

1 **Variable neuroendocrine responses to ecologically-relevant challenges in sticklebacks**

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ABSTRACT

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Variable neuroendocrine responses to ecologically-relevant challenges in sticklebacks.
PHYSIOL BEHAV 00(0) 000-000, 2006. Here, we compare the behavioral, endocrine and
neuroendocrine responses of individual sticklebacks exposed to either an unfamiliar conspecific
or to a predator. We found that the two stressors elicited a similar hypothalamic-pituitary-
interrenal response as assessed by whole-body concentrations of immunoreactive corticosteroids,
but produced quite different patterns of change in brain monoamine and monoamine metabolite
content as assessed by concentrations of serotonin (5-HT), dopamine (DA), norepinephrine (NE)
and the monoamine metabolites 5-hydroxyindole acetic acid (5-HIAA), homovanillic acid
(HVA) and 3-4-dihydroxyphenylacetic acid (DOPAC). For example, relative to baseline levels,
NE levels were elevated in individuals exposed to a predator but were lower in individuals
confronted by a challenging conspecific. Levels of monoamine neurotransmitters in specific
regions of the brain showed extremely close links with behavioral characteristics. Frequency of
attacking a conspecific and inspecting a predator were both positively correlated with
concentrations of NE. However, whereas serotonin was negatively correlated with frequency of
attacking a conspecific, it was positively associated with predator inspection. The data indicate
that the qualitative and quantitative nature of the neuroendocrine stress response of sticklebacks
varies according to the nature of the stressor, and that interindividual variation in behavioural
responses to challenge are reflected by neuroendocrine differences.

Key words: stickleback, *Gasterosteus aculeatus*, behavioral syndromes, antipredator
behavior, aggression, glucocorticoid, serotonin, stress, coping styles, individual differences

Running head: Individual differences in sticklebacks

47 INTRODUCTION

48

49 Both attacking a conspecific and confronting a potential predator are dangerous. In
50 addition to energetic costs [1], aggression can result in injury [2] and exposure to predation risk
51 while fighting [3]. Similarly, an encounter with a potential predator can impose energetic costs of
52 escape [4], injury [5] or even death. Not surprisingly, both confrontation by a challenging
53 conspecific [6-11] and exposure to a predator [12-15] elicit a neuroendocrine stress response.

54 The neuroendocrine stress response involves a coordinated activation of both the
55 hypothalamic-pituitary-adrenal (or interrenal, in the case of fishes, HPI) axis and the brain
56 monoamine neurotransmitter systems [16]. When a stimulus evokes a stress response, both
57 systems are activated by the same central mechanism, resulting in the elevation of plasma
58 corticosteroids and brain monoaminergic activity. In general, exposure to stressors is associated
59 with increased concentrations of plasma glucocorticoids and increased turnover of 5-HT to 5-
60 hydroxyindoleacetic acid (5-HIAA) [17].

61 Individual differences in behavior are often related to individual differences along both
62 axes of the stress response [18-22]. With respect to the HPA axis, individual differences in
63 aggressiveness are negatively correlated with concentrations of plasma glucocorticoids in trout
64 [23] and chickens [24]. In humans, individual differences in behaviors that are analogous to risk-
65 taking behaviors and aggression are associated with increased norepinephrine and dopamine
66 activity [25,26]. Finally, aggression and risk-taking behaviors in several species have been
67 linked to serotonin turnover. For example, individual differences in aggression are negatively
68 related to serotonin turnover in monkeys [24,27-29], trout [21] and anolis lizards [30-32].
69 However, the relationship between 5-HT, stress, the HPI axis and aggression is complex and

70 depends on the duration of the stressor. For example, in salmonids, 5-HT turnover is usually
71 positively associated with plasma ACTH and cortisol concentrations and negatively associated
72 with aggression. However, long-term stimulation of the serotonergic system has inhibitory
73 (negative) effects on the HPI axis [33] and aggression [17].

74 In previous work, we have shown that behavioral reactions to predators and competing
75 conspecifics covary at the individual level in threespined sticklebacks (*Gasterosteus aculeatus*)
76 [34-36]. While some individuals are willing to engage in behavior that appears to be dangerous,
77 such as foraging under predation risk or performing predator inspection, other individuals are
78 much more cautious around predators. Individuals that take more risks in this context are also
79 more aggressive toward conspecifics. Covariance among suites of behavioral traits is common
80 [37,38] and in several species the shy-bold continuum and the proactive-reactive axis have been
81 associated with individual differences in stress responsiveness [39]. Therefore it is possible that
82 differences in how individual sticklebacks respond to dangerous situations might be linked with
83 differences in the stress response.

84 Here, we investigated natural variation in behavioral, glucocorticoid and monoamine
85 responses of individual sticklebacks to two potentially dangerous situations. We wished to
86 establish whether wild-caught animals responding to ecologically-relevant challenges show
87 stress responses that are comparable in nature and extent to those described for laboratory
88 animals, and whether the stress response might be an underlying root of the covariance of
89 behavioral responses in sticklebacks. With this in mind, we exposed individuals to either an
90 unfamiliar conspecific or to a potential predator and recorded their behavior. Although the
91 danger of predation is greater than the danger posed by a territorial intrusion, we hypothesized
92 that both situations would induce a stress response because social stress is one of the most

93 effective stressors in inducing a high magnitude response in other animals [40]. We sampled
94 individuals at 15, 30 or 60 minutes after exposure to determine the time course of the
95 glucocorticoid and monoaminergic responses to these two threats. This design allowed us not
96 only to follow the neuroendocrine responses to these stressors through time, but also to
97 determine whether individual differences in behavioral responses to these challenges could be
98 related to underlying neuroendocrine physiology.

99

100 **METHODS**

101

102 **Overview:** Individuals were presented with one of two potential threats, either an
103 unfamiliar conspecific or a predator, hereafter referred to as ‘conspecific’ and ‘predator’,
104 respectively, and their behavior was recorded. Individuals exposed to the ‘conspecific’ or the
105 ‘predator’ were subdivided into three different treatment groups, sacrificed 15, 30 or 60 minutes
106 after exposure to the potentially threatening stimulus. Individuals were randomly assigned to a
107 treatment group prior to observing their behavior. The responses to the stressors were compared
108 across time periods and against a ‘baseline control’ group, which consisted of individuals
109 sampled directly from an undisturbed stock tank. Each treatment group comprised ten
110 individuals.

111 Subadult sticklebacks were collected from the River Endrick in January 2004 and brought
112 to the Glasgow University Field Station, Rowardennan, where all of the behavioral observations
113 were carried out. Groups of fish (n=10-40) were maintained in flow-through stock tanks (210
114 liters) at $9 \pm 2^\circ$ C and on a 14L:10D photoperiod. Fish were fed frozen bloodworms *ad libitum*
115 daily except on the day of observation, when they were unfed.

116 Behavioral observations took place in March and April 2004 in a U-shaped flume with a
117 live pike (*Esox lucius*) in either arm of the flume. Aquaria that were used for behavioral
118 observation ('observation tanks', 44 liters, 61x32x22 cm) were placed inside the flume and next
119 to a window in the flume so that the behavior of the fish could be observed. The window was
120 covered by a blind with a small opening which allowed the observer to see through the window
121 with minimal disturbance to the fish. Each observation tank contained a one-liter glass conical
122 flask, a plastic plant and a length of opaque tube (12 cm diameter, 36 cm tall) that stood
123 vertically on one side of the tank and allowed fish to be introduced into the tank with a minimum
124 of disturbance. Exterior lines on the tanks divided them into 16 equally-sized areas.

125 Each arm of the flume contained one of two live pike (46, 41cm standard length) and
126 cloth plants which served as hiding places for the pike. The compartments were fitted with a
127 removable opaque cover which created a dark, shaded area for the pike. The pike were caught by
128 hook and line in February 2004 in a small water body near the Glasgow University Field Station
129 (the Duibh Lochan). The two pike were fed dead minnows and dead sticklebacks *ad libitum*.

130

131 Procedure:

132 Fish were removed from the stock tank and placed into a settling tank (49 liters,
133 61x31x26 cm) for two nights in order to acclimate to the flume. After the acclimation period,
134 sticklebacks were netted from the settling tank and were randomly assigned to one of eight
135 treatments (see below for a description of the different treatments). The stickleback was
136 deposited into the tube in an observation tank. After 15 minutes, the tube was lifted, which
137 allowed the stickleback to swim freely around the tank. After another 15 minutes, the fish was

138 presented with either an unfamiliar conspecific or a pike, and the behavioral observation began.
139 Behavioral observations of response to an unfamiliar conspecific and predator were alternated.

140

141 Treatments:

142 **Unfamiliar conspecific:** We employed a procedure that was designed to simulate a
143 challenge to the resident fish by an intruding conspecific. Sticklebacks at this size and age (0.373
144 \pm 0.02 g, approximately 7-8 months of age) are not breeding and so do not defend breeding
145 territories, but they do display aggressive behavior during competition for food and other
146 resources and can be territorial [41]. Therefore we interpret the behavioural response of
147 sticklebacks to the unfamiliar conspecific in this experiment as a response to a potential
148 competitor for food and/or space. It is also worth considering that the sticklebacks' response to a
149 conspecific might also reflect an affiliative motivation because they were held in isolation.

150 A live conspecific (within 5mm standard length of the resident) was placed into the flask
151 in the observation tank. Seven different conspecifics were used as intruders throughout the
152 experiment. A fish was never used as an intruder more than once consecutively. The flask
153 effectively standardized the behavior of the intruder by minimizing movement. The frequency of
154 attacking the conspecific (biting) was recorded for 15 minutes after the resident first oriented to
155 the conspecific because some individuals were facing away from the flask when the intruder was
156 introduced. Latency to orient to the intruder ranged from 0.4-482.0 seconds (mean=104.6 \pm 24.7
157 s). This procedure is roughly analogous to studies with trout where a resident is challenged by an
158 intruder [23]. However, an important difference is that in the present case there is no physical
159 contact between the resident and intruder and the intruder cannot escape. We elected to use this
160 procedure to minimize stress to the intruder. After the behavioral observation, the flask

161 containing the conspecific was removed from the tank and the resident fish was sacrificed
162 according to treatment (15 minutes, 30 minutes or 60 minutes after the behavioral observation
163 was completed).

