated salwa (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	10110153
		-	+		36-62	M/F	Non-smoker; Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.		19118452
P30086		-	+		36-62 36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy		subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452

	 -								
P00558	x	x	-3,2	36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.	ss	19118452
053635	х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until specieted to two-dimensional liquid chromatography (2-DLC) and LC-tandem magnetic.		19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem managements.		
Q6P4A8				36-62	M/F	pregnancy Non-smoker; Non-	analysis.  Spectrometry (LC.MS/MS) analysis  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit  Five saliva samples each from the control and DM groups were pooled separatel their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem ma		19118452
P55058	x	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanalysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to til.  Five saliva Samples each from the control and DM groups were pooled separatel.		19118452
P36955	x	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis. subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.	ss	19118452
P03952	x	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.	ss	19118452
Q5VY30	х	х	2,2	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manahosis spectrometry (LC-MS/MS) anahosis	and	19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem magnetic process.	and	
P00747	×	×		36-62	M/F	pregnancy Non-smoker; Non- alcoholic: no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit  Five saliva samples each from the control and DM groups were pooled separatel their heads forward and allow saliva to flow into a sterile container for 5 min. These soecimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem ma		19118452
P13797		^			,.	pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separatel	and	19118452
P43034	×	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Judgicted to two-dimensional liquid chromatography (2-DLC) and LC-tandem may apply (2-DLC) and LC-tandem may analysis.  Spectrometry (LC-MS/MS) analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may apply (2-DLC) and LC-tandem may analysis.  Spectrometry (LC-MS/MS) analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may apply (2-DLC) and LC-tandem may analysis.  Spectrometry (LC-MS/MS) analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may apply (2-DLC) and LC-tandem may analysis.  Spectrometry (LC-MS/MS) analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may analysis of the subjected to two-dimensional liquid ch		19118452
Q15149	х	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis. subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.	ss	19118452
P21128	×	×		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.		19118452
P09668	х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem majayis.		19118452
	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem magnetic process.		
043490	×	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit  Five saliva samples each from the control and DM groups were pooled separatel their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem ma		19118452
P27918						pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separatel	and	19118452
Q14914	x	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until sanakvist.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt.  Five saliva samples each from the control and DM groups were pooled separatel.		19118452
Q16651	x	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may be extremely (LC-MS/MS) analysis of the control and DM groups were pooled separatel five saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit		19118452
Q06323	х	x		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	is	19118452
P25786	x	×		36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to til their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis spectrometry (LC-MS/MS) analysis		19118452
	х	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem magnetic process.		
P25787	x	×		36-62	M/F	pregnancy Non-smoker; Non- alcoholic: no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit  Five saliva samples each from the control and DM groups were pooled separatel  their heads forward and allow saliva to flow into a sterile container for 5 min. These soecimens were immediately frozen and stored at -80 °C until  subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem ma		19118452
P25789		+		36-62	M/F	pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separatel		19118452
P28066	×	х			IVI/F	alcoholic; no pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately	and	19118452
P60900	х	х		36-62	M/F	alcoholic; no pregnancy Non-smoker: Non-	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to til.  Five saliva samples each from the control and DM groups were pooled separatel.		19118452
014818	x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis. subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.	ss	19118452
Q86U62	x	x	-6,1	36-62	M/F	Non-smoker; Non- alcoholic; no pregnancy	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manahysis spectrometry (LC-MS/MS) analysis		19118452
	x	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem may be a container for 5 min. These specimens were immediately frozen and stored at -80 °C until		
P20618	x	x		36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to lit.  Five saliva samples each from the control and DM groups were pooled separatel their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dimensional liquid chromatography (2-D.C.) and LC-tandem many control and the saliva samples can be subjected to two-dime		19118452
P49721		x	+	36-62	M/F	pregnancy Non-smoker; Non- alcoholic; no	analysis.  Unstimulate Saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separatel their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at 8.0° C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem ma		19118452
P49720	×		-			pregnancy Non-smoker; Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separatel	and	19118452
P28062	х	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis. subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem manalysis.	SS	19118452

							1	Non-smoker: Non-	Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
		x	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P02760				_				pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P07237								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
						36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P30101		×	×			30-02	IVI/F	alconolic; no pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
Q15084		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q15084								pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q99497				_				pregnancy	analysis.	spectrometry (LC-MS/MS) analysis  Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q96BQ1			^			30 02	,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
Q9NUQ9		×	x			36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q3NOQ3								Non-smoker; Non-	driatysis. Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	13110432
		×	x			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q5T949				_				pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
						36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P31949			^			30 02	,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
P80511		x	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
r6U511	<del>   </del>			+				pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P29034								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
		×				36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P26447		×	х			30-02	IVI/F	alconolic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
		×	×		-2,9	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P31151	-		-					pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×		+	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P06702							,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
Q70J99		×	×			36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q/0333								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	15110452
		×	х			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q08188	-		-					pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P22735							,	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P22061		×	×			36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (z-bEc) and EC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
122001								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	15110452
		×	х			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P00734				_				pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P07988							,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
P00491		×	×		-2,1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
10491			-	-				pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P55786								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
						36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
060361		×	×			30-62	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
		x	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
000764	-		-	-				pregnancy Non-smoker: Non-	analysis.	spectrometry (LC-MS/MS) analysis  Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×		3.5	36-62	M/F	alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P14618					-,-		,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
P50395		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
CECUC			-	-				pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q6FHR0								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
						36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
		×	×			30-02	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and EC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P46940				_			_		Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	25220432
P46940		J		- 1				Non-smoker; Non-	onstinulated saliva (2011c). Briefly, at 8 a.m. (before breaklast), the subjects were asked to finde their models thoroughly with water, then to the		1 1
P46940 P51149		x	×			36-62	M/F	alcoholic; no pregnancy	belianced sawa (2011c). Briefly, at 0 a.m. (belove breakess), the sagects were asked to line their modulis third origin; with water, then to the their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452

		x	×		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20m), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt five saliva samples each from the control and DM groups were pooled separately and their heads froward and allow saliva to flow into a sterile container for 5 min. These secimens were immediately frozen and stored at -80° C unit work and allow saliva to flow into a sterile container for 5 min. These secimens were immediately frozen and stored at -80° C unit work and allow saliva to flow into a sterile container for 5 min. These secimens were immediately frozen and stored at -80° C unit work and allow samples each from the control and DM groups were pooled separately and their first samples and the samples ach from the control and DM groups were pooled separately and their first samples and the samples ach from the control and DM groups were pooled separately and their first samples and the samples and the samples and the samples and the samples are the samples and the samples and the samples are the samples and the samples and the samples are the samples and the samples are the samples and the samples are the samples are the samples and the samples are the samples and the samples are the samples are the samples and the samples are the samples are the samples are the samples and the samples are the samples and the samples are the samples and the samples are the samples a	
Q7Z3Y5							pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
					36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20m). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse when from the control and DM groups were pooled separately and their heads from ward and allow saliva to flow into a sterile container for 5 min. These occimens were immediately frozen and stored at -80°C with the saliva samples each from the control and DM groups were pooled separately and their heads from ward and allow active to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
ОЭНДЯЭ		×	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass apalysis.	19118452
2011200							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt   Five saliva samples each from the control and DM groups were pooled separately and	
		x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q15300	-		+	<u> </u>			pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to it!    Westimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were pooled separately and samples each from the control and DM groups were pooled separately and	19118452
		×	x		36-62	M/F	alcoholic: no	Unsurface a save control and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at 80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
A0N5G5							pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
					36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
A0N5G3		×	x		30-02	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applications and the container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass applications are contained from the container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass application and the container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass application and the container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass application and the container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass application and the container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass application and the container for 5 min. The container for 5 m	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt   Five saliva samples each from the control and DM groups were pooled separately and	
		x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at +80 °C until	
A2J1M4	-		+	<u> </u>			pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit.  Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
A2J1N6							pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
		×			36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These sociemens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P52565			×		30-02	IVI/F	alcoholic; no pregnancy	their neads torward and allow saliva to now into a sterile container for 5 min. These specimens were immediately frozen and stored at -80. Cuntil speciments (IC-MS/MS) analysis.	19118452
							Non-smoker; Non-	Unstimulated saliva (20mt). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	
P52566							pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tit  Five salivas samples each from the control and DM groups were pooled separately and	19118452
		×	x	1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P13489		$\longrightarrow$		1	1	1	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
		×	×	3.8	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P07998			^	3,0	30-02	IVI/F	pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and storied at -80. C until	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
000584		x	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two flows frozen and stored at -80 °C until subjected to two	19118452
000584							pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to it!  If we saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	
P02810							pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
		x	×		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q9NQ38			^		30-02	IVI/F	pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and storied at -80. C until	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	
P29508		+	+		1	1	pregnancy Non-smoker: Non-	analysis.  Inordinated value 20mil Briefly, at 8 a.m. (hefore breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till. Five salls as samples each from the control and DM groups were pooled separately and	19118452
		x	x	2,1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q4VAX6							pregnancy	analysis.  Inodemulated saliva (20mi) Briefly, at 8 a.m. (hefore breakfast), the subjects were acked to risse their mouths thoroughly with water, then to till. Five salivas asmoles each from the control and DM groups were gooled separately and	19118452
		×			36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20m.). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse where in mouths thoroughly with water, then to tilt five saliva samples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These secimens were immediately frozen and stored at -80° C until subjected to two-dimensional liquid chromatography (2-DLO) and LC-tandem mass	
P27169		^			30 02	,.	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
							Non-smoker; Non-	Unstimulated saliva (20m.L). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	
Q5VXV3		×	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	19118452
QJVAVJ		+					Non-smoker; Non-	analysis.  [Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to till life saliva smallers each from the control and DM groups were pooled separately and	13110432
		x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
075368				<u> </u>			pregnancy	analysis.  Incidentiated salica (20mi) Briefly, at 8 a.m. (hefore breakfast), the subjects were acked to rises their mouths thoroughly with water, then to till. Five salives samples each from the control and DM groups were pooled separately and	19118452
		×	×		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20m.), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse where in mouths thoroughly with water, then to tilt life saliva's asmples each from the control and DM groups were pooled separately and their heads forward and allow saliva to flow into a sterile container for 5 min. These secimens were immediately frozen and stored at -80° C unit worked and allow admit a liquid chromatography (2-DLO) and LC-tandem mass	
Q9H299							pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
					20.00		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt  Five saliva samples each from the control and DM groups were pooled separately and	
P62316		x	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	19118452
		1	1	1		1	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt five saliva samples each from the control and DM groups were pooled separately and	13110432
		×	x	1	36-62	M/F	alcoholic; no	their heads forward and allow sallwa to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P35326		<del></del>	1	<del>                                     </del>	1	1	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to it!  If we saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×	-2,9	36-62	M/F	alcoholic; no	Unstitutiated saliva (LUML), Brienty, at 8 a.m. (petroe preachast), the subjects were asked to mise their mouths thories their mouths thories and solid and their salivations and solid asked to mise their mouths thories were asked to mise their mouths thories which asked to mise asked to mise their mouths thories which asked to mise asked to mise their mouths thories which asked to mise asked to mise asked to mise their mouths thories which asked to mise asked	
Q96RM1			1	,	1		pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
				1	20.00		Non-smoker; Non-	Unstimulated saliva (20m), Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt behave the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and their behaviors of the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the control and DM groups were pooled separately and the saliva samples each from the con	
Q9UBC9		×	x		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	19118452
		1	1				Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	23110432
		x	x	1	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q14515			+	<b> </b>	1	1	pregnancy Non-smoker: Non-	analysis. spectrometry (LC-MS/MS) analysis Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt Five saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×	1	36-62	M/F	alcoholic; no	Unsumulated saliva (zourn), priemy, at a a.m. (perior ereathas), the subjects were asset to rise their modulins introdeging with water, then to uttain their heads forward and allow as aliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80°C until subjected to two-dimensional liquid chromatography (2-DLC) and C-tandem mass	
P02549			1	ļ		<u> </u>	pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
		x	×		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q13813		1 *		1	30-02	IVI/F	alconolic; no pregnancy	analysis. spectrometry (LC-MS/MS) analysis	19118452
							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
		×	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	19118452
001002					1	1	pregnancy		19118452
Q01082							Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt    Five saliva samples each from the control and DM groups were pooled separately and	
Q01082 Q59EP5		×	х		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimens were immediately frozen and stored at -80 °C until subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass specimently (LC-MS/MS) analysis	19118452

