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Niche partitioning among and within sympatric tropical seabirds revealed by stable isotope analysis

H S. Young Stanford University

D J. McCauley Stanford University

R Dirzo Stanford University

R D. Dunbar San Jose State University

Scott A. Shaffer San Jose State University, scott.shaffer@sjsu.edu

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1	STABLE ISOTOPES SUGGEST NICHE PARTITIONING AMONG
2	SYMPATRIC TROPICAL SEABIRDS
3	
4	Hillary S. Young ¹ , Douglas J. McCauley ¹ ,
5	Rodolfo Dirzo ¹ , Rob D. Dunbar ¹ , Scott A. Shaffer ²
6	
7	¹ Department of Biology, Stanford University, 371 Serra Mall, Stanford CA 94305
8	² Department of Biological Sciences, San José State University, San José, CA 95192
9	
10	
11	*Corresponding author: Email: hsyoung@stanford.edu, phone: 619-889-7520, fax: 650-
12	723-6132
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16	$\delta^{13}C, \delta^{15}N$
17	
18	

19 Abstract

20 Despite the low productivity and ephemeral and patchy nature of resources in tropical 21 waters, stable isotopic data from this study suggests that substantial resource partitioning occurs among tropical seabird communities. In this study we compared δ^{13} C and δ^{15} N 22 23 levels in feathers across eight sympatric tropical seabird species; for a subset of these 24 species we also compared isotopic levels in blood, and examined variation across years and sexes. We found that while there is low total variation in both $\delta^{13}C$ and $\delta^{15}N$ across 25 26 the eight seabird species examined, all species occupied a distinct isotopic niche when 27 both breeding and non-breeding periods were evaluated. There was only slight variation 28 in the pattern of resource partitioning between breeding and non-breeding periods. 29 Notably, there was a strong correlation between body mass, wing span, and wing loading ratios on foraging area, evaluated by δ^{13} C levels, which is also coincident with estimates 30 31 of field metabolic rate. Isotopic separation by age and year, within species was also 32 observed; however separation by sex appeared to be relatively uncommon even in 33 sexually dimorphic species. As a group, seabirds were isotopically distinct both from 34 their prey and from other marine predators. Overall, the results are generally consistent 35 with what is known about the at sea distribution and diet of these seabirds, and with 36 patters of stable isotope partitioning among these species in other locations. Still, several results, including low δ^{13} C of black noddies (*Anous minutus*), high δ^{15} N of white terns 37 (*Gygis alba*), and strong correlation of δ^{13} C to body size and metabolic rate merit further 38 39 examination. More research on isotopic cartography of tropical oceans, species specific 40 fractionation rates, and stable isotopes of prey are needed to evaluate the usefulness of 41 stable isotopes in identifying resource partitioning in tropical marine environments

43 **INTRODUCTION**

44 Open oceans in tropical environments generally have low productivity, patchy and 45 unpredictable distribution of prey, and low structural complexity (Ballance et al 1997, Longhurst and Pauly 1987, Weimerskirch 2007). High level marine predators thus face 46 47 many foraging challenges in locating prey and are generally physiologically constrained 48 for energetically efficient travel and foraging behavior (Weimerskirch et al 2004, 49 Bertrand et al 2002). For tropical seabirds this has generally limited them to foraging 50 within the first several meters of the sea surface, and they are often reliant on subsurface 51 predators to drive food to the surface, further increasing patchy nature of food resources 52 (Ballance et al 1997). Yet, despite these strong constraints, diverse predator, and 53 particularly seabird communities, occur; this leads to questions about the degree, nature, 54 and mechanisms of resource partitioning in these extremely homogenous and resource 55 poor environments. 56 Resource partitioning in tropical oceanic environments has been well documented 57 for temperate and polar seabird species (Ainley et al 1992, 1994), but remains an area of 58 much inquiry for tropical seabird species particularly in open ocean environments 59 (Harrison and Seki et al 1987, Catry et al 2009). Tropical seabird diets are more diverse 60 than their temperate and polar counterparts, and there can be a high degree of overlap in 61 diets and foraging areas (Ainley and Boekelheide 1990, Ballance et al 1997, Catry et al 62 2009). Thus, these observations have led to questions about the degree to which tropical 63 seabird species are able to partition resources. 64 There have been multiple studies of diets of seabird communities in tropical

65 environments, but these have generally been constrained to the breeding season (Ashmole

66 and Ashmole 1967, Diamond 1983, Harrison et al 1983, Catry et al 2009). The only study 67 to comprehensively explore the diet of non-breeding tropical seabird communities found higher degrees of resource portioning than found in a similar study of polar seabirds 68 69 (Spear et al 2007). They found partitioning by species, sex, age, foraging strategy, and 70 body size. However, at-sea surveys of seabird diets have some inherent limitations for 71 answering questions about resource partitioning. Since seabirds are typically sampled 72 lethally, each animal contributes only a single data point in space and time, and it is thus 73 not possible to examine changes in individual foraging behavior across space and time. 74 Also, since reproductive status cannot be confirmed, it is not possible to link foraging 75 behavior to reproductive status, although it is clear that reproductive status influences 76 foraging ability. Spatially extensive survey efforts may also cross community boundaries 77 for both seabirds and prey species, resulting in comparisons of foraging ecology of 78 species that have limited co-occurrence. Collections across longer time periods may 79 compare foraging across heterogeneous temporal periods. 80 Data on resource partitioning of tropical seabirds is also available from direct 81 comparisons of foraging behavior across species via various methods of electronic 82 tracking (Burger and Shaffer 2008, Weimerskirch et al 2005, 2006a, 2006b, Young et al. 83 2010). However, cost, size, and logistical constraints associated with these methods have 84 been generally limited to comparisons involving two or three species, with emphasis on 85 larger species during breeding periods (Ropert-Coudert and Wilson 2005). 86 The usefulness of stable isotope analysis (SIA) as a tool for understanding 87 resource partitioning is providing a low impact way to examine resource partitioning, 88 both spatially and by trophic level. Stable isotopes of nitrogen and carbon in seabird

