

Article (refereed)

Hayes, Felicity; Mills, Gina; Ashmore, Mike. 2010 How much does the presence of a competitor modify the within-canopy distribution of ozone-induced senescence and visible injury? *Water Air and Soil Pollution*, 210. 265-276. [10.1007/s11270-009-0248-9](https://doi.org/10.1007/s11270-009-0248-9)

© Springer Science+Business Media 2010

This version available <http://nora.nerc.ac.uk/4910/>

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the authors and/or other rights owners. Users should read the terms and conditions of use of this material at <http://nora.nerc.ac.uk/policies.html#access>

This document is the author's final manuscript version of the journal article, incorporating any revisions agreed during the peer review process. Some differences between this and the publisher's version remain. You are advised to consult the publisher's version if you wish to cite from this article.

The original publication is available at www.springerlink.com

Contact CEH NORA team at
noraceh@ceh.ac.uk

1 **How much does the presence of a competitor modify the**
2 **within-canopy distribution of ozone-induced senescence**
3 **and visible injury?**

4 Hayes, F.^{a*}, Mills, G.^a, Ashmore, M.^b

5

6 ^aCentre for Ecology and Hydrology, Environment Centre Wales, Deiniol Road, Bangor,
7 Gwynedd, LL57 2UW, UK.

8 ^bEnvironment Department, University of York, Heslington York, YO10 5DD, UK

9

10 *Corresponding Author: Felicity Hayes

11 Tel: 44 (0) 1248 374500

12 Fax: 44 (0) 1248 362133

13 Email: fhay@ceh.ac.uk

14

15 **Summary**

16 • Many natural vegetation species have been shown to be negatively affected by ozone.

17 This study has investigated how the presence of competing species in a community
18 affects two common responses to ozone: visible injury and senescence.

19 • Monocultures and mixtures of *Trifolium repens* and *Lolium perenne* grown in large
20 containers were exposed in solardomes to either an episodic rural ozone profile
21 (AOT40 of 12.86 ppm.h) or control conditions (AOT40 of 0.02 ppm.h) for 12 weeks.

22

- 23 • The proportion of ozone-injured or senesced leaves decreased in the order
24 upper>edge>inner canopy for *T. repens* and *L. perenne*. The presence of *L. perenne*
25 increased the proportion of ozone-injured leaves in *T. repens*, whilst the presence of *T.*
26 *repens* decreased the proportion of senesced leaves in *L. perenne*. In *L. perenne*, the
27 proportion of injured leaves at the edge and inner canopy decreased significantly when
28 grown in competition, whilst for *T. repens* the reverse effect occurred in the inner
29 canopy only.
- 30 • It is proposed that different mechanisms influence the interaction between response to
31 ozone and competitors in these species: the response of *Lolium perenne* to ozone may
32 have been related to nitrogen supply, whilst in *Trifolium repens* canopy structure was
33 more important.

34

35 **Key words**

36 Ozone; visible injury; senescence; stomatal conductance; canopy; competition

37

38 **Introduction**

39 Ambient ozone concentrations in Europe have been shown to cause significant effects on a
40 wide range of plant species. Although the effects vary between species, visible leaf injury and
41 premature senescence are frequently reported from ozone exposure studies (e.g. Bergmann et
42 al., 1999; Novak et al., 2003). In addition, approximately 80 species of semi-natural vegetation
43 have been recorded with symptoms attributed to ozone in ambient air conditions (Hayes et al.,
44 2007). There is a need to improve predications of the impacts of ambient ozone on natural
45 vegetation communities, however, many studies investigate the effects of ozone using single
46 species, and the presence of competing species in a community may affect the response to

47 ozone. Canopy structure and competition are two interlinked factors to be considered as
48 influences on the response to ozone in mixed vegetation communities. To our knowledge, no
49 other studies have investigated both of these factors together.

50

51 For some species, the magnitude of the response to ozone has been shown to be influenced by
52 competition, for example, the grass *Elymus glaucus* increased the impact of ozone exposure on
53 *Pinus ponderosa* (Anderson et al., 2001). Similarly *Poa pratensis* has been demonstrated to be
54 more sensitive to ozone (in terms of visible injury) when grown in competition with *Veronica*
55 *chamaedrys* compared to when grown as a monoculture but not when grown with other species
56 such as *Achillea millefolium* (Bender et al., 2005). In contrast, *Holcus lanatus*, *Lychnis flos-*
57 *cuculi*, *Molinia caerulea* and *Plantago lanceolata* showed no difference in response to ozone
58 when grown in monoculture compared to when grown in competition with *Agrostis capillaris*
59 (Tonneijck et al., 2004).

60

61 Presence within a canopy of vegetation may also influence the response of an individual
62 species to ozone. Few studies have investigated the changing profiles of ozone and light
63 through plant canopies, and these existing studies have tended to involve large canopies such
64 as forests (e.g. Utiyama et al., 2004). Lantinga et al. (1999) showed that PAR was dramatically
65 reduced inside a plant canopy, and in stands of a monoculture of cut-leaved coneflower
66 (*Rudbeckia laciniata* L.), ozone concentrations 20 cm above the ground were only half the
67 concentration of those just above the top of the canopy, which was approximately 1.5 – 2.0
68 metres high (Finkelstein et al., 2004). Within these large stands of cut-leaved coneflower, the
69 extent of ozone injury was lower on plants growing within the stand compared to those on the
70 edge (Chappelka et al., 2003). A similar pattern of ozone concentration within the canopy
71 occurred in the only study to investigate profiles of a grassland canopy, where leaves of low

72 growing *Trifolium repens* received approximately 30% less ozone than *Alopecurus pratensis*,
73 which itself was exposed to slightly lower ozone concentrations than those of the bulk air
74 above the canopy (Jäggi et al., 2006). Thus, there is the potential for differential effects of
75 ozone within mixed canopy grasslands.

