

Thesis for doctoral degree (Ph.D.)  
2015

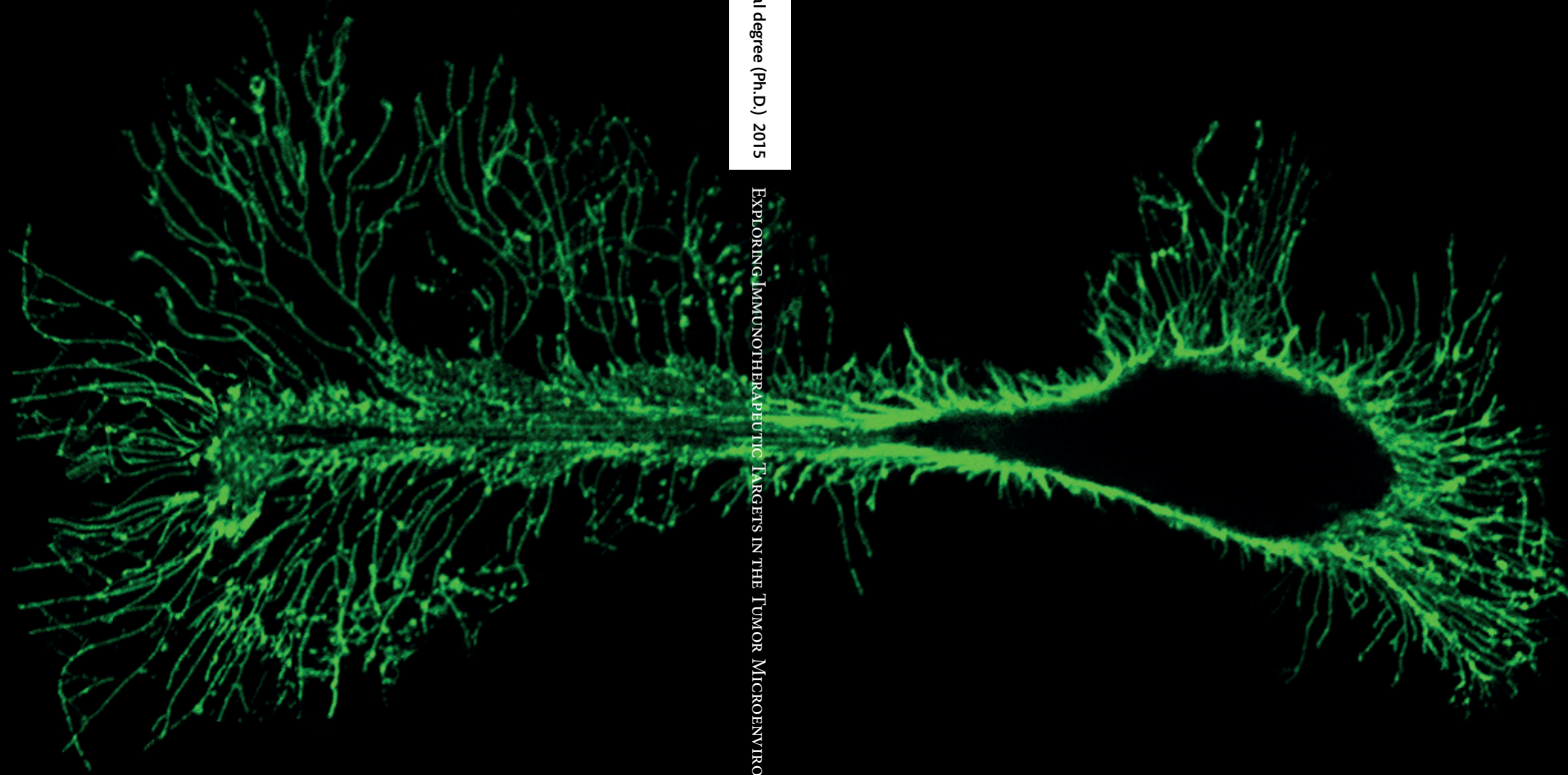
# EXPLORING IMMUNOTHERAPEUTIC TARGETS IN THE TUMOR MICROENVIRONMENT

Anna-Maria Georgoudaki

Thesis for doctoral degree (Ph.D.) 2015

EXPLORING IMMUNOTHERAPEUTIC TARGETS IN THE TUMOR MICROENVIRONMENT

Anna-Maria Georgoudaki



**Karolinska  
Institutet**



**Karolinska  
Institutet**

From The Department of Microbiology, Tumor & Cell biology  
Karolinska Institutet, Stockholm, Sweden

# **EXPLORING IMMUNOTHERAPEUTIC TARGETS IN THE TUMOR MICROENVIRONMENT**

Anna-Maria Georgoudaki



**Karolinska  
Institutet**

Stockholm 2015

Cover illustration: Fibroblast transfected to express scavenger receptor MARCO

All previously published papers were reproduced with permission from the publisher.

Published by Karolinska Institutet

Printed by AJ E-print AB

© Anna-Maria Georgoudaki, 2015

ISBN 978-91-7676-139-7



**Karolinska  
Institutet**

**Institutionen för Mikrobiologi, Tumör- och Cellbiologi**

## Exploring Immunotherapeutic Targets In The Tumor Microenvironment

**AKADEMISK AVHANDLING**

som för avläggande av medicine doktorexamen vid Karolinska Institutet offentlig  
försvaras i Welandersalen, B2:00 Karolinska Universitetssjukhuset, Solna

**Fredagen den 11 December 2015, kl. 09.30**

av

**Anna-Maria Georgoudaki**

*Huvudhandledare:*

Docent Mikael C.I. Karlsson  
Karolinska Institutet  
Institutionen för Mikrobiologi, Tumör- & Cell-  
biologi

*Bihandledare:*

Docent Benedict J. Chambers  
Karolinska Institutet  
Institutionen för Medicin, Huddinge  
Centrum för Infektionsmedicin

Professor Ola Winqvist  
Karolinska Institutet  
Institutionen för Medicin, Solna  
Enheten för Translationell Immunologi

*Fakultetsponent:*

Professor Siamon Gordon  
University of Oxford  
Sir William Dunn School of Pathology

*Betygsnämnd:*

Docent Anna Dimberg  
Uppsala University  
Institutionen för Immunologi, Genetik och  
Patologi  
Divisionen för Vaskulärbiologi

Professor Arne Östman  
Karolinska Institutet  
Institutionen för Onkologi-Patologi  
Cancer Center Karolinska

Jakob Michaëlsson, PhD  
Karolinska Institutet  
Institutionen för Medicin, Huddinge  
Centrum för Infektionsmedicin

**Stockholm 2015**





**Karolinska  
Institutet**

**Department of Microbiology, Tumor and Cell biology**

## **Exploring Immunotherapeutic Targets In The Tumor Microenvironment**

**THESIS FOR DOCTORAL DEGREE (Ph.D.)**

Publicly defended at Karolinska Institutet, Welander Hall, B2:00 Karolinska University  
Hospital, Solna

**Friday December 11th 2015, 09.30**

By

**Anna-Maria Georgoudaki**

*Principal Supervisor:*

Docent Mikael C.I. Karlsson  
Karolinska Institutet  
Department of Microbiology, Tumor & Cell  
biology

*Co-supervisor(s):*

Docent Benedict J. Chambers  
Karolinska Institutet  
Department of Medicine, Huddinge  
Center for Infectious Medicine

Professor Ola Winqvist  
Karolinska Institutet  
Department of Medicine, Solna  
Translational Immunology Unit

*Opponent:*

Professor Siamon Gordon  
University of Oxford  
Sir William Dunn School of Pathology

*Examination Board:*

Docent Anna Dimberg  
Uppsala University  
Department of Immunology, Genetics and  
Pathology  
Division of Vascular Biology

Professor Arne Östman  
Karolinska Institutet  
Department of Oncology-Pathology  
Cancer Center Karolinska

Assistant Professor Jakob Michaëlsson  
Karolinska Institutet  
Department of Medicine, Huddinge  
Center for Infectious Medicine

**Stockholm 2015**



*To my parents,  
for giving me my wings*

Σα βγεις στον πηγαμό για την Ιθάκη,  
να εύχεσαι να 'ναι μακρύς ο δρόμος,  
γεμάτος περιπέτειες, γεμάτος γνώσεις.  
Κ. Καβάφης

As you set out for Ithaka,  
hope the voyage is a long one,  
full of adventure, full of discovery.  
K. Kavafis





## ABSTRACT

The immune system has developed along with the evolution of increasingly complex cellular organisms to sustain homeostasis and protect from threats. Cancer, a detrimental side effect of increasing organismic complexity, typically sequesters the immune system and hijacks its functions for its own prosperity. Cancer immunotherapy aims to harness the intrinsic potential of the immune system for the therapeutic benefit of cancer patients.

The focus of this thesis is to identify and evaluate new immunotherapeutic targets in the tumor microenvironment, which can be modulated to restrict tumor growth and metastasis.

**Paper I** describes a novel mechanism of interaction between marginal zone macrophages (MZMs) and marginal zone B cells (MZBs) in the spleen, which can be modulated by antibodies (Abs) to scavenger receptor MARCO on MZMs. This study demonstrates that MARCO targeting diminishes antigen (Ag) uptake by MZBs, which results in reduced Ag deposition in the splenic follicles. As anti-MARCO Abs can also be found in systemic lupus erythematosus (SLE), this interaction may affect subsequent adaptive immune responses to both self- and foreign antigen.

**Paper II** identifies MARCO as a specific marker for a tumor-promoting macrophage subtype in the tumor microenvironment of mammary carcinoma, melanoma and colon carcinoma tumor models. Targeting MARCO on tumor-associated macrophages (TAMs) by Abs, inhibits tumor growth and metastasis and reprograms TAMs to an anti-tumor phenotype. Treatment with anti-MARCO mAbs in combination with checkpoint inhibitor, anti-CTLA-4 ab, may provide a promising approach for cancer immunotherapy with clinical relevance for human breast cancer and melanoma.

**Paper III** reveals the molecular mechanism through which tumor cells imitate immune cells, during epithelial-mesenchymal transition (EMT), prompting their targeted metastasis through the lymphatics. This study demonstrates that TGF- $\beta$ , a known inducer of EMT, regulates the chemotactic axis CCR7/CCL21, directing preferential lymphatic dissemination of breast cancer cells.

**Paper IV** evaluates dendritic cell-derived exosomes in a vaccination approach to re-activate adaptive anti-tumor responses. This study shows that dendritic cell-derived CD1d expressing exosomes loaded with  $\alpha$ -Galactosylceramide ( $\alpha$ -GalCer) can sensitize NKT cells and lead to subsequent activation of B cell and effector T cell responses, restricting tumor growth.

In summary, the work presented in this thesis describes novel targets in the tumor microenvironment that can be used in immunotherapeutic approaches to re-activate endogenous mechanisms of innate and adaptive immunity against cancer. Additionally, it gives new insight into gene regulatory pathways controlling metastatic tumor spread, as well as utilizes custom designed biological molecules in anti-tumor vaccination strategies.

Increasing our understanding of the intricate mechanisms regulating the immunosuppressive tumor microenvironment will reveal new knowledge and novel targets that can contribute to the design of prospective cancer immunotherapies.

## LIST OF SCIENTIFIC PAPERS

- I. Prokopec K\*, **Georgoudaki AM\***, Sohn S, Wermeling F, Grönlund H, Lindh E, Carroll MC, Karlsson MCI  
**Marginal zone macrophages regulate antigen transport by B cells to the follicle in the spleen via CD21**  
*Submitted*
- II. **Georgoudaki AM**, Prokopec K, Boura V, Hellqvist E, Sohn S, Östling J, Dahan R, Harris RA, Rantalainen M, Klevebring D, Sund M, Egyhazi Brage S, Fuxe J, Rolny C, Li F, Ravetch JV, Karlsson MCI  
**Reprogramming tumor associated macrophages by antibody targeting inhibits cancer progression and metastasis**  
*Submitted*
- III. Pang MF, **Georgoudaki AM**, Lambut L, Johansson J, Tabor V, Hagikura K, Jin Y, Jansson M, Alexander JS, Nelson CM, Jakobsson L, Betsholtz C, Sund M, Karlsson MCI, Fuxe J  
**TGF- $\beta$ 1-induced EMT promotes targeted migration of breast cancer cells through the lymphatic system by the activation of CCR7/CCL21-mediated chemotaxis**  
*Oncogene, 2015 May 11. doi: 10.1038/onc.2015.133.*
- IV. Gehrman U, Hiltbrüner S, **Georgoudaki AM**, Karlsson MCI, Näslund TI\*, Gabrielsson S\*  
**Synergistic induction of adaptive antitumor immunity by codelivery of antigen with  $\alpha$ -galactosylceramide on exosomes**  
*Cancer Research, 2013 Jul 1;73(13):3865-76*

\* denotes equal contribution

## PUBLICATIONS NOT INCLUDED IN THIS THESIS

**Georgoudaki AM**, Khodabandeh S, Puiac S, Persson CM, Larsson MK, Lind M, Hammarfjord O, Nabatti TH, Wallin RP, Yrlid U, Rhen M, Kumar V, Chambers BJ  
**CD244 is expressed on dendritic cells and regulates their functions**  
*Immunology & Cell Biology, 2015 Jul;93(6):581-90*

Lindmark E, Chen Y, **Georgoudaki AM**, Dudziak D, Lindh E, Adams WC, Loré K, Winqvist O, Chambers BJ, Karlsson MCI  
**AIRE expressing marginal zone dendritic cells balances adaptive immunity and T-follicular helper cell recruitment**  
*Journal of Autoimmunity, 2013 May;42:62-70*

# CONTENTS

1	Introduction .....	3
1.1	The immune system .....	3
1.1.1	Lymphoid organs .....	4
1.1.2	The innate immune system .....	5
1.1.3	The adaptive immune system .....	6
1.1.4	Lymphocytes .....	6
1.1.5	Myeloid cells .....	11
1.2	The immunosuppressive tumor microenvironment .....	21
1.2.1	Tumor-associated macrophages .....	22
1.3	Epithelial-Mesenchymal Transition in tumor metastasis.....	26
1.4	Cancer immunotherapy.....	28
1.4.1	Immunotherapy using monoclonal antibodies .....	28
1.4.2	Targeting tumor-associated macrophages .....	32
1.4.3	Exosomes in cancer immunotherapy.....	34
1.5	Murine models of human melanoma, breast and colon cancer .....	35
1.5.1	Mouse models of melanoma (B16) .....	35
1.5.2	Mouse model of breast cancer (4T1).....	36
1.5.3	Mouse model of colon cancer (MC38).....	36
1.5.4	Mouse model for the study of EMT and metastasis.....	37
2	The present study.....	38
2.1	Aims .....	38
2.2	Results and discussion .....	39
2.2.1	Marginal zone macrophages regulate antigen transport by B cells to the follicle in the spleen via CD21 (Paper I).....	39
2.2.2	Reprogramming tumor associated macrophages by antibody targeting inhibits cancer progression and metastasis (Paper II) .....	41
2.2.3	TGF- $\beta$ 1-induced EMT promotes targeted migration of breast cancer cells through the lymphatic system by the activation of CCR7/CCL21-mediated chemotaxis (Paper III).....	44
2.2.4	Synergistic induction of adaptive antitumor immunity by codelivery of antigen with $\alpha$ -galactosylceramide on exosomes (Paper IV).....	46
2.3	Final reflections and future perspectives.....	49
3	Popular scientific summary (English, Svenska, Ελληνικά) .....	51
4	Acknowledgements .....	56
5	References.....	61

## LIST OF ABBREVIATIONS

<b><math>\alpha</math>-GalCer</b>	alpha-galactosylceramide	<b>mAb</b>	Monoclonal antibody
<b>Ab</b>	Antibody	<b>MAPK</b>	Mitogen-activated protein kinase
<b>ADCC</b>	Antibody-dependent cellular cytotoxicity	<b>MARCO</b>	Macrophage receptor with collagenous structure
<b>ADCP</b>	Antibody-dependent cellular phagocytosis	<b>MDSC</b>	Myeloid derived suppressor cell
<b>ADP</b>	Adenosine diphosphate	<b>MHC</b>	Major histocompatibility complex
<b>Ag</b>	Antigen	<b>MHC-I/-II</b>	MHC class I/II
<b>AP-1</b>	Activator protein-1	<b>MMP</b>	Matrix metalloproteinases
<b>APC</b>	Antigen presenting cells	<b>Mo-MDSC</b>	Monocytic-MDSC
<b>ATP</b>	Adenosine triphosphate	<b>MyD88</b>	Myeloid differentiation primary response gene 88
<b>BCR</b>	B-cell receptor	<b>MZ</b>	Marginal zone
<b>CCL</b>	Chemokine (C-C motif) ligand	<b>MZB</b>	Marginal zone B cells
<b>CCR</b>	Chemokine (C-C motif) receptor	<b>MZM</b>	Marginal zone macrophages
<b>CD</b>	Cluster of differentiation	<b>M<math>\Phi</math></b>	Macrophages
<b>CD40L</b>	CD40 ligand	<b>NALP3</b>	NACHT, LRR and PYD domains-containing protein 3
<b>CDR</b>	Complementarity determining regions	<b>NF-<math>\kappa</math>B</b>	Nuclear factor kappa-light-chain-enhancer of activated B cells
<b>CRI</b>	Cancer-related inflammation	<b>NK</b>	Natural Killer
<b>CSF-1(R)</b>	Colony stimulating factor-1 (receptor)	<b>NKT<sub>(m)</sub></b>	Natural killer-like T (follicular helper) cell
<b>CTL</b>	Cytotoxic T lymphocytes	<b>NLR</b>	Nod-like receptor
<b>CTLA-4</b>	cytotoxic T-lymphocyte-associated protein 4	<b>NOD</b>	Nucleotide-binding oligomerization domain
<b>CXCR</b>	CXC chemokine receptor	<b>PAMP</b>	Pathogen associated molecular patterns
<b>DAMP</b>	Danger associated molecular pattern	<b>PD-(L)1</b>	Programmed death-(ligand) 1
<b>DC</b>	Dendritic cell	<b>pMHC</b>	Peptide-MHC complex
<b>EGF</b>	Epidermal growth factor	<b>PMN-MDSC</b>	Polymorphonuclear-MDSC
<b>EMT</b>	Epithelia-to-Mesenchymal Transition	<b>PRR</b>	Pattern recognition receptor
<b>Fab</b>	Fragment, antigen binding	<b>RLR</b>	RIG-I-like receptors
<b>Fc</b>	Fragment, crystallizable	<b>SIGN-R1</b>	Specific intracellular adhesion molecule-3-grabbing non-
<b>Fc<math>\gamma</math>R</b>	Fc gamma receptor	<b>SLE</b>	Systemic lupus erythematosus
<b>FDC</b>	Follicular dendritic cell	<b>SR</b>	Scavenger receptor
<b>FOB</b>	Follicular B cell	<b>SRCR</b>	Scavenger receptor cysteine rich
<b>GC</b>	Germinal center	<b>STAT</b>	Signal transducer and activator of transcription
<b>GITR</b>	glucocorticoid-induced TNFR-related protein	<b>TAM</b>	Tumor associated macrophages
<b>GM-CSF</b>	Granulocyte-macrophage colony-stimulating factor	<b>TAN</b>	Tumor associated neutrophils
<b>IC</b>	Immune complexes	<b>TCR</b>	T cell receptor
<b>ICD</b>	Immunogenic cell death	<b>TD</b>	Thymus-dependent
<b>IDO</b>	Indoleamine 2,3-dioxygenase	<b>T<sub>h</sub></b>	T follicular helper cell
<b>IFN</b>	Interferon	<b>TGF-<math>\beta</math></b>	Tumor growth factor beta
<b>Ig</b>	Immunoglobulin	<b>T<sub>h</sub></b>	T helper cell
<b>IL</b>	Interleukin	<b>TI</b>	Thymus-independent
<b>ILC</b>	Innate lymphoid cells	<b>TI-I/-II</b>	Thymus –independent Type I/II
<b>(i)NKT</b>	Invariant natural killer T cells	<b>TIL</b>	Tumor infiltrating lymphocytes
<b>iNOS</b>	Inducible nitric oxide synthase	<b>TLR</b>	Toll-like receptor
<b>IRF</b>	Interferon regulatory factor 1	<b>TNF(R)</b>	Tumor necrosis factor (receptor)
<b>IVIG</b>	Intravenous immunoglobulin	<b>TRAIL</b>	TNF-related apoptosis-inducing ligand
<b>KIR</b>	Killer-cell immunoglobulin-like receptors	<b>Treg</b>	T regulatory cell
<b>KO</b>	Knock out	<b>VEGF</b>	Vascular endothelial growth factor
<b>LPS</b>	Lipopolysaccharide		
<b>M-CSF</b>	Macrophage colony-stimulating factor		

# 1 INTRODUCTION

## 1.1 THE IMMUNE SYSTEM

The immune system is an ensemble of sophisticated sensors and effectors that are in place to ensure tissue homeostasis and protect from invasion by infectious pathogens. Threats can be detected by a plethora of molecular sensors, which vary depending on the context of the threat. Accordingly, these sensors also target different responders with various functional activities. The common goal of these immune responses is to restore homeostasis or eliminate threats by inducing inflammation. Although infectious pathogens are the most commonly considered instigators, inflammation also occurs during tissue injury, cancer, autoimmune disease and metabolic deregulation <sup>1</sup>.

The existence of the immune system was observed as early as 430 BC by the Athenian Thucydides who described that patients who recovered from the plague were not struck a second time or to the same extent by the disease. This observation describes the concept of immunity, which refers to the physiological state where an organism exhibits lack of susceptibility towards unwanted or noxious agents. The term originates from the Latin *immunitas*, which means “to be exempt from”. References to active immunity date back to the early 1700s, when the practice of variolation in the Ottoman Empire (Turkey) was reported in the *Philosophical Transactions* of the Royal Society of London.

In 1796, Edward Jenner, set up an experiment that would for the first time demonstrate the workings of active immunotherapy and what we today refer to as immunological memory by showing that exposure to cowpox induced protection to smallpox. In 1884 Robert Koch identified pathogens as the underlying cause of infectious diseases postulating the “Germ theory”. This would set the basis for what Louis Pasteur in 1891 termed vaccination (*fr. latin vacca*; cow), posthumously crediting Jenner for his contribution and expanding the term to include immunizations against various different pathogens. In parallel, great advances were made in the field of immunology, such as the discovery by Ilya Metchnikov in 1866 of leukocytes (white blood cells) with the capacity to ingest pathogens. He termed this process phagocytosis and the cells exerting this function phagocytes, an important component of the innate immune system <sup>2</sup>. This marked the birth of the branch of cellular immunity and earned Metchnikov the Nobel prize in 1908.

Further efforts to elucidate the mechanisms of induced immunity lead Emil von Behring and Shibasaburo Kitasato in 1890 to identify antitoxins, soluble factors in the serum, that were able to neutralize bacterial toxins. Antitoxins, later termed antibodies (Abs), could confer passive immunity to naïve individuals. Paul Ehrlich was the first to suggest the lock-and-key model describing the mechanism of molecular interactions through which Abs bound their ligands. Antibodies, together with the complement and antimicrobial peptides, constitute humoral immunity. The generation of such pathogen-specific immune responses would later be attributed to the adaptive immune system, which is acquired during the lifetime of an individual.

The above-mentioned discoveries come together to form our current view on the concept of immunology. As it is understood today, the immune system is a set of interactive networks which have evolved to eliminate potential threats (infection) and restore homeostasis (following trauma or cancer), while retaining sufficient tolerance to avoid reactivity to self (autoimmunity) or otherwise innocuous agents (allergy). A vital prerequisite for this to occur is the ability of the immune system to discriminate “self” from “non-self”. Hence, recognition is an important capacity of both the early, innate immune system and the late, specific adaptive immune system and holds the key to preserving the balance between immune tolerance and immune-mediated elimination.

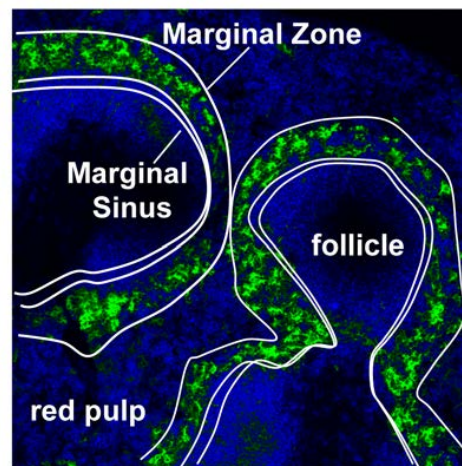
Below follows a description the two branches of the immune system, which collaborate to orchestrate immune cascades with the aim to restore immunity and homeostasis. Particular emphasis will be given to the cells and mechanisms relevant to this thesis.

### **1.1.1 Lymphoid organs**

Immune cells arise from the CD34<sup>+</sup> hematopoietic stem cell lineage whose developmental niche is primarily the bone marrow. The bone marrow, along with the fetal liver and the thymus are termed primary lymphoid organs and are sites where immune cells are generated. Upon differentiation from common progenitors, the pluripotent hematopoietic stem cells, immune cells diverge to different lineages and egress from the bone marrow. In the periphery they populate different tissues, as well as secondary lymphoid organs, which function as major hubs where immune responses are generated. The secondary lymphoid organs include the spleen, lymph nodes, Peyer’s patches, mucosa/gut-associated lymphoid tissue (MALT/GALT) and tonsils. Secondary lymphoid organs are dispersed throughout the body providing a network for efficient communication, facilitating immune interactions. These organs are interconnected by the lymphatic vessels. Some sites, however, are ”protected” from possible adverse effects of inflammatory responses and therefore immune cell presence is restricted<sup>3</sup>. These sites are known as immune privileged and include the central nervous system, testis and eyes. These sites are instead guarded by other protective mechanisms, such as immunoglobulins (Igs) and complement.

The spleen, the largest peripheral lymphoid organ, is an important site where innate immune cells migrate to orchestrate adaptive immune responses. The splenic red pulp contains macrophages, which are responsible for scavenging aging erythrocytes and uptake of antigen (Ag) from the blood. The white pulp is the lymphoid compartment of the spleen, which consists of highly organized structures, known as the B cell follicles and the surrounding T cell areas, much resembling a lymph node. B cell follicles are the sites where Ag-specific adaptive immune responses are initiated. In mice, located between the white and red pulp is the marginal sinus, which directly connects the spleen to the circulation and is in strategic proximity of the marginal zone (MZ) of the spleen surrounding the follicles. Specialized cells populating the marginal zone, the marginal zone macrophages (MZMs), the marginal zone B cells (MZBs) and dendritic cells (DCs), surveil the blood entering the marginal sinus and capture antigens Ags through pattern recognition receptors (PRRs)<sup>4,5</sup> (**Figure 1**). MZBs

shuttle Ags from the MZ into the follicles for Ag-presentation on follicular dendritic cells (FDCs) and activation of adaptive T and B cell responses through germinal center (GC) formation. The MZ is a dynamic structure where MZMs and MZBs are interdependent for their retention in the MZ and their trafficking to specialized splenic compartments upon Ag uptake <sup>6</sup>. Similar interactions are observed at other sites of immune surveillance, for example between subcapsular macrophages and follicular B cells (FOBs) in the lymph nodes, and peritoneal macrophages and B1 B cells in the peritoneal cavity. This common interaction between the two cell types suggests an important role for macrophages in regulating antigen availability and presentation by B cells, which will be addressed further in Paper I.



**Figure 1. The marginal zone of the spleen**  
Immunofluorescence staining on histological section of a mouse spleen. Green indicates MZM expressing MARCO, lining the outer side of the marginal sinus of a B cell follicle, B cells are stained with B220 (blue).

### 1.1.2 The innate immune system

The innate immune system is the body's primary line of defense. It partly consists of physical barriers, such as the skin and mucosal surfaces, and anti-microbial peptides, which act by inhibiting pathogens at sites of entry. Additionally, the innate immune system consists of cells such as monocytes, macrophages, DCs, neutrophils, eosinophils, basophils and mast cells which are of myeloid origin, but also some DCs, natural killer (NK) cells and the recently described innate lymphoid cells (ILCs), which are lymphocytes. The innate immune system is characterized by rapid and broad responses against threats. The "self" vs. "non-self" discriminating ability of the innate immune system lies in the vast array of molecular PRRs. Some PRRs of ancestral defense systems (such as Ig-like structures, scavenger receptor cystein-rich domains and Toll-like receptors) are preserved through evolution. More specifically, the mechanistic basis for the innate immune system's efficiency depends on the recognition of common conserved molecular signatures/structures on pathogens, known as pathogen-associated molecular patterns (PAMPs), by PRRs. PRRs discriminate molecular patterns signaling altered self, non-self and damage/danger <sup>7</sup>. PRRs can be divided into membrane bound; such as scavenger receptors (SR), toll-like receptors (TLR) and C-type lectin receptors, cytoplasmic; such as NOD-like receptors (NLR) and retinoic acid-inducible gene (RIG)-I-like receptors (RLR), and secreted; complement receptors and others <sup>8</sup>. Charles



Janeway proposed the Infectious non-self theory, whereby the immune system recognizes noxious foreign agents through PRRs on antigen presenting cells (APCs). Later, Polly Matzinger suggested the revised ‘Danger model’, suggesting that the immune system can distinguish between dangerous and innocuous stimuli of either self- or foreign origin <sup>9</sup>. Similarly to PAMP recognition, PRRs are also capable of recognizing endogenous components signaling ‘damage’. These signals are released into the surrounding extracellular space, actively secreted or exposed on the surface of dying, stressed or injured cells: these include calreticulin (CRT), adenosine triphosphate (ATP) and high mobility group protein B1 (HMGB1). They are known as danger-associated molecular patterns (DAMPs) and have recently been shown to function as adjuvants (generating “sterile” inflammation) for the induction of immune responses during cancer immunotherapy, a concept described as immunogenic cell death (ICD) <sup>10</sup>. Release of extracellular ATP by apoptotic cells functions as a “find me” signal, but ATP can also mediate an array of immunoregulatory responses and is thus of particular interest for this thesis. ATP release during ICD can activate the immune system, however its catabolite adenosine has the opposite effect, immunosuppression. The catabolic activity of ectonucleotidase CD39 is responsible for the reversible conversion of ATP to ADP (adenosine diphosphate) and ADP to AMP (adenosine monophosphate), while CD73 handles the irreversible conversion of AMP to adenosine <sup>11</sup>.

### **1.1.3 The adaptive immune system**

The adaptive immune system arose as a prerequisite for survival during the evolution of increasingly complex organisms and evolved to fulfill the need for a more specialized line of defense. In contrast to innate immune responses, adaptive immune responses are not self-sufficient. They depend on and are tightly interwoven with components of the innate immune system for the orchestration of functional cascades of events that aim to restore homeostasis. Another basic difference between the two branches of immunity, is the fact that adaptive immune responses are antigen-specific and encompass a memory component to previously encountered Ags. The main cellular components of the adaptive immune system are the T and B cells, which arise from common lymphoid progenitors in the bone marrow and diverge into separate lineages in the periphery. Depending on the type of antigen, different types of immune responses are orchestrated by the secretion of different cytokines, leading to the mobilization of various subsets of immune effector cells. These processes ultimately induce clonal expansion of activated T and B cells, as well as deployment to the site of inflammation where they exert various effector functions. Thus, the peripheral lymphoid organs, the spleen and the lymph nodes, are the sites where cellular components of the innate immune system activate cellular and humoral responses of adaptive immunity.

### **1.1.4 Lymphocytes**

The cells of the lymphoid lineage originate from common lymphoid progenitors in the bone marrow, which give rise to T cells,  $\gamma\delta$  T cells, NKT cells, NK cells, innate lymphoid cells (ILC), subsets of DCs and B cells. The relevant subsets for this thesis are described below.

#### 1.1.4.1 T cells

T cells are part of the lymphoid hematopoietic lineage. T lymphocyte precursors (thymocytes) originate from common lymphoid precursors in the bone marrow, which migrate to the thymus where T cell development occurs. Like the innate branch, the functionality of the adaptive immune system relies on efficient recognition. Therefore, T cells express the T cell receptor (TCR) on their surface, which is generated through complex gene rearrangements of the Tcr loci during T cell development in the thymus. The TCR consists of one  $\alpha$  and one  $\beta$ -chain, it has a single unique antigen-recognition site and binds to antigenic peptides in complex with the major histocompatibility complex (MHC) on the surface of APCs. Upon engagement of the Ag-specific TCR with its cognate Ag, the cell expressing that TCR undergoes extensive proliferation, known as clonal expansion, and part of its progeny will go on to mount a directed immune response against the specific target, whether it be an infected cell or cancer cell. Other progeny will form a memory compartment that can instantly be deployed upon re-encounter of the Ag. Consequently each individual acquires a unique repertoire of Ag-specific T cell clones depending on their antigenic encounters.

More specifically, T cell precursors in the thymus express a functional TCR as well as co-receptors CD4 and CD8. During the first step, T cells whose TCR will engage self-peptide-MHC (pMHC) complexes on the cell surface of cortical thymic epithelial cells (cTECs) are selected, while absence of this “survival” signal results in death by neglect. This process is termed positive selection and ensures the functionality of the selected clones. Selected clones undergo a second step, negative selection, during which T cells with a TCR that engages self-pMHC on the surface of medullary TECs too strongly are eliminated through apoptosis. This is known as negative selection and aims to eliminate autoreactive clones. Selection is thus, regulated by a certain threshold of activation.

Thereafter, selected naïve T cells egress from the thymus and populate peripheral lymphoid tissues, such as the spleen and lymph nodes. There, induction of adaptive immune responses relies on the activation of the innate immune system and antigen presentation. During Ag-presentation by APCs to naïve T cells in peripheral lymph nodes, three signals are required for lymphocyte activation to occur. The first signal is provided by engagement of the TCR with the peptide-MHC complex and the second signal is provided by the activated DC in the form of co-stimulatory molecules CD80 and CD86. Depending on the context of the immune response the DC will provide a third activation signal in the form of soluble mediators, such as cytokines, which will polarize T cells to different effector phenotypes. pMHC-I complexes are recognized by CD8<sup>+</sup> T cells, while pMHC-II complexes are recognized by CD4<sup>+</sup> T cells. Thus, CD8<sup>+</sup> T cells become cytotoxic effectors, while CD4<sup>+</sup> T cells polarize into various helper populations driven by the up-regulation of different transcription factors depending on the inflammatory mediators in the microenvironment (more specifically: cytokines interferon  $\gamma$  (IFN $\gamma$ ) and IL-12 drive the expression of transcription factor *T-bet* which leads to T helper 1 (T<sub>h</sub>1) polarization. Similarly, IL-4/*GATA-3* drive T<sub>h</sub>2, IL-6/IL-21/IL-23/*ROR $\gamma$ T* drive T<sub>h</sub>17 and TGF- $\beta$ /*Foxp3* drive T<sub>reg</sub> polarization).