							1	Non-smoker: Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	re saliva samples each from the control and DM groups were pooled separately and	
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P02808								pregnancy	analysis.	ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	
000391		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass ectrometry (LC-MS/MS) analysis	19118452
000391			-	-				pregnancy Non-smoker: Non-		re saliva samples each from the control and DM groups were pooled separately and	19118452
		x	×			36-62	M/F	alcoholic: no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P00441								pregnancy		ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	onstandicted stative (20112). Orienty, at 6 a.m. (before breakdast), the stablects were disked to finde their models diorodginy with water, their to the	re saliva samples each from the control and DM groups were pooled separately and	
P04179		x	х			36-62	M/F	alcoholic; no	their needs for ward and allow salva to now into a sterile container for 5 mile. These specimens were infinited active to the article and stored at the container for 5 mile.	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass ectrometry (LC-MS/MS) analysis	19118452
PU41/9		-		-				pregnancy Non-smoker: Non-		e saliva samples each from the control and DM groups were pooled separately and	19118452
		x	x			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q6UWP8								pregnancy	analysis. spe	ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	
099536		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass ectrometry (LC-MS/MS) analysis	19118452
Q99330			_					pregnancy Non-smoker: Non-		e saliva samples each from the control and DM groups were pooled separately and	19116432
		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
000560								pregnancy		ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	onstitutated salva (zonie). Briefly, at a diff. (before breaklasty, the subjects were disked to finde their models thoroughly with water, their to the	re saliva samples each from the control and DM groups were pooled separately and	
09Y490		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass ectrometry (LC-MS/MS) analysis	19118452
Q91490			_					pregnancy Non-smoker; Non-	analysis. Spe Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	re saliva samples each from the control and DM groups were pooled separately and	19116432
		x	×			36-62	M/F	alcoholic: no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q6FGD7							,	pregnancy	analysis.	ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		e saliva samples each from the control and DM groups were pooled separately and	
		x	х			36-62	M/F	alcoholic; no	their needs for ward and allow salva to now into a sterile container for 5 mile. These specimens were infinited active to the article and stored at the container for 5 mile.	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q9BRA2			_	-				pregnancy Non-smoker: Non-	dialysis.	ectrometry (LC-MS/MS) analysis re saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	alcoholic: no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P30048						****	,.	pregnancy	analysis.	ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	
		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q59E99			_					pregnancy Non-smoker: Non-	dialysis.	ectrometry (LC-MS/MS) analysis re saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	alcoholic: no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q6FGX5		^	^			30-02	141/1	pregnancy		ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	re saliva samples each from the control and DM groups were pooled separately and	
		x	×			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P37837				_				pregnancy	dialysis.	ectrometry (LC-MS/MS) analysis re saliva samples each from the control and DM groups were pooled separately and	19118452
						36-62	M/F	Non-smoker; Non- alcoholic: no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P20061						30-02	IVI/F	pregnancy		ectrometry (LC-MS/MS) analysis	19118452
120001								Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	13110432
		x	x			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P61586								pregnancy		ectrometry (LC-MS/MS) analysis	19118452
		×	×			36-62	M/F	Non-smoker; Non- alcoholic: no		re saliva samples each from the control and DM groups were pooled separately and biected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P37802						30-02	IVI/F	pregnancy		ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	
		x	x			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P55072								pregnancy	analysis.	ectrometry (LC-MS/MS) analysis	19118452
						36-62		Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q53EM5		×	×			36-62	M/F	alcoholic; no pregnancy		ectrometry (LC-MS/MS) analysis	19118452
QJJEMIJ		+						Non-smoker: Non-		re saliva samples each from the control and DM groups were pooled separately and	13110432
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P13693								pregnancy	analysis. spe	ectrometry (LC-MS/MS) analysis	19118452
		1		1				Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	
Q15631		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass ectrometry (LC-MS/MS) analysis	19118452
Q15631			_					pregnancy Non-smoker: Non-		re saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×		-2,1	36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
060235							السا	pregnancy	analysis.	ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		e saliva samples each from the control and DM groups were pooled separately and	
		x	х			36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q9UL52			_	-				pregnancy Non-smoker: Non-		ectrometry (LC-MS/MS) analysis re saliva samples each from the control and DM groups were pooled separately and	19118452
						36-62	M/F	alcoholic; no		bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P02766		^	^			30-02	141/1	pregnancy		ectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-		e saliva samples each from the control and DM groups were pooled separately and	1
		×	×		2,4	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P02766			_	_				pregnancy		ectrometry (LC-MS/MS) analysis re saliva samples each from the control and DM groups were pooled separately and	19118452
		×	×			36-62	M/F	Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and biected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q07654		l ×	×			30-02	IVI/F	alcoholic; no pregnancy		ectrometry (LC-MS/MS) analysis	19118452
20,034				+				Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and	13110432
		×	×			36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
014773								pregnancy		ectrometry (LC-MS/MS) analysis	19118452
	T				Ī	36-62	M/F	Non-smoker; Non-		re saliva samples each from the control and DM groups were pooled separately and bjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
l		×	×	1		30-02	IVI/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	ojected to two-uniterisional liquid chirolitatography (2-bEc) and Ec-tandem mass	1 1
Q9NYL9		_ ^			Į.			pregnancy	analysis and	ectrometry (LC-MS/MS) analysis	19118452

						1		Non-smoker no			
								intake of			
			×	×	+	35-65	M/F	antibiotics in the	Non-stimulated whole expectorated saliva; Subjects refrained from eating, drinking, and oral hygiene.		
			^	^		33 03	,.	last 6 months, no	Ton Jamantea Whole expectoratea Jama, Janjees renamea nom eating, animing, and oral riggene.		
000300								pregnancy or lactation		ELISA	20192865
						20.02	/=	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
P09758			х	х		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
103730								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	15110452
P78324			×	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
P78324								pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
			x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P68036			-					pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
			×	x		36-62	M/F	alcoholic: no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P61088								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
			v	v		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P22314			^	^		30-02	141/1	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
O969H8			x	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
QSUSTIO								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	13110432
O6P5S2			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q0P552							1	pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
1			x	×	10,4	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P11684	1						<b> </b>	pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
1			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q53F10	ļ	$oxed{oxed}$					<u> </u>	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
1			×	x		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q5NV91			^			30-02	141/1	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
						36-62		Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q5NV82			x	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
QSITTOL								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	15110452
075436			×	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
075436								pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
095498								pregnancy Non-smoker: Non-	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
			x	×		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P50552								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
			w.	x		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q12907			*	^		30-02	IVI/F	pregnancy	their neads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
P08670			х	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	19118452
100070								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	15110452
			×	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P18206		<del></del>					<del>                                     </del>	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P02774	<b> </b>	<b>—</b>					<u> </u>	pregnancy Non-smoker, Non-	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
1			x	×		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P04004								pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
1			x	x		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q14508			х	×		30-02	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis	19118452
								Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	
075083			x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
0/3003								pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
l			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P12956	<b> </b>	$\vdash$					<del>                                     </del>	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
Q96DA0	ļ	L .					<b> </b>	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	19118452
			×	x		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P09871							,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis	19118452
l						20.02	24/5	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
P23141			х	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
<u> </u>			İ					Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	22210432
O13126			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis	19118452
Q1312b		<del></del>	1			<b> </b>	<del>                                     </del>	pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and	19118452
l			x	×		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass	
								pregnancy	analysis	spectrometry (LC-MS/MS) analysis	19118452

