

2  
3  
4 SHORT COMMUNICATION5  
6  
7 **Immunomodulatory arsenal of nymphal ticks**8  
9  
10 K. PETERKOVÁ<sup>1</sup>, I. VANČOVÁ<sup>1</sup>, V. HAJNICKÁ<sup>1</sup>, M. SLOVÁK<sup>2</sup>,  
11 L. ŠIMO<sup>2</sup> and P. A. NUTTALL<sup>3</sup>12  
13 <sup>1</sup>Department of Xxx, Institute of Virology, Slovak Academy of Sciences, Bratislava, Slovakia, <sup>2</sup>Department of Xxx, Institute of  
14 Zoology, Slovak Academy of Sciences, Bratislava, Slovakia and <sup>3</sup>Department of Xxx, Centre for Ecology and Hydrology, Oxford, U.K.15  
16  
17 **Abstract.** Ticks have developed their own immunomodulatory mechanisms to inhibit  
18 the host inflammatory response. One of them involves the ability to subvert the cytokine  
19 network at the site of tick feeding by secreting cytokine binding molecules. Most studies  
20 have focused on the immunomodulatory prowess of adult female ticks. Here we de-  
21 scribe anti-cytokine activity in salivary gland extracts (SGEs) prepared from 2-day-fed  
22 nymphs of *Dermacentor reticulatus* Fabricius, *Ixodes ricinus* L., *Rhipicephalus appen-*  
23 *diculatus* Neumann and *Amblyomma variegatum* Fabricius. Anti-CXCL8 activity was  
24 detected in nymphs of all species. Relatively high activity against CCL2, CCL3 and  
25 CCL11 was observed in SGEs of *R. appendiculatus* and *A. variegatum* nymphs, whereas  
26 SGEs of *I. ricinus* nymphs showed comparatively high anti-interleukin-2 (-IL-2) and  
27 anti-IL-4 activities. These data show that nymphs, which epidemiologically are usually  
28 more important than adults as disease vectors, possess a range of anti-cytokine activities  
29 that may facilitate pathogen transmission.  
3031  
32 **Key words.** Cytokine inhibitors, Ixodid ticks, nymph salivary glands.33  
34  
35 Feeding ticks stay attached to their hosts for several days or  
36 weeks, depending on the species and developmental stage. The  
37 prolonged feeding period provides ample time for inflammation  
38 to promote haemostasis at the feeding site. Host immune mech-  
39 anisms may reduce the feeding success of ticks by enhancing  
40 inflammatory reactions. Ticks have developed mechanisms to  
41 subvert the host response, presumably as an adaptation to obtain  
42 larger bloodmeals that would result in increased tick fitness. In  
43 particular, the saliva of ticks has anti-inflammatory and immu-  
44 nosuppressive properties (Brossard & Wikel, 2004).45  
46 The host response to foreign antigens requires the co-ordinated  
47 action of innate and acquired components of the immune system,  
48 which is regulated by small secreted proteins known as cytokines  
49 (Borish & Steinke, 2003). Cytokines are a diverse group of solu-  
50 ble messenger proteins involved in the activation, growth, control  
51 and repair of cells, and regulation of immune events. Chemok-  
52 ines, a sub-set of cytokines, play an important role in controlling  
53 leucocyte migration. In previous studies, saliva and/or salivary  
54 gland extract (SGE) of ixodid (hard) adult tick species was shown  
55 to bind numerous cytokines (interleukin-2, IL-4 and some impor-  
56 tant chemokines) and suppress the activity of immune cells that  
57 are responsive to their stimulation. Results varied between spe-58  
59  
60 cies, and also between adult males and females of the same spe-  
61 cies (Gillespie *et al.*, 2001; Hajnická *et al.*, 2001, 2005).  
62 Manipulation of the host cytokine network by ticks provides a  
63 mechanism to help ticks feed and may also facilitate tick-borne  
64 pathogen transmission (Nuttall & Labuda, 2004).65  
66 The nymphal stage is often the most important in tick-borne  
67 pathogen transmission. Several studies have shown that nym-  
68 phal feeding induces changes to host haemostatic and immune  
69 responses, with some evidence of differences between nymphs  
70 and adults (Brossard & Wikel, 2004; Narasimhan *et al.*, 2007;  
71 Pedra *et al.*, 2007). To determine whether nymphs have immu-  
72 nomodulatory mechanisms similar to adults, we compared anti-  
73 cytokine activity in SGEs prepared from nymphs of four ixodid  
74 tick species, *Dermacentor reticulatus*, *Ixodes ricinus*, *Rhipi-*  
75 *cephalus appendiculatus* and *Amblyomma variegatum*, all of  
76 which are important vectors of tick-borne pathogens (De Vos,  
77 1981; Camus & Barre, 1992; Nambota *et al.*, 1994; Hubalek  
78 *et al.*, 1997; Labuda & Nuttall, 2004; Foldvari *et al.*, 2005;  
79 Kelly, 2006; Sreter-Lancz *et al.*, 2006; Skarphedinsson *et al.*,  
80 2007). The materials and methods used followed those de-  
81 scribed in our previous studies with adult ticks (Hajnická *et al.*,  
82 2005; Vančová *et al.*, 2006).83  
84  
85 Correspondence: Professor Pat A. Nuttall, NERC Centre for Ecology and Hydrology, Maclean Building, Benson Lane, Crowmarsh Gifford,  
86 Wallingford, Oxfordshire OX10 8BB, U.K. Tel.: + 44 1491 692560; Fax: + 44 1491 692598; E-mail: pan@ceh.ac.uk87  
88 © 2008 The Authors89  
90 Journal compilation © 2008 The Royal Entomological Society