#### 1.1.4.2 Natural Killer cells

NK cells were discovered in 1975 by Rolf Kiessling, Eva Klein and Hans Wigzell at Karolinska Institutet<sup>12,13</sup> and Ronald Heberman et al. simultaneously, at the University of Pittsburgh<sup>14,15</sup>. They are cells of lymphoid origin with the capacity to exert cytotoxic activity against virus infected or tumor cells without the need for prior activation. As such, they are attractive candidates for cancer immunotherapy<sup>16</sup>. They are potent producers of IFN $\gamma$  and have cytotoxic granules containing perforin as well as granzyme B. NK cell recognition and binding to self-pMHC complexes on the surface of other cells leads to NK cell inhibition. This is mediated by the inhibitory killer cell Ig-like receptors (KIR) in human NK cells and by the c-type lectin-like Ly49 family of receptors in mice, and by NKG2A, which is conserved between the species. NK cells are educated to recognize a vast repertoire of MHC-I loaded with self-peptides. However, in the absence of self-pMHC-I recognition, lack of inhibitory receptor signaling leads NK cell activation, according to the “missing self hypothesis”<sup>17</sup>. This can occur e.g. in an MHC-I mismatch transplantation setting or in tumors that have down-regulated MHC-I as an immune escape mechanism. NK cells also express CD16 (Fc $\gamma$ RIIIA/B in humans, Fc $\gamma$ RIII in mice), a receptor involved in antibody-dependent cellular cytotoxicity (ADCC), and can be activated by binding of the Fc-part of Abs, as well as activating receptors such as DNAM1, NKp30, NKp44, NKp46 and NKG2D, whose ligands are present on tumor cells and infected cells. Thus, the final fate of NK cell responses results from the net balance of activating vs. inhibitory signal input.

#### 1.1.4.3 Natural Killer T cells

NKT cells are innate lymphoid cells that share characteristics of both NK cells, such as expression of NK1.1 and NKG2D, and T cells as they express a TCR<sup>18</sup>. They are a small population making up 0.1% of the cells in the human circulation. The TCR of NKT cells is restricted to CD1d, an MHC-like presentation molecule which presents lipid and glycolipid Ag. Like NK cells, upon activation they secrete perforin and granzymes upon activation but they are also potent producers of cytokines. Unlike T cells, their TCR is less variable, only consisting restricted  $\alpha$  and  $\beta$  chains. There are two types of NKT cells, Type I invariant NKT cells (iNKT) that bind the prototypic glycolipid Ag  $\alpha$ -galactosylceramide ( $\alpha$ -GalCer), and Type II diverse NKT cells which bind sulfatide. The study of NKT cells has been facilitated by the generation of CD1d-tetrameres loaded with  $\alpha$ -GalCer. However, deciphering between the actions of the two subpopulations is still complex. There are two mouse models that allow the *in vivo* study of NKT cells. J $\alpha$ 18 knock-out (KO) mice are deficient in iNKT cells, while CD1d KO lack both subtypes. Thus the observed differential outcomes while comparing immune responses in these two strains has helped shed light onto the action of Type II NKT cells. In homeostasis, CD1d presents self-lipids to NKT cells. In mice, iNKT cells are classified into two major categories based on the expression of CD4 and CD8. The majority are CD4<sup>+</sup> and the remaining are CD4<sup>-</sup>CD8<sup>-</sup>, and differ in their ability to secrete cytokines<sup>19</sup>. They have been described to contribute to both pro- and anti-inflammatory innate immune responses depending on the cytokine context in their activation milieu and the lipid Ag they recognize. According to their function NKT cells

can be further classified into NKT<sub>H1</sub>, NKT<sub>H2</sub>, NKT<sub>H17</sub> and NKT<sub>FH</sub> cells, and upon activation they become rapid producers of various cytokines<sup>20</sup>. In response to  $\alpha$ -GalCer, NKT cells secrete IFN $\gamma$ , inducing the maturation of DC into APCs by providing co-stimulation via the CD40-CD40L interaction. This interaction also leads to the release of interleukin 12 (IL-12) from activated DC which further propagates CD8<sup>+</sup> T cells through DC-cross-priming via CD70<sup>21</sup>. Notably, many solid tumors express CD1d on their surface and the role of NKT cells in tumor immune surveillance is compelling, as suggested by their low number and loss of functionality in cancer patients<sup>22</sup>. Moreover, iNKT cells have been reported to kill tumor-associated macrophages, a major immunosuppressive cell type found in tumors (described in 1.2.1), thus contributing to anti-tumor immunity<sup>23</sup>. Due to this, most studies on iNKT cells in cancer immunotherapy utilize the so far best-characterized agonist,  $\alpha$ -GalCer, which has been widely explored both experimentally and recently also in clinical trials as an adjuvant in DC-based vaccine approaches<sup>24,25</sup>. However, attempts to enhance NKT cell mediated anti-tumor responses by administration of soluble  $\alpha$ -GalCer have been shown to cause anergy and therefore more controllable delivery systems and other agonistic analogues are being explored (see Paper IV)<sup>26,27</sup>. Examples of such are the co-administration of  $\alpha$ -GalCer and tumor-Ag or the adoptive transfer of  $\alpha$ -GalCer-loaded DCs or exosomes with tumor-Ag, both of which induced durable NKT cell cytokine responses<sup>28,29</sup>.

#### 1.1.4.4 B cells

B cells were discovered by Max Cooper in 1965 as a separate type of lymphocytes than T cells<sup>30,3130,3130,31</sup> that develop from common lymphocyte precursors in the bone marrow<sup>29,30</sup>. In the bone marrow, they transition from pro-B cells, late pro-B cells, large pre-B cells, small pre-B cells and immature B cells that egress the bone marrow into the blood stream and migrate to peripheral lymphoid organs where they later mature<sup>32</sup>. During their development in the bone marrow B cells interact with stromal cells and recognize Ags through the highly specific B cell receptor (BCR), which is a membrane-bound Ig that arises from the random rearrangement of the gene segments of the Ig locus. The BCR consists of two pairs of a heavy and a light chain ( $\kappa$ , kappa or  $\lambda$ , lambda) which come together to form two identical antigen-binding regions that directly bind to antigenic peptides, the complementarity determining regions (CDRs)<sup>33</sup>. Similar to T cells, the cost of the high specificity of a vast array of BCRs is the generation of autoreactive B cell clones. Upon engagement of the BCR with self-Ags, B cells are eliminated by apoptosis or undergo editing of the Ag-specificity of their BCR to avoid self-reactivity and are then released into the periphery. This gives rise to a wide repertoire of receptors specificities (up to 10<sup>8</sup> different ones), which accommodate a vast heterogeneity of Ags. Although central tolerance provides an efficient checkpoint it is not infallible, as 4% of immature B cells in the periphery are autoreactive. Several mechanisms, collectively known as peripheral tolerance, are put in place to compensate for the escape of those autoreactive clones from central selection. These include suppression by T<sub>regs</sub>, induction of anergy in the absence of co-stimulation or presence of co-inhibition, and immune privileged sites.

The main innate-like B cell subset is the B1 B cells that populate the peritoneal cavity, the MZBs lining the marginal sinus of the spleen and the FOBs, which are found in the follicles of the spleen and lymph nodes. FOBs express CXCR5, which is important for their retention in the follicles in response to CXCL13. While FOBs recirculate, MZBs are mostly regarded as resident in the MZ. MZBs comprise a rather scarce, highly specialized B cell subpopulation in the spleen. They localize in close proximity to MZMs and DCs filtering the incoming blood for particulate Ags. MZBs are generally identified as B220<sup>+</sup> CD21<sup>hi</sup> (complement receptor 2) CD23<sup>lo</sup>. Additionally they also express CD1d and can thus recognize lipid Ags. Like other innate-like cells, they are readily activated and respond with rapid induction of IgM production and plasma blast formation. MZBs and MZMs are interdependent and this cross talk is important for effective capture and coordinated early IgM responses to bacterial Ags<sup>34,35</sup>. MZBs are important for the continuous shuttling of Ags into the follicles via complement receptors and deposition on the FDCs so that GC reactions can give rise to Ag-specific adaptive effector and memory responses<sup>36,37</sup>. Ag-loaded CXCR5-expressing MZBs respond to FDC-produced CXCL13 by migrating into the follicle and depositing Ag in the form of immune complexes (IC) on FDCs for subsequent activation of FOBs. IC deposition is mediated through the interaction of complement receptors CD21 on MZBs and CD35 on FDCs. FOBs bind and internalize Ags through the BCR, process it and present it on MHC class II on their surface. The additional activation signal is delivered by an activated CD4<sup>+</sup> T helper cell which travels to the T/B border of the spleen, recognizes the peptide-MHCII complex and engages in a cognate T/B cell interaction. This leads to up-regulation of CD40L on T cells, which provides the co-stimulatory signal upon binding to CD40 on B cells, licensing B cells to enter the GC reaction and resulting in plasma cell formation and antibody production<sup>38</sup>. During the GC reaction B cells undergo class switch recombination and affinity maturation where B cells with high affinity to the Ag are selected. The selected B cells then undergo somatic hypermutation (SHM) where random mutation in the CDRs of the BCR further increase its specificity and affinity. A specific subtype of T cells, of particular importance for the GC reaction, is the T follicular helper cells (T<sub>fh</sub>). T<sub>fh</sub> initiate the GC response by providing survival, proliferation and differentiation signals to FOBs after cognate interaction. They also, secrete cytokines which determine the subclass (isotype) of the BCR and generated Abs. The resulting high affinity B cells will then differentiate into antibody-producing plasma cells or memory B cells.

Depending on the type of Ag encountered, different modes of B cell activation can occur. Antigen types include: Thymus-dependent (TD), such as TNP-KLH and Thymus-independent (TI). TD Ag activation involves the B cell receiving CD4<sup>+</sup> T cell help in the form of co-stimulatory CD40-CD40L interaction, as described above. TI Ag can be further subdivided into TI type I or TI type II. TI-I Ag (TLR-ligand polysaccharides and cytosine-phosphodiester-guanin (CpG)) lead to polyclonal B cell activation through engagement of TLR4 on the B cell surface. TI-II Ag are long polysaccharides (e.g. dextran, NP-Ficoll) that can simultaneously bind and activate multiple B cells. B cell receptors, B1 B cells and MZBs are mostly associated to TI Ag responses<sup>39</sup>, while FOB are associated to TD Ag responses. MZB cells also highly express the MHC-like molecule CD1d, which is involved in lipid Ag

presentation to NKT cells in the splenic MZ and red pulp <sup>40</sup>.

The role of B cells in cancer is disputable, with studies reporting both positive and deleterious influence of B cells on anti-tumor immunity. In a melanoma tumor model it was shown that B cells were essential for the mediation of CD4<sup>+</sup> and CD8<sup>+</sup> anti-tumor responses <sup>41</sup>. Another study describes a mechanism through which B cells drive carcinogenesis through the generation of immune complexes that bind FcγR, thus contributing to chronic inflammation <sup>42</sup>. Moreover, B cells secrete anti-inflammatory cytokines, such as IL-10 and TGF, which have profound immunosuppressive effect on other tumor-infiltrating immune cells <sup>43</sup>. The role of B cells in the regulation of macrophages in the tumor microenvironment (TME) is described in 1.2.1.

### **1.1.5 Myeloid cells**

Myeloid cells originate from common myeloid precursors in the bone marrow, which give rise to granulocytes, monocytes, macrophages and DCs. The relevant subsets for this thesis, with particular focus on macrophages, are described below.

#### *1.1.5.1 Granulocytes*

Granulocytes are cells of the myeloid hematopoietic lineage and consist of subsets with distinct morphology and function, namely eosinophils, mast cells, basophils and neutrophils; which are the most abundant subset in humans, accounting for 70% of all circulating leukocytes. Neutrophils are short-lived, rapid responders to inflammatory cues, which migrate to sites of inflammation through a process known as chemotaxis. At site, they engulf microbial pathogens in phagosomes, which then fuse with intracellular granules containing reactive oxygen species (ROS), eliminating the pathogen. Alternatively, granules containing antimicrobial enzymes are released into the extracellular space. During their high turn over rate, dying neutrophils also have the ability to externalize their nuclear content forming neutrophil extracellular traps (NETs) that capture pathogens immobilizing them to be engulfed by other phagocytes. Eosinophils, basophils and mast cells perform immune responses against extracellular pathogens by engaging IgG and IgE antibodies via their Fc receptors.

#### *1.1.5.2 Dendritic cells*

In 1973, Ralph Steinman and Zanvil Cohn discovered a novel cell type, the DCs, which since then have been described to be the fundamental cellular sensors of the immune system and the bridge between innate and adaptive immune responses (Nobel prize 2011 for R.S.) <sup>44-46</sup>. DCs belong to the myeloid lineage, they originate from the bone marrow and develop into immature tissue-specific subsets that populate and surveil the periphery for potential invaders <sup>47</sup>. Cytokines fms-like tyrosine kinase three ligand (Flt3L) and granulocyte-monocyte colony stimulating factor (GM-CSF) are important for DC differentiation. There are two major subsets of DC in mouse, the myeloid DCs (mDCs) and the plasmacytoid DCs (pDCs). mDCs (also known as conventional DCs) are subdivided into CD8α<sup>+</sup> or CD8α<sup>-</sup> and originate from precursors in the blood, while inflammatory DCs arise from circulating inflammatory

monocyte precursors<sup>48</sup>. pDCs are potent responders to TLR7 and TLR9 stimuli, such as viral RNA and bacterial CpG-ODNs (oligodeoxynucleotides), to which they respond by producing type I interferons (IFN $\alpha$ , IFN $\beta$ ).

Immature DC are phagocytic cells, which typically express high levels of Ag-presenting molecules MHC-I and -II and CD1d, integrin CD11c, intracellular adhesion molecule 1 (ICAM-1) and low levels of co-stimulatory molecules CD80, CD86 and CD40. Ag-recognition by PRRs, leads to phagocytic uptake and lysosomal degradation, upon which antigenic peptides are loaded on MHC class II molecules from the endoplasmic reticulum and transported to the cell surface of the DC for subsequent presentation to naïve T cells<sup>49</sup>. This process leads to DC maturation, at which point the expression of co-stimulatory molecules is up-regulated and IL-12 is secreted to assist T cell activation. Additionally, the expression of chemokine receptor CCR7 is up-regulated, which detects chemotactic gradients of CCL19 and CCL21, and induces DC migration to lymph nodes for Ag-presentation to occur<sup>50</sup>. CD8 $\alpha^+$  DCs are capable of Ag cross-presentation through loading on MHC class I molecules. Ag-presentation is mediated through the formation of the pMHC/TCR complex. This induces changes in the cell membrane topology and causes the rearrangement of MHC-II, and co-stimulatory molecules CD80 and CD86 on the DC and of TCR, co-receptors CD4, CD8 and co-stimulatory molecule CD28 on the T cell, assembling a unique structure at the DC/T cell contact interface, the immunological synapse<sup>51</sup>. The immunological synapse potentiates the relay of three signals that determine the outcome of the interaction: 1) pMHC/TCR-mediated T cell activation (CD4/MHC-II or CD8/MHC-I), 2) CD80/CD28 or CD86/CD28 co-stimulation and 3) cytokine production.

Due to their inherent ability to coordinate innate and adaptive responses, DCs or DC-exosomes are potential tools for therapeutic cancer vaccine approaches aiming to induce tumor-specific cytotoxic responses and induction of immunological memory (Paper IV)<sup>52</sup>.

#### *1.1.5.3 Monocytes*

Circulating monocytes in the blood can be classified into two types; the inflammatory and the resident monocytes. Inflammatory monocytes originate from the bone marrow and respond to inflammatory stimuli by differentiating into macrophages or monocyte-derived DCs (in human CD14<sup>+</sup>CD16<sup>-</sup>, in mouse CCR2<sup>+</sup>Ly6C<sup>hi</sup>CX3CR1<sup>low</sup>)<sup>53</sup>. Resident monocytes, on the other hand, are thought to originate from circulating inflammatory monocytes and are important for tissue homeostasis and repair (in human CD14<sup>+</sup>/midCD16<sup>+</sup>, in mouse CCR2<sup>-</sup>Ly6C<sup>-</sup>CX3CR1<sup>hi</sup>)<sup>54</sup>. They rely on the macrophage-colony stimulating factor (M-CSF) for their survival and differentiation. Besides the bone marrow, monocytes also populate the spleen and are responsible for local replenishment of tissue-resident macrophages. The spleen also constitutes a reservoir of inflammatory monocytes that can be easily mobilized in response to soluble mediators and recruited to distal inflammatory sites<sup>55</sup>. The release of monocytes into the circulation, as well as the recruitment to sites of inflammation, are dependent on the chemokine CCL2.

#### 1.1.5.4 Macrophages

Macrophages are tissue-resident cells of the mononuclear phagocyte system with extreme functional diversity as is evident by the various highly specialized subpopulations across the different anatomical locations of the body. The name macrophages originates from the Greek: big eaters, from *makros* “large” + *phagein* “eat”, and describes their function as professional phagocytes responsible for the uptake and clearance of pathogens and endogenous debris through a wide array of PRRs. Recent studies have unveiled a broad spectrum of functions of macrophages, which far exceed their phagocytic activity, as indicated by their important role in the regulation of tissue homeostasis, inflammation and adaptive immune responses <sup>56</sup>.

#### *Subsets of macrophages*

Until recently, tissue-resident macrophages were regarded as descendants of circulating monocytes, which derive from hematopoietic stem cell precursors in the bone marrow. However, recent fate-mapping studies have shown that most tissue resident macrophages (in the lung, liver, peritoneum, spleen red pulp, bone marrow) originate from precursors in the yolk sac or the fetal liver during embryogenesis. These populations are self-maintained independently from circulating monocytes in steady state during adulthood <sup>57</sup>.

The spleen is our largest peripheral lymphoid organ and is extensively infiltrated by many different highly specialized subsets of tissue-resident macrophages. Depending on their sub-anatomical localization in the spleen these subsets exert different functions related to Ag uptake, recycling and presentation. Red pulp macrophages are derived from the fetal liver and are responsible for clearance of aged erythrocytes and recycling of heme <sup>58</sup>. White pulp macrophages (tingible body macrophages) are responsible for the clearance of B cells licensed by FDCs in the GC. The MZ of the spleen is the interface between the blood circulation and resident lymphocytes of the spleen. More specifically, as the anatomical structure of the MZ provides a site for the blood flow to be filtered for Ags, the macrophages guarding the marginal sinus are important for the uptake and clearance of self- and foreign Ags<sup>59</sup>. Another specialized macrophage subset, the metallophilic macrophages (MMs) expressing CD169 (sialoadhesin, Siglec-1), lines the inner side of the MZ surrounding the follicle. Although the origin of MZMs is unclear, they are dependent on the nuclear liver-X receptor  $\alpha$  (LXR $\alpha$ ) and arise early during development <sup>60</sup>. Of particular interest for this thesis and more in detail described below, are the MZMs. MMs are important for the initiation of humoral responses to TD Ags <sup>61</sup>.

MZMs are characterized by their expression of PRRs, amongst which class A scavenger receptors SR-A and the MARCO (macrophage receptor with collagenous structure), as well as the C-type lectin SIGN-R1 (specific intercellular adhesion molecule-3-grabbing non-integrin receptor 1). Due to their strategic localization in the MZ in close proximity to MZBs and DCs, they have an important role in bridging innate and adaptive immune responses. More specifically, SIGN-R1, an orthologue to human DC-SIGN (dendritic cell specific intracellular adhesion molecule-3-grabbing non-integrin), on MZMs binds different glycoproteins such as the capsular polysaccharide of *Streptococcus pneumoniae* (S.



*pneumonia*)<sup>62</sup> and polysaccharide dextran<sup>63,64</sup>. SR-A and MARCO on MZMs are involved in binding and clearance of meningococci<sup>65</sup>. SIGN-R1 also enhances the clearance of apoptotic cells through complement opsonization and is important for maintaining self-tolerance<sup>66</sup>. Although expressed on the same cell, SIGN-R1 and MARCO, exhibit opposing regulation, with MARCO being up-regulated while SIGN-R1 is down-regulated upon activation<sup>67</sup>. After bacterial or viral Ag uptake, macrophages in the MZ respond by producing a variety of pro-inflammatory mediators (Type I interferons, IL-1 $\beta$  and TNF $\alpha$ ) that ensure the propagation of the immune response. However, the same macrophages play an important role in mediating the anti-inflammatory effect of intravenous Ig (IVIG), a widely used treatment for many chronic autoimmune diseases. Binding of IVIG to SIGN-R1 leads to engagement of the Fc $\gamma$  receptor IIb (Fc $\gamma$ RIIb), a negative regulator of inflammation, which leads to an increased activation threshold and thereby suppression of autoimmunity<sup>68</sup>. MZMs are important for the suppression of innate and adaptive immune responses to apoptotic cells and maintenance of peripheral tolerance<sup>69,70</sup>. Scavenger receptor MARCO is essential for the uptake, clearance and maintenance of tolerance to apoptotic cells<sup>69</sup> and presence of autoantibodies against the receptor is indicative of systemic lupus erythematosus (SLE)<sup>71,72</sup>.

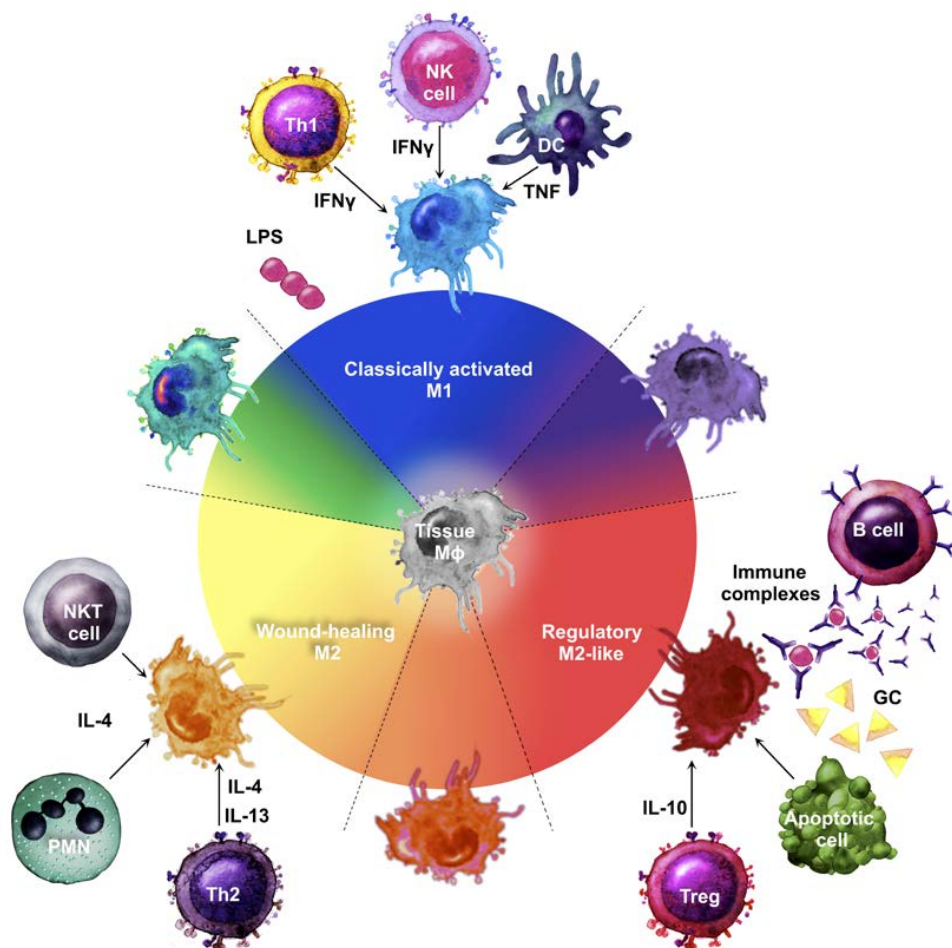
Similarly to MM, CD169<sup>+</sup> macrophages, the subcapsular sinus macrophages in the lymph nodes, are strategically located at site that allows immune surveillance and antigen capture from the lymph<sup>73</sup>. They play an important role in bridging innate and adaptive immunity as they have been shown to activate B cells by presenting opsonized Ags<sup>74,75</sup>, to cross-present apoptotic tumor cells to CD8<sup>+</sup> T cells<sup>76</sup> and to present lipid Ags to NKT cells<sup>77</sup>. During an immune response CD169<sup>+</sup> macrophages in the lymph nodes are mobilized into B cell follicles and during the GC reaction localize at the T-B border, supporting a role in Ag transport<sup>78</sup>. Medullary macrophages in the lymph nodes, which are responsible for filtering the lymph for particulate Ags, are also known to express CD169, SIGN-R1, SR-A and MARCO.

Peritoneal macrophages (originating from the fetal liver) are restricted to the transcription factor Gata-6 and upon TLR-stimulation with microbial products up-regulate scavenger receptor MARCO amongst other PRRs. Due to the easy accessibility peritoneal macrophages have been extensively studied both in their naïve state as well as activated upon administration of inflammatory agents in the peritoneal cavity.

#### *Macrophage activation and regulation*

Since macrophage function is highly context dependent, they exhibit great plasticity and diversity in phenotype and expression pattern of different molecules<sup>79,80</sup> and are very responsive to soluble mediators in their microenvironment. Thus, depending on the micro environmental context, macrophages are differently polarized along a continuous spectrum of activation states. Reflecting the effect of cytokines IFN $\gamma$  and IL-4 in inducing Th1 vs. Th2 immune responses<sup>81</sup>, macrophages were classified into classically activated (M1)<sup>82</sup> or alternatively activated (M2)<sup>83,84</sup> subsets depending on phenotype and function<sup>85</sup>. Later studies on macrophage activation performed in mouse strains with known predisposition toward either Th1 or Th2 responses, led to the distinction between the M1 and M2 activation

states based on phenotypic characteristics <sup>86,87</sup>. This classification was further developed to include several phenotypic marker and functional characteristics that distinguish the two activation states <sup>88-91</sup>. However, an increasing amount of data demanded a more informative classification based on the basic functions macrophages employ to preserve homeostasis, host defense, wound healing and immune regulation. This was further extended to include distinct transcriptional pathways regulating the different activation states <sup>92,93</sup>. Thus, in recent years, the M1/M2 dichotomy, which represents two extremes of a linear activation spectrum was replaced by a more correct representation of the polarization status as a continuous spectrum where all intermediate activations states can be encountered <sup>94</sup>. This is illustrated as a color wheel where the basic functions are depicted as primary colors and shared functions are represented by many different shades in-between, making up a spectrum of activation states that are related to different functions <sup>79</sup> (Figure 2).



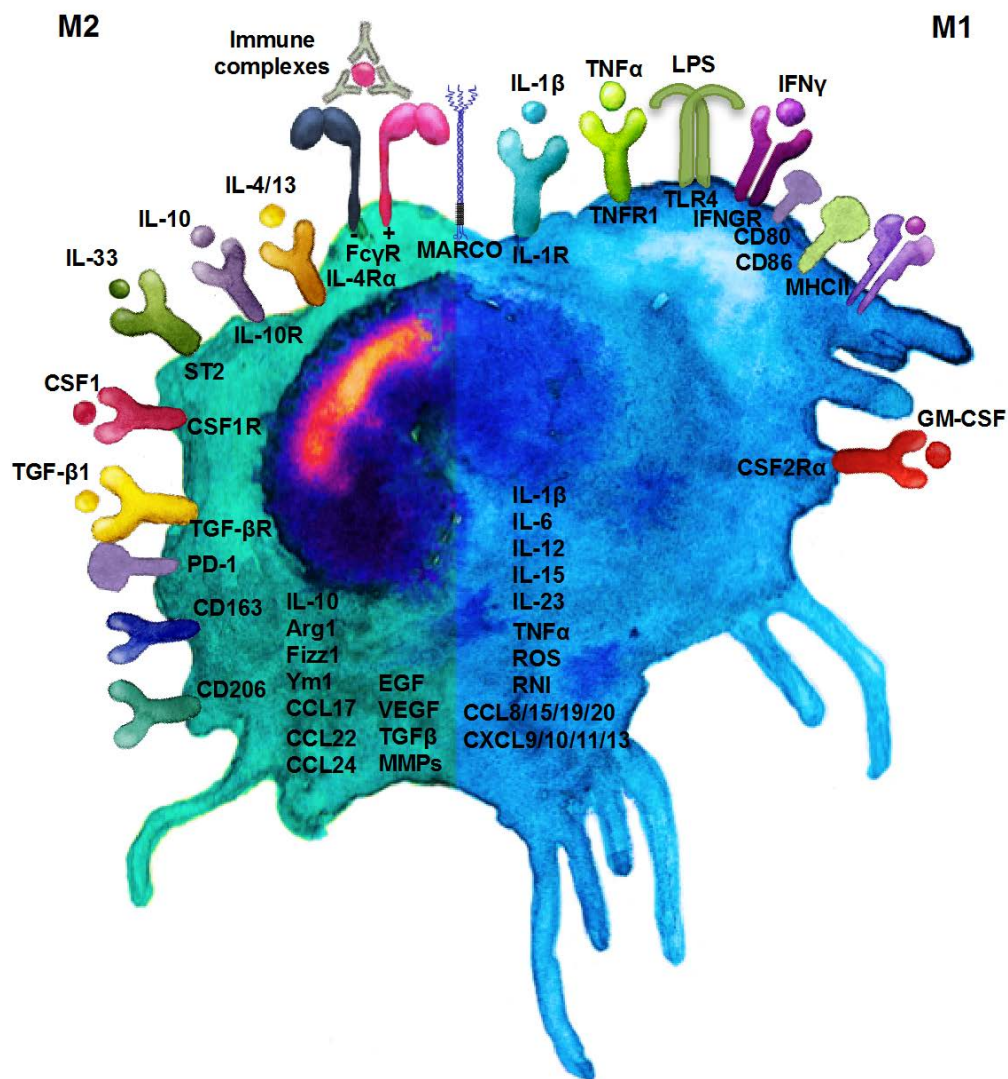
**Figure 2. The colorful wheel of macrophage polarization**

Macrophages can be polarized to a spectrum of different activation states. This is illustrated as a colorful wheel where the main macrophage activation modes are shown in primary colors: classical M1 activation in blue, wound-healing M2 in yellow and regulatory M2-like in red. Typically, M1 activation occurs in response to Th1-type cytokines, like IFN and TNF during infection, while M2 is induced during Th2-responses, by IL-4 and IL-13, e.g. in responses to parasites. M2 macrophages do not constitute a uniform population, thus intermediate colors represent activation states with shared characteristics. M2-like immunoregulatory macrophages are induced by immune complexes plus LPS or IL-1β, but also by IL-10, TGF-β and glucocorticoids. Finally, TAMs are a heterogeneous group and depending on the TME they can share characteristics of different activation states, illustrated here as secondary colors. Adapted from reference #78.

M1 (or classically activated) macrophages are considered pro-inflammatory and respond to IFN $\gamma$  and TLR stimulation, such as lipopolysaccharide (LPS), or tumor necrosis factor (TNF), produced by adaptive immune effector cells by exerting microbicidal and tumoricidal activities<sup>95,96</sup>. IFN $\gamma$ , produced by NK, NKT cells or macrophages, activates STAT1 and interferon regulatory factors (IRF1 and 8), which up-regulate the transcription of cytokine receptor genes (IL-15RA, IL-2RA and IL-6R), cell activation molecules (CD38, CD69 and CD97) and cell adhesion molecules (integrins, mucin 1). LPS binds to TLR4 which signals through myeloid differentiation primary response gene 88 (MyD88), activating transcription factors nuclear factor kappa-light-chain enhancer of activated B cells (NF- $\kappa$ B), activator protein 1 (AP-1) and signal transducer and activator of transcription 5 (STAT5) to induce the production of pro-inflammatory cytokines, chemokines and Ag presentation molecules. They also produce high levels of IL-12 and low levels of IL-10. They contain inducible nitric oxide synthase (iNOS), which leads to bacterial or cell lysis and they have antigen Ag-presenting capacity as indicated by the expression of high levels of MHC-II and co-stimulatory molecule CD86. M1 macrophages are potent producers of chemokines CCL2, CXCL9 and CXCL10<sup>97-99</sup> and pro-inflammatory cytokines IL-1 $\beta$ , IL-6, IL-23 and TNF $\alpha$ . Lately, GM-CSF (granulocyte macrophage colony-stimulating factor) was added to the arsenal of stimuli leading to M1 polarization<sup>100</sup>. GM-CSF signaling lead to the activation and nuclear translocation of STAT5, IRF5 and NF- $\kappa$ B, which ultimately leads to the production of pro-inflammatory cytokines (IL-6, IL-8, G-CSF, M-CSF, TNF and IL-1 $\beta$ ) and up-regulation of molecules associated with Ag presentation, complement- and antibody-mediated phagocytosis and migration, such as CD14, Fc $\gamma$ RIA and CD163. Thus, M1 macrophages drive a so-called Type I immunity through the secretion of mediators that attract Th1 cells, which can be beneficial in the case of tumoricidal or microbicidal activity but can also cause tissue damage<sup>101</sup>. Therefore, they are considered important mediators of host defense but are also common culprits in several autoimmune pathologies (**Figure 3**).

On the contrary, M2 (alternatively activated) macrophages can be elicited by a broader range of stimuli and have thus been subdivided further. More specifically, M2a macrophages typically express the IL-4R $\alpha$ , which is important for their polarization to M2 through binding to cytokines IL-4 and IL-13, primarily produced by eosinophils, basophils, NKT cells and macrophages, leading to STAT6 activation. M2b macrophages bind immune complexes through Fc-receptors expressed on their surface in combination with TLR stimulation or IL-1R ligands. Engagement of Fc-receptors recruits tyrosine kinase Syk, which activates PI3K (phosphoinositide 3-kinase). M2c macrophages are induced by IL-10, TGF- $\beta$  (both potent anti-inflammatory cytokines) or glucocorticoids. IL-10 is produced by macrophages in response to TLR- and glucocorticoid signaling and upon engagement of C-type lectin receptors SIGN-R1 and Dectin 1. IL-10 receptor triggering by its ligand leads to STAT3 activation, driving the expression of Fc-receptors, chemokines CXCL13 and CXCL4 as well as PRRs: formyl-peptide receptor (FPR1), TLR1, TLR8 and MARCO<sup>102</sup>. Tumor-associated macrophages (TAMs) are induced by tumor-derived factors, such as IL-6, LIF (leukemia inhibitory factor) and MCF (macrophage chemotactic factor), and are hard to categorize to one specific M2 category as they vary depending on the TME<sup>97,103,104</sup>. M-

CSF has also been shown to shift macrophages to an M2 polarization. Contrary to M1, M2 macrophages produce low levels of IL-12 and high levels of IL-10, express Ym1 (Chil3) and Fizz1 (Retnl $\alpha$ ) and produce Arginase 1 which inhibits T cell proliferation by depleting arginine from the microenvironment<sup>105</sup>. They promote so called Type 2 immunity which supports wound healing and tumor progression. They express low levels of MHC-II and promote immunosuppression through production of CCL22, which attracts Tregs<sup>106</sup>. They also express programmed death ligand 1 (PD-L1), which antagonizes activated T cells by binding to its receptor programmed death 1 (PD-1) on their surface and inducing apoptosis<sup>107</sup>. They are considered anti-inflammatory and pro-tumorigenic.