Q6XQN6	×	×		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis		19118452
QBXQINB				00.00		pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and		19118452
043278	×	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis		19118452
	x	x		36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
P06753						pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis  Five saliva samples each from the control and DM groups were pooled separately and		19118452
	x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
P07910				-	-	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and	-	19118452
P11413	х	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		19118452
P11413					1	pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and		19118452
P25788	x	х		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis		19118452
	×			36-62	M/F	Non-smoker; Non- alcoholic: no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These soccimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
P31946		^		30-02	IVI/F	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis		19118452
	×	×		36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
P54819					,.	pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and		19118452
	x	x		36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
P58499					1	pregnancy Non-smoker: Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and		19118452
	x	x	4,8	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
P60953						pregnancy Non-smoker; Non-	analysis.  Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and		19118452
P67936	х	×	-3,8	36-62	M/F	alcoholic; no	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis		19118452
107330						Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and		13110432
Q08209	×	×		36-62	M/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass spectrometry (LC-MS/MS) analysis		19118452
				36-62	M/F	Non-smoker; Non-	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
Q12805	*	^		30-02	IVI/F	alcoholic; no pregnancy	their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until analysis.	spectrometry (LC-MS/MS) analysis		19118452
	×	×	-2.2	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	Five saliva samples each from the control and DM groups were pooled separately and subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
Q13231			-,-		,.	pregnancy	analysis.	spectrometry (LC-MS/MS) analysis Five saliva samples each from the control and DM groups were pooled separately and		19118452
	x	x	2,6	36-62	M/F	Non-smoker; Non- alcoholic; no	Unstimulated saliva (20mL). Briefly, at 8 a.m. (before breakfast), the subjects were asked to rinse their mouths thoroughly with water, then to tilt their heads forward and allow saliva to flow into a sterile container for 5 min. These specimens were immediately frozen and stored at -80 °C until	subjected to two-dimensional liquid chromatography (2-DLC) and LC-tandem mass		
Q14624						pregnancy	analysis. Patients were asked to retrain from drinking, eating,	spectrometry (LC-MS/MS) analysis		19118452
						Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch			
	х	×	-91,53	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was			
A1XBS5						pregnancy	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)		22314925
AIABSS							and practicing any oral hygiene habits 2 hours before sample	chromatography, tandem mass spectromed y (25-te-wis) wis)		22314923
						Non-smoker; Non-	collection. All sample collection was completed before lunch			
	x	×	17,47	55-75	MF	alcoholic; no pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid		
095969							aliquots of 250 ml and immediately frozen in liquid nitrogen.  Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)		22314925
							and practicing any oral hygiene habits 2 hours before sample			
	x	×	-2,44	55-75	MF	Non-smoker; Non- alcoholic; no	collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was			
						pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid		
P01036					-		aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	-	22314925
						Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample			
	х	×	-4,27	55-75	MF	alcoholic; no	collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was			
P01037						pregnancy	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)		22314925
PU1037							Patients were asked to retrain from drinking, eating,	chromatography/tanuem mass spectrometry (2D-tC-wis/wis)		22314925
						Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch			
	x	×	-6,63	55-75	MF	alcoholic; no pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was			
P01833						pregnancy	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)		22314925
							Patients were asked to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample			
	×		-1.58	55-75	MF	Non-smoker; Non- alcoholic: no	collection. All sample collection was completed before lunch			1
	*	*	-1,30	33-73	IVIF	pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid		
P01834							aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)		22314925
							and practicing any oral hygiene habits 2 hours before sample			
	x	x	-1,52	55-75	MF	Non-smoker; Non- alcoholic; no	collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was			
						pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid		1
P01842		<u> </u>	<b>!</b>	<u> </u>	<u> </u>	ļ	aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)		22314925

								and practicing any oral hygiene habits 2 hours before sample				
							Non-smoker; Non-	collection. All sample collection was completed before lunch				
		x	×	1,52	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was				
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid			
P01857						-		aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	_	<u> </u>	22314925
								and practicing any oral hygiene habits 2 hours before sample				
				-5,40	55-75	MF	Non-smoker; Non- alcoholic; no	collection. All sample collection was completed before lunch				
		^	^	-5,40	33-73	1011	pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was	d label for a differential annual			
P01876							, ,,	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)			22314925
								Patients were asked to retrain from drinking, eating,	, (22 23)			
							Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch				
		x	×	-5,77	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was				
							pregnancy	Collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid			
P02533								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)			22314925
								Patients were asked to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample				
							Non-smoker; Non-	collection. All sample collection was completed before lunch				
		х	×	-5,00	55-75	MF	alcoholic; no	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was				
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid			
P0DJI8								aliquots of 250 ml and immediately frozen in liquid nitrogen.  Tratients were asked to refrain from grinking, eating.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)			22314925
								and practicing any oral hygiene habits 2 hours before sample			1	
							Non-smoker; Non-	collection. All sample collection was completed before lunch			1	
		x	x	-5,00	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was		N4,N4-	1	
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	dimethylas	1	
PODJI9								aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	paragine	Selected saliv	22314925
								and practicing any oral hygiene habits 2 hours before sample				
						MF	Non-smoker; Non-	collection. All sample collection was completed before lunch				
		x	×	4,08	55-75	MF	alcoholic; no pregnancy	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was				
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid			
P02768								aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	_	<del>                                     </del>	22314925
								and practicing any oral hygiene habits 2 hours before sample				
				-1,64	55-75	MF	Non-smoker; Non- alcoholic; no	collection. All sample collection was completed before lunch				
		^	*	-1,04	33-73	IVIF	pregnancy	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was		N-linked		
P02787							pregnancy	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	and O- linked	Selected saliv	22314925
PU2/0/								Patients were asked to remain from drinking, eating,	Ciriomatography, tandem mass spectromed y (20-cc-wis) wisj	illiked	Selected Sally	22314923
							Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample				
		×	×	4,02	55-75	MF	alcoholic; no	collection. All sample collection was completed before lunch		N-linked		
				.,			pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	N-linked glycosylate		
P04745								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	d	Selected saliv	22314925
								Patients were asked to refrain from drinking, eating,				
							Non-smoker: Non-	and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch				
		x	×	-2,41	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was				
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid			
P05976								aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)			22314925
							Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch				
		x	x	16,91	55-75	MF	alcoholic; no	(9 am-12 pm). 84 ml of unstimulated whole saliva sample was				
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid		1	
P08118								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)			22314925
		Ţ	Ţ		1			Patients were asked to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample			1	
							Non-smoker; Non-	collection. All sample collection was completed before lunch				
		х	x	-1,72	55-75	MF	alcoholic; no	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was		N-linked		
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	glycosylate	:	
P12273								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	d	Selected saliv	22314925
								and practicing any oral hygiene habits 2 hours before sample				
							Non-smoker; Non-	collection. All sample collection was completed before lunch			1	
		×	×	2,27	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was		N-linked		
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	glycosylate	:	
P19961	<b>-</b>				<b> </b>	-	<del>                                     </del>	aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	d	Selected saliv	22314925
								and practicing any oral hygiene habits 2 hours before sample			1	
				1.78	55-75	MF	Non-smoker; Non-	collection. All sample collection was completed before lunch			1	
		x	×	1,/8	55-/5	IVIF	alcoholic; no pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was			1	
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid			0004405-
P19961			-		1		1	aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	+	Selected saliv	22314925
								and practicing any oral hygiene habits 2 hours before sample			1	
		×	×	-1,56	55-75	MF	Non-smoker; Non-	collection. All sample collection was completed before lunch			1	
		*	*	-1,30	33-73	IVIF	alcoholic; no pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was	d label for all figures and a second as a second as a second at the seco		1	
	1						p01	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)		1	22314925
P25311												