1

2

3

4

5

Nymphs were allowed to feed on rabbits, on which complete engorgement takes approximately 5–8 days for *D. reticulatus* nymphs, 3–6 days for *I. ricinus*, 5–9 days for *R. appendiculatus*, and 5–9 days for *A. variegatum* (Honzáková, 1971; Jones et al., 1988). As anti-cytokine activity of ixodid species is most consistently detected after feeding commences but prior to engorgement, nymphs were collected when they had completed approximately 2 days of feeding (Vančová et al., 2006). Approximately 300 partially fed nymphs of each species were collected and SGEs prepared as described previously (Slovák et al., 2000). The total amounts of protein from 10 nymphs obtained from two independent feeding sessions were 2.8 µg and 3.9 µg in *D. reticulatus*, 5.1 µg and 3.5 µg in *I. ricinus*, 5.5 µg and 6.8 µg in *R. appendiculatus*, and 11.6 µg and 9.7 µg in *A. variegatum*. Pooled SGE was prepared as 10, 5, 2.5, 1 or 0.5 nymphal equivalents per 5 µl. Salivary gland extracts were screened by ELISA for activity against human CXCL8, CCL2, CCL3, CCL5, CCL11, IL-2 and IL-4 using commercial ELISA kits obtained from R&D Systems (Xxx) and/or Bender MedSystems Diagnostics (Xxx), as described previously, with duplicate assays of each sample (Hajnická et al., 2005). The results represent the means obtained with the two batches of SGEs derived from independent feeding sessions. A reduction in the detectable level of a particular cytokine, compared with the control, was interpreted as evidence of putative cytokine binding activity.

The SGEs of all the nymphal species reduced the level of CXCL8 (Fig. 1). The highest levels of inhibition were shown by *R. appendiculatus* and *D. reticulatus*. Thus there was no correlation between the total protein content of SGE from each species and the levels of inhibitory activity (*D. reticulatus* had the lowest protein content and *R. appendiculatus* the second highest). Relatively high activity against CCL2, CCL3 and CCL11 was observed in *R. appendiculatus* and *A. variegatum* nymphs, whereas activity was barely detectable in *D. reticulatus* and undetectable in *I. ricinus*. Only *A. variegatum* showed significant levels of activity with CCL5. Anti-IL-2 activity was detected in SGE of *I. ricinus* nymphs and low levels of activity in SGE of *D. reticulatus* nymphs, whereas anti-IL-4 was demonstrated in SGE of *I. ricinus* and *R. appendiculatus*. Thus nymphs of four ixodid tick species showed contrasting patterns of anti-cytokine activity after 2 days of feeding on rabbits. Similar results were obtained using murine (rather than human) cytokines (data not shown), reflecting the high degree of amino acid identity between mammalian cytokines and the likelihood that anti-cytokine activity is effective irrespective of (mammalian) host species.