76

77 Models of ozone fluxes to natural vegetation communities have been developed (e.g. Bassin et
78 al., 2004, Emberson et al., 2000, 2001, Simpson et al., 2003). These models currently include
79 the influence of environmental variables such as temperature on stomatal conductance and
80 therefore ozone fluxes. Use of a mechanistic model of canopy development of *Lolium perenne*
81 demonstrated the importance of simulation of canopy growth compared to a fixed seasonal
82 profile of leaf area index (Ashmore et al., 2007), however these models do not currently
83 account for differential ozone uptake within different portions of a plant canopy, or differential
84 uptake by different species or functional types.

85

86 In this study, responses of plants grown in monoculture were compared to the responses when
87 grown in mixture, using *Trifolium repens* and *Lolium perenne* as model species that respond to
88 ozone by the development of ozone injury and senescence. Detailed measurements of visible
89 injury and senescence were carried out at different positions in the canopy to investigate
90 whether the presence of a competitor modifies the extent and location of damage within the
91 canopy. Effects in *Trifolium repens* were related to within canopy variation in stomatal
92 conductance.

93

94 **Materials and Methods**

95 **Plant material**

96 Plant material was vegetatively propagated from *Lolium perenne* and *Trifolium repens* plants
97 from turf samples of pasture managed for silage near Edinburgh, UK (Grid reference
98 NT245642). Plants originating from different parents were randomised between different
99 competition and ozone treatments. Individual plants were established for approximately eight
100 weeks before monocultures and mixtures of plants were established for ozone exposure.

101 **Experimental design**

102 Large containers (35.5 cm x 45 cm x 25 cm deep), with holes for drainage, were lined with
103 perforated plastic sheeting to prevent roots from growing out through the bottom and filled
104 with multipurpose compost ('Gem' tub and planter).

105

106 In each pot twelve plants were planted in an evenly spaced arrangement, consisting of four
107 central plants surrounded by eight additional plants. In each mixture, the four central plants
108 were *Trifolium repens* and the eight surrounding plants were *Lolium perenne*. Three pots each
109 of the *Lolium perenne* and *Trifolium repens* monocultures, and three pots of the *Lolium*
110 *perenne* and *Trifolium repens* mixture were randomly allocated to each of four solardomes.

111 Plants were exposed in the solardomes for twelve weeks, starting on 26th July 2002. The
112 exposure period was divided into two harvest periods. Plants were cut back on 6th September,
113 the intermediate harvest, and 16th October, when the final harvest occurred. Plants were kept
114 well-watered throughout the experiment using a mist irrigation system, with additional
115 watering by hand during periods of warm weather.

116

117 **Ozone exposure**

118 Four solardomes were used for exposure. Ozone was generated from oxygen using an ozone
119 generator (Wallace and Tiernan). Ozone concentrations were measured every 30 minutes in
120 each solardome using an ozone analyser (Dasibi 1003-AH) which sampled ozone for a
121 minimum of 3.5 minutes from each solardome using a computer controlled sample selector.
122 Two solardomes were used as controls, with ozone added to charcoal-filtered air using
123 computer controlled (LabView version 6) mass flow controllers to give continuous ozone
124 concentrations in each dome of 30 ppb ($O_3(30)$). An episodic rural ozone profile
125 ($O_3(30+\text{peaks})$) was given over the course of each week to the two other domes. The ozone
126 exposure was programmed to reach a maximum concentration of 80 ppb on days 1 and 4, and a
127 maximum concentration of 100 ppb on days two and three. Ozone concentrations increased
128 from 30 ppb to the daily maximum over the course of 2 hours, remained at the daily maximum
129 for 6 hours, then decreased back down to 30 ppb over the course of 2 hours. Ozone
130 concentrations were programmed to remain at 30 ppb at all other times.

131

132 **Visual assessments**

133 Visual estimates of senescence and ozone-specific injury, apparent as white or pale yellow
134 stipples on the leaf surface, were made for whole pots, because the individual plants had grown
135 together and could not be separated. Leaves were classified as either senesced or injured if
136 >25% of the leaf was senesced or injured respectively, otherwise they were classified as
137 healthy. For *Lolium perenne* senescence of leaves started at the leaf tip and progressed along
138 the leaf blade. The length of the senesced portion (in mm) of the leaf blade was measured on a
139 sub-sample of twenty randomly chosen leaves in each pot.

140 **Harvests**

141 All plants were cut back to a height of 7 cm on 6th September and 16th October, after exposure
142 to the ozone regime for six weeks and 12 weeks respectively. The plants were harvested in
143 separate layers: material growing outside the pot perimeter, material greater than 14 cm above
144 soil level, and plant material between 7 cm and 14 cm above soil level. At the final harvest an
145 additional layer with plant material 0 to 7 cm above soil level was also used. Fresh plant
146 material from each layer was sorted into the component species at the time of harvest. Healthy
147 and ozone-injured leaves of *Trifolium repens* were separated. *Lolium perenne* was sorted into
148 healthy leaves and senesced leaves. Plant material was dried at 65°C for a minimum of 4 days
149 before biomass was determined.

150 **Stomatal conductance measurements**

151 Measurements of stomatal conductance were made on *Trifolium repens* using a porometer
152 (Delta-T AP4) on days of stable meteorological conditions after exposure to the ozone regime
153 for 10/11 weeks. Measurements of stomatal conductance in the upper canopy (where leaves
154 were in full sunlight) and the inner canopy (where leaves were more shaded) were taken, using
155 six leaves (two per pot) for each canopy position in every solardome.