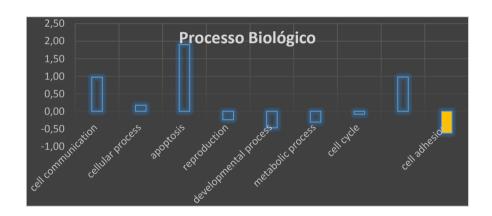
								Patients were asked to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample		
							Non-smoker; Non-	collection. All sample collection was completed before lunch		
		x	x	-1,59	55-75	MF	alcoholic; no	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was		
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
P31025								aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
								and practicing any oral hygiene habits 2 hours before sample		
							Non-smoker; Non-	collection. All sample collection was completed before lunch		
		x	x	3,23	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was		
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
P48723								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
							1	Patients were asked to refrain from drinking, eating,		
							Non-smoker: Non-	and practicing any oral hygiene habits 2 hours before sample		
		×	×	-63.41	55-75	MF	alcoholic; no	collection. All sample collection was completed before lunch		
							pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
P55042								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
							1	Patients were asked to retrain from drinking, eating,	, , , , , , , , , , , , , , , , , , , ,	
							Non smokers Non	and practicing any oral hygiene habits 2 hours before sample		
		×	×	-2,17	55-75	MF	Non-smoker; Non- alcoholic; no	collection. All sample collection was completed before lunch		
		_ ^	^	2,1,	33.73		pregnancy	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was		
Q53EP0							pregnancy	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
Q53EP0	+		-		+	1		auquots or 250 mi and immediately frozen in inquio nitrogen. Patients were asked to refrain from drinking, eating.	chromatography/tandem mass spectrometry (20-LC-Ms/Ms)	22314925
				1	1		1	and practicing any oral hygiene habits 2 hours before sample		
							Non-smoker; Non-	collection. All sample collection was completed before lunch		
	1 1	x	×	-9,81	55-75	MF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was	1	1
				1	1	1	pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
Q5F1R6	$\vdash$			1	1	1	<b></b>	aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
	1 1			1	1	1	1	and practicing any oral hygiene habits 2 hours before sample	1	
				1	1	1	Non-smoker; Non-	collection. All sample collection was completed before lunch		
		x	×	-9,18	55-75	MF	alcoholic; no	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was		
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
Q8NGY0								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
								Patients were asked to retrain from drinking, eating,		
							Non-smoker; Non-	and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch		
		x	×	-4,43	55-75	MF	alcoholic; no	(9 am=12 pm). B4 ml of unstimulated whole saliva sample was		
				1			pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
Q8TAX7								aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
							1	Patients were asked to retrain from drinking, eating,	, , , , , , , , , , , , , , , , , , , ,	
							Non-smoker: Non-	and practicing any oral hygiene habits 2 hours before sample		
		×	×	13.17	55-75	MF	alcoholic; no	collection. All sample collection was completed before lunch		
		_ ^	^	13,17	33.73		pregnancy	(9 am-12 pm). B4 ml of unstimulated whole saliva sample was		
							pregnancy	collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
Q99102	+ +		+	+	1			anduots or 250 mill and infineductery in ober nin induot introgen. Patients were asked to retrain from drinking, eating,	Chromatography/tandem mass spectrometry (20-EC-Wis/Wis)	22314925
								and practicing any oral hygiene habits 2 hours before sample		
				2.46	55.75	MF	Non-smoker; Non-	collection. All sample collection was completed before lunch		
		x	×	-2,16	55-75	IVIF	alcoholic; no	(9 am–12 pm). B4 ml of unstimulated whole saliva sample was		
							pregnancy	collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid	
	L				1					
Q9HC84	1					1	<b>↓</b>	aliquots of 250 ml and immediately frozen in liquid nitrogen.	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
Q9HC84								Patients were asked to retrain from drinking, eating,	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
Q9HC84							Non-smoker; Non-	aliquots of 250 ml and immediately frozen in liquid nitrogen.  Padents were assect or retrain from drinking, eather grang, and practicing any oral hygiene habits 2 hours before sample Collection. All ample collection was completed before funch	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
Q9HC84		x	x	-1,82	55-75	MF	alcoholic; no	Patients were asked to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample	chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
		x	x	-1,82	55-75	MF		Patients were asked to retrain from drinking, eating, and practicing any oral hyglene habits 2 hours befor sample collection. All sample collection was completed before lunch (9 am-12 pm). 84 ml of unstimulated whole saliva sample was collected and centrifuged. The supermatant was placed into	chromatography/tandem mass spectrometry (2D-LC-Ms/Ms)  d label-free differential protein expression analysis using multidimensional liquid	
Q9HC84 Q9HC84		х	x	-1,82	55-75	MF	alcoholic; no	Patients were asked to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen.		22314925 22314925
		х	x	-1,82	55-75	MF	alcoholic; no	Patients were asked to rerain from drinking, eating, and practicing any oral hyglene habits 2 hours before sample collection. All sample collection was completed before funch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to rerain from drinking, eating,	d label-free differential protein expression analysis using multidimensional liquid	
		x	x	-1,82	55-75	MF	alcoholic; no	Patients were asked to rerrain from drinking, eating, and practicing any oral hyglene habits 2 hours befor sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernation was placed into alliquots of 250 ml and immediately frozen in liquid nitrogen. Pratients were asked to retrain from drinking, eating, and practicing any oral hyglene habits 2 hours before sample	d label-free differential protein expression analysis using multidimensional liquid	
		x	x	-1,82 6,54	55-75 55-75	MF	alcoholic; no pregnancy	Patients were asked to rerrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 dam-12 pm.) 84 mil of unstributated whole salive sample was collected and centrifuged. The supernatant was placed into aliquots of 250 mil and immediately frozen in liquid nitrogen. Patients were asked to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch	d label-free differential protein expression analysis using multidimensional liquid	
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							alcoholic; no pregnancy Non-smoker; Non- alcoholic; no	Patients were asked to rerain from drinking, eating, and practicing any oral hygiene habitis 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). 84 ml of unstimulated whole saliva sample was collected and centrifuged. The supermatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to rerain from drinking, eating, and practicing any oral hygiene habitis 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). 84 ml of unstimulated whole saliva sample was collected and centrifuged. The supermatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	
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Q9HC84 Q9UIV8		x	x	6,54	55-75	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non-	Patients were asset to rerrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm), 8 ml of or institulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen.  Patients were assect to rerrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Trainens were assect to rerain mortin unitang, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm). B4 ml of unstimulated whole saliva sample was collection. All sample collection was completed before lunch (9 am—12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid	22314925 22314925
Q9HC84		x	x	6,54	55-75	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	Patients were asset to reram from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from drinking, editing, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and contrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from training, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection in liquid nitrogen. Patients were asset to retrain from training, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925
Q9HC84 Q9UIV8		x	x	6,54	55-75	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	Patients were asset to reram from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 dam-12 pm.) & mol of unstributed whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to Teram from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm.) & ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to Teram from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm.) & ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to Teram from drinking, eating, and practicing and practicing and practicing and and immediately frozen in liquid nitrogen. Patients were asset to Teram from drinking, eating, and practicing and yor and hygiene habits 2 hours before sample	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid	22314925 22314925
Q9HC84 Q9UIV8		x	x	1,90	55-75 55-75	MF	alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-smo	Patients were asset to rerain from drinking, eating, and practicing any oral hyglene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from drinking, eating, and practicing any oral hyglene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to trenam from trunking, eating, and practicing any oral hyglene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to trenam from trunking was sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to trenam from trunking eating, and practicing and practic	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid	22314925 22314925
Q9HC84 Q9UIV8		x	x	6,54	55-75	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	Patients were asset to reram from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 dam—12 pm). Bt mild or minimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen. Tracticing were asset to reram from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm). Bt ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Tracticing any oral hygiene habits 2 hours before sample collection. All sample collection must sample collection mor mining, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm). Bt ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Tracticing were asset to reram from divinge, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm). Bt ml of unstimulated whole saliva sample was collection. All sample collection was completed before lunch (9 am—12 pm). At ml of unstimulated whole saliva sample was collection. All sample collection was completed before lunch (9 am—12 pm). At ml of unstimulated whole saliva sample was	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925
Q9HC84  Q9UIV8  Q9Y5J9		x	x	1,90	55-75 55-75	MF	alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-smo	Patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset or rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Process were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before funch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from drinking sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen. Patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925 22314925
Q9HC84  Q9UIV8  Q9Y5J9		x	x	1,90	55-75 55-75	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	Patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 dam—12 pm). Bet mild or milder was placed into aliquots of 250 ml and immediately frozen in liquid introgen. Patients were asset to Terrain room officially and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am—12 pm). Bet mild of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to Terrain room officially asset to the consideration of the co	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925
Q9HC84  Q9UIV8  Q9Y5J9		x	x	1,90	55-75 55-75 55-75	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	i patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from trunking leading, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from trunking leading.  and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen. Patients were asset to rerain from drinking, eating.  and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to refain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925 22314925
Q9HC84 Q9UIV8 Q9Y5J9		x	x	1,90	55-75 55-75	MF	alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy	ratents were asset to rerrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm), 84 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were assets to retrain room orinking, eating, and practicing any or all hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm), 84 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were assets to retrain room uniming, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm), 84 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were assets to retrain room orinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection or morinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection or morinking, eating, and practicing any oral hygiene habits 2 hours before sample collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to retrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. Patients were asked to refrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. Patients were asked to refrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. Patients were asked to refrain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collec	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925 22314925 22314925
Q9HC84		x	x	1,90	55-75 55-75 55-75	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	i patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from trunking leading, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from trunking leading.  and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid introgen. Patients were asset to rerain from drinking, eating.  and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am–12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asked to refain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection of 250 ml and immediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925 22314925
Q9HC84  Q9UIV8  Q9Y5J9  Q6CRY5		x	x	1,90	55-75 55-75 55-75	MF MF	alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy	ratents were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Tratents were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Tratents were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Tratents were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and immediately frozen in liquid nitrogen. Patients were asset to rerain from drinking, eating, and practicing any oral hygiene habits 2 hours before sample collection. All sample collection was completed before lunch (9 am-12 pm). B4 ml of unstimulated whole saliva sample was collected and centrifuged. The supernatant was placed into aliquots of 250 ml and mediately frozen in liquid nitrogen.	d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)  d label-free differential protein expression analysis using multidimensional liquid chromatography/tandem mass spectrometry (2D-LC-MS/MS)	22314925 22314925 22314925 22314925