For *I. ricinus* and *A. variegatum*, nymphal anti-cytokine profiles were similar to those recorded for adults, whereas adult *D. reticulatus* showed a much greater repertoire of anti-cytokine activity compared with conspecific nymphs (Table 1). The differences between species and between stages may reflect differences in host preference. However, *I. ricinus* has probably the most catholic 'taste', but appears to have the poorest anti-cytokine repertoire. An alternative explanation may be that anti-cytokine activity reflects the size of the mouthparts and/or the duration of feeding. For example, *A. variegatum* has large mouthparts that penetrate deep into the dermis, and takes

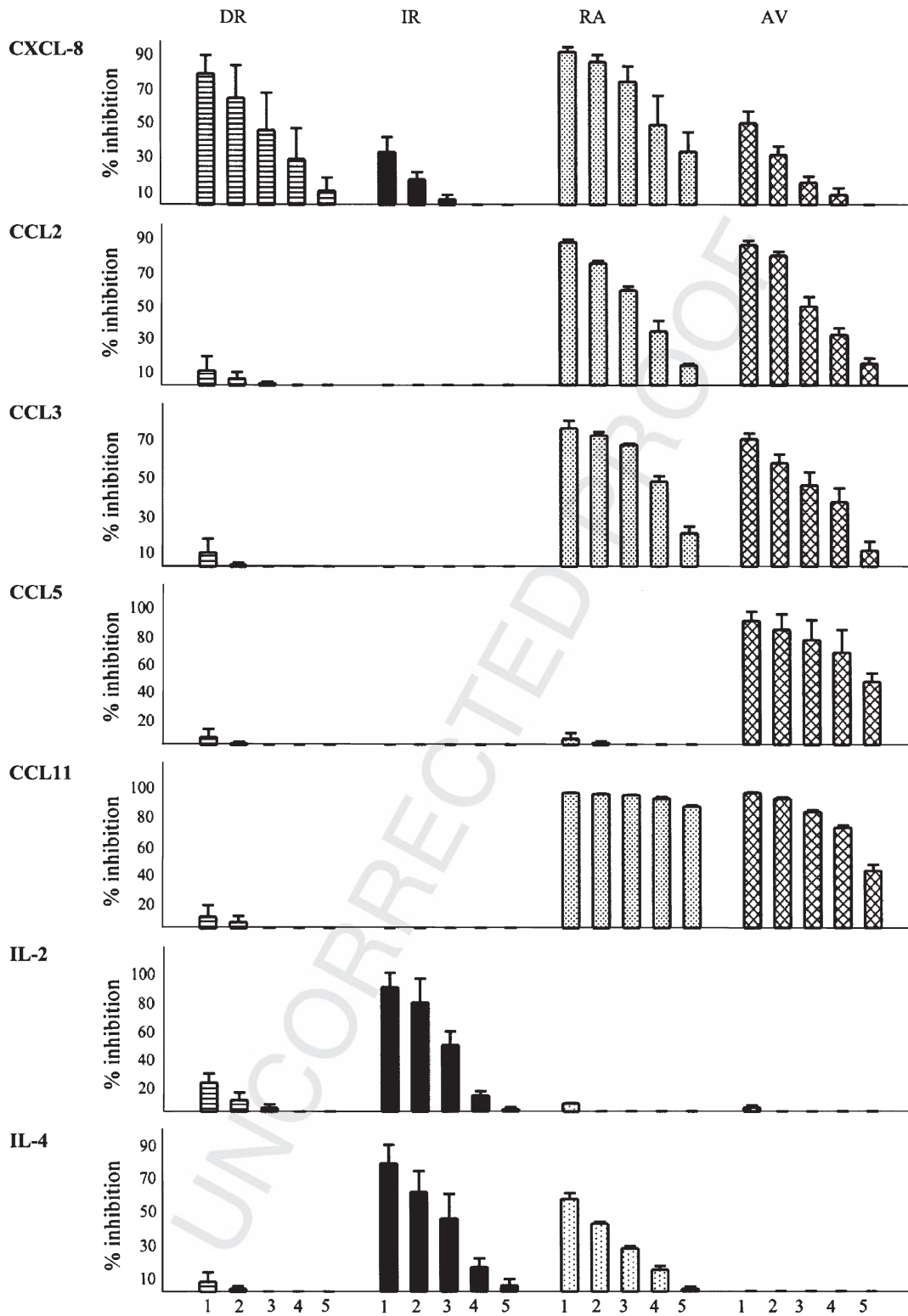
a comparatively long time to reach engorgement (Stewart et al., 1998). The mechanics and physiology of *A. variegatum* feeding may antagonize different cytokines to those provoked by a species such as *I. ricinus*, which has much smaller mouthparts and engorges faster.