164 **Predator:** This procedure was designed to simulate a potential predatory threat by a live
165 pike. We lured the pike into a chamber situated next to the observation tank by removing cover
166 over the pike. In general, the pike willingly swam into the chamber, seeking cover. A removable
167 opaque divider was situated between the observation aquarium and the predator chamber. To
168 start the behavioural observation, the divider separating the observation aquarium from the
169 chamber was gently lifted, allowing the stickleback a clear view of the pike on the other side of
170 the glass. The behavior of the individual stickleback was observed for 15 minutes after the
171 divider was removed and the following behaviors were recorded: predator inspection (swimming
172 next to and orienting to the mouth of the pike) and time orienting (body facing toward the pike).
173 Whether the pike moved or oriented to the stickleback during the observation was also recorded.
174 After the behavioral observation, the opaque divider separating the chamber from the
175 observation aquarium was replaced and the fish was sacrificed according to treatment (15
176 minutes, 30 minutes or 60 minutes after the behavioral observation completed). In order to
177 eliminate any olfactory cues that might affect subsequent behavioral observations, the water in
178 each of the observation tanks was replaced after each behavioral observation.

179 The two pike used in this study did not differ in behavior and movement of the pike
180 during the observation period did not have a statistically detectable effect on either the behavior
181 or the physiology of the sticklebacks (all $P > 0.05$).

182 **Baseline control:** Each day, for ten days, a single stickleback was netted from a stock
183 tank and sacrificed immediately to contribute to a baseline control value for neuroendocrine and

184 hormonal measurements. These fish were collected at the same time as individuals in the
185 treatment groups to minimize the amount of disturbance in the stock tank.

186 **Settling tank control:** At the end of each observation day, 1-2 remaining individuals in
187 the 'settling tank' were quickly netted from the settling tank and sacrificed immediately. This
188 group (n=10) was analyzed for corticosteroids to determine whether transfer and housing in the
189 flume produced a stress response. However, it is important to note that this group does not
190 control for the effect of isolation. We did not detect a difference in whole-body between the
191 settling tank control and the baseline control and therefore did not analyze this treatment group
192 further (Figure 1, $F_{1,18}=0.488$, $P=0.494$).

193

194 Tissue collection

195 Fish were quickly killed by decapitation. The head and body were immediately weighed,
196 the brain dissected out within three minutes and mounted in Tissue-Tek (Sakura). The brain and
197 body were immediately frozen on dry ice and stored at -80°C until physiological analyses. A
198 small amount of tissue from the tail fin was placed in 80% ethanol for DNA extraction for sex
199 determination. Tissue was collected between 0800 and 1800 hours. As in [42], we found no
200 evidence for circadian changes in whole-body cortisol ($r=0.045$, $F_{1,58}=0.118$, $P=0.773$).

201

202 Steroid determination

203 Corticosteroids were assessed by measurement of solvent-extractable immunoreactivity in
204 whole-body homogenates. Corticosteroids were extracted from the tissue by homogenization in
205 ethyl acetate (5:1 volume:carcass weight). Recovery of steroids from homogenized tissue was
206 assessed by adding 50 μl radio-labelled cortisol tracer to homogenized tissue and equilibrating for

207 one hour before extractions. Immunoreactive steroids were quantified in 20-100 μ l aliquots of
208 ethyl acetate extracts of whole-body homogenates using a validated cortisol radioimmunoassay
209 procedure as described previously [43-46]. We used the rabbit polyclonal antibody to cortisol
210 produced by the IgG Corporation and supplied by Campro Scientific (code IgG-F-2).
211 A standard curve of 0-800 pg cortisol per tube was used.

212 We quantified cortisol in whole-body homogenates rather than plasma because successful
213 extraction of the brain for monoamine analyses required that it be dissected out and frozen as
214 soon as possible, which precluded rapid blood sampling from the body. The whole-body
215 homogenate method measures cortisol in multiple body compartments. Therefore in addition to
216 measuring plasma concentrations of cortisol, this method also detects cortisol derivatives in the
217 liver and gall bladder that might have cross-reacted with the antibody [47]. This does not detract
218 from the ability of this method to detect the onset of a stress response, because corticosteroids
219 are synthesized de novo and not stored prior to release. This method has been employed
220 previously to monitor the stress response in fish from which, because of their small size, blood
221 samples could not be obtained, including juvenile trout [48], zebra fish [49] and sticklebacks
222 [46]. Simultaneous measurement of plasma cortisol and whole-body cortisol in fish exposed to
223 acute and chronic stressors has confirmed that the method is appropriate for detecting stress-
224 induced changes in HPI activity [48]. Hereafter we refer to concentrations of corticosteroids we
225 measured on whole body preps as ng/g of 'whole-body cortisol'.

226

227 Analysis of brain monoamines

228 Brains were sectioned in a frozen state on a cryostat and mounted on glass slides.
229 Sections of 300 μ m thickness were cut in the coronal plane. Brain-punch microdissection was

230 performed as described by [30]. The hypothalamus, telencephalon and region posterior to the
231 hypothalamus ('reticular formation') were identified for punching.

232 Punches from each of these three regions were collected and homogenized in 50 μ l ice-
233 cold 4% perchloric acid containing 40 ng/ml DHBA (dihydroxybenzamine) as internal standard,
234 using an MSE 100-W ultrasonic disintegrator. Samples were then centrifuged at 13000rpm for
235 10 minutes at 4°C and the supernatants were analyzed for serotonin (5-HT), dopamine (DA) and
236 norepinephrine (NE) and their metabolites 5-hydroxyindoleacetic acid (5-HIAA), 3,4-
237 dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA) using high performance
238 liquid chromatography with electrochemical detection [50] immediately, or stored at -80°C for
239 no more than two days prior to analysis. Pellets were stored at -80°C for subsequent analysis of
240 protein content in an Eppendorf Biophotometer by a pre-made program measuring absorbance at
241 280nm. The monoamines and monoamine metabolites were quantified using standard solutions
242 and corrected for recovery of the internal standard using HPLC software (CSW, DataApex Ltd.,
243 the Czech republic). The concentration of monoamines and monoamine metabolites is expressed
244 as ng per mg protein.

245 We did not detect strong differences between brain regions in concentrations of brain
246 monoamines: the only effect that we detected was that levels of DA ($F_{2,81}=3.36$, $P=0.04$), 5-
247 HIAA ($F_{2,81}=4.57$, $P=0.013$) and 5-HT ($F_{2,81}=5.21$, $P=0.007$) were significantly lower in the
248 reticular formation in the 'predator' treatment (Table 1). Therefore we summed the concentration
249 of each monoamine across regions and focused our subsequent analysis of treatment differences
250 on the whole-brain values. However, the failure to detect strong region-specific differences
251 should not be overinterpreted because we did not have the resolution to detect fine-scale

252 differences. Other studies have found region-specific differences in monoamine turnover during
253 aggression [32].

254 A decrease in the concentration of a monoamine neurotransmitter could reflect a
255 reduction in the release of the neurotransmitter (decrease in activity) or an increase in turnover to
256 its metabolite (increase in activity). Therefore, it is preferable to use the ratio of the parent
257 neurotransmitter to its metabolite (5-HIAA:5-HT, DOPAC:DA AND HVA:DA) as an index of
258 neurotransmitter activity. However, we were unable to quantify the NE metabolite, 3-methoxy-4-
259 hydroxyphenylglycol (MHPG) in any of the samples as a consequence of non-identified
260 interfering peaks. In addition, in some samples the monoamines (especially 5-HIAA and 5-HT)
261 became degraded during the sampling procedure, resulting in our failure to detect 5-HIAA. This
262 was particularly a problem for the ‘conspecific’ treatments (Table 1). Samples with undetectable
263 levels of a monoamine were omitted from that analysis.

264 Here, we report data on the concentration of both the parent monoamine and metabolite,
265 and we focus our interpretation on differences between treatment groups, rather than on the
266 functional significance of absolute levels.

267

268 Determining genetic sex

269 DNA was extracted from each fin clip and genetic sex was determined by genotyping
270 each individual for a male-specific genetic marker validated for sticklebacks [51].

271

272 Data analysis

273 We compared the behavioral and physiological responses of sticklebacks to an unfamiliar
274 conspecific and a predator across time using general linear models except when data were non-

275 normal. We tested for the effects of sex, body size, time and treatment on each of the dependent
276 variables (behavior, whole-body cortisol and brain monoamines in the different regions). We did
277 not detect sex differences in behavior, whole-body cortisol or brain monoamines and therefore
278 did not analyze this factor further (all $P > 0.4$). The least-squares difference post-hoc test was used
279 to test for differences between groups, except when the distribution was non-normal, in which
280 case we tested for differences between treatments using the nonparametric Mann-Whitney U test.

281 Pearson correlations were used to test for statistically significant relationships between
282 variables when the data were normally distributed; otherwise, Spearman rank correlation
283 statistics were computed. Because the same behavioral data was used to test for associations with
284 brain monoamine concentrations, we used the sequential Bonferroni procedure to correct for
285 multiple tests. Briefly, for each brain region within a treatment group, we replaced the
286 correlation statistics with their corresponding P-values and then ranked them from smallest to
287 largest. Results that were significant ($P < 0.05$) after the sequential Bonferroni procedure are
288 reported [52]. All tests were two-tailed.

289 All of the procedures were carried out according to institutional guidelines and in
290 accordance with the U.K. Animals (Scientific Procedures) Act of 1986.

291

292 **RESULTS**

293 Behavioural and physiological responses to an unfamiliar conspecific

294 Presentation of an unfamiliar conspecific elicited a behavioral response; on average,
295 individuals approached the intruder 8 times and attacked 11 times within the observation period.
296 However, individuals differed in their behavioral reaction to the simulated intrusion; while one
297 individual attacked the conspecific over 40 times, other individuals spent most of their time

298 hiding, and scarcely left the refuge. Body size explained some of this individual variation; bigger
299 fish were more aggressive toward their size-matched opponents (number of attacks: $r=0.433$,
300 $P=0.024$, $n=27$). All of the fish oriented to and approached the conspecific and one-half of the
301 fish attacked it at least once.