									Patients were instructed to expectorate every 6 h into pre-labeled sterile collection tubes. Unstimulated whole resting saliva (approximately 3 ml)		
								Non-smoker; Non-	was obtained from each		
			x	x	1,21	.7 years old; fe	MF	alcoholic; no	patient by passive expectoration at 0, 6, 12, 18, 24, 30, 36 and 42 h. Patients were asked to collect approximately 3 ml of whole saliva without the Quantikine Human EGF Immunoassay		
P01133								pregnancy	assistance of chewing material. No time limit was enforced to collect the requested volume of material. Samples were stored at y20°C until evaluated.		10989322
P01133						<b> </b>			evaluated. Salwa samples were taken from ST patients each with or without diabetes type Q. All samples were taken without oral salivary gland stimulation.	-	10989322
					0.50		MF	Non-smoker; Non-	The patients were instructed to salivate directly into Falcon tubes (Falcon fT ml. Germany) including fTTul RL (QV.Qf.Qd) buff er with protease		
			x	x	9,50		IVIF	alcoholic; no pregnancy	Inhibitors (Complete cocktail tablets, Roche,		
P10645								pregnancy	Germany) for preserving proteins from degradation processes.	biomarker fo	20192923
									(UW) saliva, stimulated		
								Non-smoker; Non-	parotid (SP) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
				x	1,30	34-65	MF	alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a		
								pregnancy	mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of		
P01876									active only swapping the lateral surfaces of the choligae winto. In Individual Casa you are stated as a state of the choligae winto. In Individual Casa you are stated as a state of the choligae winto. In Individual Casa you are stated as a state of the choligae winto. In Individual Casa you are stated as a state of the choligae winto  and the choligae wintow		11014514
010/0									Subjects were requested not to ear, orms or perform oral mygrene measures after midnight the might before saliva collections. Orisinmulated whole	1	11014514
									(LWM) saliva, stimulated particulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
					1.20	34-65	MF	Non-smoker; Non- alcoholic: no	Jamusou (37) and some collected using a modified carfson-Crittendencup (13) and US and SS saliva was collected using a modified carfson-Crittendencup (13) and US and SS saliva was collected with a micropipette connected to a BLISA		
				х	1,26	34-65	IVIF	pregnancy	mini-vacuum pump (14). Salivary stimulation was		
								pregnancy	achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of		
P61626	х								saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).		11014514
									Subjects were requested not to ear, units or perform oral hygiene measures after mininght the night before sanva conections. Oristinulated whose		
								Non-smoker: Non-	parotid (5P) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
				x	1,70	34-65	MF	alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a		
					, ,			pregnancy	mini-vacuum pump (14). Salivary stimulation was		
									achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of		
P02788	X					+			Saliva was as follows: UW (5 min); US (5 min); SP (5 min a firer 2 min to establish a flow); SS (3 min).  Subjects were requised not to ead, ormix or permitted more assured to the months of the mont		11014514
							l		(UW) saliva, stimulated		
							l	Non-smoker; Non-	parotid (SP) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
				х	1,25	34-65	MF	alcoholic; no	a.m. 5º saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum opmo (14). Salivary stimulation was mini-vacuum opmo (14). Salivary stimulation was		
								pregnancy	mini-vacuum pump (14), Salvary sumulaution was a salvary sumular pump (14), Salvary sumulaution was a salvary sumular pump (14), Salvary sumulaution was a salvary sumular pump (14), S		
P02768	x								sallwa was as follows: UN (5 min); US (5 min); SF (5 min after 2 min to establish a flow); SS (3 min).		11014514
									subjects were requested not to eat, orink or perform oral nygiene measures after midnight the night before saliva collections. Oristimulated whole		
									(LWM) saliva, stimulated particulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
				×	2.28	34-65	MF	Non-smoker; Non- alcoholic: no	James of the properties of the		
				×	2,28	34-65	IVIF	pregnancy	mini-vacuum pump (14). Salivary stimulation was		
								pregnancy	achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of		
P22079	х					1			saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).		11014514
									Subjects were required to ear, unit or perform of an hygiene measures after miningire the right before sanks conections. Orsumble of whole (UW) salive, stimulated		
								Non-smoker: Non-	parotid (SP) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
				x	3,86	34-65	MF	alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a		
								pregnancy	mini-vacuum pump (14). Salivary stimulation was		
P05164									achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% MyV) citric acid every 30 s. The sequence and times of collection of sallow axes as follows: UN (5 mily, 18° follows: UN (5 mily, 18° follows: 10 MY 5 mily, 18° f		11014514
PU3104						+			subjects were requested not to eat, urink or perform or at nygrene measures after mining it the night before saliva collections. Onstimulated whole		11014514
									(UW) saliva, stimulated		
								1			
								Non-smoker; Non-	parotid (SP) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 and 10:00 from the collected from the col		
		x		x	1,25	34-65	MF	alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a		
		×		x	1,25	34-65	MF		a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was		
P01876		х		x	1,25	34-65	MF	alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a		11014514
P01876		х		x	1,25	34-65	MF	alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by wabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were requested not or exact unmix or periorm or an register measures arter miningir to or ingir upon exams collections. Unstimulated whole		11014514
P01876		x		х	1,25	34-65	MF	alcoholic; no pregnancy	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Surjects where requested into to eat, unink or perform oral riggerie measures after minding it the night before saliva collections. Onsumulated whole (UW) saliva, stimulated		11014514
P01876		x		x	1,25		MF	alcoholic; no pregnancy Non-smoker; Non-	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by wabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were requested not or exact unmix or periorm or an register measures arter miningir to or ingir upon exams collections. Unstimulated whole		11014514
P01876		x		x		34-65 34-65		alcoholic; no pregnancy	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a rachieved by swabbling the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivar was as follows: UW (5 min); US (5 min); SP (5 min are removed and though soft and the salivar collection of salivary was as follows: UW (5 min); US (5 min); SP (5 min are removed and though soft and though soft and though soft and the salivary of the salivar		11014514
		x		x				alcoholic; no pregnancy Non-smoker; Non- alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were requisestern to tree act, unnix or periorm branching the minimal tree might be one and the salivary of		
P01876		x		x				alcoholic; no pregnancy Non-smoker; Non- alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a rachieved by swabbling the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivar was as follows: UW (5 min); US (5 min); SP (5 min are removed and though soft and the salivar collection of salivary was as follows: UW (5 min); US (5 min); SP (5 min are removed and though soft and though soft and though soft and the salivary of the salivar		11014514
		x		x				alcoholic; no pregnancy Non-smoker; Non- alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were requisestern to tree act, unnix or periorm branching the minimal tree might be one and the salivary of		
		×		x				alcoholic; no pregnancy Non-smoker; Non- alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a achieved by swabbling the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min a fater 2 min to establish a flow); SS (3 min).  Subjects were requested to true act, units or periorition var register miseauries a term emiliagnite the might be better a salva collections. Unsumulated winder (UW) saliva, stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbling the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min a fater 2 min to establish a flow); SS (3 min).  Subjects were requested to tree can, unmix or periorition var register membranes are are mininger to mingrite tree might before a saliva collections. Unsumulated wrone (UW) saliva, stimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00		
		x		x x				alcoholic; no pregnancy Non-smoker; Non- alcoholic; no pregnancy	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min.) Se (5 min.) Se (5 min.) for salivary stimulation was a surface of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was a surface of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was a collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary was refreshed to a collection of salivary was refreshed to the collection of salivary was collected with a micropipette connected to a collection of salivary was collected with a micropipette collection of salivary was collected with a micropipette solice where 8:00 and 10:00 are salivary was collected (SS) and unstimulated (US) submandibular/sublingual salivary was collected with a micropipette connected to a salivary was collected using a modified Carlson-Crittenden cup (13) and US and SS salivary was collected with a micropipette connected to a salivary was collected using a modified Carlson-Crittenden cup (13) and US and SS salivary was collected with a micropipette connected to a salivary was collected with a micropipette connected to a salivary was collected with a micropipette connected to a salivary was collected with a micropipette connected to a salivary was collected with a micropipette connected to a salivary was collected with a micropipette connected to a salivary was collected with a micr		
					1,15	34-65	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non-	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); 2F (5 min after 2 min to establish a flow), SS (3 min). Surjects were requested to compare the control of the salivary of the control of the con		
P61626					1,15	34-65	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min), 58 (6 min) in the castables in flowly, \$6 (7 min) in the saliva was as follows: UW (5 min), 58 (6 min) in the castables in flowly, \$6 (7 min) in the saliva was as follows: UW (5 min), 58 (6 min) in the saliva was as follows: UW (5 min), 58 (6 min) in the saliva was as follows: UW (5 min), 58 (6 min) in the saliva was as follows: UW (5 min), 58 (6 min) in the saliva was as follows: UW (5 min), 58 (5 min) in the saliva was as follows: UW (5 min), 58 (5 min) in the saliva was as follows: UW (5 min), 58 (5 min after 2 min to establish a flow), 55 (3 min).  Subjects were requested unto use and under the saliva was as follows: UW (5 min), 58 (5 min after 2 min to establish a flow), 55 (3 min).  Subjects were requested in the saliva was as follows: UW (5 min), 58 (5 min after 2 min to establish a flow), 55 (3 min).  Subjects were requested in the saliva was as follows: UW (5 min), 58 (5 min after 2 min to establish a flow), 55 (3 min).  Subjects were requested in the saliva was as follows: UW (5 min), 58 (5 min after 2 min to establish a flow), 55 (3 min).  Subjects were requested unto the saliva was collected from each subject between 8:00 and 10:00 an		11014514
					1,15	34-65	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a stokewed by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min), 5F (6 min after 2 min to establish a flow), 5S (2 min).  Subjects were requested unit or eat, units or perform oral rygene measures after mortigint units regin units or saliva was collected (SP) and stimulated (SP) and stimulated (SP) and stimulated (SP) and stimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was sollected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Suppers were requested units of the surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish af low); SS (3 min).		
P61626					1,15	34-65	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); 2F (5 min after 2 min to establish a flow), SS (3 min).  The particle (5P) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SF (6 min after 2 min to establish a flow); SS (3 min).  Subjects were requested unit or ear, units or perform oral rygene measures after mutigint ore right openers saliva collected to sing a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was an experiment of the salivary stimulation was acid to salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation		11014514
P61626		x		x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Superctive feer requested to the case, unknown periorition was registered in the properties of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min) representation to each surface so the construction of salivary was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min) representation to the activity of the salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s.		11014514
P61626					1,15	34-65	MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  (UW) saliva, stimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SP (6 min after 2 min to establish a flow); SS (3 min).  Subjects were requested unit to each, unit or periorin oral regient measures after multigrit the right between 8:00 and 10:00 a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Saliva was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Saliva was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Saliva was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).		11014514
P61626		x		x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Superctive feer requested to the case, unknown periorition was registered in the properties of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min) representation to each surface so the construction of salivary was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min) representation to the activity of the salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s.		11014514
P61626		x		x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SF (5 min after 2 min to establish a flow), SS (3 min).  granting (19) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SF (6 min after 2 min to establish a flow); SS (3 min).  Supects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was a collected using a modified Carlson-Crittenden cup (13) and uS and SS saliva was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SF (6 min after 2 min to establish a flow); SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was collected with a micropipette connected to a minivazuum pump (14). Salivary stimulation was		11014514
P61626		x		x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was as followed by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min)s, EV, 6 min sfer 2 min to establish a flowly. SG 2 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was as follows: UW (5 min)s, EV, 6 min sfer 2 min to establish a flowly. Sci 2 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected with a micropipette connected to		11014514 11014514
P61626 P02788		x		x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow), SS (3 min).  The particle (SP) and stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SP (6 min after 2 min to establish a flow); SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  ELISA  ELIS		11014514 11014514
P61626		x	x	x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. 9P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a solice with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a solice with salivary stimulation was s		11014514 11014514
P61626 P02788		x	x	x	1,15	34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no pregnancy	a.m. 9P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was as followed by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min)s, 190, 6 min after 2 min to establish a flowly, SS (2 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flowly, SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flowly, SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flowly, SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connect		11014514 11014514
P61626 P02788		x	×	x x	1,15 1,59 1,42	34-65 34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non-alcoholic; no pregnancy	a.m. SP saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a follows: UW (S min); US (S min); SP (S min after 2 min to establish a flow); SS (3 min).  ELISA		11014514 11014514
P02788 P02768		x	x x	x x	1,15 1,59 1,42	34-65 34-65 34-65	MF MF	alcoholic; no pregnancy  Non-smoker; Non- alcoholic; no	a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was as follows: UW (5 min), 5P (5 min after 2 min to establish a flow), 5S (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulated (SS) and unstimulated (US) submandibular/sublingual saliva were collected from each subject between 8:00 and 10:00 a.m. 5P saliva was collected using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was achieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were requested using a modified Carlson-Crittenden cup (13) and US and SS saliva was collected with a micropipette connected to a mini-vacuum pump (14). Salivary stimulation was a chieved by swabbing the lateral surfaces of the tongue with 0.1 mol/L (2% w/v) citric acid every 30 s. The sequence and times of collection of saliva was as follows: UW (5 min); US (5 min); SP (5 min after 2 min to establish a flow); SS (3 min).  Subjects were asked not to eat, smoke or drink (except water) for an overnight fast prior to collected of the pumps (swaps) and the salivary stimulation was achieved		11014514 11014514 11014514 17244479