156 **Chlorophyll content**

157 Chlorophyll content (chlorophyll a + b) of leaves of *Trifolium repens* was measured using a
158 SPAD meter (CCM-200, ADC Bioscientific Ltd., UK) after exposure to the ozone treatment
159 for one week and ten weeks. ‘Typical’ leaves were used; therefore some ozone injury was
160 present in some cases. The chlorophyll index, in relative units, given by the SPAD meter, were
161 calibrated for *Trifolium repens* following determination of chlorophyll content by extraction
162 with acetone and measurement of light absorption at wavelengths 470, 646 and 663 nm,
163 according to the equations of Lichtenthaler and Wellburn (1983). The relationship between

164 chlorophyll index and measured chlorophyll (mg g^{-1} fresh weight) had an r^2 of 0.90 (data not
165 presented) and was:

166

167 Chlorophyll content (mg g^{-1} FW) = (chlorophyll index * 30.448) + 417

168

169 **Statistical analysis**

170 For each parameter, values were averaged to provide a mean per solardome prior to subsequent
171 analysis. Statistical analysis was based on these dome means. Visible injury and senescence
172 data were arcsine transformed prior to analysis. Oneway ANOVA (Minitab version 14) was
173 used for analysis of stomatal conductance data. Other comparisons were made in Genstat
174 (version 8) using split-plot or split-split plot ANOVA. The main plot was ozone treatment and
175 the sub-plots were monoculture/mixture. Sub-sub-plots of canopy position were used where
176 appropriate.

177 **Results**

178 **Ozone concentrations**

179 The mean AOT40 for the two domes exposed to the $\text{O}_3(30+\text{peaks})$ episodic ozone regime was
180 9.98 ppm.h during the first harvest interval, and 11.89 ppm.h during the second harvest
181 interval, giving a total of 21.86 ppm.h over the 12 week exposure period (Table 1). The
182 difference in AOT40 between the two replicate $\text{O}_3(30+\text{peaks})$ solardomes was less than 2% for
183 each harvest interval. In the two replicate $\text{O}_3(30)$ solardomes, the mean AOT40 over the
184 exposure period was less than 0.02 ppm.h. 24-hour mean, 12-hour mean and 12-hour mean of
185 episode days also show small differences between the replicate solardomes (Table 1).

186

187 **Influence of *Lolium perenne* on visible injury on clover**

188 Visible injury caused by ozone on *Trifolium repens* was apparent first as small, yellow flecks
189 on the leaves. As the severity increased, the extent of chlorosis increased until eventually the
190 leaf was dry and curled. Visible injury symptoms caused by ozone were first observed on the
191 clover plants after one week of exposure. Very little non-specific senescence (<1 % of leaves)
192 was observed on *T. repens* leaves during the experiment; any senescence that corresponded
193 with the presence of ozone injury symptoms was recorded as “visible injury”.

194

195 At the intermediate harvest, a visual assessment of the O₃(30+peaks) treated *Trifolium repens*
196 plants growing in monoculture showed that 69% of leaves per pot had visible injury symptoms
197 compared to only 0.5% in the O₃(30) treatment (p<0.001). Similar proportions of injury were
198 observed when *Trifolium repens* was grown in combination with *Lolium perenne* - 67%
199 injured leaves in O₃(30+peaks) compared to 0% injured in O₃(30) (p<0.001). At the final
200 harvest the proportion of injured *Trifolium repens* leaves per pot in the O₃(30+peaks) treatment
201 was significantly higher when grown in the mixture compared to when grown in monoculture
202 (77% compared to 67%, p<0.01). There was also an interaction between ozone treatment and
203 whether the plants were grown in monoculture or in mixture (p<0.01), with a larger difference
204 in the extent of visible injury between O₃(30) and O₃(30+peaks) if the plants were grown in
205 mixture with *Lolium perenne*.

206

207 The proportion of injured leaves was also quantified by biomass. Separation of leaves into
208 those that were healthy and those that were injured at the intermediate harvest showed that
209 differences in the biomasses of both healthy leaves and ozone injured leaves were significantly
210 affected by ozone in *Trifolium repens* growing both as a monoculture and as part of the
211 mixture (Table 2). The biomass of injured leaves was approximately two thirds of the total leaf

212 biomass in O₃(30+peaks) treated plants, whereas the biomass of injured leaves was negligible
213 in O₃(30) plants. At the final harvest the total leaf biomass and the biomass of both healthy
214 and injured leaves were significantly affected by ozone in *Trifolium repens* growing both as a
215 monoculture and as part of the mixture (Table 2). The proportion of injured leaves was
216 negligible in O₃(30) treated plants and approximately 80% of the total leaf biomass in
217 O₃(30+peaks) treated plants (Table 2). Due to the difference in the number of *Trifolium repens*
218 plants per pot in the monoculture and mixture, statistical comparison was based on the
219 proportion of injured leaves relative to healthy leaves, rather than the actual biomass. This
220 showed that there was no significant interaction between ozone treatment and whether the
221 plants were grown in monoculture or in mixture.

222

223 The proportion of injured leaves was different in the different regions of the canopy (Figure 1).
224 At the intermediate harvest the highest proportion of injured leaves was in the plant material
225 growing at the edge of the canopy – plant material growing outside the pot perimeter (75% of
226 leaves were injured, p<0.05). The proportion of injured leaves was lower above 14cm – the
227 upper canopy (67%) and lowest in the inner canopy (52%) – plant material between 7cm and
228 14cm. The pattern was similar in the monoculture, and there were no significant effects of
229 whether the plants were grown in monoculture or in mixture, or any significant interaction
230 between this and the ozone treatment.