302 Interaction with the unfamiliar conspecific quickly produced a glucocorticoid response
303 (Figure 1). Whole-body cortisol levels were highest 15 minutes after the simulated intrusion and
304 then returned to baseline levels by 30 minutes.

305 The serotonergic system was quickly suppressed in response to the presence of the
306 unfamiliar conspecific, as indicated by reduced whole-brain levels of 5-HT (Figure 2A, Table 1).

307 Dopamine turnover to DOPAC was elevated 60 minutes following the aggressive
308 interaction (Figure 2C and 2D), while levels of norepinephrine were consistently low (Figure
309 2F).

310 Individual differences in concentrations of brain monoamines were related to differences
311 among individuals in aggressiveness. Individuals with lower hypothalamic 5HT were more
312 aggressive ($r=-0.806$, $P=0.016$, $n=8$, Figure 3A), while norepinephrine ($r=0.883$, $P=0.020$, $n=6$,
313 Figure 3B) and DOPAC ($r=0.815$, $P=0.048$, $n=6$, Figure 3C) were positively associated with
314 aggressiveness.

315

316 Behavioural and physiological responses to a predator

317 When presented with the pike, most individuals inspected the predator at least once and
318 oriented to it more than nine times. As in the ‘conspecific’ treatment, individuals differed in their
319 behavior: some individuals inspected the pike as many as seven times during the 15-minute
320 observation period, while others spent the entire observation period hiding in the refuge.

321 Exposure to the predator elicited a significant glucocorticoid response within 15 minutes
322 which reached a maximum 60 minutes after exposure to the predator (Figure 1). Concentrations
323 of DOPAC fell at 60 minutes (Figure 2D) while concentrations of HVA increased at 15 minutes
324 (Figure 2E), indicating that predator-induced stress stimulated the rapid turnover of DA to HVA.

325 Activity under predation risk and predator inspection behavior (both of which potentially
326 involve a risk of predation) were positively associated with neurotransmitter concentrations. For
327 example, individuals with greater levels of NE engaged in riskier behavior ($r=0.766$, $P=0.027$,
328 $n=8$, Figure 4A). Serotonin turnover was also associated with predator inspection behavior: the
329 number of predator inspections was significantly positively correlated with hypothalamic
330 serotonin ($r=0.928$, $P=0.003$, $n=7$, Figure 4B) and negatively correlated with whole-brain
331 serotonergic activity ($r=-0.669$, $P=0.049$, $n=9$, Figure 4C).

332

333 Comparing responses to the conspecific and predator

334 Both confrontation by a conspecific and exposure to a predator elicited a cortisol
335 response, but the time course of the cortisol response differed between treatments (Figure 1), as
336 evidenced by the significant interaction between time and treatment ($F_{2,58}=5.5$, $P=0.006$).
337 Moreover, the magnitude (average across the three time periods) of the cortisol response was
338 greater to the predator compared to a conspecific (Conspecific: 47 ± 4.97 ng/g, Predator: 72 ± 8.24
339 ng/g, $P=0.002$).

340 Relative to the conspecific treatment, NE (Figure 2F) and to a lesser extent, DA (Figure
341 2C) were higher in the predator treatments.

342

343 **DISCUSSION**

344

345 In this experiment, we tested the hypothesis that both the HPI axis and brain
346 monoaminergic systems are activated in response to fighting with an unfamiliar conspecific and
347 exposure to a predator. While other studies have found links between these systems in laboratory
348 animals, the results from this study extends these findings to wild-caught animals that were
349 confronted by ecologically relevant challenges [28,53]. We found that both stressors elicited a
350 similar HPI response, but produced very different patterns of change in monoamine content.

351 Our design permitted us to determine the time course of the neuroendocrine response to
352 these stressors and to ascertain whether individual differences in behavioral responses to the
353 stressors were related to underlying physiology. We showed that not only do these challenges
354 elicit a neuroendocrine response, but that different behavioral responses of individuals were
355 related to their particular neuroendocrine profiles.

356

357 The cortisol response to a conspecific and predator were broadly similar, but exposure to a
358 predator was more stressful

359

360 During the present study, both confrontation with an unfamiliar conspecific and exposure
361 to a predator resulted in activation of the HPI axis and significant alterations in the levels of
362 brain monoamines in sticklebacks. These results are consistent with other studies which have
363 shown that both confrontation by a challenging conspecific [10,23] and exposure to a predator
364 [54] elicit a neuroendocrine stress response in fishes.

365 In the present study both exposure to a conspecific or to a predator resulted in highly
366 significant increases in whole-body cortisol concentrations within 15 minutes relative to controls.
367 In the conspecific-exposed group, whole-body cortisol levels were statistically indistinguishable
368 from control fish after 30 minutes and remained so at 60 minutes. In contrast, whole-body
369 cortisol concentrations in the predator-exposed group remained highly elevated after 60 minutes,
370 significantly exceeding levels attained after 15 minutes. We interpret these data to indicate that
371 the magnitude of the initial response to both stressors was similar, resulting in similar whole-
372 body cortisol concentrations at 15 minutes, but that the HPI axis in the predator-exposed fish
373 remained active for longer, resulting in a greater accumulation of whole-body cortisol with time.
374 The overall significant difference in total cortisol between the two treatment groups detected
375 across all time points indicates a quantitative difference in the response of the fish to the two
376 stressors.

377 Other studies have found evidence for a more rapid recovery to baseline cortisol levels
378 following less threatening situations compared to more threatening situations [55]. A longer-
379 lasting cortisol response to threat of predation as compared to other stressors has been
380 documented in stonechats [56] and rodents [57,58]. Therefore in this experiment, we hypothesize
381 that the different time course of the cortisol response to a competitor versus to a predator is
382 related to the perceived magnitude of the two different challenges. Sticklebacks are social fish,
383 and frequently interact with other sticklebacks in shoals. Because encounters with conspecifics
384 are frequent, natural selection might have favored individuals which do not mount a severe stress
385 response to frequent interactions with conspecifics, and should favor individuals which recover
386 quickly from fights. In contrast, encounters with predators are less frequent and more threatening

387 than encounters with conspecifics, so selection might have favored individuals with a greater and
388 longer-lasting stress response.

389 The levels of whole-body cortisol detected in unstressed sticklebacks during the present
390 study were similar to those previously reported for this species (2 – 8 ng g⁻¹; [46]) and levels
391 detected in the stressed fish in the present study, although slightly higher, were also broadly
392 consistent with previous observations (50 ng g⁻¹;[46]). The difference in magnitude of whole-
393 body cortisol levels between this and previous studies may be related to the nature of the
394 stressor.

395 Links between stress-induced blood cortisol levels and behavioral traits have been shown
396 in fish [10,23], mammals [59] and reptiles [9]. However, while exposure to both stressors elicited
397 a behavioral and whole-body cortisol response in the treatment groups, we did not detect a
398 relationship at the individual level between concentrations of whole-body cortisol and behavior.
399 It is possible that our method might not have had the resolution to detect fine-scale individual
400 differences.

401 We did not detect any sex differences in whole-body cortisol. The stress response in
402 vertebrates, including fish [60], is modulated by gonadal steroids with androgens suppressing
403 and estrogens enhancing corticosteroid responsiveness [61]. However, the fish employed in this
404 study were not reproductively active and it is therefore unsurprising that no sex-dependent
405 differences in stress response were observed.

406

407 The monoamine responses to a conspecific and a predator were qualitatively different

408 Whereas the cortisol response was broadly similar across stressors, the monoamines
409 showed a differential response across the two stressors, some being suppressed in response to a
410 conspecific but elevated in response to the predator.

411 For example, relative to the control group, concentrations of NE were consistently *higher*
412 in the ‘predator’ treatments, and *lower* in the ‘conspecific’ treatments. Without data on the NE
413 metabolite, MHPG, we cannot distinguish if reduced concentrations reflect a reduction in NE
414 release (decrease in NE activity) or an increased turnover to MHPG (increase in NE activity).
415 However, at an individual level we found that NE was consistently associated with risk-taking
416 behaviors in both kinds of situations: NE was positively correlated with aggressive behaviors as
417 well as predator inspection behaviors. These positive correlations suggest that more bold or
418 aggressive individuals were more ‘aroused’, active or uninhibited, results which are consistent
419 with other studies showing positive relationships between NE activity and behavioral impulsivity
420 in monkeys [28] and sensation seeking in humans [62]. The fact that serotonin and NE had
421 opposite relationships with risk-taking behaviors in this experiment is consistent with the
422 observation that 5-HT and catecholamines can have antagonistic effects on behavior [17].

423

424 Associations between serotonin, risk-taking behaviors and aggression

425 In agreement with other studies which have shown that risk-taking behaviors are
426 negatively associated with brain serotonergic activity [24,27-29], we found that risk-taking
427 behaviors performed while under predation risk (e.g. inspection) were negatively correlated with
428 serotonin turnover to 5-HIAA (Figure 4C).

429 Our results support the view that 5-HT has an inhibitory effect on aggressive behavior
430 [16,54]. We found a negative relationship at the individual level between concentrations of 5-HT
431 and aggressive behavior, and that confrontation by an unfamiliar conspecific resulted in lower 5-
432 HT. Other studies have shown that winners of agonistic interactions have up-regulated brain 5-
433 HT activity [21,30-32]. One possible explanation for this different pattern is that in our
434 experiment, there was no physical contact between the resident and the intruder because the
435 intruders were confined to a flask. As a result, the resident fish were unable to complete their
436 attacks and therefore might not be analogous to the winners in the forementioned studies. We
437 remain provisional in our interpretation of these results because 5-HIAA was degraded in many
438 of the samples in the ‘conspecific’ treatments, preventing us from calculating serotonin turnover
439 in those treatments. However, it is worth noting that while more aggressive behaviors were
440 negatively associated with serotonin (Figure 3A), risk-taking behavior under predation risk
441 showed the opposite pattern – it was *positively* correlated with 5HT (Figure 4B), and negatively
442 associated with serotonin turnover to 5-HIAA (Figure 4C).

