



METABOLOMICS

The X-windows High-Throughput Mass Spectroscopy Pipeline

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****BioEngineering Lab**

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University of Minho
Braga - Portugal



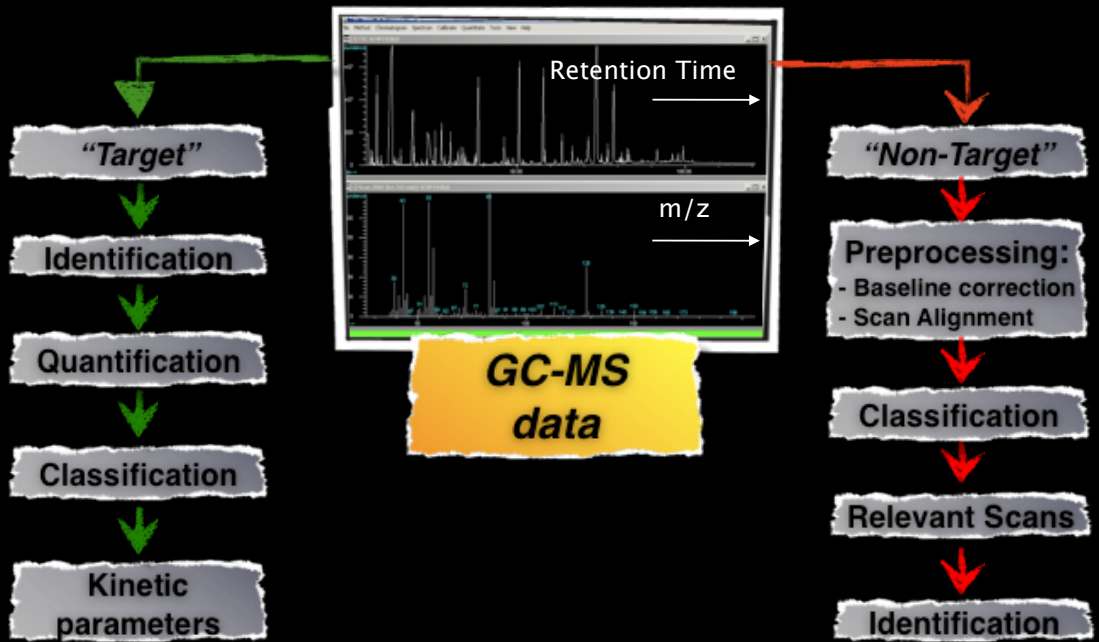
METABOLOMICS
High-Throughput Mass Spectroscopy

In This Talk:

1. Metabolomics: Target vs Non-Target Approaches
2. Mass Spectroscopy Signal Processing
3. The X – Metabolomics Architecture
4. Software Features
5. High-throughput examples
6. Software Demonstration



1. Metabolomics: Target vs Non-Target Approaches





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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing

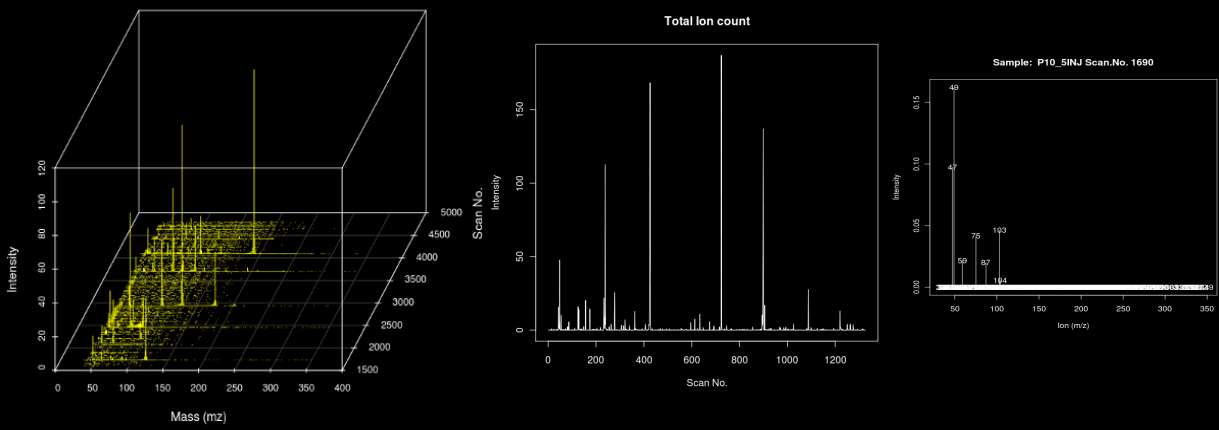
- 2.1. The MS Chromatographic Signal
- 2.2. Main Approaches: Bioinformatics Vs Chemometrics
- 2.3. Pre-Processing
- 2.4. Feature Extraction
- 2.4. Chromatographic Alignment
- 2.5. Robust Peak Recognition
- 2.6. Identification and Composition
- 2.7. High-throughput MS Bioinformatics



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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing

2.1. The MS Chromatographic Signals





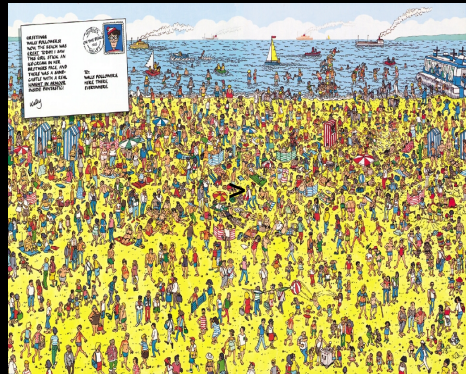
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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing

2.1. The MS Chromatographic Signals



Target
Approach



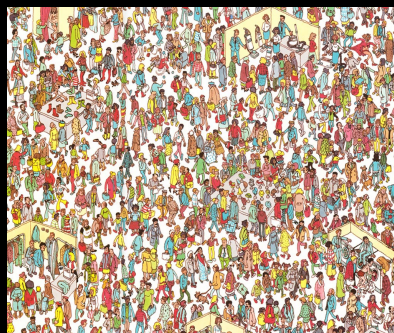
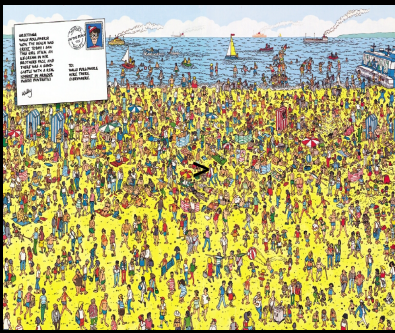
Holistic
Approach



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2. Mass Spectroscopy Signal Processing

2.1. The MS Chromatographic Signals



High-Throughput
MS Signal Processing

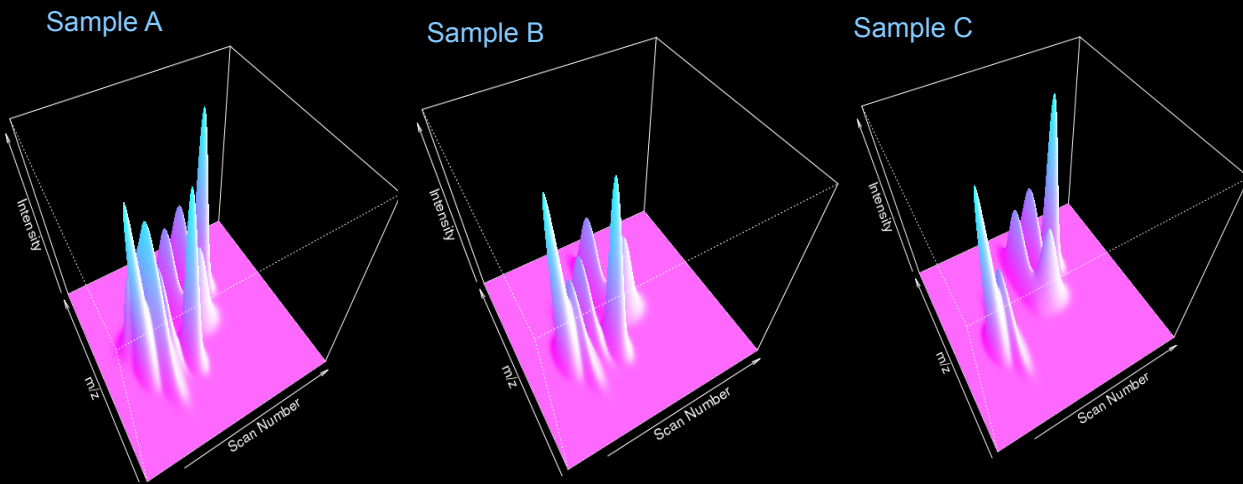


Holistic Approach
Complex Systems
Systems Biology
Systems Chemistry



2. Mass Spectroscopy Signal Processing

2.1. The MS Chromatographic Signals

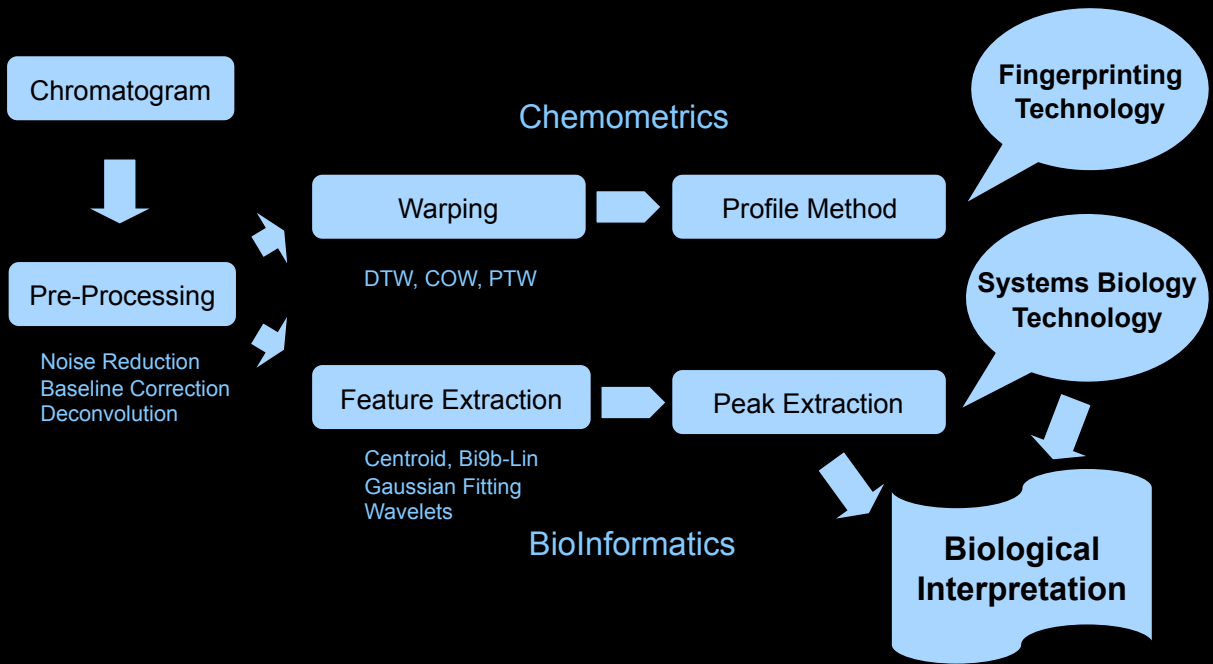


'In-Silico' Processing:
Pre-process, Extract, Recognize, Identify, Quantify, Analyse