89 tissue reflect those values seen in their prey and give insight into origin and type of prey 90 consumed. Stable carbon isotopes of seabirds and seabird prey show evidence of an 91 increasing enrichment in inshore as compared to offshore feeding animals in both tropical 92 and temperate areas (Hobson et al 2004, Cherel et al 2008). Stable nitrogen isotopes 93 increase in a predictable matter with each trophic exchange, and thus indicate trophic 94 position of a consumer (Vanderklift and Ponsard 2003). Crucial to seabird studies, since 95 different tissues integrate these stable isotopes over different periods of time, a single bird 96 can provide integrated information on diet over a period of time ranging from days to 97 even years, depending on the tissue or tissues analyzed (Dalerum and Angerbjörn 2005). 98 Multiple studies have now used these SIA to understand resource partitioning 99 among seabird communities (REFS). SIA have now yielded important insights into subtle 100 changes in foraging ecology based on reproductive stage (e.g. Awkerman et al 2007), age 101 (e.g. Forero et al 2002), colony location (e.g. Jaquemet et al 2008), sex (e.g. Bearhop et al 102 2006), migratory patterns (e.g. Cherel et al 2000), as well as other factors (e.g. Cherel et 103 al 2005, Phillips et al 2009). These studies have been consistent with data observed via 104 direct tracking both in temperate and tropical systems (Phillips et al 2009, Weimerskirch 105 et al 2009b, Young et al 2010). Since SIA allows for fairly robust, low impact method to 106 resolve spatial and trophic separation among species, it is a promising tool to examine 107 resource partitioning among seabirds in tropical environments, particularly for those 108 species (e.g. terns, noddies, and small petrels) that are too small for current tracking 109 methods. There have been a few studies that used SIA to examine resource partitioning 110 on a community scale in the tropics (Cherel et al 2008, Catry et al 2008, Kojadinovic 111 2008). Those studies, all conducted in Southern Indian Ocean, reached very different

112 conclusions, both in the degree of resource partitioning and the usefulness of isotopes as 113 a tool for identifying foraging patterns in the tropics. Our study site, Palmyra Atoll 114 (equatorial Pacific), is quite different from the other studies in that Palmyra is far (1000's 115 km) from any continental shelf or coastal habitat, and has lower heterogeneity in oceanic 116 conditions (productivity, SST, bathymetry) and thus may more typify open ocean 117 conditions experienced by many tropical seabirds (Weimerskirch et al 2005, Catry et al 118 2008). 119 The aims of the present study are 1) determine if there is significant isotopic 120 partitioning in tropical seabirds living in an open ocean environment; 2) compare SIA 121 results to data from conventional stomach content analyses, at-sea surveys, and known 122 natural history to evaluate the usefulness of SIA as a tool for examining resource 123 partitioning in among sympatric tropical seabirds; 3) compare patterns of isotopic 124 partitioning in blood and feathers to evaluate differences in partitioning in breeding (from 125 blood) and non-breeding (from feathers)periods; and 4) compare isotopic levels among 126 different age classes and sexes of the same species and over multiple consecutive years. 127 Specifically, we examined stable isotopes of carbon and nitrogen from nine species of 128 sympatric seabirds at Palmyra Atoll. This included multiple congeneric species, and 129 species with similar foraging strategies, where fine-scale niche partitioning might be

- 130 particularly important for mitigating competitive interactions.
- 131

132 <u>Methods</u>

- 133 Our research was conducted at Palmyra Atoll National Wildlife Refuge (5.867° N,
- 134 162.067° W). Palmyra Atoll is a low-lying tropical atoll located in the Line Island chain

135	of the central Pacific Ocean. It is situated at the boundary of the eastern cool tongue and
136	western warm pools of the Pacific, on the boundary of the intertropical convergence zone
137	(Longhurst and Pauly 1987). Palmyra is composed of a ring of calcium carbonate
138	derived islets encircling three saltwater lagoons. The land of Palmyra is predominantly
139	forested; P. grandis and T. argentea forest provide extensive nesting habitat for tree
140	nesting birds and occasional herbaceous and bare patches (including two maintained
141	areas) serve as nesting areas for ground nesting birds (Young et al 2010). The
142	surrounding waters are uniformly low in productivity (mean of 0.14 mg chlorophyll a
143	/m ³), warm (mean sea surface temperature of 21.3 $^{\circ}$ C), and deep (except in immediate
144	vicinity of the Line Island chain, surrounding waters are > 1000 m). The seabird
145	community at Palmyra consists of 10 breeding species from 1) Order Charadriiformes -
146	sooty terns Sterna fuscata (125,000-220,000 pair), white terns Gygis alba (~200 pair),
147	brown noddies Anous stolidus (~500 pair), black noddies A. minutus (~1000 pair), and 2)
148	Order Pelecaniformes - greater frigatebird Fregata minor (~250 pair), red-footed boobies
149	Sula sula (~2500 pairs), brown booby S. leucogaster (~400 pairs), masked booby S.
150	dactylatra (~35 pairs), and red-tailed tropicbirds Phaethon rubricauda (~150 pairs), and
151	white-tailed tropicbirds P. lepturus (~10 pairs) (Fefer 1987, Young et al unpublished
152	data). At Palmyra these birds breed asynchronously throughout the year.
153	
154	Sample collection
155	We collected feather samples from 9 of the 10 species that breed at the atoll (all except

