Exploiting the bead injection LOV approach to carry out spectrophotometric assays in wine: Application to the determination of iron

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Keywords: Solid phase spectrometry Sequential injection lab-on-valve Bead injection Iron Wine A sequential injection lab-on-valve (SI-LOV) system was used to develop a new methodology for the determination of iron in wine samples exploiting the bead injection (BI) concept for solid phase extraction and spectrophotometric measurement. Nitrilotriacetic Acid (NTA) Superflow resin was used to build the bead column of the flow through sensor. The iron (III) ions were retained by the bead column and react with SCN⁻ producing an intense red colour. The change in absorbance was monitored spectrophotometrically on the optosensor at 480 nm. It was possible to achieve a linear range of 0.09–5.0 mg L⁻¹ of iron, with low sample and reagent consumption; 500 μ L of sample, 15 μ mol of SCN⁻, and 9 μ mol of H₂O₂, per assay. The proposed method was successfully applied to the determination of iron in wine, with no previous treatment other than dilution, and to other food samples.

1. Introduction

The chemical composition of wine is very variable and complex; besides the main organic compounds such as ethanol, sugars, organic acids, polyphenols and proteins, wine also contains several inorganic species dissolved [1–3]. Some trace metals can also affect the organoleptic characteristics of the wine, including the flavour, freshness, aroma, colour, and taste [1]. Moreover, the content of some metals can be used to prove wine authenticity and quality. It is known that concentrations higher than 5 mg L^{-1} can cause several changes on the stability of the product and can promote oxidation and wine aging [4,5]. The natural concentration in wine depends principally on the type of the soil where the grapes are produced [2,6-8]. The official method recommended for the determination of iron in alcoholic beverages by OIV and AOAC is based on atomic absorption spectrometry (AAS), either with flame or with furnace atomisation, and is the most widely used method for this determination [3,9].

Molecular absorption spectrophotometry has been the most common detection method used to quantify chemical components [10], but not so common in wine analysis. This might be due to a number of reasons associated with the complexity of the wines. To begin with, metals can be present in wines in different forms, either free or bound to different ligands. Therefore, if the total content is aimed, metals have to be released before or during the colour reaction, either by simple acidification for labile species or by carrying out drastic mineralisation procedures. There are also potential interferences in the colour reactions with a major difficulty arising from the intrinsic colour of the wine. Different flow analysis approaches, like reversed flow [11], merging zones [12], multi-site detection [13], or use of in-line dialysis [14], have been proposed to tackle this problem. Even so, if the background absorption is very high, the first two approaches may not be efficient, while the use of in-line dialysis might induce a too high dilution. Additionally, in dialysis flow analysis systems, it is difficult to match the diffusion yield of standards and samples, which make difficult to obtain accurate calibration procedures.

Solid phase extraction (SPE) can be an attractive alternative to the above-mentioned processes, because it might provide the preconcentration of the analyte and the discard of interfering agents. In the traditional SPE concept, the detection will occur on the eluted phase; however there is a partial loss of preconcentration capabilities gained in the sorption step [15]. To enhance sensitivity, solid phase spectrometry (SPS) has been developed based on the direct measurement of the light attenuation of adsorbent particles packed in an optical cell, in which the target analyte is concentrated and coloured [10]. Considering that the measurement occurs without elution, SPS turns to be a more sensitive method than the conventional SPE. SPS can be performed by two different approaches: batch or flow method. To employ the batch method, besides being very time-consuming, above-average skills are required to pack the solid particles into the cell. The flow methods are more suitable since they can easily measure the light attenuation in the adsorbed species in the flow-through cell with considerable reduction of the volumes used. However, the implementation of SPE/SPS in flow systems can be tricky, as the solid phase can become too much

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Fig. 1. (a) Configuration of the BI-LOV manifold. (b) Position of the bead column in the flow cell. SP, syringe pump; PP, peristaltic pump; HC, holding coil; FC, flow cell; SCN⁻, 1.5 M; H₂O₂, 0.3 M; W, waste; ST, PTFE stopper, P, PEEK plug; B, beads; FO, fibre optics.