Soon after tissue damage, specific leucocyte subsets emigrate from the circulation into the affected area. These leucocytes function as the primary line of host defence in the destruction of micro-organisms and initiation of tissue repair. The histopathology of tick-bite lesions shows that, depending on the tick species and host species, the predominant cells infiltrating attachment sites are neutrophils (in mammals) or heterophils (in non-mammals), eosinophils and basophils (Latif et al., 1990; Szabo & Bechara, 1999; Van der Heijden et al., 2005). Neutrophils are the first infiltrating cell type in the dermis; their migration to inflammatory sites is directed by the chemokine CXCL8. The molecular structure of CXCL8 has been determined for various vertebrate species and shown to be similar. Indeed, the most ancient chemokine, found in a primitive group of vertebrates, resembles mammalian CXCL8, indicating high conservation of this chemokine since the evolution of early vertebrates (Najakshin et al., 1999). The chemotactic ability of human CXCL8 is not species-specific; granulocytes from many vertebrate species migrate to this chemokine *in vitro* (Röt, 1991). Polymorphonuclear neutrophils inform and shape immune responses. Thus it is perhaps not surprising that all the ixodid species and stages showed anti-CXCL8 activity, presumably indicating the importance to ixodid ticks of controlling neutrophil activity. Even adult female *A. variegatum*, which has low anti-CXCL8 activity at 5 days of feeding, shows comparatively higher activity earlier in feeding (Vančová et al., 2006).

The CC chemokines have pleiotropic activities; they are potent attractants for monocytes, eosinophils, basophils, natural killer cells and memory T cells (Laing & Secombes, 2004). The importance of cells of the host immune system infiltrating the tick feeding site resides in their ability to produce cytokines that modulate the downstream response (Falcone et al., 2001). Subversion of the activity of the four CC chemokines examined appears important for the two larger nymphal species, *R. appendiculatus* and *A. variegatum*.

Because of the relatively long duration of tick blood-feeding, ticks must suppress host immune reactions at all levels. The main function of IL-2 is to stimulate the growth and cytotoxic response of activated T lymphocytes. In addition, IL-2 is implicated in the development, homeostasis and function of natural killer cells. For nymphal *I. ricinus* in particular, the results suggest the importance of suppressing one or more of these functions.