- 156 white-tailed tropicbirds). Samples were collected from breeding adults (either incubating
- 157 or chick rearing), with the exception of samples from white terns, where reproductive

158	status was unknown. All samples were collected in July 2009, except for sooty terns,
159	which did not breed in summer 2009. Samples from sooty terns were thus collected in
160	July 2008. For red-footed boobies, additional samples were collected from breeding
161	adults in two previous years (July 2007 and July 2008) for interannual comparisons. For
162	five species we also analyzed feathers collected from chicks in 2009. Feather tissue from
163	brown noddies and wedge-tailed shearwaters (not a breeding species at Palmyra) were
164	not included in statistical analyses due to small sample sizes. Birds were sexed using
165	voice, plumage, and/or molecular sexing methods (Young et al 2010). We used
166	unabraided underwing contour feathers (Jaeger et al 2009). Body masses of remaining
167	species, plus wingspan and wingloading for all species were estimated from Hertel and
168	Ballance (1999) and Spear and Ainley (1999). Field metabolic rate (kJ/day) was
169	estimated from allometric equations for Pelicaniformes, Charadriformes, and
170	Procellariformes (Shaffer in review).
171	For the three boobies, the frigatebird, and the sooty tern, blood was also collected
172	from a brachial vessel from a subset of the individuals that were sampled for feathers.
173	Muscle tissue (breast) was collected from red-footed boobies, masked boobies, black
174	noddies, and sooty terns (chicks only) found dead in the breeding colony.
175	Diet samples were collected when spontaneously regurgitated. The best-
176	preserved specimens of the common Exocetidae (flying fish) and Ommastrephidae
177	(flying squids) from these diet samples were used for the isotopic analyses. Muscle
178	tissue of Clupeiformes (herring and anchovy), small baitfish common in the diets of many
179	of these species (Catry et al 2009) was directly collected from below a seabird feeding
180	aggregation immediately off the atoll. Muscle tissue from pelagic fish predators (wahoo,

Hillary 2/13/10 9:17 PM

Comment [1]: Scott – I don't mention the black noddy wingspan measurements by Melinda. Do you think I should?

- 181 Acanthocybium solandri; yellowfin tuna, Thunnus albacores) was collected from animals
- 182 captured from within 3km of the reef immediately surrounding the atoll. Sthenoteuthis
- 183 *spp.* (jumbo flying squid) was captured approximately 700 km from the atoll.
- 184

185 Sample preparation and isotopic analyses

186 Feathers were washed in DI water, dried at 60°C for storage, and subsequently cut into

187 fine pieces for analysis. Blood, diet, and muscle tissue samples were all preserved frozen

188 at -80°C. They were then freeze dried and ground to a fine powder. We did not extract

189 lipid from any tissues as C:N ratios were always less than 4.0, and usually less than 3.5,

190 suggesting lipid levels were low across all samples (Post et al 2007).

191 Stable isotopic ratios of C and N were analyzed at the Stanford Stable Isotope

192 Biogeochemistry Laboratory (SIBL) using a Thermo Finnegan Delta-Plus XP IRMS.

193 Replicate laboratory standards of graphite (USGS 24), ammonium sulfate (IAEA N1),

and acetelanalide internal to each run show analytical error of less than 0.2‰ for both C

195 and N.

196

197 Isotopic interpretations

198 The interpretation of carbon isotope values presented in this paper, are based on the

199 assumption that the established inshore/offshore gradient of carbon-13 in seabird diets

200 (Cherel and Hobson 2007, Graham et al 2009) is likely the primary driver for changes in

201 stable carbon isotopes observed in this study. Benthic to pelagic gradients in carbon

202 isotopes may partially cause this pattern; however the steep drop off in waters

203 immediately surrounding Palmyra, and the fact that all seabirds in this study consume

204	prey found on or near the ocean surface, make it unlikely to be the primary driver of
205	differences in carbon isotopes. Changes in carbon isotopes due to trophic level
206	differentiation are generally small in seabirds, and given the limited range of $\delta^{15}N$
207	measured and the lack of correlation between $\delta^{15}N$ and $\delta^{13}C$ observed in this study, it is
208	not likely to be an important explanatory factor here. Variation in $\delta^{15}N$ within a tissue
209	type is likely primarily due to sequential enrichment in consumer tissues, such that $\delta^{15}N$
210	is interpreted as a measurement of trophic position. However, since there are established
211	δ^{15} N gradients on large scales across the Pacific Ocean (Graham et al 2009) we also
212	consider the possibility that $\delta^{15}N$ changes could be caused by these spatial gradients.
213	We further assume that there is no size or age specific fractionation of either
214	carbon (Δ C) or nitrogen (Δ N) isotopes within seabirds (Cherel et al 2005). We do not
215	directly compare $\delta^{15}N$ or $\delta^{13}C$ across tissue types, given different istopic signatures and
216	fractionation rates of these tissues (Cherel et al 2005). The period of isotopic integration
217	in blood is assumed to be days to weeks, such that blood taken from a breeding bird is
218	assumed to give diet information on the breeding period (Hobson and Clark 1992a,
219	1992b). Since feathers are usually molted after reproduction, and are inert thereafter,
220	feather samples were assumed to represent the composition of the diet during the
221	nonbreeding period (Bearhop et al 2002). Muscle tissue likely integrates over
222	intermediate time periods (4-6 weeks) (Hobson and Clark 1992a, 1992b).
223	
224	Statistical analyses