packed and increase overpressure. Additionally, if it is not renewed, it might become saturated, either with the analyte or with the interfering compounds. To overcome this problem, instead of reusing the particles, it would be interesting to renew the optosensor. This is quite feasible using sequential injection, lab-on-valve approach (SI-LOV) [16,17]. The association of SIA with SPS leads us to the concept of bead injection (BI), based on the introduction and trapping of the functionalized beads in the flow conduit [18]. Beads are used as a solid surface to extract the analyte or to accommodate a chemical reaction. A precise volume of sample can be transfused through the bead surface allowing analyte separation and concentration. In the BI format, initially a small amount of micro-spheres is injected into the flow channel, where the beads are trapped and perfused by analyte solutions and auxiliary reagents. The reaction takes place on the bead surface allowing real time monitoring, directly on the solid phase, SPS, or on the eluted liquid phase, SPE [16]. When the bead column is placed in the detection zone it is labelled as flowthrough sensor [19]. In some applications, after the measurement, the beads can be discharged, occurring physical regeneration, or in other cases the sensor surface can be renewed, being considered as chemical regeneration. The physical regeneration of the sensor is an advantage since there is no need for the elution step and its applicability is not limited by the lifetime of the sensor, at the same time any possible contamination is also eliminated. The major interest of using these sensors is the increase on sensitivity and selectivity. The sensitivity is augmented since the analyte is retained in the detection zone, and the selectivity also increases since interfering species are excluded from the bead column, therefore they do not develop any signal. In SPS assay, when the chemical reaction takes place on the bead surface, the baseline must be established after the application of the sample mixture to the bead column. In this way the interference of the colour of the sample can be also reduced. The main difficulty of this approach is to obtain a solid phase that can be applied to the separation, concentration and determination of the chemical species of interest [18].

Beads of different material nature are commercially available and have become useful as surface media for retaining reagents, analytes or reaction products. Sephadex is usually used for the linkage of reagents; Sepharose 4B; agarose; and Aminolink, which provide adequate bead support for bioligand immobilization; Cytopore is the most usual for cell linkage [20]. For the determination of metallic ions several types of beads have been used; Sephadex [19] and Chelex-100 [21] beads have been applied for total iron determinations. The present work employs a commercial available "Nitrilotriacetic Acid (NTA) Superflow resin" which can isolate and concentrate dissolved iron at low pH [22]. This resin was originally designed for high throughput sample clean-up procedures based on the affinity chromatography concept. Recently, NTA Superflow resin was applied for the pre-concentration of total iron in sea [22,23] and river [24] water samples.

The herein proposed methodology exploits the concept of solid phase spectrometry in a SI-LOV format to circumvent some difficulties associated with complex matrices in spectrophotometric measurements, namely a high background sample absorption; it was applied to the determination of iron (III) in wines. At low pH (ca. 2) the Fe (III) ions are retained in the bead column and react with thiocyanate reagent. As a result of the reaction, an intense red colour is produced and is spectrophotometrically monitored at 480 nm.

2. Experimental

2.1. Reagents and solutions

All solutions were prepared from analytical grade reagents, and deionised water (conductivity < 0.1 μ S cm⁻¹) was used throughout the work.

The thiocyanate solution was prepared by dissolving 11.4 g of NH₄SCN in 100 mL of water. The hydrogen peroxide solution (0.3 M) was prepared from the stock solution (Perhydrol, 30% H₂O₂). The iron (III) stock standard solutions (up to 5 mg L⁻¹) were prepared by diluting commercial 1000 mg L⁻¹ iron atomic absorption standards (iron standard solution, iron (III) nitrate, VWR-Spectrosol) in 0.01 M HCl solution. The iron (II) stock standard solution (100 mg L⁻¹) was prepared by dissolving (NH₄)₂Fe(SO₄)₂·6H₂O in 0.01 M HCl solution.

The bead suspension used was a dilution in water to half (w/w) of the commercial stock solution (NTA Superflow resin, highly cross-linked 6% agarose, $60-160 \,\mu$ m of bead diameter, 50% suspension in 30% ethanol, 30510, Qiagen).

2.2. Samples

The wine samples analysed (red table, white table and port wine) were purchased in a local market. When necessary, dilution was carried out as sample pre-treatment. The other samples, olive and soybean brines were purchased in a local market, and had no other sample pre-treatment than filtration followed by dilution, when necessary.