231

232 At the final harvest the proportion of injured leaves in the monoculture was not significantly
233 different in the different regions of the canopy. There was much less growth outside of the pot
234 perimeter during the second harvest interval (data not presented). In addition, although there
235 was reduced leaf biomass at the final harvest compared to the intermediate harvest (Table 2),

236 the canopy height was the same (data not presented) indicating that the canopy was much more
237 open during the second harvest interval.

238

239 The proportion of injured leaves in the inner canopy (7 – 14cm) was higher in plants growing
240 in mixture with *Lolium perenne* compared to those of the monoculture, where the proportions
241 of injured leaves were 81% and 63% in the mixture and monoculture respectively at the final
242 harvest (Figure 1, $p < 0.01$). There was also an interaction between ozone exposure and whether
243 the plants were grown in monoculture or in mixture for the proportion of injured leaves in the
244 inner canopy ($p < 0.05$), with ozone treatment corresponding with an increased proportion of
245 injured leaves in the mixture. There were no significant differences and no interaction between
246 ozone exposure and whether plants were grown in monoculture or in mixture for the proportion
247 of injured leaves in the upper canopy or the canopy edge.

248 **The influence of *Trifolium repens* on senescence of *Lolium perenne***

249 In contrast to *T.repens*, *L. perenne* responded to ozone by the development of non-specific
250 senescence; no ozone-specific injury was observed during the course of the experiment.

251

252 The large difference in the extent of senescence of $O_3(30+\text{peaks})$ treated *Lolium perenne*
253 compared to $O_3(30)$ was significant at both harvests (Table 3, $p < 0.05$ at each harvest). In the
254 $O_3(30+\text{peaks})$ treatment at the intermediate harvest, the proportion of senesced leaves was
255 approximately 50% for plants growing in the monoculture and in the mixture. At the final
256 harvest, there was a further increase in senescence of plants in the $O_3(30+\text{peaks})$ treatment in
257 the monoculture, to 68%, but a reduction in senescence for plants in mixture with *Trifolium*
258 *repens* to 28%. There was also significantly less senescence of *Lolium perenne* when grown as
259 a mixture compared to as a monoculture in the $O_3(30)$ treatment (0% vs 28%, $p < 0.001$).

260 However, there was no significant interaction between ozone treatment and whether the plants
261 were grown in monoculture or in mixture at either harvest.

262

263 In *Lolium perenne* plants, senescence started at the tip of the leaf blade and progressed back
264 towards the main plant. The extent of the senesced portion of leaf (in mm) was significantly
265 increased in O₃(30+peaks) treated plants compared to O₃(30) plants for both the monoculture
266 and the mixture at both harvests (Table 3, p<0.05). As with the proportion of senesced leaves,
267 the extent of senescence of both O₃(30+peaks) and O₃(30) treated plants was significantly less
268 in the mixture compared to the monoculture at both harvests (p < 0.001 in each case). Again,
269 there was no significant interaction between ozone treatment and whether the plants were
270 grown in monoculture or in mixture at either harvest.

271

272 The biomass of healthy leaves and senesced leaves were not affected by ozone at the
273 intermediate harvest (Table 4), and there was no significant difference in the proportion of
274 senesced leaves of plants grown in monoculture compared to those grown in mixture. The
275 senesced biomass was approximately four-times greater in the O₃(30+peaks) treatment in the
276 monoculture (p<0.01) and approximately two-times greater in the mixture (p<0.1, Table 4).
277 There was no significant interaction between ozone treatment and whether the plants were
278 grown in monoculture or in mixture.

279

280 At the final harvest there was a significant effect of ozone on the biomass of the senesced
281 leaves (p<0.01, Table 4). There was also a large reduction (80%) in the biomass of healthy
282 leaves in the O₃(30+peaks) treatment of the monoculture (p<0.05), whereas the biomass of
283 healthy leaves in the mixture was not significantly affected by ozone treatment (Table 4).

284

285 There was a significant effect of canopy position on the proportion of senesced leaves of
286 *Lolium perenne* ($p < 0.01$ at each harvest; Figure 2). The proportion of senesced leaves of
287 *Lolium perenne* was much lower in the inner canopy than in the upper canopy or canopy edge
288 for plants growing in both the monoculture and the mixture ($p < 0.01$ in both cases). The
289 proportion of senesced leaves of *Lolium perenne* was also much lower overall in the mixture
290 than in the monoculture, although this difference was only statistically significant at the
291 intermediate harvest ($p < 0.01$). However there was no significant interaction between ozone
292 treatment and whether the plants were grown in monoculture or in mixture.

293 **Within-canopy variation in stomatal conductance**

294 There were no significant differences in stomatal conductance of *Trifolium repens* in the
295 monoculture compared to in mixture with *Lolium perenne* (data not presented). However,
296 there was reduced stomatal conductance in the inner canopy compared to the upper canopy of
297 *Trifolium repens* monocultures in both $O_3(30)$ ($p < 0.05$) and $O_3(30+\text{peaks})$ treatments ($p < 0.05$,
298 Table 5). There were also significant differences between the $O_3(30)$ and $O_3(30+\text{peaks})$
299 treatments, with increased stomatal conductance in the inner canopy of $O_3(30+\text{peaks})$ treated
300 plants compared to $O_3(30)$ ($p < 0.05$). There were no significant differences in stomatal
301 conductance between treatments in the upper canopy.