P05231		x	x	0,00	35-65	MF	Non-smoker; Non- alcoholic; no pregnancy	Subjects refrained from eating, drinking, and oral hygiene for 2 hours prior to saliva collection.	ELISA	20192865
P04745		x	x	1,14	45-64	MF	The study included patientes with habits of smoking and alcohol consumption	Subjects were told not to eat or smoke for one hour prior to the examination.	Boehringer MPR 3 a-amylase EPS method	11005153
P02768		x	x	1,19	45-64	MF	The study included patientes with habits of smoking and alcohol consumption	Subjects were told not to eat or smoke for one hour prior to the examination.	colorimetric methods	11005153
P04745		x	х	1,24	38-66	MF	Non-smoker; Non- alcoholic; no	Subjects were asked not to eat, smoke or drink (except water) for an overnight fast prior to collection of saliva samples. Their diets were similar with respect to protein content and uptake of fat and carbohydrates. Socioeconomical status was similar for both groups (survey data).	amylase activity was determined by the colorimetric method of Winn-Deen and coworkers	17244479
P04745		v	x	1,25	38-66	MF	alcoholic; no	Subjects were asked not to eat, smoke or drink (except water) for an overnight fast prior to collection of saliva samples. Their diets were similar with respect to protein content and uptake of fat and carbohydrates. Socioeconomical status was similar for both groups (survey data).	amylase activity was determined by the colorimetric method of Winn-Deen and coworkers	17244479
Q9UBU3		x	x	-1,23	38-66	MF	alcoholic; no	Subjects were asked not to eat, smoke or drink (except water) for an overnight fast prior to collection of sailwa samples. Their diets were similar with respect to protein content and uptake of fat and carbohydrates. Socioeconomical status was similar for both groups (survey data).	Ghrelin was measured in unstimulated total saliva using the same human-ghrelin- RIA kit, which is designed to test any biological fluid with sufficient levels of the peptide to be determined.	17244479

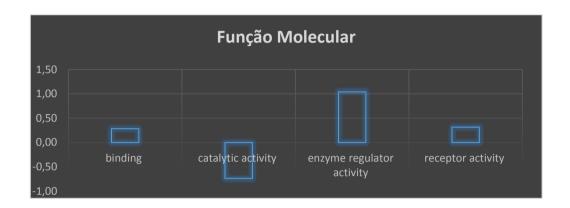
	Oraloma	Diabetes	Diabetes	Diferença	
Biological Process	Normal	esperado	observado	fracional	P value
cell communication	544	6,598	13	0,97	0,007013492
cellular process	912	11,061	13	0,18	0,287905016
apoptosis	142	1,722	5	1,90	0,026555207
reproduction	109	1,322	1	-0,24	0,741568258
developmental process	469	5,688	3	-0,47	0,942642667
metabolic process	1307	15,852	11	-0,31	0,977046763
cell cycle	269	3,263	3	-0,08	0,648398234
immune system process	459	5,567	11	0,98	0,01466063
cell adhesion	219	2,656	1	-0,62	0,938322452
Total Oraloma normal	2391				
Total Diabetes 1	29				

 $<sup>^{</sup>st}$  Log of fractional difference (observed vs expected) is calculated for extesting list as: (# genes for the category - # genes expected)/ # genes expected



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	Oraloma	Diabetes	Diabetes	Diferença	
Função Molecular	Normal	esperado	observado	fracional	
binding	965	11,70431	15	0,28	0,1452
catalytic activity	946	11,47386	3	-0,74	0,999912
enzyme regulator activity	202	2,450021	5	1,04	0,093208
receptor activity	188	2,280217	3	0,32	0,401688



<sup>\*</sup> Log of fractional difference (observed vs expected) is calculated for  $\varepsilon$  testing list as: (# genes for the category - # genes expected)/ # gene expected

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	Oraloma	Diabetes
Pathway	Normal	esperado
Alzheimer disease-presenilin pathway	23	0,28
Apoptosis signaling pathway	23	0,28
Beta1 adrenergic receptor signaling pathway	16	0,19
Beta2 adrenergic receptor signaling pathway	16	0,19
5HT1 type receptor mediated signaling pathway	13	0,16
Endothelin signaling pathway	17	0,21
Enkephalin Release	13	0,16
GABA-B receptor II signaling	10	0,12
Hedgehog signaling pathway	3	0,04
	50	0.70
Gonadotropin releasing hormone receptor pathway	58	•
Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pa	=	,
Inflammation mediated by chemokine and cytokine signaling pathway	74	-/
Interleukin signaling pathway	14	- /
Histamine H2 receptor mediated signaling pathway	9	,
Interferon-gamma signaling pathway	4	-,
Metabotropic glutamate receptor group I pathway	3	,
Metabotropic glutamate receptor group II pathway	16	0,19
Metabotropic glutamate receptor group III pathway	12	0,15
Muscarinic acetylcholine receptor 2 and 4 signaling pathway	16	0,19
Transcription regulation by bZIP transcription factor	4	0,05
Wnt signaling pathway	43	0,52

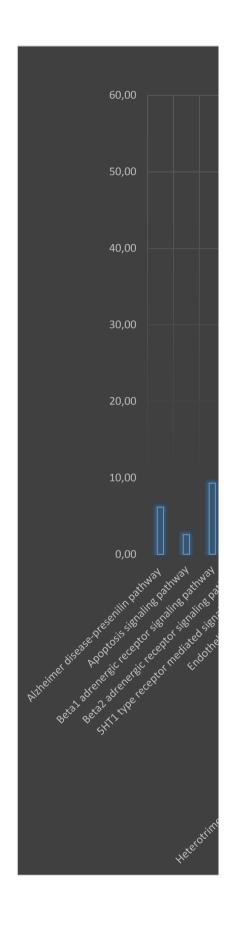
Devido ao nº elevado de Pathways relativo ao OralOma Normal (125), apresentei apenas os comuns com

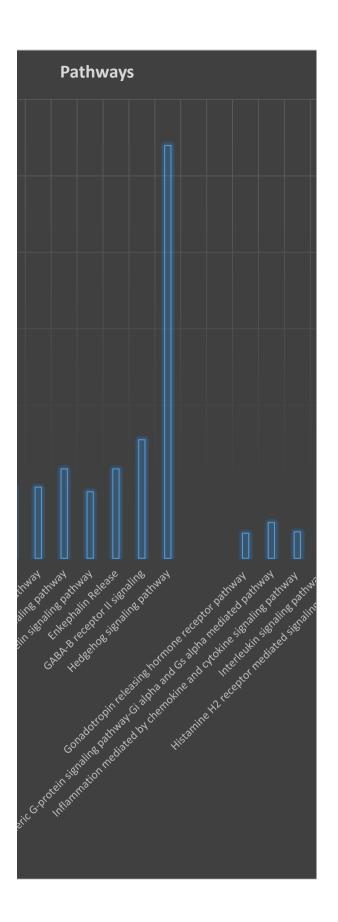
 $<sup>^{\</sup>ast}$  Log of fractional difference (observed vs expected) is calculated for eac testing list as: (# genes for the category - # genes expected)/ # genes expected

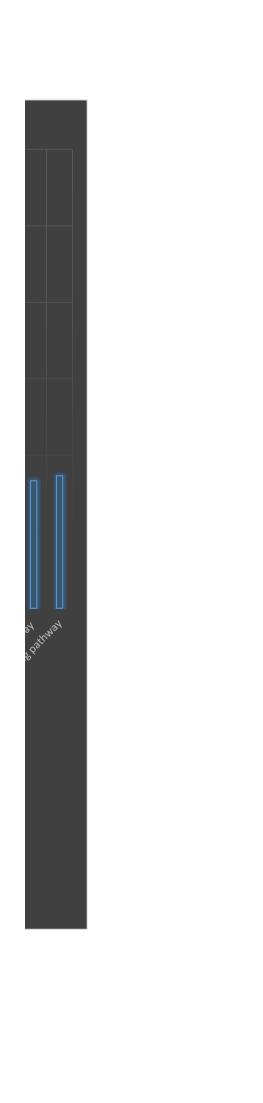
Diabetes	Diferença	
observado	fracional	
2	6,17	0,03
1	2,58	0,24
2	9,31	0,02
2	9,31	0,02
2	11,68	0,01
2	8,70	0,02
2	11,68	0,01
2	15,49	0,01
2	53,97	0,00
3	3,26	0,03
2	4,69	0,05
4	3,46	0,01
3	16,67	0,00
2	17,32	0,01
1	19,61	0,05
2	53,97	0,00
2	9,31	0,02
2	12,74	0,01
2	9,31	0,02
2	40,22	0,00
1	0,92	0,41

## a Diabetes

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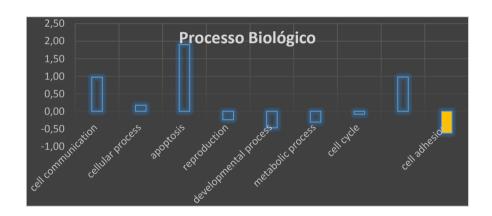






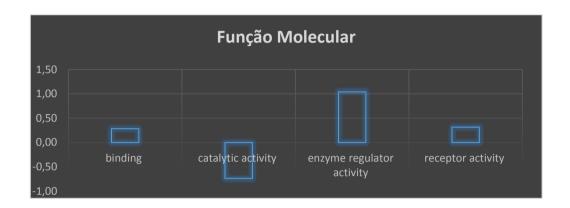
	Oraloma	Diabetes	Diabetes	Diferença	
Biological Process	Normal	esperado	observado	fracional	P value
cell communication	544	6,598	13	0,97	0,007013492
cellular process	912	11,061	13	0,18	0,287905016
apoptosis	142	1,722	5	1,90	0,026555207
reproduction	109	1,322	1	-0,24	0,741568258
developmental process	469	5,688	3	-0,47	0,942642667
metabolic process	1307	15,852	11	-0,31	0,977046763
cell cycle	269	3,263	3	-0,08	0,648398234
immune system process	459	5,567	11	0,98	0,01466063
cell adhesion	219	2,656	1	-0,62	0,938322452
Total Oraloma normal	2391				
Total Diabetes 1	29				