- 225 To examine differences in resource partitioning among species, we used multivariate
- analysis of variance (MANOVA), with subsequent univariate ANOVA tests of δ^{13} C and

- 227 δ^{15} N difference, with post-hoc Tukey HSD analyses. Statistical analyses were performed
- 228 in JMP 7 (SAS Institute, Cary, NC, USA). When necessary to meet assumptions of
- 229 normality, data was transformed using Box-Cox transformation. All figures/tables depict
- 230 untransformed data. All mean values are shown with ± 1 SD, also untransformed.
- 231

232 **Results**

233 Species comparisons

- 234 The nine species of adult seabirds showed significant overall isotopic segregation in
- feather samples (MANOVA, Wilks' lambda, $F_{16,288}$ = 12.49, p < 0.0001; Fig 1A).
- 236 Univariate tests of feathers also show significant difference both by δ^{13} C (ANOVA, $F_{7,145}$
- 237 = 30.41, p < 0.0001) and by δ^{15} N (ANOVA, $F_{7,145}$ = 5.20, p < 0.0001). Results from post
- 238 hoc Tukey pairwise comparisons show significant differences in δ^{13} C among most
- 239 species, although there is some overlap (Table 1). For $\delta^{15}N$, the three booby species
- 240 were all significantly different from the white tern and the great frigatebird; there are no
- 241 other significant differences (Table 1).

242 The five species sampled for blood also showed significant overall isotope

- 243 segregation (MANOVA, Wilks' lambda, $F_{8,90} = 36.63$, p < 0.0001). Univariate tests
- showed significant differences both by δ^{13} C (ANOVA, $F_{4,46} = 129.10$, p < 0.0001) and by

245 δ^{15} N (ANOVA, $F_{4,46}$ = 30.41, p < 0.0001). In post hoc analyses, all seabird species

- 246 partitioned separately for δ^{13} C, except for greater frigatebirds, which were
- 247 indistinguishable from either masked or red-footed boobies. For $\delta^{15}N$, greater
- 248 frigatebirds and brown boobies were distinct from the other three species (Fig 1B).

All seabirds differed from one another in at least one of the two stable isotopes measured in blood or feathers. The total variation in values among species was between 1.2‰ (in feathers and 1.8‰ (in blood) in δ^{13} C and between 2.5‰ (in feathers) and 2.3‰ (in blood) in δ^{15} N. The pattern of trophic partitioning among species changed somewhat between blood and feathers, with great frigatebirds having a relatively lower δ^{13} C, in feathers compared to blood, and brown boobies had a relatively higher δ^{13} C in feathers compared to blood.

256 Comparison of δ^{13} C by mean body mass per species yielded a significant positive 257 relationship (R² = 0.67, *p* < 0.01), with more enriched δ^{13} C for larger birds (Fig 2A). The 258 relationship improved significantly when δ^{13} C was compared to wing loading (R² = 0.82, 259 *p* < 0.01), where birds with lower wing loading were more enriched with δ^{13} C (Fig 2B). 260 There was also a strong positive relationship between estimated field metabolic rate and 261 δ^{13} C (R² = 0.74, *p* < 0.01). There were no significant relationships between any of the 262 above variables and δ^{15} N values.

Although there were few muscle tissue samples per species, there were significant differences in isotope levels between species in this tissue. Black noddies had lower $\delta^{15}N$ in muscles than sooty terns (F_{3,12} = 4.64, P = 0.02). For $\delta^{13}C$ black noddies were significantly more depleted than either red-footed or masked boobies.

267

268 Effects of year, age, and sex within species

269 There was a slight but significant interannual difference in δ^{13} C for breeding adult red-

footed boobies (MANOVA, Wilks' Lambda, $F_{4,126} = 4.10$, p < 0.01), where boobies in

271 2007 had slightly higher δ^{13} C values than birds in either 2008 or 2009 (ANOVA, $F_{2,50}$ =

272	3.61, p = 0.03; Fig 3). No such trend was detected in δ^{15} N. There were slight differences
273	in feather isotope levels across age classes for red-footed boobies within 2009
274	(MANOVA, Wilks' lambda, $F_{4,44} = 3.21$, p = 0.02), where chicks had slightly higher
275	δ^{15} N values ($F_{2,23} = 5.49$, p = 0.01) than either adult or juvenile birds. No such trend was
276	apparent in δ^{13} C.
277	Of the five species for which adult and chick feathers were compared, four
278	showed significantly higher δ^{15} N values in chicks than in adults (black noddy, $t = 2.47, p$
279	= 0.02, df = 24; red-footed booby t = 2.92, p = 0.01, df = 18; red-tailed tropic bird, t =
280	5.63, $p < 0.0001$, df = 39; brown booby $t = 2.71$, $p = 0.01$, df = 29) and one, great
281	frigatebirds showed marginally significant increases ($t = 1.82$, $p = 0.08$, df = 24; Fig 4).
282	Only one species, red-tailed tropic birds showed significant differences in $\delta^{13}C$ by age,
283	with adult birds having lower δ^{13} C than chicks ($t = 5.63, p < 0.0001, df = 39$).
284	Comparisons by sex (among adult birds from the same year) were conducted for
285	red-footed, masked, and brown boobies, as well as greater frigatebirds. The only species
286	that showed significant differences by sex were brown boobies where males had lower
287	δ^{13} C (t = 4.23, p < 0.001, df = 14) and lower δ^{15} N (t = 2.40, p = 0.03, df = 14) than
288	females.