302

303 Corresponding measurements of PAR, measured at the same time as stomatal conductance
304 using a light sensor on the head of the leaf clip of the porometer, indicated that the PAR was
305 different in the different regions of the canopy. PAR was reduced by 88% and 77% in the
306 inner canopy compared to the upper canopy in the $O_3(30)$ and $O_3(30+\text{peaks})$ treatments
307 respectively (Table 5). The PAR in the inner canopy was significantly higher for canopies that
308 received the $O_3(30+\text{peaks})$ treatment compared to $O_3(30)$, $p < 0.01$, however, there was no

309 difference in the relationship between PAR and stomatal conductance between the two ozone
310 treatments (data not presented).

311
312 Stomatal conductance was not related to leaf age. For *Trifolium repens* there was no difference
313 in stomatal conductance of different age leaves along a stolon (i.e. between Leaf 1 the newest
314 fully expanded leaf, Leaf 2 and Leaf 3) in either the O₃(30) or O₃(30+peaks) treatments (data
315 not presented).

316
317 There were no significant differences between ozone treatments in the stomatal conductance of
318 upper canopy leaves of *Lolium perenne* after exposure for 2, 4 or 10 weeks (data not
319 presented).

320 **Within-canopy variation in chlorophyll content**

321 Chlorophyll content of upper canopy leaves was reduced by approximately 12% in leaves of
322 *Trifolium repens* that had been exposed to O₃ (30+peaks) compared to the O₃(30) treatment
323 (p<0.05, Figure 3). However, there were no significant differences between ozone treatments
324 for leaves of the inner canopy.

325
326 There were no differences in the chlorophyll content of leaves of different ages in the O₃(30)
327 treatment, however, there was a significant decrease in the chlorophyll content with increasing
328 leaf age in the O₃(30+peaks) treatment (Figure 4), which corresponded with an increased
329 extent of ozone damage in older leaves. There were no significant differences in chlorophyll
330 content of plants grown in monoculture compared to plants grown in mixture (data not
331 presented).

332 **Discussion**

333 By using two model species representing grasses and legumes, this study has revealed that the
334 presence of a competitor modifies the extent and canopy distribution of two important
335 responses to ozone: visible injury and senescence.

336

337 Overall, a higher proportion of leaves were injured by ozone when *T. repens* was grown in
338 competition with *L. perenne* than when grown in monoculture, with this effect most significant
339 in the inner canopy leaves. Increased sensitivity to ozone when grown in competition has
340 previously been demonstrated on *Poa pratensis* (Bender et al., 2005), where *P. pratensis*
341 developed more ozone injury when grown with competing species such as *Veronica*
342 *chamaedrys* than when grown alone. In contrast, *L. perenne* was not affected as severely by
343 ozone when growing in combination with *T. repens* compared to when growing in
344 monoculture. Indeed, senescence was reduced in the mixture in both the O₃(30) and
345 O₃(30+peaks) treatments, we speculate that in *L. perenne*, since nitrogen transfer from clover
346 to grass in grass-clover swards has been demonstrated in several studies e.g. Sincik & Acikgoz
347 (2007) and Goodman (1988) there is likely to have been an increased availability of nitrogen to
348 *Lolium perenne* when it was grown with *Trifolium repens*. It has been shown that for some
349 species, e.g. *Trifolium subterraneum*, increased nitrogen supply can partially counterbalance
350 the effects of ozone exposure (Sanz et al., 2005). Some studies have shown that levels and
351 activity of Rubisco were reduced following ozone exposure (Pell et al., 1997). Increased
352 nitrogen availability may have increased turnover of the Rubisco enzyme in *L. perenne*,
353 reducing leaf senescence.

354

355 The reduced chlorophyll content of *Trifolium repens*, which corresponds with increased visible
356 injury, implies that there is a reduced capacity for photosynthesis following ozone exposure for

357 this species, which may have contributed to reduced plant growth (Hayes et al., in press). The
358 proportion of leaves showing visible injury symptoms in *Trifolium repens* varied according to
359 the position of the leaf in the plant canopy, with reduced injury in the inner canopy. This
360 corresponded with reduced stomatal conductance in the inner canopy compared to the upper
361 canopy. At the intermediate harvest, the proportion of leaves of *Trifolium repens* that had
362 visible injury symptoms was lower in the inner canopy than in the upper canopy and the
363 canopy edge. This pattern was not as pronounced at the final harvest, which may have been
364 because there was less growth between the intermediate harvest and the final harvest, resulting
365 in a more open canopy. This would allow increased light and ozone penetration into the inner
366 canopy during the second harvest interval, reducing the differences in microclimate between
367 the upper canopy/canopy edge compared to the inner canopy at the final harvest.

368

369 There was increased overall ozone leaf injury at the final harvest than at the intermediate
370 harvest (using the proportion of injured leaves, quantified by biomass), even though the
371 AOT40 value during the two harvest intervals was similar. This could have been due to the
372 more open canopy, allowing greater penetration of ozone and light. However, this effect was
373 also seen on the upper canopy and canopy edge leaves, so may have been due to a
374 cumulative/carry-over effect of ozone on the plants. Cumulative effects caused by ozone on
375 plant biomass have previously been shown for *Trifolium repens* (Fumagalli et al., 2003,
376 Nussbaum et al., 1995). In these two studies, regrowth in subsequent growth periods was
377 affected and the biomass differences were better related to the cumulative ozone than to the
378 ozone dose from an individual growth period only. However, these cumulative effects have
379 been shown only in biomass and not for visible injury on leaves produced in a subsequent
380 growth period, as in this study.