443 Overall, these data provide evidence that the response of fish to stressors is not identical
444 regardless of the nature of the challenge, but rather that the response varies according to the
445 magnitude, frequency and predictability of the stressor, as is the case for other vertebrates
446 [56,63]. Further studies on individual variation in responses to different stressors would benefit
447 from repeated sampling of the same physiological measures on the same individuals. While it is
448 currently a challenge to measure brain monoamines noninvasively, noninvasive methods for
449 measuring glucocorticoids in fish [64] are a promising alternative. In addition, the roles played
450 by upstream elements of the stress response such as corticotropin releasing hormone (CRH) and
451 variation in the binding characteristics of corticosteroid receptors and corticotropin binding

452 proteins should also be investigated [65]. Given that other studies have shown that inter-
453 individual differences in stress responsiveness have a high heritable component [66], further
454 investigation will provide insight into the mechanisms that have produced adaptive, heritable
455 behavioral variation in sticklebacks in diverse ecological settings.

456

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466

467

468 **REFERENCES**

469

- 470 1. Thorpe, K.E.; A.C. Taylor; F.A. Huntingford. How costly is fighting? Physiological
471 effects of sustained exercise and fighting in swimming crabs, *Necora puber*. *Animal*
472 *Behaviour*. 1995, 50: 1657-1666.
- 473 2. Neat, F.C.; A.C. Taylor; F.A. Huntingford. Proximate costs of fighting in male cichlid
474 fish: the role of injuries and energy metabolism. *Animal Behaviour*. 1998, 55: 875-882.

- 475 3. Diaz-Uriarte, R. Anti-predator behaviour changes following an aggressive encounter in
476 the lizard *Tropidurus hispidus*. Proceedings of the Royal Society of London Series B-
477 Biological Sciences. 1999, 266(1437): 2457-2464.
- 478 4. Ydenberg, R.C.; L.M. Dill. The economics of fleeing from predators. Advances in the
479 Study of Behavior. 1986, 16: 229-249.
- 480 5. Lin, Z.H.; Y.F. Qu; X. Ji. Energetic and locomotor costs of tail loss in the Chinese skink,
481 *Eumeces chinensis*. Comparative Biochemistry and Physiology a-Molecular &
482 Integrative Physiology. 2006, 143(4): 508-513.
- 483 6. Elofsson, U.O.E.; L. Mayer; B. Damsgard; S. Winberg. Intermale competition in sexually
484 mature arctic charr: Effects on brain monoamines, endocrine stress responses, sex
485 hormone levels, and behavior. General and Comparative Endocrinology. 2000, 118(3):
486 450-460.
- 487 7. Fox, H.E.; S.A. White; M.H.F. Kao; R.D. Fernald. Stress and dominance in a social fish.
488 Journal of Neuroscience. 1997, 17(16): 6463-6469.
- 489 8. Winberg, S.; O. Lepage. Elevation of brain 5-HT activity, POMC expression, and plasma
490 cortisol in socially subordinate rainbow trout. American Journal of Physiology-
491 Regulatory Integrative and Comparative Physiology. 1998, 43(3): R645-R654.
- 492 9. Summers, C.H.; M.J. Watt; T.L. Ling; G.L. Forster; R.E. Carpenter; W.J. Korzan; J.L.
493 Lukkes; O. Overli. Glucocorticoid interaction with aggression in non-mammalian
494 vertebrates: Reciprocal action. European Journal of Pharmacology. 2005, 526(1-3): 21-
495 35.

- 496 10. Sloman, K.A.; N.B. Metcalfe; A.C. Taylor; K.M. Gilmour. Plasma cortisol
497 concentrations before and after social stress in rainbow trout and brown trout.
498 *Physiological and Biochemical Zoology*. 2001, 74: 383-389.
- 499 11. Knapp, R.; M.C. Moore. Hormonal responses to aggression vary in different types of
500 agonistic encounters in male tree lizards. *Hormones and Behavior*. 1995, 29: 85-105.
- 501 12. Cockrem, J.F.; B. Silverin. Sight of a predator can stimulate a corticosterone response in
502 the great tit. *General and Comparative Endocrinology*. 2002, 125: 248-255.
- 503 13. Scheuerlein, A.; T.J. Van't Hof; E. Gwinner. Predators as stressors? Physiological and
504 reproductive consequences of predation risk in tropical stonechats. *Proceedings of the*
505 *Royal Society Biological Sciences Series B*. 2001, 268: 1575-1582.
- 506 14. Eilam, D.; T. Dayan; S. Ben-Eliyahu; I. Schulman; G. Shefer; C.A. Hendries. Differential
507 behavioural and hormonal responses of voles and spiny mice to owl calls. *Animal*
508 *Behaviour*. 1999, 58: 1085-1093.
- 509 15. Winberg, S.; G.E. Nilsson. Roles of brain monoamine neurotransmitters in agonistic
510 behaviour and stress reactions, with particular reference to fish. *Comparative*
511 *Biochemistry and Physiology C Comparative Pharmacology and Toxicology*. 1993,
512 106(3): 597-614.
- 513 16. Winberg, S.; A. Nilsson; P. Hylland; V. Soderstrom; G.E. Nilsson. Serotonin as a
514 regulator of hypothalamic-pituitary-interrenal activity in teleost fish. *Neuroscience*
515 *Letters*. 1997, 230(2): 113-116.
- 516 17. Johnsson, J.I.; S. Winberg; K.A. Sloman. Social interactions. In: K.A. Sloman, R.W.
517 Wilson, and S. Balshine, *Behavior: Interactions with fish physiology. A volume of Fish*
518 *Physiology*, Editor^Editors. 2006, Elsevier.

- 519 18. Campbell, T.; S. Lin; C. DeVries; K. Lambert. Coping strategies in male and female rats
520 exposed to multiple stressors. *Physiology and Behavior*. 2003, 78: 495-504.
- 521 19. Ebner, K.; C.T. Wotjak; R. Landgraf; M. Engelmann. Neuroendocrine and behavioral
522 response to social confrontation: residents versus intruders, active versus passive coping
523 styles. *Hormones and Behavior*. 2005, 47: 14-21.
- 524 20. Clement, T.S.; V. Parikh; M. Schrumph; R.D. Fernald. Behavioral coping strategies in a
525 cichlid fish: the role of social status and acute stress response in direct and displaced
526 aggression. *Hormones and Behavior*. 2005, 47: 336-342.
- 527 21. Øverli, O.; C. Sorensen; G.E. Nilsson. Behavioral indicators of stress-coping style in
528 rainbow trout: do males and females react differently to novelty? *Physiology and*
529 *Behavior*. 2006, 87: 506-512.
- 530 22. Schjolden, J.; A. Stoskhus; S. Winberg. Does individual variation in stress responses and
531 agonistic behavior reflect divergent stress coping strategies in juvenile rainbow trout?
532 *Physiological and Biochemical Zoology*. 2005, 78: 715-723.
- 533 23. Øverli, O.; S. Winberg; T.G. Pottinger. Behavioral and neuroendocrine correlates of
534 selection for stress responsiveness in rainbow trout - a review. *Integrative & Comparative*
535 *Biology*. 2005, 45(3): 463-474.
- 536 24. van Hierden, Y.M.; S.M. Korte; E.W. Ruesink; C.G. van Reenen; B. Engel; G.A.H.
537 Korte-Bouws; J.M. Koolhaas; H.J. Blokhuis. Adrenocortical reactivity and central
538 serotonin and dopamine turnover in young chicks from a high and low feather-pecking
539 line of laying hens. *Physiology & Behavior*. 2002, 75(5): 653-659.

- 540 25. Ballenger, J.C.; R.M. Post; D.C. Jimerson; C.R. Lake; D. Murphy; M. Zuckerman; C.
541 Cronin. Biochemical correlates of personality traits in normals - an exploratory study.
542 Personality and Individual Differences. 1983, 4(6): 615-625.
- 543 26. Depue, R.A. Neurobiological factors in personality and depression. European Journal of
544 Personality. 1995, 9(5): 413-439.
- 545 27. Ferrari, P.F.; P. Palanza; S. Parmigiani; R.M.M. de Almeida; K.A. Miczek. Serotonin and
546 aggressive behavior in rodents and nonhuman primates: Predispositions and plasticity.
547 European Journal of Pharmacology. 2005, 526(1-3): 259-273.
- 548 28. Fairbanks, L.A.; M.B. Fontenot; J.E. Phillips-Conroy; C.J. Jolly; J.R. Kaplan; J.J. Mann.
549 CSF monoamines, age and impulsivity in wild grivet monkeys (*Cercopithecus aethiops*
550 *aethiops*). Brain Behavior and Evolution. 1999, 53(5-6): 305-312.
- 551 29. Mehlman, P.T.; J.D. Higley; I. Faucher; A.A. Lilly; D.M. Taub; J. Vickers; S.J. Suomi;
552 M. Linnoila. Low Csf 5-Hiaa concentrations and severe aggression and impaired impulse
553 control in nonhuman primates. American Journal of Psychiatry. 1994, 151(10): 1485-
554 1491.
- 555 30. Korzan, W.J.; T.R. Summers; C.H. Summers. Monoaminergic activities of limbic regions
556 are elevated during aggression: Influence of sympathetic social signaling. Brain
557 Research. 2000, 870(1-2): 170-178.
- 558 31. Summers, C.H.; T.R. Summers; M.C. Moore; W.J. Korzan; S.K. Woodley; P.J. Ronan; E.
559 Hoglund; M.J. Watt; N. Greenberg. Temporal patterns of limbic monoamine and plasma
560 corticosterone response during social stress. Neuroscience. 2003, 116: 553-563.
- 561 32. Summers, C.H.; W.J. Korzan; J.L. Lukkes; M.J. Watt; G.L. Forster; O. Øverli; E.
562 Höglund; E.T. Larson; P.J. Ronan; J.M. Matter; T.R. Summers; K.J. Renner; N.