**Figure 3. Receptors and ligands involved in macrophage activation and produced mediators**

The main receptor/ligand pairs responsible for M1 activation are IFN $\gamma$ -R, LPS/TL4 and TNF $\alpha$ -R, leading to the production of pro-inflammatory cytokines and cytotoxic mediators. M2 activation is induced by signaling of IL-4 and IL-13 through IL-4R $\alpha$  and IL-33 via ST2, inducing anti-inflammatory factors and regulatory enzymes. Additional receptor/ligand pairs are involved in intermediary activation states that can be encountered depending on the microenvironmental context of activation.

### *Scavenger receptors on macrophages*

Scavenger receptors (SR) are PRRS expressed by phagocytes, with wide ligand specificity for both endogenous and foreign molecules. SRs comprise a range of receptors grouped together based on their ability to bind polyanionic (negatively charged) ligands<sup>108</sup> or based on the fact that they contain a highly conserved version of the Ig domain known as the scavenger receptor cystein-rich (SRCR) domain<sup>109</sup>. Based on their multi-domain structure they have been classified into eight different classes (A-H). Due to the wide range of ligand specificity they are involved in both homeostatic regulation as well as in various inflammatory disease settings and have been extensively studied in the contexts of infection, atherosclerosis and autoimmune disease<sup>110-115 116</sup>.

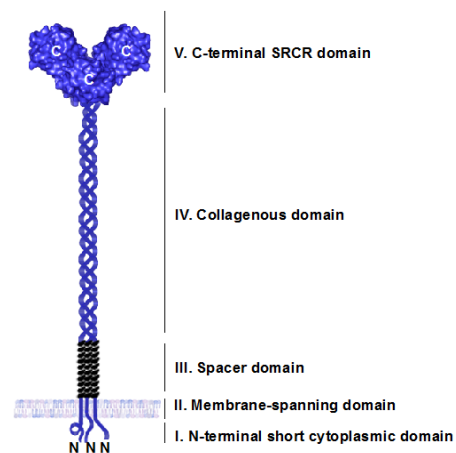
However, an increasing number of studies report the involvement of different scavenger receptors also in cancer<sup>117</sup>. For example, Class A scavenger receptor A (SR-A), which is closely related to MARCO and is expressed on macrophages, has been implicated to promote ovarian and pancreatic cancer<sup>118,119</sup>. Moreover, it has been shown to negatively regulate Ag-specific antitumor immunity by limiting Ag cross-presentation<sup>120</sup>. In a separate study, targeted depletion of SR-A-expressing leukocytes inhibited peritoneal ovarian tumor progression<sup>121</sup>. SR-A can signal via the receptor tyrosine kinase (Mertk), which is important in the uptake of apoptotic cells<sup>122</sup>. Mertk was recently associated to M2 polarization and promotion of anti-inflammatory responses in cancer<sup>123</sup>. This could be linked to increased loads of apoptotic tumor cells in the TME. SR-A deficiency, on the other hand, led to a shift in M1 polarization of TAMs and delayed tumor growth in a lymphoma model<sup>124</sup>.

Another member of the scavenger receptor family, CD163, has been associated with an M2 macrophage phenotype in cancer<sup>125</sup>. CD163 correlates to negative prognosis and poor patient survival in breast cancer<sup>126,127</sup>, adult T cell leukemia/lymphoma<sup>128</sup> and rectal cancer<sup>129</sup>.

The various functions of scavenger receptor MARCO are described in detail below, while consequences of its targeting on macrophages through monoclonal antibodies in the context of sterile- and cancer-related inflammation are discussed in Paper I and II.

### *Scavenger receptor MARCO*

Class A scavenger receptor MARCO was cloned by Karl Tryggvasson at Karolinska Institutet in 1995. MARCO is a membrane bound receptor, which structurally consists of a disulphide-bonded trimer with a partly collagenous structure (**Figure 4**)<sup>130</sup>. The 210kDa protein is composed of three glycosylated subunits and consists of five domains; the N-terminal short cytoplasmic domain I, the membrane-spanning domain II, and domains III, IV and V which are extracellular. Domain III is a 75-residue spacer domain that separates the collagenous domain IV from the plasma membrane.



**Figure 4. Schematic representation of the five domains of the MARCO structure.**

The SRCR domain is adapted from the RCSB Protein data bank. PDB ID: 2OY3 (reference #132)

The collagenous domain contains 270 amino acids forming an overall collagenous structure which is only interrupted at a single site and is involved in trimerization<sup>131</sup>. The interruption by an Ala-Gly-Lys sequence suggests a hinge region in the triple helix of the MARCO molecule. The C-terminal end domain V of MARCO contains six cysteine residues making up the scavenger receptor SRCR domain and proximally the ligand-binding domain<sup>132</sup> (**Figure 4**).

Although structurally closely related to SR-A, MARCO exhibits different regulation<sup>133</sup>. For example, both SR-A and MARCO bind CpG-ODNs. However, MARCO-binding leads to an activating response in macrophages characterized by TLR9-mediated NO production and IL-12 secretion, while SR-A-binding lead to negative regulation of IL-12 responses<sup>134</sup>. Also, in contrast to SR-A expression, which is inducible but not constitutive, expression of MARCO is dependent on TLR4-signaling<sup>135,136</sup>. MARCO is constitutively expressed on distinct subsets of macrophages restricted to certain tissue compartments. MARCO is expressed on the macrophages of the MZ of the spleen (MZM) and medullary regions of the lymph nodes<sup>130</sup>. Its expression on this highly specialized subset of macrophages, located in strategic proximity to DCs and B cells in the marginal zone, highlights MARCO as an important receptor for the capture and clearance of antigens and apoptotic cells from the blood. Inability to clear apoptotic cells from the circulation is associated with increased risk for developing SLE<sup>137,138</sup>. Also, mice lacking MARCO, have a defective MZ architecture, show impaired responses to T cell-independent (T-I) type 2 Ag, produce elevated levels of autoantibodies to DNA and develop symptoms of SLE<sup>71,139</sup>. Additionally, autoantibodies to MARCO have been found early on in mouse models for SLE before the onset of disease and the receptor is down-regulated on MZMs<sup>140</sup>. Autoantibodies to MARCO have similarly been found in SLE patients however it is unclear whether they are involved in driving the disease<sup>71</sup>.

As for most SR, its ligands are negatively charged polyanionic molecules. More specifically, MARCO has been shown to bind low density lipoprotein (LDL) through its SRCR domain<sup>141</sup>, as well as bacterial agents such as *Esheria coli*, *Staphylococcus aureus*<sup>130</sup> and LPS<sup>142</sup>, but also un-opsonized particles, such as titanium dioxide (TiO<sub>2</sub>), ferric oxide (Fe<sub>2</sub>O<sub>3</sub>), silica (CsiO<sub>2</sub>) and latex beads<sup>64,143-145</sup>.

The fact that MARCO has a very short intracellular domain suggests that the receptor requires a signaling partner to relay its downstream effects. Although little is known about the exact mechanism of action and interaction partners it is clear that MARCO has a role in modulating inflammatory responses. Several studies have attempted to elucidate this by studying its function on macrophages. More specifically, it was shown that cooperation of MARCO, TLR2 and CD14 induces macrophage cytokine responses to *Mycobacterium tuberculosis*<sup>146</sup>, while MARCO co-operation with TLR2 and nucleotide binding oligomerization domain-containing 2 (NOD 2) is important for clearance of *Streptococcus pneumoniae*<sup>147</sup>. In alveolar macrophages, uteroglobin-related protein 1 (UGRP1) was identified as a ligand for MARCO<sup>148</sup>. Moreover, it has been suggested that MARCO is negatively regulated by FcR $\gamma$  through the recruitment of Src homology region 2 domain-containing phosphatase 1 (SHP-1) during *Esheria coli* binding to Fc $\gamma$ RIII, resulting in

decreased phagocytosis and enhanced production of TNF $\alpha$  <sup>149</sup>. Also, ligand internalization by MARCO decreases surface-sensed TLR4 responses, while simultaneously enhancing intracellular TLR3, NOD2 and NACHT, LRR and PYD domains-containing protein 3 (NALP3, inflammasome) <sup>150</sup>. Similarly, MARCO has been shown to modulate TLR-induced DC activation, possibly providing a bridge between innate and adaptive immunity <sup>151</sup>. While MARCO is important for the clearance of *Pneumococcus pneumoniae* infection <sup>152</sup>, it has a deleterious role mediating uptake of the intracellular parasite *Leishmania major* <sup>153</sup> and suppressing early inflammatory responses to *Influenza A* viral infection <sup>154</sup> as well as facilitating HSV-1 infection and spread <sup>155</sup>.

Although primarily a MZM receptor, MARCO can be rapidly up-regulated on other macrophages and DCs after activation by bacterial products <sup>156,157</sup>. LPS binding to TLR4 leads to up-regulation of both MARCO (and SR-A, in both an MyD88-dependent and independent ways, and leads to B cell activation <sup>158</sup>. Up-regulation of MARCO expression is associated with cytoskeletal rearrangements <sup>159</sup>, which induce drastic morphological changes to cells, such as the formation of lamellipodia and dendritic-like processes. These result in loss of cell adhesion and decreased migration, processes that are necessary for the engulfment of particles <sup>160</sup>. Another study by Grolleau *et al.* reported that the inducible up-regulation of MARCO on DCs after pulsing with tumor cell lysate is associated with increased phagocytic capacity <sup>161</sup>. Here, targeting MARCO with a monoclonal antibody led to enhanced DC motility and anti-tumor activity in a mouse model of melanoma <sup>162</sup>. In a follow-up study, it was shown that MARCO deficient tumor-pulsed DCs responded to CCL21 with an increased migratory capacity to draining lymph nodes of B16 melanoma tumors, leading to improved anti-tumor IFN $\gamma$  T cell responses <sup>163</sup>. Similarly, lack of MARCO led to enhanced DC migration to lymph nodes and increased production of pro-inflammatory mediators in the context of airway inflammation <sup>164</sup>.

Human MARCO was recently identified and found to be highly similar to the mouse molecule <sup>165</sup>. Studies on MARCO expression in humans show a wider distribution pattern of the receptor on several tissue macrophage subsets. However it should be noted that expression of the receptor was characterized on samples from septic patients and may therefore not be representative of the steady state <sup>166</sup>. Although extensively studied in infection and autoimmunity, the role of MARCO in cancer still remains poorly understood. In the context of cancer, the role of MARCO is so far contradictory, correlating with better prognosis in follicular lymphomas <sup>167</sup>, but with poor prognosis in human breast cancer <sup>168</sup>. MARCO has been suggested to be expressed on a subpopulation of macrophages in the tumor stroma with immunosuppressive activity, however its role is unknown <sup>169</sup>. Paper II describes an attempt to shed light on the so far diffuse role of MARCO in cancer. Data acquired from other physiological or pathophysiological settings so far pinpoint MARCO as an important regulator of responses to both self- and exogenous antigens and systemic inflammation <sup>170</sup>.

## 1.2 THE IMMUNOSUPPRESSIVE TUMOR MICROENVIRONMENT

Cancer is a multifactorial disease where a number of mechanisms converge to initiate, sustain and promote malignancy. The hallmarks of cancer progression include evasion of apoptosis, genome instability and mutation, sustained angiogenesis, limitless replicative potential, self-sufficiency in growth signals, insensitivity to growth inhibitors, avoiding immune destruction, as well as invasion and metastasis. However, tumor cells do not act alone in the processes of tumorigenesis, tumor progression and metastasis. They are rather part of a permissive microenvironment established by the concerted actions of different cell types and stromal components, such as cancer-associated fibroblasts, pericytes, myeloid cells, lymphocytes, endothelial cells and extracellular matrix <sup>171</sup>. Recently, tumor-promoting inflammation was recognized as the 10<sup>th</sup> hallmark of cancer development <sup>172,173</sup>. It is generally characterized by the presence of inflammatory cells and mediators (e.g. chemokines, cytokines) in the tumor stroma, tissue remodeling, angiogenesis and tissue repair. The key cellular players in cancer-related inflammation are the tumor-infiltrating leukocytes (TILs); cells of the myeloid lineage exhibiting a great degree of plasticity and diversity, such as TAMs and the related myeloid-derived suppressor cells (MDSCs), tumor-associated neutrophils (TANs), mast cells and eosinophils <sup>98</sup>. TILs are present in most tumors to varying degrees. Although initially the presence of TILs was considered beneficial, accumulating evidence suggested that tumor-derived factors imposed phenotypic and functional changes on TILs, which altered them to in fact promote tumorigenesis.

TAMs, TANs and the related MDSCs, but also T regulatory cells (Tregs) are responsible for imposing a generalized immunosuppressive milieu within the tumor stroma <sup>174-176</sup>. Immunosuppression is achieved through a variety of mechanisms, such as up-regulation of cell surface receptors, secretion of cytokines and chemokines, and different enzymatic activities.

Shimon Sakaguchi first identified T<sub>regs</sub> as a naturally occurring subset of CD4<sup>+</sup> T cells. They are characterized as CD4<sup>+</sup> CD25<sup>hi</sup> and rely on the transcription factor FoxP3 for their development. Other surface markers that are typically expressed on T<sub>regs</sub> include CTLA-4, glucocorticoid-induced TNF receptor (GITR) and lymphocyte activation gene-3 (LAG-3). T<sub>regs</sub> are recruited to tumors through the CCR4/CCL17, CCL22, CCR8/CCL1 and CCR6/CCL20 chemokine axes to promote suppress anti-tumor immunity <sup>177</sup>. Moreover, tumors promote T<sub>reg</sub> expansion by producing TGF- $\beta$ . Studies have shown that large numbers of Tregs in the tumor, but not in the draining lymph nodes, inversely correlate with survival of patients with ovarian carcinoma. Therapeutic approaches targeting Tregs for elimination, such as targeting GITR or CD25, have shown promising results heaving immunosuppression and unleashing anti-tumor CD8<sup>+</sup> T cells <sup>178,179</sup>.

Although less explored than TAMs in the context of cancer, tumor-associated neutrophils (TANs) are also important mediators of cancer-related inflammation and tumor progression <sup>180</sup>. TANs are attracted to the inflammatory TME by chemokines, such as tumor-derived CXCL8 and epithelial cell-derived CXCL1, 2 and 3. On site, depending on the local cytokine milieu they can be polarized to an N1 anti-tumorigenic phenotype with the ability



to kill tumor cells and block metastasis or to an N2 pro-tumorigenic phenotype <sup>181</sup>. Blockade of TGF- $\beta$ -signaling inhibits N2 polarization <sup>182</sup>. While primarily thought to originate from granulocytic bone marrow precursors, they have also been shown to accumulate in the spleen of tumor-bearing mice, from where they are later deployed to the tumor <sup>183</sup>. N2 TANs have a variety of immunomodulatory functions, which can have profound implications on anti-tumor immunity. These include but are not limited to, induction of a tolerogenic DC phenotype <sup>184</sup>, affecting NK cell development <sup>185</sup> as well as B cell survival and maturation <sup>186</sup>.

MDSCs are a group of heterogeneous immature myeloid precursors with profound capacity to suppress effector T cells responses. Under physiological conditions they are present in very few numbers and rapidly differentiate into granulocytes, macrophages and immature DCs. Normally they account for 20-30% of all cells in the bone marrow, and 2-4% in the spleen, while they are absent from lymph nodes. However, pathological inflammation caused by cancer, infection, autoimmune disease or trauma, halts their differentiation and induces expansion of the immature cells with immunosuppressive capacity. They are subdivided into monocytic (Mo-MDSC) and polymorphonuclear (PMN-MDSC), with distinct functions in tumors. Mo-MDSC represent ca 20-30%, while PMN-MDSC are 70-80% of the total MDSCs in a tumor. In mice, Mo-MDSCs are characterized as CD11b<sup>+</sup>Gr-1<sup>+</sup>Ly6C<sup>high</sup>Ly6G<sup>low</sup>, while PMN-MDSCs are CD11b<sup>+</sup>Gr-1<sup>+</sup>Ly6C<sup>low</sup>Ly6G<sup>high</sup>. MDSCs employ various mechanisms to impose immunosuppression and these may differ between the periphery and the tumor site. The suppressive capacity of PMN-MDSCs is mediated by Arg1, while for Mo-MDSC iNOS production is more important <sup>187</sup>. They have been shown to produce chemokines, such as CCL3, CCL4, and CCL5, to attract CCR5<sup>+</sup> T<sub>regs</sub> in a mouse melanoma model <sup>188</sup>. MDSC can also induce the de novo generation of T<sub>regs</sub> through production of IL-10 and TGF- $\beta$ . <sup>189</sup>. In lymphoma MDSCs induced Tregs through Arg1 and Ag presentation <sup>190</sup>. The importance of heaving immunosuppression to achieve a successful immunotherapeutic effect in cancer treatment is becoming more and more apparent. Therefore, strategies to eliminate, block the expansion and recruitment or differentiate MDSCs have become the subject of numerous investigations.

### **1.2.1 Tumor-associated macrophages**

Already in 1863 Virchow described the presence of infiltrating leukocytes in tumors and proposed a link between cancer and inflammation. Indeed, as many as 25% of all cancers are thought to arise as a consequence of chronic smouldering inflammation <sup>191-195</sup>. TAMs are a major component of tumor-promoting inflammation and increased numbers have been associated with poor prognosis in a variety of human cancers, and breast cancer in particular <sup>127,196</sup>. In breast cancer, high infiltration of TAMs has been associated with hormone receptor negative status, including triple negative tumors, which is the type of breast cancer with the poorest clinical outcome <sup>197-199</sup>. However, our understanding of the distinct phenotypes of TAMs, their distribution in the tumor stroma and the connection to pathological outcome is still limited, thus further investigation is warranted.

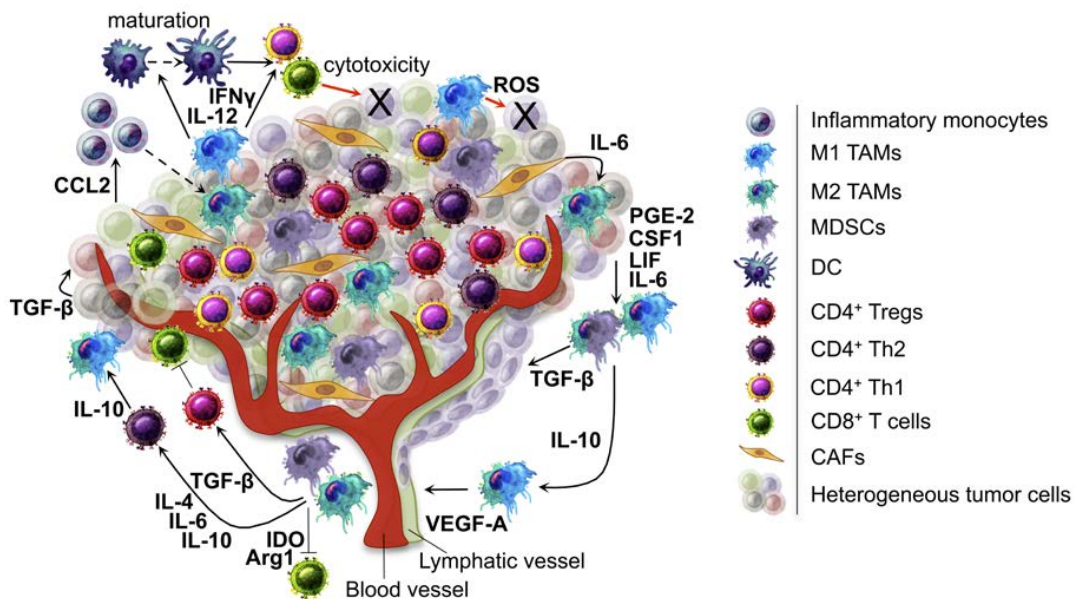
The origin of TAMs is still disputed and several different sources, known to give rise to different macrophage subpopulations with distinct functions have been suggested over the years (embryonic yolk sac, bone marrow, spleen). In experimental models of mammary carcinoma and lung adenocarcinoma models, most TAM populations found in the tumors originated from Ly6C<sup>+</sup> circulating monocytic precursors<sup>200</sup>. However, there is considerable debate regarding the origins of the monocytic precursors, some supporting the idea that they originate from the bone marrow and others suggesting that they arise from the extramedullary hematopoiesis in the spleen<sup>183</sup> that gives rise to an easily mobilized reservoir of pro-inflammatory monocytic precursors. However, the contribution of the latter compartment seems to be minor<sup>201</sup>. Finally, Mo-MDSC and PMN-MDSC are able to differentiate into mature myeloid cells; macrophages and neutrophils respectively. However, in cancer they are skewed to a pathological phenotype, which can give rise to TAMs or TANs<sup>202,203</sup>.

TAMs are recruited to the inflammatory TME, where they are sequestered by tumor-derived factors, cytokines and chemokines produced by other inflammatory cells and by hypoxia, all together contributing to a pro-tumorigenic inflammatory microenvironment<sup>204,205</sup>. Tumors secrete pro-inflammatory mediators, such as IL-6 and TNF, which contribute to the conditioning of the inflammatory TME<sup>195</sup>. This induces the production of chemokines (CCL2, CCL5, CCL22, CXCL1, CXCL8, CXCL12 and MCP) by various immune cell types that will attract macrophages to the inflammatory tumor site and promote their production of immunosuppressive cytokines, such as IL-10 and CSF-1, as well as up-regulate the expression of scavenger receptors SR-A and CD163<sup>99,119</sup>. In addition, tumor cell and TAM crosstalk through CCR2 and CX3CR1 further drives tumor progression and metastasis<sup>206,207</sup>. Moreover, tumor-derived prostaglandin E2 and TGF- $\beta$  promote a distinct M2 TAM phenotype, the major regulator of which is NF- $\kappa$ B, the downstream signal transducer of the TLR4-MyD88 signaling pathway<sup>208</sup>. This generates a self-propagating circle of pro-tumorigenic inflammation<sup>209</sup>.

T<sub>regs</sub> in particular, are potent inducers of TAMs as they produce abundant amounts of IL-10, which promotes the expression of CD163 and CCL18, suppresses the production of pro-inflammatory cytokines and expression of MHCII<sup>210</sup>. CD4<sup>+</sup> T<sub>h</sub>2 helper cells have been implicated in the promotion of M2 TAMs through the production of IL-4<sup>211</sup>. Also, it has been shown, both *in vitro* and *in vivo* that B cells producing IL-10 profoundly affect the phenotype of macrophages shifting them to M2 polarization<sup>212,213</sup>. Moreover, B cells can remotely control macrophages through their production of Abs to tumor-related proteins. These Abs form immune complexes, which are taken up by macrophages through Fc-receptors, resulting in M2 polarization<sup>214</sup>. MDSCs, physically interact with TAMs and suppress macrophage-derived IL-12 in an IL-10-dependent manner, leading to enhanced immunosuppression<sup>215</sup>. As a result, TAMs evolve into different specialized subsets with distinct functions, that ultimately support tumor growth and progression through various mechanisms<sup>119,169,216-218</sup>.

Upon recruitment to the inflammatory TME, TAMs produce factors that support tumor cell proliferation, neovascularization and metastasis<sup>219</sup> (**Figure 5**). They have been found to

assist invasion of the primary tumor into the surrounding tissue but also intravasation and further dissemination of tumor cells. This molecular polarization is reflected in the acquisition of a distinct gene expression signature of M2-like TAMs<sup>220-222</sup>. These have an anti-inflammatory phenotype; suppressing adaptive immunity (TGF- $\beta$ , IL-10)<sup>174,200</sup>, promoting tumor growth (EGF), driving the EMT of invading tumor cells (TGF- $\beta$ )<sup>223</sup>, tissue remodeling (MMP, cathepsin proteases)<sup>224</sup> and angiogenesis (VEGF; vascular endothelial growth factor). TAMs are primarily of M2-like phenotype, but some can have a pro-inflammatory M1-like phenotype, which can elicit potent anti-tumor inflammatory responses. As for macrophages, the M1/M2 polarization axis is however a mere simplification. In reality, tumors are complex heterogeneous tissues and therefore different regions of the tumor may have distinct microenvironments. As a result, the macrophage infiltrate may vary greatly with regards to phenotype and function depending on their localization. Another level of complexity is added depending on the type of cancer and the permissiveness of its microenvironment. Consequently, macrophage plasticity and diversity create a dynamic environment which evolves with tumor progression, where specialized subpopulations of macrophages support different functions<sup>225</sup>. The major functions of the different TAM subsets are described below.



**Figure 5. Function of TAMs and MDSCs in the immunosuppressive tumor microenvironment**

Heterogeneous tumor cells in the TME produce various factors that promote immunosuppression. For example, tumor-derived IL-6, LIF, PGE-2 and CSF1, drive the differentiation of MDSCs to M2 TAMs and the polarization of M1 TAMs to M2 TAMs. In turn, MDSCs and M2 TAMs secrete TGF- $\beta$  and pro-angiogenic factors that drive the angiogenesis and migration of metastatic tumor cells. The immunosuppressive phenotype of M2 TAMs are further sustained by tumor-derived and autocrine TGF- $\beta$  and CAF-derived IL-6. MDSCs and M2 TAMs produce TGF, driving T<sub>regs</sub> expansion and inhibition of CD8<sup>+</sup> T cell anti-tumor responses. They also typically express the enzymes IDO and Arg1, which have a profound inhibitory effect on CD8<sup>+</sup> T cells. Moreover, MDSCs and TAMs secrete IL-4, IL-6 and IL-10, which drive T<sub>H2</sub> T cell differentiation. CD4<sup>+</sup> T<sub>H2</sub> cells augment the anti-inflammatory cytokine IL-10 leading to further M1 to M2 TAM polarization. Additionally, tumor-derived chemokine CCL2 attracts inflammatory monocytes to the TME, providing a replenishing source of TAM precursors. Altogether, these mechanisms establish an immunosuppressive TME and inhibit the production of ROS and pro-inflammatory IL-12 and IFN $\gamma$  by M1 TAMs, leading to diminished cytotoxic activity against the tumor.

#### 1.2.1.1 TAMs in chemoresistance and angiogenesis

In a recent study, DeNardo *et al.* highlighted the prognostic value of a certain TAM-related tumor signature (high TAM count, high CD4<sup>+</sup> T cell count and low CD8<sup>+</sup> T cell count) and associated that to poor response to chemotherapy in breast cancer patients<sup>226</sup>. This could be due to chemotherapy-induced CSF1 release by the tumor which leads to macrophage recruitment, as has been shown to be the case in mouse mammary tumors, or it could be related to the immunosuppressive effect of TAMs on CD8<sup>+</sup> T cells. Moreover they found that CSF-1-targeting only depleted macrophages in poorly vascularized areas of the tumors. Hypoxia, is known to drive the recruitment of angiopoietin 2 receptor (Tie2) -expressing macrophages<sup>227,228</sup>. Tie2<sup>+</sup> macrophages up-regulate transcription factors HIF-1 $\alpha$  and -2 $\alpha$ , which control VEGF-A production and suppress T cell function<sup>229-231</sup>. TAMs are known to have an angiogenic function, which leads to the formation of poorly perfused leaky vessels<sup>204,225,232,233</sup>. Therefore, the authors speculate that by depleting those macrophages the overall tumor vasculature is normalized, allowing for better delivery of the chemotherapeutic agent<sup>232,234</sup>.

#### 1.2.1.2 TAMs in tumor cell invasion and metastasis

The process of metastasis involves several steps, during which tumor cells invade the surrounding normal tissues and extracellular matrix (ECM), intravasate into the blood or lymphatic vessels and travel to distal sites where they extravasate and seed micrometastatic niches. Recent studies demonstrate that TAMs originate from circulating inflammatory monocytes expressing chemokine receptor CCR2, which respond to tumor-derived CCL2 by migrating to the site of tumor inflammation<sup>235</sup>. There they differentiate into TAMs lining the invasive front of tumor cells and assist many of the afore-mentioned processes through the production of a wide array of mediators. It has been shown that colony-stimulating factor 1 (CSF-1), an important factor not only for macrophage differentiation, but also for promoting tumor invasion and metastasis is produced by TAMs, which in turn drives the production of the tumor attractant epidermal growth factor (EGF). Additionally, macrophage inhibitory factor (MIF) promotes tumor cell invasiveness and trafficking. TAMs also produce platelet-derived growth factor (PDGF) to support tumor cell proliferation. Moreover, TAMs produce matrix-degrading enzymes such as matrix metalloproteases (MMPs), cathepsins and proteases, which break down ECM and basement membrane to facilitate the invasion of tumor cells into the surrounding normal tissue. The detrimental role of TAMs in tumor progression has also been shown in macrophage deficient mice, where metastatic disease could be avoided in their absence. Alternatively, metastasis could be abrogated by inducing the polarization of M2-like TAMs to an M1-like phenotype. Finally, it has been shown that macrophages are important for “conditioning” distal sites for the seeding of micrometastases. Thus, targeting M2 TAMs in the TME could limit tumor metastasis (Paper II)<sup>236</sup>.

#### 1.2.1.3 TAMs and inhibition of antitumor immunity

In order to inhibit T cell mediated anti-tumor immunity, TAMs up-regulate the expression

of ligands for the inhibitory receptors programmed cell death protein (PD-1 or B7-H1) and cytotoxic T-lymphocyte Ag 4 (CTLA-4), on their surface. Under physiological conditions these molecules function as safety switches expressed on activated T effector cells when the need to resolve inflammation occurs. Engagement of PD-1 and CTLA-4 through their respective ligands (PD-L1/-L2 and CD80/CD86), inhibits effector cells from exerting cytotoxic functions through induction of apoptosis. PD-L1 is constitutively expressed on immune cells and is up-regulated on TAMs by IL-10 and TNF $\alpha$  <sup>107</sup>. In contrast, PD-L2 is exclusively expressed on APCs. Its expression on monocytes and macrophages is induced by CSF-1, IL-4 and IFN $\gamma$  <sup>237</sup>. Both PD-L1 and PD-L2 are up-regulated by TAMs and MDSCs <sup>238,239</sup>. Another such molecule that is expressed by TAMs and has a suppressive effect on the function of T cells by arresting cell the cycle is B7-H4, which is induced in macrophages by IL-6 and IL-10 <sup>240</sup>.

Lack of co-stimulation is another mechanism TAMs employ to render T cells non-responsive. CD86 is constitutively expressed on APCs in low amounts and is up-regulated upon activation, while CD80 is expressed only upon APC activation. CD80 and CD86 are expressed on M1 TAMs <sup>241-243</sup> but are down-regulated on M2 TAMs as a result of hypoxia <sup>244</sup> and by their interaction with MDSCs <sup>245</sup>.

As previously mentioned, M2 TAMs are potent producers of IL-10 but have very low levels of IL-12, an essential cytokine for CD8<sup>+</sup> cytotoxic T cells activation, NK cell tumoricidal activity <sup>246</sup> and DC Ag presentation <sup>247</sup> (**Figure 5**). As a result, in the absence IL-12, the TME is skewed towards a pro-tumorigenic Th2 immune response, where IL-10 and TGF- $\beta$  impose generalized immunosuppression. IL-10 leads to the expansion of natural T<sub>regs</sub> and in combination with TGF- $\beta$  induces the up-regulation of FoxP3 and the generation of induced T<sub>regs</sub>, which in turn further suppress CD8<sup>+</sup> T cell function <sup>248</sup>. TGF- $\beta$  inhibits CD8<sup>+</sup> cytotoxic function in vivo <sup>249,250</sup> and Th1/Th2 CD4 functions by interfering with lineage-determining transcription factors <sup>251,252</sup>. Additionally, TAMs secrete chemokine CCL22, which recruits CCR4<sup>+</sup> T<sub>regs</sub> <sup>106</sup> and CCL20 that attracts CCR6<sup>+</sup> T<sub>regs</sub> <sup>253</sup>. TAMs also employ the enzymatic activity of arginase (encoded by the *Arg1* gene), a hallmark of M2 polarization in itself, to suppress T cells by depleting L-arginine, which is essential for the re-expression of the CD3 $\zeta$  chain after TCR down-regulation upon activation <sup>254,255</sup>.

An additional mechanism utilized by TAMs is regulation through the enzyme indolamine 2,3-dioxygenase (IDO). IDO depletes tryptophan from the environment thus leading to cell cycle arrest and cessation of T cell proliferation and effector functions <sup>256</sup>. Alternatively, IDO can be expressed by APCs, which capture and present tumor Ag in draining lymph nodes to naïve T cells, leading to tolerization against the tumor <sup>257,258</sup>. Additionally, tryptophan breakdown by-products, such as kynurenine, are also cytotoxic to T cells, which might also be a reason for inhibition of T cell proliferation.

### **1.3 EPITHELIAL-MESENCHYMAL TRANSITION IN TUMOR METASTASIS**

Malignant invasion requires tumor cells to undergo EMT, a process recently shown to be assisted by M2 TAMs <sup>259-261</sup>. EMT is a biological process during which a polarized

epithelial cell loses its apical-basal polarity, loses contact to the basement membrane and acquires a mesenchymal phenotype associated with increased migratory, invasive and anti-apoptotic properties. EMT is absolutely essential for several physiological and pathophysiological processes, such as implantation, embryogenesis and organ development (Type 1 EMT), tissue regeneration and fibrosis (Type 2 EMT), as well as tumor growth and cancer progression (Type 3 EMT). EMT is governed by the activation of a highly preserved distinct genetic pattern, including *Snail/Slug*, *Twist*, *Six1*, *Cripto*, *Tgf-b*, and *Wnt/ $\beta$ -catenin*, across different species<sup>262</sup>.

In cancer progression, tumor cells undergoing EMT are found at the invasive front of the tumor and are thought to drive metastasis through intravasation into blood and lymphatic vessels, extravasation and seeding of micrometastases at distal sites, at which point the tumor cells revert to a mesenchymal-to-epithelial transition (MET). However, not all cancer cells can undergo EMT and of the ones that will, not all will survive to seed distal metastases. An interesting conundrum in the case of breast cancer is that, while only a few cancer progenitor cells can give rise to metastases, a high genetic diversity is observed in metastatic breast cancer cells<sup>263</sup>. This discrepancy was explained by the fact that heterogeneity in cancer is not the result of random genetic aberrations, but is rather orchestrated by the evolutionarily conserved genetic programs that make up EMT<sup>264,265</sup>.