290 *Comparisons to prey and other marine predators*

291 Comparisons among seabirds, other marine predators, and prey only included the five292 species for which blood data was available. As a group, seabirds were segregated in

- trophic space both from their prey and from other large pelagic predators (MANOVA,
- Wilks' Lambda, $F_{6,298} = 23.26$, p < 0.0001; Fig 5). Seabirds and predatory fish (wahoo,

295 yellowfin tuna) had higher δ^{15} N than prey (flying fish, squid, and anchovies) or than large 296 predatory invertebrates (jumbo flying squid) ($F_{6,149}$ =15.87, p < 0.0001). The patterns of 297 δ^{13} C was different, with the large predatory fish having higher δ^{13} C than seabirds, seabird 298 prey, or predatory invertebrates ($F_{6,149}$ = 64.93, p < 0.0001).

299 Post-hoc analysis on a species by species comparison, showed less clear patterns 300 of partitioning among seabirds and their prey. With regard to prey, diet items from seabird stomachs (flying fish, squid) had large SD (\pm XX), particularly in δ^{15} N, and were 301 significantly elevated in δ^{15} N over anchovy. Squid were indistinguishable in δ^{15} N from 302 any of the seabird species, although δ^{13} C distinguished them from all seabird species 303 304 except red-footed boobies and greater frigatebirds. Flying-fish were indistinguishable in 305 either parameter from red-footed and masked boobies. 306 Seabirds had relatively little overlap with other pelagic predators. Only the brown 307 boobies and yellowfin tuna, and red-footed boobies and jumbo flying squid were 308 indistinguishable from each other in both δ^{13} C and δ^{15} N. 309

310 **DISCUSSION**

311 *Resource partitioning by species*

312 The eight sympatric seabird species studied each occupy a distinct ecological niche

313 across the breeding and non-breeding periods. The degree of partitioning observed in

314 non breeding period was greater than that detailed by at-sea surveys and diet analyses

315 (Surman and Wooler 2003, Spear et al 2007).

316 The patterns observed in δ^{13} C in non-breeding period were generally consistent

317 with data from tracking and at-sea surveys, where sooty terns and greater frigatebirds

318 being highly pelagic, white terns and red-footed boobies less pelagic, followed red-tailed 319 tropicbirds, brown and masked boobies, the least pelagic (Ballance et al 1997, Jaquemet 320 et al 2005). The one surprising result based on carbon isotope levels was the highly 321 pelagic signal of black noddies (i.e. values were more negative than that of any other 322 species). This species is generally considered to be an opportunistic nearshore feeder that 323 is often seen foraging near jacks and in lagoons (Ashmole 1968, Seki and Harrison 1989). 324 Yet values of carbon isotope levels in lagoons at Palmyra are particularly elevated, and 325 black noddy isotope levels do not resemble those of reef jacks at Palmyra (McCauley et al unpublished data). Given the small total range of δ^{13} C observed, controlled 326 327 measurements of species-specific fractionation rates would be helpful to confirm that 328 species-specific fractionation rates do not drive these patterns (Becker et al 2007). Likewise, while δ^{13} C maps available for the equatorial Pacific Ocean do not suggest high 329 330 variation in δ^{13} C around this region, that is based on limited sampling near Palmyra 331 (Graham et al 2007); better isotopic sampling of oceans in this region would help 332 interpret these results. Species level changes in δ^{13} C were highly correlated to body mass, wing loading 333 334 and metabolic rates; small species with high metabolic rates and species with low wing 335 loading exhibit a more pelagic signature than larger species and species with high wing 336 loading. Generally, birds with low body mass and high wing loading should have low 337 costs of flight, perhaps enabling a more pelagic lifestyle (Pennycuick 1989). However, 338 since these factors covary with metabolic rate, it is also possible that metabolic rate 339

drives this pattern.

340	Nitrogen isotopes from non-breeding periods showed the greater frigatebird to
341	have elevated $\delta^{15}N$ levels over many species including all the boobies (consistent with
342	Cherel et al. 2008). Although direct analysis of diets of non-breeding greater frigatebirds
343	is quite similar to red-footed boobies, these birds also often consume pulli of sooty terns
344	and noddies, potentially explaining this variation (Schreiber and Hensley 1986, Megyesi
345	and Griffin 1996, Spear et al 2007). Kleptoparasatism which seems to be of particular
346	importance in non-breeding birds could potentially explain these elevated $\delta^{15}N$ values, as
347	these partially digested food items may have elevated $\delta^{15}N$ levels (Gilardi 1994). As in
348	the Seychelles (Catry et al 2008), the white tern also showed high $\delta^{15}N$ levels. While
349	Catry et al suggested that this might be reason to discount results of $\delta^{15}N$, the consistency
350	across studies perhaps merits further consideration for biologically valid explanations. It
351	is possible that these high levels may be due to the large portion of its diet (>40%)
352	composed of small, predatory Scombridae (Euthynnus sp; Spear et al 2007). This
353	predatory species might well be higher in δ^{15} N than flying fish and squid dominating diet
354	of other species; direct measurements of $\delta^{15}N$ of these prey would be necessary to resolve
355	this. The three booby species, which feed primarily on flying fish and squid (Schreiber
356	and Hensley 1986, Spear et al 2007) show particularly low $\delta^{15}N$ values. Controlled
357	studies of species specific fractionation patterns would help understand if physiological
358	or ecological factors drive δ^{15} N patterns (Becker et al 2007)
359	Examination of blood samples, representing the breeding interval, also showed
360	distinct niches for each of the species examined. There were small changes in foraging