381 The structure of the canopy is also important in influencing the impact of the ozone exposure.
382 O₃(30+peaks) treated *Trifolium repens* had a more open canopy due to reduced leaf biomass
383 and the leaves curling due to ozone injury. Similarly, reduced leaf-area index of a soybean
384 (*Glycine max*) canopy has been demonstrated due to increased senescence following ozone
385 exposure (Dermody et al., 2006). Differences in leaf-area index have been related to
386 differences in penetration of PAR through plant canopies (Shulski et al., 2004). In the current
387 study the microclimate of the canopy was altered following ozone exposure and light levels of
388 the inner canopy were higher than those from the O₃(30) treatment. Other factors such as
389 temperature and windspeed may also have been affected, but were not measured. In this study,
390 the difference in stomatal conductance between the upper and inner canopy of *Trifolium repens*
391 was reduced in the O₃(30+peaks) treatment compared to O₃(30) and this corresponded to less
392 dense leaf growth giving a more open canopy in the O₃(30+peaks) treatment. This would
393 reduce the differences in microclimatic conditions between the upper and inner canopy,
394 particularly for light. Models of stomatal conductance in response to climatic conditions have
395 shown a strong influence of light (e.g. Emberson et al., 2000), and in the current study the
396 differences in stomatal conductance between the upper and inner canopy were attributed to
397 differences in light conditions rather than alterations in the relationship between stomatal
398 conductance and light. It is also possible that chronic exposure to ozone increased the
399 sluggishness of stomata of the inner canopy leaves as found in other studies (Mills et al., in
400 press; Paoletti, 2005).

401

402 Stomatal conductance of *Trifolium repens* was similar to that of *Lolium perenne*, indicating
403 that differences in sensitivity to ozone of the two species are not linked to stomatal
404 conductance. There was no evidence that the stomata of *Trifolium repens* in comparable upper
405 canopy leaves were being closed by ozone treatment, in contrast to the assumptions made by

406 Sitch et al. (2007), where models predicted further increases in atmospheric carbon dioxide
407 concentrations due to ozone induced stomatal closure. However, in this study measurements of
408 stomatal conductance were only carried out on 'non-episode days', when the ozone
409 concentration was the same (approximately 30 ppb) in the two treatments. It is possible that
410 plants may respond to high ozone concentrations by closing their stomata during the period of
411 exposure only.

412

413 This study has shown that interspecific interactions can modify the response to ozone of both
414 *T. repens* and *L. perenne*, with the direction of the interaction dependant on the species. In
415 addition, within-canopy variations in the response to ozone occur, with inner canopy leaves
416 having less response to ozone. The influence of neighbouring species and the effects these
417 species have on the canopy and microclimate should be considered in future studies. There is a
418 need for studies on more complex plant communities to further investigate whether species are
419 as sensitive to ozone as predicted from experiments on monocultures and binary mixtures, and
420 to further investigate the role of microclimate in influencing the response to ozone.

421

422 **Acknowledgements**

423 This work was funded by the Centre for Ecology and Hydrology Integrating Fund Initiative.

424

425 **References**

426

427 **Anderson CP, Hogsett WE, Plocher M, Rodecap KD, Lee EH. 2001.** Blue wild-rye grass
428 competition increases the effect of ozone on ponderosa pine seedlings. *Tree Physiology*
429 **21:**319-327.

430

431 **Ashmore MR, Buker P, Emberson LD, Terry AC, Toet S. 2007.** Modelling stomatal ozone
432 flux and deposition to grassland communities across Europe. *Environmental Pollution*
433 **146(3):659-670.**
434

435 **Bassin S, Kolliker R, Cretton C, Bertossa M, Widmer F, Bungener P, Fuhrer E. 2004.**
436 Intra-specific variability of ozone sensitivity in *Centaurea jacea* L., a potential bioindicator for
437 elevated ozone concentrations. *Environmental Pollution* **131(1):1-12.**
438

439 **Bender J, Krause GHM, Grünhage L, Jäger HJ, Köllner B, Weigel HJ. 2005.** Practical
440 and ecological considerations in ozone risk assessments. In: Wieser G, Tausz M. (Eds.)
441 Proceedings on the workshop ‘Critical Levels of Ozone: Further applying and developing the
442 flux-based concept’. Obergurgl, Austria. pp 67-71.
443

444 **Bergmann E, Bender J, Weigel HJ. 1999.** Ozone threshold doses and exposure-response
445 relationships for the development of ozone injury symptoms in wild plant species. *New*
446 *Phytologist* **144(3):423-435.**
447

448 **Chappelka AH, Neufeld HS, Davison AW, Somers GL, Renfro JR. 2003.** Ozone injury on
449 cutleaf coneflower (*Rudbeckia laciniata*) and crown-beard (*Verbesina occidentalis*) in Great
450 Smoky Mountains National Park. *Environmental Pollution* **125(1):53-59.**
451

452 **Dermody O, Long SP, DeLucia EH. 2006.** How does elevated CO₂ or ozone affect the leaf-
453 area index of soybean when applied independently? *New Phytologist* **169(1):145-155.**
454

455 **Emberson LD, Ashmore MR, Cambridge HM, Simpson D, Tuovinen JP. 2000.** Modelling
456 stomatal ozone flux across Europe. *Environmental Pollution* **109**(3):403-413.
457

458 **Emberson LD, Ashmore MR, Murray F, Kuylenstierna JCI, Percy KE, Izuta T, Zheng Y,**
459 **Shimizu H, Sheu BH, Liu CP and others. 2001.** Impacts of air pollutants on vegetation in
460 developing countries. *Water Air and Soil Pollution* **130**(1-4):107-118.
461

462 **Finkelstein PL, Davison AW, Neufeld HS, Meyers TP, Chappelka AH. 2004.** Sub-canopy
463 deposition of ozone in a stand of cutleaf coneflower. *Environmental Pollution* **131**(2):295-303.
464