The adaptive immune system has evolved two types of immune cells, Th1 and Th2, as the system supervisors (Kidd, 2003). Th1 cells are predominantly involved in the type-1 pathway of cellular immunity, whereas Th2 cells drive the type-2 pathway of humoral immunity. Th2 differentiation is a central process in the protection against parasites such as helminths. Tick infestation also results in a Th2 immune response, as shown by the cytokine profile induced in murine lymph node cells (Ferreira & Silva, 1999). IL-4 is a key cytokine in the induction of Th2 immunity, mediating B-cell activation. Activated



**Fig. 1.** Anti-cytokine activities of salivary gland extract (SGE) obtained from 2-day-fed nymphs of *Dermacentor reticulatus* (DR), *Rhipicephalus appendiculatus* (RA) and *Amblyomma variegatum* (AV) ticks. Salivary gland extracts equivalent to 10.0, 5.0, 2.5, 1.0 or 0.5 nymphs (labelled 1, 2, 3, 4, 5, respectively) were pre-incubated with 50 pg of each cytokine for 90 min before ELISA analysis. Results are expressed as percentage reduction of OD reading compared with control.

14

19



- Ferreira, B.R. & Silva, J.S. (1999) Successive tick infestations selectively promote a T-helper 2 cytokine profile in mice. *Immunology*, **96**, 434–439.
- Foldvari, G., Hell, E. & Farkas, R. (2005) *Babesia canis canis* in dogs from Hungary: detection by PCR and sequencing. *Veterinary Parasitology*, **127**, 221–226.
- Gillespie, R.D., Dolan, M.C., Piesman, J. & Titus, R.G. (2001) Identification of an IL-2 binding protein in the saliva of the Lyme disease vector tick, *Ixodes scapularis*. *Journal of Immunology*, **166**, 4319–4327.
- Hajnická, V., Kocáková, P., Sláviková, M., Slovák, M., Gašperík, J., Fuchsberger, N. & Nuttall, P.A. (2001) Anti-interleukin-8 activity of tick salivary gland extracts. *Parasite Immunology*, **23**, 483–489.
- Hajnická, V., Vančová, I., Kocáková, P. *et al.* (2005) Manipulation of host cytokine network by ticks: a potential gateway for pathogen transmission. *Parasitology*, **130**, 333–342.
- Honzáková, E. (1971) Development of some tick species under standard laboratory conditions. *Folia Parasitologica*, **18**, 357–363.
- Hubalek, Z., Sixl, W., Halouzka, J. & Mikulaskova, M. (1997) Prevalence of *Francisella tularensis* in *Dermacentor reticulatus* ticks collected in adjacent areas of the Czech and Austrian Republics. *Central European Journal of Public Health*, **5**, 199–201.
- Jones, L.D., Davies, C.R., Steele, G.M. & Nuttall, P.A. (1988) The rearing and maintenance of ixodid and argasid ticks in the laboratory. *Animal Technology*, **39**, 99–106.
- Kapsenberg, M.L. (2003) Dendritic-cell control of pathogen-driven T-cell polarization. *Nature Reviews Immunology*, **3**, 984–993.
- Kasama, T., Miwa, Y., Isozaki, T., Odai, T., Adachi, M. & Kunkel, S.L. (2005) Neutrophil-derived cytokines: potential therapeutic targets in inflammation. *Current Drug Targets Inflammation and Allergy*, **4**, 273–279.
- Kelly, P.J. (2006) *Rickettsia africae* in the West Indies. *Emerging Infectious Diseases*, **12**, 224–226.
- Kidd, P. (2003) Th1/Th2 balance: the hypothesis, its limitations, and implications for health and disease. *Alternative Medicine Review*, **8**, 223–246.
- Labuda, M. & Nuttall, P.A. (2004) Tick-borne viruses. *Parasitology*, **129**(Suppl.), 221–245.
- Laing, K.J. & Secombes, C.J. (2004) Chemokines. *Developmental and Comparative Immunology*, **28**, 443–460.