 $<sup>^{</sup>st}$  Log of fractional difference (observed vs expected) is calculated for extesting list as: (# genes for the category - # genes expected)/ # genes expected



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	Oraloma	Diabetes	Diabetes	Diferença	
Função Molecular	Normal	esperado	observado	fracional	
binding	965	11,70431	15	0,28	0,1452
catalytic activity	946	11,47386	3	-0,74	0,999912
enzyme regulator activity	202	2,450021	5	1,04	0,093208
receptor activity	188	2,280217	3	0,32	0,401688



<sup>\*</sup> Log of fractional difference (observed vs expected) is calculated for  $\varepsilon$  testing list as: (# genes for the category - # genes expected)/ # gene expected

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	Oraloma	Diabetes
Pathway	Normal	esperado
Alzheimer disease-presenilin pathway	23	0,28
Apoptosis signaling pathway	23	0,28
Beta1 adrenergic receptor signaling pathway	16	0,19
Beta2 adrenergic receptor signaling pathway	16	0,19
5HT1 type receptor mediated signaling pathway	13	0,16
Endothelin signaling pathway	17	0,21
Enkephalin Release	13	0,16
GABA-B receptor II signaling	10	0,12
Hedgehog signaling pathway	3	0,04
Gonadotropin releasing hormone receptor pathway	58	0,70
Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pa	29	0,35
Inflammation mediated by chemokine and cytokine signaling pathway	74	0,90
Interleukin signaling pathway	14	0,17
Histamine H2 receptor mediated signaling pathway	9	0,11
Interferon-gamma signaling pathway	4	0,05
Metabotropic glutamate receptor group I pathway	3	0,04
Metabotropic glutamate receptor group II pathway	16	0,19
Metabotropic glutamate receptor group III pathway	12	0,15
Muscarinic acetylcholine receptor 2 and 4 signaling pathway	16	0,19
Transcription regulation by bZIP transcription factor	4	0,05
Wnt signaling pathway	43	0,52

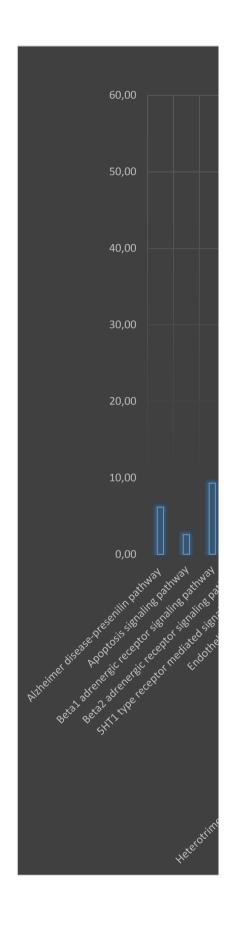
Devido ao nº elevado de Pathways relativo ao OralOma Normal (125), apresentei apenas os comuns com

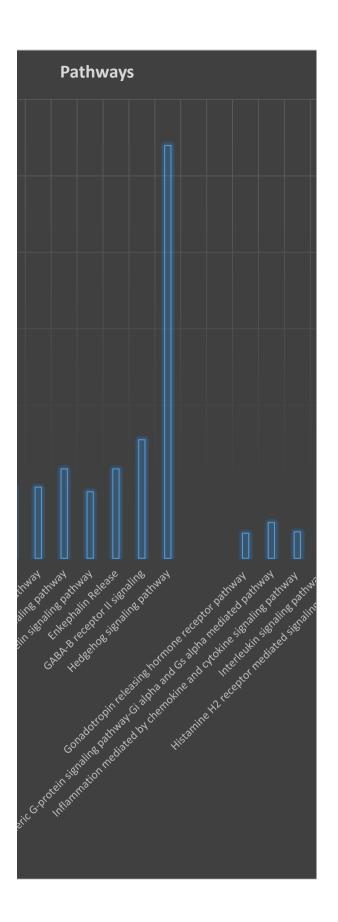
 $<sup>^{\</sup>ast}$  Log of fractional difference (observed vs expected) is calculated for eac testing list as: (# genes for the category - # genes expected)/ # genes expected

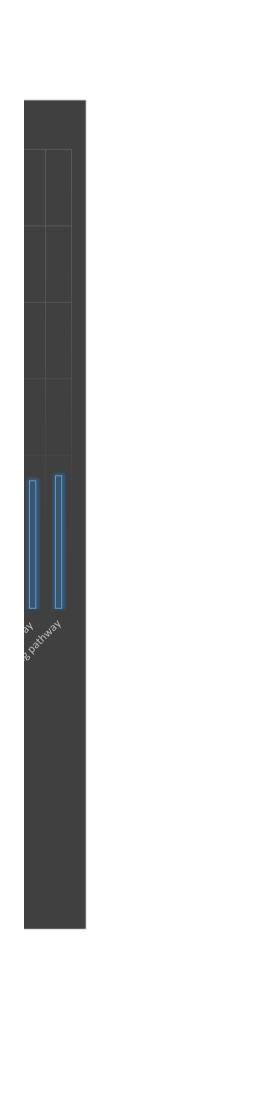
Diabetes	Diferença	
observado	fracional	
2	6,17	0,03
1	2,58	0,24
2	9,31	0,02
2	9,31	0,02
2	11,68	0,01
2	8,70	0,02
2	11,68	0,01
2	15,49	0,01
2	53,97	0,00
3	3,26	0,03
2	4,69	0,05
4	3,46	0,01
3	16,67	0,00
2	17,32	0,01
1	19,61	0,05
2	53,97	0,00
2	9,31	0,02
2	12,74	0,01
2	9,31	0,02
2	40,22	0,00
1	0,92	0,41

## a Diabetes

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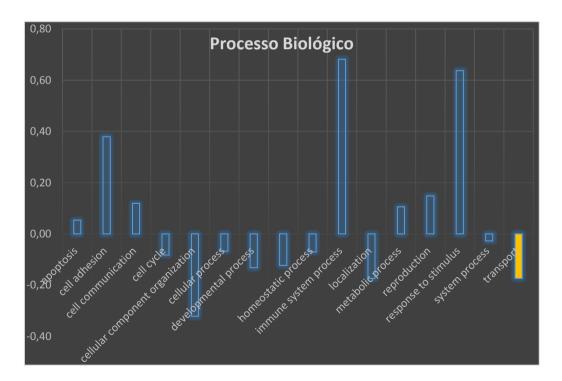






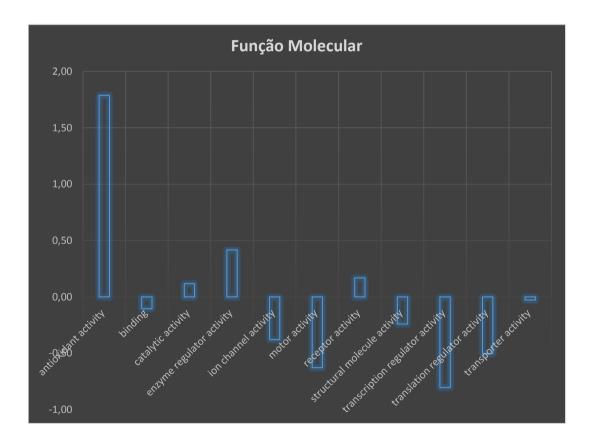
	Ovalant	Diabetes	Diabatas	Diferen	
Dialogical Drasses	Oraloma		Diabetes	Diferença	
Biological Process	Normal	0	<u>observado</u>	fracional	value
apoptosis	127	22,780	24	0,05	0,43
cell adhesion	194	34,797	48	0,38	0,02
cell communication	503	90,221	101	0,12	0,11
cell cycle	237	42,510	39	-0,08	0,74
cellular component organization	304	54,527	37	-0,32	1,00
cellular process	843	151,206	141	-0,07	0,86
developmental process	411	73,720	64	-0,13	0,91
generation of precursor metabolites and energ	89	15,964	14	-0,12	0,73
homeostatic process	30	5,381	5	-0,07	0,63
immune system process	398	71,388	120	0,68	0,00
localization	27	4,843	4	-0,17	0,71
metabolic process	1261	226,181	250	0,11	0,01
reproduction	68	12,197	14	0,15	0,34
response to stimulus	286	51,299	84	0,64	0,00
system process	281	50,402	49	-0,03	0,61
transport	459	82,329	68	-0,17	0,97
Total Oraloma normal	2336				
Total Diabetes	419				

 $<sup>^{</sup>st}$  Log of fractional difference (observed vs expected) is calculated for each testing list as: (# genes for the category - # genes expected)/ # genes expected



	a	esperad	Diabetes	a	
Função Molecular	Normal	o	observado	fracional	P value
antioxidant activity	16	2,87	8	1,79	0,01
binding	910	163,22	146	-0,11	0,96
catalytic activity	909	163,04	182	0,12	0,03
enzyme regulator activity	181	32,47	46	0,42	0,01
ion channel activity	27	4,84	3	-0,38	0,86
motor activity	30	5,38	2	-0,63	0,97
receptor activity	167	29,95	35	0,17	0,19
structural molecule activity	426	76,41	58	-0,24	0,99
transcription regulator activity	86	15,43	3	-0,81	1,00
translation regulator activity	45	8,07	4	-0,50	0,96
transporter activity	149	26,73	26	-0,03	0,59
Total Oraloma normal	2336				
Total Diabetes	419				

<sup>\*</sup> Log of fractional difference (observed vs expected) is calculated for each testing list as: (# genes for the category - # genes expected)/ # genes expected