2. Mass Spectroscopy Signal Processing

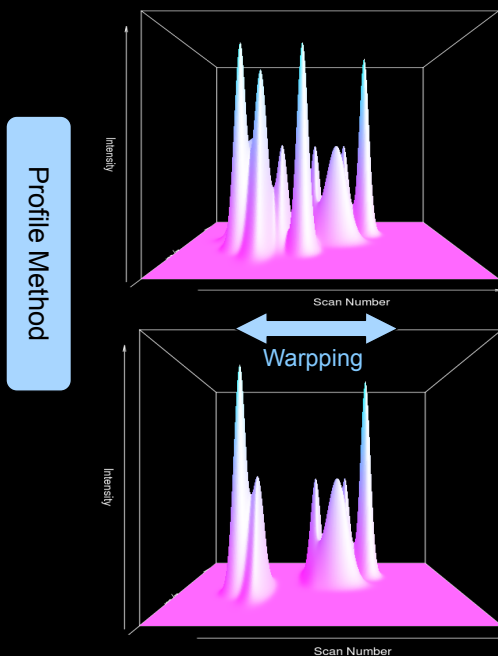
2.2. Main Approaches: Bioinformatics Vs Chemometrics





2. Mass Spectroscopy Signal Processing

2.2. Main Approaches: Chemometrics



Warp the chromatogram respecting an objective function:

1. Dynamic Time Warping: $j = \text{argmin} (y_i - y_r)^2$

2. Correlation Time Warping: $j = \text{argmax} \text{corr}(Y_i/Y_r)$

e.g. Which Reference Chromatogram to use with complex biological samples?

Only robust with low-rank Chromatography – Low complexity of samples → Not for Metabolomics!

Not For Complex Chromatograms!

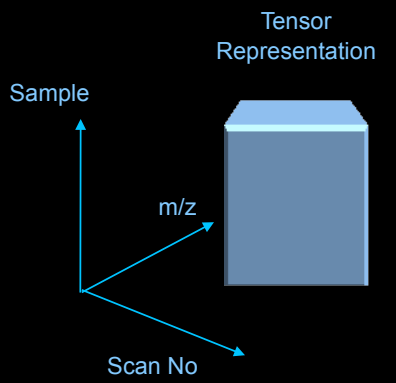
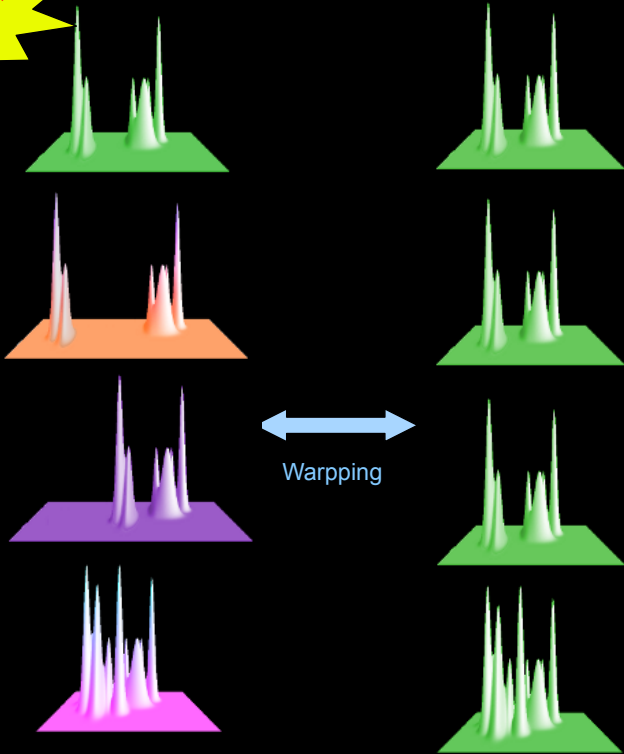


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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing



Profile Method



Multi-Dimensional Linear Algebra

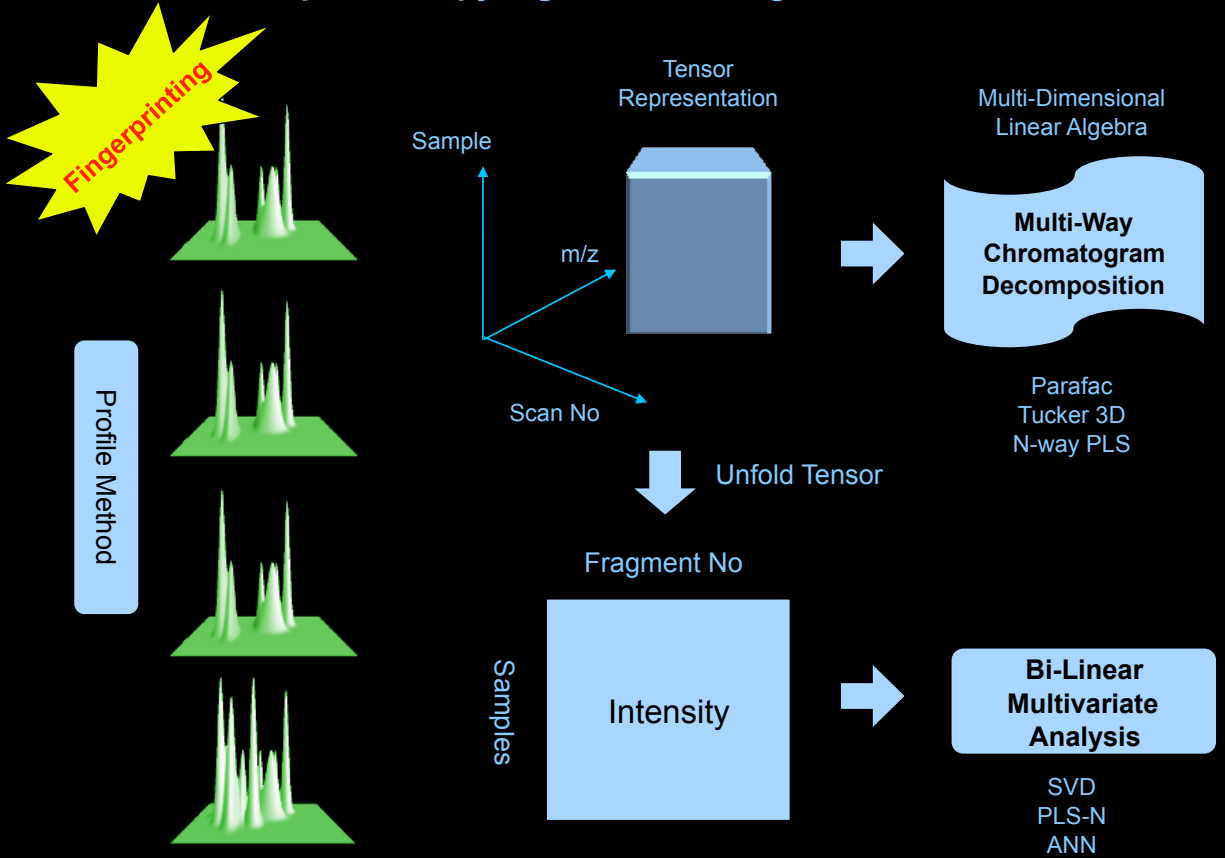
Multi-Way Chromatogram Decomposition

Chemometrics Approach



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High-Throughput Mass Spectroscopy

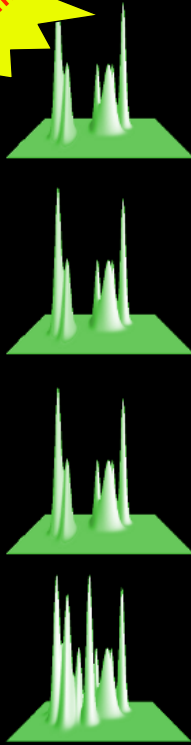
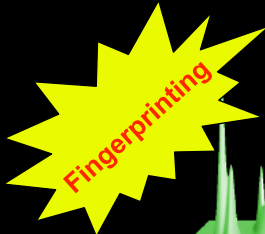
2. Mass Spectroscopy Signal Processing





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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing



Appropriate for
Process
Analytical
Technology

Profile Methods

Not Appropriate
For Complex
Biological
Systems

Slow Convergence

Fingerprinting Only

Low Rank Only

MS Artifacts

Chrom. Artifacts

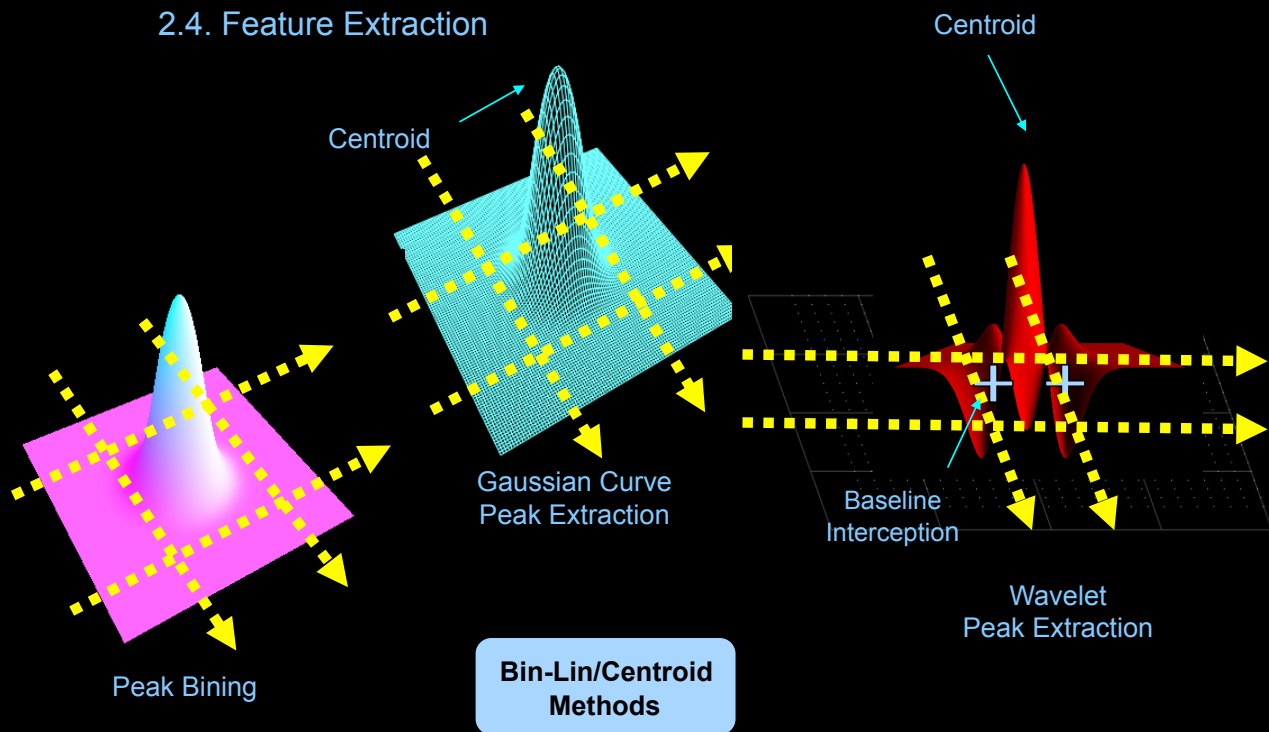
**Difficult to provide High-Throughput Metabolomic
Information for Systems Biology/Chemistry
and Complex Systems Approaches**

Feature Extraction



2. Mass Spectroscopy Signal Processing

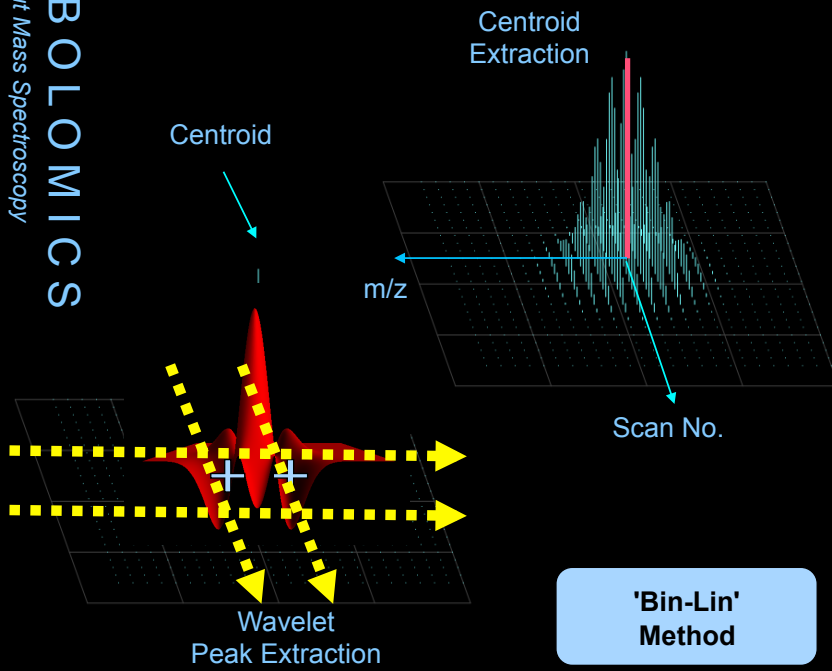
2.4. Feature Extraction





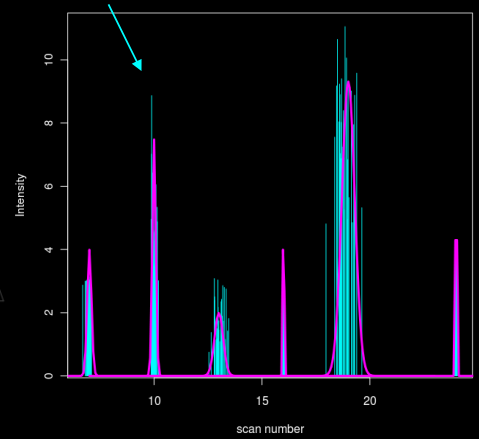
2. Mass Spectroscopy Signal Processing

2.4. Feature Extraction



'Equal'
Sample
Centroids

Peak Grouping



Assuming Centroid
Scan No is Normally
Distributed!



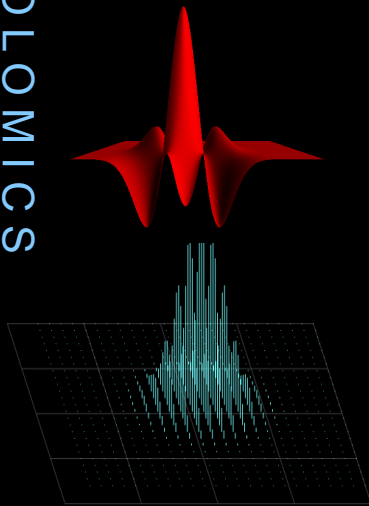
2. Mass Spectroscopy Signal Processing

2.4. Feature Extraction

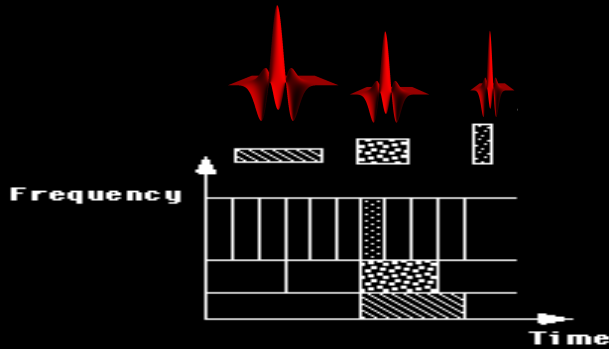
Wavelets:

- Representation of a signal by a new orthonormal space basis given by non-stationary oscillating waveforms;
- Discontinuities and sharp peaks;
- The Mexican Hat Wavelet:

$$\psi(t) = \frac{1}{\sqrt{2\pi}\sigma^3} \left(1 - \frac{t^2}{\sigma^2} \right) e^{-\frac{t^2}{2\sigma^2}}$$



Wavelet
Peak Extraction



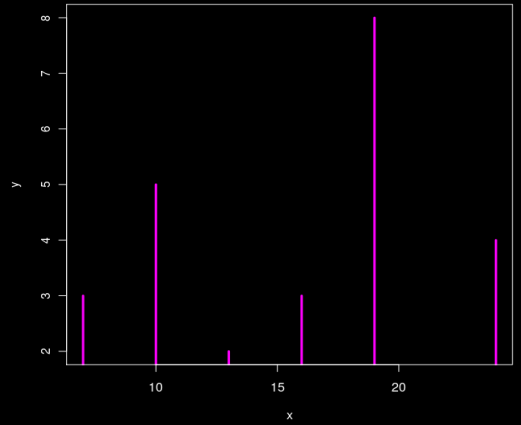
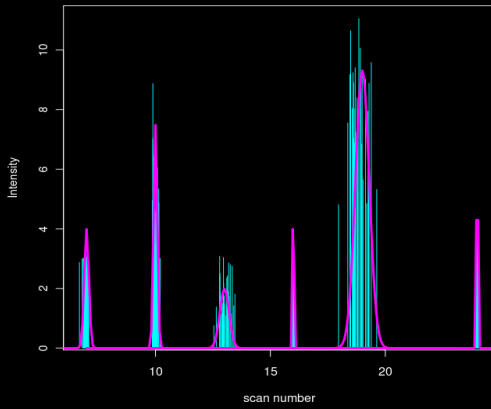
**Multi-Scale
Chromatogram
Decomposition**