- Latif, A.A., Maina, J.N., Dhadialla, T.S. & Nokoe, S. (1990) Histological reactions to bites of *Amblyomma variegatum* and *Rhipicephalus appendiculatus* (Acari: Ixodidae) fed simultaneously on naïve or sensitized rabbits. *Journal of Medical Entomology*, **27**, 316–323.
- Min, B., Le Gros, G. & Paul, W.E. (2006) Basophils: a potential liaison between innate and adaptive immunity. *Allergy International*, **55**, 99–104.
- Najakshin, A.M., Mechetina, L.V., Alabyev, B.Y. & Taranin, A.V. (1999) Identification of an IL-8 homolog in lamprey (*Lampetra fluviatilis*): early evolutionary divergence of chemokines. *European Journal of Immunology*, **29**, 375–382.
- Nambota, A., Samui, K., Sugimoto, C., Kakuta, T. & Onuma, M. (1994) Theileriosis in Zambia: aetiology, epidemiology and control measures. *Japanese Journal of Veterinary Research*, **42**, 1–18.
- Narasimhan, S., DePonte, K., Marcantonio, N. *et al.* (2007) Immunity against *Ixodes scapularis* salivary proteins expressed within 24 hours of attachment thwarts tick feeding and impairs *Borrelia* transmission. *PLoS ONE*, **2**, e451.
- Nuttall, P.A. & Labuda, M. (2004) Tick–host interactions: saliva-activated transmission. *Parasitology*, **129**(Suppl.), 177–190.
- Pedra, J.H., Narasimhan, S., DePonte, K., Maracantonio, N., Kantor, F.S., Fikrig, E. (2007) Disruption of the salivary protein 14 in *Ixodes scapularis* nymphs and impact on pathogen acquisition. *American Journal of Tropical Medicine and Hygiene*, **75**, 677–682.
- Rot, A. (1991) Chemotactic potency of recombinant human neutrophil attractant/activation protein-1 (interleukin-8) for polymorphonuclear leucocytes of different species. *Cytokine*, **3**, 21–27.
- Skarphedinsson, S., Lyholm, B.F., Ljungberg, M., Sogaard, P., Kolmos, H.J. & Nielsen, L.P. (2007) Detection and identification of *Anaplasma phagocytophilum*, *Borrelia burgdorferi*, and *Rickettsia helvetica* in Danish *Ixodes ricinus* ticks. *Acta Pathologica, Microbiologica et Immunologica*, **115**, 225–230.
- Slovák, M., Hajnická, V., Labuda, M. & Fuchsberger, N. (2000) Comparison of the protein profiles of salivary gland extracts derived from three species of unfed and partially fed ixodid ticks analysed by SDS-PAGE. *Folia Parasitologica*, **47**, 67–71.
- Sreter-Lancz, Z., Szell, Z., Kovacs, G., Egyed, L., Marialigeti, K. & Sreter, T. (2006) Rickettsiae of the spotted-fever group in ixodid ticks from Hungary: identification of a new genotype ('Candidatus Rickettsia kotlanii'). *Annals of Tropical Medicine and Parasitology*, **100**, 229–236.
- Stewart, J.R., Burgdorfer, W. & Needham, G. (1998) Evaluation of three commercial tick removal tools. *Wilderness and Environmental Medicine*, **9**, 137–142.
- Szabo, M.P. & Bechara, G.H. (1999) Sequential histopathology at the *Rhipicephalus sanguineus* tick feeding site on dogs and guinea pigs. *Experimental and Applied Acarology*, **23**, 915–928.
- Vančová, I., Slovák, M., Hajnická, V. *et al.* (2006) Differential anti-chemokine activity of *Amblyomma variegatum* adult ticks during blood-feeding. *Parasite Immunology*, **29**, 169–177.
- Van der Heijden, K.M., Szabo, M.P., Egami, M.I., Pereira, M.C. & Matushima, E.R. (2005) Histopathology of tick-bite lesions in naturally infested capybaras (*Hydrochoerus hydrochaeris*) in Brazil. *Experimental and Applied Acarology*, **37**, 245–255.
- Webb, L.M.C. & Alcamí, A. (2005) Virally encoded chemokine binding proteins. *Mini Reviews in Medicinal Chemistry*, **5**, 833–848.