2.3. Apparatus

The SI-LOV system (FIAlab-3500, FIAlab Instruments, Medina, WA, USA) presented in Fig. 1, consisting of a bi-directional syringe pump (2500 μ L of volume), a holding coil, a bi-directional variable speed peristaltic pump, and a lab-on-valve manifold mounted on the top of a six-port multi-position valve, was used. As detection system, a USB 2000 Ocean Optics, charge coupled device (CCD) spectrophotometer equipped with fibre optics (FIA-P200-SR; id: 200 μ m), and a DH-2000-BAL Mikropack, UV/VIS/NIR light source, was used for flow programming and data acquisition. The bead column was built between the two optical fibres (Fig. 1b) and a plug (small piece of yellow PEEK tubing, # 1536, Upchurch Scientific).

2.4. Flow procedure

The SI-BI-LOV flow procedure is summarised in Table 1. The initial steps of A–C, consisted in the preparation of the bead column in the flow cell, where 40 μ L of the bead suspension was packed in the flow cell by 80 μ L of carrier at 10 μ L s⁻¹. Afterwards, the sample and the oxidant were aspirated in a following sequence: 10 μ L of H₂O₂; 250 μ L of sample; 10 μ L of H₂O₂; 250 μ L of sample; 10 μ L of H₂O₂; 250 μ L of sample; 10 μ L of H₂O₂. Then, the sample mixture perfused the bead column in the flow cell; afterwards the reference scan was performed to establish the absorbance baseline. The change in the absorbance was monitored at 480 nm while 10 μ L of SCN⁻ drove through the bead column. After the measurement, the physical regeneration of the flow-through sensor was carried out by the aspiration of the beads to the holding coil and subsequent discharging to waste.

For cleaning the sample circuit, the analytical cycle was stopped and diluted acid-and-base solutions were sequentially passed through the flow cell, followed by water for rinsing. At the end of the washing cycle, the blank absorbance was registered and, if this value approached the original one, the system was assumed to be cleaned.

2.5. Reference method

A reference method was performed to corroborate the results obtained by the developed method. The method used was a Portuguese reference method [25] for the determination of iron content in alcoholic beverages and spirits by atomic absorption spectrophotometry.

3. Results and discussion

3.1. Study of the flow system

Several physical parameters like sample and reagent volumes, the aspiration sequence and the flow rates used in the flow system, were studied.

The repeatability of bead packing is an essential parameter to assure reproducible and good quality data. The first study was to select the strategy of how to introduce the bead suspension in the flow system. The option was made to keep the bead suspension circulating in a closed circuit connected to the port 5 (Fig. 1) of the LOV system. It is essential to maintain a well-stirred bead suspension, since aspiration of beads from an unstirred sediment results in blockage of the bead aspiration channel. As the nature of the beads can tolerate magnetic stirring, this was the chosen procedure to maintain the beads suspended. A rotating container (placed in a Vortex shaker) was also tested and the repeatability showed no significant difference when compared to the magnetic stirring. The NTA Superflow beads have a bead structure with relatively high percentage of agarose, this composition offers the beads low flow resistance, and easy handling possibilities inside the flow system. Still, the physical integrity of the beads had to be followed during the method development. The beads in agitation and circulation were kept under visual control, with the use of a microscope, and only after 4 days were possible to observe about 7% more of broken beads when comparing with the freshly prepared suspension. Therefore the replacement of the bead suspension could be done weekly. Some physical parameters of the beads packing like: volume of the beads suspension, carrier volume and flow rate were studied, since the light-path geometry must allow sufficient light to penetrate the bead layer and at the same time the target molecules must be on the optical path. The volume of bead suspension was studied in a range from 20 to 40 µL, while the carrier volume was studied in a range from 45 to 100 μ L with a flow rate of bead packing of 2, 5 and 10 µL s⁻¹. This study was carried out based on monitoring the increase of absorbance at 225 nm. The minimum volume of bead suspension used to construct the bead column was $25 \,\mu$ L; up to this volume of suspension there were no sufficient beads to make a reproducible sensor. The best loading conditions for the column preparation was selected based on the time required for establishing a reproducible bead column; it was set as 80 µL of carrier used to propel 40 μ L of bead suspension at 10 μ L s⁻¹.