2. Mass Spectroscopy Signal Processing

2.4. Feature Extraction

Synchronization



Centroid Retention Time
Correction Among Samples

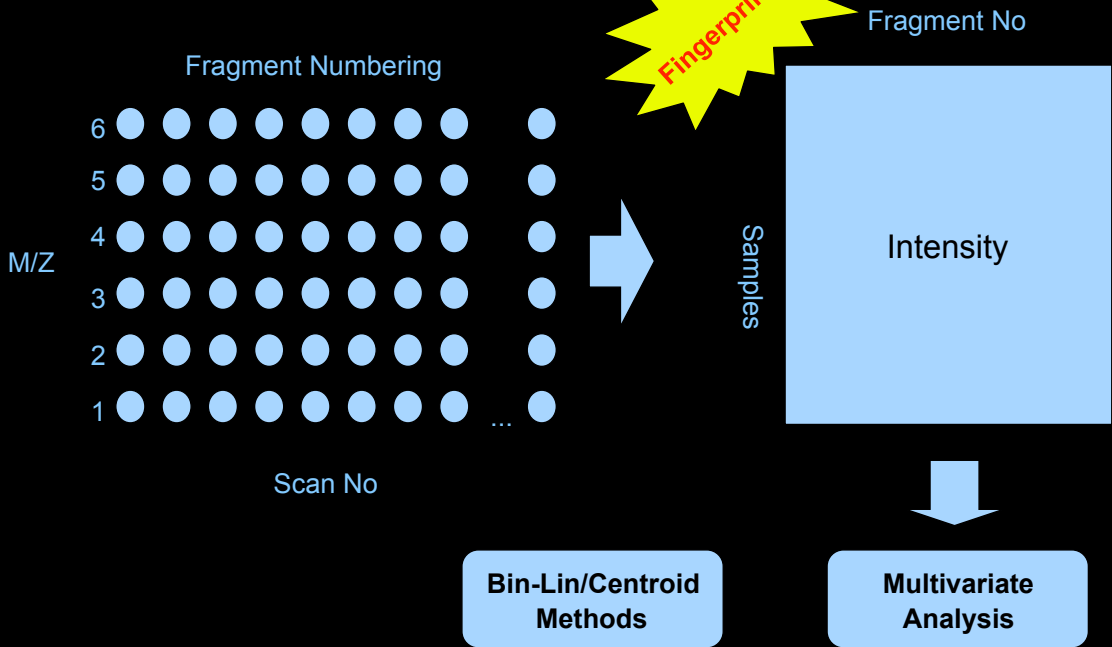
**Bin-Lin/Centroid
Methods**

Interpolation functions
To correct deviation in
'non-common' centroids



2. Mass Spectroscopy Signal Processing

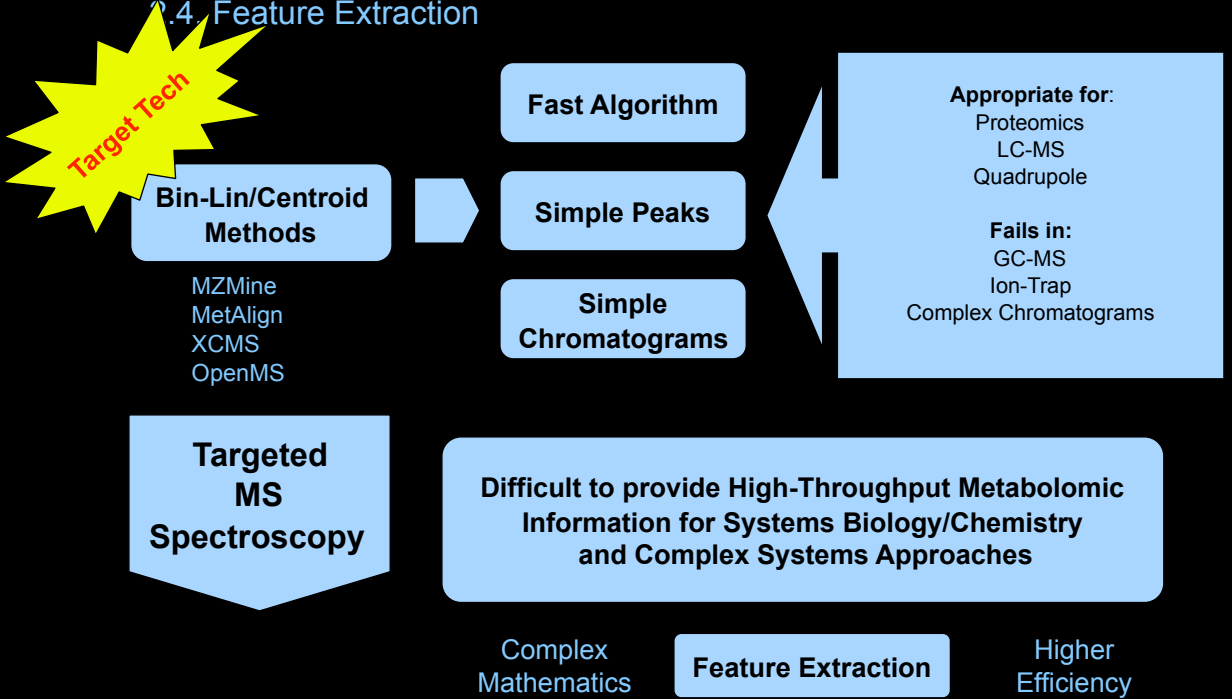
2.4. Feature Extraction





2. Mass Spectroscopy Signal Processing

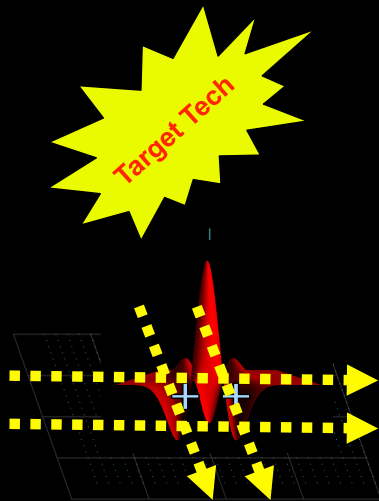
2.4. Feature Extraction



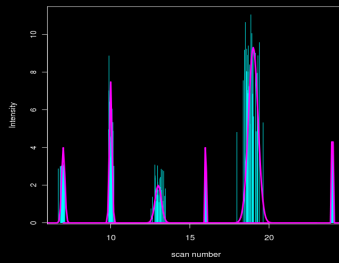


2. Mass Spectroscopy Signal Processing

2.4. Feature Extraction

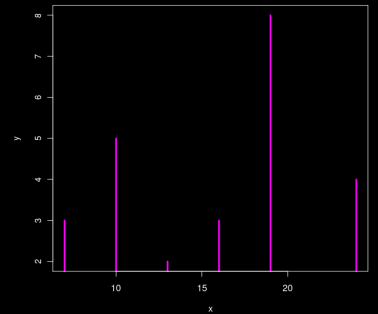


Failure in Multiscale
Extraction
(e.g. loss of small
peaks)



Failure
In Compound Grouping
(e.g. Loss of Compounds)

**Bin-Lin/Centroid
Methods**



Failure
In Chromatogram
Reconstruction
(e.g. Colapse of adjacent
Scans, loss of fragments)



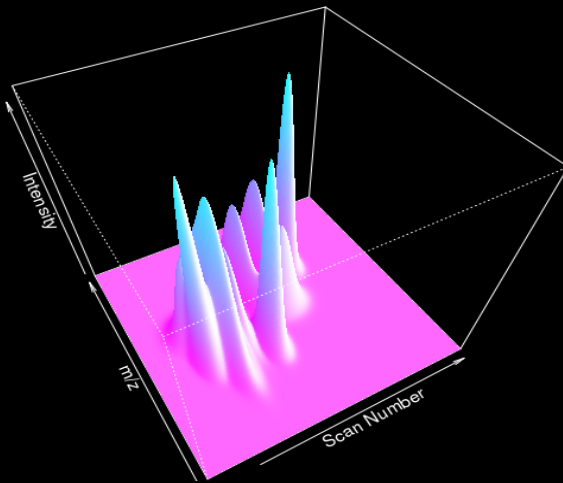
2. Mass Spectroscopy Signal Processing

2.4. Feature Extraction

How to provide
High-Throughput
Metabolomic
Information?

Signal Processing
Feature Extraction

Standards for 'In-silico'
High-Throughput Chromatography
In Metabolomics



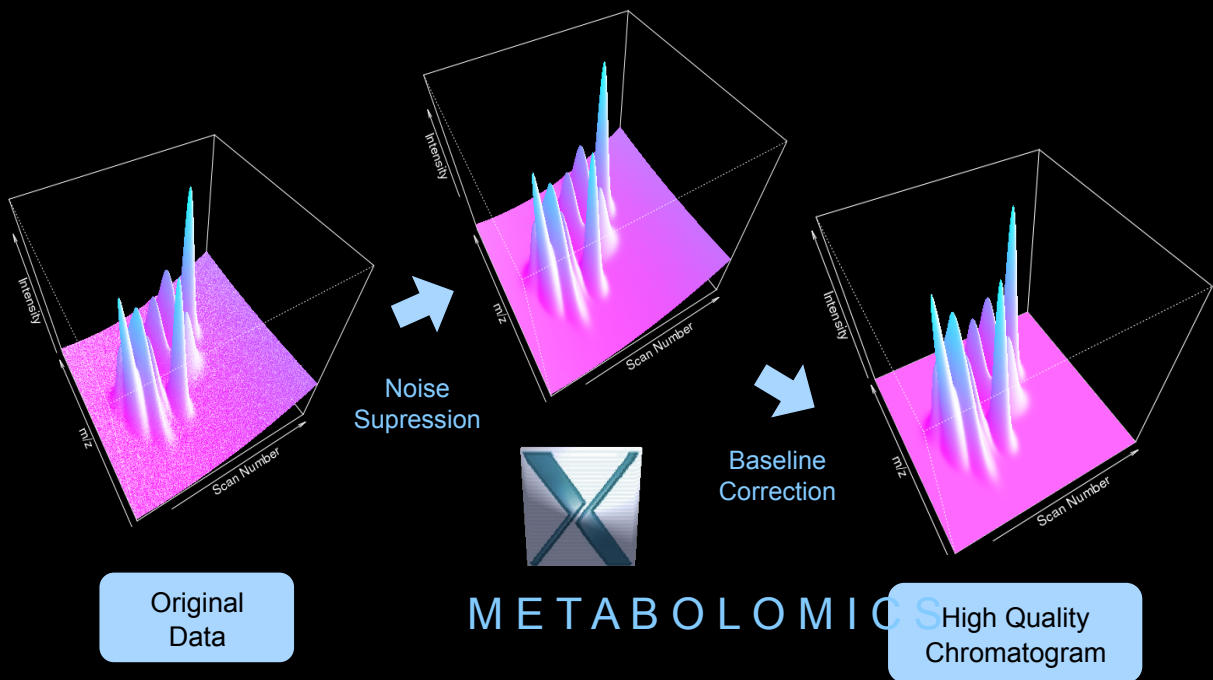
High Quality Signals
Peaks High Definition
High Peak Concentration
Filters
Deconvolution
Recognize,
Identify, Quantify,
Analyse, Quality Control
Biological Interpretation