- 563 Greenberg. Does serotonin influence aggression? Comparing regional activity before and
564 during social interaction. *Physiol Biochem Zool.* 2005, 78: 679-694.
- 565 33. Lepage, O.; O. Tottmar; S. Winberg. Elevated dietary intake of l-tryptophan counteracts
566 the stress-induced elevation of plasma cortisol in rainbow trout. *Journal of Experimental*
567 *Biology.* 2002, 205: 3679-3687.
- 568 34. Bell, A.M.; J.A. Stamps. The development of behavioural differences between
569 individuals and populations of stickleback. *Animal Behaviour.* 2004, 68: 1339-1348.
- 570 35. Bell, A.M. Differences between individuals and populations of threespined stickleback.
571 *Journal of Evolutionary Biology.* 2005, 18: 464-473.
- 572 36. Huntingford, F.A. The relationship between inter- and intra-specific aggression. *Animal*
573 *Behaviour.* 1976, 24: 485-497.
- 574 37. Sih, A.; A.M. Bell; J.C. Johnson. Behavioral syndromes: an ecological and evolutionary
575 overview. *Trends in Ecology & Evolution.* 2004, 19(7): 372-378.
- 576 38. Sih, A.; A.M. Bell; J.C. Johnson; R. Ziemba. Behavioral syndromes: an integrative
577 overview. *Quarterly Review of Biology.* 2004, 79: 241-277.
- 578 39. Koolhaas, J.M.; S.M. Korte; S.F. De Boer; B.J. Van Der Vegt; C.G. Van Reenen; H.
579 Hopster; I.C. De Jong; M.A.W. Ruis; H.J. Blokhuis. Coping styles in animals: Current
580 status in behavior and stress-physiology. *Neuroscience & Biobehavioral Reviews.* 1999,
581 23(7): 925-935.
- 582 40. Tamashiro, K.L.K.; M.M.N. Nguyen; R.R. Sakai. Social stress: From rodents to primates.
583 *FRONTIERS IN NEUROENDOCRINOLOGY.* 2005, 26: 27-40.
- 584 41. Bakker, T.C.M. Aggressiveness in sticklebacks (*Gasterosteus aculeatus*) a behavior-
585 genetic study. *Behaviour.* 1986, 98(1-4): 1-144.

- 586 42. Audet, C.; G.J. Fitzgerald; H. Guderley. Photoperiod effects on plasma cortisol levels in
587 *Gasterosteus aculeatus*. *General and Comparative Endocrinology*. 1986, 61(1): 76-81.
- 588 43. Pottinger, T.G.; T.A. Moran; J.A.W. Morgan. Primary and secondary indices of stress in
589 the progeny of rainbow trout (*Oncorhynchus mykiss*) selected for high and low
590 responsiveness to stress. *Journal of Fish Biology*. 1994, 44(1): 149-163.
- 591 44. Øverli, O.; T.G. Pottinger; T.R. Carrick; E. Øverli; S. Winberg. Brain monoaminergic
592 activity in rainbow trout selected for high and low stress responsiveness. *Brain Behavior*
593 *and Evolution*. 2001, 57(4): 214-224.
- 594 45. Øverli, O.; T.G. Pottinger; T.R. Carrick; E. Øverli; S. Winberg. Differences in behaviour
595 between rainbow trout selected for high- and low-stress responsiveness. *Journal of*
596 *Experimental Biology*. 2002, 205(3): 391-395.
- 597 46. Pottinger, T.G.; T.R. Carrick; W.E. Yeomans. The three-spined stickleback as an
598 environmental sentinel: effects of stressors on whole-body physiological indices. *Journal*
599 *of Fish Biology*. 2002, 61(1): 207-229.
- 600 47. Pottinger, T.G.; T.A. Moran; P.A. Cranwell. The biliary accumulation of corticosteroids
601 in rainbow trout, *Oncorhynchus mykiss*: a stable indicator of chronic stress. *Fish*
602 *Physiology and Biochemistry*. 1992, 10: 55-66.
- 603 48. Pottinger, T.G.; E.M. Musowe. The corticosteroidogenic response of brown and rainbow
604 trout alevins and fry during a 'critical' period. *General and Comparative Endocrinology*.
605 1994, 95: 350-362.
- 606 49. Ramsay, J.M.; G.W. Feist; Z.M. Varga; M. Westerfield; M.L. Kent; C.B. Schreck.
607 Whole-body cortisol is an indicator of crowding stress in adult zebrafish, *Danio rerio*.
608 *Aquaculture*. 2006, 258(1-4): 565-574.

- 609 50. Øverli, O.; C.A. Harris; S. Winberg. Short-term effects of fights for social dominance and
610 the establishment of dominant-subordinate relationships on brain monoamines and
611 cortisol in rainbow trout. *Brain Behavior and Evolution*. 1999, 54: 263-275.
- 612 51. Peichel, C.; J. Ross; C. Matson; M. Dickson; J. Grimwood; J. Schmutz; R. Myers; S.
613 Mori; D. Schluter; D. Kingsley. The master sex-determination locus in threespine
614 sticklebacks in on a nascent Y chromosome. *Current Biology*. 2004, 14: 1416-1424.
- 615 52. Rice, W.R. Analyzing tables of statistical tests. *Evolution*. 1989, 43: 223-225.
- 616 53. Kaplan, J.R.; J. Phillips-Conroy; M.B. Fontenot; C.J. Jolly; L.A. Fairbanks; J.J. Mann.
617 Cerebrospinal fluid monoaminergic metabolites differ in wild anubis and hybrid (*Anubis*
618 *hamadryas*) baboons: Possible relationships to life history and behavior.
619 *Neuropsychopharmacology*. 1999, 20(6): 517-524.
- 620 54. Winberg, S.; A.A. Myrberg; G.E. Nilsson. Predator exposure alters brain serotonin
621 metabolism in bicolour damselfish. *Neuroreport*. 1993, 4(4): 399-402.
- 622 55. Djordjevic, J.; G. Cvijic; V. Davidovic. Different activation of ACTH and corticosterone
623 release in response to various stressors in rats. *Physiological Research*. 2003, 52: 67-72.
- 624 56. Canoine, V.; T.J. Hayden; K. Rowe; W. Goymann. The stress response of european
625 stonechats depends on the type of stressor. *Behaviour*. 2002, 139: 1303-1311.
- 626 57. Blanchard, R.J.; J.N. Nikulina; R.S. Sakai; C. McKittrick; B. McEwen; D.C. Blanchard.
627 Behavioral and endocrine change following chronic predatory stress. *Physiology &*
628 *Behavior*. 1998, 63: 561-569.
- 629 58. Adamec, R.E.; T. Shallow. Lasting effects on rodent anxiety of a single exposure to a cat.
630 *Physiology and Behavior*. 1993, 54: 101-109.

- 631 59. Steimer, T.; P. Driscoll. Divergent stress responses and coping styles in
632 psychogenetically selected roman high-(RHA) and low-(RLA) avoidance rats:
633 Behavioural, neuroendocrine and developmental aspects. *Stress*. 2003, 6: 87-100.
- 634 60. Pottinger, T.G.; P.H.M. Balm; A.D. Pickering. Sexual maturity modifies the
635 responsiveness of the pituitary-interrenal axis to stress in male rainbow trout. *General and*
636 *Comparative Endocrinology*. 1995, 98: 311-320.
- 637 61. Pottinger, T.G.; T.R. Carrick; S.E. Hughes; P.H.M. Balm. Testosterone, 11-
638 ketotestosterone and estradiol-17_β modify baseline and stress-induced interrenal and
639 corticotropic activity in trout. *General and Comparative Endocrinology*. 1996, 104: 284-
640 295.
- 641 62. Gerra, G.; P. Avanzini; A. Zaimovic; R. Sartori; C. Bocchi; M. Timpano; U. Zambelli; R.
642 Delsignore; F. Gardini; E. Talarico; F. Brambilla. Neurotransmitters, neuroendocrine
643 correlates of sensation-seeking temperament in normal humans. *Neuropsychobiology*.
644 1999, 39(4): 207-213.
- 645 63. Silverin, B. Behavioural and hormonal responses of the pied flycatcher to environmental
646 stressors. *Animal Behaviour*. 1998, 55: 1411-1420.
- 647 64. Scott, A.P.; M. Pinillos; T. Ellis. Why measure steroids in fish plasma when you can
648 measure them in water? in *Perspectives in Comparative Endocrinology: Unity and*
649 *Diversity. 14th International Congress of Comparative Endocrinology*. 2001. Sorrento,
650 Italy: Monduzzi Editore S.p.A. -Medimond Inc.: Italy.
- 651 65. Sapolsky, R.M.; L.M. Romero; A.U. Munck. How do glucocorticoids influence stress
652 responses? Integrating permissive, suppressive, stimulatory and preparative actions.
653 *Endocrine Reviews*. 2000, 21(1): 55-89.

654 66. Pottinger, T.G.; T.R. Carrick. Modification of the plasma cortisol response to stress in
655 rainbow trout by selective breeding. *General and Comparative Endocrinology*. 1999,
656 116(1): 122-132.

657

658

659 **Figure legends**

660

661 Figure 1. Whole-body cortisol in the different treatments. Statistically similar means share the
662 same letter.

663

664 Figure 2. Whole-brain concentrations of brain monoamines in different treatments. Statistically
665 similar means share the same letter. (A) 5-HT; (B) 5-HIAA; (C) DA; (D) DOPAC; (E) HVA; (F)
666 NE.

667

668 Figure 3. Correlations between monoamine concentrations and aggressive behavior (attacks). (A)
669 Hypothalamic 5-HT 60 minutes after a fight; (B) NE in reticular formation 15 minutes after a
670 fight; (C) Telencephalic DOPAC 30 minutes after a fight.

671

672 Figure 4. Correlations between monoamine concentrations and behavior under predation risk.
673 (A) Telencephalic NE 60 minutes after exposure and time orienting to the predator; (B)
674 Hypothalamic 5-HT 60 minutes after exposure and predator inspections; (C) Whole-brain 5-
675 HIAA:5-HT ratio 15 minutes after exposure and predator inspections.

