The correspondence section is a public forum and, as such, is not peer-reviewed. EHP is not responsible for the accuracy, currency, or reliability of personal opinion expressed herein; it is the sole responsibility of the authors. EHP neither endorses nor disputes their published commentary.

Good Laboratory Practices Are Not Synonymous with Good Scientific Practices, Accurate Reporting, or Valid Data

doi:10.1289/ehp.0901495

View metadata, citation and similar papers at <u>core.ac.uk</u>

and she defended the reliance on Good Laboratory Practices (GLP) in animal studies concerning risks posed by chemicals. Her commentary, however, provides additional evidence that her research on BPA is flawed and that GLP can be unreliable.

The key evidence can be found in her treatment of the effect of BPA on prostate weight (Tyl et al. 2008). This effect is important because Tyl's data on adult prostate in mice has been used by the chemical industry-which has funded all of Tyl's research on BPA-and the Food and Drug Administration (FDA) to conclude that BPA has no effect at low doses. Indeed, Tyl argued that the weight of the evidence supports her findings that BPA is safe (all industry-funded studies report no low-dose effects of BPA). In contrast, > 200 studies in experimental animals, all funded by government agencies, have reported significant effects of BPA at low doses that are relevant to human and ecologic exposures (vom Saal et al. 2007).

We (Myers et al. 2009) concluded that prostate weights reported by Tyl et al. (2008) were abnormally high in control males, suggesting either that the dissections were done improperly, that control animals were exposed to a contaminating estrogen, or that their prostates were diseased. This would render the results invalid and therefore inappropriate to use in assessing BPA safety. It would also provide insights as to why, despite many other studies showing adverse effects of exposure to BPA at low doses (vom Saal et al. 2007), Tyl et al. (2008) detected none.

To counter this criticism, Tyl (2009) presented a table (her Table 2) of mouse prostate weights from other laboratories. The data she presented in fact show that no other laboratory measuring prostate weight in mice has reported mean weights as high as those reported by Tyl et al. (2008), except in old male mice with diseased prostates. Tyl's table cites data from research published by Heindel et al. (1995) previously conducted at her own institution, Research Triangle Institute, although she did not acknowledge this. The mean prostate weight reported by Heindel et al. (1995) for 16- to 17-week-old CD-1 male mice was 48 mg, which is similar to most other findings, but contrasts sharply with Tyl's mean prostate weights of 74 mg for the F_1 males in her BPA study (these males were identified in Table 1 of Tyl's commentary as being examined at 18 weeks of age).

Table 2 of Tul (2000) also includes data

in 23-week-old CD-1 mice. However, this study involved comparing data from two laboratories, and Tyl omitted from her table the data from the second laboratory that reported a mean prostate weight of 35 mg in 23-week-old CD-1 males. Morrissey et al. (1988) observed that the laboratory reporting the mean of 58 mg also had a higher standard deviation and lower statistical sensitivity than the laboratory reporting the 35 mg mean prostate weight. In studies in which prostate weight is high, such as that of Tyl et al. (2008), the findings are suspect in that the abnormally high prostate weight data show a poor relationship to other male reproductive organs (Morrissey et al. 1988). This strongly suggests that nonprostatic tissue has been included when prostate weights are abnormally high in the absence of disease.

Tyl's discussion of prostate weight effects also suggests that studies identified as GLP may not adhere to the strict record-keeping goals to which GLP aspires, undermining one of the arguments used for the value of GLP over research funding by the National Institutes of Health, which rarely follows the costly GLP guidelines. In the original publication, Tyl et al. (2008) reported that F1 retained males were necropsied at approximately 14 weeks of age. In Table 1 of her commentary (Tyl 2009), Tyl stated that these males were 18 weeks of age at necropsy. However, in testimony before the FDA Science Board BPA Subcommittee hearing on 16 September 2008 (FDALive.com 2008), Tyl stated that these males were 24 weeks of age at necropsy as an explanation for their high prostate weights. Tyl assured the FDA panel that since "the difference in age influences growth rate and growth of organs, the comparison [of 12- and 24-week-old males] is specious, it is comparing apples and oranges." In fact, Tyl's data in Table 1 of her commentary (Tyl 2009) show no relationship between age and body weight. The inconsistencies in Tyl's FDA testimony, which could have had a significant impact on a regulatory decision concerning BPA, and the data concerning the age at tissue collection, prostate weights, and body weights presented in Table 1 of her

commentary are disturbing, and indicate that a thorough review of original data in Tyl et al. (2008) by scientific experts is warranted.

In addition to serving on the faculty of the University of Missouri, F.S.v.S. is CEO of XenoAnalytical LLC, a small private laboratory that performs assays of xenobiotic compounds, and he has prepared reviews concerning the health effects of bisphenol A for nongovernmental organizations and corporations. J.P.M. is CEO/Chief Scientist for Environmental Health Sciences (EHS), a non-

brought to you by 🗓 CORE

provided by University of Missouri: MOspace

org/about.html) to support EHS's mission to advance public understanding of environmental health sciences.

Frederick S. vom Saal

University of Missouri Columbia, Missouri E-mail: vomsaalf@missouri.edu

John Peterson Myers

Environmental Health Sciences Charlottesville, Virginia

REFERENCES

- FDALive.com. 2008. Meeting of the Bisphenol A Subcommittee of the Science Board. Meeting Date: 9/16/08-9/16/08. Available: http://www.fdalive.com/pastmeetings. cfm?committeekey=61 [accessed 21 October 2009].
- Heindel JJ, Chapin RE, George J, Gulati DK, Fail PA, Barnes LH, et al. 1995. Assessment of the reproductive toxicity of a complex mixture of 25 groundwater contaminants in mice and rats. Fundam Appl Toxicol 25(1):9–19.
- Morrissey RE, Lamb JC, Schwetz BA, Teague JL, Morris RW. 1988. Association of sperm, vaginal cytology, and reproductive organ weight data with results of continuous breeding reproduction studies in Swiss (CD-1) mice. Fundam Appl Toxicol 11(2):359–371.
- Myers JP, vom Saal FS, Akingbemi BT, Arizono K, Belcher S, Colborn T, et al. 2009. Why public health agencies cannot depend on good laboratory practices as a criterion for selecting data: the case of bisphenol A. Environ Health Perspect 117:309–315.
- Tyl RW. 2009. Basic exploratory research versus guidelinecompliant studies used for hazard evaluation and risk assessment: bisphenol A as a case study. Environ Health Perspect 117:1644–1651.
- Tyl RW, Myers C, Marr M, Sloan CS, Castillo N, Veselica MM, et al. 2008. Two-generation reproductive yoxicity study of dietary bisphenol A (BPA) in CD-1(r) (Swiss) mice. Toxicol Sci 104(2):362–384.
- vom Saal FS, Akingbemi BT, Belcher SM, Birnbaum LS, Crain DA, Eriksen M, et al. 2007. Chapel Hill bisphenol A expert panel consensus statement: integration of mechanisms, effects in animals and potential to impact human health at current levels of exposure. Reprod Toxicol 24(2):131–138.

Good Laboratory Practices: Tyl Responds

doi:10.1289/ehp.0901495R

I am responding to the comments by vom Saal and Myers on my commentary (Tyl 2009), which was in response to their commentary (Myers et al. 2009). Our dietary BPA mouse study was performed under Organisation for Economic Co-operation and