TGF- $\beta$  has been implicated as a major inducer of EMT in cancer through at least two pathways. Of importance for this thesis is the p38 mitogen activated protein kinase (MAPK)-mediated autocrine TGF- $\beta$ -induced EMT, which leads to preferential dissemination of tumor cells through the lymphatics in a targeted manner<sup>266</sup>. Besides being produced by tumor cells, TGF- $\beta$  can be readily produced by several cell types in the inflammatory TME, including TAMs, tumor-associated DCs, MDSCs and T<sub>regs</sub><sup>267</sup> (**Figure 5**). As previously mentioned TGF- $\beta$  is considered a master regulator of the processes leading up to the conditioning of the TME and the transitioning of the tumor cells for metastatic spread. An EMT permissive TME is similar to a wound unable to heal, characterized by increased enzymatic activity, matrix remodeling, inflammation, pathological tissue regeneration and scarring<sup>268</sup>. The conditioning effect of TGF- $\beta$  on the TME relies on imposing immunosuppression by affecting the recruitment, polarization and production of inflammatory mediators by immune cells<sup>269</sup>. Tumor-derived TGF- $\beta$  is known to attract and polarize macrophages and neutrophils to M2 and N2 activation states respectively. This loop is sustained by the induction of TGF- $\beta$  production by M2 TAMs. TAMs have been pinpointed as the main drivers of tumor progression, as they assist invasion from the primary tumor into the surrounding tissue by the use of enzymes that specialize in breaking down extracellular matrix (matrix metalloproteases, MMPs), suppress anti-tumor immune responses, produce pro-tumorigenic growth factors and mediators and promote EMT in a NF- $\kappa$ B-dependent manner<sup>223,270</sup>. TGF- $\beta$  further promotes the overall immunosuppressive milieu by shifting recruited CD4<sup>+</sup> precursors to Th2 responses and by inducing the de novo generation of T<sub>regs</sub> through the up-regulation of FoxP3<sup>271,272</sup>. Together, an increasing amount of evidence supports a central role for TAMs

in sustaining and promoting TGF- $\beta$ -driven EMT and facilitating tumor cell metastasis<sup>273</sup>. During inflammation DCs are capable of migrating to lymph nodes through the lymphatics in response to chemotactic gradients of molecular cues in order to initiate immune responses<sup>274</sup>. In study III we investigate the link between TGF- $\beta$ -induced tumor cell EMT and the acquisition of an immune cell-like phenotype, which directs lymphatic dissemination and distal metastasis of tumor cells.

#### **1.4 CANCER IMMUNOTHERAPY**

The field of cancer immunotherapy was born in 1891, when William Coley, a surgeon at the New York Cancer Hospital (currently Memorial-Sloan Kettering Cancer Center), injected live or inactivated bacteria in order to mimic the spontaneous regression observed in sarcoma patients who had experienced infection by *Streptococcus pyogenes*<sup>275,276</sup>. The theory that “Coley’s toxins” activated Metchnikoff’s phagocytes to induce inflammation and kill bystander tumor cells was gaining momentum amongst immunologists, albeit not amongst clinical oncologists who turned to the more traditional treatments options of surgery, radio- and chemotherapy.

Harnessing but also unleashing the natural resources of the immune system for the benefit of cancer patients has long been the goal of vigorous research in the field of cancer immunotherapy<sup>277</sup>. The design and combination of sophisticated therapeutics, which overcome tolerance mechanisms and immunosuppression to induce anti-tumor responses incorporating a long-term memory component, is the goal to successful treatment of metastatic disease. Passive immunotherapy approaches consisting of Abs or adoptive transfer of donor effector T cells rely on the induction of an immediate therapeutic effect without the need for prior activation. Active immunotherapy, on the other hand, presents much greater challenges.

Active immunotherapy includes several prerequisites. DCs need to take up tumor-specific Ag against which central or peripheral tolerance has not been established. Upon Ag encounter the DC would also have to receive a second co-stimulatory signal, which would allow it to mature into a professional APC. After maturation DCs migrate to draining lymph nodes where they present the tumor-antigenic peptides in order to elicit Ag-specific T effector responses. DCs may also induce antibody responses, or activate NK and NKT cells. Finally, activated lymphocytes need to exert their function at the tumor site, at which point the suppressive inflammatory tumor environment provides a new level of complexity to the already daunting challenges. Therefore, attempts to simultaneously activate immune effectors and modulate components of the inflammatory TME into a less immunosuppressive state are of great importance for successful immunotherapy.

##### **1.4.1 Immunotherapy using monoclonal antibodies**

In 1975 Milstein and Köhler successfully fused B cells from the spleen of an immunized mouse with a myeloma cell line and generated an immortal antibody-producing cell line that produced large amounts of mAbs against the specific Ag<sup>278</sup>. The discovery of the hybridoma

technique was going to revolutionize life science research. To understand the different modes of action and potential of mAbs I have included a brief introduction into the basic molecular aspects of antibodies and Fc receptors.

Abs or Igs are divided into five classes based on the sequence of the heavy chain constant regions; IgA, IgD, IgE, IgG and IgM. Abs are structurally composed of two identical heavy (H) and two identical light (L) chains, which interact through disulfide bonds to form a Y-shaped molecule. They consist of two functional subunits, the Fab (Fragment, Ag binding) domain, which is Ag specific, and the Fc (Fragment, crystallizable) domain, which is responsible for mediating effector functions through its interaction with Fc receptors or complement. The variable region of the Fab domain contains three hypervariable complementarity-determining regions (CDRs), which form the Ag-binding site. The most common antibody class used in cancer immunotherapies is the IgG, which can be further subdivided into subclasses/isotypes IgG1-IgG4 in humans and IgG1, IgG2a, IgG2b and IgG3 in mice. These bind different FcRs with varying affinities and specificities. Abs undergo post-translational modifications during which they are differently glycosylated. This has implications for the effector functions mediated by the Fc part as the glycan alters its binding affinity to FcRs.

There are two types of FcRs, type I FcRs are transmembrane glycoproteins of the Ig superfamily and include Fc $\gamma$ Rs, while type II FcR belong to the c-type lectin family and include DC-SIGN/SIGN-R1 (CD209) and CD23. Depending on the structural conformation of the Fc, Abs show a particular preference for either Type I or Type II FcRs. Of particular interest for this thesis are type I FcRs, which will therefore be described in more detail. Upon antibody binding, activating Fc $\gamma$ Rs transduce signals through immunoreceptor tyrosine-based activation motifs (ITAMs) and inhibitory Fc $\gamma$ Rs through immunoreceptor tyrosine-based inhibitory motifs (ITIMs). In mice there are three activating Fc receptors, namely Fc $\gamma$ RI, Fc $\gamma$ RIII and Fc $\gamma$ RIV and one inhibitory, Fc $\gamma$ RIIB. In humans the activating Fc $\gamma$ RI (CD64), Fc $\gamma$ RIIA (CD32A), Fc $\gamma$ RIIC (CD32C), Fc $\gamma$ RIIIA (CD16A), Fc $\gamma$ RIIIB (CD16B) and the inhibitory Fc $\gamma$ RIIB (CD32B) are present. Fc $\gamma$ R are expressed on myeloid cells, such as monocytes, macrophages, DCs, basophils and mast cells, which can express both activating and inhibitory Fc $\gamma$ Rs. More specifically, monocytes and macrophages express all Fc $\gamma$ Rs, neutrophils mainly express the inhibitory Fc $\gamma$ RIIB and the activating Fc $\gamma$ RIII and Fc $\gamma$ RIV, while DCs express Fc $\gamma$ RI, Fc $\gamma$ RIIB and Fc $\gamma$ RRIII. In the lymphoid lineage, NK cells, express only the activating receptor Fc $\gamma$ RIII, whereas B cells only express the inhibitory receptor Fc $\gamma$ RIIB. Studies on the therapeutic effects and the mechanisms of action of Abs have become more informative after the development of a mouse model in which murine FcRs have been replaced by their human counterparts, faithfully recapitulating the human FcR-expression profile on hematopoietic cells<sup>279</sup>. Ab-mediated inflammatory signaling through FcRs has been implicated in the physiological processes of Ag presentation and B cell selection, but also in the pathophysiological contexts of infection, autoimmune disease, and cancer. Growing understanding of the underlying mechanisms governing these responses has led to advances in the generation of antibody therapeutics, such as the optimization of Ab Fc-



design, the customized targeting to particular FcR and enhancement of the binding affinity of Abs by introduction of Fc modification, in order to modulate antibody effector functions to clinical benefit. Such approaches have been utilized in vaccinations against infectious diseases and the development of cytotoxic or immunomodulatory Abs for the treatment of autoimmune and neoplastic diseases<sup>280</sup>.

#### *1.4.1.1 Monoclonal antibody therapeutics for cancer*

The use of monoclonal antibodies in cancer therapy can be direct, targeting molecules expressed by the tumor itself known as tumor Ags, blocking signaling cascades that are important for tumor survival and progression, inducing apoptosis or immunogenic cell death<sup>281</sup>. Tumors have, however, evolved to overcome such blockades, for example by down-regulating the expression of the targeted tumor-associated Ags (TAA) or selecting clones lacking TAA expression<sup>282</sup>. Thus, the need for immunomodulatory Abs targeting the immunosuppressive TME, aiming to evoke long-lasting anti-tumor immune responses emerged. These approaches aim to bridge innate and adaptive effector mechanisms and to establish immunological memory against the tumor<sup>283</sup>.

These complex processes boil down to the following basic modes of action for Ab therapeutics used in anti-cancer therapies: i) Abs that recognize TAA and target tumor cells for elimination by Ab-dependent cellular cytotoxicity (ADCC) or Ab-dependent cellular phagocytosis (ADCP), ii) Abs stimulating receptor activity, inducing downstream signaling of receptors expressed on the surface of tumor cells or accessory cells in the TME, iii) Abs blocking receptor signaling, iv) agonistic immunomodulatory Abs targeting activation molecules on immune effector cells to induce anti-tumor responses, and v) antagonistic immunomodulatory Abs blocking inhibitory pathways in order to unleash anti-tumor immune responses. The last are known as checkpoint immunotherapies and have received immense recognition for their therapeutic effects, revolutionizing the field of cancer immunotherapy.

#### *Antibody-dependent cellular cytotoxicity/-phagocytosis*

One of the main functions of immunotherapeutic Abs is to exert cytotoxic or phagocytic activity by engaging secondary immune effector mechanisms. This is mediated through ADCC or ADCP, processes during which Abs bound to their ligands interact with activating FcR on innate immune effector cells (most commonly NK cells, eosinophils, monocytes and macrophages) to mediate target cell lysis or engulfment. The cytotoxic capacity of Abs depends on the isotype, which dictates the specificity for different activating FcR<sup>284,285</sup>. More specifically, mouse IgG2a preferentially binds FcγRI with high affinity, FcγRIV with intermediate affinity and FcγRIII with low affinity, while IgG2b binds FcγRIII and FcγRIV. In contrast, IgG1 binds inhibitory FcγRIIb with high affinity and to a lesser extent FcγRIII and thus has low/no cytotoxic activity<sup>286</sup>. Thus, IgG2a and IgG2b isotypes have higher cytotoxic potency than IgG1 and IgG3. A recent promising development in the field of immunotherapeutic approaches utilizing mAb is the engineering of the Fc part of mAbs to enhance or alter their binding affinity or specificity to different FcR and in this way affect their mode of action. ADCC is one of the main modes of action of mAbs in cancer

immunotherapy the efficacy of which can be improved. Enhancement of ADCC could be achieved through the introduction of modifications in the Fc part of mAbs, which lead to increased binding affinity to FcγRIIIA<sup>287</sup>. In fact *in vivo* studies in FcR deficient mice have highlighted the importance of specific Fc-FcR interactions in mediating the anti-tumor effect<sup>288-290</sup>. Moreover, a recent study in FcγR-humanized showed that while FcγRIIIA-binding on macrophages is essential for the ADCC activity of human IgG1 Abs, binding to FcγRIIA on DCs is responsible for the induction of a potent vaccinal effect against the tumor<sup>291</sup>. Moreover, FcR alleles attributing increased binding affinity to activating FcR result in increased ADCC and directly correlate with enhanced clinical responses to mAb therapy in patients<sup>292-296</sup>. There are currently several mAb targeting tumor-associated cell surface differentiation Ag, growth factors or molecules involved in angiogenesis that have been approved for clinical use in the treatment of cancer. These targets are: ERBB2 (trastuzumab), EGFR (cetuximab, panitumumab), VEGF (bevacizumab), CD20 (rituximab, ofatumumab, ibritumomab, tositumomab), CD30 (brentuximab), CD33 (gemtuzumab) and CD52 (alemtuzumab). The anti-CD20 mAb (Rituximab) targets B cells causing their depletion through an FcR-dependent manner. Besides being a useful tactic to eliminate malignant B cells, this mAb can also be used to deplete B cells in tumor settings where they exert a tumor-promoting effect, as has been shown in pancreatic cancer<sup>297</sup>.

#### *Immunomodulatory antibodies*

Immunomodulatory Abs can target activating molecules on the surface of immune cells. CD40, a member of the tumor necrosis factor receptor (TNFR) family, is a co-stimulatory molecule, which is expressed on APCs such as DCs, B cells, monocytes and macrophages<sup>298</sup>. CD40 triggering by the anti-CD40 mAb Dacetuzumab leads to up-regulation of co-stimulatory molecules, induces cytokine production and enhances Ag presentation. Particularly in B cells, it promotes maturation, GC formation, Ig-isotype switching and affinity maturation. Its mode of action depends on the inhibitory FcγRIIb, and is similar for other members of the TNFR family such as CD95, DR4 (TNFRSF10A) and DR5 (TNFRSF10B)<sup>299,300</sup>. DR3 (TNFRSF25) is expressed on activated CD8<sup>+</sup> T cells and interacts with TL1A, a TNF-like cytokine, on CD4<sup>+</sup> T cells to promote inflammation and NK cell activation. Crosslinking of DR3 leads to enhanced cytotoxic NK cell activity. Another member of the TNFR family, CD27, is a co-stimulatory molecule expressed on T, B and NK cells. Binding of CD27 to its ligand CD70 is important for the effector functions of these cells. Crosslinking of the receptor by mAb leads to enhanced immune effector functions. Another target receptor is CD137 (4-1BB), which is expressed on activated T cells, T<sub>regs</sub>, NK cells, NKT cells, DCs, neutrophils and monocytes. Crosslinking of CD137 promotes expansion of T cell populations, CD8<sup>+</sup> T cell survival, NK cell proliferation and IFNγ production<sup>301</sup>. Also, targeting activated T cells through their IL-2Rα chain by the anti-CD25 mAb (Daclizuman), promotes T cell proliferation. Moreover, CD25 is highly expressed by T<sub>regs</sub> which leads to their transient depletion, leading to increased numbers of effector T cells and decreased immunosuppression<sup>178</sup>.

A particular sub-category of immunotherapeutic Abs, known as immune checkpoint therapies, has shown great promise, evoking durable clinical responses, in patients with metastatic disease, and has revolutionized the field of cancer immunotherapy in the last decade. Checkpoint inhibitors aim to block regulatory pathways in T cells that inhibit anti-tumor responses leading to enhanced co-stimulation<sup>302</sup>. The first checkpoint therapeutic to be approved by the FDA in 2011 for the treatment of metastatic melanoma was anti-CTLA-4 (cytotoxic T lymphocyte antigen 4) mAb, Ipilimumab. Ipilimumab targets CTLA-4, a negative regulator of T cell activation, which is up-regulated on activated T cells and functions as a safety switch to attenuate the response. CTLA-4 is a homologue of CD28 and binds the same ligands, CD80 and CD86, with much higher affinity. Contrary to the co-stimulatory effect of CD28, CTLA-4 cross-linking to its ligands leads to inhibition of the proliferation of activated T cells<sup>303,304</sup>. Validation of the mechanism of action of Abs blocking the interaction between CTLA-4 and CD80 was performed in numerous experimental settings, in combination with various treatments that would expose TAA and make them readily available for uptake by APCs<sup>305-308</sup>. Antibody-mediated blocking of CTLA-4-mediated suppression unleashed effector T cell activation in an FcR-independent manner. At the same time, it induced FcR-dependent elimination of Tregs by intratumoral macrophages<sup>309,310</sup>.

Two other immune checkpoint Abs were recently approved by the FDA in 2014. The anti-PD-1 mAb Pembrolizumab and Nivolumab target programmed death receptor 1 (PD-1), which is expressed on activated T cells and B cells and provides a potent inhibitory signal when bound to its ligands PD-L1 and PD-L2 expressed on APCs or tumor cells<sup>311</sup>. Blocking this interaction with mAb interferes with signaling through the TCR leading to T cell inactivation<sup>312</sup>.

The ligand for PD-1, PD-L1, can be up-regulated on tumor cells as an immune escape mechanism by inducing T cell death. Targeting PD-L1 with mAb inhibits binding to PD-1 and CD80, rescuing effector T cells. Clinical trials using Ipilimumab and Pembrolizumab/Nivolumab or anti-PD-L1 as a combination therapy are on-going and hold great promise.

There are several inhibitory receptors on lymphocytes that have been explored as potential targets for Ab-mediated immunotherapy. For instance, OX40 a negative regulator of lymphocyte proliferation and cytokine production on activated lymphocytes, GITR, and KIR2DL1/L2/L3 and KIR2DS1/S2) on NK cells which inhibit NK cytotoxic activity<sup>313-316</sup>. New members of the immune checkpoints family are continuously emerging and are being evaluated as monotherapies and in combination with already established therapies. These aim to provide a blockade of inhibitory pathways, such as LAG-3, VISTA, BTLA and co-stimulation of activating pathways ICOS<sup>317-322</sup>.

#### **1.4.2 Targeting tumor-associated macrophages**

Modulation of the immunosuppressive myeloid compartment has become a crucial factor in determining the success of cancer immunotherapy and is currently the subject of vigorous

investigation. Intensive efforts continue to identify candidate molecules targeting different aspects of the development and function of TAMs. These have three main goals: 1) to decrease the number of precursors recruited to the tumors by inducing apoptosis or maturation or by inhibiting their trafficking, 2) to block molecular mechanisms employed by TAMs to inhibit lymphocyte tumoricidal activity, and 3) to eliminate TAMs or re-program them to an M1 phenotype. The last category is of particular interest for this thesis and some examples are mentioned below.

In a mammary carcinoma model, blockade of the CCL2/CCR2 axis lead to decreased recruitment of inflammatory monocytes and macrophages and as a result inhibited metastatic spread <sup>323</sup>. Combinatorial treatment with clodronate-liposomes and VEGF-neutralizing Abs decreased certain TAM subpopulations and led to a reduction in tumor-associated DC and blood vessel density <sup>324</sup>. Targeting legumain, a stress protein and a member of the asparaginyl endopeptidase family, on TAMs, lead to CD8<sup>+</sup> cytotoxic T cell responses against TAMs and decreased pro-angiogenic factors, such as TGF- $\beta$ , TNF $\alpha$ , MMP-9 and VEGF, thus constricting angiogenesis and metastasis of breast carcinomas <sup>325</sup>. Adenoviral delivery of CCL16, TLR9 stimulation with CpG DNA and anti-IL-10R Abs led to TAM reprogramming from M2 to M1 phenotype leading to tumor de-bulking, followed by DC migration to draining lymph nodes for priming of adaptive responses <sup>326</sup>. Treatment with soluble IL-12 led to a phenotypic switch to M1 TAMs, characterized by a reduction in the levels of anti-inflammatory IL-10, TGF- $\beta$  and CCL2 and increased levels of TNF, IL-15 and IL-18, and concomitant NK and T cytotoxic cell activation in melanoma, lung and colon carcinoma <sup>327</sup>. Similarly, adoptive transfer of tumor-specific cytotoxic T cells (CTLs) or chimeric-Ag receptor T cells both engineered to release IL-12 shifted the TAM polarization to M1 <sup>328,329</sup>. Targeting of NF- $\kappa$ B, the master regulator of the immunosuppressive phenotype of TAMs, through its downstream signaling kinase I $\kappa$ B re-educated TAMs to the M1 phenotype in ovarian carcinoma <sup>330</sup>. Treatment of mice bearing mammary carcinoma 4T1-Neu with the chemotherapeutic agent docetaxel induced MDSC apoptosis and reprogramming of TAMs to M1 <sup>331</sup>. In another study, HRG (Host-produced histidine-rich glycoprotein) down-regulated PIGF (placental growth factor) and skewed TAM polarization to M1 phenotype promoting anti-tumor responses and vessel normalization <sup>332</sup>. Anti-CD40 Abs in combination with IL-2 reprogrammed TAMs and up-regulated iNOS in TAMs of lung metastases but not in the primary tumor <sup>333</sup>. In patients with pancreatic cancer agonistic anti-CD40 Abs activated macrophages to infiltrate pancreatic tumors and exert tumoricidal activity in combination with chemotherapeutic agent gemcitabine <sup>334</sup>. CSF-1 is an important cytokine for macrophage survival and M2 polarization. Use of CSF-1R antagonist inhibited the expansion of MDSCs and TAMs and their recruitment to lung and prostate tumors <sup>335</sup>. Blockade of CSF-1R and cKIT receptor tyrosine kinase led to reduced TAM recruitment in a mammary carcinoma model <sup>226</sup>. In a mouse model of glioblastoma multiforme, inhibition of CSF-1R altered TAM polarization to M1 and blocked tumor growth <sup>336</sup>. Inhibiting TAMs and their precursor inflammatory monocytes through targeting of CSF-1R and CCR2 reduced the number of tumor-infiltrating TAMs, but also reprogrammed the remaining TAMs, thus reducing metastasis and increased anti-tumor T cell responses in a pancreatic tumor model <sup>337</sup>. This

TAM-reprogramming approach has shown great promise in improving responses to T cell checkpoint immunotherapies using anti-PD-1 and anti-CTLA-4 Abs<sup>338</sup>. In another study, targeting of CSF-1R enhanced anti-tumor responses of adoptively transferred T cells<sup>339</sup>. In Paper II we use mAbs to target scavenger receptor MARCO in order to reprogram M2 TAMs in the TME.

### **1.4.3 Exosomes in cancer immunotherapy**

The release of extracellular vesicles is a way of intercellular communication employed by many different cell types. Exosomes are 30-100nm vesicles consisting of a lipid bilayer membrane that derive from the late endosomal compartment. They are shed from different types of cells, including APCs and tumor cells, during both physiological and pathophysiological conditions. Exosomes can carry membranous proteins on their surface that are involved in membrane transport, fusion, adhesion, Ag-presentation and immune stimulation. They can also contain cytosolic proteins, lipids and RNA molecules. Depending on the cell of origin and its activation state, their composition may vary. As a result, they have various immunomodulatory activities, which also depend on their cargo. These range from cell-cell communication to immunomodulation, and acting as shuttles for Ag-presentation, transfer of proteins, mRNA and microRNA.

The observation that exosomes originating from APCs, such as DCs and B cells, carry MHC-I and -II as well as co-stimulatory molecules on their surface, suggested that they are involved in direct Ag-presentation to T cells. In fact, in a mouse model of melanoma tumor-peptide pulsed DC-derived exosomes exhibited potent immunostimulatory and anti-tumor capacity leading to activation of T cell effector functions and tumor growth inhibition<sup>340</sup>. Alternatively, exosomes can indirectly activate T cell by enhancing the Ag-presenting capacity of DCs. It has been shown that exosomes that are taken up by immature DCs transfer their antigenic load to endogenous MHC molecules, which are then transported to the surface of the DC and present Ag in the classical manner to stimulate T cells<sup>341</sup>.

Tumor-derived exosomes have been suggested to both promote and suppress Ag-specific and non-specific anti-tumor responses. For example, tumor-derived exosomes are enriched for receptors that induce T cell apoptosis, such as TNF-related apoptosis-inducing ligand (TRAIL) and CD95, leading to suppression. Additionally, they can promote the immunosuppressive TME as they are enriched in factors such as prostaglandin E2 and TGF- $\beta$  that drive the generation of MDSCs and M2 polarization of TAMs<sup>342</sup>. Also, as carriers of mRNA and microRNA molecules, exosomes have the capacity to regulate the transcription of different genes in the recipient cells. For example tumor-derived exosomes carrying miR-21 can bind to TLR7 and 8 activating macrophages in the TME to produce TNF, IL-6 and thereby lead to increased tumor growth and metastasis<sup>343</sup>. Tumor-derived exosomes also facilitate tumor invasion and metastatic spread by conditioning the distal locations for the seeding of micrometastases, as has been shown in the case of melanoma-derived exosomes that accumulate in draining lymph nodes<sup>344,345</sup>.

However, exosomes carrying tumor-Ags have also been shown to activate DCs to induce potent CTL anti-tumor responses<sup>346</sup>. Moreover, experimental approaches have tried to generate immunogenic exosomes by pulsing APCs in vitro with tumor-derived Ags, aiming to induce Ag-specific immune responses against the tumor. It has been shown that activation of B cells is necessary for the efficient induction of T cell responses to exosome-bound Ags<sup>347</sup>. Additionally, exosomes have also been described as potent inducers of inflammatory cascades and could thus be used as adjuvants as they carry a wide array of stimuli that lead to immune activation. These can be inflammatory mediators (cytokines such as IL-1 $\beta$ ), microbial Ags that are ligands for PRRs and TNF-related proteins (FasL, TRAIL, CD40L).

Immunotherapeutic approaches focus on attempts to modulate exosome composition, release and cell targeting capacity, as well as modulating the activity of the APC that they originate from by cytokines, Ag-stimulation, gene transfer and microRNA pulsing, as has been reviewed elsewhere<sup>348,349</sup>. In Paper IV we use DC-derived exosomes loaded with  $\alpha$ GC and the model Ag OVA in a vaccination approach to induce Ag-specific anti-tumor responses.

## **1.5 MURINE MODELS OF HUMAN MELANOMA, BREAST AND COLON CANCER**

Breast cancer is the most common cancer in women (second most common cancer overall), with 1.7 million new cases and representing 25% of all cancers in women in 2012. Melanoma accounts for 1.6% of all cancers and colon cancer is the third most common cancer, representing 9.7% of all cancer cases in 2012, (source: www.wcrf.org).

Studies on murine tumor models have shown that metastatic patterns of different cancer cell lines are not random, but rather site specific and depend on the site of injection of the tumor cells, thus taking into account the local tissue-specific microenvironment and supporting the use of orthotopic models in cancer research<sup>350,351</sup>. However, this is not always an easy task. For several reasons the below-mentioned models were assessed as valid to explore the hypotheses described in this thesis.

### **1.5.1 Mouse models of melanoma (B16)**

The B16 melanoma cell line was derived from a spontaneous C57BL/6 tumor and is a widely used model in melanoma studies<sup>352,353</sup>. It is known for its aggressive growth and being difficult to treat. The cells can either be inoculated subcutaneously in the flank of a mouse to generate a primary tumor or intravenously to simulate a metastatic model of melanoma in the lung. B16 exists in many different variants, the difference being their metastatic potential. The most commonly used is the B16.F10, which metastasizes from the primary subcutaneous tumor to the lymph nodes and lungs. B16 is a rather plastic cell line and the degree of pigmentation may vary, however this does not seem to influence the expression of different TAAs. B16 expresses many of the TAAs that human melanomas express, making it a good model system for the human disease. One of the TAAs expressed on the surface of B16 cells is glycoprotein 75 or TRP-1. The surface molecule gp75, is targeted by Abs of the TA99 IgG2a clone (used in Paper II) which when bound trigger its internalization and mediate ADCC by NK cells<sup>354</sup>. This antibody clone is used as a golden standard (positive control) in

many immunotherapy settings tested in the B16 model. The B16 cell line exists in many functional variants genetically modified to express different genes of interest, such as GFP, luciferase, Flt3L, GM-CSF and soluble or membrane-bound ovalbumin. This makes it a very versatile and useful model system. B16 expresses low levels of MHC-I<sup>355</sup> and MHC class II expression can be induced by IFN $\gamma$ <sup>356</sup>. B16 is a low- or non-immunogenic tumor. The B16 melanoma model is a widely used model with many modified version that allow flexibility in functional read-outs, such as the OVA-expressing and the luciferase-tagged variants that are used in Papers II and IV.

### **1.5.2 Mouse model of breast cancer (4T1)**

The 4T1 mammary carcinoma cell line was derived from a cell line that originated from a spontaneous tumor of a MMTV Balb/c mouse, which was transplanted to a C3H mouse<sup>357,358</sup>. It is a typical triple negative mammary carcinoma cell line with a mesenchymal phenotype (ER<sup>-</sup>/PR<sup>-</sup>/HER2<sup>-</sup>), the most challenging to treat type of breast cancer. As shown in Paper II, the MARCO receptor is highly expressed in triple negative human breast cancer, making 4T1 a relevant experimental model to study the effect of MARCO-targeting. 4T1 was selected for being easy to culture, highly tumorigenic when inoculated into mice, resistant to 6-thioguanine and for its capacity to metastasize to distal sites. There are several reasons that make the 4T1 cell line a suitable model for human breast cancer. Firstly, it can be readily introduced orthotopically into the mammary fat pad of mice and can metastasize from the primary tumor site to several sites (lymph nodes, bone, brain liver and lungs) in a pattern similar to human mammary carcinomas. Moreover, after inoculation the disease process progresses over a period of several weeks modeling the human setting. Much like many human cancers, 4T1 tumors are poorly immunogenic, which means that treatment with irradiated tumor cells fails to mount a memory response against a secondary introduction of the same tumor. 4T1 cells express MHC-I but not MHC-II, making them good targets for CD8<sup>+</sup> CTLs and NK cells. Finally, as metastatic disease is of great importance to study, a great advantage comes from being able to excise the primary tumor in order to generate a model for metastatic disease simulating the human setting where surgical removal of the primary tumor proceed any other treatment step.

### **1.5.3 Mouse model of colon cancer (MC38)**

The MC38 colon adenocarcinoma cell line was established in a C57BL/6 mouse using dimethyl-hydrazine (DMH). DMH 0.2mg/mouse was injected subcutaneously (s.c.) weekly for 7 moths. Formed colon tumors were excised and passaged and the ones that survived the first passage went on to serial transplants. The MC38 tumor was the only small tumor to survive the first passage and was transplanted by trocar to the axillary region. MC38 was characterized as a grade III adenocarcinoma of the colon, which is highly metastatic to the lung<sup>359</sup>. However, intraperitoneal transplantation gave rise to hepatic metastases, indicating that the site of implantation affects the metastatic pattern. MC38 has been extensively used in studies of checkpoint therapies with anti-CTLA-4, anti-PD-1 and anti-PD-L1 Abs, and thus

was assessed as a relevant model to compare combination therapies of anti-CTLA-4 and anti-MARCO Abs in Paper II.

#### **1.5.4 Mouse model for the study of EMT and metastasis**

To study the complexity of the metastatic process, where the evaluation of tumor cell motility and the ability to colonize distal sites is imperative, perhaps the most physiological assay is the injection of tumor cells sub-cutaneously or in the footpad. This route of injection gives direct access into the lymphatics, where the first stop is the popliteal lymph node. This model was used to compare the migratory capacity of cell lines with different EMT profiles in Paper III. EpH4 is a non-invasive non-migratory cell line derived by immortalization of balb/c mammary epithelial cells <sup>360</sup>. EpRas cells were generated by transducing the parental EpH4 cells with a retroviral vector expressing v-Ha-ras <sup>361</sup>. Upon stimulation with TGF- $\beta$  these cells gain migratory capacity. Finally, EPXT cells are stably in EMT under the influence of oncogenic Ras and autocrine TGF- $\beta$ -signaling.



## 2 THE PRESENT STUDY

### 2.1 AIMS

This thesis aims to identify novel candidates in the tumor microenvironment for targeted immunotherapy of cancer.

Specific aims:

**Paper I** - To investigate the impact of monoclonal antibodies targeting scavenger receptor MARCO on marginal zone macrophages and adaptive immune responses.

**Paper II** - To modulate the immunosuppressive tumor microenvironment into induction of anti-tumor immune responses by using monoclonal antibodies to scavenger receptor MARCO targeting tumor-associated macrophages.

**Paper III** - To investigate the role of TGF- $\beta$ 1-induced epithelia-mesenchymal transition on lymphatic metastasis of cancer cells.

**Paper IV** - To evaluate the immunomodulatory role of exosomes as cancer vaccines, triggering tumor-specific adaptive immune responses.

## 2.2 RESULTS AND DISCUSSION

### 2.2.1 Marginal zone macrophages regulate antigen transport by B cells to the follicle in the spleen via CD21 (Paper I)

The presence of anti-MARCO autoantibodies has previously been reported in SLE patients, however, their role in the pathogenesis of autoimmune disease is still unknown. In this study we investigate the possible regulatory effects induced by the anti-MARCO Abs on other immune cells in responses to foreign Ags. MZMs are strategically positioned at the marginal sinus of the spleen where they capture incoming Ag from the circulation via scavenger receptor MARCO and interact with MZBs. Based on this, we therefore set out to investigate the effect of anti-MARCO Abs on the MZM/MZB interplay.

To study the effects of MARCO-crosslinking on the MZM/MZB interaction, a rat anti-mouse MARCO Ab was injected intravenously (i.v.) in wildtype (wt) mice. The Ab selectively bound to MARCO-expressing MZMs in the MZ without depleting them. This resulted in the gradual loss of CD21 (complement receptor 2) on MZBs, and to a lesser extent on other B cell populations, already one hour after administration. The loss of CD21 could not be attributed to local complement activation, blocking of antibody binding, or internalization of CD21. It was also solely targeting CD21, leaving its alternative splice-variant, CD35 on FDCs unaffected. A similar CD21 loss was observed also with polyclonal mouse serum containing Abs against MARCO, thus excluding inter-species cross-reactivity. Since MARCO can be up-regulated in response to LPS-stimulation, we investigated the involvement of MyD88-, TLR2-, TLR4- and TLR9-signalling in mediating CD21 loss on MZBs upon MARCO triggering. However, these pathways were not involved and we could also exclude the involvement of Fc $\gamma$ Rs.

Due to the speed of the response (observed CD21-loss already 1 hour post administration), we hypothesized that shedding of CD21 from the surface of MZBs could be the underlying mechanism of the CD21 loss. This is known to occur on B cells and soluble CD21 can be detected in several inflammatory and autoimmune conditions. Purinergic enzyme receptors P2X7 and P2Y have the capacity to cleave CD21 in response to extracellular ATP. Based on that, ATP-release after anti-MARCO Ab addition to peritoneal MARCO<sup>+</sup> macrophage cultures *in vitro*, was investigated. Extracellular ATP could be detected already 5 minutes post stimulation. Moreover, addition of extracellular ATP to splenocyte cultures led to a decrease in CD21 levels on MZBs. Increased levels of extracellular ATP are associated with pro-inflammatory activation of macrophages, referred to as M1 polarization. This suggests that MARCO-engagement leads to macrophage activation. To further investigate the effect of the released ATP by the macrophages we assessed the expression of CD39, an ectonucleotidase involved in the catabolism of pro-inflammatory extracellular ATP to ADP. This metabolic reaction facilitates the transition of macrophages from a pro-inflammatory to an anti-inflammatory regulatory state, thus functioning as a homeostatic switch during inflammation<sup>362</sup>. Interestingly, 24 hours after stimulation of macrophages with anti-MARCO *in vitro*, CD39-expression increased to levels similar to those induced by the addition of extracellular ATP. This could be regarded as a compensatory mechanism

aiming to regulate macrophage activation in response to immunogenic stimuli, such as ATP.

It has previously been shown that MZBs take up Ag and migrate back and forth to the follicle depositing Ag on the FDCs. CD21 is an important receptor for opsonization of foreign Ags and generation of immune complexes. To assess the impact of CD21 loss in response to anti-MARCO engagement on Ag-shuttling, mice were immunized with high molecular weight dextran, as a model Ag. Dextran is dependent on CD21 for its uptake by MZBs, making it an ideal model-Ag to study complement receptor-dependent Agtransportation. Interestingly, pre-treatment with anti-MARCO Abs significantly reduced binding of dextran to MZBs, due to the decrease in CD21. Importantly, it also decreased the amount of dextran that was deposited on the FDCs. Reconstitution of CD19-deficient mice (lacking MZBs) with splenic B cells made it possible to tract the migration of CD19<sup>+</sup> MZBs upon Ag-stimulation. No differences were observed, suggesting that the ability of MZBs to migrate to the follicles of the spleen in response to dextran immunization was not affected by the anti-MARCO Ab treatment. In conclusion, MARCO-crosslinking affects the amount of Ag that can be captured by MZBs due to the loss of CD21, as well as Ag deposition on FDCs in the follicles, while leaving the shuttling capacity of MZBs intact.