As this work is based on a solid phase reaction, the easiest way to establish the maximum binding capacity of the beads was to study the effect of the sample loading volume. The volume of sample was varied in a range from 100 to 600 μ L. This study was performed with a 10 mg L⁻¹ Fe³⁺ standard solution, 1.5 M of SCN⁻ and 0.3 M of H₂O₂. Over 500 μ L of sample, there was no significant increase on the analytical signal; therefore, 500 μ L was the volume of sample chosen for the work. In this case, the maximum load of Fe³⁺ corresponds to 250 mg Fe³⁺ L⁻¹ of commercial beads suspension.

The flow rate used to propel the sample towards the bead column was studied in a range from 3 to $15 \,\mu L \, s^{-1}$. The analytical signal increased with the increase of the flow rate up to $9 \,\mu L \, s^{-1}$. When the flow rate was augmented from 9 to $11 \,\mu L \, s^{-1}$, it was possible to observe an increase on the analytical signal of 10%, but at the same time the relative standard deviation of the signal increased from 2% to 5%, therefore as compromise between increase on the analytical signal and repeatability $9 \,\mu L \, s^{-1}$ was chosen for further work.

With the physical conditions defined, the effect of the concentration of the reagents on the analytical signal was studied. Calibration curves up to 8 mg L^{-1} of Fe³⁺ were performed. The concentration of SCN⁻ was set at 1.5 M and the volume of the reagent was altered to study its effect on the sensitivity of the method. When the quantity is augmented from 7.5 to 15 µmol per assay the sensitivity increased about 25%. Over this value, the linearity of the calibration curve is deteriorated; therefore the chosen quantity was 15 µmol per assay.

Given that the thiocyanate reacts with iron (III), it is necessary to ensure that the iron is in the oxidized form, thus an in-line oxidation is performed. Therefore, three steps were added to the analytical cycle, in order to sandwich the sample between oxidant (hydrogen peroxide) plugs. Since the plug of sample is long ($500 \,\mu$ L) it was divided into two, and the oxidant was aspirated before, in the middle, and after the sample. The volumes of oxidant were tested for 10 and 20 μ L per plug. Different aspiration sequences were studied using a standard of 4.8 mg L⁻¹ Fe³⁺ and 0.4 M hydrogen peroxide. It was possible to reduce the volume of the oxidant to 10 μ L per plug since no significant loss on the analytical signal was observed. The concentration of oxidant was also studied in a range from 0.04 to 0.5 M and the analytical signal increased with the concentration of hydrogen peroxide up to 0.3 M. The analytical cycle was repeated with a 4.8 mg L⁻¹ of Fe²⁺ solution and the results were similar to

Table 1Flow protocol sequence of the developed method.

Step	Description	Volume (µL)	Flow rate ($\mu L s^{-1}$)	Selection valve position
А	Aspirate carrier to SP	1000	250	_
В	Aspirate beads to HC	40	20	5
С	Propel beads to FC	80	10	2
D	Aspirate H ₂ O ₂ to HC	10	50	4
E	Aspirate sample to HC	250	50	3
F	Aspirate H ₂ O ₂ to HC	10	50	4
G	Aspirate sample to HC	250	50	3
Н	Aspirate H ₂ O ₂ to HC	10	50	4
Ι	Send sample mixture to FC	1100	9	2
J	Reference scan, absorbance scanning	_	-	-
K	Aspirate SCN ⁻ to HC	10	50	6
L	Send SCN ⁻ to FC	75	3	2
M	Aspirate beads from FC, by reversed flow	100	300	2
Ν	Dispense beads to waste	425	300	1

those obtained with a standard of 4.8 mg L^{-1} of Fe³⁺, therefore the 0.3 M concentration was chosen for further work.

The standard solutions of iron were prepared at pH 2. As wine samples have pH values around 3.5, the effect of the sample acidity on the reaction was also studied. Calibration curves were performed with standard solutions prepared in acidic medium at pH of 2 and at 3.5. The values obtained for the sensitivity of the curves at pH 2 and 3.5 showed no significant difference.