Accepted 7 January 2008

# Author Query Form

**Journal: Medical and Veterinary Entomology**

**Article: mve\_726**

Dear Author,

During the copy-editing of your paper, the following queries arose. Please respond to these by marking up your proofs with the necessary changes/additions. Please write your answers on the query sheet if there is insufficient space on the page proofs. Please write clearly and follow the conventions shown on the attached corrections sheet. If returning the proof by fax do not write too close to the paper's edge. Please remember that illegible mark-ups may delay publication. Many thanks for your assistance.

Query No.	Query	Remark
1	Au: insert name of department.	
2	Au: insert name of department.	
3	Au: insert name of department.	
4	Au: Please provide the department name (if any) for all the affiliations.	
5	Au: Please note that in text citation 'Vančova <i>et al.</i> (2006) has been changed to Vančová <i>et al.</i> (2006) as per the list. Please check.	
6	Au: insert town, state (abbreviation), country for R&D Systems.	
7	Au: insert town, state (abbreviation), country for Bender MedSystems Diagnostics.	
8	Au: Please note that in text citation 'van der Heijden <i>et al.</i> (2005) has been changed to Van der Heijden <i>et al.</i> (2005) as per the list. Please check.	
9	Au: Please note that spelling of author name 'Kurtenbach' in text citation 'Dizij & Kurtenbach (1995)' has been changed to 'Kurtnebach' as per the list. Please check.	
10	Au: give VEGA in full.	
11	Au: Please note that 'Kapsenberg (2003)' has not been cited in text. Please cite it in text or delete from the list.	
12	Au: Please note that 'Kasama <i>et al.</i> (2005)' has not been cited in text. Please cite it in text or delete from the list.	
13	Au: please check and correct journal title.	

---

14 Au: define OD.

---

15 Au: ‡ does not appear in the table; please correct.

---

16 Au: define N.

---

17 Au: define F.

---

18 Au: define M.

---

19 Au: Figure 1 is in low resolution with poor quality. Please provide high resolution figure at 300 dpi. For more information about supplying electronic artwork, please see the journal webpage or our electronic artwork guidelines at <http://www.blackwellpublishing.com/authors/digill.asp>.

# MARKED PROOF

## Please correct and return this set

Please use the proof correction marks shown below for all alterations and corrections. If you wish to return your proof by fax you should ensure that all amendments are written clearly in dark ink and are made well within the page margins.

<i>Instruction to printer</i>	<i>Textual mark</i>	<i>Marginal mark</i>
Leave unchanged	... under matter to remain	Ⓟ
Insert in text the matter indicated in the margin	∧	New matter followed by ∧ or ∧ <sup>Ⓢ</sup>
Delete	/ through single character, rule or underline or ┌───┐ through all characters to be deleted	Ⓞ or Ⓞ <sup>Ⓢ</sup>
Substitute character or substitute part of one or more word(s)	/ through letter or ┌───┐ through characters	new character / or new characters /
Change to italics	— under matter to be changed	↙
Change to capitals	≡ under matter to be changed	≡
Change to small capitals	≡ under matter to be changed	≡
Change to bold type	~ under matter to be changed	~
Change to bold italic	≈ under matter to be changed	≈
Change to lower case	Encircle matter to be changed	≡
Change italic to upright type	(As above)	⊕
Change bold to non-bold type	(As above)	⊖
Insert 'superior' character	/ through character or ∧ where required	Υ or Υ under character e.g. Υ or Υ
Insert 'inferior' character	(As above)	∧ over character e.g. ∧
Insert full stop	(As above)	⊙
Insert comma	(As above)	,
Insert single quotation marks	(As above)	ʹ or ʸ and/or ʹ or ʸ
Insert double quotation marks	(As above)	“ or ” and/or ” or ”
Insert hyphen	(As above)	⊥
Start new paragraph	┌	┌
No new paragraph	┐	┐
Transpose	┌┐	┌┐
Close up	linking ○ characters	○
Insert or substitute space between characters or words	/ through character or ∧ where required	Υ
Reduce space between characters or words		↑