Ag deposition on FDCs is important both for the initiation of the GC reaction and consequently also humoral responses. Naturally, we next investigated how the loss of CD21 on MZBs and the consequent reduction of Ag deposition on FDCs affected the subsequent immune response to those Ags. For that, immunization with TI Ag NP-dextran was performed after anti-MARCO treatment, at which point the expression of CD21 was already lost. Analysis of the subsequent humoral response showed significantly lower levels of Ag-specific IgM Abs in mice pretreated with anti-MARCO Abs. This is in line with the fact that dextran is primarily bound by MZBs, which respond best to TI Ags.

Next, we analyzed the humoral response against the TD Ag NP-CGG after anti-MARCO injection. In line with our previous results, anti-MARCO injected mice showed a reduction in GC B cell numbers compared to controls. This was further reflected in the antibody response in treated mice, where lower IgM, IgG1 and IgG3 levels were observed. Thus, anti-MARCO Abs have a pronounced effect on the humoral immune response against both TI and TD Ags. This suggests that the elevated levels of anti-MARCO autoantibodies found in SLE patients could contribute to the increased risk of infection observed in these patients. An additional contribution of anti-MARCO Abs to the pathology of SLE could be that defective complement receptor expression on B cells leads to inefficient clearance of apoptotic cells, which causes a break in self-tolerance.

In summary, this paper reports a novel mechanism through which, resident MZM in the spleen regulate innate and adaptive immune responses to foreign Ags. MZM activation through PRR MARCO by Abs leads to ATP-release, cleavage of CD21 on MZBs and reduced Ag deposition on FDCs. Since MARCO can also bind self-Ags, a similar mechanism could limit their deposition on FDCs. It would therefore be interesting to investigate how this pathway can be manipulated to regulate unwanted adaptive immune

responses to self-Ags or to enhance protective immunity to prevent infection. Finally, crosslinking of MARCO with Abs led to ATP-release, which is associated with M1 macrophage polarization. This knowledge led to the hypothesis that MARCO can be targeted by Abs to induce a switch to a pro-inflammatory phenotype, for example in the setting of cancer.

### **2.2.2 Reprogramming tumor associated macrophages by antibody targeting inhibits cancer progression and metastasis (Paper II)**

Tumors are complex tissues consisting of various types of cells besides the tumor cells. TAMs are a heterogeneous population of macrophages and make up the majority of the tumor-infiltrating inflammatory myeloid cells that account for the immunosuppressive microenvironment of tumors.

Based on the fact that MARCO expression is restricted to certain macrophage subpopulations, we wanted to investigate whether MARCO was also expressed on TAMs in the TME. We screened tumor sections of mammary adenocarcinoma, melanoma and colon adenocarcinoma tumor model and identified the presence of MARCO on macrophages in the tumor stroma. MARCO-expression was only present in a fraction of the F4/80<sup>+</sup> cells in the tumor stroma, suggesting that the receptor is restricted to a subtype of macrophages. To further elucidate the nature of the subset of macrophages that expressed MARCO, we sorted the different subsets by flow cytometry. We could identify expression of MARCO on the CD11b<sup>+</sup> Ly6C<sup>lo</sup> MHCII<sup>lo</sup> subset of tumor infiltrating TAMs. This subset showed characteristic expression of typical M2 markers (*arg1*, *fizz1*, etc), as well as Cx3cr1 and low expression of MHC-II, as observed by qPCR.

In an effort to identify what is driving the up-regulation of MARCO on TAMs, we set up an *in vitro* system for polarization of bone marrow-derived macrophages by cytokines or tumor supernatant. Interestingly MARCO expression was up-regulated in M2 and tumor supernatant polarized cultures, supporting our *in vivo* findings.

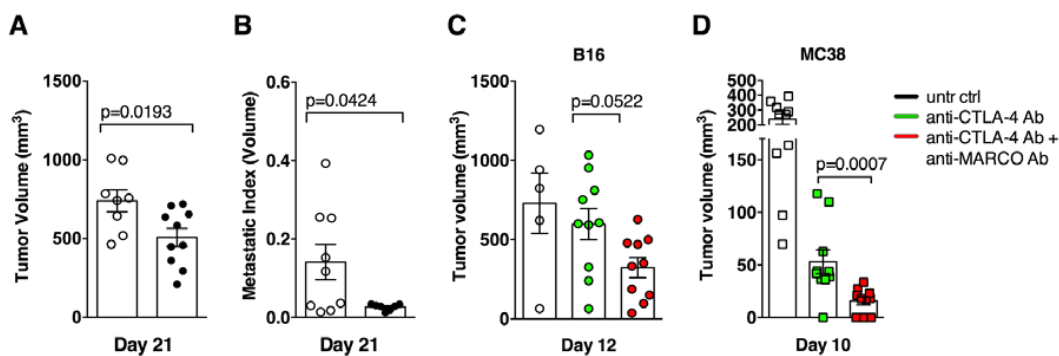
Based on the fact that, IL-10 and TGF- $\beta$  are the major immunosuppressive cytokines in the TME and can promote M2 polarization of macrophages, we investigated whether they could also drive the expression of MARCO. *In vitro* cultures of a peritoneal macrophage cell line were stimulated with IL-10 or TGF- $\beta$  and indeed, MARCO expression increased in response to both these cytokines.

Having identified MARCO as a specific marker for M2 TAMs, we generated mAbs to MARCO, and evaluated their use in an immunotherapy setting to the three previously mentioned tumor models. Anti-MARCO Abs could infiltrate the tumor and bind to MARCO<sup>+</sup>F4/80<sup>+</sup> cells. Surprisingly, anti-MARCO treatment led to decreased primary and metastatic tumor growth in the mammary adenocarcinoma model 4T1 (**Figure 6A and B**). Moreover, we found that anti-MARCO treatment led to an increase in M1 TAMs and a decrease in M2 TAMs. This was accompanied by an increased GC B cell response in the tumor-draining lymph node and an increased CD4<sup>+</sup>h/CD8<sup>+</sup> T cell ratio in the tumor, suggesting the treatment increased tumor immunogenicity. We did not observe any other

significant difference caused by the anti-MARCO treatment when we examined different lymphocyte populations infiltrating the tumors (NK, B and T cells, DC, neutrophils) or Mo-MDSC and PMN-MDSC in the spleens of tumor-bearing mice.

To further validate the effect of anti-MARCO treatment, we used another tumor model, the B16 melanoma. Also here, treatment with anti-MARCO Abs inhibited tumor growth, to a similar degree as TA99 Ab, an antibody known to induce ADCC to B16. Moreover, the anti-MARCO effect was absent in MARCO-deficient tumor bearing animals, suggesting the observed tumor decrease in the treated wt mice was MARCO-specific. Similar alterations were observed in the different tumor infiltrating leukocyte populations as in the 4T1 model, more specifically a decrease in inflammatory macrophages in the tumor and an increase in CD4<sup>+</sup>/CD8<sup>+</sup> T cells, CD4<sup>+</sup>/Tregs, OVA-specific CD8<sup>+</sup> T cells and OVA-specific IgG2a and b.

Additionally, the effect of anti-MARCO treatment was assessed in combination with other mAb immunotherapies. When combined with TA99 mAbs or immune checkpoint anti-CTLA-4 Abs we observed an additive tumor growth inhibiting effect. In particular, anti-MARCO Ab and anti-CTLA-4 Ab combination treatments decreased tumor growth of B16 melanoma and MC38 colon tumor-bearing mice compared to anti-CTLA-4 Ab monotherapy (**Figure 6C and D**).



**Figure 6. Targeting MARCO by mAbs inhibits *in vivo* tumor growth and metastasis**

4T1 (mammary carcinoma), B16 (melanoma) or MC38 (colon carcinoma) cells were injected s.c. in mice followed by Ab treatment. Tumor growth was monitored by manual measurements. (A) Tumor volume of 4T1 mammary carcinoma after anti-MARCO Ab treatment (full circles) compared to untreated controls (empty circles), on day 21 after tumor inoculation (B) Metastatic index (number of lung tumor colonies/primary tumor volume) of 4T1 tumors after anti-MARCO (full circles) compared to untreated controls (empty circles), on day 21 (C) Tumor volume of B16 melanoma untreated, anti-CTLA-4 Ab+Gvax and anti-CTLA-4 Ab +Gvax+anti-MARCO Ab-treated mice, day 12 (D) Volume of MC38 colon cancer untreated, anti-CTLA-4 Ab and anti-CTLA-4 Ab+anti-MARCO Ab-treated mice, day 10. Described also in Paper II: Fig. 3 and 4.

As many immunomodulatory Ab therapeutics require FcγR involvement to exert their function, we generated mouse anti-MARCO Ab variants/mutants with null FcγR-binding and observed that the tumor growth inhibiting effect was FcγR-dependent. Utilizing KO mice deficient for all or only for activating FcγR we could narrow the effect down to the involvement of the inhibitory FcγRIIb. Interestingly, high expression of FcγRIIb correlated

with the macrophage subpopulations exhibiting high MARCO expression, namely the *in vitro* M2 BMDM and tumor sorted M2 TAMs, further strengthening our findings. This suggests that the anti-tumor activity of anti-MARCO Abs is dependent on the ability of the Fc-part of the anti-MARCO Ab to engage the inhibitory FcγRIIB.

To investigate the potential application of the anti-MARCO Ab treatment in the clinical cancer setting we investigated the expression profile of MARCO by gene expression and immunofluorescence on human breast cancer and metastatic melanoma biopsies. In the case of human breast cancer, using the TCGA and KI/Clinseq datasets we found that higher MARCO expression correlated with the basal (triple negative) subgroup of patients. In the case of melanoma, higher MARCO expression was observed amongst in the distal metastases. Thus in both cancers, MARCO expression is associated with more aggressive and metastatic cancer types.

To further interpret MARCO expression with reference to the metastatic cancer profile, we compared MARCO expression relative to an M2 TAM gene signature in the human breast cancer and melanoma datasets. Interestingly, MARCO-expression correlated with many M2 genes as well as with the expression of FcγRIIB, suggesting that MARCO is expressed in tumors with a high TAM content. Furthermore, we analyzed MARCO expression relative to an EMT gene signature, and observed a correlation with the expression of known EMT regulators *mmp9*, *snail* and *twist*.

Finally, we verified MARCO expression on tumor sections from human breast cancer and melanoma patients by immunofluorescence staining. In human breast cancer, we observed a higher infiltration of CD68<sup>+</sup> macrophages in the stroma of the basal triple negative breast cancer compared to the ER<sup>+</sup>/PR<sup>+</sup> subgroup. In addition, MARCO was present on CD68<sup>+</sup> macrophages and correlated with the presence of the M2 markers CD163, as well as CD206, strengthening our hypothesis that MARCO is a marker for M2 TAMs. These data suggest that MARCO is a clinically relevant marker for distinguishing aggressive metastatic breast cancer and melanoma, and provides a novel candidate for targeted immunotherapy using Abs.

Overall, this study identifies a novel marker for a subset of immunosuppressive TAMs, which correlates with metastatic gene signatures in clinical samples of human breast cancer and melanoma. Pre-clinical models show a promising decrease in primary and metastatic tumor growth, as well as re-programming of TAMs from M2 towards M1 phenotype and accompanied by increased tumor-immunogenicity. Moreover, combinatorial treatment using anti-MARCO Ab and checkpoint therapy using anti-CTLA-4 Ab showed an impressive inhibition of tumor growth and increased overall survival. Altogether, this suggests that immunotherapeutic approaches aiming at repolarizing the immunosuppressive TME offer a promising new approach to cancer immunotherapy.

### **2.2.3 TGF- $\beta$ 1-induced EMT promotes targeted migration of breast cancer cells through the lymphatic system by the activation of CCR7/CCL21-mediated chemotaxis (Paper III)**

Metastatic cancer disease presents the greatest challenge in treating cancer. Tumor cells have the ability to disseminate through lymphatic and the blood circulation during metastatic spread. However, it is still unknown whether lymphatic dissemination is a targeted process. TGF- $\beta$ 1 is an abundant cytokine in the TME and has been associated with metastatic disease in many cancers. TGF- $\beta$ 1-signaling is a known activator of a genetic program known as epithelial-mesenchymal transition (EMT), which induces a migratory phenotype in tumor cells. Based on this, we hypothesized that TGF- $\beta$ -induced EMT could be connected to lymphatic dissemination of tumor cells.

To investigate whether TGF- $\beta$ -induced EMT predisposes tumor cells to metastasize through the lymphatics, we used a mouse model frequently used to study DC trafficking to draining popliteal lymph nodes (PLN), by injecting tumor cells into the hind footpad of mice. For this, three versions of a mammary epithelial cell line, with different EMT profiles, were used. EpH4 is non-metastatic in response to TGF- $\beta$ 1, EpRas originates from EpH4 but has constitutive oncogenic Ras-signaling and undergoes EMT in response to TGF- $\beta$ 1, and EpXT is derived from EpH4 but has constitutive expression of oncogenic Ras- and TGF- $\beta$ 1-signaling and is in stable EMT. Fluorescently labeled versions of the cell lines were injected into the footpads of mice to assess the migratory capacity to the draining PLN. EpXT cells formed tumors at the site of injection but also acquired a migratory capacity and were detected in the draining PLN after 2 days. They were primarily located in the subcapsular sinuses, indicating entrance through afferent lymphatic vessels. In contrast, non-migratory EpH4 cells formed small tumors only at the site of injection and could not be detected in lymph nodes. EpRas cells gained migratory capacity only upon pre-treatment with TGF- $\beta$ 1. Thus, induction of EMT by TGF- $\beta$ 1-signaling induced a migratory phenotype on the tumor cells, which led to lymphatic metastasis.

Moreover, immunofluorescence stainings for blood and lymphatic vessels in EpXT footpad tumors showed that blood vessels were evenly dispersed within the tumors, while lymphatic vessels clustered in certain regions. The areas of lymphatic vessel clustering were associated with more invasive/migratory tumor cell morphology and showed evidence of intravasation. This suggested that EMT might induce lymphatic metastasis in a targeted manner. To investigate this, an *in vitro* 3D matrix co-culture system was used to study migration towards lymphatic endothelial cells (LECs), or vascular endothelial cells (VECs). Interestingly, tumor cells undergoing EMT acquired an elongated morphology and formed clusters of invasive cells that preferentially migrated towards LECs, compared to VECs. Non-EMT cells, on the other hand, did not migrate. Taken together, these results suggest that mammary tumor cells that have undergone TGF- $\beta$ -induced EMT have an increased migratory capacity and preferentially migrate in a targeted manner towards lymphatic vessels, as opposed to blood vessels.

It is well established that upon activation by inflammatory stimuli in the periphery, DCs use

chemokine receptor CCR7 to respond to a chemokine gradient of its ligand CCL21. The CCR7/CCL21 chemotactic axis guides DC migration through lymphatic vessels in order to ultimately travel to draining lymph nodes where they initiate adaptive immune responses. Based on the fact that breast cancer cells can also express CCR7, we hypothesized that CCR7 could have a similar role in guiding the lymphatic dissemination of EMT cells. Indeed, both TGF- $\beta$ 1-induced EMT models showed up-regulated expression of CCR7, compared to the non-EMT EpH4, suggesting that TGF- $\beta$  driven CCR7 expression is dependent on EMT. To investigate the CCR7/CCL21 chemotactic axis we set up transwell invasion assays, in which mammary cancer cells that had undergone TGF- $\beta$ -induced EMT were seeded in the upper chamber and recombinant CCL21, was added in the lower chamber as a chemoattractant. Tumor cells that had been treated with TGF- $\beta$ 1 migrated more efficiently towards the CCL21 gradient, compared to untreated cells. When a neutralizing anti-CCR7 Ab was added to the cultures, the CCR7/CCL21 interaction was blocked leading to reduced targeted migration. Similarly, reduced migration of EpXT cells was observed in the in vitro 3D migration assay towards LECs in the presence of the neutralizing anti-CCR7 antibody, further strengthening our hypothesis. Moreover, in order to assess if this was the case also in vivo, we introduced small interfering RNA to silence CCR7 in EpXT cells and injected these cells into the mouse footpad. Interestingly, CCR7-siRNA-treated EpXT cells expressed lower levels of CCR7 and exhibited reduced migration to the PLN compared to their untreated counterparts. These data demonstrate that the targeted migration of TGF- $\beta$ 1-induced EMT mammary cancer cells to the lymphatics depends on the interaction of CCR7 with CCL21.

TGF- $\beta$ 1-signaling during EMT can be induced through two independent pathways, a Smad-dependent and a P38 MAPK-dependent pathway. In order to elucidate the underlying molecular mechanisms governing CCR7 up-regulation during TGF- $\beta$ -induced EMT, we utilized molecular inhibitors to those pathways. Surprisingly, both Smad3 and p38 MAPK inhibitors led to decreased CCR7 mRNA levels during TGF- $\beta$ 1-induced EMT in mammary cancer cells. However, when assessing CCR7 protein levels only p38 MAPK had significant inhibitory effect, suggesting that factors operating downstream of p38 MAPK signaling were involved in inducing CCR7 expression in EMT cells. By gene expression analysis we were able to show that AP-1 factors c-Jun and JunB are responsible for the CCR7 overexpression as a result of TGF- $\beta$ 1-induction in EMT in mammary cancer cells. Moreover, induction of JunB is P38 MAPK dependent. These results highlight the P38 MAPK pathway as the underlying mechanism of CCR7-mediated chemoattraction and lymphatic dissemination of mammary cancer cells having undergone TGF- $\beta$ 1-induced EMT. It also demonstrates that P38 MAPK blockade can reverse EMT, thus highlighting it as a novel target for pharmacological inhibition to abrogate lymphatic spread of mammary cancer cells. Interestingly, TGF- $\beta$ 1-induced EMT mammary cancer cells that retain endogenous TGF- $\beta$ 1 production can induce the secretion of CCL21 by lymphatic endothelia vessels, thus potentiating their own metastatic spread. This can be regarded as a feed back loop driving the lymphatic spread of malignant cells. Finally, histological evaluation of human breast cancer tissue sections revealed that CCR7-expression is



associated with tumor cells in invasive areas of the tumors with characteristic low expression of E-cadherin, suggesting they have undergone EMT. Moreover, analysis of microarray data from a cohort of human breast cancers indicated that high expression of CCR7 and CCL21 correlates with an EMT gene signature.

Overall, the results show that tumor cells use a similar mechanism as DCs to gain migratory capacity through the lymphatic system in a targeted manner. Furthermore, it identifies p38 MAPK as a useful candidate for targeted inhibition to limit EMT and lymphatic dissemination of tumor cells. TGF- $\beta$ 1-signaling plays a crucial role in driving EMT and the up-regulation of CCR7. As previously mentioned, TGF- $\beta$ 1 is highly expressed by several cell types present in the suppressive TME, such as TAMs and T<sub>regs</sub>. Targeting TGF- $\beta$ -producing cells in the TME for reprogramming, with approaches such as the one described in Paper II, may thus provide additional ways to restrict lymphatic dissemination of cancer cells.

#### **2.2.4 Synergistic induction of adaptive antitumor immunity by codelivery of antigen with $\alpha$ -galactosylceramide on exosomes (Paper IV)**

NKT cells are potent regulators of innate and adaptive immune responses. Moreover in the context of cancer, they have been implicated in the maturation of DCs facilitating subsequent CD8<sup>+</sup> T cell anti-tumor responses, as well as in the killing of immunosuppressive TAMs. Their function is however often compromised in cancer. Thus attempts to re-activate anergized NKT cells are of great interest in cancer immunotherapy.

NKT cells recognize lipid Ags presented by the MHC-like molecule CD1d.  $\alpha$ -GalCer is a super-agonist of NKT cell activation, however when administered in a soluble form it leads to NKT cell anergy. Exosomes derived from APCs, and DCs in particular, can express CD1d and have been explored in immunotherapeutic approaches aiming to activate adaptive immune responses towards tumor-Ag.

To assess the capacity of exosomes to activate NKT cells, we generated exosomes from bone marrow-derived DCs from wt and CD1d-KO mice, lacking NKT cells. We next loaded exosomes with  $\alpha$ -GalCer and the model Ag ovalbumin (OVA) and assessed NKT cell activation in vitro. OVA-loaded exosomes (OVA/exo) or  $\alpha$ -GalCer-loaded exosomes ( $\alpha$ -GalCer/exo) were added to splenocytes cultures from Val4 mice, transgenic for the iNKT cell receptor. CD1d-proficient  $\alpha$ -GalCer/exo induced greater proliferation of iNKT cells compared to CD1d-deficient  $\alpha$ -GalCer/exo, suggesting iNKT cell activation is in part dependent on the presence of CD1d on the surface of exosomes. OVA/exo failed to induce NKT activation, indicating that an activating ligand, in this case  $\alpha$ -GalCer, for the NKT cell TCR is necessary for their activation. CD1d-proficient  $\alpha$ -GalCer/exo also induced greater IL-4, IFN $\gamma$  and IL-17A production by splenocytes, compared to the low levels observed in response to CD1d-deficient  $\alpha$ -GalCer/exo stimulation and no cytokine production by OVA/exo stimulation. The results suggest, that  $\alpha$ -GalCer/exo potently stimulate NKT cell activation, proliferation and cytokine production in part, but not exclusively, through CD1d and require exosomal  $\alpha$ -GalCer.

In an attempt to translate our findings to an *in vivo* system, we generated DC-derived exosomes loaded with  $\alpha$ GC and OVA ( $\alpha$ -GalCer-OVA/exo) or the CD8<sup>+</sup> T-cell-specific OVA-peptide SIINFEKL ( $\alpha$ -GalCer-SIINFEKL/exo). These were injected i.v. in wt recipient mice, and proliferation was assessed after 7 days. Splenic NKT cell proliferation was only observed in response to exosomes loaded with both  $\alpha$ -GalCer and the Ag/Ag-peptide, suggesting  $\alpha$ -GalCer is required for efficient NKT cell proliferation. Up-regulation of the activation marker CD69 was observed already after 1 day, while proliferation proceeded until day 5 after stimulation and was accompanied by IFN $\gamma$  production. IL-4 production was observed only during the first 3 days. iNKT cell activation also led to an early activation and proliferation of DCs, NK, and  $\gamma\delta$  T cells. Our findings suggest that  $\alpha$ GC-loaded exosomes are potent inducers of iNKT cell responses, as well as DC, NK- and  $\gamma\delta$  T-cell activation and proliferation *in vivo*.

$\alpha$ -GalCer/exo stimulation led to increased OVA-specific CD8<sup>+</sup> T-cell proliferation, to a greater extent compared to  $\alpha$ -GalCer-SIINFEKL/exo, which is in line with previous data where SIINFEKL/exo did not sufficiently stimulate proliferation of OVA-specific CD8<sup>+</sup> T cells, due to the lack of B-cell co-stimulation. Moreover,  $\alpha$ -GalCer-OVA/exo immunization generated increased numbers of SIINFEKL-specific IFN $\gamma$  producing cells in a CD1d-dependent manner. These results show that  $\alpha$ -GalCer-loaded exosomes boost Ag-specific CD8<sup>+</sup> T-cell responses via iNKT cells *in vivo* in a CD1d-dependent manner. Additionally,  $\alpha$ -GalCer-OVA/exo immunization induced proliferation of CD4<sup>+</sup> T cells and T follicular helper (T<sub>fh</sub>) cells in an iNKT cell-dependent manner, as the response was lower in CD1d-deficient mice. In line with the role of T<sub>fh</sub> in humoral responses, we detected increased numbers GC B cells and plasma cells and increased levels of OVA-specific IgG2c Abs.

Interestingly, when  $\alpha$ -GalCer-OVA/exo were compared to soluble  $\alpha$ -GalCer and OVA, they were more potent activators of adaptive immune responses as observed by increased activation of  $\gamma\delta$  T cells, CD4<sup>+</sup> T cells, and OVA-specific CD8<sup>+</sup> T cells. A second boost injection of  $\alpha$ -GalCer-OVA/exo significantly augmented GC B-cell responses as reflected in the increased levels of OVA-specific IgG Abs. These results highlight  $\alpha$ -GalCer-loaded exosomes as more potent adjuvants and Ag-delivery systems for the initiation of adaptive immune responses.

An additional advantage to the use of  $\alpha$ -GalCer-OVA/exo in vaccination, was the observed lack of NKT cell anergy induction compared to administration of soluble  $\alpha$ -GalCer. Importantly, in serial vaccinations only  $\alpha$ -GalCer-OVA/exo induced a prolonged second wave IFN $\gamma$ -response, which also resulted in an increase in OVA-specific CD8<sup>+</sup> T cells, when compared to soluble  $\alpha$ -GalCer and OVA vaccination. This suggests that  $\alpha$ -GalCer-OVA/exo have a more potent long-term immunostimulatory effect, making them promising vaccination vehicles.

Based on the previous finding that exosome co-delivery of glycolipid and protein Ag boosted adaptive immune responses without inducing iNKT-cell anergy, we investigated the potential use of  $\alpha$ -GalCer-OVA/exo in tumor immunotherapy. Utilizing an OVA-

expressing B16 melanoma s.c. tumor model we similarly compared the efficiency of  $\alpha$ -GalCer-OVA/exo to induce adaptive anti-tumor responses, relative to independent administration of  $\alpha$ -GalCer and OVA.  $\alpha$ -GalCer-OVA/exo significantly inhibited tumor growth and prolonged the survival of tumor-bearing mice. This was attributed to increased numbers of tumor-infiltrating CD8<sup>+</sup> OVA-specific T cells and increased serum levels of OVA-specific IgG in  $\alpha$ -GalCer-OVA/exo treated mice compared to  $\alpha$ -GalCer plus OVA treated mice. Together these results show that vaccination with  $\alpha$ -GalCer-OVA-loaded exosomes can provide a promising immunotherapeutic strategy to induce potent adaptive anti-tumor responses.

### 2.3 FINAL REFLECTIONS AND FUTURE PERSPECTIVES

Harnessing the inherent potential of the immune system for the therapeutic benefit of cancer patients has long been the goal of intensive research in the field of cancer immunotherapy. As previously mentioned, active immunotherapy poses great challenges but holds greater potential with regards to obtaining durable responses in cancer patients. A plethora of pre-clinical and clinical studies suggest that activation of early innate immune responses appears to be a prerequisite for the induction of potent adaptive immunity with a long-term memory component. Thus, combinatorial treatment approaches targeting multiple components of the TME are more favorable. Therefore, the focus of this thesis has been to identify and evaluate novel targets in the TME, which can be modulated to induce the activation of anti-tumor immune responses and to ultimately constrict tumor growth and metastasis.

Paper I describes a mechanism regulating the interplay between MZMs and MZBs in the spleen, through the use of Abs to scavenger receptor MARCO. This study provides insight into how manipulating this interaction can affect Ag presentation and adaptive immune responses to foreign but also to self-Ags. It also identifies a novel tool to regulate macrophage activation, with implications for future immunotherapeutic approaches aiming to regulate innate immune responses or to increase protective immunity. More specifically, the use of anti-MARCO Ab triggers the release of extracellular ATP by marginal zone macrophages. In the TME, a shift towards a higher extracellular ATP concentration can be crucial as it drives anti-tumor responses. Extracellular ATP is taken up by purinergic P2X7 receptors on DCs, leading to inflammasome activation and increased production of pro-inflammatory cytokines IL-1 $\beta$  and IL-18<sup>363</sup>. IL-1 $\beta$  together with Ag presentation leads to increased NK cell proliferation IFN $\gamma$ -production and activation of CD8<sup>+</sup> tumoricidal T cells. ATP also induces macrophage maturation and M1 polarization, and increased secretion of IFN $\gamma$  and IL-17 by T cells. In contrast, adenosine leads to reduced production of IL-12 and increased levels of IL-6, IL-8, IL-10, TGF- $\beta$  and VEGF by DCs, reduced NK cell cytotoxicity, M2 macrophage polarization, decreased cytotoxic T cell proliferation and effector function, and finally increased generation of T<sub>regs</sub>.

Paper II identifies scavenger receptor MARCO as a specific marker for the immunosuppressive M2 TAM phenotype. MARCO is expressed on TAMs in the TME of mammary carcinoma, melanoma and colon carcinoma tumor models. Use of Abs to target MARCO significantly inhibited tumor growth and metastasis, and increased survival in combination treatments with checkpoint therapy with anti-CTLA-4 Ab. This effect is attributed to reprogramming of TAMs from an immunosuppressive M2 towards an immunostimulatory M1 phenotype characterized by the production of pro-inflammatory cytokines. This is in line with the findings of Paper I, and suggests that anti-MARCO Ab could use the same mechanism of ATP-release to stimulate the polarization of M2 TAMs to M1, thus relieving immunosuppression in the TME. Finally, it identifies MARCO as a clinically relevant target in highly invasive and metastatic subtypes of human breast cancer and melanoma. There, MARCO expression correlates with an M2 TAM gene profile and

increased expression of EMT-associated genes. EMT, as is further discussed in Paper III, is a process involved in the acquisition of migratory properties by tumor cells. This suggests that TAMs, and in particular MARCO-expressing TAMs, can be a link explaining the more invasive phenotypes of those cancers, but also provides a tangible target for immunotherapy. Further studies investigating the relevance of MARCO in other human cancers as well as elucidating the underlying mechanism of action and pathways involved in MARCO-signaling are warranted and will increase our understanding of its role in the TME. These will evaluate new combinatorial immunotherapeutic treatments and provide a solid basis for future translation of these findings into the clinical setting.

Paper III elucidates the mechanism through which TGF- $\beta$ 1, a major immunosuppressive cytokine in the TME and the driving force for EMT, regulates targeted lymphatic metastasis of breast cancer cells. TGF- $\beta$  controls the CCR7/CCL21 chemotactic axis that guides tumor cells to metastasize through the lymphatic system in an EMT-dependent manner. This study highlights TGF- $\beta$ 1 as a candidate for targeted pharmacological inhibition in an approach to restrict lymphatic metastasis of tumor cells. As was already mentioned TGF- $\beta$ 1 is one of the major immunosuppressive cytokines produced by TAMs. Additionally, macrophages have been implicated in the induction of EMT in pancreatic tumor cells tumor cells and promote invasion and metastasis of mammary carcinoma cells by inducing an inflammatory signature that facilitates metastasis <sup>364</sup>. Although metastasis-promoting TAMs may be a different subtype from the immunosuppressive TAMs, combining the knowledge acquired from Paper II and III to treat cancer with anti-MARCO Ab in combination with molecular inhibitors of TGF- $\beta$  may significantly abrogate metastatic disease.

Paper IV explores the use of DC-derived  $\alpha$ -GalCer-loaded exosomes as a vaccination approach. This study demonstrates that CD1d<sup>+</sup> DC-derived exosomes expressing loaded with  $\alpha$ -GalCer can be used as adjuvants and are potent inducers of iNKT, CD8<sup>+</sup> and CD4<sup>+</sup> T and B cell responses that ultimately restrict tumor growth. In a similar manner one could envisage the design of anti-MARCO Ab-carrying exosomes specifically targeting M2 TAMs, containing immunostimulatory receptors or microRNA125, -155 or -378, to induce M1 polarization. Overall, exosomes provide a flexible vehicle for delivery of "messages" which can be customized to the disease setting and the target of interest.

In conclusion, the immunotherapy approaches described in this thesis provide new insight into mechanisms of innate and adaptive immune regulation, identify novel targets to reprogram the immunosuppressive TME, elucidate genetic programs driving lymphatic metastasis, as well as harness endogenous cellular products and amplify their modulatory capacity. Altogether, this thesis aims to contribute with novel insight into the inner workings of several components of our immune system, which can be used in the design of future approaches to cancer immunotherapy.

### 3 POPULAR SCIENTIFIC SUMMARY

#### English

The term cancer was coined by the Father of Medicine, the Greek physician Hippocrates of Kos (460-370 BC), who first used the words "karkinos" and "karkinoma" to describe tumors. In greek, the word karkinos means crab and refers to the similarity observed between the branch-like spread of tumors and the shape of a crab. Although primary cancers can be treated by several different means, it is the metastatic cancer disease that is the ultimate elusive culprit that makes cancer an incurable disease. However, recent advances in the field of cancer immunotherapy, in particular, have lead to new treatment that awaken the patient's own immune system to fight off cancer.

The immune system is a complex network of cells and molecules that are responsible for protecting us against pathogens. In the case of an invasion the immune system recruits specialized cells to the site of invasion in order to mount a defense response to combat the invader. This phenomenon is known as inflammation. Cancers can also mobilize a similar type of inflamantion, recruiting immune cells to sites of tumor growth. However, immune cells in the cancer microenvironment fail to recognize cancer as a threat and instead are sequestered by the cancer to support its growth and metastasis. Strategies to awaken the body's own immune system to fight cancer are therefore of great importance. This thesis focuses on identifying new targetd to activate the immune system to combat cancer.

Paper I, studies two types of immune cells, the macrophages and the B cells, located in the spleen. Macrophages (form the greek: makros large and phagein eat) are the big eaters of our immune system. They are responsible for taking up and discarding bacteria, dead cells, and metabolic biproducts form our blood circulation which would otherwise harm us if they reached large amounts. They are therefore found in strategy location in our bodies where the blood flows through and can be filtered by macrophages, such as the spleen. This janitorial function is mediated, amongst other, by the so called scavenger receptor MARCO, a molecule found on the surface of macrophages. B cells are a type of immune cells that have the capacity to recognize pathigens and generate specific responses against them by producing target-seeking molecule, antibodies. MARCO can however be falsely attacjed by target seeking molecules, antibodies, as is the case for example in the autoimmune disease SLE. This study shows that, simulating SLE by administering antibodies against MARCO has a profound negative effekt on the function of B cells and restricts the specific responses they generate agains pathogens. This may provide an explanation as to why SLE patients suffer from recurrent infections.

While macrophages have an important defense and janitorial fuction, they are also the major culprits of inflammation that drives cancer growth and metastasis. In paper II, we found that some macrophages, in the tumor bed of breast and colon cancer as well as melanoma (skin cancer), express MARCO. Thus, we used the same target-seeking molecules, antibodies, as before to target MARCO on macrophages that are present in the tumor, in order to re-activate them against the tumors. Interestingly, antibodies found a subpopulation of macrophages in the tumor bed. Serial treatments with MARCO antibodies, as well as with antibodies that activate other immune cells, lead to the re-education of macrophages. This way they started producing molecules which activated other immune cells against the tumor, instead of suppressing them. This lead to significantly less tumor growth and metastasis. The presence of MARCO is associated with the most aggressive types of breast cancer and melanoma, which cause metastasis and are hard to treat. This study shows that MARCO is a target that is present in three different types of cancer and that antibodies can be used as a specific tools to target MARCO-macrophages in the tumor. This may provide a useful future cancer therapy.