3.2. Study of interfering species

The study of potential interfering species was performed considering the typical composition of wine samples. The effect of ethanol was studied in a range up to 20% (v/v) and there was no significant variation of the sensitivity of the method. Since wine has a complex matrix, the standards were prepared in a wine model solution containing 10% (v/v) of ethanol, $3 \, g \, L^{-1}$ of sugars (40:60; glucose:fructose), $7 \, g \, L^{-1}$ of glycerol, 100 mg L^{-1} of Mg²⁺, 100 mg L^{-1} of Ca²⁺, 50 mg L^{-1} of Na⁺ and $4 \, g \, L^{-1}$ of tartaric acid, to evaluate the effect of matrix interference in the determination. There was no significant difference found on the sensitivity obtained in the analysis of standards prepared in water and for the ones prepared in wine model solution, as it can be concluded from the values of the sensitivity obtained, 0.134 \pm 0.008 and 0.137 \pm 0.010, respectively.

3.3. Figures of merit of the method

The performance of the developed method was evaluated in terms of reagent and sample consumption, determination rate and dynamic range. The method presented a sample consumption of $500 \,\mu\text{L}$ per assay and a consumption of SCN⁻ and H₂O₂ of 15 μ mol and 9 μ mol per assay, respectively. It was possible to achieve a linear range up to 5.0 mg L⁻¹ with a detection and quantification limits [26] of 0.02 and 0.09 mg L⁻¹, respectively.

These figures of merit of the developed method are summarized in Table 2.

The operational stability of the developed system was calculated by performing the calibration procedure under identical physical and chemical conditions between different working days. A calibration curve: Absorbance = 0.138 (\pm 0.006) × [Fe³⁺] (mg L⁻¹) – 0.004 (\pm 0.024); the values in parentheses are the standard deviation for *n* = 8 calibration curves obtained from 5 standards injected 5 times each and performed in a period of five months. Stability can be considered satisfactory, as this data reflects different batches of beads and reagents and daily preparation of standard solutions.

3.4. Sample analysis

A total of 12 (five white and seven red) table wines and four Port (two white and two tawny) wine samples were analysed. No other sample pre-treatment than dilution was required, for some of the samples. To evaluate the accuracy of the developed method, the reference method was also performed. The results obtained in the sample analysis are presented in Table 3.

The results obtained for the developed method are in good agreement with those obtained for the reference method. A linear relationship: $C_{\text{developed meth.}} = mC_{\text{comp. meth.}} + b$, was established for the samples analysed, and the values obtained for the intercept (*b*) and for the slope (*m*) were $-0.025 (\pm 0.470)$ and $0.973 (\pm 0.101)$, respectively. The values presented in parentheses are the limits of the 95% confidence intervals [26].

The previous table indicates that every red table wine sample had to be diluted. The main reason for this is not the high level of iron present but the natural colour of the matrix. Given that this is a spectrophotometric assay, the baseline absorbance must be established after the sample mixture contacts the bead column. For red table wines, this was not possible due to the natural colour of the matrix. The possibility to use a reduced sample volume as an alternative to the off-line dilution of the sample was tested. A red table wine with a volume of 150 μ L, as opposed to 500 μ L, was introduced in the system, packed between two plugs of 10 µL of H₂O₂. Even though a decrease on the sensitivity of the method of about 50% was observed, the result obtained for the analysis of a red table wine sample for the developed method (1.45 ± 0.19) was in good agreement with the result obtained for the reference method (1.4 ± 0.1) . However, for other red table wine samples the same protocol, did not allow to establish the baseline of the measurement. As a further reduction in the sample volume would decrease too much the sensitivity, we concluded that this approach could not be a general solution for every red wine.

Table 2	
Figures of merit of the developed method.	

Parameter	Value
Reagent consumption per assay	
SCN-	15 μmol
H_2O_2	9 µmol
Sample	500 µL
Waste production per assay	2.5 mL
Determination rate	$20 det. h^{-1}$
Dynamic range	$0.09-5.0 \mathrm{mg}\mathrm{L}^{-1}$
LOD	$0.02 mg L^{-1}$
LOQ	$0.09 \text{mg} \text{L}^{-1}$

Table 3

Comparison of the results obtained for the analysis of wine samples by the reference and the developed procedures.