Figure 1

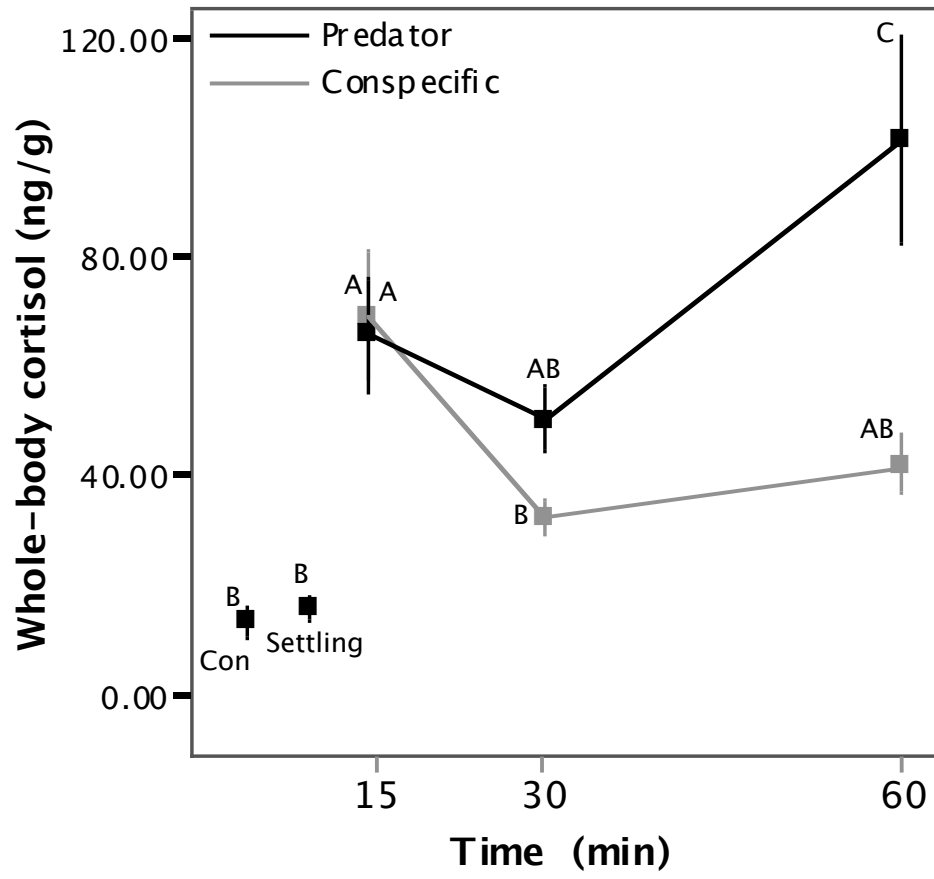
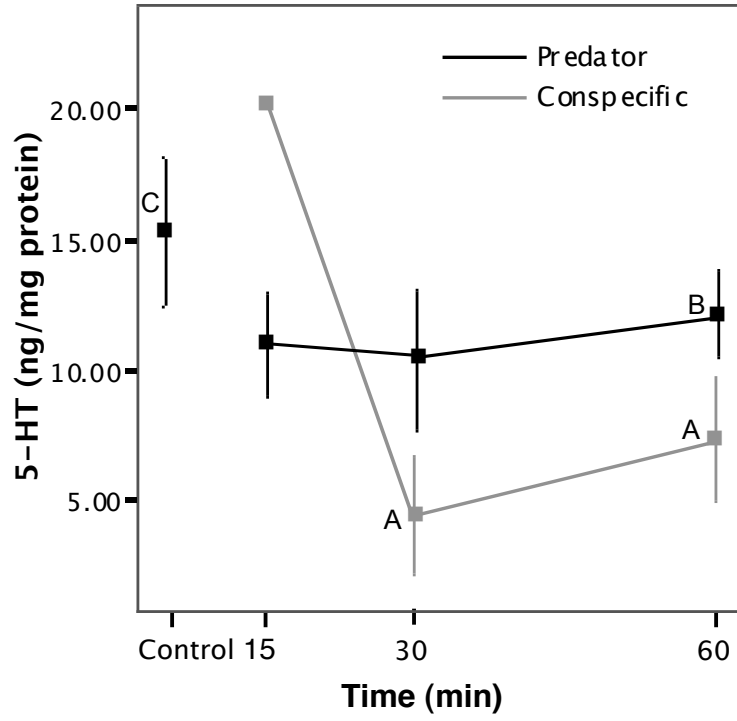
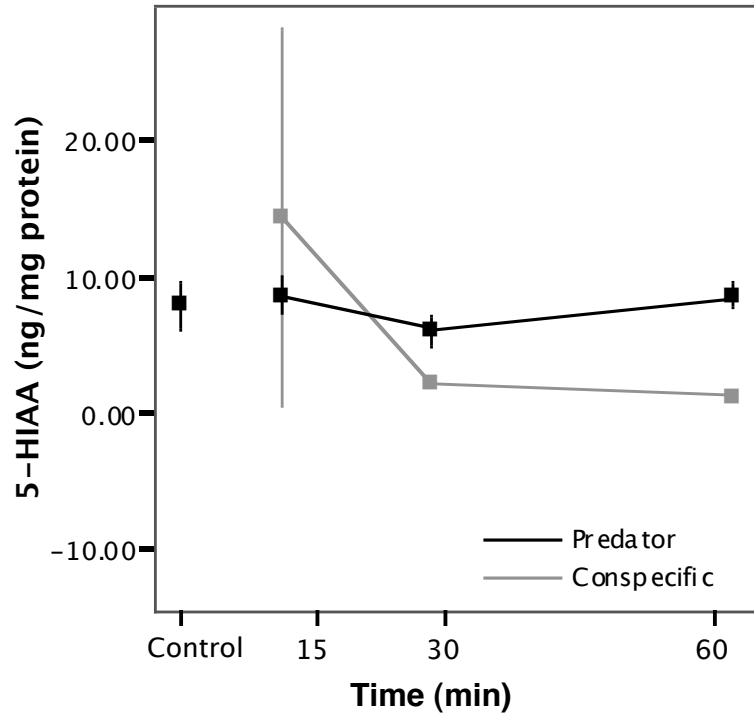


Figure 2

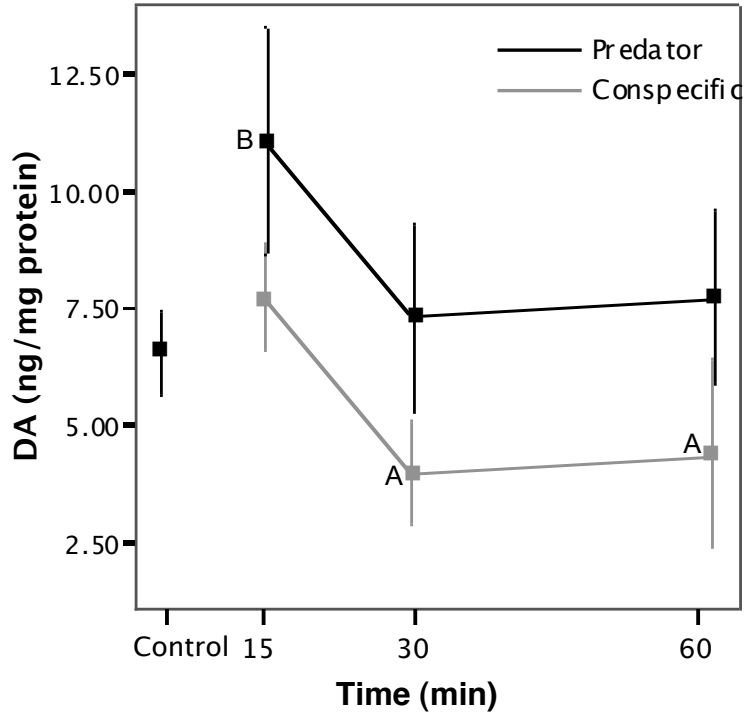
(A)



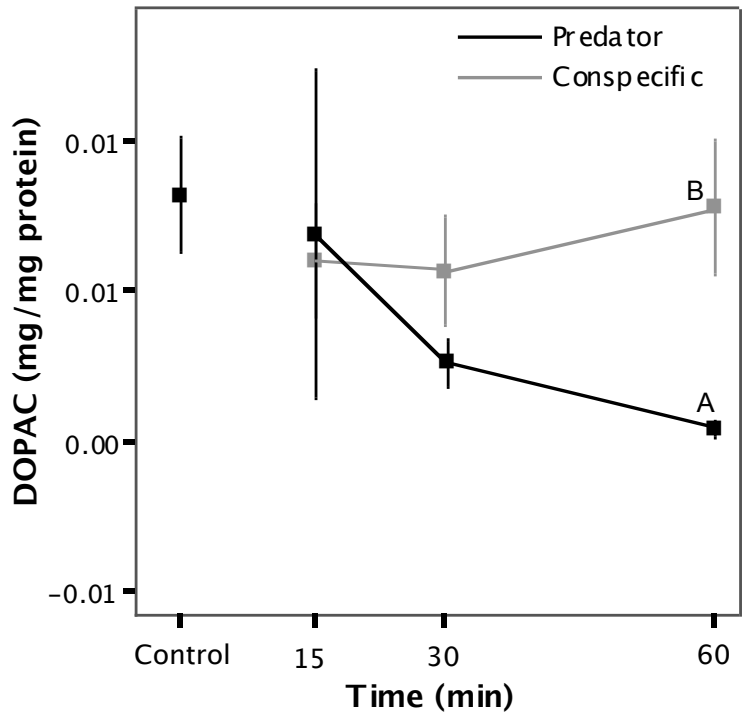
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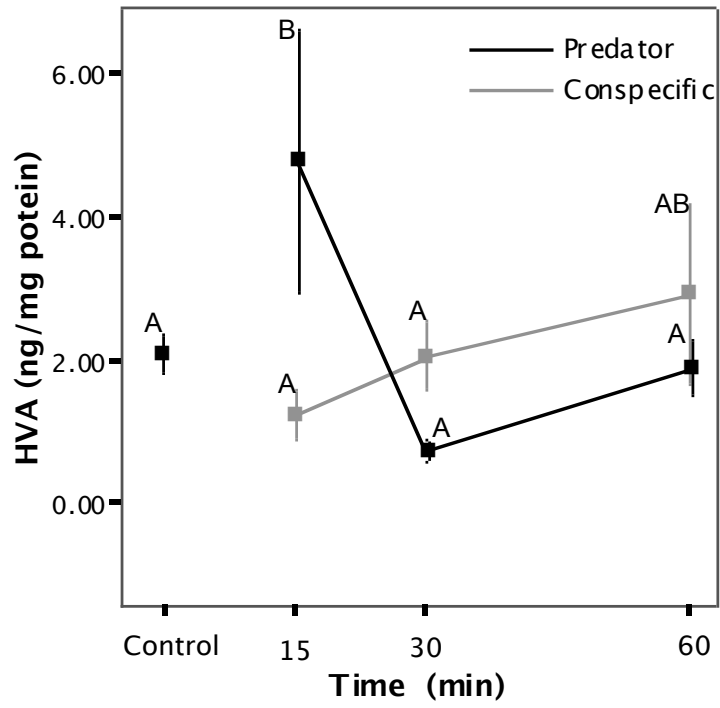
(C)