Cancer cells can grow locally or travel thourgh the blood and lymphatic system to distal sites of the body where they seed metastases. Not all cancer cells become metastatic, but the ones

that do have to activate a particular gene program in order to gain the capacity to migrate. This gene program is normally active during fetal development, when cells need to travel to different sites of the developing foetus in order to form different organs, and is known as epithelia-mesenchymal transition (EMT). The wrongful activation of EMT is attributed to the production of the signaling molecule TGF- $\beta$  by cancer cells and other sequestered immune cells, such as macrophages, in the tumor bed. In paper III we studied the effect of TGF- $\beta$  on breast cancer cells. Indeed breast cancer cells that only grew locally, changed form and gained a migratory capacity because of TGF- $\beta$ . Interestingly they mimicked another type of immune cell, the dendritic cells, whose primary role is to take up pathogens and travel through the lymphatic system to major immune cell hubs, where they present the pathogen and activate specific immune responses against it. In a similar way, TGF- $\beta$  activating of EMT, lead to the guided entry of cancer cells into the lymphatic system and gave them access to travel to distal sites of the body where they could seed new tumors.

Exosomes are small vesicles that are released by many different types of cells in our bodies. They act as a means of communication, transporting various molecular messages between different cells. Because of this they can be used to load specific/custom molecules which can target specific immune cells and different types of immune responses. In paper IV, we used exosomes generated from dendritic cells, which are known to present foreign molecules to activated immune responses. These were loaded with  $\alpha$ -GalCer, an activator of a type of immune cell known as NKT cells. Vaccination using exosomes loaded with  $\alpha$ -GalCer lead to a more efficient activation of NKT cells, but also several other immune cells, than what  $\alpha$ -GalCer alone would have caused. This was particularly important in the tumor setting, where exosomes loaded with  $\alpha$ -GalCer as well as a molecule expressed by the tumor, activated immune cells against the tumor leading to less tumor growth.

In summary, this thesis identifies new targets in the tumor microenvironment, which can be used to improve the design of future cancer therapeutics for the benefit of cancer patients.

### **Svenska**

Ordet cancer kommer ursprungligen ifrån läkekonstens fader, den grekiska läkaren Hippokrates av Kos (460-370 f.Kr.), som först använde termerna "karkinos" och "karkinoma" för att beskriva tumörer. Det grekiska ordet karkinos betyder kräfta och hänvisar till likheten som observerats mellan spridningsgrenarna i tumörer och formen på en kräfta. Även om primärtumörer kan behandlas på flera olika sätt, är det den metastaserande cancersjukdomen som är den ultimata skyldige som gör cancer till en obotlig sjukdom. Men de senaste framstegen i forskningsfältet om cancer immunterapi, i synnerhet, har lett till nya behandlingar som aktiverar patientens egna immunförsvar till att kämpa mot cancer.

Immunsystemet är ett komplext nätverk av celler och molekyler som är ansvariga för att skydda oss mot patogener. När en invasion inträffar rekryterar immunsystemet specialiserade celler till platsen för invasionen sker för att att bekämpa inkräktaren. Detta fenomen kallas inflammation. Cancer kan också mobilisera en liknande typ av inflammation, dvs rekrytera immunceller till platser av tumörtillväxt. Men immunceller i cancer mikromiljön lyckas inte känna igen tumören som ett hot och istället angrips de av tumören för att stödja dess tillväxt och metastasering. Strategier för att väcka kroppens egna immunsystem för att bekämpa cancer är därför av stor betydelse. Denna avhandling fokuserar på att identifiera nya målmolekyler som kan aktivera immunsystemet för att bekämpa cancer.

I studie I, studeras två typer av immunceller, makrofager och B-celler, som befinner sig i mjälten. Makrofager (från grekiska: makros stor och phagein äta) är storätarna i vårt immunsystem. De är ansvariga för att ta upp och gör sig av med bakterier, döda celler och metaboliska biprodukter från blodcirkulationen som annars skulle skada oss om de uppnådde stora mängder. Därför befinner makrofager sig i strategiska positioner i våra kroppar där blodet flyter igenom och kan filtreras, såsom i mjälten. Denna vaktmästerifunktion medieras,

bland annat, av den så kallade scavenger receptor MARCO, en molekyl som finns på makrofagernas yta. B celler är en typ av immunceller som har kapaciteten att känna igen patogener och generera specifika svar mot dem genom att producera målsökande molekyler, antikroppar. MARCO kan dock felaktigt bli attackerad av dessa antikroppar, vilket är fallet till exempel i den autoimmuna sjukdomen SLE. Denna studie visar att simulering av SLE genom att administrera antikroppar mot MARCO har en stark negativ effekt på B cellers funktion och begränsar det specifika svaret som genereras mot patogener. Detta kan ge en förklaring till varför SLE-patienter lider av återkommande infektioner.

Medan makrofager har en viktig försvars- och vaktmästerifunction, de är också de stora skyldiga till inflammationen som driver cancertillväxt och metastasering. I studie II, fann vi att vissa makrofager, i tumörbädden av bröst- och tjocktarmscancer, samt melanom (hudcancer), uttrycker MARCO. Därför använde vi samma antikroppar som tidigare, mot MARCO på makrofager som finns i tumören, för att återaktivera dem mot tumörer. Intressant nog fann dessa antikroppar en subpopulation av makrofager i tumörbädden. En serie behandlingar med MARCO antikroppar, liksom med antikroppar som aktiverar andra immunceller, ledde till omskolning av makrofagerna. På så sätt började de producera molekyler som aktiverade andra immunceller mot tumören, i stället för att undertrycka dem. Detta ledde till significant mindre tumörtillväxt och metastas. Närvaron av MARCO är förknippad med de mest aggressiva typer av bröstcancer och melanom, som orsakar metastaser och är svåra att behandla. Denna studie visar att MARCO är ett mål som finns i tre olika typer av cancer och att antikroppar riktade mot denna kan användas som särskilda verktyg för att omvandla MARCO-makrofager i tumören. Detta kan därför bli en användbar framtida cancerterapi.

Cancerceller kan växa lokalt eller färdas genom blodet och det lymfatiska systemet till avlägsna ställen i kroppen där de utsäder metastaser. Inte alla cancerceller blir metastaserande, men de som gör det måste aktivera ett särskilt genetiskt program för att få förmågan att migrera. Detta genetiska program är normalt aktivt under fosterutvecklingen, då celler behöver resa till olika platser i det växande fostret för att bilda olika organ, och är känt som epitel-mesenkymal övergång (EMÖ). Den felaktiga aktivering av EMÖ i cancer är beroende av produktionen av signaleringsmolekylen TGF- $\beta$  utav cancerceller och andra sekvestrerade immunceller, såsom makrofager, i tumörbädden. I studie III, studerar vi effekten av TGF- $\beta$  på bröstcancerceller. Vi observerade att bröstcancerceller som endast växte lokalt, ändrade form och fick en vandrande kapacitet på grund av TGF- $\beta$ . Intressant nog, härjade de en annan typ av immunceller, de dendritiska cellerna, vars främsta roll är att ta upp patogener och färdas genom det lymfatiska systemet till stora immuncellsnav, där de presenterar patogener och aktiverar specifika immunsvaret mot dem. På ett liknande sätt, leder TGF- $\beta$  aktiverad EMÖ, till en guidad migrering av cancerceller som ger inträde till lymfsystemet och ger dem möjlighet att färdas till avlägsna ställen i kroppen där de kan utsäda nya tumörer.

Exosomer är små blåsor som frigörs av många olika typer av celler i våra kroppar. De fungerar som ett kommunikationsmedel, som transporterar olika molekylära meddelanden mellan olika celler. På grund av detta kan de användas för att laddas med specifika molekyler som kan riktas mot särskilda immunceller och ge upphov till olika typer av immunsvaret. I studie IV, använde vi exosomer som genereras från dendritiska celler, som är kända för att presentera främmande molekyler för att aktivera immunsvaret. Dessa laddades med  $\alpha$ -GalCer, en molekyl som aktiverar en typ av immunceller som kallas NKT celler. Vaccination med hjälp av exosomer laddade med  $\alpha$ -GalCer ledde till en mer effektiv aktivering av NKT celler, men även av flera andra immunceller, än vad  $\alpha$ -GalCer enbart skulle ha orsakat. Detta var särskilt viktigt i tumörmiljön, där exosomer laddade med  $\alpha$ -GalCer samt en molekyl som uttrycks av tumören, aktiverade immunceller mot tumören och ledde till mindre tumörtillväxt.



Sammanfattningsvis, identifierar denna avhandling nya mål i cancernmikromiljön som kan användas för att förbättra framtida cancerterapi till förmån av cancerpatienter.

### **Ελληνικά**

Η λέξη καρκίνος ορίστηκε από τον Πατέρα της Ιατρικής, τον Έλληνα ιατρό Ιπποκράτη τον Κώο (460-370 π.Χ.), ο οποίος χρησιμοποίησε για πρώτη φορά τους όρους "καρκίνος" και "καρκίνωμα" για να περιγράψει όγκους. Ο συμβολισμός βασίζεται στην ομοιότητα που παρατηρείται μεταξύ των διακλαδώσεων εξάπλωσης των όγκων και του σχήματος του καρκίνου. Αν και οι πρωτογενείς όγκοι μπορούν να αντιμετωπιστούν με αρκετούς διαφορετικούς τρόπους, η μεταστατική ασθένεια του καρκίνου είναι ο τελικός ένοχος που καθιστά τον καρκίνο ανίατη νόσο. Ωστόσο, πρόσφατες πρόοδοι ειδικά στον τομέα της ανοσοθεραπείας του καρκίνου, έχουν οδηγήσει στην ανάπτυξη νέων θεραπειών που ξυπνούν το ανοσοποιητικό σύστημα του ασθενούς έτσι ώστε αυτό να καταπολεμήσει τον καρκίνο.

Το ανοσοποιητικό σύστημα είναι ένα πολύπλοκο δίκτυο κυττάρων και μορίων που είναι υπεύθυνα για την προστασία μας έναντι των παθογόνων. Σε περίπτωση παθογόνου εισβολής το ανοσοποιητικό σύστημα στρατολογεί εξειδικευμένα κύτταρα στην συγκεκριμένη τοποθεσία προκειμένου να ενεργοποιήσει μια ανοσοαπόκριση για την καταπολέμηση του εισβολέα. Αυτό το φαινόμενο είναι γνωστό ως φλεγμονή. Ο καρκίνος μπορεί επίσης να κινητοποιήσει έναν παρόμοιο τύπο φλεγμονής, προσελκύνοντας κύτταρα του ανοσοποιητικού συστήματος στους τόπους ανάπτυξης του όγκου. Εντούτοις, τα κύτταρα του ανοσοποιητικού συστήματος στο μικροπεριβάλλον του καρκίνου αποτυγχάνουν να αναγνωρίσουν τον καρκίνο ως απειλή και αντίθετα χειραγωγούνται από τον καρκίνο έτσι ώστε να σθνεισφέρουν στην ανάπτυξη και μετάστασή του. Η ανάπτυξη στρατηγικών για την αφύπνιση του ανοσοποιητικού συστήματος του οργανισμού για την καταπολέμηση του καρκίνου είναι, επομένως, ύψιστης σημασίας. Η συγκεκριμένη διατριβή εστιάζει στον εντοπισμό νέων στόχων για την ενεργοποίηση του ανοσοποιητικού συστήματος ως προς την καταπολέμηση του καρκίνου.

Η πρώτη μελέτη, εστιάζεται σε δύο τύπους κυττάρων του ανοσοποιητικού συστήματος, τα μακροφάγα και τα Β λεμφοκύτταρα, που βρίσκονται στο σπλήνα. Τα μακροφάγα (από το: Μακρός και φαγείν) είναι αδηφάγα κύτταρα του ανοσοποιητικού συστήματος. Είναι υπεύθυνα για την ανάληψη και την απόρριψη βακτηρίων, νεκρών κυττάρων και μεταβολικών παραπροϊόντων από την κυκλοφορία του αίματος, που διαφορετικά θα μας έβλαπταν αν έφταναν μεγάλες ποσότητες. Ως εκ τούτου, βρίσκονται σε στρατηγικές τοποθεσίες, όπου το αίμα εισέρχεται και διηθείται από τα μακροφάγα, όπως ο σπλήνας. Αυτή η καθαριστική λειτουργία των μακροφάγων εξασκεείται, μεταξύ άλλων, από το λεγόμενη υποδοχέα-καθαριστή MARCO, που βρίσκεται στην επιφάνεια των κυττάρων αυτών. Τα Β λεμφοκύτταρα είναι ένας τύπος κυττάρων του ανοσοποιητικού που έχουν την ικανότητα να αναγνωρίζουν και να παράγουν ειδικές αποκρίσεις ενάντια στα παθογόνα, κυρίως μέσω της παραγωγή αντισωμάτων. Ο MARCO μπορεί ωστόσο να προσδεθεί λανθασμένα από αντισώματα, όπως συμβαίνει για παράδειγμα στην αυτοάνοση ασθένεια του συστηματικού ερυθματώδους λύκου (ΣΕΛ). Αυτή η μελέτη δείχνει ότι, η προσομοίωση του ΣΕΛ μέσω της χορήγησης αντισωμάτων εναντίον του MARCO, έχει σοβαρές αρνητικές επιπτώσεις στη λειτουργία των Β λεμφοκυττάρων και περιορίζει τις ειδικές ανοσοαποκρίσεις που εξασκούν ενάντια στα παθογόνα. Αυτό μπορεί να αποτελεί μια εξήγηση ως προς το γιατί οι ασθενείς με ΣΕΛ υποφέρουν από υποτροπιάζουσες λοιμώξεις.

Ενώ τα μακροφάγα έχουν σημαντική αμυντική και καθαριστική λειτουργία, είναι και οι κύριοι ένοχοι της φλεγμονής που οδηγεί στην ανάπτυξη και την μετάσταση του καρκίνου. Στη δεύτερη μελέτη, ανακαλύψαμε ότι ορισμένα μακροφάγα, στον καρκίνο του μαστού και του παχέος εντέρου, καθώς και του μελανώματος (καρκίνου του δέρματος), εκφράζουν τον υποδοχέα MARCO. Έτσι, χρησιμοποιήθηκαν τα ίδια αντισώματα όπως και στην προηγούμενη μελέτη για την στόχευση του MARCO σε μακροφάγα εντός του όγκου, ώστε να ενεργοποιηθούν κατά αυτού. Τα αντισώματα στοχεύουν έναν υποπληθυσμό μακροφάγων

στο καρκινικό περιβάλλον. Σειριακές θεραπείες με αντισώματα κατά του MARCO, καθώς επίσης και με αντισώματα που ενεργοποιούν άλλα κύτταρα του ανοσοποιητικού, οδηγούν στην επανεκπαίδευση των μακροφάγων. Έτσι επάγουν την παραγωγή μορίων τα οποία ενεργοποιούν άλλα κύτταρα του ανοσοποιητικού έναντι του όγκου, αντί να τα καταστέλλουν. Αυτό οδηγεί σε σημαντικά μικρότερη ανάπτυξη και μετάσταση του καρκίνου. Η παρουσία του MARCO συνδέεται με τις πιο επιθετικές μορφές καρκίνου του μαστού και του μελανώματος, οι οποίες προκαλούν μετάσταση. Αυτή η μελέτη αποδεικνύει ότι ο υποδοχέας MARCO είναι ένας νέος μοριακός στόχος, σε τρεις διαφορετικούς τύπους καρκίνου και ότι τα αντισώματα μπορούν να χρησιμοποιηθούν ως εργαλεία για την ειδική στόχευση MARCO-μακροφάγων στον καρκίνο. Ως εκ τούτου, η προκειμένη στρατηγική στόχευσης μπορεί μελλοντικά να αποτελέσει μια χρήσιμη θεραπεία κατά του καρκίνου.

Τα καρκινικά κύτταρα μπορούν να αναπτυχθούν σε τοπικό επίπεδο ή να ταξιδέψουν μέσω του αίματος και του λεμφικού συστήματος σε απόμακρα σημεία του σώματος όπου δημιουργούν μεταστάσεις. Τα μεταστατικά καρκινικά κύτταρα, έχουν ενεργοποιήσει ένα συγκεκριμένο γονιδιακό πρόγραμμα προκειμένου να αποκτήσουν την ικανότητα να μεταναστεύσουν. Το συγκεκριμένο γονιδιακό πρόγραμμα ενεργοποιείται υπό κανονικές συνθήκες κατά τη διάρκεια της εμβρυϊκής ανάπτυξης, όταν τα κύτταρα ταξιδεύουν σε διαφορετικές τοποθεσίες του αναπτυσσόμενου εμβρύου, προκειμένου να σχηματίσουν διάφορα όργανα, και είναι γνωστό ως επιθηλιακή-μεσεγγυματική μετάβαση (EMM). Η λανθασμένη ενεργοποίηση του EMM αποδίδεται στην παραγωγή του μορίου-σηματοδότη TGF- $\beta$  από τα καρκινικά και από άλλα χειραγωγημένα κύτταρα του ανοσοποιητικού, όπως τα μακροφάγα στο καρκινικό μικροπεριβάλλον. Στην Τρίτη μελέτη, ερευνούμε την επίδραση του TGF- $\beta$  στα κύτταρα του καρκίνου του μαστού. Πράγματι, τα καρκινικά κύτταρα που αναπτύχθηκαν μόνο σε τοπικό επίπεδο, άλλαξαν μορφολογία και απέκτησαν μεταναστευτικές ικανότητες λόγω του TGF- $\beta$ . Επίσης, ενστερνίζονται χαρακτηριστικά ενός άλλου τύπου κυττάρων του ανοσοποιητικού, των δενδριτικών κυττάρων, πρωταρχικός ρόλος των οποίων είναι η ανάλυση παθογόνων και η μετακίνηση μέσω του λεμφικού συστήματος σε κομβικά σημεία του ανοσοποιητικού, όπου παρουσιάζουν τα παθογόνα και να ενεργοποιούν ειδικές ανοσοαποκρίσεις. Με παρόμοιο τρόπο, ο TGF- $\beta$  οδηγεί στην είσοδο των καρκινικών κυττάρων στο λεμφικό σύστημα, δίνοντάς τους πρόσβαση σε απόμακρες τοποθεσίες του σώματος όπου μπορούν να συνεισφέρουν στη δημιουργία νέων όγκων.

Τα εξωσώματα είναι μικρά κυστίδια που απελευθερώνονται από πολλούς διαφορετικούς τύπους κυττάρων στο σώμα μας. Δρουν ως μέσο επικοινωνίας, μεταφέροντας μοριακά μηνυτάτα μεταξύ κυττάρων. Λόγω αυτού, μπορούν να χρησιμοποιηθούν για την φόρτωση μορίων τα οποία στοχεύουν συγκεκριμένα κύτταρα του ανοσοποιητικού και διαφορετικού τύπου ανοσοποιητικές αποκρίσεις. Στη μελέτη IV, χρησιμοποιήσαμε εξωσώματα από δενδριτικά κύτταρα, τα οποία είναι γνωστό ότι παρουσιάζουν ξένα μόρια για την ενεργοποίηση ανοσοαντιδράσεων. Αυτά φορτώθηκαν με  $\alpha$ -GalCer, ένα λιπίδιο που αποτελεί ενεργοποιητή ενός τύπου ανοσοκυττάρων, γνωστά ως NKT κύτταρα. Ο εμβολιασμός πειραματοζώων χρησιμοποιώντας εξωσώματα φορτωμένα με  $\alpha$ -GalCer οδηγεί σε μια πιο αποτελεσματική ενεργοποίηση των NKT κυττάρων, καθώς επίσης και άλλων κυττάρων του ανοσοποιητικού συστήματος, από ό, τι θα προκαλούσε απομονωμένα το  $\alpha$ -GalCer. Αυτό αποτελεί ιδιαίτερα σημαντικό εύρημα όσον αφορά τον καρκίνο, καθώς εξωσώματα φορτωμένα με  $\alpha$ -GalCer μαζί με κάποιο μόριο που εκφράζεται από τον καρκινικό όγκο, ενεργοποίησαν κύτταρα του ανοσοποιητικού συστήματος εναντίον του καρκίνου περιορίζοντας την ανάπτυξή του.

Εν κατακλείδι, η συγκεκριμένη διατριβή προσδιορίζει νέους στόχους στο μικροπεριβάλλον του καρκίνου, οι οποίοι μπορούν να χρησιμοποιηθούν για τη βελτίωση του σχεδιασμού μελλοντικών θεραπειών, προς όφελος των ασθενών με καρκίνο.

## 4 ACKNOWLEDGEMENTS

Kungshamra 02.46, November 1<sup>st</sup> 2015

All of this would not have been possible if it weren't for all the amazing people I met along my journey. I would like to say Ευχαριστώ από καρδιάς, Thank you with all my heart, to:

My supervisor, **Micke**, for taking that phone call that brought me into your group as a PhD student, for your enthusiasm and optimism, for creating opportunities, believing in me and encouraging me to explore my independent creative (sometimes crazy) thinking. And for venturing into the field of tumor immunology with me!

My co-supervisor, **Benedict**, for being there for me since the start, for teaching me the basics of immunology and for loving the fun about research. For all history lessons and fun facts!

My co-supervisor, **Ola**, for your ever-positive attitude, your enthusiasm for research and for homemade allergy tests!

**Annika**, for welcoming me into the L2:04 community and for always encouraging people to reach their greatest potential.

I would like to thank all my co-authors and collaborators, in particular: **Jonas Fuxe** and **Mei-Fong Pang**, for a great collaboration in two projects and for the countless hours by the microscope. **Bob Harris**, for a great two-way collaboration and your advice and expertise in MΦ. **Charlotte Rolny**, for your knowledge on TAMs and helpful contribution to my project. **Timo Pikkarainen**, for your help with the elusive MARCO curing my early days in the world of scavengers. **Malin Sund, and Mattias Rantalainen**, your help has been invaluable in interpreting the significance of our findings for the human cancers.

**Jeff Ravetch**, for the opportunity to visit your lab at Rockefeller and for your distinct scientific perspective. **Rony Dahan**, for your invaluable help with my project and for fun times in NYC. **Fubin Li**, for teaching me the magic of cloning, the chinese meaning of greek words and for the candy supply during late days in the lab.

Several people made my stay at Rockefeller University possible and enjoyable. A special thank you to **Patrick Smith**, for making my stay at Rockefeller practically possible, for all the fun stories and for showing me the greek hoods of NYC! **Dave DiLillo** for lymph node excavations, science talk and food talk. **Meghan DiLillo** for the greatest company at Brandy's (I'll be back soon!) and of course all your help with paper work chaos. **Taia Wang** for fun times in NYC and Stockholm, **Jad Maamary** for fun times sharing lebanease culture, **Ruben Peraza** for breakfast discussions on everything and anything and **Prisca Gell** for always wearing a smile. **Stelios Bournazos**, for being my greek buddy in NYC, for chats over an espresso or two and always having an optimized solution to everything! **Nina Papavasiliou** and **Erec Stebbins** for taking me and Adil in for a greek Christmas away from home. **Linda** and **Olta Molla**, for your friendship and fun times in NYC and Stockholm.

**Tracy McGaha**, for welcoming me to Augusta, it was a great experience in the American South, and **Ted Johnsson** for help and expertise in the cancer therapy field.

**Evren**, for telling me to give 200%, for challenging me from the 1<sup>st</sup> day I set foot in your lab and for teaching me even when suffering from a broken back. **Marie & Birgitta**, for nice company at work and great dinners.

**Le Groupe** members, you are all amazing and I want to thank you for all the little things that made my every day these past years, long lab days, lunch-breaks and fikas, AWs, boule tournaments, conferences, MTC pubs and film clubs. **Fredrik**, for (still 8 years later) sharing MARCO-expertise and NYC tips. **Sara**, for giving me valuable insider tips on how things work when I first started in the lab. **Yunying**, for your caustic humor and for drawings of NK cells in tumor sections. **Kajsa**, for being my partner in crime in mission “almost impossible, but we made it!” MARCO, for teaching me to stain spleen sections by drawing animals on top of each other ☺ and for chats over fika. **Mattias**, for your refreshing positive attitude (dä ä fantastiskt bra!) and for helping me not forget how to speak norrländska. **Emma**, for your fun and bubbly character and for always taking care off us, **Eva**, for bringing reinforcements to the cancer immunotherapy front and for always being up for a fika. **Vanessa** and **Silke** for joining forces at the final stretch of my PhD and for hanging in there during my crazy all-day-and-night-long experiments.

The Croatia Crew: **Amanda** (MandaPand) for being my bench n’ Beer buddy, I miss our talks, **Thomas** (Thompa – Yeah Baby!) always being up for a cuba libre and being a good listener, **Carin** (Hurtbullen) for “Heja heja” support and always being so helpful and knowing, and **Anton** for “killer” parties and your alternative humor. That week will last a lifetime and words can/should not describe. Road tripping and Pivo! Anchor-baby! SweetChilOMyayayaya.. These past 2 years, you kept me “(in)sane”. Soon it’s your turn to take the mic!

The WASP-neighbors, **Lisa** for always wearing a smile and for your good scientific input, **Marisa** for your contagious laughter, back-to-back **Anton** (Αντωνάκη), for letting me pull your leg but also kicking back, **Marton** for the being the energizer of the corridor, **Nik** for greek/russian discussions and spoiling us with sweets, **Magda-Liz** for joining forces as CRPs, **Ming**, **Mariana**, **Jamie** and **Paul** for the nice lunch company and outings.

Past and present MTCers for the good old times and the new. Especially **Nicolas R**, **Emma**, **André**, **Frank** and **Danica**. **Nilla’s** and **Gerry’s groups** for being nice corridor-neighbors and for fun MTC pubs. **Maggan**, **Anna-Karin**, **Kenth**, **Helen**, **Torunn**, **Emelie**, **Elin & co.**, at the MTC research facility, **Birgitta** at FlowCyt, **Helene**, **Mia**, **Kristina** and **Lina** at HR/finance, **David**, **Magnus**, **Per** and **Torbjörn** in the service group and **Åsa** at education, for all your help and for making MTC a great workplace.

Legendary **Versailles** office, aforementioned members (T.A.C.K), **Nina** and **Mattias E**, you are an unmatched combination of characters!

Everyone at L2:04 for making it such a great environment! **Ulf** for singing down the lab isles, **Evelina** for delicious cakes and DC collaborations, **Susanne** and **Guro** for engaging conversations during meetings and dinners, **Hans** for your humor and limericks, **Erik** for fun apps and nice photos, **Jonas** for fun facts, **John** for the T cell-side-of-things, **Anne-**

**Laure** for nice company at dinners and grills, **Sang** for teaching me to make dumplings, **Agneta** for company at cell culture, **Inga-Lill & Catharina** for all your help and support, **Gerd** for your happy smile, **Ali** for always being interested to know how things are going, **Lill-Emma** for all the fun going back to being lab mates at Biomed – it’s been a long time!, **Anna Z** for organizing the best lab summer party together, **Marianne and Titti** for finding I’m allergic to...wait for it...mice!, **Neda** for always having a good story to tell, **Tanja** for fun AWs and fika chats. A special thank you to **Malin Winerdal**, for kindly letting me use your beautiful illustrations to make immunology colorful.

**Stefanie** and **Anna A**, for always being there to enjoy a glass of wine, a fika or a nice dinner, relax and talk about anything and everything. **Tiiu** and **Camilla**, for all the fun times in Stockholm, NYC and Boston, for dinners, chats and always a friendly ear. **Patricia, Eduardo** and **Isabelle**, for a heart-warming Thanksgiving in Boston. **Cindy**, a big thank you for being my person in NYC, we had a lot of funny moments I will never forget (pumpkin-flavored coffee upon arrival!), looking forward to many more! **Kiran, Indira** and **Satya**, for being great friends that one can count on in- and outside of the lab. **Arnika**, for corridor chats, fikas and parties, always fun to hang out! **Joanna** my fellow Kosian-half-swede-office-mate for all that we share and for our “lab-καφεδάκια” and **Milind** (honorary greek) for always asking a thoughtful “Τι κάνεις;” and for awesome parties.

**Lindsay & Steven**, my Augusta family, for Southern hospitality, Po’ boys and krispy kreme and for your friendship.

**Frida**, for always being first and most excited at a dinner party! **Maija**, for always being ready to catch a flight to meet and share all the nice moments. **Najla**, for having the biggest heart out there, keep up the good work! **Filip**, for being the 1st person I met at Biomed when I first arrived in Sweden and for your friendship all these years. **Su** for all the memorable moments, and for lately sharing the understanding of the extremely unique status: grass-widow-on-wrong-side-of-Atlantic-trying-to-defend ☺. **Nico**, for your brilliant humor and for being a true friend. **Emilie**, karotaki mou we share so much since 2005, we’re in each others heads! Your friendship and support those past years has meant a lot, I look forward to more cherished memories from the other side of the pond of course not avoiding bubbles!

**Athanasia and Claes**, for greek food and open hearts. **Giorgos & Tina**, for being shiny happy people. **Agaristi**, my first greek friend in Stockholm, for the good old times in MTC back in 2007. **Thodoris**, for nice get-togethers in Stockholm and NYC and at KI conferences. **Dimitris, Sevi & Yakinthos** for being lovely neighbours and friends, for delicious greek dinners and chocolate souflé. **Eirini**, for being you and for bringing laughter even when the sky is grey, I’m counting on you to hopp on a flight to come find me wherever I am.

**Ayse, Mehmet, Ahu, Özge, Duygu** and **Süleyman**, for being great friends to spend time with in Istanbul and abroad, serefe!

**My Kos-ians! Πόπη & Κώστα, Άννα & Σταύρο, Λευτέρη, Κώστα, Ειρήνη, Γιάννη** (ποντικάκι πράσινο δεν πρόλαβα τελικά να φτιάξω), **Λευούδι & Δημήτρη**, όποτε γυρίζω στο νησί είναι σχεδόν σαν να μην πέρασε μια μέρα, εκτός φυσικά από την τριήμερη αναπροσαρμογή στα ελληνικά, αλλά είπαμε...πολλά χρόνια στη Σουηδία! Σύντομα, η βαλίτσα θα πάει ακόμη πιο μακριά. Είστε, όμως, σταθέρες αξίες, κι ας μας χωρίζουν χιλιόμετρα.

Till min familj och nära vänner i Sverige, **Mormor Sigrid & Morfar Erik, Roger, Lena, Johan & André, Inger, Tord, Stefan, Marie, Johan, Tilde, Alfred & Signe**. Tack för all hjälp i starten av denna svenska resa med flytlass från Forsbacka till Flemingsberg, boende på 5\* Lingvalls och Springs och för att ni har funnits där för mig när jag har saknat min familj på Kos. **Mia & Agne** för att ni är så fina inombords och för att ni alltid tittar in när ni har vägarna förbi Stockholm. **Ingrid & Nikos** med familj, ett stort tack för att ni hjälpte mig från första början när jag flyttade till Sverige, för goda middagar i Stockholm och härliga somrar på Kos.

**Ülker, Günay and Fidan** (How are you? Very well thank you!), çok tesekkürler for being my second family and always being there loving and supporting me.

**Tolga**, I met you as "Adil's Bro". Abi, we have worked, partied, laughed, drunk, celebrated, fought, gotten nerdy and probably also cried together. Well guess what, I am "The Yenge" and you're my Bro too, like it or not. Thanks for your friendship and support.

A big thank you to my sister **Alexandra** for being "my Force" and taking care of me for the past months of thesis writing, also for cropping, and layering in photoshop, you're a savior. I couldn't have done it without you.

**Mamma Margaretha & Pappa Stavros. Mamma**, när jag var liten sa du åt mig att sikta mot stjärnorna för då kanske jag iaf kommer halvvägs. Du lärde mig att göra det jag vill, och "så är det" bara. Tack för att ni alltid finns där för mig. Saknar er. **Μπαμπά**, από μικρή μου έλεγες «Μάθε τέχνη κι άστηνε κι άμα πεινάσεις πιάστηνε», έτσι έμαθα τουριστικά, ξένες γλώσσες, μέχρι και χτισίματα μαζί σου, τελικά με κέρδισε η έρευνα! Μου έδειξες ότι αν βάλω κάτι σκοπό, θα το καταφέρω και γι' αυτό σε ευχαριστώ. Σας αγαπώ και μου λείπετε. Ni är bäst!

Last but not least, my husband **Adil Doganay Duru**. Life is kinda like Baris Manco's song that we were listening to yesterday late at night when you stayed up to help and keep me company writing this thesis. "Egri egri, dogru dogru" (straight lines, curved lines), things go fine, but sometimes they go wrong. One, seldom knows what lies in the path ahead, my path unexpectedly, but fortunately, led me to you. The only thing I know for sure is that whenever I need you, you are here for me, to bend the curve together. Thank you for your support and love, I couldn't have done it without you. Let the adventures begin on a new continent, The Moro Mous together again. Seni seviyorum!

The Swedish Cancer Foundation, The Erik and Edith Fernström foundation, the Nicholson fellowship and the Karolinska Institutet research grant for financial support.