Sample	Ref. met. (mg L ⁻¹) ^a	SI-LOV $(mg L^{-1})^b$	RD% ^c	Off-line treatment
White table wine 1	1.2 ± 0.1	1.4 ± 0.1	16.7	None
White table wine 2	2.1 ± 0.1	2.1 ± 0.1	0.0	None
White table wine 3	9.0 ± 0.4	9.2 ± 0.7	2.2	Dil. $4 \times$
White table wine 4	6.1 ± 0.2	5.4 ± 0.1	-11.5	Dil. $4 \times$
White table wine 5	2.2 ± 0.1	1.9 ± 0.2	-13.6	None
Red table wine 1	4.0 ± 0.2	4.1 ± 0.6	2.5	Dil. $4 \times$
Red table wine 2	5.2 ± 0.4	5.0 ± 0.1	-3.8	Dil. $4 \times$
Red table wine 3	5.5 ± 0.2	4.9 ± 0.6	-10.9	Dil. $4 \times$
Red table wine 4	5.5 ± 0.4	5.4 ± 0.4	-1.8	Dil. $4 \times$
Red table wine 5	5.4 ± 0.1	5.0 ± 0.1	-7.4	Dil. $4 \times$
Red table wine 6	5.3 ± 0.4	6.0 ± 0.8	13.2	Dil. 4×
Red table wine 7	4.9 ± 0.2	4.3 ± 0.5	-12.2	Dil. $4 \times$
Port, white 1	2.7 ± 0.2	2.6 ± 0.3	-3.7	None
Port, white 2	2.7 ± 0.4	2.6 ± 0.1	-3.7	None
Port, tawny 1	4.1 ± 0.4	3.9 ± 0.2	-4.9	Dil. 2×
Port, tawny 2	2.0 ± 0.2	2.1 ± 0.8	5.0	None

^a Mean and standard deviation for n = 3.

^b Mean and standard deviation for n = 5.

^c Relative deviation.

As it can be concluded from Table 3, samples of red table wines and port wines presented distinct and typical concentration levels, therefore other samples were also tested to assess the adaptability of the method to samples of non-wine origin and in different concentration ranges. A certified reference water sample and two different types of brines, olive and soybean, were analysed and the results obtained for the developed method were comparable to those obtained for the reference method (Table 4 in ESI).

4. Conclusions

The use of a lab-on-valve format with bead injection solid phase extraction proved to be a useful tool for the development of a molecular absorption spectrometry alternative to the atomic absorption spectrometry (AAS) reference method for the determination of metals in complex matrices like wine. The proposed procedure also enabled to quantify the total content of the metal, with much simpler equipment. Based on the results (*viz*: good agreement with the reference atomic absorption procedure), it can be concluded that total iron is determined. This can be also deduced from the experimental conditions: the pH, the presence of an oxidant and a strong complexant and the additional off-line dilution of the samples contribute to the shift of the chemical equilibrium in a way that most of the iron originally present in a complexed form may be released, and made available in the form of Fe³⁺.

This flow system incorporating this optosensor also demonstrated to be a useful strategy to avoid interferences from complex matrices with high background absorption values, in a spectrophotometric method. This was possible by using the bead injection procedure to separate the analyte from the matrix; taking advantage of the use of programmable flow. The chosen NTA resin proved to display characteristics appropriate for bead injection. The overall strategy could be adapted to other spectrophotometric determinations in complex matrices.

The results obtained for the analysis of several different food samples are in good agreement with the results obtained for the reference method. When comparing similar approaches [19,27,28] (Table 5 in ESI) the applicability of the developed SIA-BI approach was illustrated using various sample matrixes and in a highly automated fashion.

Regarding the main limitations of the proposed method, we can point out the need for sample dilution if some red wines are analysed. Another difficulty is the cleaning of the flow system after the analysis of red table wines; however, with additional port(s) on the SI-LOV device, the cleaning protocol could be easily incorporated into the analytical cycle.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.talanta.2011.01.041.

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