361 areas in non-breeding as opposed to breeding periods, with greater frigatebirds and brown

362 boobies looking comparatively less pelagic than during non-breeding interval. For the

363	two species for which tracking data is available at Palmyra (red-footed and masked
364	booby), the δ^{13} C results are high consistent with tracking results, even though the
365	differences in foraging distances was not extremely large (Young et al in press).
366	The relative positions of $\delta^{13}C$ and $\delta^{15}N$ in both breeding and non-breeding periods
367	observed at Palmyra were highly consistent with those observed in Europa Island and in
368	the Seychelles (Catry et al 2008, Cherel et al 2008) suggesting that niche partitioning is
369	consistent in very different parts of the species' ranges. The absolute values of $\delta^{13}C$ were
370	also consistent in both studies in nonbreeding interval, but slightly depleted at Palmyra
371	during breeding interval, perhaps due to more oceanic location of Palmyra itself which
372	might lead to lower $\delta^{13}C$ values (Graham et al 2009). In contrast, absolute values of $\delta^{15}N$
373	observed at Palmyra were greatly elevated (by about 2‰) over that observed at Europa in
374	both breeding and non-breeding periods. This suggests a higher baseline of nitrogen at
375	Palmyra than at Europa; similar results were seen in comparison of Seychelles to
376	Mozambique Channel (Jaquemet et al 2008). This is consistent with latitudinal variations
377	of δ^{15} N in isotopic cartography. This suggests that while it is possible to compare
378	relative $\delta^{15}N$ positions across studies, to compare absolute values of $\delta^{15}N$ strong good
379	knowledge of baseline δ^{15} N is needed (Graham et al 2009).
380	There was a relatively few samples of muscle tissue, but patterns were fairly
381	consistent with that seen in feathers and blood. Only the $\delta^{15}N$ of black noddies were
382	significantly different (lower in δ^{15} N).
383	

Hillary 2/15/10 12:30 PM **Comment [2]:** What about dropping muscle tissue altogether? It's not really adding much and it will cut length a little...

384 Partitioning within species

Comparison of adult feathers to chick feathers shows elevated levels of $\delta^{15}N$ in 385 386 chicks across all five species examined. This varies from the pattern seen for greater 387 frigatebirds in Europa Island, but is consistent with results seen for sooty terns at the 388 same site (Cherel 2008). This could reflect a shift to higher trophic level prey items 389 during the breeding period, selective feeding of food items to young, age specific 390 fractionation, or some effect of regurgitation of food. Other studies have shown 391 differential provisioning of chicks with higher quality food, or different trophic level food 392 sources (Hodum and Hobson 2000, Cherel 2008). While seabird studies have not 393 documented changes in ΔN by age, this has been seen in other taxa and could be a viable 394 explanation (Roth and Hobson 2000). Even without different ΔN , by feeding on partially digested food, they may be incorporating δ^{15} N from their parents' bodies, thus explaining 395 higher δ^{15} N values. Direct comparison of adult and chick blood and diets (not taken 396 here) would help resolve this. The lack of any shift in δ^{13} C from adults to chicks was 397 398 unexpected given that breeding places constraints on seabird foraging distances, but is 399 consistent with lack of change seen in Seychelles (Catry et al 2008).

400 We saw relatively little resource partitioning by sex. Of the four species, all 401 exhibiting reverse sexual dimorphism, for which sex differences in foraging were 402 examined, we saw differences only for one (brown booby). The larger females of brown 403 boobies showed higher δ^{15} N and δ^{13} C, indicating higher trophic level and less pelagic 404 food sources. This is consistent with other evidence of niche partitioning by sex in brown 405 boobies (Gilardi 1992, Weimerskirch et al 2009b). Likewise, the lack of resource 406 partitioning by sex in red-footed boobies, and masked boobies is also consistent other 407 studies (Weimerskirch et al 2009a, Young et al in press), although sexual differences

408 have been observed in the red-footed booby in other locations (Weimerskirch et al

409 2006b).

410 In the one species for which we compared data across multiple years, we saw 411 small but significant differences in δ^{13} C for one of the three years. This may point to 412 small variability in food sources across time even in tropical resources. While it is not 413 always possible to gather simultaneously, this suggests cautions in interpreting data from 414 seabirds gathered different years in isotopic analyses.