(D)



(E)



(F)

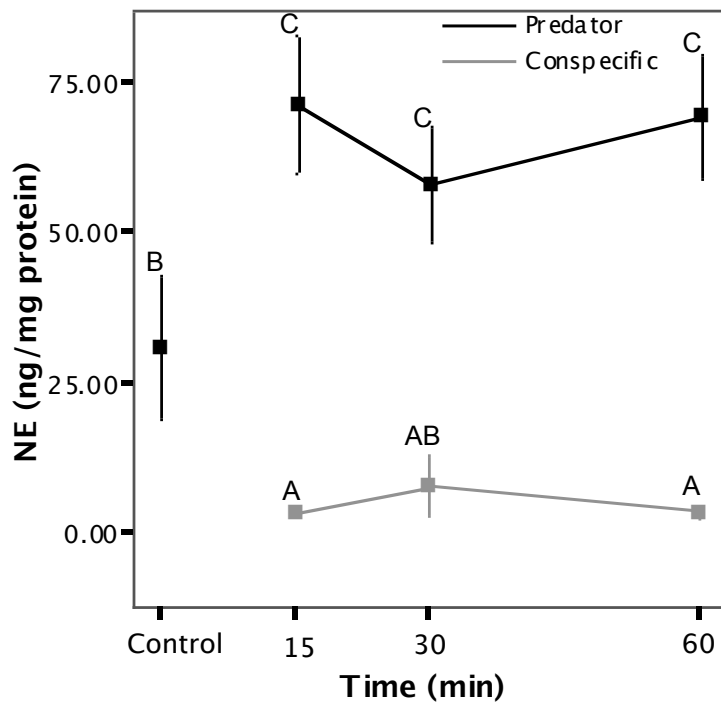
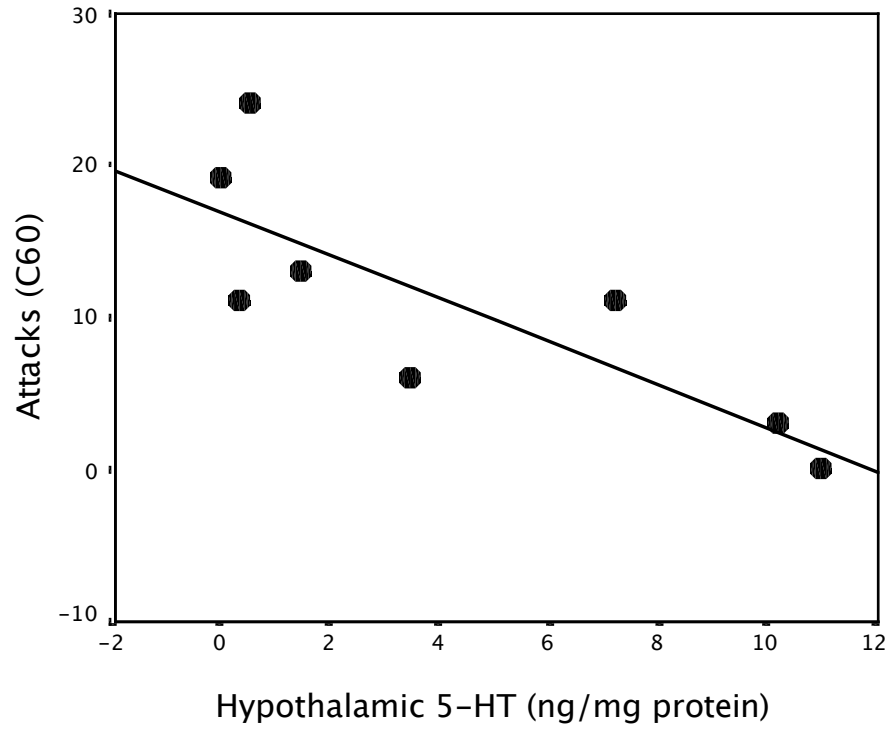
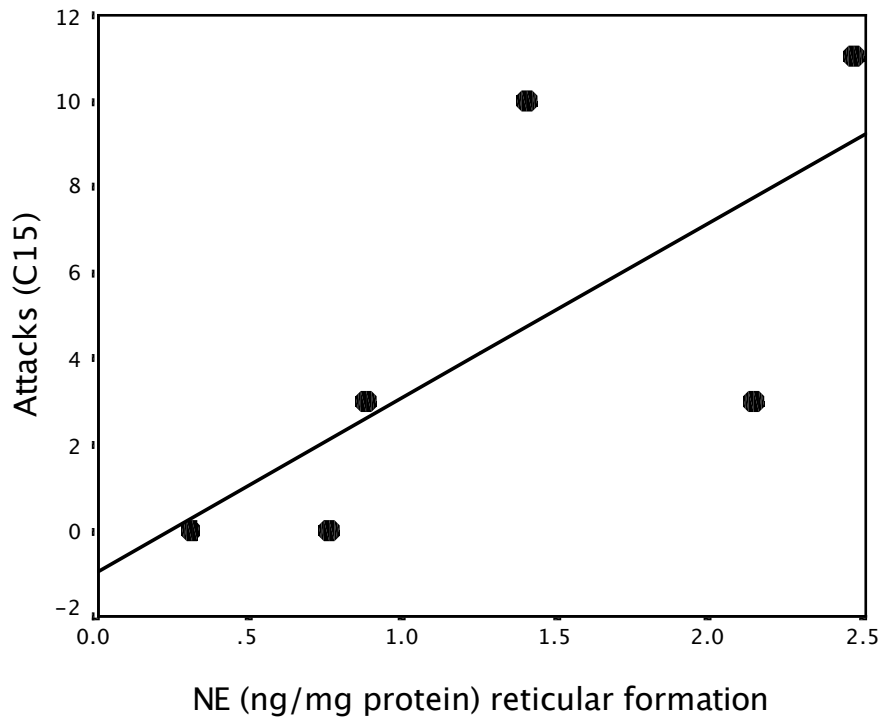


Figure 3

(A)



(B)



(C)

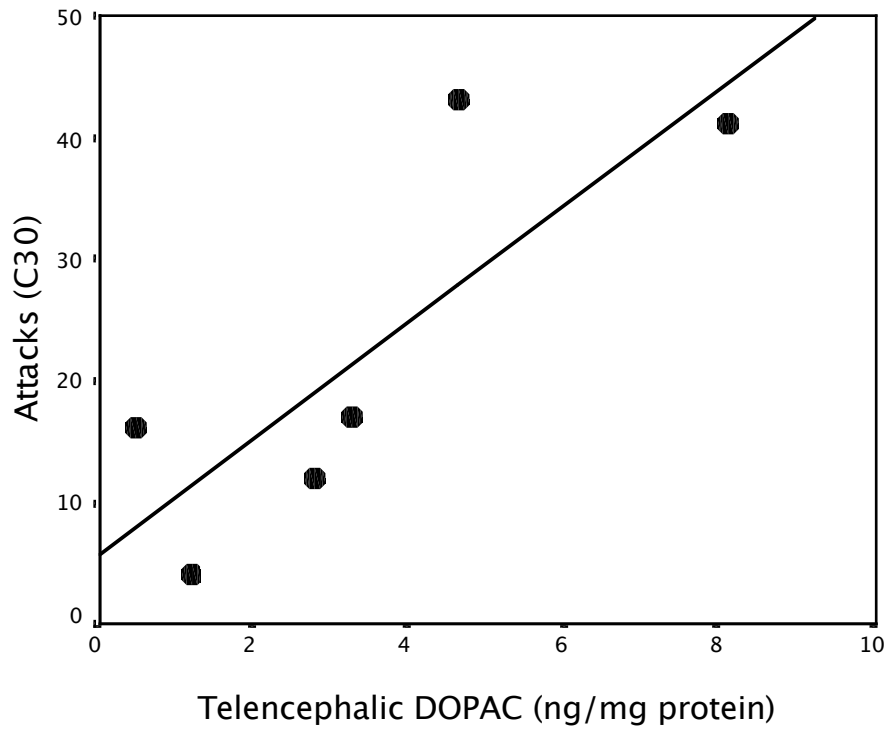
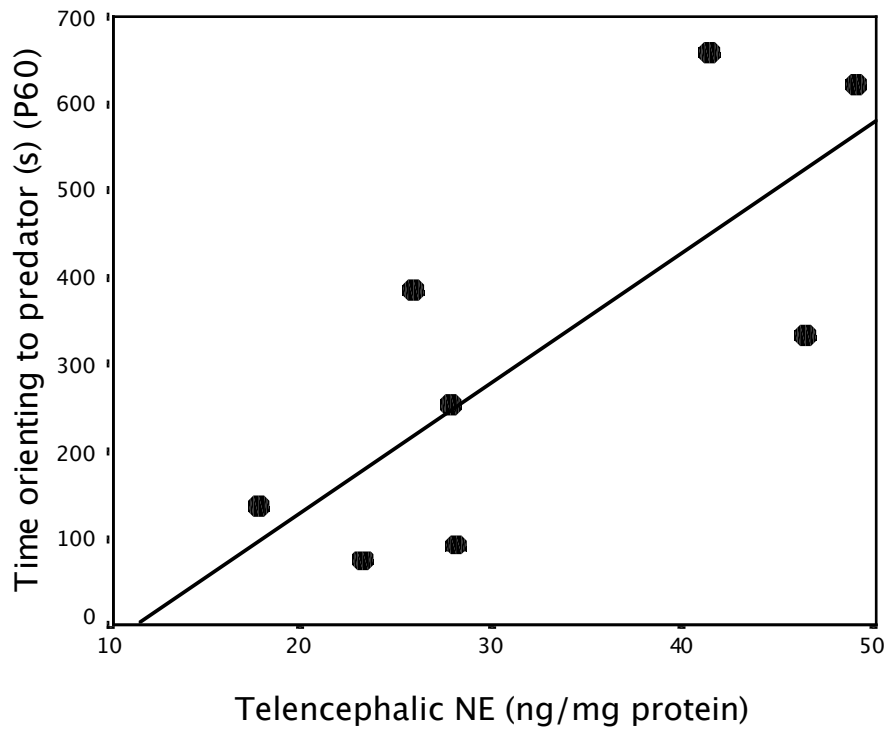
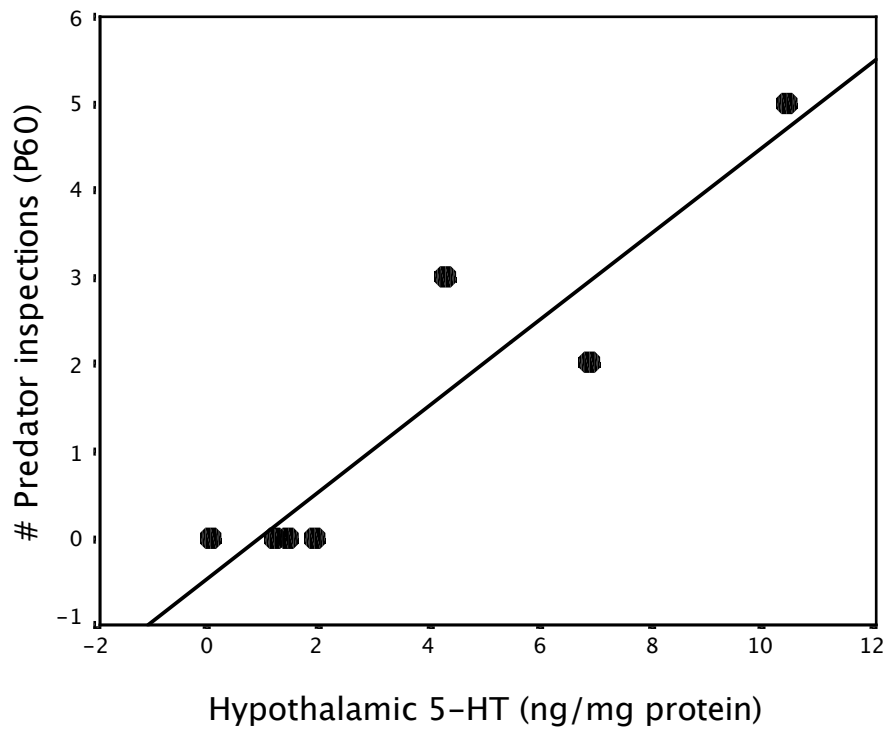