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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing

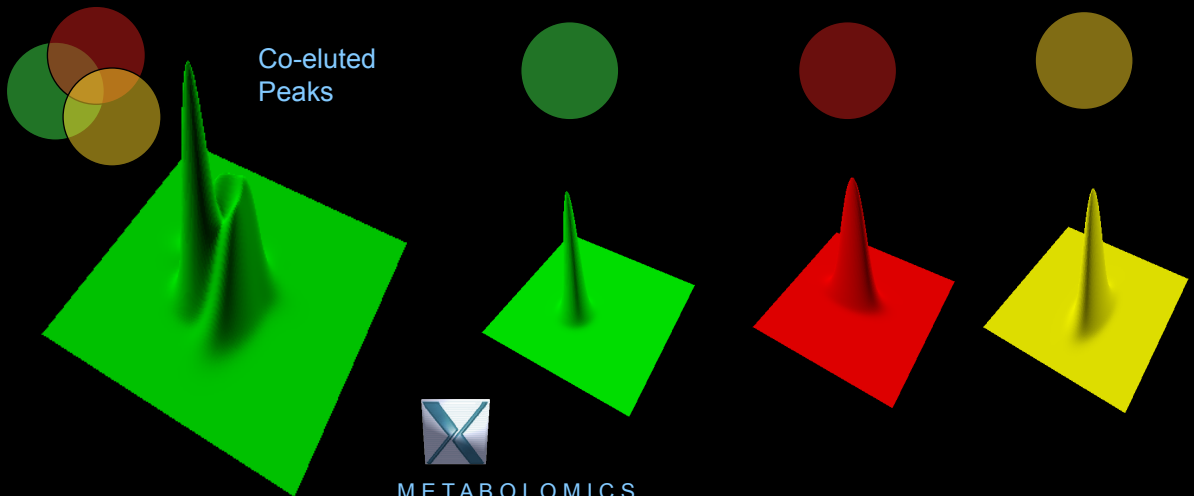




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High-Throughput Mass Spectroscopy

2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing



Convolution
Detection

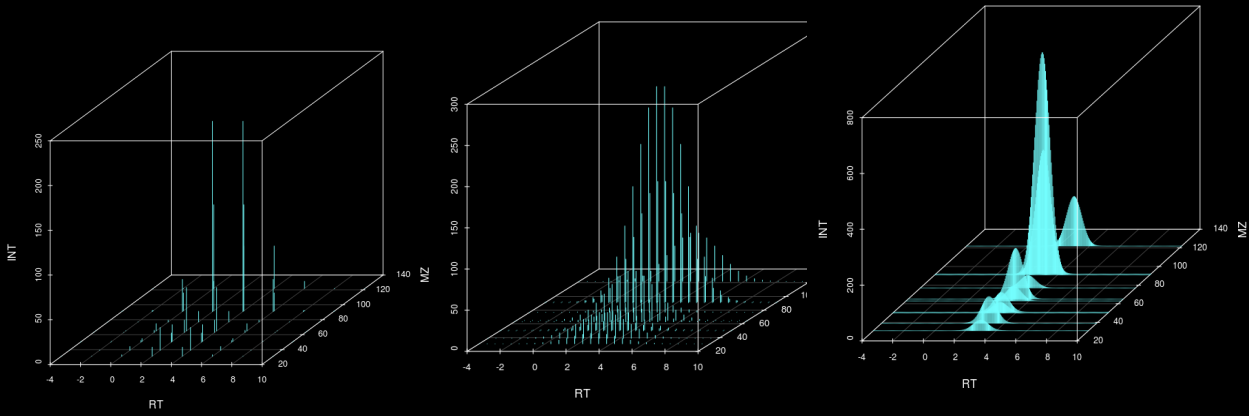
'In-Silico' Deconvolution for
High-Resolution Feature Extraction



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2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing



**Scan Rate Influence on
High-Resolution Feature Extraction**



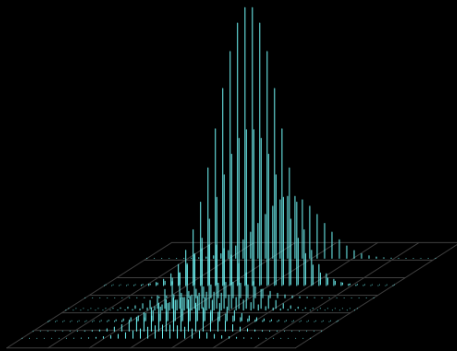
**Minimum of
5 to 8 scans**



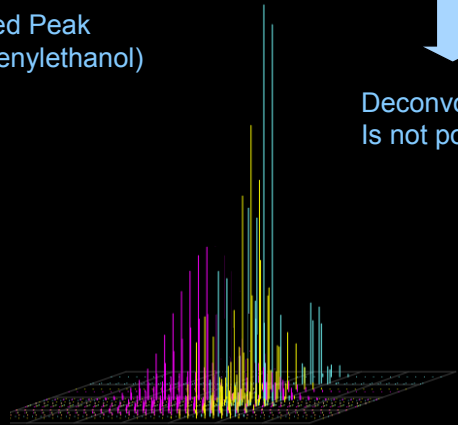
2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing

Regular Peak
(e.g. phenylethanol)



Saturated Peak
(e.g. phenylethanol)



Feature Randomization
During on-Trap
Saturation

Deconvolution
Is not possible

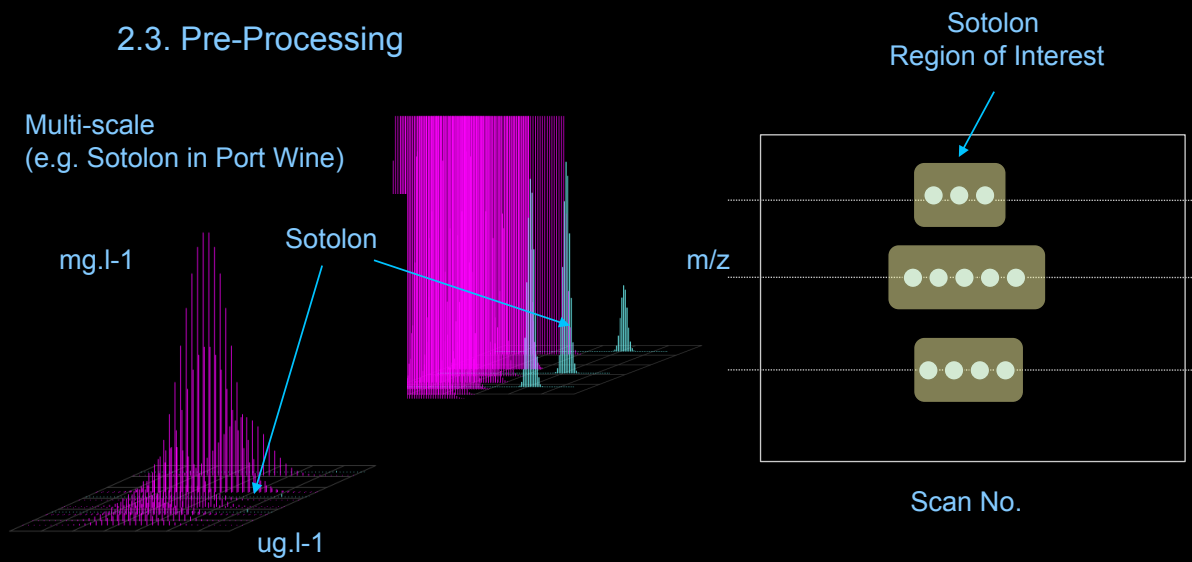
Saturation

Saturation
Filter



2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing



Multi-scale Feature Extraction

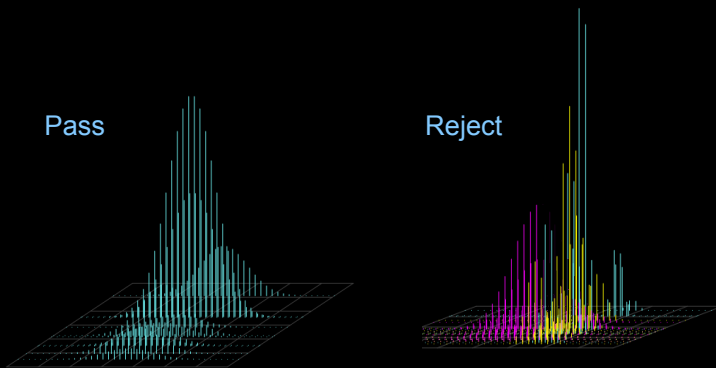


Increase in
High-Throughput
Detail



2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing



Fuzzy Filtering
using Sinkhorn
Factorization!!!

Peak = $D_1 A D_2$
(perform until
convergence)

- Inside a sample and
Between samples!!!

Feature Self-Consistency Test



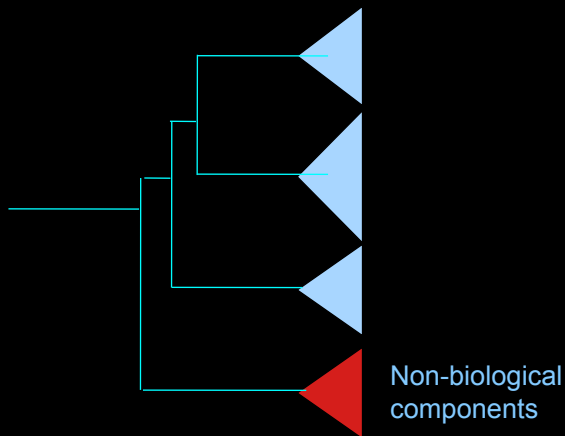
Only
Robust Peaks
Pass



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2. Mass Spectroscopy Signal Processing

2.3. Pre-Processing



Correlation Between fragments to obtain The clusters of Non-biological components

Check each cluster According to Tikunov et al (2005)

Clean Non-Biological Components



Robust Databases



METABOLOMICS
High-Throughput Mass Spectroscopy

3. The X-Metabolomics Architecture

- 3.1. Operating Systems
- 3.2. Software Compatibility
- 3.3. File Formats
- 3.4. Supported Equipments
- 3.5. Software Architecture in a Nutshell

In: Castro, CC, Teixeira, JA, Silva-Ferreira, AC, Martins, RC. 2009. X-Metabolomics: A high-throughput GC-MS metabolomics pipeline for Saccharomyces cerevisiae. BMC Bioinformatics, Submitted.