## 5 REFERENCES

1. Medzhitov R. Inflammation 2010: new adventures of an old flame. *Cell*. 2010;140(6):771-776.
2. Metschnikoff E. Lecture on Phagocytosis and Immunity. *British medical journal*. 1891;1(1570):213-217.
3. Niederkorn JY. See no evil, hear no evil, do no evil: the lessons of immune privilege. *Nature immunology*. 2006;7(4):354-359.
4. Zandvoort A, Timens W. The dual function of the splenic marginal zone: essential for initiation of anti-TI-2 responses but also vital in the general first-line defense against blood-borne antigens. *Clinical and experimental immunology*. 2002;130(1):4-11.
5. Kraal G, Mebius R. New insights into the cell biology of the marginal zone of the spleen. *International review of cytology*. 2006;250:175-215.
6. Karlsson MC, Guinamard R, Bolland S, Sankala M, Steinman RM, Ravetch JV. Macrophages control the retention and trafficking of B lymphocytes in the splenic marginal zone. *J Exp Med*. 2003;198(2):333-340.
7. Areschoug T, Gordon S. Pattern recognition receptors and their role in innate immunity: focus on microbial protein ligands. *Contributions to microbiology*. 2008;15:45-60.
8. Janeway CA, Jr., Medzhitov R. Innate immune recognition. *Annu Rev Immunol*. 2002;20:197-216.
9. Matzinger P. Tolerance, danger, and the extended family. *Annual review of immunology*. 1994;12:991-1045.
10. Garg AD, Nowis D, Golab J, Vandenabeele P, Krysko DV, Agostinis P. Immunogenic cell death, DAMPs and anticancer therapeutics: an emerging amalgamation. *Biochimica et biophysica acta*. 2010;1805(1):53-71.
11. Eltzschig HK, Sitkovsky MV, Robson SC. Purinergic signaling during inflammation. *N Engl J Med*. 2012;367(24):2322-2333.
12. Kiessling R, Klein E, Pross H, Wigzell H. "Natural" killer cells in the mouse. II. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Characteristics of the killer cell. *European journal of immunology*. 1975;5(2):117-121.
13. Kiessling R, Klein E, Wigzell H. "Natural" killer cells in the mouse. I. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Specificity and distribution according to genotype. *European journal of immunology*. 1975;5(2):112-117.
14. Herberman RB, Nunn ME, Lavrin DH. Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic acid allogeneic tumors. I. Distribution of reactivity and specificity. *International journal of cancer Journal international du cancer*. 1975;16(2):216-229.
15. Herberman RB, Nunn ME, Holden HT, Lavrin DH. Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic and allogeneic tumors. II. Characterization of effector cells. *International journal of cancer Journal international du cancer*. 1975;16(2):230-239.
16. Sutlu T, Alici E. Natural killer cell-based immunotherapy in cancer: current insights and future prospects. *Journal of internal medicine*. 2009;266(2):154-181.
17. Ljunggren HG, Karre K. In search of the 'missing self': MHC molecules and NK cell recognition. *Immunology today*. 1990;11(7):237-244.
18. Bendelac A, Savage PB, Teyton L. The biology of NKT cells. *Annual review of immunology*. 2007;25:297-336.
19. Gumperz JE, Miyake S, Yamamura T, Brenner MB. Functionally distinct subsets of CD1d-restricted natural killer T cells revealed by CD1d tetramer staining. *The Journal of experimental medicine*. 2002;195(5):625-636.
20. Coquet JM, Chakravarti S, Kyriakopoulos K, et al. Diverse cytokine production by NKT cell subsets and identification of an IL-17-producing CD4-NK1.1- NKT cell population. *Proceedings of the National Academy of Sciences of the United States of America*. 2008;105(32):11287-11292.
21. Taraban VY, Martin S, Attfield KE, et al. Invariant NKT cells promote CD8+ cytotoxic T cell responses by inducing CD70 expression on dendritic cells. *Journal of immunology*. 2008;180(7):4615-4620.
22. Crough T, Purdie DM, Okai M, Maksoud A, Nieda M, Nicol AJ. Modulation of human Valpha24(+)Vbeta11(+) NKT cells by age, malignancy and conventional anticancer therapies. *British journal of cancer*. 2004;91(11):1880-1886.
23. Song L, Asgharzadeh S, Salo J, et al. Valpha24-invariant NKT cells mediate antitumor activity via killing of tumor-associated macrophages. *The Journal of clinical investigation*. 2009;119(6):1524-1536.
24. Shimizu K, Kurosawa Y, Taniguchi M, Steinman RM, Fujii S. Cross-presentation of glycolipid from tumor cells loaded with alpha-galactosylceramide leads to potent and long-lived T cell mediated immunity via dendritic cells. *The Journal of experimental medicine*. 2007;204(11):2641-2653.
25. McEwen-Smith RM, Salio M, Cerundolo V. The regulatory role of invariant NKT cells in tumor immunity. *Cancer immunology research*. 2015;3(5):425-435.
26. Parekh VV, Wilson MT, Olivares-Villagomez D, et al. Glycolipid antigen induces long-term natural killer T cell anergy in mice. *The Journal of clinical investigation*. 2005;115(9):2572-2583.
27. Wojno J, Jukes JP, Ghadbane H, et al. Amide analogues of CD1d agonists modulate iNKT-cell-mediated cytokine production. *ACS chemical biology*. 2012;7(5):847-855.
28. Fujii S, Shimizu K, Kronenberg M, Steinman RM. Prolonged IFN-gamma-producing NKT response induced with alpha-galactosylceramide-loaded DCs. *Nature immunology*. 2002;3(9):867-874.
29. Fujii S, Shimizu K, Smith C, Bonifaz L, Steinman RM. Activation of natural killer T cells by alpha-galactosylceramide rapidly induces the full maturation of dendritic cells in vivo and thereby acts as an adjuvant for combined CD4 and CD8 T cell immunity to a coadministered protein. *The Journal of experimental medicine*. 2003;198(2):267-279.
30. Cooper MD, Peterson RD, Good RA. Delineation of the Thymic and Bursal Lymphoid Systems in the Chicken. *Nature*. 1965;205:143-146.
31. Cooper MD, Raymond DA, Peterson RD, South MA, Good RA. The functions of the thymus system and the bursa system in the chicken. *The Journal of experimental medicine*. 1966;123(1):75-102.
32. LeBien TW, Tedder TF. B lymphocytes: how they develop and function. *Blood*. 2008;112(5):1570-1580.

33. Reth M. Antigen receptors on B lymphocytes. *Annual review of immunology*. 1992;10:97-121.
34. Koppel EA, Litjens M, van den Berg VC, van Kooyk Y, Geijtenbeek TB. Interaction of SIGIRR expressed by marginal zone macrophages with marginal zone B cells is essential to early IgM responses against *Streptococcus pneumoniae*. *Molecular immunology*. 2008;45(10):2881-2887.
35. You Y, Myers RC, Freeberg L, et al. Marginal zone B cells regulate antigen capture by marginal zone macrophages. *Journal of immunology*. 2011;186(4):2172-2181.
36. Cinamon G, Zachariah MA, Lam OM, Foss FW, Jr., Cyster JG. Follicular shuttling of marginal zone B cells facilitates antigen transport. *Nature immunology*. 2008;9(1):54-62.
37. Arnon TI, Horton RM, GrigoroVA IL, Cyster JG. Visualization of splenic marginal zone B-cell shuttling and follicular B-cell egress. *Nature*. 2013;493(7434):684-688.
38. Gatto D, Brink R. The germinal center reaction. *The Journal of allergy and clinical immunology*. 2010;126(5):898-907; quiz 908-899.
39. Martin F, Oliver AM, Kearney JF. Marginal zone and B1 B cells unite in the early response against T-independent blood-borne particulate antigens. *Immunity*. 2001;14(5):617-629.
40. Barral P, Sanchez-Nino MD, van Rooijen N, Cerundolo V, Batista FD. The location of splenic NKT cells favours their rapid activation by blood-borne antigen. *EMBO J*. 2012;31(10):2378-2390.
41. DiLillo DJ, Yanaba K, Tedder TF. B cells are required for optimal CD4+ and CD8+ T cell tumor immunity: therapeutic B cell depletion enhances B16 melanoma growth in mice. *J Immunol*. 2010;184(7):4006-4016.
42. de Visser KE, Korets LV, Coussens LM. De novo carcinogenesis promoted by chronic inflammation is B lymphocyte dependent. *Cancer Cell*. 2005;7(5):411-423.
43. Inoue S, Leitner WW, Golding B, Scott D. Inhibitory effects of B cells on antitumor immunity. *Cancer Res*. 2006;66(15):7741-7747.
44. Steinman RM, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. I. Morphology, quantitation, tissue distribution. *The Journal of experimental medicine*. 1973;137(5):1142-1162.
45. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature*. 1998;392(6673):245-252.
46. Steinman RM, Banchereau J. Taking dendritic cells into medicine. *Nature*. 2007;449(7161):419-426.
47. Liu K, Nussenzweig MC. Origin and development of dendritic cells. *Immunological reviews*. 2010;234(1):45-54.
48. Shortman K, Naik SH. Steady-state and inflammatory dendritic-cell development. *Nature reviews Immunology*. 2007;7(1):19-30.
49. Steinman RM. The dendritic cell system and its role in immunogenicity. *Annual review of immunology*. 1991;9:271-296.
50. von Andrian UH, Mempel TR. Homing and cellular traffic in lymph nodes. *Nature reviews Immunology*. 2003;3(11):867-878.
51. Alarcon B, Mestre D, Martinez-Martin N. The immunological synapse: a cause or consequence of T-cell receptor triggering? *Immunology*. 2011;133(4):420-425.
52. Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. *Nature reviews Cancer*. 2012;12(4):265-277.
53. Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nat Rev Immunol*. 2005;5(12):953-964.
54. Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes, macrophages, and dendritic cells. *Science*. 2010;327(5966):656-661.
55. Swirski FK, Nahrendorf M, Etzrodt M, et al. Identification of splenic reservoir monocytes and their deployment to inflammatory sites. *Science*. 2009;325(5940):612-616.
56. Epelman S, Lavine KJ, Randolph GJ. Origin and functions of tissue macrophages. *Immunity*. 2014;41(1):21-35.
57. Hashimoto D, Chow A, Noizat C, et al. Tissue-resident macrophages self-maintain locally throughout adult life with minimal contribution from circulating monocytes. *Immunity*. 2013;38(4):792-804.
58. Haldar M, Kohyama M, So AY, et al. Heme-mediated SPI-C induction promotes monocyte differentiation into iron-recycling macrophages. *Cell*. 2014;156(6):1223-1234.
59. Aichele P, Zinke J, Grode L, Schwendener RA, Kaufmann SH, Seiler P. Macrophages of the splenic marginal zone are essential for trapping of blood-borne particulate antigen but dispensable for induction of specific T cell responses. *Journal of immunology*. 2003;171(3):1148-1155.
60. N AG, Guillen JA, Gallardo G, et al. The nuclear receptor LXRalpha controls the functional specialization of splenic macrophages. *Nat Immunol*. 2013;14(8):831-839.
61. Buiting AM, De Rover Z, Kraal G, Van Rooijen N. Humoral immune responses against particulate bacterial antigens are dependent on marginal metallophilic macrophages in the spleen. *Scandinavian journal of immunology*. 1996;43(4):398-405.
62. Kang YS, Kim JY, Bruening SA, et al. The C-type lectin SIGN-R1 mediates uptake of the capsular polysaccharide of *Streptococcus pneumoniae* in the marginal zone of mouse spleen. *Proceedings of the National Academy of Sciences of the United States of America*. 2004;101(1):215-220.
63. Kang YS, Yamazaki S, Iyoda T, et al. SIGN-R1, a novel C-type lectin expressed by marginal zone macrophages in spleen, mediates uptake of the polysaccharide dextran. *International immunology*. 2003;15(2):177-186.
64. Palecanda A, Paulauskis J, Al-Mutairi E, et al. Role of the scavenger receptor MARCO in alveolar macrophage binding of unopsonized environmental particles. *The Journal of experimental medicine*. 1999;189(9):1497-1506.
65. Mukhopadhyay S, Chen Y, Sankala M, et al. MARCO, an innate activation marker of macrophages, is a class A scavenger receptor for *Neisseria meningitidis*. *European journal of immunology*. 2006;36(4):940-949.
66. Prabagar MG, Do Y, Ryu S, et al. SIGN-R1, a C-type lectin, enhances apoptotic cell clearance through the complement deposition pathway by interacting with C1q in the spleen. *Cell death and differentiation*. 2013;20(4):535-545.
67. Groeneveld PH, Erich T, Kraal G. The differential effects of bacterial lipopolysaccharide (LPS) on splenic non-lymphoid cells demonstrated by monoclonal antibodies. *Immunology*. 1986;58(2):285-290.



68. Anthony RM, Wermeling F, Karlsson MC, Ravetch JV. Identification of a receptor required for the anti-inflammatory activity of IVIG. *Proceedings of the National Academy of Sciences of the United States of America*. 2008;105(50):19571-19578.
69. McGaha TL, Chen Y, Ravishankar B, van Rooijen N, Karlsson MC. Marginal zone macrophages suppress innate and adaptive immunity to apoptotic cells in the spleen. *Blood*. 2011;117(20):5403-5412.
70. Miyake Y, Asano K, Kaise H, Uemura M, Nakayama M, Tanaka M. Critical role of macrophages in the marginal zone in the suppression of immune responses to apoptotic cell-associated antigens. *The Journal of clinical investigation*. 2007;117(8):2268-2278.
71. Wermeling F, Chen Y, Pikkarainen T, et al. Class A scavenger receptors regulate tolerance against apoptotic cells, and autoantibodies against these receptors are predictive of systemic lupus. *The Journal of experimental medicine*. 2007;204(10):2259-2265.
72. Chen XW, Shen Y, Sun CY, Wu FX, Chen Y, Yang CD. Anti-class a scavenger receptor autoantibodies from systemic lupus erythematosus patients impair phagocytic clearance of apoptotic cells by macrophages in vitro. *Arthritis research & therapy*. 2011;13(1):R9.
73. Martinez-Pomares L, Gordon S. CD169+ macrophages at the crossroads of antigen presentation. *Trends in immunology*. 2012;33(2):66-70.
74. Martinez-Pomares L, Gordon S. Antigen presentation the macrophage way. *Cell*. 2007;131(4):641-643.
75. Phan TG, Green JA, Gray EE, Xu Y, Cyster JG. Immune complex relay by subcapsular sinus macrophages and noncognate B cells drives antibody affinity maturation. *Nature immunology*. 2009;10(7):786-793.
76. Asano K, Nabeyama A, Miyake Y, et al. CD169-positive macrophages dominate antitumor immunity by crosspresenting dead cell-associated antigens. *Immunity*. 2011;34(1):85-95.
77. Barral P, Polzella P, Bruckbauer A, et al. CD169(+) macrophages present lipid antigens to mediate early activation of iNKT cells in lymph nodes. *Nature immunology*. 2010;11(4):303-312.
78. Martinez-Pomares L, Kosco-Vilbois M, Darley E, et al. Fc chimeric protein containing the cysteine-rich domain of the murine mannose receptor binds to macrophages from splenic marginal zone and lymph node subcapsular sinus and to germinal centers. *The Journal of experimental medicine*. 1996;184(5):1927-1937.
79. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. *Nature reviews Immunology*. 2008;8(12):958-969.
80. Wermeling F, Karlsson MC, McGaha TL. An anatomical view on macrophages in tolerance. *Autoimmunity reviews*. 2009;9(1):49-52.
81. Coffman RL. Origins of the T(H)1-T(H)2 model: a personal perspective. *Nature immunology*. 2006;7(6):539-541.
82. Mackaness GB. Cellular resistance to infection. *The Journal of experimental medicine*. 1962;116:381-406.
83. Stein M, Keshav S, Harris N, Gordon S. Interleukin 4 potentially enhances murine macrophage mannose receptor activity: a marker of alternative immunologic macrophage activation. *The Journal of experimental medicine*. 1992;176(1):287-292.
84. Doyle AG, Herbein G, Montaner LJ, et al. Interleukin-13 alters the activation state of murine macrophages in vitro: comparison with interleukin-4 and interferon-gamma. *European journal of immunology*. 1994;24(6):1441-1445.
85. Gordon S. Alternative activation of macrophages. *Nature reviews Immunology*. 2003;3(1):23-35.
86. Mills CD, Kincaid K, Alt JM, Heilman MJ, Hill AM. M-1/M-2 macrophages and the Th1/Th2 paradigm. *Journal of immunology*. 2000;164(12):6166-6173.
87. Jenkins SJ, Allen JE. Similarity and diversity in macrophage activation by nematodes, trematodes, and cestodes. *Journal of biomedicine & biotechnology*. 2010;2010:262609.
88. Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nature immunology*. 2010;11(10):889-896.
89. Mantovani A, Sica A, Locati M. Macrophage polarization comes of age. *Immunity*. 2005;23(4):344-346.
90. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends in immunology*. 2002;23(11):549-555.
91. Mantovani A, Biswas SK, Galdiero MR, Sica A, Locati M. Macrophage plasticity and polarization in tissue repair and remodelling. *The Journal of pathology*. 2013;229(2):176-185.
92. Lawrence T, Natoli G. Transcriptional regulation of macrophage polarization: enabling diversity with identity. *Nat Rev Immunol*. 2011;11(11):750-761.
93. Murray PJ, Allen JE, Biswas SK, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity*. 2014;41(1):14-20.
94. Martinez FO, Gordon S. The M1 and M2 paradigm of macrophage activation: time for reassessment. *F1000prime reports*. 2014;6:13.
95. Martinez FO, Gordon S, Locati M, Mantovani A. Transcriptional profiling of the human monocyte-to-macrophage differentiation and polarization: new molecules and patterns of gene expression. *Journal of immunology*. 2006;177(10):7303-7311.
96. Nau GJ, Richmond JF, Schlesinger A, Jennings EG, Lander ES, Young RA. Human macrophage activation programs induced by bacterial pathogens. *Proceedings of the National Academy of Sciences of the United States of America*. 2002;99(3):1503-1508.
97. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends in immunology*. 2004;25(12):677-686.
98. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature*. 2008;454(7203):436-444.
99. Balkwill FR. The chemokine system and cancer. *The Journal of pathology*. 2012;226(2):148-157.
100. Fleetwood AJ, Dinh H, Cook AD, Hertzog PJ, Hamilton JA. GM-CSF- and M-CSF-dependent macrophage phenotypes display differential dependence on type I interferon signaling. *J Leukoc Biol*. 2009;86(2):411-421.

101. Sica A, Porta C, Morlacchi S, et al. Origin and Functions of Tumor-Associated Myeloid Cells (TAMCs). *Cancer microenvironment : official journal of the International Cancer Microenvironment Society*. 2012;5(2):133-149.
102. Park-Min KH, Antoniv TT, Ivashkiv LB. Regulation of macrophage phenotype by long-term exposure to IL-10. *Immunobiology*. 2005;210(2-4):77-86.
103. Hao NB, Lu MH, Fan YH, Cao YL, Zhang ZR, Yang SM. Macrophages in tumor microenvironments and the progression of tumors. *Clinical & developmental immunology*. 2012;2012:948098.
104. Duluc D, Delneste Y, Tan F, et al. Tumor-associated leukemia inhibitory factor and IL-6 skew monocyte differentiation into tumor-associated macrophage-like cells. *Blood*. 2007;110(13):4319-4330.
105. Rodriguez PC, Ochoa AC. T cell dysfunction in cancer: role of myeloid cells and tumor cells regulating amino acid availability and oxidative stress. *Seminars in cancer biology*. 2006;16(1):66-72.
106. Curiel TJ, Coukos G, Zou L, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nature medicine*. 2004;10(9):942-949.
107. Kuang DM, Zhao Q, Peng C, et al. Activated monocytes in peritumoral stroma of hepatocellular carcinoma foster immune privilege and disease progression through PD-L1. *The Journal of experimental medicine*. 2009;206(6):1327-1337.
108. Pluddemann A, Neyen C, Gordon S. Macrophage scavenger receptors and host-derived ligands. *Methods*. 2007;43(3):207-217.
109. Sarrias MR, Gronlund J, Padilla O, Madsen J, Holmskov U, Lozano F. The Scavenger Receptor Cysteine-Rich (SRCR) domain: an ancient and highly conserved protein module of the innate immune system. *Critical reviews in immunology*. 2004;24(1):1-37.
110. Jordo ED, Wermeling F, Chen Y, Karlsson MC. Scavenger receptors as regulators of natural antibody responses and B cell activation in autoimmunity. *Molecular immunology*. 2011;48(11):1307-1318.
111. Kzhyshkowska J, Neyen C, Gordon S. Role of macrophage scavenger receptors in atherosclerosis. *Immunobiology*. 2012;217(5):492-502.
112. Palm NW, Medzhitov R. Pattern recognition receptors and control of adaptive immunity. *Immunological reviews*. 2009;227(1):221-233.
113. Gough PJ, Gordon S. The role of scavenger receptors in the innate immune system. *Microbes and infection / Institut Pasteur*. 2000;2(3):305-311.
114. Kodama T, Doi T, Suzuki H, Takahashi K, Wada Y, Gordon S. Collagenous macrophage scavenger receptors. *Current opinion in lipidology*. 1996;7(5):287-291.
115. Areschoug T, Gordon S. Scavenger receptors: role in innate immunity and microbial pathogenesis. *Cellular microbiology*. 2009;11(8):1160-1169.
116. Canton J, Neculai D, Grinstein S. Scavenger receptors in homeostasis and immunity. *Nat Rev Immunol*. 2013;13(9):621-634.
117. Yu X, Guo C, Fisher PB, Subjeck JR, Wang XY. Scavenger Receptors: Emerging Roles in Cancer Biology and Immunology. *Adv Cancer Res*. 2015;128:309-364.
118. Neyen C, Pluddemann A, Mukhopadhyay S, et al. Macrophage scavenger receptor a promotes tumor progression in murine models of ovarian and pancreatic cancer. *Journal of immunology*. 2013;190(7):3798-3805.
119. Hagemann T, Wilson J, Burke F, et al. Ovarian cancer cells polarize macrophages toward a tumor-associated phenotype. *Journal of immunology*. 2006;176(8):5023-5032.
120. Wang XY, Facciponte J, Chen X, Subjeck JR, Repasky EA. Scavenger receptor-A negatively regulates antitumor immunity. *Cancer research*. 2007;67(10):4996-5002.
121. Bak SP, Walters JJ, Takeya M, Conejo-Garcia JR, Berwin BL. Scavenger receptor-A-targeted leukocyte depletion inhibits peritoneal ovarian tumor progression. *Cancer research*. 2007;67(10):4783-4789.
122. Todt JC, Hu B, Curtis JL. The scavenger receptor SR-A I/II (CD204) signals via the receptor tyrosine kinase Merck during apoptotic cell uptake by murine macrophages. *Journal of leukocyte biology*. 2008;84(2):510-518.
123. Graham DK, DeRyckere D, Davies KD, Earp HS. The TAM family: phosphatidyserine sensing receptor tyrosine kinases gone awry in cancer. *Nat Rev Cancer*. 2014;14(12):769-785.
124. Komohara Y, Takemura K, Lei XF, et al. Delayed growth of EL4 lymphoma in SR-A-deficient mice is due to upregulation of nitric oxide and interferon-gamma production by tumor-associated macrophages. *Cancer science*. 2009;100(11):2160-2166.
125. Komohara Y, Ohnishi K, Kuratsu J, Takeya M. Possible involvement of the M2 anti-inflammatory macrophage phenotype in growth of human gliomas. *J Pathol*. 2008;216(1):15-24.
126. Shabo I, Stal O, Olsson H, Dore S, Svanvik J. Breast cancer expression of CD163, a macrophage scavenger receptor, is related to early distant recurrence and reduced patient survival. *International journal of cancer Journal international du cancer*. 2008;123(4):780-786.
127. Medrek C, Ponten F, Jirstrom K, Leandersson K. The presence of tumor associated macrophages in tumor stroma as a prognostic marker for breast cancer patients. *BMC cancer*. 2012;12:306.
128. Komohara Y, Niino D, Saito Y, et al. Clinical significance of CD163(+) tumor-associated macrophages in patients with adult T-cell leukemia/lymphoma. *Cancer science*. 2013;104(7):945-951.
129. Shabo I, Olsson H, Sun XF, Svanvik J. Expression of the macrophage antigen CD163 in rectal cancer cells is associated with early local recurrence and reduced survival time. *International journal of cancer Journal international du cancer*. 2009;125(8):1826-1831.
130. Elomaa O, Kangas M, Sahlberg C, et al. Cloning of a novel bacteria-binding receptor structurally related to scavenger receptors and expressed in a subset of macrophages. *Cell*. 1995;80(4):603-609.
131. Kraal G, van der Laan LJ, Elomaa O, Tryggvason K. The macrophage receptor MARCO. *Microbes and infection / Institut Pasteur*. 2000;2(3):313-316.
132. Ojala JR, Pikkarainen T, Tuuttila A, Sandalova T, Tryggvason K. Crystal structure of the cysteine-rich domain of scavenger receptor MARCO reveals the presence of a basic and an acidic cluster that both contribute to ligand recognition. *The Journal of biological chemistry*. 2007;282(22):16654-16666.

133. Jozefowski S, Arredouani M, Sulahian T, Kobzik L. Disparate regulation and function of the class A scavenger receptors SR-A/II and MARCO. *Journal of immunology*. 2005;175(12):8032-8041.
134. Jozefowski S, Sulahian TH, Arredouani M, Kobzik L. Role of scavenger receptor MARCO in macrophage responses to CpG oligodeoxynucleotides. *Journal of leukocyte biology*. 2006;80(4):870-879.
135. Mukhopadhyay S, Peiser L, Gordon S. Activation of murine macrophages by *Neisseria meningitidis* and IFN-gamma in vitro: distinct roles of class A scavenger and Toll-like pattern recognition receptors in selective modulation of surface phenotype. *Journal of leukocyte biology*. 2004;76(3):577-584.
136. Pluddemann A, Mukhopadhyay S, Sankala M, et al. SR-A, MARCO and TLRs differentially recognise selected surface proteins from *Neisseria meningitidis*: an example of fine specificity in microbial ligand recognition by innate immune receptors. *Journal of innate immunity*. 2009;1(2):153-163.
137. Casciola-Rosen LA, Anhalt G, Rosen A. Autoantigens targeted in systemic lupus erythematosus are clustered in two populations of surface structures on apoptotic keratinocytes. *The Journal of experimental medicine*. 1994;179(4):1317-1330.
138. Kim SJ, Gershov D, Ma X, Brot N, Elkon KB. Opsonization of apoptotic cells and its effect on macrophage and T cell immune responses. *Annals of the New York Academy of Sciences*. 2003;987:68-78.
139. Chen Y, Pikkarainen T, Elomaa O, et al. Defective microarchitecture of the spleen marginal zone and impaired response to a thymus-independent type 2 antigen in mice lacking scavenger receptors MARCO and SR-A. *Journal of immunology*. 2005;175(12):8173-8180.
140. Rogers NJ, Lees MJ, Gabriel L, et al. A defect in Marco expression contributes to systemic lupus erythematosus development via failure to clear apoptotic cells. *Journal of immunology*. 2009;182(4):1982-1990.
141. Chen Y, Sankala M, Ojala JR, et al. A phage display screen and binding studies with acetylated low density lipoprotein provide evidence for the importance of the scavenger receptor cysteine-rich (SRCR) domain in the ligand-binding function of MARCO. *The Journal of biological chemistry*. 2006;281(18):12767-12775.
142. Sankala M, Brannstrom A, Schulthess T, et al. Characterization of recombinant soluble macrophage scavenger receptor MARCO. *The Journal of biological chemistry*. 2002;277(36):33378-33385.
143. Thakur SA, Hamilton R, Jr., Pikkarainen T, Holian A. Differential binding of inorganic particles to MARCO. *Toxicological sciences : an official journal of the Society of Toxicology*. 2009;107(1):238-246.
144. Arredouani MS, Palecanda A, Koziel H, et al. MARCO is the major binding receptor for unopsonized particles and bacteria on human alveolar macrophages. *Journal of immunology*. 2005;175(9):6058-6064.
145. Hamilton RF, Jr., Thakur SA, Mayfair JK, Holian A. MARCO mediates silica uptake and toxicity in alveolar macrophages from C57BL/6 mice. *The Journal of biological chemistry*. 2006;281(45):34218-34226.
146. Bowdish DM, Sakamoto K, Kim MJ, et al. MARCO, TLR2, and CD14 are required for macrophage cytokine responses to mycobacterial trehalose dimycolate and *Mycobacterium tuberculosis*. *PLoS pathogens*. 2009;5(6):e1000474.
147. Dorrington MG, Roche AM, Chauvin SE, et al. MARCO is required for TLR2- and Nod2-mediated responses to *Streptococcus pneumoniae* and clearance of pneumococcal colonization in the murine nasopharynx. *Journal of immunology*. 2013;190(1):250-258.
148. Bin LH, Nielson LD, Liu X, Mason RJ, Shu HB. Identification of uteroglobin-related protein 1 and macrophage scavenger receptor with collagenous structure as a lung-specific ligand-receptor pair. *Journal of immunology*. 2003;171(2):924-930.
149. Pinheiro da Silva F, Aloulou M, Skurnik D, et al. CD16 promotes *Escherichia coli* sepsis through an FcR gamma inhibitory pathway that prevents phagocytosis and facilitates inflammation. *Nature medicine*. 2007;13(11):1368-1374.
150. Mukhopadhyay S, Varin A, Chen Y, Liu B, Tryggvason K, Gordon S. SR-A/MARCO-mediated ligand delivery enhances intracellular TLR and NLR function, but ligand scavenging from cell surface limits TLR4 response to pathogens. *Blood*. 2011;117(4):1319-1328.
151. Kissick HT, Dunn LK, Ghosh S, Nechama M, Kobzik L, Arredouani MS. The scavenger receptor MARCO modulates TLR-induced responses in dendritic cells. *PLoS one*. 2014;9(8):e104148.
152. Arredouani MS, Kobzik L. The structure and function of marco, a macrophage class A scavenger receptor. *Cellular and molecular biology*. 2004;50 Online Pub:OL657-665.
153. Gomes IN, Palma LC, Campos GO, et al. The scavenger receptor MARCO is involved in *Leishmania major* infection by CBA/J macrophages. *Parasite immunology*. 2009;31(4):188-198.
154. Ghosh S, Gregory D, Smith A, Kobzik L. MARCO regulates early inflammatory responses against influenza: a useful macrophage function with adverse outcome. *American journal of respiratory cell and molecular biology*. 2011;45(5):1036-1044.
155. MacLeod DT, Nakatsuji T, Yamasaki K, Kobzik L, Gallo RL. HSV-1 exploits the innate immune scavenger receptor MARCO to enhance epithelial adsorption and infection. *Nature communications*. 2013;4:1963.
156. van der Laan LJ, Dopp EA, Haworth R, et al. Regulation and functional involvement of macrophage scavenger receptor MARCO in clearance of bacteria in vivo. *Journal of immunology*. 1999;162(2):939-947.
157. van der Laan LJ, Kangas M, Dopp EA, et al. Macrophage scavenger receptor MARCO: in vitro and in vivo regulation and involvement in the anti-bacterial host defense. *Immunology letters*. 1997;57(1-3):203-208.
158. Chen Y, Wermeling F, Sundqvist J, et al. A regulatory role for macrophage class A scavenger receptors in TLR4-mediated LPS responses. *European journal of immunology*. 2010;40(5):1451-1460.
159. Granucci F, Petralia F, Urbano M, et al. The scavenger receptor MARCO mediates cytoskeleton rearrangements in dendritic cells and microglia. *Blood*. 2003;102(8):2940-2947.
160. Pikkarainen T, Brannstrom A, Tryggvason K. Expression of macrophage MARCO receptor induces formation of dendritic plasma membrane processes. *The Journal of biological chemistry*. 1999;274(16):10975-10982.
161. Grolleau A, Misek DE, Kuick R, Hanash S, Mule JJ. Inducible expression of macrophage receptor Marco by dendritic cells following phagocytic uptake of dead cells uncovered by oligonucleotide arrays. *Journal of immunology*. 2003;171(6):2879-2888.

162. Matsushita N, Komine H, Grolleau-Julius A, Pilon-Thomas S, Mule JJ. Targeting MARCO can lead to enhanced dendritic cell motility and anti-melanoma activity. *Cancer immunology, immunotherapy : CII*. 2010;59(6):875-884.
163. Komine H, Kuhn L, Matsushita N, Mule JJ, Pilon-Thomas S. Examination of MARCO activity on dendritic cell phenotype and function using a gene knockout mouse. *PLoS one*. 2013;8(7):e67795.
164. Arredouani MS, Franco F, Imrich A, et al. Scavenger Receptors SR-AI/II and MARCO limit pulmonary dendritic cell migration and allergic airway inflammation. *Journal of immunology*. 2007;178(9):5912-5920.
165. Elshourbagy NA, Li X, Terrett J, et al. Molecular characterization of a human scavenger receptor, human MARCO. *European journal of biochemistry / FEBS*. 2000;267(3):919-926.
166. Elomaa O, Sankala M, Pikkariainen T, et al. Structure of the human macrophage MARCO receptor and characterization of its bacteria-binding region. *The Journal of biological chemistry*. 1998;273(8):4530-4538.
167. Harjunpaa A, Taskinen M, Nykter M, et al. Differential gene expression in non-malignant tumour microenvironment is associated with outcome in follicular lymphoma patients treated with rituximab and CHOP. *British journal of haematology*. 2006;135(1):33-42.
168. Bergamaschi A, Tagliabue E, Sorlie T, et al. Extracellular matrix signature identifies breast cancer subgroups with different clinical outcome. *The Journal of pathology*. 2008;214(3):357-367.
169. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell*. 2010;141(1):39-51.
170. Martinez VG, Moestrup SK, Holmskov U, Mollenhauer J, Lozano F. The conserved scavenger receptor cysteine-rich superfamily in therapy and diagnosis. *Pharmacological reviews*. 2011;63(4):967-1000.
171. Pietras K, Ostman A. Hallmarks of cancer: interactions with the tumor stroma. *Exp Cell Res*. 2010;316(8):1324-1331.
172. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell*. 2000;100(1):57-70.
173. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674.
174. Coussens LM, Pollard JW. Leukocytes in mammary development and cancer. *Cold Spring Harbor perspectives in biology*. 2011;3(3).
175. Gajewski TF, Schreiber H, Fu YX. Innate and adaptive immune cells in the tumor microenvironment. *Nature immunology*. 2013;14(10):1014-1022.
176. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nature reviews Immunology*. 2012;12(4):253-268.
177. Nishikawa H, Sakaguchi S. Regulatory T cells in tumor immunity. *Int J Cancer*. 2010;127(4):759-767.
178. Onizuka S, Tawara I, Shimizu J, Sakaguchi S, Fujita T, Nakayama E. Tumor rejection by in vivo administration of anti-CD25 (interleukin-2 receptor alpha) monoclonal antibody. *Cancer research*. 1999;59(13):3128-3133.
179. Turk MJ, Guevara-Patino JA, Rizzuto GA, Engelhorn ME, Sakaguchi S, Houghton AN. Concomitant tumor immunity to a poorly immunogenic melanoma is prevented by regulatory T cells. *The Journal of experimental medicine*. 2004;200(6):771-782.
180. Galdiero MR, Garlanda C, Jaillon S, Marone G, Mantovani A. Tumor associated macrophages and neutrophils in tumor progression. *Journal of cellular physiology*. 2013;228(7):1404-1412.
181. Granot Z, Henke E, Comen EA, King TA, Norton L, Benezra R. Tumor entrained neutrophils inhibit seeding in the premetastatic lung. *Cancer cell*. 2011;20(3):300-314.
182. Fridlender ZG, Sun J, Kim S, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer cell*. 2009;16(3):183-194.
183. Cortez-Retamozo V, Etzrodt M, Newton A, et al. Origins of tumor-associated macrophages and neutrophils. *Proceedings of the National Academy of Sciences of the United States of America*. 2012;109(7):2491-2496.
184. Eken C, Gasser O, Zenhausern G, Oehri I, Hess C, Schifferli JA. Polymorphonuclear neutrophil-derived ectosomes interfere with the maturation of monocyte-derived dendritic cells. *Journal of immunology*. 2008;180(2):817-824.
185. Jaeger BN, Donadieu J, Cognet C, et al. Neutrophil depletion impairs natural killer cell maturation, function, and homeostasis. *The Journal of experimental medicine*. 2012;209(3):565-580.
186. Roosnek E, Burjanadze M, Dietrich PY, Matthes T, Passweg J, Huard B. Tumors that look for their springtime in APRIL. *Critical reviews in oncology/hematology*. 2009;72(2):91-97.
187. Movahedi K, Guillemins M, Van den Bossche J, et al. Identification of discrete tumor-induced myeloid-derived suppressor cell subpopulations with distinct T cell-suppressive activity. *Blood*. 2008;111(8):4233-4244.
188. Schlecker E, Stojanovic A, Eisen C, et al. Tumor-infiltrating monocytic myeloid-derived suppressor cells mediate CCR5-dependent recruitment of regulatory T cells favoring tumor growth. *Journal of immunology*. 2012;189(12):5602-5611.
189. Huang B, Pan PY, Li Q, et al. Gr-1+CD115+ immature myeloid suppressor cells mediate the development of tumor-induced T regulatory cells and T-cell anergy in tumor-bearing host. *Cancer research*. 2006;66(2):1123-1131.
190. Serafini P, Mgebhoff S, Noonan K, Borrello I. Myeloid-derived suppressor cells promote cross-tolerance in B-cell lymphoma by expanding regulatory T cells. *Cancer research*. 2008;68(13):5439-5449.
191. Coussens LM, Werb Z. Inflammation and cancer. *Nature*. 2002;420(6917):860-867.
192. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet*. 2001;357(9255):539-545.
193. Karin M, Greten FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nature reviews Immunology*. 2005;5(10):749-759.
194. Greten FR, Eckmann L, Greten TF, et al. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell*. 2004;118(3):285-296.
195. Balkwill F, Charles KA, Mantovani A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer cell*. 2005;7(3):211-217.
196. Mahmoud SM, Lee AH, Paish EC, Macmillan RD, Ellis IO, Green AR. Tumour-infiltrating macrophages and clinical outcome in breast cancer. *Journal of clinical pathology*. 2012;65(2):159-163.
197. Zhang Y, Cheng S, Zhang M, et al. High-infiltration of tumor-associated macrophages predicts unfavorable clinical outcome for node-negative breast cancer. *PLoS one*. 2013;8(9):e76147.