Figure 4

(A)



(B)



(C)

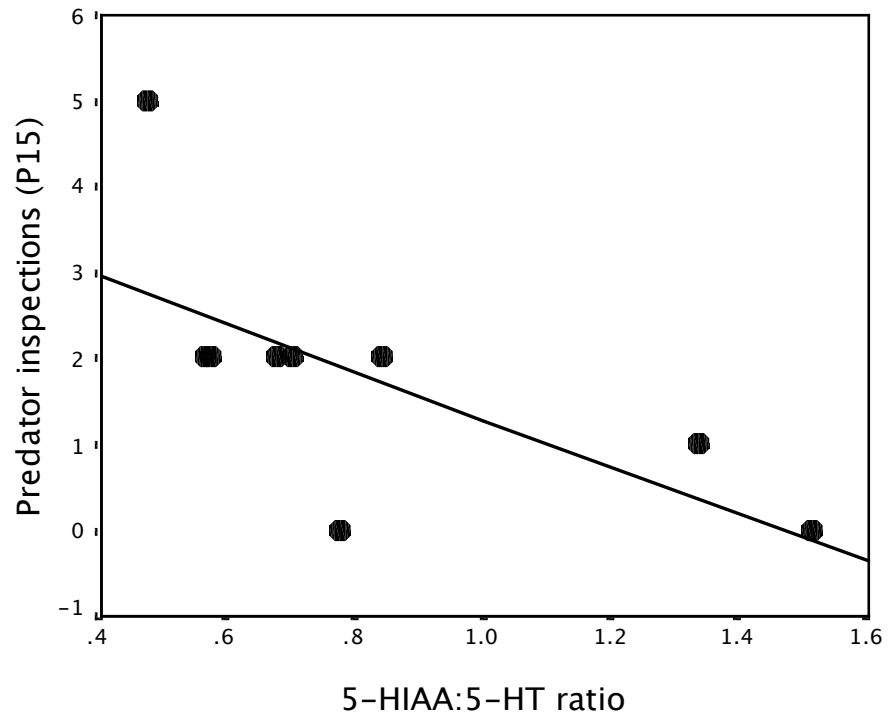


Table 1. Concentrations (ng/mg protein) of monoamines in the different brain regions for the different treatments. Statistics are presented as mean \pm sd. Sample sizes are in parentheses.

	<i>NE</i>	<i>DOPAC</i>	<i>5-HIAA</i>	<i>DA</i>	<i>HVA</i>	<i>5-HT</i>
HYPOTHALAMUS						
Control	10.48 \pm 12.91(10)	4.04 \pm 3.76(6)	3.16 \pm .737(4)	2.27 \pm 1.57(10)	0.84 \pm 0.50(10)	5.83 \pm 4.32(10)
Conspecific						
15 min	1.16 \pm 0.59(7)	3.60 \pm 4.71(3)	und	3.16 \pm 1.82(8)	0.78 \pm .55(5)	und
30 min	0.84 \pm 0.83(6)	3.03 \pm 3.40(3)	und	2.04 \pm 1.91(5)	1.00 \pm 1.13(6)	2.27 \pm 2.54(3)
60 min	1.44 \pm 2.47(8)	2.65 \pm 3.19(3)	und	4.43 \pm 5.01(4)	2.68 \pm 3.38(7)	4.26 \pm 4.56(8)
Predator						
15 min	25.57 \pm 15.63(10)	2.62 \pm .94(6)	3.96 \pm 1.77(9)	4.11 \pm 4.07(10)	1.13 \pm 1.34(10)	4.67 \pm 2.48(10)
30 min	21.36 \pm 10.53(8)	4.24 \pm .56(5)	2.07 \pm 0.74(8)	2.92 \pm 3.09(8)	0.21 \pm 0.14(7)	4.81 \pm 3.45(8)
60 min	27.86 \pm 8.21(8)	1.07 \pm 1.07(4)	3.33 \pm 1.56(8)	3.36 \pm 2.73(8)	0.58 \pm 0.41(8)	3.75 \pm 3.44(8)
RETICULAR FORMATION						
Control	9.04 \pm 11.20(10)	6.10 \pm 2.73(6)	2.57 \pm 1.41(4)	1.84 \pm 1.08(9)	0.41 \pm 0.17(10)	3.19 \pm 2.45(10)
Conspecific						
15 min	1.32 \pm .84(6)	2.54 \pm 2.47(8)	und	3.27 \pm 1.58(6)	0.84 \pm .49(5)	20.30 \pm 0(1)
30 min	0.83 \pm .56(6)	4.38 \pm 3.24(5)	und	1.98 \pm 0.80(7)	0.95 \pm 0.53(7)	1.80 \pm 1.54(3)
60 min	1.08 \pm 0.71(8)	4.49 \pm 4.18(7)	und	1.57 \pm 1.21(4)	0.87 \pm 0.69(8)	2.07 \pm 2.36(8)
Predator						
15 min	21.51 \pm 10.78(10)	und	2.06 \pm .79(8)	2.26 \pm 1.04(10)	1.45 \pm 1.90(10)	2.11 \pm 1.37(10)
30 min	15.53 \pm 6.40(8)	und	1.46 \pm .66(8)	2.18 \pm 2.14(8)	0.15 \pm 0.14(7)	2.10 \pm 1.83(8)
60 min	19.88 \pm 8.26(9)	und	1.98 \pm .70(8)	1.32 \pm 1.23(9)	0.69 \pm .96(8)	2.58 \pm 1.45(8)
TELENCEPHALON						
Control	11.44 \pm 14.59(10)	3.60 \pm 1.80(6)	3.43 \pm 1.59(5)	2.63 \pm 1.29(10)	0.84 \pm 0.49(10)	6.36 \pm 4.46(10)
Conspecific						
15 min	1.36 \pm 0.68(8)	2.14 \pm 2.23(8)	14.39 \pm 19.61(2)	2.11 \pm 0.72(8)	0.64 \pm .82(3)	und
30 min	7.60 \pm 16.94(8)	3.41 \pm 2.74(6)	2.12 \pm 0(1)	1.31 \pm 1.13(6)	0.75 \pm 0.53(8)	5.15 \pm 2.10(2)
60 min	1.54 \pm 2.17(8)	6.48 \pm 2.88(6)	1.24 \pm 0(1)	1.68 \pm 2.37(4)	0.45 \pm 0.71(8)	1.92 \pm 3.56(10)
Predator						
15 min	24.18 \pm 12.70(10)	53.64 \pm 0(1)	3.01 \pm 2.40(9)	4.71 \pm 3.61(10)	2.20 \pm 2.59(10)	4.75 \pm 3.42(9)
30 min	21.07 \pm 14.45(8)	und	2.53 \pm 2.79(8)	2.20 \pm 1.41(8)	0.42 \pm 0.26(8)	3.52 \pm 3.93(8)
60 min	29.24 \pm 14.30(10)	0.37 \pm 0(1)	4.04 \pm 2.38(9)	3.87 \pm 2.77(10)	0.86 \pm 0.71(10)	6.59 \pm 4.18(9)