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High-Throughput Mass Spectroscopy

3. The X-Metabolomics Architecture

3.1. Operating Systems - UNIX Like Platforms

Currently Available for:



Ubuntu Linux:
8.04 (Discont.)
8.10 (Stable)
9.04 (Stable)
9.10 (Testing)



Mac OS X
Snow Leopard



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3. The X-Metabolomics Architecture

3.2. Software Compatibility



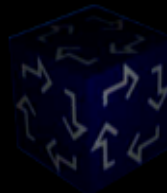
Xorg



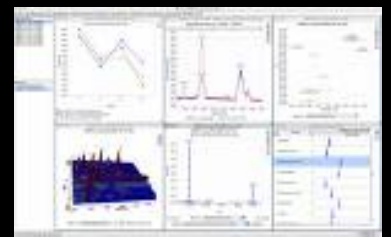
R-project



Bioinformatics Platforms



OpenPAT
Plug-In



MZmine

MetAlign



Markup Languages
and DataBases

Mass Spectroscopy
Processing Software



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3. The X-Metabolomics Architecture

3.3. File Formats

ASCII Text Files

MZ XML (Proteomics)

NetCDF (Preferential!!!)
(Use Mass Transit for conversion)



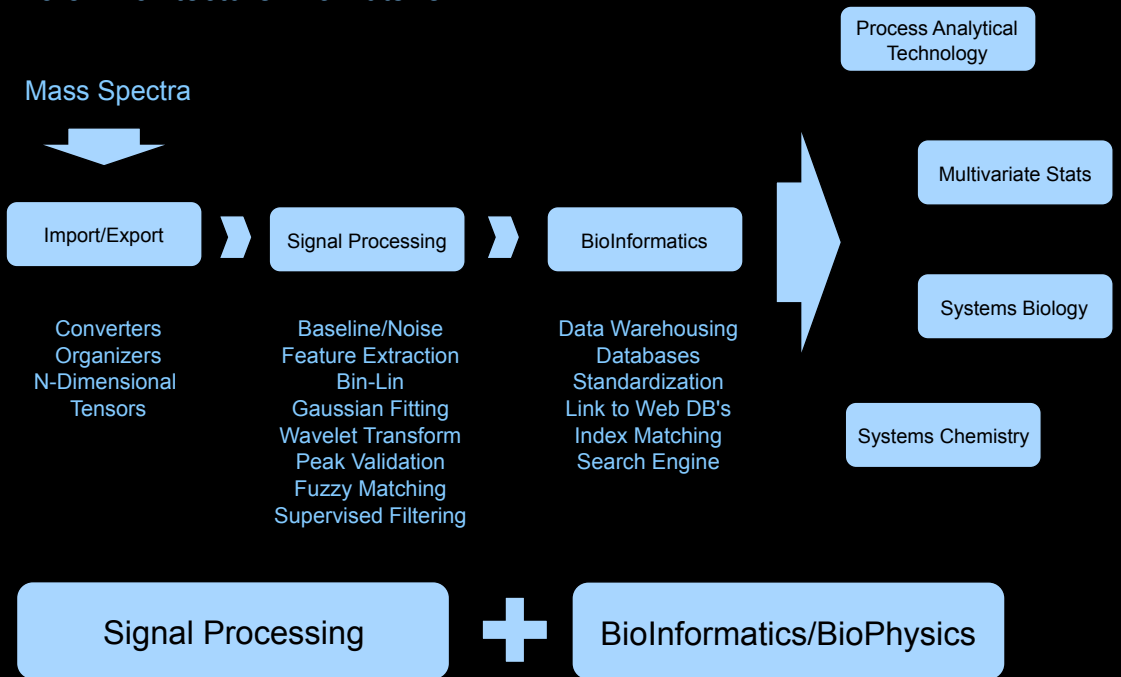
NetCDF (network Common Data Form)
Multi-Dimensional Array-oriented Scientific Data
Useful for Complex MS/MS datasets



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High-Throughput Mass Spectroscopy

3. The X-Metabolomics Architecture

3.5. Architecture in a nutshell





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High-Throughput Mass Spectroscopy

4. Software Features

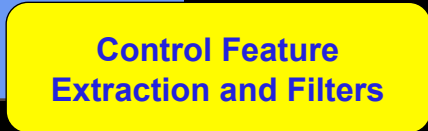
- 4.1. Configuration
- 4.2. File Import/Export
- 4.3. Pre-processing
- 4.4. Filtering
- 4.4. Fingerprinting Diagnostics
- 4.5. MS Quality Control Charts
- 4.6. Unsupervised Metabolomics
- 4.7. Identification and Composition Table
- 4.8. Co-Expression Pathway Analysis
- 4.9. Time Course Metabolomic Analysis



4. Software Features

4.1. Configuration

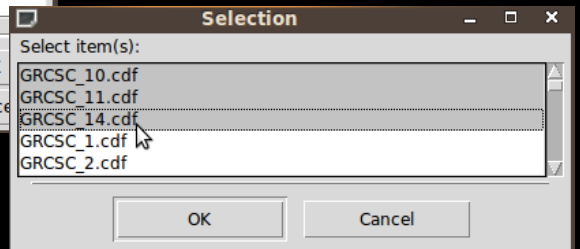
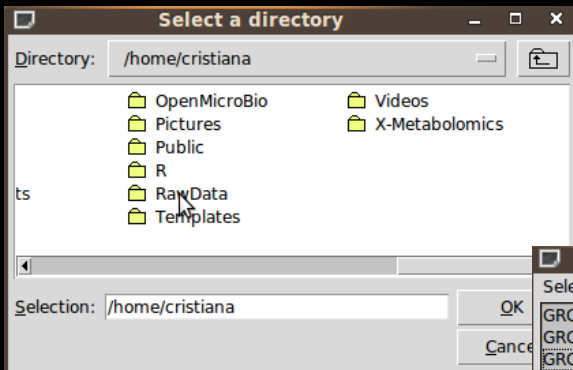
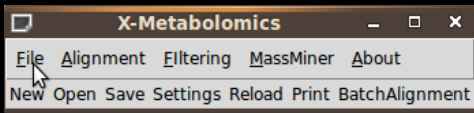
```
>> ShowXMetabolomicsSettings()
[1] "You are trying to use ShowXMetabolomicsSettings()"
      BinLin CentWave Group
fwhm      1000      NaN      NaN
mzdiff      1      NaN      NaN
ppm      nan      4.0      NaN
snthresh  nan      0.1      NaN
peakwidthmin nan      15.0     NaN
peakwidthmax nan      60.0     NaN
scanrangemin nan      100.0    NaN
scanrangemax nan      4000.0   NaN
bw      nan      NaN      1.00
minfrac  nan      NaN      0.10
minsamp  nan      NaN      2.00
mzwid    nan      NaN      0.01
max      nan      NaN      50.00
sleep    nan      NaN      0.00
Flag3D   FALSE     NaN      NaN
Filter   0.01      2.0      4.00
```





4. Software Features

4.2. File Import

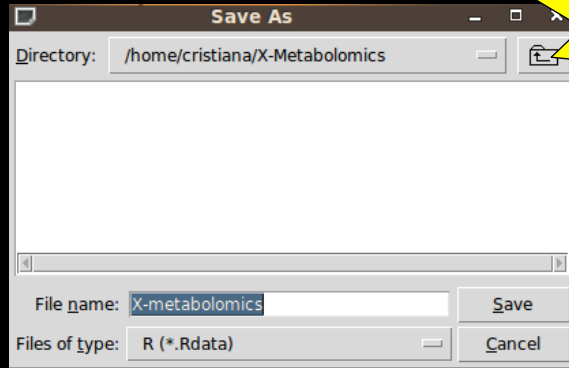
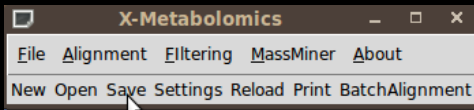




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4. Software Features

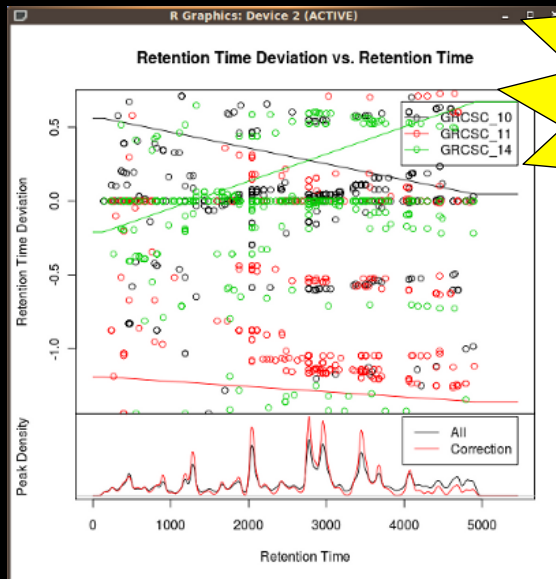
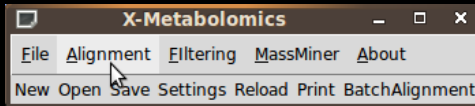
4.2. File Export