465 **Fumagalli I, Mignanego L, Mills G. 2003.** Ozone biomonitoring with clover clones: yield
466 loss and carryover effect under high ambient ozone levels in northern Italy. *Agriculture*
467 *Ecosystems & Environment* **95**(1):119-128.
468

469 **Goodman PJ. 1988.** Nitrogen fixation, transfer and turnover in upland and lowland grass-
470 clover swards, using ¹⁵N isotope dilution. *Plant and Soil* **112**(2):247-254.
471

472 **Hayes F, Mills G, Harmens H, Norris D. 2007.** Evidence of widespread ozone damage to
473 vegetation in Europe (1990-2006). Cambridgeshire, UK. ISBN: 978-0-9557672-1-0.
474

475 **Hayes F, Mills G, Ashmore M. In Press.** Effects of ozone on inter- and intra-species
476 competition and photosynthesis in mesocosms of *Lolium perenne* and *Trifolium repens*.
477 *Environmental Pollution* doi 10.1016/j.envpol.2008.07.002
478

479 **Jäggi M, Ammann C, Neftel A, Fuhrer J. 2006.** Environmental control of profiles of ozone
480 concentration in a grassland canopy. *Atmospheric Environment* **40**(28): 5496-5507.
481

482 **Lantinga EA, Nassiri M, Kropff MJ. 1999.** Modelling and measuring vertical light
483 absorbtion within grass-clover mixtures. *Agricultural and Forest Meteorology* **96**:71-83.
484

485 **Lichtenthaler HK, Wellburn AR. 1983.** Determinations of total carotenoids and chlorophylls
486 *a* and *b* of leaf extracts in different solvents. *Biochemical Society Transactions* **603**:591–592.
487

488 **Mills G, Hayes F, Wilkinson S, Davies W. In press.** Chronic exposure to increasing
489 background ozone impairs stomatal functioning in grassland species. *Global Change Biology*
490

491 **Novak K, Skelly JM, Schaub M, Krauchi N, Hug C, Landolt W, Bleuler P. 2003.** Ozone
492 air pollution and foliar injury development on native plants of Switzerland. *Environmental*
493 *Pollution* **125**(1):41-52.
494

495 **Nussbaum S, Geissmann M, Fuhrer J. 1995.** Ozone exposure-response relationships for
496 mixtures of perennial ryegrass and white clover depend on ozone exposure patterns.
497 *Atmospheric Environment* **29**(9):989-995.
498

499 **Paoletti E. 2005.** Ozone slows stomatal response to light and leaf wounding in a
500 Mediterranean evergreen broadleaf, *Arbutus unedo*. *Environmental Pollution*, **134**, 439-445.
501

502 **Pell EJ, Schlagnhauser CD, Arteca RN. 1997.** Ozone-induced oxidative stress: Mechanisms
503 of action and reaction. *Physiologia Plantarum* **100**(2):264-273.

504

505 **Sanz J, Muntifering RB, Bermejo V, Gimeno BS, Elvira S. 2005.** Ozone and increased
506 nitrogen supply effects on the yield and nutritive quality of *Trifolium subterraneum*.

507 *Atmospheric Environment* **39**(32):5899-5907.

508

509 **Shulski MD, Walter-Shea EA, Hubbard KG, Yuen GY, Horst G. 2004.** Penetration of
510 photosynthetically active and ultraviolet radiation into alfalfa and tall fescue canopies.

511 *Agronomy Journal* **96**(6):1562-1571.

512

513 **Simpson D, Tuovinen JP, Emberson L, Ashmore MR. 2003.** Characteristics of an ozone
514 deposition module II: Sensitivity analysis. *Water Air and Soil Pollution* **143**(1-4):123-137.

515

516 **Sincik M, Acikgoz E. 2007.** Effects of white clover inclusion on turf characteristics, nitrogen
517 fixation, and nitrogen transfer from white clover to grass species in turf mixtures.

518 *Communications in soil science and plant analysis* **38** (13-14):1861-1877.

519

520 **Sitch S, Cox PM, Collins WJ, Huntingford C. 2007.** Indirect radiative forcing of climate
521 change through ozone effects on the land-carbon sink. *Nature* **448**(7155):791-U4.

522

523 **Tonneijck AEG, Franzaring J, Brouwer G, Metselaar K, Dueck TA. 2004.** Does
524 interspecific competition alter effects of early season ozone exposure on plants from wet
525 grasslands? Results of a three-year experiment in open-top chambers. *Environmental Pollution*

526 **131**(2):205-213.

527

528 **Utiyama M, Fukuyama T, Maruo YY, Ichino T, Izumi K, Hara H, Takano K, Suzuki H,**
529 **Aoki M. 2004.** Formation and deposition of ozone in a red pine forest. *Water Air and Soil*
530 *Pollution* **151**(1-4):53-70.
531

532 Table 1: Ozone exposure characteristics for the O₃(30) and O₃(30+peaks) treatments. Standard
 533 errors are shown in brackets

		First harvest interval	Second harvest interval
AOT40 (ppm.h)	O ₃ (30)	0.02 (0.02)	0 (0)
	O ₃ (30+peaks)	9.98 (0.10)	11.89 (0.08)
24 hour mean (ppb)	O ₃ (30)	28.0 (1.4)	27.1 (1.5)
	O ₃ (30+peaks)	41.8 (0.9)	46.2 (1.0)
12 hour mean (episode days, ppb)	O ₃ (30)	27.8 (1.4)	28.9 (1.4)
	O ₃ (30+peaks)	65.1 (0.0)	61.4 (0.0)

534

535

536 Table 2: Biomass of injured and healthy leaves of *Trifolium repens* at the intermediate and
 537 final harvests from the O₃(30) and O₃(30+peaks) treatments of plants growing in monoculture
 538 and in mixture. Standard errors are shown in brackets. ***/**/* indicates significant
 539 differences at p<0.001, p<0.01 and p<0.05 respectively.