415

416 *Comparisons with prey and predators*

417 We saw clear distinctions among seabirds, their prey, and other marine predators. 418 The relatively little overlap between pelagic predators and seabirds on a species by 419 species basis was unexpected, given that many of these seabirds feed so heavily in flocks 420 over schools of predators. The primary differences in seabirds and other predatory fish 421 was δ^{13} C and may reflect integration of benthic food sources into fish diets. However, it 422 could also represent other differences in diet, as other studies of seabird and predator 423 diets have shown substantial variation, even though the forage together (Catry et al 424 2009). The difference in δ^{15} N between *Sthenoteuthis spp.* and seabirds may reflect lower 425 trophic diet of Sthenoteuthis spp (Shchetinnikov 1992). Seabirds were also not isotopically distinct from individual prey types in δ^{15} N. 426 which was unexpected given known fractionation rates for seabirds between 3 and 5 ‰ 427 428 (REFS). This may be due to use of muscle/mantle from diet samples rather than whole animals; whole fish have been shown to have lower δ^{15} N levels (REFS). This may also 429 430 be due to partial digestion of diet samples analyzed. Although diet samples selected

431 appeared to be in excellent condition, and interior muscle samples were, minimizing 432 potential contamination with seabird digestive enzymes, SD was very high in these samples and overall δ^{15} N levels were high. Muscle from Clupeiformes from beneath bird 433 foraging flocks did not show this elevated δ^{15} N or the high SD. We suggest care should 434 435 be used in determining diet items based on items from prey items gathered via stomach 436 contents. Analysis of whole additional diet items gathered directly, and sorted to lower 437 taxanomic levels would be a good resource for future use of isotopes in diet analyses of 438 tropical seabirds. This would also help resolve lingering questions about the merit of δ^{15} N in assessing food sources of tropical seabirds (Catry et al 2008) 439

440

441

Stable isotopes as a tool for evaluating resource partitioning in tropical seabirds

442 There has been some discussion about the merit of using stable carbon and 443 nitrogen in examining niche partitioning among tropical seabirds in open ocean sites. In thus study, we find high consistency in species specific patterns of δ^{15} N and δ^{13} C across 444 445 tissue types in this study, and between this study and other studies despite widely 446 different habitats (Catry et al 2008, Cherel et al 2008). For the two species for which 447 tracking data is available at this site, stable isotope levels are consistent (both within and 448 across species) with tracking data and appear to be able to detect small scale changes in 449 foraging (Young et al *in press*). Alignment of tracking, at-sea surveys, and isotope data 450 is also seen from various work in the Mozambique Channel (Jaquemet et al 2005, Cherel 451 et al 2008). Also, for the great majority of analyses, patterns of partitioning are in 452 keeping with the data available from at-sea surveys and stomach content analyses. All of these factors appear to provide evidence for reliability of isotopes for identifying foragingpatterns of tropical species.

However the anomalously low δ^{13} C of black noddies and the strong correlation of 455 δ^{13} C to field metabolic rate, does raise possibility that species specific fractionation rates 456 457 might be an alternative explanation for significant variation observed in isotopes across 458 species (repeated across both space and time). Other potential explanations (i.e. 459 nutritional stress causing differential fractionation, mixing of benthic δ^{13} C signals, local variation in δ^{13} C and δ^{15} N) would be unlikely to be consistent across spatially and 460 461 temporally distinct studies. However analyses of prey item isotope levels, controlled 462 laboratory experiments of species specific fractionation rates, and better isotopic maps for 463 the region would help confirm the interpretations presented here. 464 465 **Conclusions** 466 In general this study supports the idea that there is high niche partitioning among 467 tropical seabirds even in ocean environments. Resource partitioning is not complete 468 unless considered across both breeding and non-breeding periods. Partitioning appears to 469 occur within species, as well as among species – with different isotopic signatures 470 observed by sex, age, and year. Sexual partitioning of resources did not appear to be 471 common even among species that had relatively high sexual dimorphism, consistent with 472 tracking data from this site (Young et al *in press*). Total trophic range of seabird diet was 473 small, and both seabirds and their diet showed substantially higher levels of δ^{15} N in this 474 study than in studies from other sites despite apparently similar diets, pointing to

475 potential variability in space in this value. There was a strong correlation between body

476 size, wing loading, and metabolic rate on δ^{13} C; there are multiple possible explanations

477 for this pattern.

478 While variability in both isotopes was smaller in this study than in comparable 479 polar or temperate studies, the results suggest that niche partitioning is at least as 480 prevalent in this system. This is despite more limited foraging techniques of tropical 481 seabirds studied here, and despite patchier resources in the tropical ocean. The apparent 482 ability to detect small differences in foraging changes via isotopes in tropical 483 environments, continues to further suggestions of others that this minimally invasive tool 484 can offer powerful insight to niche partitioning in tropical seabirds. 485 486 **ACKNOWLEDGEMENTS** 487 We thank the National Science Foundation, the National Geographic Society, the 488 Stanford Vice Provost Office for Undergraduate Education summer field studies grants, 489 the Stanford Gabilan Graduate Fellowship, and the Woods Environmental Institute for 490 financial support. For logistical and material support we thank US Fish and Wildlife 491 Service (Palmyra Atoll National Wildlife Refuge), The Nature Conservancy, and the

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495 Anderegg, C. Depkin, A. Briggs, M. deGraff, P. de Salles, T. Jen, E. Hoffman, C.

496 Burniske, N. Wenner, C. Hanson, L. Palumbi, and T. Robbins.

497

Hillary 2/15/10 12:25 PM **Comment [3]:** Could cut this entire paragraph, basically just summarizes previous statements...

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Hillary 1/29/10 5:50 PM Comment [4]: Weimerskirch et al 2007 reference missing.