4. Software Features

4.3. Pre-processing

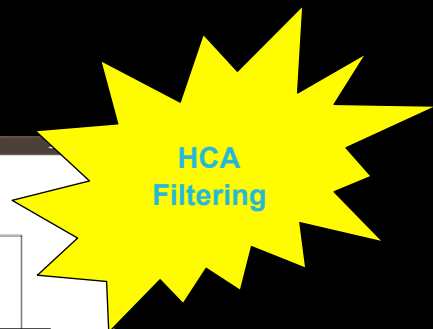
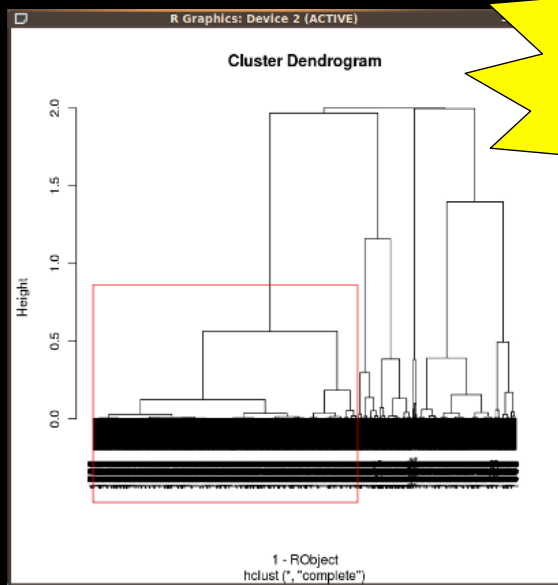
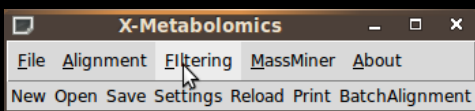


Synchronization



4. Software Features

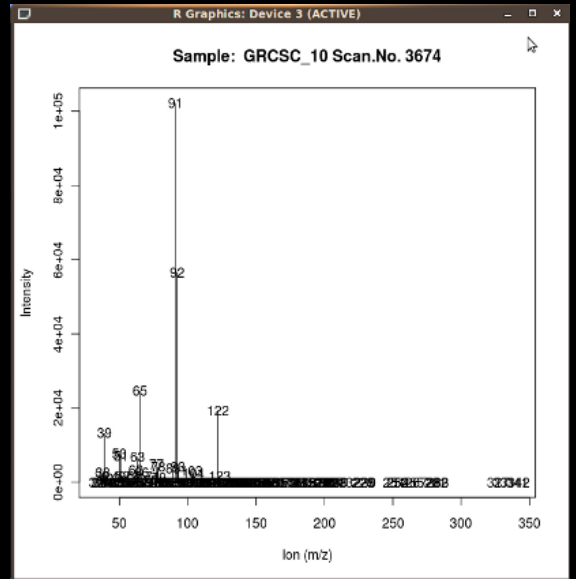
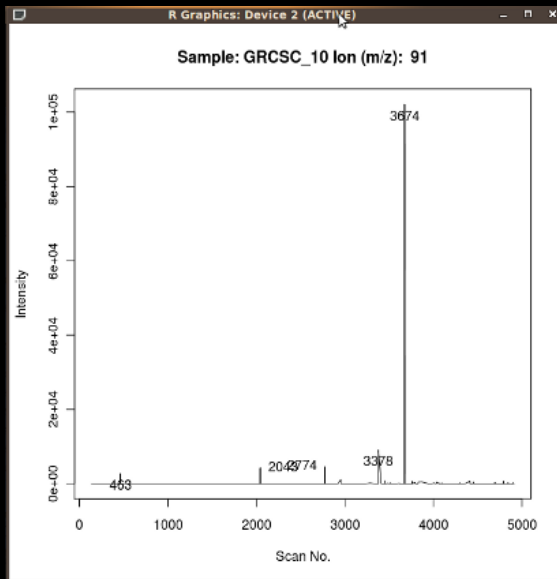
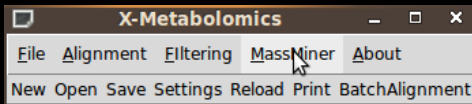
4.4. Filtering





4. Software Features

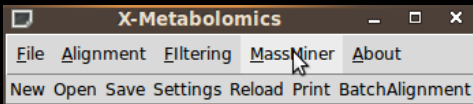
4.5. Fingerprinting Diagnostics



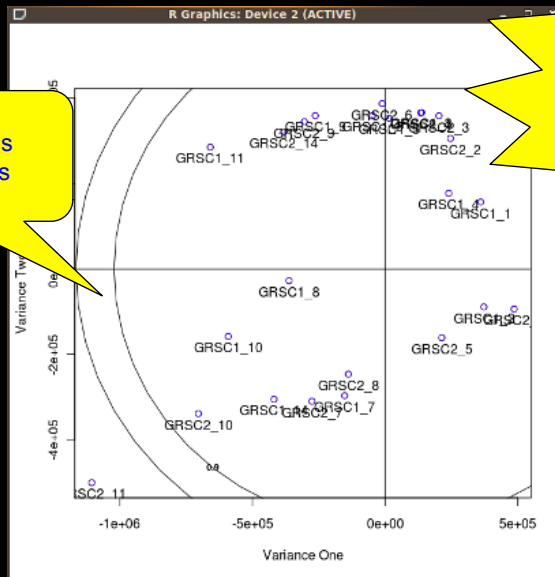


4. Software Features

4.6. MS Quality Control Charts



Control Methods and Equipments



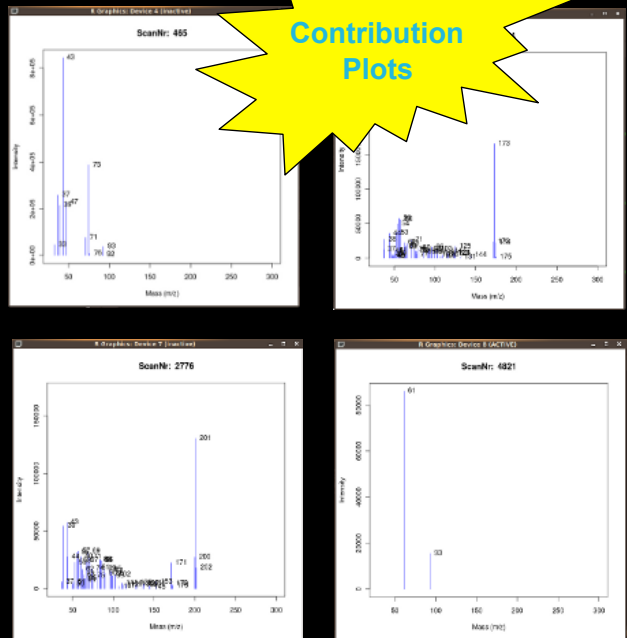
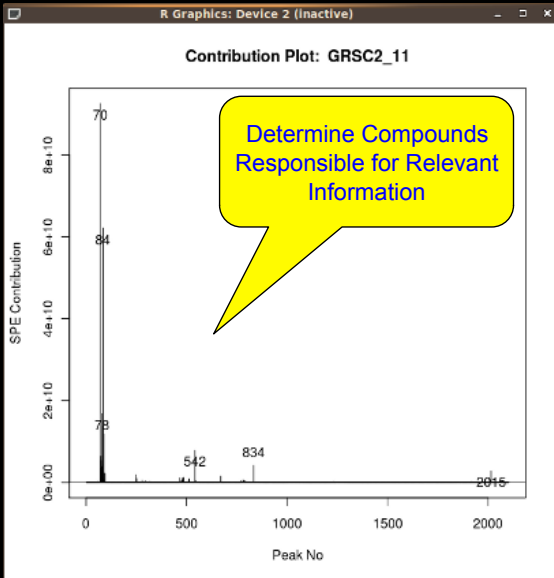
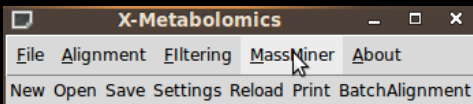
Multivariate Chart Control



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4. Software Features

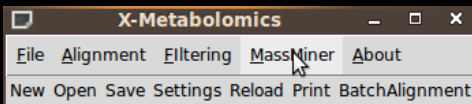
4.7. Unsupervised Metabolomics





4. Software Features

4.8. Identification and Composition Table



Scan.Nr	Mass	Intensity	Mean	RelatPercent	Mean	/(%)
2055	189	281	3631663	100.000000	1117	73
1983	189	208	68337	100.000000	1117	267
10	190	207	100.000000	100.000000	1117	44
11	192	209	100.000000	100.000000	1118	251
12	220	208	100.000000	100.000000	1118	45
13	222	207	100.000000	100.000000	1146	39
14	281	100.000000	100.000000	100.000000	1146	41
7	42	100.000000	100.000000	100.000000	1146	57
9	46	95.0408619	100.000000	100.000000	1146	71
4	4661684	0.4661684	100.000000	100.000000	1186	58
4.4929697	86.8334385	4.4929697	100.000000	100.000000	1186	49
13.1665615	13.1665615	86.8334385	100.000000	100.000000	1186	50
		13.1665615	100.000000	100.000000	1186	51
			100.000000	100.000000	1186	59
			100.000000	100.000000	1186	86
			100.000000	100.000000	1186	87

Relevant Peaks
For Further Identification

Automatic Composition
Systems Biology
Systems Chemistry

Identification
Table!!!