		Intermediate harvest		Final harvest	
		Healthy (g)	Injured (g)	Healthy (g)	Injured (g)
Monoculture	O ₃ (30)	70.2 (7.0)	0.04 (0.0)	48.6 (1.9)	0 (0)
	O ₃ (30+peaks)	13.2 (1.8)	31.4 (4.1)	5.7 (0.4)	23.4 (1.6)
Mixture	O ₃ (30)	45.1 (2.0)	1.2 (0.7)	43.1 (2.7)	0 (0)
	O ₃ (30+peaks)	10.3 (0.9)	20.3 (1.7)	2.8 (1.1)	15.4 (0.1)
Significance of ozone treatment		***	***	***	**
¹ Significance of mixture vs monoculture			ns		*
^{1,2} Significance of interaction			ns		ns

540 ¹Using the proportion of injured to healthy leaves.

541 ²Significance of the interaction between whether plants are grown in monoculture or mixture
 542 and ozone treatment.

543

544 Table 3: Senescence of *Lolium perenne* at the intermediate and final harvests from the O₃(30)
 545 and O₃(30+peaks) treatments of plants growing in monoculture and in mixture. Standard
 546 errors are shown in brackets. ***, * and (*) indicate differences at p<0.001, p<0.05 and p<0.1
 547 respectively.

		Intermediate Harvest		Final Harvest	
		Senescence	Senescence	Senescence	Senescence
		(%)	(mm from	(%)	(mm from
			tip)		tip)
Monoculture	O ₃ (30)	9 (8.9)	25.8 (20.9)	28 (10.0)	28 (6.7)
	O ₃ (30+peaks)	52 (5.0)	96.7 (16.7)	68 (5.0)	74 (0.8)
Mixture	O ₃ (30)	4 (2.8)	14.0 (8.4)	0 (0)	0.5 (0.3)
	O ₃ (30+peaks)	49 (4.1)	61.0 (19.2)	28 (3.3)	45 (2.5)
Significance of ozone treatment		*	*	*	*
Significance of mixture vs monoculture		ns	***	*	***
Significance of interaction ¹		ns	ns	ns	ns

548 ¹Significance of the interaction between whether plants are grown in monoculture or mixture
 549 and ozone treatment.

550

551 Table 4: Biomass of senesced and healthy leaves of *Lolium perenne* at the intermediate and
 552 final harvests from the O₃(30) and O₃(30+peaks) treatments of plants growing in monoculture
 553 and in mixture. Standard errors are shown in brackets. **/* indicates significant differences at
 554 p<0.01 and 0.05 respectively.

		Intermediate harvest		Final harvest	
		Healthy (g)	Senesced (g)	Healthy (g)	Senesced (g)
Monoculture	O ₃ (30)	16.3 (4.8)	1.4 (0.3)	5.8 (4.1)	1.5 (1.1)
	O ₃ (30+peaks)	10.0 (2.9)	5.8 (1.4)	1.2 (0.3)	2.0 (0.2)
Mixture	O ₃ (30)	12.6 (2.1)	2.5 (0.4)	4.6 (4.1)	0 (0)
	O ₃ (30+peaks)	11.5 (3.1)	4.8 (0.9)	3.7 (1.7)	1.8 (0.2)
Significance of ozone treatment		ns	ns	ns	**
Significance of mixture vs monoculture		ns	*	ns	ns
Significance of interaction ¹		ns	ns	ns	ns

555 ¹ Significance of the interaction between whether plants are grown in monoculture or mixture
 556 and ozone treatment.

557

558 Table 5: Stomatal conductance and PAR of *Trifolium repens* (monoculture) leaves from the
 559 inner and upper canopy. Standard errors are shown in brackets. ** and * indicate significant
 560 differences between ozone treatments at $p < 0.01$ and $p < 0.05$ respectively.

	Inner Canopy		Upper Canopy	
	O ₃ (30)	O ₃ (30+peaks)	O ₃ (30)	O ₃ (30+peaks)
Stomatal Conductance (mmol m ⁻² s ⁻¹)	66 (7)	119 (7) *	338 (44)	291 (9)
PAR (μmol m ⁻² s ⁻¹)	94 (13)	220 (0) **	814 (176)	951 (13)

561

562

563 Figure 1: Percentage of injured leaves (determined by biomass) of *Trifolium repens* in
564 different regions of the canopy at the intermediate harvest (A) and final harvest (B) from the
565 O₃(30+peaks) treatment of plants growing in monoculture and in mixture. Bars are standard
566 errors. ** indicates a significant difference at p<0.01.

567

568 Figure 2: Percentage of senesced leaves (determined by biomass) of *Lolium perenne* in
569 different regions of the canopy at the intermediate harvest (A) and final harvest (B) from the
570 O₃(30+peaks) treatment of plants growing in monoculture and in mixture. Bars are standard
571 errors. * indicates a significant difference at p<0.05.

572

573 Figure 3: Chlorophyll content of leaves from the inner and upper canopy of *Trifolium repens*
574 exposed to O₃(30) or O₃(30+peaks). Bars are standard errors. * indicates significant
575 differences at p<0.05.

576

577 Figure 4: Chlorophyll content of leaves of *Trifolium repens* exposed to O₃(30) or
578 O₃(30+peaks). Leaves were numbered from Leaf 1 (newest fully expanded leaf) to Leaf 3 (3rd
579 newest fully expanded leaf). Bars are standard errors. * indicates significant differences at
580 p<0.05.