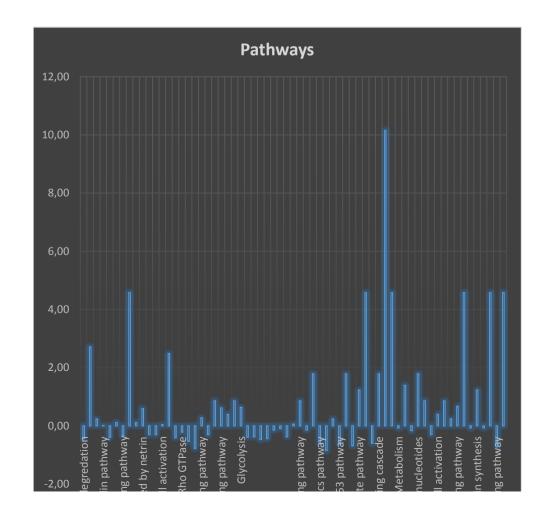


Pathway	Oraloma Normal	Diabetes esperado	Diabetes observado	Diferença fracional	P value
5-Hydroxytryptamine degredation	10	1,79	1,00	-0,44	0,83
Adenine and hypoxanthine salvage pathway	3	0,54	2,00	2,72	0,10
Alzheimer disease-amyloid secretase pathway	9	1,61	2,00	0,24	0,48
Alzheimer disease-presenilin pathway	22	3,95	4,00	0,01	0,56
Angiogenesis	28	5,02	3,00	-0,40	0,88
Angiotensin II-stimulated signaling through G proteins and beta-arrestin	15	2,69	3,00	0,12	0,50
Apoptosis signaling pathway	25	4,48	3,00	-0,33	0,83
Asparagine and aspartate biosynthesis	2	0,36	2,00	4,58	0,05
ATP synthesis	5	0,90		0,12	0,59
Axon guidance mediated by netrin	7	1,26	2,00	0,59	0,36
Axon guidance mediated by semaphorins	8	1,43	1,00	-0,30	0,76
Axon guidance mediated by Slit/Robo	8	1,43	1,00	-0,30	0,76
B cell activation	16	2,87		0,05	0,55
Blood coagulation	16	2,87			0,00
Cadherin signaling pathway	19	3,41	2,00	-0,41	0,86
Cytoskeletal regulation by Rho GTPase	50	8,97		-0,22	0,79
De novo purine biosynthesis	12	2,15	1,00	-0,54	0,88
Dopamine receptor mediated signaling pathway	25	4,48	1,00	-0,78	0,99
EGF receptor signaling pathway	35	6,28	8,00	0,27	0,29
Endothelin signaling pathway	16	2,87	2,00	-0,30	0,78
FAS signaling pathway	9	1,61	3,00	0,86	0,22
FGF signaling pathway	31	5,56	9,00	0,62	0,11
Fructose galactose metabolism	4	0,72	1,00	0,39	0,51
Glutamine glutamate conversion	3	0,54	1,00	0,86	0,42
Glycolysis	17	3,05	5,00	0,64	0,19
Gonadotropin releasing hormone receptor pathway	55	9,87		-0,39	0,93
Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pathwa		4,84	3,00	-0,38	0,86
Heterotrimeric G-protein signaling pathway-Gq alpha and Go alpha mediated pathw		3,77	2,00	-0,47	0,89
Heterotrimeric G-protein signaling pathway-rod outer segment phototransduction	10	1,79	1,00	-0,44	0,83
Huntington disease	59	10,58	9,00	-0,15	0,73
Inflammation mediated by chemokine and cytokine signaling pathway	74	13,27		-0,10	0,68
Insulin/IGF pathway-mitogen activated protein kinase kinase/MAP kinase cascade	9	1,61		-0,38	0,80
Integrin signalling pathway	84	15,07		0,06	0,44
Interferon-gamma signaling pathway	3	0,54	1,00	0,86	0,42
Interleukin signaling pathway	13	2,33	2,00	-0,14	0,68
N-acetylglucosamine metabolism	2	0,36	1,00	1,79	0,30
Nicotine pharmacodynamics pathway	14	2,51	1,00	-0,60	0,92
Nicotinic acetylcholine receptor signaling pathway	36	6,46	1,00	-0,85	1,00
p38 MAPK pathway	9	1,61	2,00	0,24	0,48
p53 pathway	15	2,69	1,00	-0,63	0,93
Parkinson disease	36	6,46	18,00	1,79	0,00
PDGF signaling pathway	18	3,23	1,00	-0,69	0,96
Pentose phosphate pathway	5	0,90	2,00	1,23	0,23
Phenylalanine biosynthesis	2	0,36	2,00	4,58	0,05
PI3 kinase pathway	14	2,51	1,00	-0,60	0,92
Plasminogen activating cascade	8	1,43	4,00	1,79	0,06
Purine metabolism	1	0,18		10,15	0,01
Pyridoxal phosphate salvage pathway	1	0,18	1,00	4,58	0,16
. ,	1	0,10	1,00	4,50	0,10

Pyrimidine Metabolism	6	1,08	1,00	-0,07	0,66
Pyruvate metabolism	7	1,26	3,00	1,39	0,13
Ras Pathway	20	3,59	3,00	-0,16	0,70
Salvage pyrimidine deoxyribonucleotides	2	0,36	1,00	1,79	0,30
Salvage pyrimidine ribonucleotides	6	1,08	2,00	0,86	0,29
Synaptic_vesicle_trafficking	8	1,43	1,00	-0,30	0,76
T cell activation	20	3,59	5,00	0,39	0,29
TCA cycle	6	1,08	2,00	0,86	0,29
TGF-beta signaling pathway	9	1,61	2,00	0,24	0,48
Toll receptor signaling pathway	10	1,79	3,00	0,67	0,27
Tyrosine biosynthesis	2	0,36	2,00	4,58	0,05
Ubiquitin proteasome pathway	24	4,30	4,00	-0,07	0,62
Vasopressin synthesis	5	0,90	2,00	1,23	0,23
VEGF signaling pathway	12	2,15	2,00	-0,07	0,63
Vitamin B6 metabolism	1	0,18	1,00	4,58	0,16
Wnt signaling pathway	41	7,35	2,00	-0,73	0,99
Xanthine and guanine salvage pathway	2	0,36	2,00	4,58	0,05
Total Oraloma normal	2336				

Devido ao nº elevado de Pathways relativo ao OralOma Normal (125) , apresentei apenas os comuns com a Diabetes

\* Log of fractional difference (observed vs expected) is calculated for each testing list as: (# genes for the category - # genes expected)/ # genes expected

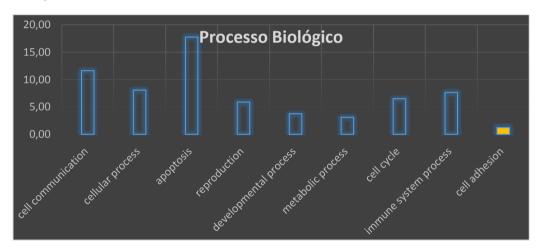


Alzheimer disease-presenil
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Salvage pyrimidine N
T cel
Toll receptor signalir

Wnt signalir

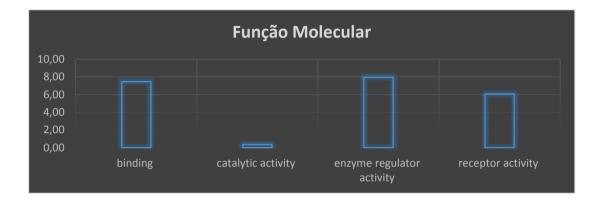
		DMT1	DMT1	Diferença	
Biological Process	DMT2	esperado	observado	fracional	P value
cell communication	85	1,031	13	11,61	0,00
cellular process	118	1,431	13	8,08	0,04
apoptosis	22	0,267	5	17,74	0,02
reproduction	12	0,146	1	5,87	0,57
developmental process	52	0,631	3	3,76	0,72
metabolic process	221	2,680	11	3,10	0,96
cell cycle	33	0,400	3	6,50	0,40
immune system process	105	1,274	11	7,64	0,09
cell adhesion	37	0,449	1	1,23	0,93
response to stimulus	72	0,873	8	8,16	0,11
Total Oraloma normal	419				
Total Diabetes 1	29				

<sup>\*</sup> Log of fractional difference (observed vs expected) is calculated for each testing list as: (# genes for the category - # genes expected)/ # genes expected





Função Molecular				Diferença fracional	p-value
binding	146	1,7708072	15	7,47	0,05
catalytic activity	182	2,2074446	3	0,36	1,00
enzyme regulator activity	46	0,5579256	5	7,96	0,21
receptor activity	35	0,4245086	3	6,07	0,44

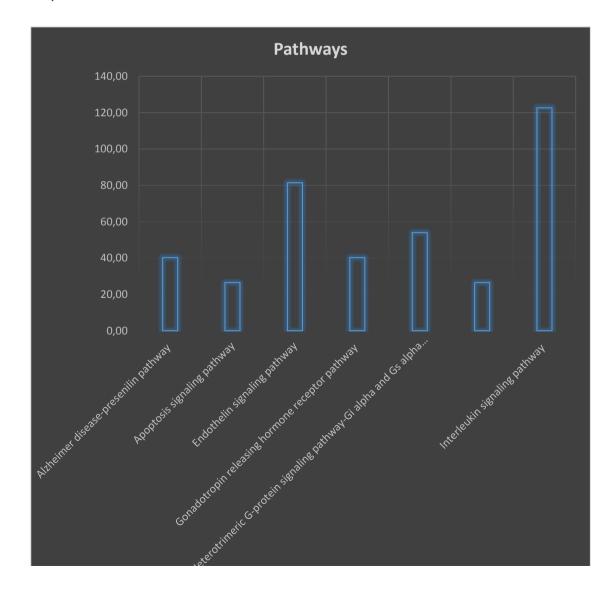


<sup>\*</sup> Log of fractional difference (observed vs expected) is calculated for each testing list as: (# genes for the category - # genes expected)/ # genes expected

		DMT1
Pathway	DMT2	esperado
Alzheimer disease-presenilin pathway	4	0,05
Apoptosis signaling pathway	3	0,04
Endothelin signaling pathway	2	0,02
Gonadotropin releasing hormone receptor pathway	6	0,07
Heterotrimeric G-protein signaling pathway-Gi alpha and Gs alpha mediated pa	3	0,04
Inflammation mediated by chemokine and cytokine signaling pathway	12	0,15
Interleukin signaling pathway	2	0,02
Interferon-gamma signaling pathway	1	0,01
Wnt signaling pathway	2	0,02
	419	
	29	

Devido ao nº elevado de Pathways relativo ao OralOma Normal (125), apresentei apenas os comuns com a Diabetes

\* Log of fractional difference (observed vs expected) is calculated for each testing list as: (# genes for the category - # genes expected)/ # genes expected



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DMT1	Diferença	
observado	fracional	
2	40,22	0,03
1	26,48	0,19
2	81,45	0,01
3	40,22	0,01
2	53,97	0,02
4	26,48	0,01
3	122,67	0,00
1	81,45	0,07
1	40,22	0,13