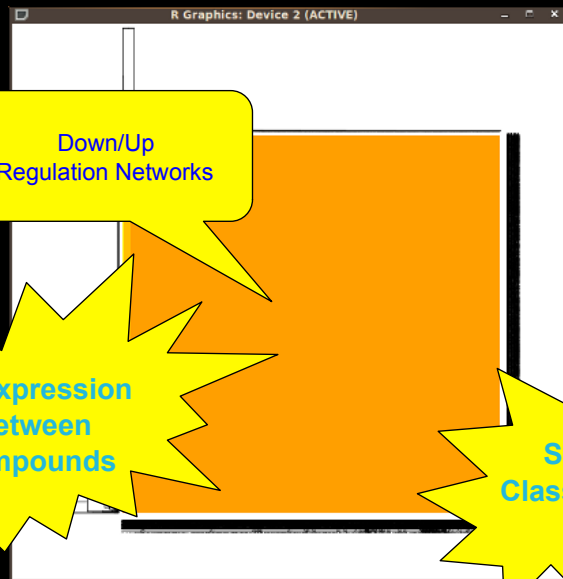
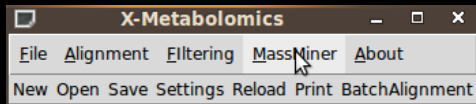
Composition
Table

Post-Processing by
Bioinformatics

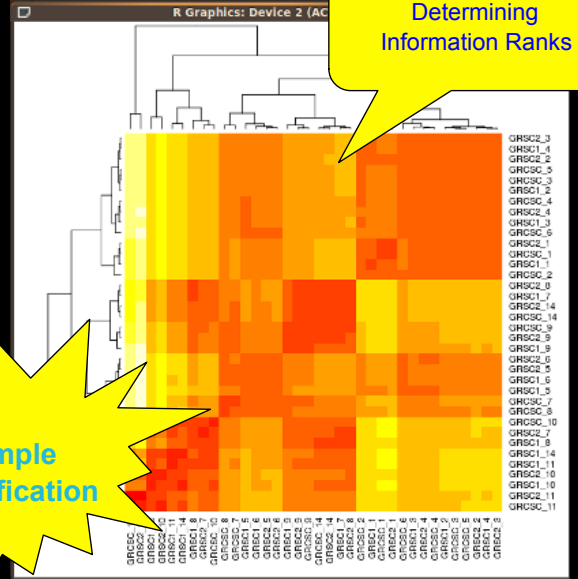


4. Software Features

4.9. Co-Expression Pathway Analysis



Co-expression
Between
Compounds



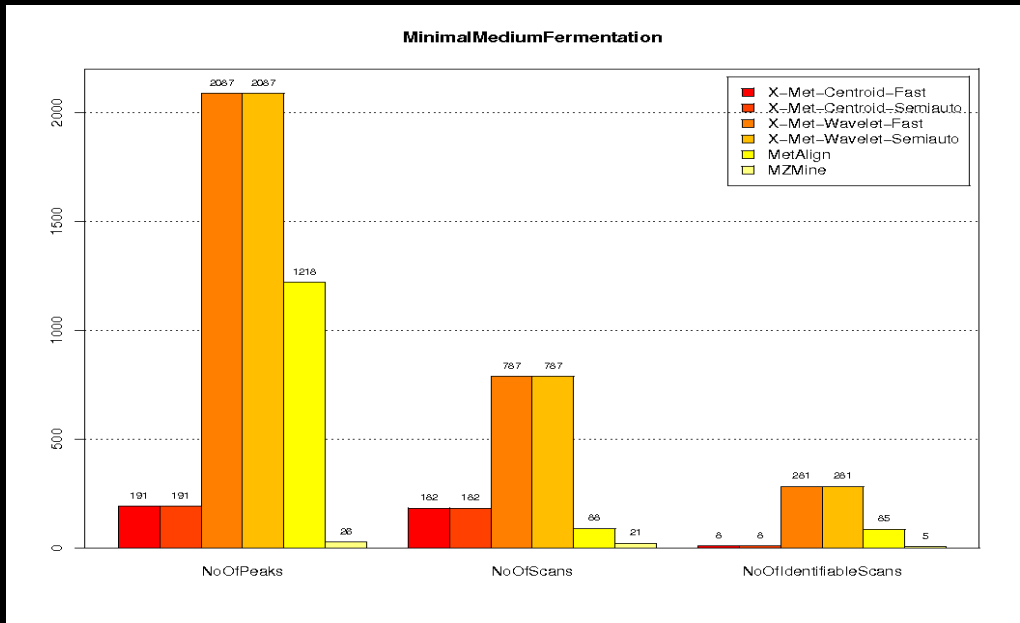
Determining
Information Ranks

Sample
Classification



5. Benchmarks

Minimum Media *Saccharomyces Cerevisiae* Growth

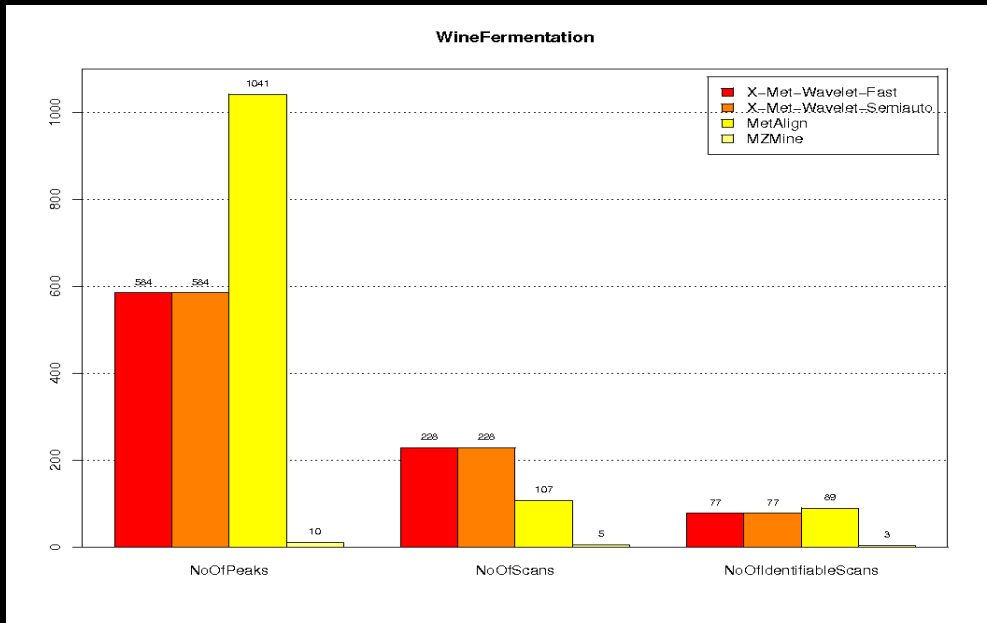


In: Castro, CC, Silva-Ferreira, AC., Teixeira, JA, Martins, RC. 2009. X-Metabolomics: A High-throughput GC-MS metabolomics pipeline for *Saccharomyces cerevisiae*. BMC Bioinformatics, Submitted.



5. Benchmarks

Wine Fermentation

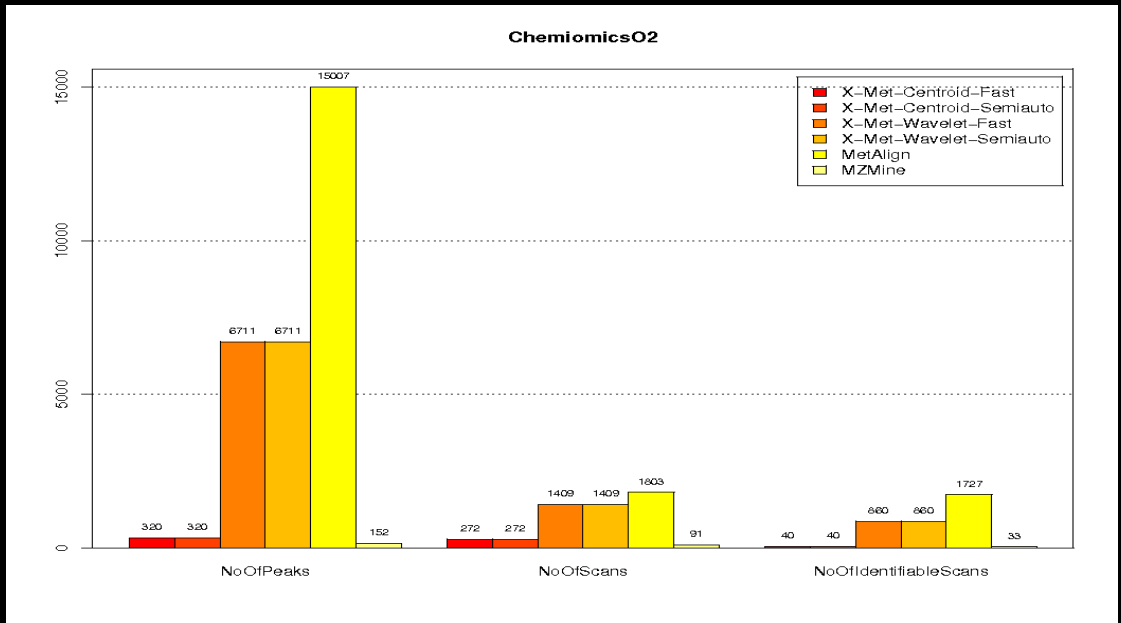


In: Castro, CC, Silva-Ferreira, AC, Teixeira, JA, Martins, RC. 2009. X-Metabolomics: A High-throughput GC-MS metabolomics pipeline for *Saccharomyces cerevisiae*. BMC Bioinformatics, Submitted.



5. Benchmarks

Madeira Wine GC-MS Data



In: Castro, CC, Silva-Ferreira, AC, Teixeira, JA, Martins, RC. 2009. X-Metabolomics: A High-throughput GC-MS metabolomics pipeline for *Saccharomyces cerevisiae*. BMC Bioinformatics, Submitted.



METABOLOMICS
High-Throughput Mass Spectroscopy

5. Benchmarks

Conclusions:

X-Metabolomics and **MetAlign** were developed for Metabolomics and Metabonomics research – **Out-perform all other software**, which were mainly **developed for Proteomics**

Performance of **X-Metabolomics** is in most cases similar to **MetAlign** with well defined peaks

X-Metabolomics out-performs MetAlign when chromatograms exhibit ion-trap artifacts (due to X-metabolomics filters)

Further developments in «In-Silico» Chromatography are only possible by the development of new feature extraction methods!



METABOLOMICS
High-Throughput Mass Spectroscopy

6. Software Demonstration

Minimum Media Fermentation Time Course Analysis

In: Silva-Ferreira, AC, Gunning, C., Castro, CC, Teixeira, JA,, Martins, RC. 2009. A non-target approach for time-course *Saccharomyces cerevisiae* oxidative response by GC-MS and Cyclic Voltammetry. Metabolomics, Submitted.