198. Campbell MJ, Tonlaar NY, Garwood ER, et al. Proliferating macrophages associated with high grade, hormone receptor negative breast cancer and poor clinical outcome. *Breast cancer research and treatment*. 2011;128(3):703-711.
199. Gwak JM, Jang MH, Kim DI, Seo AN, Park SY. Prognostic value of tumor-associated macrophages according to histologic locations and hormone receptor status in breast cancer. *PLoS one*. 2015;10(4):e0125728.
200. Movahedi K, Laoui D, Gyssemans C, et al. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. *Cancer research*. 2010;70(14):5728-5739.
201. Shand FH, Ueha S, Otsuji M, et al. Tracking of intertissue migration reveals the origins of tumor-infiltrating monocytes. *Proceedings of the National Academy of Sciences of the United States of America*. 2014;111(21):7771-7776.
202. Sica A, Bronte V. Altered macrophage differentiation and immune dysfunction in tumor development. *The Journal of clinical investigation*. 2007;117(5):1155-1166.
203. Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. *Journal of immunology*. 2008;181(8):5791-5802.
204. Bingle L, Brown NJ, Lewis CE. The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. *The Journal of pathology*. 2002;196(3):254-265.
205. Ruffell B, Affara NI, Coussens LM. Differential macrophage programming in the tumor microenvironment. *Trends in immunology*. 2012;33(3):119-126.
206. Kitamura T, Qian BZ, Song D, et al. CCL2-induced chemokine cascade promotes breast cancer metastasis by enhancing retention of metastasis-associated macrophages. *J Exp Med*. 2015;212(7):1043-1059.
207. Schmall A, Al-Tamari HM, Herold S, et al. Macrophage and cancer cell cross-talk via CCR2 and CX3CR1 is a fundamental mechanism driving lung cancer. *Am J Respir Crit Care Med*. 2015;191(4):437-447.
208. Torroella-Kouri M, Silvera R, Rodriguez D, et al. Identification of a subpopulation of macrophages in mammary tumor-bearing mice that are neither M1 nor M2 and are less differentiated. *Cancer research*. 2009;69(11):4800-4809.
209. Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nature reviews Cancer*. 2004;4(1):71-78.
210. Tiemessen MM, Jagger AL, Evans HG, van Herwijnen MJ, John S, Taams LS. CD4+CD25+Foxp3+ regulatory T cells induce alternative activation of human monocytes/macrophages. *Proceedings of the National Academy of Sciences of the United States of America*. 2007;104(49):19446-19451.
211. DeNardo DG, Barreto JB, Andreu P, et al. CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. *Cancer cell*. 2009;16(2):91-102.
212. Popi AF, Lopes JD, Mariano M. Interleukin-10 secreted by B-1 cells modulates the phagocytic activity of murine macrophages in vitro. *Immunology*. 2004;113(3):348-354.
213. Wong SC, Pauax AL, Chittezhath M, et al. Macrophage polarization to a unique phenotype driven by B cells. *European journal of immunology*. 2010;40(8):2296-2307.
214. Andreu P, Johansson M, Affara NI, et al. FcRgamma activation regulates inflammation-associated squamous carcinogenesis. *Cancer cell*. 2010;17(2):121-134.
215. Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S. Cross-talk between myeloid-derived suppressor cells and macrophages subverts tumor immunity toward a type 2 response. *Journal of immunology*. 2007;179(2):977-983.
216. Allavena P, Mantovani A. Immunology in the clinic review series; focus on cancer: tumour-associated macrophages: undisputed stars of the inflammatory tumour microenvironment. *Clinical and experimental immunology*. 2012;167(2):195-205.
217. Mantovani A, Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Current opinion in immunology*. 2010;22(2):231-237.
218. Riboldi E, Porta C, Morlacchi S, Viola A, Mantovani A, Sica A. Hypoxia-mediated regulation of macrophage functions in pathophysiology. *International immunology*. 2013;25(2):67-75.
219. Lucas T, Abraham D, Aharinejad S. Modulation of tumor associated macrophages in solid tumors. *Frontiers in bioscience : a journal and virtual library*. 2008;13:5580-5588.
220. Biswas SK, Gangi L, Paul S, et al. A distinct and unique transcriptional program expressed by tumor-associated macrophages (defective NF-kappaB and enhanced IRF-3/STAT1 activation). *Blood*. 2006;107(5):2112-2122.
221. Ojalvo LS, King W, Cox D, Pollard JW. High-density gene expression analysis of tumor-associated macrophages from mouse mammary tumors. *The American journal of pathology*. 2009;174(3):1048-1064.
222. Schmieder A, Michel J, Schonhaar K, Goerdts S, Schledzewski K. Differentiation and gene expression profile of tumor-associated macrophages. *Seminars in cancer biology*. 2012;22(4):289-297.
223. Bonde AK, Tischler V, Kumar S, Soltermann A, Schwendener RA. Intratumoral macrophages contribute to epithelial-mesenchymal transition in solid tumors. *BMC cancer*. 2012;12:35.
224. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nature medicine*. 2013;19(11):1423-1437.
225. Lewis CE, Pollard JW. Distinct role of macrophages in different tumor microenvironments. *Cancer research*. 2006;66(2):605-612.
226. DeNardo DG, Brennan DJ, Rexhepaj E, et al. Leukocyte complexity predicts breast cancer survival and functionally regulates response to chemotherapy. *Cancer discovery*. 2011;1(1):54-67.
227. Leek RD, Lewis CE, Whitehouse R, Greenall M, Clarke J, Harris AL. Association of macrophage infiltration with angiogenesis and prognosis in invasive breast carcinoma. *Cancer research*. 1996;56(20):4625-4629.
228. Murdoch C, Giannoudis A, Lewis CE. Mechanisms regulating the recruitment of macrophages into hypoxic areas of tumors and other ischemic tissues. *Blood*. 2004;104(8):2224-2234.
229. Talks KL, Turley H, Gatter KC, et al. The expression and distribution of the hypoxia-inducible factors HIF-1alpha and HIF-2alpha in normal human tissues, cancers, and tumor-associated macrophages. *The American journal of pathology*. 2000;157(2):411-421.

230. Squadrito ML, De Palma M. Macrophage regulation of tumor angiogenesis: implications for cancer therapy. *Molecular aspects of medicine*. 2011;32(2):123-145.
231. Doedens AL, Stockmann C, Rubinstein MP, et al. Macrophage expression of hypoxia-inducible factor-1 alpha suppresses T-cell function and promotes tumor progression. *Cancer research*. 2010;70(19):7465-7475.
232. Pucci F, Venneri MA, Biziato D, et al. A distinguishing gene signature shared by tumor-infiltrating Tie2-expressing monocytes, blood "resident" monocytes, and embryonic macrophages suggests common functions and developmental relationships. *Blood*. 2009;114(4):901-914.
233. De Palma M, Lewis CE. Cancer: Macrophages limit chemotherapy. *Nature*. 2011;472(7343):303-304.
234. Stockmann C, Doedens A, Weidemann A, et al. Deletion of vascular endothelial growth factor in myeloid cells accelerates tumorigenesis. *Nature*. 2008;456(7223):814-818.
235. Franklin RA, Liao W, Sarkar A, et al. The cellular and molecular origin of tumor-associated macrophages. *Science*. 2014;344(6186):921-925.
236. Josephs DH, Bax HJ, Karagiannis SN. Tumour-associated macrophage polarisation and re-education with immunotherapy. *Front Biosci (Elite Ed)*. 2015;7:293-308.
237. Loke P, Allison JP. PD-L1 and PD-L2 are differentially regulated by Th1 and Th2 cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2003;100(9):5336-5341.
238. Belai EB, de Oliveira CE, Gasparoto TH, et al. PD-1 blockage delays murine squamous cell carcinoma development. *Carcinogenesis*. 2014;35(2):424-431.
239. Duraiswamy J, Freeman GJ, Coukos G. Therapeutic PD-1 pathway blockade augments with other modalities of immunotherapy T-cell function to prevent immune decline in ovarian cancer. *Cancer research*. 2013;73(23):6900-6912.
240. Kryczek I, Zou L, Rodriguez P, et al. B7-H4 expression identifies a novel suppressive macrophage population in human ovarian carcinoma. *The Journal of experimental medicine*. 2006;203(4):871-881.
241. Ding L, Linsley PS, Huang LY, Germain RN, Shevach EM. IL-10 inhibits macrophage costimulatory activity by selectively inhibiting the up-regulation of B7 expression. *Journal of immunology*. 1993;151(3):1224-1234.
242. Flores Villanueva PO, Reiser H, Stadecker MJ. Regulation of T helper cell responses in experimental murine schistosomiasis by IL-10. Effect on expression of B7 and B7-2 costimulatory molecules by macrophages. *Journal of immunology*. 1994;153(11):5190-5199.
243. Kennedy BC, Showers CR, Anderson DE, et al. Tumor-associated macrophages in glioma: friend or foe? *Journal of oncology*. 2013;2013:486912.
244. Lahat N, Rahat MA, Ballan M, Weiss-Cerem L, Engelmayer M, Bitterman H. Hypoxia reduces CD80 expression on monocytes but enhances their LPS-stimulated TNF-alpha secretion. *Journal of leukocyte biology*. 2003;74(2):197-205.
245. Ostrand-Rosenberg S, Sinha P, Beury DW, Clements VK. Cross-talk between myeloid-derived suppressor cells (MDSC), macrophages, and dendritic cells enhances tumor-induced immune suppression. *Seminars in cancer biology*. 2012;22(4):275-281.
246. Vivier E, Ugolini S, Blaise D, Chabannon C, Brossay L. Targeting natural killer cells and natural killer T cells in cancer. *Nature reviews Immunology*. 2012;12(4):239-252.
247. Shurin GV, Ouellette CE, Shurin MR. Regulatory dendritic cells in the tumor immunoenvironment. *Cancer immunology, immunotherapy : CII*. 2012;61(2):223-230.
248. Murai M, Turovskaya O, Kim G, et al. Interleukin 10 acts on regulatory T cells to maintain expression of the transcription factor Foxp3 and suppressive function in mice with colitis. *Nature immunology*. 2009;10(11):1178-1184.
249. Gorelik L, Flavell RA. Immune-mediated eradication of tumors through the blockade of transforming growth factor-beta signaling in T cells. *Nature medicine*. 2001;7(10):1118-1122.
250. Thomas DA, Massague J. TGF-beta directly targets cytotoxic T cell functions during tumor evasion of immune surveillance. *Cancer cell*. 2005;8(5):369-380.
251. Gorelik L, Fields PE, Flavell RA. Cutting edge: TGF-beta inhibits Th type 2 development through inhibition of GATA-3 expression. *Journal of immunology*. 2000;165(9):4773-4777.
252. Li MO, Sanjabi S, Flavell RA. Transforming growth factor-beta controls development, homeostasis, and tolerance of T cells by regulatory T cell-dependent and -independent mechanisms. *Immunity*. 2006;25(3):455-471.
253. Liu J, Zhang N, Li Q, et al. Tumor-associated macrophages recruit CCR6+ regulatory T cells and promote the development of colorectal cancer via enhancing CCL20 production in mice. *PLoS one*. 2011;6(4):e19495.
254. Rodriguez PC, Zea AH, DeSalvo J, et al. L-arginine consumption by macrophages modulates the expression of CD3 zeta chain in T lymphocytes. *Journal of immunology*. 2003;171(3):1232-1239.
255. Rodriguez PC, Quiceno DG, Zabaleta J, et al. Arginase I production in the tumor microenvironment by mature myeloid cells inhibits T-cell receptor expression and antigen-specific T-cell responses. *Cancer research*. 2004;64(16):5839-5849.
256. Munn DH, Shafizadeh E, Attwood JT, Bondarev I, Pashine A, Mellor AL. Inhibition of T cell proliferation by macrophage tryptophan catabolism. *The Journal of experimental medicine*. 1999;189(9):1363-1372.
257. Munn DH, Mellor AL. IDO and tolerance to tumors. *Trends in molecular medicine*. 2004;10(1):15-18.
258. Munn DH, Mellor AL. The tumor-draining lymph node as an immune-privileged site. *Immunological reviews*. 2006;213:146-158.
259. Fan QM, Jing YY, Yu GF, et al. Tumor-associated macrophages promote cancer stem cell-like properties via transforming growth factor-beta1-induced epithelial-mesenchymal transition in hepatocellular carcinoma. *Cancer letters*. 2014;352(2):160-168.
260. Liu CY, Xu JY, Shi XY, et al. M2-polarized tumor-associated macrophages promoted epithelial-mesenchymal transition in pancreatic cancer cells, partially through TLR4/IL-10 signaling pathway. *Laboratory investigation; a journal of technical methods and pathology*. 2013;93(7):844-854.
261. Singh R, Shankar BS, Sainis KB. TGF-beta1-ROS-ATM-CREB signaling axis in macrophage mediated migration of human breast cancer MCF7 cells. *Cellular signalling*. 2014;26(7):1604-1615.

262. Micalizzi DS, Farabaugh SM, Ford HL. Epithelial-mesenchymal transition in cancer: parallels between normal development and tumor progression. *Journal of mammary gland biology and neoplasia*. 2010;15(2):117-134.
263. Park SY, Gonen M, Kim HJ, Michor F, Polyak K. Cellular and genetic diversity in the progression of in situ human breast carcinomas to an invasive phenotype. *The Journal of clinical investigation*. 2010;120(2):636-644.
264. Sarkar S, Horn G, Moulton K, et al. Cancer development, progression, and therapy: an epigenetic overview. *International journal of molecular sciences*. 2013;14(10):21087-21113.
265. Meacham CE, Morrison SJ. Tumour heterogeneity and cancer cell plasticity. *Nature*. 2013;501(7467):328-337.
266. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *The Journal of clinical investigation*. 2009;119(6):1420-1428.
267. Yang L, Huang J, Ren X, et al. Abrogation of TGF beta signaling in mammary carcinomas recruits Gr-1+CD11b+ myeloid cells that promote metastasis. *Cancer cell*. 2008;13(1):23-35.
268. Dvorak HF. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *The New England journal of medicine*. 1986;315(26):1650-1659.
269. Fuxe J, Karlsson MC. TGF-beta-induced epithelial-mesenchymal transition: a link between cancer and inflammation. *Seminars in cancer biology*. 2012;22(5-6):455-461.
270. Su S, Liu Q, Chen J, et al. A positive feedback loop between mesenchymal-like cancer cells and macrophages is essential to breast cancer metastasis. *Cancer cell*. 2014;25(5):605-620.
271. Fu S, Zhang N, Yopp AC, et al. TGF-beta induces Foxp3 + T-regulatory cells from CD4 + CD25 - precursors. *American journal of transplantation : official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*. 2004;4(10):1614-1627.
272. Moo-Young TA, Larson JW, Belt BA, et al. Tumor-derived TGF-beta mediates conversion of CD4+Foxp3+ regulatory T cells in a murine model of pancreas cancer. *Journal of immunotherapy*. 2009;32(1):12-21.
273. Wyckoff J, Wang W, Lin EY, et al. A paracrine loop between tumor cells and macrophages is required for tumor cell migration in mammary tumors. *Cancer research*. 2004;64(19):7022-7029.
274. Randolph GJ, Angeli V, Swartz MA. Dendritic-cell trafficking to lymph nodes through lymphatic vessels. *Nature reviews Immunology*. 2005;5(8):617-628.
275. Coley WB. The treatment of malignant tumors by repeated inoculations of erysipelas. With a report of ten original cases. 1893. *Clinical orthopaedics and related research*. 1991(262):3-11.
276. Coley WB. The Treatment of Inoperable Sarcoma by Bacterial Toxins (the Mixed Toxins of the Streptococcus erysipelas and the Bacillus prodigiosus). *Proceedings of the Royal Society of Medicine*. 1910;3(Surg Sect):1-48.
277. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. *Nature*. 2011;480(7378):480-489.
278. Kohler G, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature*. 1975;256(5517):495-497.
279. Smith P, DiLillo DJ, Bournazos S, Li F, Ravetch JV. Mouse model recapitulating human Fc gamma receptor structural and functional diversity. *Proceedings of the National Academy of Sciences of the United States of America*. 2012;109(16):6181-6186.
280. Pincetic A, Bournazos S, DiLillo DJ, et al. Type I and type II Fc receptors regulate innate and adaptive immunity. *Nature immunology*. 2014;15(8):707-716.
281. Scott AM, Wolchok JD, Old LJ. Antibody therapy of cancer. *Nature reviews Cancer*. 2012;12(4):278-287.
282. Rabinovich GA, Gabrilovich D, Sotomayor EM. Immunosuppressive strategies that are mediated by tumor cells. *Annual review of immunology*. 2007;25:267-296.
283. DiLillo DJ, Ravetch JV. Fc-Receptor Interactions Regulate Both Cytotoxic and Immunomodulatory Therapeutic Antibody Effector Functions. *Cancer immunology research*. 2015;3(7):704-713.
284. Hamaguchi Y, Xiu Y, Komura K, Nimmerjahn F, Tedder TF. Antibody isotype-specific engagement of Fc gamma receptors regulates B lymphocyte depletion during CD20 immunotherapy. *The Journal of experimental medicine*. 2006;203(3):743-753.
285. Nimmerjahn F, Ravetch JV. Fc gamma receptors: old friends and new family members. *Immunity*. 2006;24(1):19-28.
286. Nimmerjahn F, Ravetch JV. Divergent immunoglobulin g subclass activity through selective Fc receptor binding. *Science*. 2005;310(5753):1510-1512.
287. Lazar GA, Dang W, Karki S, et al. Engineered antibody Fc variants with enhanced effector function. *Proceedings of the National Academy of Sciences of the United States of America*. 2006;103(11):4005-4010.
288. Clynes RA, Towers TL, Presta LG, Ravetch JV. Inhibitory Fc receptors modulate in vivo cytotoxicity against tumor targets. *Nature medicine*. 2000;6(4):443-446.
289. Zhang M, Zhang Z, Garmestani K, et al. Activating Fc receptors are required for antitumor efficacy of the antibodies directed toward CD25 in a murine model of adult t-cell leukemia. *Cancer research*. 2004;64(16):5825-5829.
290. Uchida J, Hamaguchi Y, Oliver JA, et al. The innate mononuclear phagocyte network depletes B lymphocytes through Fc receptor-dependent mechanisms during anti-CD20 antibody immunotherapy. *The Journal of experimental medicine*. 2004;199(12):1659-1669.
291. DiLillo DJ, Ravetch JV. Differential Fc-Receptor Engagement Drives an Anti-tumor Vaccinal Effect. *Cell*. 2015;161(5):1035-1045.
292. Cartron G, Dacheux L, Salles G, et al. Therapeutic activity of humanized anti-CD20 monoclonal antibody and polymorphism in IgG Fc receptor Fc gamma RIIIa gene. *Blood*. 2002;99(3):754-758.
293. Weng WK, Levy R. Two immunoglobulin G fragment C receptor polymorphisms independently predict response to rituximab in patients with follicular lymphoma. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2003;21(21):3940-3947.

294. Weng WK, Czerwinski D, Timmerman J, Hsu FJ, Levy R. Clinical outcome of lymphoma patients after idiotype vaccination is correlated with humoral immune response and immunoglobulin G Fc receptor genotype. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2004;22(23):4717-4724.
295. Musolino A, Naldi N, Bortesi B, et al. Immunoglobulin G fragment C receptor polymorphisms and clinical efficacy of trastuzumab-based therapy in patients with HER-2/neu-positive metastatic breast cancer. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2008;26(11):1789-1796.
296. Bibeau F, Lopez-Crapez E, Di Fiore F, et al. Impact of Fc{gamma}RIIa-Fc{gamma}RIIIa polymorphisms and KRAS mutations on the clinical outcome of patients with metastatic colorectal cancer treated with cetuximab plus irinotecan. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2009;27(7):1122-1129.
297. Koizumi M, Hiasa Y, Kumagi T, et al. Increased B cell-activating factor promotes tumor invasion and metastasis in human pancreatic cancer. *PLoS One*. 2013;8(8):e71367.
298. Elgueta R, Benson MJ, de Vries VC, Wasiuk A, Guo Y, Noelle RJ. Molecular mechanism and function of CD40/CD40L engagement in the immune system. *Immunological reviews*. 2009;229(1):152-172.
299. Li F, Ravetch JV. Inhibitory Fc{gamma} receptor engagement drives adjuvant and anti-tumor activities of agonistic CD40 antibodies. *Science*. 2011;333(6045):1030-1034.
300. Li F, Ravetch JV. A general requirement for Fc{gamma}RIIB co-engagement of agonistic anti-TNFR antibodies. *Cell cycle*. 2012;11(18):3343-3344.
301. Kohrt HE, Colevas AD, Houot R, et al. Targeting CD137 enhances the efficacy of cetuximab. *The Journal of clinical investigation*. 2014;124(6):2668-2682.
302. Postow MA, Harding J, Wolchok JD. Targeting immune checkpoints: releasing the restraints on anti-tumor immunity for patients with melanoma. *Cancer journal*. 2012;18(2):153-159.
303. Walunas TL, Lenschow DJ, Bakker CY, et al. CTLA-4 can function as a negative regulator of T cell activation. *Immunity*. 1994;1(5):405-413.
304. Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. *The Journal of experimental medicine*. 1995;182(2):459-465.
305. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science*. 1996;271(5256):1734-1736.
306. Hurwitz AA, Yu TF, Leach DR, Allison JP. CTLA-4 blockade synergizes with tumor-derived granulocyte-macrophage colony-stimulating factor for treatment of an experimental mammary carcinoma. *Proceedings of the National Academy of Sciences of the United States of America*. 1998;95(17):10067-10071.
307. van Elsas A, Hurwitz AA, Allison JP. Combination immunotherapy of B16 melanoma using anti-cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) and granulocyte/macrophage colony-stimulating factor (GM-CSF)-producing vaccines induces rejection of subcutaneous and metastatic tumors accompanied by autoimmune depigmentation. *The Journal of experimental medicine*. 1999;190(3):355-366.
308. Waitz R, Fasso M, Allison JP. CTLA-4 blockade synergizes with cryoablation to mediate tumor rejection. *Oncoimmunology*. 2012;1(4):544-546.
309. Chambers CA, Kuhns MS, Egen JG, Allison JP. CTLA-4-mediated inhibition in regulation of T cell responses: mechanisms and manipulation in tumor immunotherapy. *Annual review of immunology*. 2001;19:565-594.
310. Quezada SA, Peggs KS, Curran MA, Allison JP. CTLA4 blockade and GM-CSF combination immunotherapy alters the intratumor balance of effector and regulatory T cells. *The Journal of clinical investigation*. 2006;116(7):1935-1945.
311. Freeman GJ, Long AJ, Iwai Y, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *The Journal of experimental medicine*. 2000;192(7):1027-1034.
312. Greenwald RJ, Freeman GJ, Sharpe AH. The B7 family revisited. *Annual review of immunology*. 2005;23:515-548.
313. Buchan SL, Manzo T, Flutter B, et al. OX40- and CD27-Mediated Costimulation Synergizes with Anti-PD-L1 Blockade by Forcing Exhausted CD8+ T Cells To Exit Quiescence. *Journal of immunology*. 2015;194(1):125-133.
314. Linch SN, Redmond WL. Combined OX40 ligation plus CTLA-4 blockade: More than the sum of its parts. *Oncoimmunology*. 2014;3:e28245.
315. Le Mercier I, Chen W, Lines JL, et al. VISTA Regulates the Development of Protective Antitumor Immunity. *Cancer research*. 2014;74(7):1933-1944.
316. Wang L, Rubinstein R, Lines JL, et al. VISTA, a novel mouse Ig superfamily ligand that negatively regulates T cell responses. *The Journal of experimental medicine*. 2011;208(3):577-592.
317. Goldberg MV, Drake CG. LAG-3 in Cancer Immunotherapy. *Current topics in microbiology and immunology*. 2011;344:269-278.
318. Triebel F, Jitsukawa S, Baixeras E, et al. LAG-3, a novel lymphocyte activation gene closely related to CD4. *The Journal of experimental medicine*. 1990;171(5):1393-1405.
319. Woo SR, Turnis ME, Goldberg MV, et al. Immune inhibitory molecules LAG-3 and PD-1 synergistically regulate T-cell function to promote tumoral immune escape. *Cancer research*. 2012;72(4):917-927.
320. Fourcade J, Sun Z, Benallaoua M, et al. Upregulation of Tim-3 and PD-1 expression is associated with tumor antigen-specific CD8+ T cell dysfunction in melanoma patients. *The Journal of experimental medicine*. 2010;207(10):2175-2186.
321. Fourcade J, Sun Z, Pagliano O, et al. PD-1 and Tim-3 regulate the expansion of tumor antigen-specific CD8(+) T cells induced by melanoma vaccines. *Cancer research*. 2014;74(4):1045-1055.
322. Derre L, Rivals JP, Jandus C, et al. BTLA mediates inhibition of human tumor-specific CD8+ T cells that can be partially reversed by vaccination. *The Journal of clinical investigation*. 2010;120(1):157-167.
323. Qian BZ, Li J, Zhang H, et al. CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. *Nature*. 2011;475(7355):222-225.



324. Zeisberger SM, Odermatt B, Marty C, Zehnder-Fjallman AH, Ballmer-Hofer K, Schwendener RA. Clodronate-liposome-mediated depletion of tumour-associated macrophages: a new and highly effective antiangiogenic therapy approach. *British journal of cancer*. 2006;95(3):272-281.
325. Luo Y, Zhou H, Krueger J, et al. Targeting tumor-associated macrophages as a novel strategy against breast cancer. *The Journal of clinical investigation*. 2006;116(8):2132-2141.
326. Guiducci C, Vicari AP, Sangaletti S, Trinchieri G, Colombo MP. Redirecting in vivo elicited tumor infiltrating macrophages and dendritic cells towards tumor rejection. *Cancer research*. 2005;65(8):3437-3446.
327. Watkins SK, Egilmez NK, Suttles J, Stout RD. IL-12 rapidly alters the functional profile of tumor-associated and tumor-infiltrating macrophages in vitro and in vivo. *Journal of immunology*. 2007;178(3):1357-1362.
328. Kerkar SP, Goldszmid RS, Muranski P, et al. IL-12 triggers a programmatic change in dysfunctional myeloid-derived cells within mouse tumors. *The Journal of clinical investigation*. 2011;121(12):4746-4757.
329. Chmielewski M, Kopecky C, Hombach AA, Abken H. IL-12 release by engineered T cells expressing chimeric antigen receptors can effectively Muster an antigen-independent macrophage response on tumor cells that have shut down tumor antigen expression. *Cancer research*. 2011;71(17):5697-5706.
330. Hagemann T, Lawrence T, McNeish I, et al. "Re-educating" tumor-associated macrophages by targeting NF-kappaB. *The Journal of experimental medicine*. 2008;205(6):1261-1268.
331. Kodumudi KN, Woan K, Gilvary DL, Sahakian E, Wei S, Djeu JY. A novel chemoimmunomodulating property of docetaxel: suppression of myeloid-derived suppressor cells in tumor bearers. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2010;16(18):4583-4594.
332. Rolny C, Mazzone M, Tugues S, et al. HRG inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through downregulation of PlGF. *Cancer cell*. 2011;19(1):31-44.
333. Weiss JM, Ridnour LA, Back T, et al. Macrophage-dependent nitric oxide expression regulates tumor cell detachment and metastasis after IL-2/anti-CD40 immunotherapy. *The Journal of experimental medicine*. 2010;207(11):2455-2467.
334. Beatty GL, Chiorean EG, Fishman MP, et al. CD40 agonists alter tumor stroma and show efficacy against pancreatic carcinoma in mice and humans. *Science*. 2011;331(6024):1612-1616.
335. Priceman SJ, Sung JL, Shaposhnik Z, et al. Targeting distinct tumor-infiltrating myeloid cells by inhibiting CSF-1 receptor: combating tumor evasion of antiangiogenic therapy. *Blood*. 2010;115(7):1461-1471.
336. Pyonteck SM, Akkari L, Schuhmacher AJ, et al. CSF-1R inhibition alters macrophage polarization and blocks glioma progression. *Nature medicine*. 2013;19(10):1264-1272.
337. Mitchem JB, Brennan DJ, Knolhoff BL, et al. Targeting tumor-infiltrating macrophages decreases tumor-initiating cells, relieves immunosuppression, and improves chemotherapeutic responses. *Cancer research*. 2013;73(3):1128-1141.
338. Zhu Y, Knolhoff BL, Meyer MA, et al. CSF1/CSF1R blockade reprograms tumor-infiltrating macrophages and improves response to T-cell checkpoint immunotherapy in pancreatic cancer models. *Cancer research*. 2014;74(18):5057-5069.
339. Mok S, Koya RC, Tsui C, et al. Inhibition of CSF-1 receptor improves the antitumor efficacy of adoptive cell transfer immunotherapy. *Cancer research*. 2014;74(1):153-161.
340. Zitvogel L, Regnault A, Lozier A, et al. Eradication of established murine tumors using a novel cell-free vaccine: dendritic cell-derived exosomes. *Nature medicine*. 1998;4(5):594-600.
341. Montecalvo A, Shufesky WJ, Stolz DB, et al. Exosomes as a short-range mechanism to spread alloantigen between dendritic cells during T cell allorecognition. *J Immunol*. 2008;180(5):3081-3090.
342. Valenti R, Huber V, Filipazzi P, et al. Human tumor-released microvesicles promote the differentiation of myeloid cells with transforming growth factor-beta-mediated suppressive activity on T lymphocytes. *Cancer Res*. 2006;66(18):9290-9298.
343. Fabbri M, Paone A, Calore F, et al. MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. *Proc Natl Acad Sci U S A*. 2012;109(31):E2110-2116.
344. Peinado H, Aleckovic M, Lavotshkin S, et al. Melanoma exosomes educate bone marrow progenitor cells toward a prometastatic phenotype through MET. *Nat Med*. 2012;18(6):883-891.
345. Hood JL, San RS, Wickline SA. Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis. *Cancer Res*. 2011;71(11):3792-3801.
346. Andre F, Scharzt NE, Movassagh M, et al. Malignant effusions and immunogenic tumour-derived exosomes. *Lancet*. 2002;360(9329):295-305.
347. Qazi KR, Gehrman U, Domange Jordo E, Karlsson MC, Gabrielsson S. Antigen-loaded exosomes alone induce Th1-type memory through a B-cell-dependent mechanism. *Blood*. 2009;113(12):2673-2683.
348. Robbins PD, Morelli AE. Regulation of immune responses by extracellular vesicles. *Nat Rev Immunol*. 2014;14(3):195-208.
349. Lo Cicero A, Stahl PD, Raposo G. Extracellular vesicles shuffling intercellular messages: for good or for bad. *Curr Opin Cell Biol*. 2015;35:69-77.
350. Fidler IJ, Hart IR. Biological diversity in metastatic neoplasms: origins and implications. *Science*. 1982;217(4564):998-1003.
351. Talmadge JE, Singh RK, Fidler IJ, Raz A. Murine models to evaluate novel and conventional therapeutic strategies for cancer. *The American journal of pathology*. 2007;170(3):793-804.
352. Fidler IJ, Gersten DM, Budmen MB. Characterization in vivo and in vitro of tumor cells selected for resistance to syngeneic lymphocyte-mediated cytotoxicity. *Cancer research*. 1976;36(9 pt.1):3160-3165.
353. Fidler IJ, Bucana C. Mechanism of tumor cell resistance to lysis by syngeneic lymphocytes. *Cancer research*. 1977;37(11):3945-3956.
354. Hara I, Takechi Y, Houghton AN. Implicating a role for immune recognition of self in tumor rejection: passive immunization against the brown locus protein. *The Journal of experimental medicine*. 1995;182(5):1609-1614.
355. Li M, Xu F, Muller J, Hearing VJ, Gorelik E. Ecotropic C-type retrovirus of B16 melanoma and malignant transformation of normal melanocytes. *International journal of cancer Journal international du cancer*. 1998;76(3):430-436.
356. Bohm W, Thoma S, Leithauser F, Moller P, Schirmbeck R, Reimann J. T cell-mediated, IFN-gamma-facilitated rejection of murine B16 melanomas. *Journal of immunology*. 1998;161(2):897-908.

357. Dexter DL, Kowalski HM, Blazar BA, Fligiel Z, Vogel R, Heppner GH. Heterogeneity of tumor cells from a single mouse mammary tumor. *Cancer research*. 1978;38(10):3174-3181.
358. Aslakson CJ, Miller FR. Selective events in the metastatic process defined by analysis of the sequential dissemination of subpopulations of a mouse mammary tumor. *Cancer research*. 1992;52(6):1399-1405.
359. Corbett TH, Griswold DP, Jr., Roberts BJ, Peckham JC, Schabel FM, Jr. Tumor induction relationships in development of transplantable cancers of the colon in mice for chemotherapy assays, with a note on carcinogen structure. *Cancer research*. 1975;35(9):2434-2439.
360. Fialka I, Schwarz H, Reichmann E, Oft M, Busslinger M, Beug H. The estrogen-dependent c-JunER protein causes a reversible loss of mammary epithelial cell polarity involving a destabilization of adherens junctions. *The Journal of cell biology*. 1996;132(6):1115-1132.
361. Oft M, Peli J, Rudaz C, Schwarz H, Beug H, Reichmann E. TGF-beta1 and Ha-Ras collaborate in modulating the phenotypic plasticity and invasiveness of epithelial tumor cells. *Genes & development*. 1996;10(19):2462-2477.
362. Zanin RF, Braganhol E, Bergamin LS, et al. Differential macrophage activation alters the expression profile of NTPDase and ecto-5'-nucleotidase. *PLoS One*. 2012;7(2):e31205.
363. Ghiringhelli F, Apetoh L, Tesniere A, et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1beta-dependent adaptive immunity against tumors. *Nature medicine*. 2009;15(10):1170-1178.
364. Qian BZ, Zhang H, Li J, et al. FLT1 signaling in metastasis-associated macrophages activates an inflammatory signature that promotes breast cancer metastasis. *J Exp Med*. 2015;212(9):1433-1448.