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Scott Shaffer 2/10/10 10:21 PM Deleted: -

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FEATHERS: SEABIRDS	n	$\delta^{15}N$	$\delta^{13}C$	C:N
Black noddy	18	14.79 1.90 (AB)	-16.87 0.32 (A)	3.34 0.07
Brown noddy †	3	16.35 0.31	-16.12 0.25	3.37 0.01
Sooty tern	21	14.7 2.80 (AB)	-16.30 ± 0.28 (B)	3.23 0.03
White tern	20	16.20 ± 1.69 (A)	-15.91 ± 0.29 (C)	3.16 0.04
Brown booby	21	14.08 1.39 (B)	-15.43 0.31 (DE)	3.18 0.03
Masked booby	15	14.17 0.98 (B)	-15.22 0.24 (E)	3.18 0.03
Red-footed booby	12	13.85 1.25 (B)	-15.73 0.34 (CD)	3.22 0.03
Great frigatebird	19	16.03 ± 1.09 (A)	-16.26 0.20 (B)	3.27 0.03
Red-tailed tropicbird	26	15.35 ± 1.70 (AB)	-15.61 ± 0.27 (D)	3.14 0.03
Wedge-tailed shearwater +	1	9.31	-16.62	3.34
BLOOD: SEABIRDS				
Sooty tern	17	14.29 ± 1.52 (A)	-18.02 ± 0.13 (A)	3.33 0.11
Brown booby	10	15.67 0.72 (B)	-17.20 0.12 (B)	3.30 0.03
Masked booby	9	14.21 ± 0.61 (A)	-17.15 ± 0.18 (C)	3.40 0.06
Red-footed booby	11	13.79 ± 0.44 (A)	-17.42 ± 0.09 (D)	3.31 0.04
Great frigatebird	5	16.07 0.72 (B)	-17.20 0.12 (CD)	3.41 0.03
MUSCLE: SEABIRDS				
Black noddy	3	10.89 ± 0.17 (A)	-18.34 0.02 (A)	3.79 0.04
Masked booby	3	14.09 ± 0.01 (A)	-16.88 0.45 (B)	3.34 0.12
Red-footed booby	7	13.46 ± 1.44 (A)	-17.48 0.37 (AB)	3.89 0.07
Sooty tern (chick)	4	14.57 ± 4.05 (A)	-17.50 0.54 (AB)	3.47 ± 0.24
MUSCLE: PREDATORS & PREY	/			
Clupeiformes	15	11.00 0.46	-17.80 0.46	3.35 0.09
Exocetidae	26	13.06 2.39	-17.34 0.29	2.66 0.05
Ommastrephidae	12	14.64 1.28	-17.61 1.02	3.26 0.22
Acanthocybium solandri	15	14.53 3.15	-16.21 1.24	3.15 0.06
Thunnus albacores	25	14.63 0.96	-16.68 0.36	3.16 0.15
Sthenoteuthis spp.	11	12.82 1.02	-17.62 0.27	3.18 0.05

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667	Table 1: Stable isotopic ratios of carbon and nitrogen and mass ratio of C:N in feathers,
668	blood, and muscle tissue of seabirds, and muscles of other pelagic predators and prey
669	Letters following mean values denote significant differences among species in post-hoc
670	analyses, values (within tissue type and isotopic ratio) not connected with the same letter
671	are significantly different. Species marked with † are not considered in statistical
672	comparisons due to small sample size. Values are mean \pm SD
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Figure 1: Stable carbon and nitrogen isotope values (mean ± SD) from eight seabird
species from Palmyra Atoll. Panel A shows values from feathers; solid black symbols
denote those species for which isotopic values of blood are also presented panel B. BB =
brown booby, BN = black noddy, MB = masked booby, GF = great frigatebrid, RFB =

red-footed booby, RTT = red-tailed tropicbird, ST = sooty tern, WT = white tern.



- 684
- 685
- 686 Figure 2: Stable isotope values of carbon compared to (a) body mass (b) wing loading
- and (c) estimated field metabolic rate. Species codes are the same as in Figure 1, plus
- 688 BRN = Brown noddy; WSH = Wedge-tailed shearwater. Statistics here include all
- 689 species depicted.

Hillary 2/13/10 5:58 PM

Comment [5]: Probably can drop at least one of these as they all covary – but which?

Hillary 1/29/10 11:42 AM

Comment [6]: In text I report values w/o brownnoddy and wedge tailed shearwater. Should I clarify? Or remove? Keep consistent?



693 Figure 3: A comparison of stable carbon and nitrogen isotope values for feathers from

694 red-footed boobies across multiple years and age classes. The first letter indicates age

695 class (A= adult, I = immature, C = chick) and the number indicates the year samples were

⁶⁹⁶ collected (2007, 2008, or 2009).





699 Figure 4: Stable carbon and nitrogen isotope values of feathers compared across age 700 classes for multiple seabird species. Adults (A) are indicated by unfilled shapes and 701 chicks (C) are indicated by filled shapes. Different species are indicated by codes (as 702 same as in Figure 1) and shape.



707 Figure 5: Stable carbon and nitrogen isotope values of seabirds (filled circles), their prey

708 (open squares), and other pelagic predators around Palmyra Atoll (grey triangles). AS=

709 *Acanthocybium solandri* (wahoo); TA = *Thunnus albacores* (yellowfin tuna); E =

710 Exocetidae (flying fish, various); C = Clupeiformes (anchovy and herring); S =

711 Sthenoteuthis spp. (jumbo flying squid, various). Seabird codes are the same as in Fig 1.

712